

## Whole-Genome Sequence of Mycoplasma bovis Strain Ningxia-1

Peng Sun,<sup>a,b</sup> Haifeng Luo,<sup>a</sup> Xin Zhang,<sup>a</sup> Jingyi Xu,<sup>a</sup> Yanan Guo,<sup>a</sup> Shenghu He<sup>a</sup>

<sup>a</sup>School of Agriculture, Ningxia University, Yinchuan, China

<sup>b</sup>Virginia-Maryland Regional College of Veterinary Medicine, University of Maryland–College Park, College Park, Maryland, USA

**ABSTRACT** A genome sequence of the *Mycoplasma bovis* Ningxia-1 strain was tested by Pacific Biosciences (PacBio) single-molecule real-time (SMRT) sequencing technology. The strain was isolated from a lesioned calf lung in 2013 in Pengyang, Ningxia, China. The single circular chromosome of 1,033,629 bp shows differences between complete *Mycoplasma bovis* genome in insertion-like sequences (ISs), integrative conjugative elements (ICEs), lipoproteins (LPs), variable surface lipoproteins (VSPs), pathogenicity islands (PAIs), etc.

Mycoplasma bovis is the main cause of bovine respiratory disease syndrome. At present, the pathogen is prevalent worldwide, which causes huge economic losses to national cattle industries (1). In 2008, *M. bovis* was isolated from beef cattle in Hubei, China (2). Other places in China, such as Xinjiang, Ningxia, Chongqing, Guizhou, and Qingdao, later reported *M. bovis* isolated from beef cattle and dairy cows (3).

The whole-genome sequence of the *M. bovis* Ningxia-1 strain was isolated in 2013 from the lesioned lung of a beef calf, which was tested using single-molecule real-time (SMRT) technology (4), resulting in approximately 1,262-fold final sequence coverage. The genome of *M. bovis* Ningxia-1 contains a single circular chromosome of 1,033,629 bp, with a GC content of 29.33%. A total of 754 open reading frames (ORFs) were identified, totaling 845,928 bp (maximum length, 9,981 bp; minimum length, 114 bp), and occupied 81.84% of the whole genome, with an average length of 1121 bp and a mean GC content of 29.77%. A total of 577 coding sequence (CDS) genes could be classified into clusters of orthologous groups (COG) families, which have 19 functional categories. Seventy-four pseudogenes were predicted by GeneMarkS+ (5). The genome encodes 6 rRNA and 34 tRNA genes, representing all 20 amino acids.

We found 60 insertion-like sequence (IS) elements that comprised three distinct categories (https://www-is.biotoul.fr). A cluster of 8 variable surface protein (VSP)-related ORFs were found in the genome. Sixty-six lipoproteins (LPs) revealed signatures denoting distinct mutation-based mechanisms of phase variation (6). Of three predicted pathogenicity islands (PAIs) (7) with a lower GC content (8), PAI-1 (463,228 to 485,927) encoded one single-stranded DNA-binding protein and one conjugal transfer protein, TraE, and contained a transposase at the 5' end. PAI-2 (541731 to 556809) presented an IS4 and IS30 family transposase at the 3' end and contained an IS1634 family transposase gene. A total of two nucleotidyl transferase AbiEii/AbiGii toxin family proteins and abortive phage infection proteins in PAI-3 (968,698 to 977,477) were without an IS element but still denoted a pathogenicity island. These characteristics may contribute to the emergence of bacterial pathogens with new virulence properties (9).

**Accession number(s).** This whole-genome sequence assembly has been deposited at GenBank under the accession no. CP023663.

December 2017 **Published** 25 January 2018 **Citation** Sun P, Luo H, Zhang X, Xu J, Guo Y, He S. 2018. Whole-genome sequence of

Received 29 November 2017 Accepted 5

Mycoplasma bovis strain Ningxia-1. Genome Announc 6:e01367-17. https://doi.org/10.1128/ genomeA.01367-17. **Copyright** © 2018 Sun et al. This is an open-

access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Shenghu He, heshenghu308@163.com.

## ACKNOWLEDGMENTS

This work was sponsored by the China Science and Technology of Ningxia support program (grant 412-0164).

We thank Wei Yangyang from GeneDenovo, Guangzhou, China, for his assistance with data analysis.

## REFERENCES

- Tamang MD, Nam HM, Jang GC, Kim SR, Chae MH, Jung SC, Byun JW, Park YH, Lim SK. 2012. Molecular characterization of extended-spectrum-betalactamase-producing and plasmid-mediated AmpC beta-lactamaseproducing *Escherichia coli* isolated from stray dogs in South Korea. Antimicrob Agents Chemother 56:2705–2712. https://doi.org/10.1128/AAC .05598-11.
- Qi J, Guo A, Cui P, Chen Y, Mustafa R, Ba X, Hu C, Bai Z, Chen X, Shi L, Chen H. 2012. Comparative geno-plasticity analysis of *Mycoplasma bovis* HB0801 (Chinese isolate). PLoS One 7:e38239. https://doi.org/10.1371/ journal.pone.0038239.
- Zhou W, Li N, Wei X, Hu J, Xia T, Wang Z, Peng Y. 2012. The diagnosis of mycoplasma pneumoniae in a beef cattle farm in Chongqing, China. Prev Vet Med 34:326–328.
- Bleidorn C. 2016. Third generation sequencing: technology and its potential impact on evolutionary biodiversity research. Syst Biodivers 14: 1–8. https://doi.org/10.1080/14772000.2015.1099575.

- Besemer J, Lomsadze A, Borodovsky M. 2001. GeneMarkS: a self-training method for prediction of gene starts in microbial genomes. Implications for finding sequence motifs in regulatory regions. Nucleic Acids Res 29:2607–2618. https://doi.org/10.1093/nar/29.12.2607.
- Wise KS, Calcutt MJ, Foecking MF, Röske K, Madupu R, Methé BA. 2011. Complete genome sequence of *Mycoplasma bovis* type strain PG45 (ATCC 25523). Infect Immun 79:982–983. https://doi.org/10.1128/IAI.00726-10.
- Schmidt H, Hensel M. 2004. Pathogenicity islands in bacterial pathogenesis. Clin Microbiol Rev 17:14–56. https://doi.org/10.1128/CMR.17.1.14-56 .2004.
- Li Y, Zheng H, Liu Y, Jiang Y, Xin J, Chen W, Song Z. 2011. The complete genome sequence of *Mycoplasma bovis* strain Hubei-1. PLoS One 6:e20999. https://doi.org/10.1371/journal.pone.0020999.
- 9. Lee CA. 1996. Pathogenicity islands and the evolution of bacterial pathogens. Infect Agents Dis 5:1–7.