RESEARCH





A comprehensive assessment of urinary iodine concentration and thyroid hormones in New Zealand schoolchildren: a cross-sectional study

Sheila A Skeaff^{1*}, Christine D Thomson¹, Noela Wilson² and Winsome R Parnell¹

Abstract

Background: Insufficient iodine in children's diets is of concern because thyroid hormones are needed for normal growth and development, particularly of the brain. This study aimed to carry out a comprehensive assessment of the iodine status of New Zealand schoolchildren using a range of biochemical indices suitable for populations (i.e. urinary iodine concentration) and individuals (i.e. thyroid hormones).

Methods: The New Zealand National Children's Nutrition Survey was a cross-sectional survey of a representative sample of schoolchildren aged 5-14 years. Children were asked to provide a casual urine sample for the determination of urinary iodine concentration (UIC) and a blood sample for the determination of thyroglobulin (Tg), Thyroid Stimulating Hormone (TSH), free thyroxine (fT4) and free triiodothyronine (fT3).

Results: The median UIC was 68 μ g/L (n = 1153), which falls between 50-99 μ g/L indicative of mild iodine deficiency. Furthermore, 29% of children had an UIC <50 μ g/L and 82% had an UIC <100 μ g/L. The median Tg concentration was 12.9 μ g/L, which also falls between 10.0-19.9 μ g/L indicative of mild iodine deficiency. The Tg concentration of children with an UIC <100 μ g/L was 13.9 μ g/L, higher than the 10.3 μ g/L in children with an UIC <100 μ g/L (P = 0.001). The mean TSH (1.7 mU/L), fT4 (14.9 pmol/L), and fT3 (6.0 pmol/L) concentrations for these mildly iodine deficient New Zealand children fell within normal reference ranges.

Conclusions: The UIC and Tg concentration indicate that New Zealand schoolchildren were mildly iodine deficient according to WHO/UNICEF/ICCIDD, and both are suitable indices to assess iodine status in populations or groups. The normal concentrations of TSH, fT4 and fT3 of these children suggest that these thyroid hormones are not useful indices of mild iodine deficiency.

Keywords: lodine, lodine deficiency, Urinary iodine concentration, Children, Thyroid hormones

Background

New Zealand has a long history of iodine deficiency, with endemic goiter first reported in the 1920s. This is not surprising given that New Zealand soils are low in iodine and, despite being surrounded by the ocean, the typical diet contains small quantities of fish and other seafood. It was the work of Hercus and Purves who compared the median urinary iodine excretion from nine goitrous regions of New Zealand (i.e. 24-62 μ g/day) with non-goitrous parts of Australia (i.e. 80-147 μ g/day) and the Pacific Island of Western Samoa (i.e. 172 μ g/day), which finally prompted

¹Department of Human Nutrition, University of Otago, Dunedin 9054, New Zealand the iodization of table salt to 50 mg/kg in 1939 [1]. By the 1950s, the incidence of goiter had fallen from 20-30% to less than one percent [2]. Because iodized salt was not used in manufactured foods, the regular consumption of dairy products (contaminated by iodophors used in milk processing) and iodized salt added at the table and in cooking is believed to be responsible for the adequate iodine intakes reported in New Zealand between the 1960s and the 1980s with urinary iodine excretion ranging from 120-220 μ g/day [3-6].

Iodine deficiency is defined by the World Health Organization (WHO) as a population median urinary iodine concentration (UIC) that falls below 100 μ g/L, while a median UIC of 50–99 μ g/L, 20-49 μ g/L, and <20 μ g/L



© 2012 Skeaff et al.; licensee BioMed Central Ltd. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/2.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

^{*} Correspondence: sheila.skeaff@otago.ac.nz

Full list of author information is available at the end of the article

indicates mild, moderate, and severe iodine deficiency, respectively [7]. Mild iodine deficiency has re-emerged in Australia [8], and most recently, the United Kingdom [9], caused by a change in food patterns. In the 1990s there were reports in New Zealand of iodine deficiency in volunteer samples of adults [10-12] and in children aged 8-10 years (yr) living in two cities [13]. A lack of iodine in the diets of children is of concern, as sufficient quantities of thyroid hormones are needed for normal growth and development, particularly of the brain.

The measurement of the UIC in a single or casual urine sample is the most commonly used index to assess iodine status in schoolchildren. In addition, blood constituents such as thyroid stimulating hormone (TSH), thyroglobulin (Tg), thyroxine (T4), and tri-iodothyronine (T3) can also be measured. Severe to moderate iodine deficiency lowers circulating T4 concentrations, resulting in a subsequent rise in TSH. If iodine deficiency becomes severe enough, a decrease in T3 concentration may also be seen. Although a median UIC between 50-100 µg/L is used to classify mild iodine deficiency, the thyroid hormone concentrations of such populations typically fall within normal reference ranges. There is growing evidence that Tg is a more sensitive index of mild iodine deficiency than TSH, T4, and T3 [14-16], however, Tg is not routinely measured in cross-sectional surveys. The aim of this study was to assess the iodine status of a representative sample of New Zealand schoolchildren using a range of biochemical indices.

Subjects and methods

The Children's Nutrition Survey was a cross-sectional survey of a nationally representative sample of New Zealand school children aged 5-14 year conducted between March and December 2002. Children were categorised into three ethnic groups based on parental report; Māori, Pacific, and New Zealand European & all Other ethnicities (NZEO). A two-stage school-based sampling frame was used to recruit children with over-sampling of Māori and Pacific children to allow for ethnic specific analysis. In brief, schools were randomly selected from the Ministry of Education rolls and children from these schools were randomly selected in proportion to the number of children on the school roll. Socio-demographic data, height, weight, and a 24-hr diet recall were collected from participating children; because the iodine content of foods was neither complete nor reliable in the New Zealand Food Composition Database, iodine intakes were not determined. Full ethical approval was obtained from 13 regional health ethics committees. Informed written consent was obtained from parents or guardians of the children. Further details of the survey methodology and a summary of the key results can be found elsewhere [17]. Blood and urine samples were collected from a subset of urban children who agreed and whose parents consented

to the sampling; children at the 66 rural schools were not included due to concerns that the length of time required to transport samples to the laboratory would affect blood stability. Sampling weights were based on the provision of a blood sample and not on participation in the survey. Non-fasting venous blood samples were drawn into vacutainers (Becton Dickinson, Frankton Lakes, NJ, USA), refrigerated immediately after collection, and couriered to a central laboratory for processing the following morning. After centrifugation, serum and plasma samples were stored at -80° C until analysis. Casual or spot urine samples were collected from children during school hours, refrigerated, and couriered to the Department of Human Nutrition, University of Otago, the next day. Urine samples were stored at -20° C until analysis.

UIC was determined using a modification of the method of Pino et al. [18] at the Department of Human Nutrition, University of Otago. An external reference standard (Seronorm Trace Elements Urine, Sero As, Norway) was analysed with each batch of samples giving a mean iodine concentration of 93 μ g/L (expected: 97 μ g/L) with a coefficient of variation (CV) of 8.4% (n = 213).

Serum TSH was determined by Southern Community Laboratories using an ADVIA Centaur two-site sandwich immunoassay (Bayer Health Care Diagnostika, Vienna, Austria). The reference range was 0.10-150 mU/L. Analysis of a supplied external reference standard (Ligand Plus) resulted in a CV of 5.1% (n = 291). Serum Tg concentration was determined using a DiaSorin radioimmunoassay (DiaSorin, Saluggia, Italy) in the Department of Human Nutrition, University of Otago. The reference range was 0.3-500 µg/L. Analysis of a pooled internal control gave a CV of 8.5% (n = 20). Because these serum samples had been used to assess other nutrients prior to iodine, we measured the effect of up to four freeze-thaw cycles on serum TSH and Tg and found there were no differences (n = 18).

Plasma free T4 (fT4) and free T3 (fT3) concentrations were determined by Southern Community Laboratories using an ADVIA Centaur fT4 and fT3, respectively, competitive chemiluminescent immunoassay (Bayer Health Care Diagnostika, Vienna, Austria). Plasma was used to determine fT4 and fT3 concentrations as there was insufficient serum for analysis of all four blood constituents (i.e. Tg, TSH, fT4 and fT3). Prior to analysis for this study, we tested the recovery of fT4 and fT3 in plasma, which was 100-106% and 97-107%, respectively, compared to serum values obtained from the same subjects. The range of detection of the fT4 assay was 1.3-155 pmol/L and that of the fT3 assay was 0.3-30.8 pmol/L. Analysis of an external reference standard (Ligand Plus) resulted in a CV of 8.1% (n = 261) and 5.7% (n = 297) for fT4 and fT3 assays, respectively.

All statistical analyses were carried out using the statistical package STATA 11.1 (StataCorp LP. College Station, TX, USA). We used survey commands to account for the survey design, weighting, and clustering. Socioeconomic status was determined by grouping children using the New Zealand Index of Deprivation scale, an index based on the residential address of the child [19]. The scale is divided into five quintiles, with quintile 1 representing the least deprived area and quintile 5 the most deprived area. Children allocated to quintile 1 and 2 were categorized as high socioeconomic status, children allocated to quintile 3 and 4 were categorized as medium socioeconomic status, and children allocated to guintile 5 as low socioeconomic status. Log transformation was carried out for data that was not normally distributed (i.e. UIC, TSH and Tg data) and geometric means reported. Linear regression analysis was used to determine the effect of age (5-7 yr, 8-10 yr, 11-14 yr), sex (boy, girl), ethnicity (Māori, Pacific, NZEO), socioeconomic status (high, medium, low), and UIC (<50 $\mu g/L$ and <100 µg/L) on Tg, TSH, fT4, and fT3. Bonferroni adjusted P-values are reported for post-hoc comparisons. All tests were two-sided and the level of significance set at P < 0.05.

Results

Two hundred schools were asked to participate in the survey and 172 agreed; of the 4728 children recruited, 3275 participated. A urine sample was collected from 1796 children and a blood sample from 1927 children. Serum samples were used to determine the concentration of a number of other nutrients (i.e. 25-hydroxyvitamin D, selenium) prior to the analysis of the thyroid hormones; surplus serum or plasma was available for complete thyroid hormone (i.e. TSH, Tg, fT4 and fT3) analysis from 1212 children. The Deprivation Index (i.e. socioeconomic status) of 59 children was not known and these children were excluded from the analysis, resulting in a final sample size of 1153 children for which both urinary iodine and thyroid hormone data are available. There were no statistically significant differences between those children who gave a blood sample and those who did not by sex, age, socioeconomic status, and ethnicity.

The median (25th, 75th percentile) UIC of New Zealand children was 68 (50, 95) μ g/L (Table 1), which falls within the range 50–99 μ g/L that WHO/UNICEF/ICCIDD categorize as mild odine deficiency [7]. Furthermore, 29% of children had a UIC <50 μ g/L and 82% had UIC <100 μ g/L, above than the 20% and 50%, respectively, recommended by WHO/UNICEF/ICCIDD [7]. UIC did not differ by sex, age, socioeconomic status, or ethnicity. The median (25th, 75th percentile) Tg concentration for New Zealand children was 12.9 (8.1, 19.9) μ g/L, which falls between 10.0-19.9 μ g/L also indicative of mild iodine

deficiency according to WHO/UNICEF/ICCIDD (Table 1) [7]. Tg concentration did not differ by sex, socioeconomic status, or ethnicity (Table 2). There was a significant effect of age, such that increasing age category decreased Tg concentration by 14% (P < 0.001). Children 5-7 yr had a higher (geometric) mean Tg concentration than children 11-14 yr (16.3 vs 11.2 μ g/L; *P* = 0.002) and children 8-10 yr had a higher (geometric) mean than children 11-14 yr (13.8 vs 11.2 μ g/L; P = 0.031); there was no significant difference in the (geometric) mean Tg concentration between 5-7 yr and 8-10 yr children. The concentration of Tg at all percentiles was higher in children 5-7 yr compared to older children (Table 3). The (geometric) mean Tg of children with a UIC <50 µg/L was significantly higher than in children with an UIC $\geq 50 \ \mu g/L$ (15.6 vs 12.3 $\mu g/L$; P = 0.001) (Table 4). The (geometric) mean Tg of children with an UIC $<100 \mu g/L$ was significantly higher than in children with an UIC $\geq 100 \ \mu g/L$ (13.9 vs 10.3 $\mu g/L$; P < 0.001) (Table 4). The (geometric) mean (95% Confidence Interval (CI)) TSH concentration of the children was 1.7 (1.6-1.8) mU/L (Table 1). There were no significant differences in TSH concentration with regard to sex, age, socioeconomic category, or ethnicity (Table 2). There were also no differences in TSH concentration between children with an UIC <50 µg/L or UIC <100 µg/L (Table 4).

The mean (95% CI) fT4 concentration for New Zealand children was 14.9 (14.6-15.2) pmol/L (Table 1); fT4 concentration did not differ by sex and socioeconomic status (Table 5). There was a significant effect of age, such that increasing age category resulted in a 0.49 pmol/L decrease in fT4 concentration (Table 5). The mean fT4 concentration of children aged 5-7 yr (15.4 pmol/L) was significantly higher than children aged 11-14 yr (14.4 pmol/L; P = 0.013), but was not different from children aged 8-10 yr (15.0 pmol/L; P = 0.441); there was no difference in the fT4 concentration between children aged 8-10 yr and 11-14 yr (P = 0.146). There was also a significant effect of ethnicity (P = 0.002) (Table 5). NZEO children had a significantly lower mean fT4 concentration (14.6 pmol/L) than Māori children (15.4 pmol/L; P = 0.010) or Pacific children (15.6 pmol/L; P = 0.002); there was no significant difference between the fT4 concentrations of Māori or Pacific children. The 2.5th percentile of fT4 was highest in children aged 5-7 yrs but lower in NZEO children compared to Māori or Pacific children (Table 3). The mean (95% Confidence Interval) fT3 concentration for New Zealand children was 6.0 (5.9-6.1) pmol/L (Table 1); fT3 concentration did not differ by sex, age, socioeconomic status and ethnicity (Table 5). The mean fT3 concentration of children with an UIC $<50 \mu g/L$ was significantly higher than in children with an UIC \geq 50 µg/L (6.1 vs 5.9 pmol/L; P = 0.002) (Table 4). The (geometric) mean Tg of children with an UIC $<100 \mu g/L$ was significantly higher

	UIC	UIC (µg/L)		Tg (μg/L)		TSH (mu/L)		Free T4 (pmol/L)		Free T3 (pmol/L)	
n	Median	25th,75th	Median	25th,75th	Mean	95% Cl	Mean	95% Cl	Mean	95% Cl	
1153	68	50,95	12.9	8.1,19.9	1.7	1.6,1.8	14.9	14.6,15.2	6.0	5.9,6.1	
611	69	50,94	12.6	17.9,19.2	1.7	1.6,1.8	14.8	14.5,15.1	6.0	5.9,6.0	
542	67	55,96	13.4	8.5,20.5	1.7	1.6,1.9	15.0	14.5,15.4	6.0	5.9,6.1	
315	63	50,88	16.2	10.4,24.2	1.6	1.5,1.8	15.4	14.9,15.8	6.0	5.9,6.1	
438	67	50,94	12.5	8.0,19.0	1.8	1.7,1.9	15.0	14.6,15.5	6.0	5.9,6.1	
400	75	51,106	11.1	7.4,16.4	1.7	1.6,1.8	14.4	13.9, 14.9	5.9	5.9,6.1	
567	70	50,98	12.8	8.0,19.9	1.7	1.6,1.8	15.2	14.7,15.7	6.0	5.9,6.2	
347	68	50,93	12.4	8.0,20.4	1.6	1.5,1.7	15.0	14.5,15.5	5.9	5.9,6.0	
239	66	50,93	13.2	9.0,19.2	1.8	1.7,1.9	14.6	14.1,15.1	6.0	5.9,6.2	
333	66	50,93	13.5	8.8,21.2	1.7	1.6,1.8	14.6	14.3,15.0	6.0	5.9,6.1	
338	67	50,87	13.0	8.3,20.1	1.6	1.5,1.8	15.4	14.9,15.9	6.0	5.9,6.1	
482	73	50,105	12.4	7.8,18.0	1.7	1.6,1.8	15.6	15.2,16.1	6.0	5.9,6.2	
	n 1153 611 542 315 438 400 567 347 239 333 338 482	N Median 1153 68 611 69 542 67 315 63 438 67 400 75 567 70 347 68 239 66 333 66 338 67 482 73	UIC (μg/L)Median25th,75th11536850,956116950,945426755,963156350,884386750,944007551,1065677050,983476850,933336650,933386750,874827350,105	$\begin{tabular}{ c c c } & UIC (\mu g/L) & Tg \\ \hline Median & 25th,75th & Median \\ 1153 & 68 & 50,95 & 12.9 \\ \hline 611 & 69 & 50,94 & 12.6 \\ 542 & 67 & 55,96 & 13.4 \\ \hline 315 & 63 & 50,88 & 16.2 \\ 438 & 67 & 50,94 & 12.5 \\ 400 & 75 & 51,106 & 11.1 \\ \hline 567 & 70 & 50,98 & 12.8 \\ 347 & 68 & 50,93 & 12.4 \\ 239 & 66 & 50,93 & 13.2 \\ \hline 333 & 66 & 50,93 & 13.5 \\ 338 & 67 & 50,87 & 13.0 \\ 482 & 73 & 50,105 & 12.4 \\ \hline \end{tabular}$	UIC (μg/L) Tg (μg/L) Median 25th,75th Median 25th,75th 1153 68 50,95 12.9 8.1,19.9 611 69 50,94 12.6 17.9,19.2 542 67 55,96 13.4 8.5,20.5 315 63 50,88 16.2 10.4,24.2 438 67 50,94 12.5 80,19.0 400 75 51,106 11.1 7.4,16.4 567 70 50,98 12.8 80,19.9 347 68 50,93 12.4 80,20.4 333 66 50,93 13.2 90,19.2 333 66 50,93 13.5 8.8,21.2 338 67 50,87 13.0 8.3,20.1 482 73 50,105 12.4 7.8,180	UIC (μg/L) Tg (μg/L) TSH Median 25th,75th Median 25th,75th Median 25th,75th Mean 1153 68 50,95 12.9 8.1,19.9 1.7 611 69 50,94 12.6 17.9,19.2 1.7 542 67 55,96 13.4 8.5,20.5 1.7 315 63 50,88 16.2 10.4,24.2 1.6 438 67 50,94 12.5 8.0,19.0 1.8 400 75 51,106 11.1 7.4,16.4 1.7 567 70 50,98 12.8 8.0,19.9 1.7 347 68 50,93 12.4 8.0,20.4 1.6 239 66 50,93 13.2 9.0,19.2 1.8 333 66 50,87 13.0 8.3,20.1 1.6 482 73 50,105 12.4 7.8,18.0 1.7	UIC ($\mu\mu$ /L)Tg ($\mu\mu$ /L)TSH (mu/L)nMedian25th,75thMedian25th,75thMean95% Cl11536850,9512.98.1,19.91.71.6,1.86116950,9412.617.9,19.21.71.6,1.85426755,9613.485,20.51.71.6,1.93156350,8816.210.4,24.21.61.5,1.84386750,9412.58.0,19.01.81.7,1.94007551,10611.17.4,16.41.71.6,1.83476850,9312.48.0,20.41.61.5,1.73336650,9313.29.0,19.21.81.7,1.93336650,9313.58.8,21.21.71.6,1.83386750,8713.08.3,20.11.61.5,1.84827350,10512.47.8,18.01.71.6,1.8	UIC (µg/L) Tg (µg/L) TSH (mu/L) Free T n Median 25th,75th Median 25th,75th Mean 95% CI Mean 1153 68 50,95 12.9 8.1,19.9 1.7 1.6,1.8 14.9 611 69 50,94 12.6 17.9,19.2 1.7 1.6,1.8 14.8 542 67 55,96 13.4 8.5,20.5 1.7 1.6,1.9 15.0 315 63 50,88 16.2 10.4,24.2 1.6 1.5,1.8 15.4 438 67 50,94 12.5 8.0,19.0 1.8 1.7,1.9 15.0 400 75 51,106 11.1 7.4,16.4 1.7 1.6,1.8 15.2 347 68 50,93 12.4 8.0,19.9 1.7 1.6,1.8 15.2 333 66 50,93 13.2 9.0,19.2 1.8 1.7,1.9 14.6 333 67 50,87 13.0 8.3,20.	UIC ($\mu g/L$)Tg ($\mu g/L$)TSH ($\dots L$)Free T4 ($\dots d/L$)nMedian25th,75thMean95% CIMean95% CI11536850,9512.98.1,19.91.71.6,1.814.914.6,15.26116950,9412.617.9,19.21.71.6,1.814.814.5,15.15426755,9613.48.5,20.51.71.6,1.814.814.5,15.43156350,8816.2104,24.21.61.5,1.815.414.9,15.84386750,9412.580,19.01.81.7,1.915.014.6,15.54007551,10611.17.4,16.41.71.6,1.815.214.7,15.75677050,9812.880,19.91.71.6,1.815.214.7,15.73476850,9312.480,20.41.61.5,1.715.014.5,15.52396650,9313.29.0,19.21.81.7,1.914.614.1,15.13336650,9313.588,21.21.71.6,1.814.614.3,15.03386750,8713.083,20.11.61.5,1.815.414.9,15.93386750,8713.083,20.11.61.5,1.815.414.9,15.94827350,10512.47.8,18.01.71.6,1.815.615.2,1.6	UIC ($\mu g/L$)Tg ($\mu g/L$)TSH ($m u/L$)Free T ($\mu m n/L$)Free T 3nMedian25th,75thMead95% ClMean95% ClMean11536850,9512.98.1,19.91.71.6,1.814.914.6,15.26.06116950,9412.617.9,19.21.71.6,1.814.814.5,15.16.05426755,9613.48.5,20.51.71.6,1.814.814.5,15.46.03156350,8816.210.4,24.21.61.5,1.815.414.9,15.86.04386750,9412.58.0,19.01.81.7,1.915.014.6,15.56.04007551,10611.17.4,16.41.71.6,1.814.413.9,14.95.95677050,9812.88.0,19.91.71.6,1.814.413.9,14.95.95476850,9312.48.0,20.41.61.5,1.715.014.5,15.55.92396650,9313.58.8,21.21.71.6,1.814.614.3,15.06.03336650,9313.58.8,21.21.71.6,1.814.614.3,15.06.03386750,8713.08.3,20.11.61.5,1.815.414.9,15.96.04827350,10512.47.8,18.01.71.6,1.815.615.2,1.66.0	

Table 1 Urinary iodine (UIC), Thyroglobulin (Tg), TSH, free T4, and free T3 concentration by sex, age, socioeconomic status, and ethnicity in New Zealand children

All data adjusted for sampling weights and presented as median with 25^{th} ,75th percentile or mean with 95% Confidence Interval; geometric mean given for TSH. Tg concentration of 5-7 yr was higher than 11-14 yr (P = 0.002), and Tg concentration of 8-10 yr was higher than 11-14 yr (P = 0.031). Free T4 concentration of 5-7 yr higher than 11-14 yr (P = 0.013). Free T4 concentration of NZEO lower than Māori (P = 0.010) and Pacific (P = 0.002). Socioeconomic status = SES, New Zealand European and Other Ethnicities = NZEO.

than in children with an UIC $\geq 100 \ \mu g/L$ (6.0 vs 5.9 pmol/L; P = 0.001) (Table 4).

Discussion

The median UIC of children in this study was 68 μ g/L, which falls between 50–99 μ g/L, and according to WHO/UNICEF/ICCIDD, indicates that New Zealand children were mildly iodine deficient. Mild iodine deficiency has also been reported in Australian schoolchildren with a median UIC ranging from 74-143 μ g/L [8], and more recently in 14-15 yr girls from the United Kingdom with a median UIC of 80 μ g/L [9]. In a smaller study in New Zealand on a random sample of 300 schoolchildren 8-10 yr, a similar median UIC of 66 μ g/L was found [13]. In this earlier study, blood samples were not obtained from children but thyroid volume was

Table 2 Effect of sex, age, socioeconomic status and ethnicity on Thyroglobulin (Tg) and TSH concentration

	Тд		TSH		
	Ratio (95% Cl) ^a	Р	Ratio (95% Cl)	Р	
Sex	1.14 (0.96,1.36)	0.134	1.03 (0.96,1.13)	0.366	
SES	1.03 (0.97,1.11)	0.415	1.03 (0.98,1.07)	0.270	
Age Category	0.86 (0.78,0.93)	0.000	1.02 (0.96,1.08)	0.596	
Ethnicity	1.05 (0.98,1.13)	0.142	1.02 (0.99, 1.06)	0.212	

^aRatio (95% Confidence Interval) of log-transformed data by regression analysis.

Socioeconomic Status = SES.

measured by ultrasonography. Using the 1997 cut-offs for thyroid volume, which were the only available reference data at that time [20], 10% of the children in this smaller study had goiter; when the data were reanalyzed using revised cut-offs for thyroid volume [21], approxi-

Table 3 Thyroglobulin (Tg), TSH, free T4, and free T3 concentrations by percentile in New Zealand children

	Percentile								
	n	2.5	5	10	50	90	95	97.5	
Tg (µg/L)	1153	2.9	4.3	5.4	12.9	28.8	34.5	40.9	
5-7 yrs	315	4.0	5.3	6.1	16.2	32.5	40.4	46.7	
8-10 yrs	438	2.7	4.3	5.4	12.5	26.7	33.3	33.7	
11-14 yrs	400	2.8	3.8	5.0	11.1	26.2	33.7	37.9	
TSH (mU/L)	1153	0.7	0.8	1.0	1.7	2.9	3.5	4.1	
Free T4 (pmol/L)	1153	11.7	12.2	12.7	15.0	18.2	19.3	20.3	
5-7 yrs	315	12.4	12.7	13.2	15.3	18.5	20.0	20.9	
8-10 yrs	551	12.0	12.4	12.9	15.2	18.4	19.0	20.3	
11-14 yrs	400	11.1	11.6	12.4	14.4	17.2	18.2	19.7	
NZEO	333	11.2	12.0	12.4	14.4	17.3	18.5	20.3	
Māori	338	11.7	12.4	12.8	15.2	18.4	19.3	20.2	
Pacific	482	11.7	12.3	12.9	15.4	18.5	19.5	20.6	
Free T3 (pmol/L)	1153	4.9	5.1	5.3	6.0	6.7	7.0	7.1	

For each indice, data is presented for entire sample and by category only for those variables shown to be significant in regression analysis. New Zealand European and Other Ethnicities = NZEO.

	T ((1)		T CU (11/1)		E T (10)		F T2 (14)	
	Ig (μg/L)	Р	ISH (mU/L)	Р	Free 14 (pmol/L)	Р	Free 13 (pmol/L)	Р
	Mean (95% Cl)		Mean (95% Cl)		Mean (95% Cl)		Mean (95% Cl)	
UIC < 50 (µg/L)	15.6 (14.0,17.3)	0.001	1.7 (1.6, 1.9)	0.669	14.7 (14.3,15.2)	0.271	6.1 (6.0,6.2)	0.002
UIC ≥ 50 (µg/L)	12.3 (11.1, 13.5)		1.7 (1.6, 1.8)		15.0 (14.6, 15.3)		5.9 (5.8, 6.0)	
UIC < 100 (µg/L)	13.9 (12.8, 14.9)	0.000	1.7 (1.6, 1.8)	0.236	14.9 (14.5, 15.2)	0.630	6.0 (6.0, 6.1)	0.001
UIC ≥ 100 (µg/L)	10.3 (8.8, 12.1)		1.8 (1.6,1.9)		15.0 (14.5, 15.5)		5.9 (5.8, 6.0)	

Table 4 Concentration of thyroid indices if urinary iodine concentration (UIC) <50 µg/L or ≥50 µg/L and <100 µg/L or >100 µg/l

Tg and TSH values presented as geometric mean (95% Confidence Interval) and free T4 and T3 values presented as mean (95% Confidence Interval) as determined by regression analysis. Values of 50 µg/L and 100 µg/L based on ICCIDD/UNICEF/WHO [7] recommended cut-offs for UIC. Thyroglobulin = Tg.

mately 30% of the children were classified as having goiter [22]. Although thyroid volume was not measured in the national survey reported here, the similar method of recruitment and median UIC between the earlier study of 8-10 yr olds and this survey suggests that up to a third of New Zealand children would have had goiter in 2002.

The median Tg concentration of children in this survey of 12.9 μ g/L falls within the range of 10.0-19.9 μ g/L, and further supports the results of the urinary analysis that New Zealand children were mildly iodine deficient. In healthy individuals free of thyroid disease, Tg increases with an increase in thyroid volume, thus Tg is an indirect index of goiter. The elevated Tg found in this survey substantiates our view that a large proportion of New Zealand children would have had goiter when the survey was conducted. We found a significant inverse relationship between age and Tg concentration. Djemli et al. [23] also found that Tg concentration decreased with age in children, in contrast to Zimmermann et al. who reported that the difference in Tg with respect to age was small [16]. Of particular interest in this study was the finding that New Zealand children with an UIC $\geq 100 \ \mu g/L$ had a Tg concentration of 10.3 μ g/L, providing evidence that the 10 μ g/L cut-off proposed by WHO/UNICEF/ICCIDD for classifying mild iodine deficiency is appropriate for population studies of schoolchildren.

Because of the large intra- and inter-individual variation in UIC [24], a limitation of UIC is that it can only be used to assess the iodine status of a population, but not of the

Table 5 Effect of sex, age, socioeconomic status and ethnicity on Free T4 and Free T3 concentration

	Free T4 (pmol	l/L)	Free T3 (pmol/L)			
	β (95% Cl) ^a	Р	β (95% Cl)	Р		
Sex	0.16 (-0.29,0.62)	0.482	0.03 (-0.07,0.14)	0.511		
SES	-0.31 (-0.64, 0.03)	0.072	-0.02 (-0.08,0.09)	0.870		
Age Category	-0.49 (-0.15, -0.84)	0.004	-02.02 (-0.09,0.05)	0.570		
Ethnicity	-0.43 (-0.17,-0.70)	0.002	-0.01 (-0.07,0.07)	0.860		

 ${}^{a}\beta$ coefficient (95% Confidence Interval) by regression analysis.

Socioeconomic = SES.

individuals in that population. There is considerable interest, worldwide, in the development of an index of iodine status that can be used to assess iodine deficiency and its severity in individuals. Indeed, compared to other micronutrients such as iron and many vitamins, there is currently no biochemical measure to diagnose mild iodine deficiency in an individual. Soldin et al. used data from NHANES III and found that there was no relationship between UIC and TSH or T4 concentration [25]. Serum Tg concentration holds promise as an index of individual iodine status, but further studies are needed to determine its' specificity and sensitivity with regard to iodine deficiency. Furthermore, Tg concentration can be affected by a number of factors including interassay variability and the presence of Tg antibodies (Tg-Ab), which can elevate the concentration of Tg. Zimmermann et al. [16] developed a dried blood spot method for Tg determination, however, to our knowledge, this method is not widely used nor been reproduced in other countries. Regardless of whether Tg can be used to identify iodine deficiency in an individual, our study shows that the median Tg concentration of a group of children can be used as an index of iodine status for a population, and if the dried blood spot method was used, could eliminate the need to collect urine samples under difficult field conditions.

The (geometric) mean TSH concentration of New Zealand schoolchildren of 1.7 mU/L was similar to values published in two other studies of children [23,26] but lower than that reported in a 6-14 year old Austrian hospitalbased pediatric population [27]. In contrast, higher median TSH concentrations were observed in American children 12-19 yr from NHANES III (i.e. girls: 1.3 mU/L and boys: 1.5 mU/L) [28]. Most studies have found that TSH decreased with age, however, we did not observe an age effect in New Zealand schoolchildren. Furthermore, we did not find TSH concentration differed by sex, although both age and sex specific reference ranges for TSH do exist.

The mean fT4 concentration of children in this survey was 14.9 pmol/L and there was a significant effect of both age and ethnicity on fT4. A decrease in fT4 with increasing age has been observed in most other studies of children [23,26,29]. As with TSH, there are small

differences in the actual mean fT4 concentration for children obtained from different studies, including our New Zealand data, however, this likely reflects methodological differences with the types of assays used to determine fT4. To our knowledge, this is the first study to find an effect of ethnicity on fT4 concentration, such that Caucasian children (i.e. NZEO) had a lower fT4 than Māori or Pacific children. The mean fT3 concentration of children in this survey was similar to other studies [27,29]. We found no effect of age or sex on fT3 concentration, in agreement with the findings of Soldin et al. [29], however, Kapelari et al. [27] did report a small decline in fT3 concentration with increasing age. In this survey, New Zealand schoolchildren with a low UIC (i.e. either $<50 \ \mu g/L$ or $<100 \ \mu g/L$) had significantly higher fT3 concentration than children with a UIC above these cut-offs, however, this difference was only 0.1 pmol/L and is unlikely to have biological significance. The mean fT3, fT4, and TSH concentrations for these children fell within normal reference ranges, which was not unexpected as changes in these indices only occur in moderate to severe iodine ficiency. In studies whose aim is to assess iodine status in schoolchildren, particularly in developed countries where iodine deficiency is unlikely to be severe, only measures of UIC and Tg are required.

This survey is one of the largest recent studies of the iodine status of schoolchildren published to date, but does have limitations. The serum samples had undergone up to four freeze-thaw cycles prior to analysis, although we found that freeze-thaw cycles had no effect on either TSH or Tg concentration. We used plasma samples for the determination of fT3 and fT4, which had a recovery >97% for both fT3 and fT4 compared to serum samples from the same individuals. We did not measure Tg-Ab in our study due to limited financial resources. Zimmermann et al. suggests that screening for Tg-Ab may not be necessary as a number of studies report a low prevalence (i.e. <5%) of Tg-Ab in children [16].

Conclusions

This survey has comprehensively examined the iodine status of a representative sample of schoolchildren using a wide range of biochemical indices. Both the median UIC and Tg fell within the range indicative of mild iodine deficiency as recommended by WHO/UNICEF/ICCIDD; this study shows that both are good population indices of mild iodine deficiency. Little is known about the consequences of mild iodine deficiency in childhood as most research has focused on the effects of moderate and severe iodine deficiency. Given the role of thyroid hormones in the brain, a lack of iodine in childhood could affect the developing brain. A cross-sectional study of Spanish children found that children with an UIC <100 μ g/L were at greater risk of having an IQ below 70

[30]. An intervention study conducted in New Zealand of mildly iodine deficient 10-13 yr old children found that an improvement in iodine status as a result of daily iodine supplementation for seven months also improved aspects of cognition [31]. In response to the widespread mild iodine deficiency observed in the current survey, the New Zealand government introduced the mandatory fortification of bread with iodized salt in late 2009; no other commercial food products are fortified with iodine. An improvement in iodine status of New Zealand children, confirmed by either an increase in median UIC or a decrease in median Tg concentration, has yet to be confirmed.

Abbreviations

CI: Confidence Interval; CV: coefficient of variation; fT3: free tri-iodothyronine; fT4: free thyroxine; ICCIDD: International Committee for the Control of Iodine Deficiency Disorders; NZEO: New Zealand European and Other Ethnicities; Tg: thyroglobulin; Tg-Ab: thyroglobulin antibodies; TSH: thyroxine; UIC: urinary iodine concentration; UNICEF: United Nations Children's Fund; WHO: World Health Organization; yr: year.

Competing interests

The authors declare they have no competing interests.

Acknowledgements

The study was funded by the New Zealand Ministry of Health. We would like to thank the children and their parents for taking part in this survey. We would like to thank the other principal investigators of the Children's Nutrition Survey: Robert Scragg (University of Auckland), David Schaff (University of Auckland), and Eljon Fitzgerald (Massey University).

Author details

¹Department of Human Nutrition, University of Otago, Dunedin 9054, New Zealand. ²LINZ Activity Unit, University of Otago, Dunedin 9054, New Zealand.

Authors' contributions

WRP and NW were involved in the planning, implementation and data collection for the Children's Nutrition Survey. SAS and CDT were involved in the planning of the iodine component of the survey, and supervised the analysis of the blood and urine samples. SAS undertook the statistical analysis. The manuscript was drafted by SAS with contributions from CDT, NW, and WRP. All authors commented on the final draft. All authors read and approved the final manuscript.

Received: 10 February 2012 Accepted: 8 May 2012 Published: 8 May 2012

References

- Purves HD: The aetiology and prophylaxis of endemic goitre and cretinism. New Zealand Medical Journal 1974, 80:477–479.
- Mann J, Aitken E: The re-emergence of iodine deficiency in New Zealand. New Zealand Medical Journal 2003, 116(1170).
- 3. North KAK, Fraser S: lodine Intake as revealed by urinary iodine excretion. New Zealand Medical Journal.
- Simpson FO, Thaler BI, Paulin JM, Phelan EL, Cooper GJS: lodine excretion in a salt-restriction trial. New Zealand Medical Journal 1984, 97:890–893.
- Cooper GJS, Croxson MS, Ibbertson HK: Iodine intake in an urban environment: a study of urine iodide excretion in Auckland. New Zealand Medical Journal 1984, 97:142–145.
- Ford HC, Johnson LA, Feek CM, Newton JD: Iodine intake and the seasonal incidence of thyrotoxicosis in New Zealand. *Clinical Endocrinology* 1991, 34:179–181.
- World Health Organization: WHO/UNICEF/ICCIDD Assessment of iodine deficiency disorders and monitoring their elimination. 3rd edition. Geneva: a guide for programme managers; 2007.

- Li M, Eastman CJ, Waite KV, Ma G, Zacharin MR, Topliss DJ, Harding PE, Walsh JP, Ward LC, Mortimer RH, et al: Are Australian children iodine deficient? Results of the Australian National Iodine Nutrition Study. Medical Journal of Australia 2006, 184:165–169.
- Vanderpump MPJ, Lazarus JH, Smyth PP, Laurberg P, Holder RL, Boelaert K, Franklyn JA: lodine status of UK schoolgirls: a cross-sectional survey. Lancet 2011, 377:2007–2012.
- Thomson CD, Smith TE, Butler KA, Packer MA: An evaluation of urinary measures of iodine and selenium status. *Journal of Trace Elements in Medicine and Biology* 1996, 10:214–222.
- Thomson CD, Colls AJ, Conaglen J, MacCormack M, Stiles M, Mann J: Iodine status of New Zealand residents as assessed by urinary iodide excretion and thyroid hormones. *British Journal of Clinical Nutrition* 1997, 78:901–912.
- Thomson CD, Packer MA, Butler JA, Duffield AJ, O'Donaghue KL, Whanger PD: Urinary selenium and iodine during pregnancy and lactation. *Journal* of Trace Elements in Medicine & Biology 2001, 14:210–217.
- Skeaff SA, Thomson CD, Gibson RS: Mild iodine deficiency in a sample of New Zealand schoolchildren. European Journal of Clinical Nutrition 2002, 56:1169–1175.
- Vejbjerg P, Knudsen N, Perrild H, Laurberg P, Carlé A, Pedersen I, Rasmussen LB, Ovesen L, Jørgensen T: Thyroglobulin as a marker of iodine nutrition status in the general population. *European Journal of Endocrinology* 2009, 161:475–481.
- Rasmussen LB, Ovesen L, Bulow I, Jorgenson T, Knudsen N, Laurberg P, Perrild H: Relations between various measures of iodine intake and thyroid volume, thyroid nodularity, and serum thyroglobulin. *American Journal of Clinical Nutrition* 2002, 76:1069–1076.
- Zimmermann MB, Moretti D, Chaouki N, Torresani T: Development of a dried whole-blood spot thyroglobulin assay and its evaluation as an indicator of thyroid status in goitrous children receiving iodised salt. *American Journal of Clinical Nutrition* 2003, 77:1453–1458.
- Parnell W, Scragg R, Wilson N, Schaaf D, Fitzgerald E: NZ Food NZ Children: Key Results of the 2002 National Children's Nutrition Survey. Wellington: Ministry of Health; 2003.
- Pino S, Fang SL, Braverman L: Ammonium persulfate: A safe alternative oxidising reagent for measuring urinary iodine. *Clinical Chemistry* 1996, 42:239–243.
- Salmond C, Crampton R: NZDEP2001 Index of Deprivation. Wellington School of Medicine: University of Otago; 2002.
- Delange F, Benker G, Caron P, Eber O, Ott W, Peter F, Podoba J, Simescu M, Szybinsky Z, Vertongen F, et al: Thyroid volume and urinary iodine in European schoolchildren: standardization of values for assessment of iodine deficiency. European Journal of Endocrinology 1997, 136:180–187.
- Zimmermann MB, Hess SY, Molinari L, de Benoist B, Delange F, Braverman LE, Fujieda K, Ito Y, Jooste PL, Moosa K, *et al*: New reference values for thyroid volume by ultrasound in iodine-sufficient schoolchildren: a World Health Organization/Nutrition for Health and Development Iodine Deficiency Study Group Report. *American Journal of Clinical Nutrition* 2004, 79:231–237.
- 22. Skeaff SA: The iodine status of vulnerable groups in New Zealand. University of Otago, Department of Human Nutrition: PhD thesis; 2004.
- Djemli A, Van Vliet G, Belgoudi J, Lambert M, Delvin EE: Reference intervals for free thyroxine, total triiodothyronine, thyrotropin and thyroglobulin for Quebec newborns, children and teenagers. *Clinical Biochemistry* 2004, 37:328–330.
- Konig F, Andersson M, Hotz K, Aeberli I, Zimmerman MB: Ten repeat collections for urinary iodine from spot samples or 24-hour samples are needed to estimate individual iodine status in women. *Journal of Nutrition* 2011, 141:2049–2054.
- Soldin OP, Tractenberg RE, Pezzullo JC: Do thyroxine and thyroid;stimulating hormone levels reflect urinary iodine concentrations? *Therapeutic Drug Monitoring* 2005, 27:178–185.
- Zurakowski D, Di Canzio J, Majzoub JA: Pediatric reference values for serum thyroxine, tri-iodothyronine, thyrotropin, and free thyroxine. *Clinical Chemistry* 1999, 45:1087–1091.
- Kapelari K, Kirchlechner C, Hogler W, Schweitzer K, Virgolini I, Moncayo R: Pediatric reference intervals for thyroid hormone levels from birth to adulthood: a retrospective study. BMC Endocrine Disorders 2008, 8:15.
- Hollowell JG, Staehling NW, Flanders DW, Hannon H, Gunter EW, Spencer CA, Braverman LE: Serum TSH, T4, and thyroid antibodies in the United States population (1988 to 1994): National Health and Nutrition

- Soldin SJ, Cheng LL, Lam LY, Werner A, Le AD, Soldin OP: Comparison of FT4 with log TSH on the Abbott Architect ci8200: pediatric reference intervals for free thyroxine and thyroid-stimulating hormone. *Clinica Chimica Acta* 2010, 411:250–252.
- Santiago-Fernandez P, Torres-Barahona R, Muela-Martinez JA, Rojo-Martinez G, Garcia-Fuentes E, Garriga MJ, Garcia Leon A, Soriguer F: Intelligence quotient and iodine intake: a cross-sectional study in children. *Journal of Clinical Endocrinology & Metabolism* 2004, 89:3851–3857.
- Gordon RC, Rose MC, Skeaff S, Gray A, Morgan K, Ruffman T: Iodine supplementation improves cognition in mildly iodine deficient children. *American Journal of Clinical Nutrition* 2009, 90:1264–1271.

doi:10.1186/1475-2891-11-31

Cite this article as: Skeaff *et al.*: A comprehensive assessment of urinary iodine concentration and thyroid hormones in New Zealand schoolchildren: a cross-sectional study. *Nutrition Journal* 2012 11:31.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

) BioMed Central

Submit your manuscript at www.biomedcentral.com/submit