RESEARCH



The impact of serum ferritin on overall survival following resection in patients with intrahepatic cholangiocarcinoma

Laura Schwenk^{1,2} • Carlos Wolf¹ • Felix Dondorf^{1,2} • Oliver Rohland^{1,2,3} • Aladdin Ali-Deeb^{1,2} • Utz Settmacher^{1,2} • Falk Rauchfuß^{1,2}

Received: 12 January 2025 / Accepted: 9 May 2025 © The Author(s) 2025

Abstract

Purpose The global incidence of intrahepatic cholangiocarcinoma is increasing. Surgical resection remains the gold standard treatment. However, the long-term prognosis remains dismal. The role of serum ferritin in malignant diseases has not been fully elucidated. This study aimed to evaluate the relationship between preoperative serum ferritin levels and patient outcomes.

Methods In our retrospective study, we analyzed data from 95 patients who underwent liver resection for intrahepatic cholangiocarcinoma at Jena University Hospital between 2009 and 2023. Comprehensive clinical and pathological data, along with the correlation between Serum ferritin and clinicopathological parameters, were systematically analyzed and compared. Survival rates were determined using the Kaplan-Meier method.

Results The optimal preoperative serum ferritin cut-off value for overall survival was 303.1 μ g/L, with an area under the curve of 0.697 (95% CI (0.592–0.801; P<0.001). The 1-, 3-, and 5-year survival rates were 74.7%, 50.5%, and 43.2%, respectively. Patients with elevated preoperative SF levels demonstrated significantly worse overall survival compared to the low SF group (50.9% vs. 4.5%; P<0.001). SF had a significant impact on recurrence rates (P<0.001). The overall recurrence rate in the high-SF group was 67,3%, compared to 43,5% in the low-SF group.

Conclusion Elevated preoperative serum ferritin levels are associated with significantly worse overall and recurrence-free survival in patients with intrahepatic cholangiocarcinoma. Serum ferritin could serve as a valuable adjunct to the tumor marker CA 19-9.

Keywords Intrahepatic cholangocarcinoma · Serum ferritin · Liver resection · Overall survival

Laura Schwenk and Carlos Wolf contributed equally to this work.

□ Laura Schwenk

laura.schwenk@med.uni-jena.de

Carlos Wolf

Carlos.Wolf@uni-jena.de

Felix Dondorf

Felix.Dondorf@med.uni-jena.de

Oliver Rohland

Oliver.Rohland@med.uni-jena.de

Aladdin Ali-Deeb

Published online: 21 May 2025

Aladdin.Ali-Deeb@med.uni-jena.de

Utz Settmacher Utz.Settmacher@med.uni-jena.de

Falk Rauchfuß

Falk.Rauchfuss@med.uni-jena.de

- Department of General, Visceral and Vascular Surgery, Jena University Hospital, Am Klinikum 1, 07740 Jena, Germany
- ² Comprehensive Cancer Center Central Germany– Campus Jena, Jena, Germany
- Interdisciplinary Center for Clinical Research (IZKF), Jena University Hospital, Jena, Germany



166 Page 2 of 15 Langenbeck's Archives of Surgery (2025) 410:166

Introduction

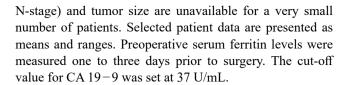
The incidence of intrahepatic cholangiocarcinoma (iCCA) has been increasing globally in recent years [1, 2]. ICCA is the second most common primary liver cancer after hepatocellular carcinoma, accounting for approximately 10–15% of all primary liver malignancies [2]. Diagnosis often occurs at an advanced stage, leaving palliative chemotherapy as the only treatment option, with dismal survival rates below 10% [3]. Surgical resection remains the gold standard for treatment, with reported overall survival rates of up to 34% [4, 5]. Identifying new prognostic factors that influence survival in patients with iCCA is therefore of increasing importance. However, laboratory-based surrogate markers remain insufficiently explored and evaluated to date.

The glycoprotein ferritin was first discovered in 1937 by Victor Laufberger, who isolated a novel protein from the spleen of horses [6]. Years later, ferritin was also detected in human serum. Ferritin is primarily known for its role in iron metabolism as an iron storage protein. The exact secretion pathway of SF remains unclear, although hepatocytes, macrophages, and Kupffer cells are known to secrete ferritin. Elevated SF levels are not only observed in cases of iron overload but also in infections, organ dysfunction, liver diseases, and chronic illnesses. Its exact pathophysiological function is still not fully understood. The role of SF in malignancies has been increasingly investigated and debated in recent years. The association of serum ferritin with malignancies and its potential role as a useful tumor marker was first postulated in 1977 by Hazard and Drysdale [7]. Since then, numerous studies have demonstrated a correlation between elevated SF levels and malignancies [8–12]. However, no studies to date have investigated the impact of SF on iCCA. Therefore, the aim of our study was to evaluate the relationship between preoperative serum ferritin levels and patient outcomes.

Materials and methods

The data of patients who underwent liver resection for iCCA at the University Hospital in Jena between 2009 and 2023 were evaluated. The following parameters were analyzed: preoperative SF levels, overall survival (OS), 3-year and 5-year survival rates, recurrence rates, and tumor-specific information such as tumor staging (TNM 8th edition), size and differentiation.

Tumor markers and general patient data, based on clinical, surgical, and pathological findings, were collected from the hospital's SAP database (SAP Global Corporate Affairs, Walldorf, Germany). Due to incomplete pathological results, exact data regarding the tumor staging (T-stage,



Statistical analysis

The optimal cut-off value for preoperative serum ferritin (SF) was determined using the Receiver Operating Characteristic (ROC) curve. The point on the ROC curve with both the maximum sensitivity and specificity was selected as the best cut-off value to define elevated SF levels.

Survival curves were generated using the Kaplan–Meier method, and group differences in overall survival were assessed using the log-rank test. To identify independent prognostic factors for overall survival, a multivariate Cox proportional hazards regression analysis was performed. Variables included in the model were selected based on clinical relevance and univariate analysis results. The final model comprised preoperative serum ferritin, T-stage, G-stage, tumor size, N-stage, albumin, and CA19-9. Preoperative SF levels were dichotomized using the ROC-derived cut-off of 303.1 μg/L, with levels>303.1 μg/L classified as elevated. Hazard ratios (HRs) with 95% confidence intervals (CIs) and corresponding *p*-values were calculated. The proportional hazards assumption was verified using log-minuslog plots and time-dependent covariates, where applicable.

All statistical analyses were performed using SPSS Statistics (IBM Corp., Armonk, NY, USA, Version 29.0.1.0 (171)). The initial collection of patient data was carried out using Microsoft Excel (Microsoft Corporation, Redmond, WA, USA, Version 16.16.27 (201012)). A *p*-value < 0.05 was considered statistically significant.

Additionally, a literature review was conducted, and the findings were discussed in the context of our results. The following search terms were used: "serum ferritin," "resection," and "intrahepatic cholangiocarcinoma." The electronic databases included PubMed, Google Scholar, and MEDLINE.

Results

A total of 95 patients were included in the analysis. The mean age at the time of diagnosis was 64 years (range: 37–81), and the mean age at the time of resection was 65 years (range: 38–81). Fifty patients (52.6%) were male, and 45 patients (47.4%) were female. Patient characteristics are summarized in Table 1.



Langenbeck's Archives of Surgery (2025) 410:166 Page 3 of 15 166

Table 1 Characteristics of patients			
Variables	Entire Cohort		
	(n=95)		
Age at the time of first diagnosis (years, mean,	64 (37–81)		
range)	65 (38–81)		
Age at the time of resection (years, mean, range)	03 (36–61)		
Gender (Male, Female, n (%))	50 (52.6), 45		
(,, (/)	(47.4)		
BMI (kg/m ² , mean, range)	26.3 (18–39)		
Liver disease (yes, n (%))			
Steatosis	39 (41.1)		
Fibrosis	8 (8.4)		
Cirrhosis	4 (4.2)		
Operation time (minutes, mean, range)	229.4 (85-437)		
G Stage (n (%))			
G1 and G2	52 (54.7)		
G3	43 (45.3)		
N Stage (n (%))			
N1/Nx	43 (47.8)		
N0	47 (52.2)		
T Stage (n (%))			
T1	39 (41.5)		
T2	39 (41.5)		
T3	11 (11.7)		
T4	5 (5.3)		
Tumor size (cm, mean, range)	7.1 (1.7–19.5)		
Tumor size (n (%))			
≥5 cm	70 (74.5)		
<5 cm	24 (25.5)		
Tumor localization (n (%))			
bilobar	11 (11.6)		
right lobe	39 (41.1)		
left lobe	37 (38.9)		
central	8 (8.4)		
Tumor lesions (n (%))			
1	55 (57.9)		
2	11 (11.6)		
3	8 (8.4)		
multiple	21 (22.1)		
CA 19–9 highest (U/ml)	5825.69		
G. 10.0 (/ /0/)	(8.6–278739)		
CA19-9 (n (%))	(4 (67.4)		
elevated	64 (67.4)		
normal	31 (32.6)		
Albumin (g/L, mean, range)	36.22 (19–63)		
Albumin	75 (70.9)		
>30 g/L	75 (79.8)		
≤30 g/L	19 (20.2)		
Ferritin (μg/L, mean, range)	764.70 (24-12107)		

Prognostic cut-off value for SF

The optimal preoperative SF cut-off value for overall survival was determined using ROC analysis.

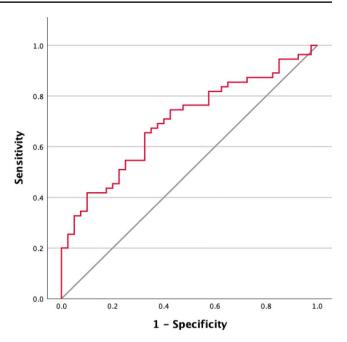


Fig. 1 Receiver operating characteristics curves to evaluate the optimal cut-off value of serum ferritin for overall survival

The best cut-off value for SF was identified as 303.1 µg/L, with an area under the curve (AUC) of 0.697, a 95% confidence interval (CI) ranging from 0.592 to 0.801 (P<0.001), a sensitivity of 65.5%, and a specificity of 67.5% (Fig. 1). Based on this threshold, patients were stratified into either a low-SF group or a high-SF group for further analysis. A comparison of patient characteristics between the high and low serum ferritin groups is presented in Table 2.

Survival analysis

Total cohort

The mean follow-up time from the date of resection was 32.6 months (range: 0.03-151 months). A total of 55 patients had died during the follow-up period, resulting in an OS rate of 42.1%. The 1-, 3-, and 5-year survival rates were 74.7%, 50.5%, and 43.2%, respectively.

A positive lymph node status (P=0.010), poor tumor differentiation (P=0.040), advanced T stage (P=0.001), elevated preoperative serum ferritin levels (P < 0.001), decreased albumin levels (≤ 30 g/L) (P=0.013), and increased tumor size (P=0.016) were all significantly correlated with poorer overall survival (Table 3).

High and low SF

Patients with elevated preoperative SF levels demonstrated significantly worse overall survival compared to the low SF group (50.9% vs. 4.5%; *P*<0.001) (Table 3; Fig. 2).



 Table 2 Comparison of patient characteristics between the high and low serum ferritin groups

	High SF group (>303.1 μg/L)	Low SF group (≤303.1 µg/L)		
Total number $(n(\%))$	49 (51.6)	46 (48.4)		
Age at the time of first diagnosis (years, mean, range)	64 (38–81)	66 (37–80)		
Age at the time of resection (years, mean, range)	64 (38–81)	66 (38–80)		
Gender (Male, Female, n)	Male: 27	Male: 23		
	Female: 22	Female: 23		
BMI (kg/m ² , mean, range)	25.9 (18–34)	26.7 (18–39)		
Liver disease (yes)	,	,		
Steatosis	20	19		
Fibrosis	5	3		
Cirrhosis	1	3		
Operation time (minutes, mean, range)	245 (85–437)	213 (88–371)		
Small for sized	243 (63–437)	213 (88–371)		
Yes	7	2		
		2		
No	42	44		
G Stage (n)		25		
G1 and G2	25	27		
G3	24	19		
N Stage (n)				
N1/Nx	29	14		
N0	18	29		
T Stage (n)				
T1	16	23		
T2	21	18		
T3	9	2		
T4	3	2		
Tumor size (cm, mean, range)	7.7 (1.7–19.5)	6.48 (2.5–13)		
Tumor size (n)	,			
≥5 cm	41	29		
<5 cm	8	16		
Tumor localization (n)				
bilobar	7	4		
right lobe	18	21		
left lobe	18	19		
central	6	2		
Tumor lesions (n)	O	2		
	23	32		
1				
2	8	3		
3	4	4		
multiple	14	7		
CA 19-9 highest (U/ml)	8623.6 (8.9-278739)	2845.315 (8.6-99731)		
CA19-9 (n)				
elevated	34	30		
normal	15	16		
Albumin (g/L, mean, range)	34.1 (19–47)	38.37 (26–63)		
Albumin				
>30 g/L	31	44		
≤30 g/L	17	2		
Ferritin (µg/L, mean, range)	1333.69 (207-12107)	158.60 (24–299)		
Relapse				
Yes	33	20		
No	16	26		
Overall Survival				
alive	13	27		
deceased	36	19		



 Table 3 Results of the Kaplan-Meier analysis

Variables	Univariate Analysis		Multivariate Analysis			
	N (%)	OS	P-value	HR	95%CI	P-value
N-Stage (Fig 2a)			0.010	1.770	0.939-3.336	0.077
N0	47 (52.2)	39.7%				
N1/Nx	43 (47.8)	21.4%				
N-Stage and SF (Fig 2b)			< 0.001			
Low SF and N0	29 (32.2)	56.3%				
Low SF and N1/Nx	14 (15.6)	44.9%				
High SF and N0	18 (20)	9.5%				
High SF and N1/Nx	29 (32.2)	7.2%				
G-Stage (Fig 3a)	- (-)		0.040	1.398	0.744-2.625	0.298
G1/G2	52 (54.7)	38.8%				
G3	43 (45.3)	16.8%				
G-Stage and SF (Fig 3b)	15 (15.5)	10.070	< 0.001			
High SF and G1/G2	25 (26.3)	8.2%	10.001			
High SF and G3	24 (25.3)	0%				
Low SF and G1/G2	27 (28.4)	60.6%				
Low SF and G3	19 (20)					
	19 (20)	32.4%	0.001			0.026
T-Stage (Fig 4a)	20 (41.5)	46.00/	0.001			0.026
T1	39 (41.5)	46.9%				
T2	39 (41.5)	20.6%				
T3	11 (11.7)	14.1%				
T4	5 (5.3)	0%	0.004			
T-Stage and SF (Fig 4b)			< 0.001			
Low SF and T1	23 (24.5)	62.1%				
High SF and T1	16 (17)	0%				
Low SF and T2	18 (19.1)	40.4%				
High SF and T2	21 (22.3)	0%				
Low SF and T3	2 (2.1)	50%				
High SF and T3	9 (9.6)	13.3%				
Low SF and T4	2 (2.1)	50%				
High SF and T4	3 (3.2)	0%				
Preoperative SF (Fig 5)			< 0.001	2.849	1.408 - 5.762	0.004
Low SF ($\leq 303.1 \mu\text{g/L}$)	46 (48.4)	50.9%				
High SF (>303.1 μg/L)	49 (51.6)	4.5%				
Preoperative Albumin (Fig 6a)			0.013	1.094	0.538 - 2.227	0.804
Albumin (>30 g/L)	75 (79.8)	33.5%				
Albumin (≤30 g/L)	19 (20.2)	9.9%				
Preoperative Albumin and SF (Fig 6b)			< 0.001			
High SF and Albumin (>30 g/L)	31 (33)	0%				
High SF and Albumin (≤30 g/L)	17 (18.1)	6.4%				
Low SF and Albumin (≤30 g/L)	2 (2.1)	100%				
Low SF and Albumin (>30 g/L)	44 (46.8)	49.1%				
Tumor size (Fig 7a)	()		0.016	0.679	0.288 - 1.600	0.376
≥5 cm	70 (74.5)	20.7%				0.0.0
<5 cm	24 (25.5)	59%				
Tumor size and SF (Fig 7b)	24 (23.3)	3770	< 0.001			
Low SF and tumor < 5 cm	16 (17)	76%	10.001			
Low SF and tumor ≥ 5 cm	29 (30.9)	40.4%				
High SF and tumor < 5 cm	8 (8.5)	43.8%				
High SF and tumor<5 cm	8 (8.3) 41 (43.6)	43.8%				
_	41 (43.0)	4.070	0.072	0.000	0.460 1.756	0.774
CA 19-9	64 (67 4)	21 20/	0.072	0.908	0.469– 1.756	0.774
High CA 19 – 9	64 (67.4)	21.3%				
Normal CA 19 – 9	31 (32.6)	45%	-0.001			
CA 19–9 and SF (Fig 8)	16/160	6701	< 0.001			
Low SF and normal CA 19-9	16 (16.8)	67%				



Table 3 (continued)

Variables	Univariate Analysis			Multivariate Analysis		
	N (%)	OS	P-value	HR	95%CI	P-value
Low SF and high CA 19-9	30 (31.6)	40.6%				
High SF and normal CA 19-9	15 (15.8)	0%				
High SF and high CA 19-9	34 (35.8)	4.5%				

Patients with elevated serum ferritin levels and positive lymph node status demonstrated significantly poorer overall survival compared to those with low serum ferritin levels (P=<0.001) (Fig. 2b). The negative impact of elevated SF was also evident in relation to the following variables: G-stage (P=<0.001) (Fig. 3b), T-stage (P=<0.001) (Fig. 4b), preoperative albumin levels (P=0.013) (Fig. 6b), tumor size (P=<0.001) (Fig. 7b), and CA 19-9 levels (P=<0.001) (Fig. 8). The results of the Kaplan-Meier analysis are presented in Table 3.

Multivariate analysis

Of the total 95 patients, 88 (92.6%) were included in the multivariable Cox regression model (Table 3). Seven patients were excluded due to missing values in one or more covariates.

In the multivariate Cox regression analysis including ferritin, T-stage, G-stage, tumor size, N-stage, albumin, and CA19-9, elevated preoperative serum ferritin remained a significant independent prognostic factor for overall survival (HR=2.85; 95% CI: 1.41-5.76; p=0.004). T-stage also demonstrated a statistically significant association with survival (p=0.026), particularly T-stage 1 (HR=2.16; p=0.025). Although T-stage 3 showed a higher hazard ratio (HR=2.76), it did not reach statistical significance (p=0.120). N-stage showed a trend toward significance (HR=1.77; p=0.077), while G-stage, tumor size, albumin, and CA19-9 were not significantly associated with overall survival (p>0.05 for all).

Relapse rate

By the final follow-up date, 53 patients (55.8%) experienced confirmed recurrence of iCCA. The 1-, 3-, and 5-year recurrence-free survival rates were 55.8%, 46.3%, and 44.2%, respectively. Intrahepatic recurrences were the most frequently observed.

Among patients with elevated serum ferritin levels (>303.1 μ g/L), 33 of 49 (67.3%) developed recurrence. In the low-SF group (\leq 303.1 μ g/L), recurrence was observed in 20 of 46 patients (43.5%). The difference in recurrence rates between the two groups was statistically significant (P<0.001) (Table 4).



In our study, we were able to demonstrate a significant association between elevated preoperative SF levels and overall survival (OS) as well as recurrence rates in patients with iCCA. The OS rate was 42.1%, with 1-, 3-, and 5-year survival rates of 74.7%, 50.5%, and 43.2%, respectively. Elevated preoperative SF levels were significantly associated with worse overall survival (low SF group: 50.9% vs. high SF group: 4.5%; P < 0.001) and higher recurrence rates (low SF group: 43,5% vs. high SF group: 67,3%; *P*<0.001) (Fig. 5). In 1977, Hazard and Drysdale first reported a potential association between elevated SF levels and malignant diseases [7]. Since then, SF has increasingly gained attention as a potential biomarker, with numerous studies demonstrating a correlation between elevated SF levels and OS and recurrence rates in various malignancies, including neuroblastoma [13], glioblastoma [14], renal cell carcinoma [15], melanoma [16], non-small cell lung cancer [17], Hodgkin lymphoma [18], pancreatic cancer [9, 19, 20], breast cancer [21], colorectal carcinoma [12], and hepatocellular carcinoma [22–24]. Kalousová et al. demonstrated in their study that elevated SF levels are associated with poorer prognosis in patients with pancreatic cancer. Their analysis of 57 patients revealed that SF is a significant independent predictor of mortality in both univariate (P < 0.001) and multivariate (P=0.002) analyses [9]. A study published in 2010 by Kukulj et al. [17] investigated the relationship between iron and inflammatory markers and OS in 125 male patients with non-small cell lung cancer. Their findings revealed that more than half of the patients presented with significantly elevated SF levels at the time of diagnosis, while serum iron levels were below the reference range. The authors concluded that increased ferritin expression in tumor tissue and elevated SF levels are more likely indicative of a response to acute inflammation, oxidative stress, and localized toxicity within tumor tissue rather than systemic iron overload. The impact of SF has also been examined in primary liver tumors. Facciorusso et al. demonstrated that elevated SF levels are a negative prognostic factor for both OS and recurrence rates in patients with HCC undergoing percutaneous radiofrequency ablation [22]. Patients with elevated SF levels (>244 ng/ml) exhibited significantly poorer survival (31 months) compared to patients with lower SF levels (\leq 244 ng/ml; 78 months; P<0.001). Furthermore, elevated



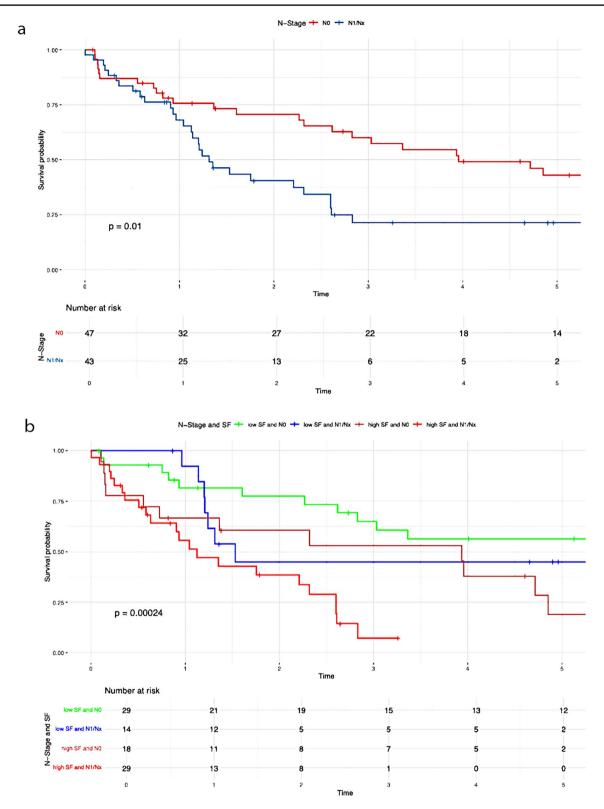
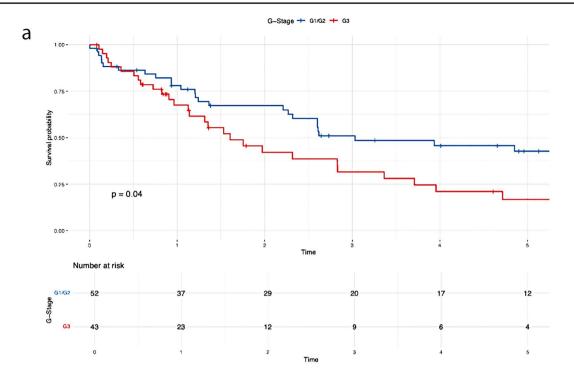


Fig. 2 Overall survival N-Stage. (a) N-Stage (b) N-Stage and SF

166 Page 8 of 15 Langenbeck's Archives of Surgery (2025) 410:166



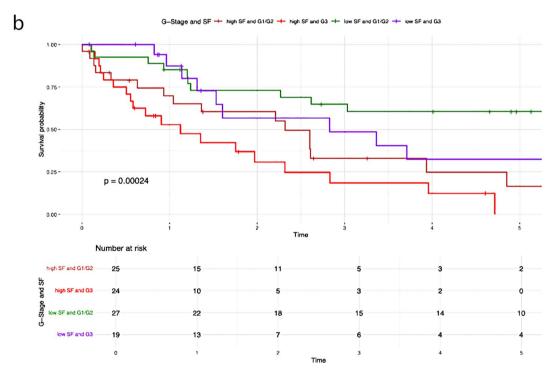


Fig. 3 Overall survival G-Stage. (a) G-Stage (b) G-Stage and SF

SF was significantly associated with reduced recurrence-free survival (55 months vs. 15 months; P<0.001). Similar findings were reported by Wu et al. [25]. Their 2019 study included 427 HCC patients who underwent curative liver

resection for hepatocellular carcinoma. The optimal SF cutoff value for OS was identified as 267 ng/mL. The analysis demonstrated that preoperative SF levels independently predicted both OS and recurrence-free survival, regardless



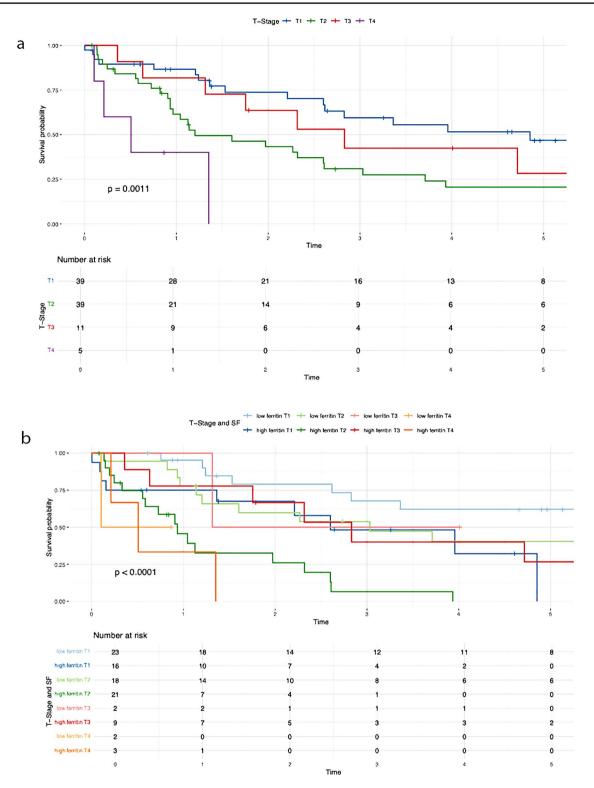


Fig. 4 Overall survival T-Stage. (a) T-Stage (b) T-Stage and SF

of other prognostic factors. Patients with lower SF levels exhibited better OS (P=0.001) and RFS (P<0.001). The 5-year OS was 45.2% for patients with low SF compared to 29% for those with high SF. Although the impact of SF

has been extensively studied in various malignancies, there is still limited research examining the association between elevated SF levels and OS or recurrence rates in patients with iCCA. Xun et al. conducted a retrospective analysis



Table 4 Recurrence rates

Group	Total	Recurrence	Rate percent
Low SF	46	20	43.5
High SF	49	33	67.5
Total	95	53	55.8

P-value: < 0.001

of prognostic risk factors in 104 iCCA patients. Their findings revealed that ferritin concentration was an independent risk factor for survival, as shown in both univariate and multivariate analyses. Patients with high preoperative ferritin levels had a 1.7-fold increased risk of mortality compared to those with normal ferritin levels [26]. Similarly, Ma et al. confirmed that ferritin serves as a predictor for both disease-free survival and OS in iCCA patients. They concluded that ferritin could complement CA19-9 in stratifying survival outcomes, particularly for patients with small-duct-type iCCA [27]. These findings are consistent with ours. Although CA19-9 alone did not have a statistically significant impact on OS in our analysis (P=0.072), likely due to the small sample size, a significant correlation (P < 0.001) (Fig. 8) was observed when preoperative serum ferritin levels were included. The OS for patients with elevated SF and CA19-9 levels was 4.5%, compared to 67% for those with low SF and normal CA19-9 levels (Fig. 8). This supports the notion that preoperative SF may serve as a valuable complement to CA19-9 as a tumor marker. While SF has been identified as a prognostic risk factor by several authors, an optimal cut-off value remains undefined. Therefore, we calculated the best cut-off value for SF using ROC analysis. The optimal cut-off value was determined to be 303.1 μ g/L, with a sensitivity of 65.5% and a specificity of 67.5% (Fig. 1). Similarly, Facciorusso et al. [22] and Wu et al. [25] identified optimal SF cut-off values of 244 ng/mL and 267 ng/mL, respectively. Although our cut-off value of 303.1 μ g/L differs slightly, it is important to note that our study focused on patients with iCCA rather than HCC. This could explain the higher cut-off value identified in our study.

In our study, we were also able to demonstrate that an increase in preoperative SF is significantly associated with worse OS (50.9% vs. 4.5%; P<0.001) and a higher recurrence rate (67,3% vs. 43.5%; P<0.001).

Additionally, a correlation was observed between elevated SF concentrations and tumor size as well as tumor differentiation (Figs. 2b and 3b). The significant impact of tumor size on survival in patients with iCCA has been repeatedly demonstrated [28, 29]. Sapisochin even showed that patients with iCCA and a tumor diameter of less than 2 cm had excellent survival rates even after liver transplantation [30]. This correlation is also reflected in our results. The overall survival was 59% for patients with a tumor diameter < 5 cm, compared to 20.7% for those with a tumor

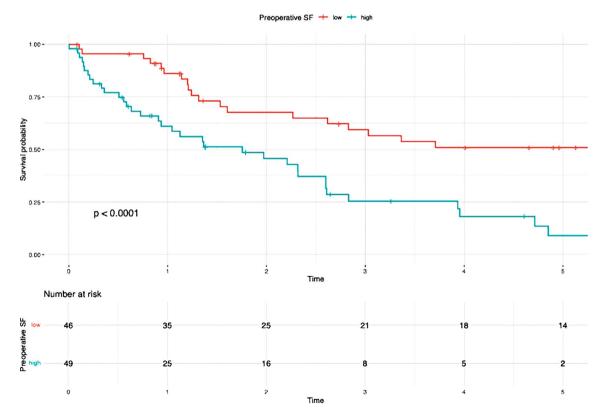


Fig. 5 Overall survival Preoperative SF



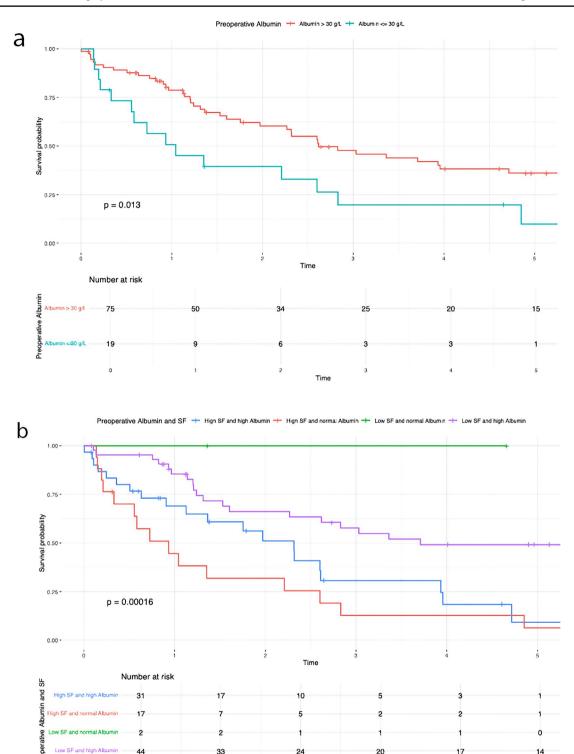


Fig. 6 Overall survival preoperative Albumin. (a) preoperative Albumin (b) preoperative Albumin and SF

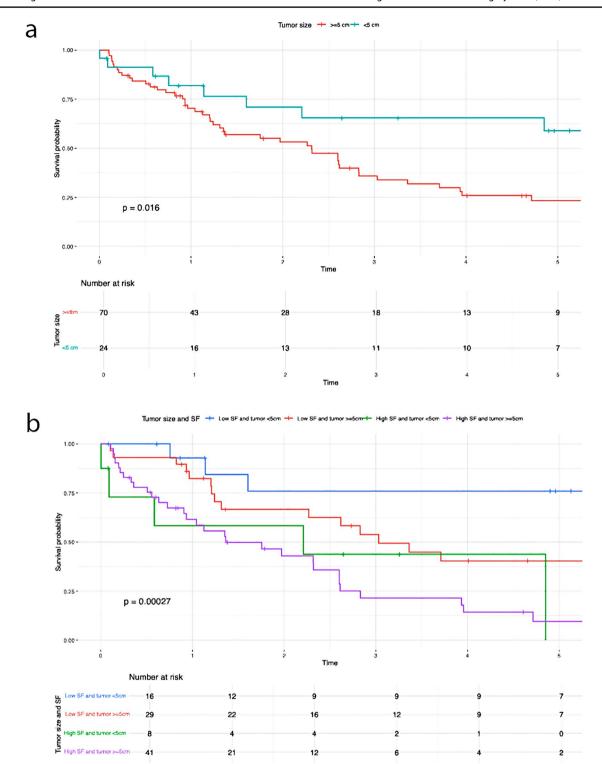
diameter ≥ 5 cm (P=0.016), as shown in Table 2. It is noteworthy that the overall survival of patients assigned to the low SF group was 76% (tumor diameter ≤ 5 cm) and 40.4% (tumor diameter ≥ 5 cm). In contrast, in the case of high

preoperative SF, the overall survival decreased to 43.8% (tumor diameter ≤ 5 cm) and 4.8% (tumor diameter ≥ 5 cm) (P < 0.001). Both for surgical resections and in the context of liver transplantation, a correlation between poor tumor

Time



166 Page 12 of 15 Langenbeck's Archives of Surgery (2025) 410:166



Time

Fig. 7 Overall survival Tumor size. (a) Tumor size (b) Tumor size and SF

differentiation and high recurrence or low overall survival rates in iCCA patients has been demonstrated [31–33]. Our findings support these results. Patients with poor tumor differentiation (G3) showed a significantly worse OS (16.8%)

compared to those with G1/G2 differentiation (38.8%) (P=0.040). Interestingly, a significant decrease in OS was also observed for patients in the high SF group with a G1/G2 tumor (OS: 8.2%) (Fig. 3b). In cases of high preoperative SF



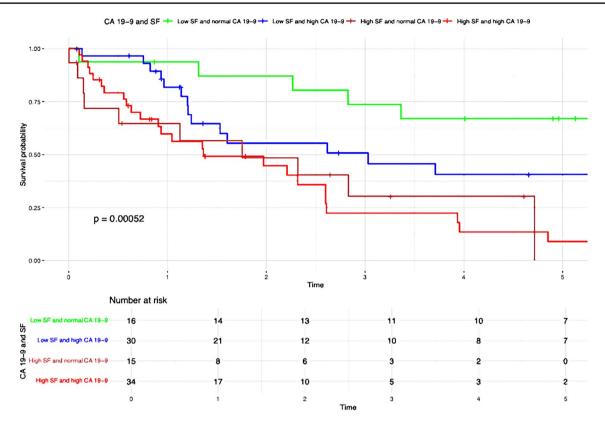


Fig. 8 Overall survival CA19-9 and SF

and a G3 tumor, overall survival dropped to 0%. The exact pathological mechanism of SF remains unknown. However, it is known that elevated SF concentrations do not necessarily correlate with an increase in transaminases, suggesting that the rise in SF is not a response to liver cell damage [24]. Similarly, an increase in SF does not seem to reflect changes in the body's iron stores [17, 34]. Tappin et al. suggested that the increase in SF is partially due to a local release in the tumor area, at least in patients with breast tumors. After surgical resection, the SF level decreased by about 50%, which underscores the connection between tumor mass and high SF [35]. Although the impact of SF on tumorigenesis is still not fully understood, there are indications that extracellular ferritin can directly enhance proliferation in cancer cells, increase angiogenesis, and suppress lymphocyte responses [36–38]. Overall, these effects could contribute to tumor development and thus provide a possible explanation for why ferritin secretion plays a direct role in promoting and maintaining tumor progression. Nonetheless, our study has several limitations. Firstly, as a retrospective study, it is subject to inherent biases that cannot be entirely eliminated. Furthermore, the sample size was limited, and data were obtained from a single center. As such, the results should be interpreted with caution and require validation through larger, multicenter studies to ensure broader applicability. Furthermore, no postoperative assessment of serum ferritin levels was performed. As a result, dynamic changes in serum ferritin could not be evaluated. This limitation may serve as the basis for future studies.

Conclusion

In summary, elevated preoperative SF levels in patients with iCCA are associated with significantly worse OS and higher recurrence rates. The optimal cutoff value for SF was 303.1 μ g/l, with a sensitivity of 65.5% and a specificity of 67.5%. Although our results are statistically significant, further studies are required to investigate the exact pathomechanism of SF.

Author contributions Study conception and design: L.S., C. W., F. R., U. S.; Acquisition of data: L.S., C.W., F.R.; Analysis and interpretation of data: L.S., C.W., U.S., F.R.; Drafting of manuscript: L.S., C.W., F.R.; Critical revision of manuscript: L.S., C.W., O.R., F.D., A.AD., U.S., F. R. All authors reviewed the manuscript.

Funding Open Access funding enabled and organized by Projekt DEAL.

This research received no external funding.

Data availability No datasets were generated or analysed during the current study.



166 Page 14 of 15 Langenbeck's Archives of Surgery (2025) 410:166

Declarations

Competing interests The authors declare no competing interests.

Informed consent Informed consent was obtained from all subjects involved in the study.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit https://creativecommons.org/licenses/by/4.0/.

References

- Leitlinienprogramm Onkologie der Arbeitsgemeinschaft der Wissenschaftlichen Medizinischen Fachgesellschaften e. V. (AWMF) DKeVDudSDKD (2024) Leitlinienprogramm Onkologie (Deutsche Krebsgesellschaft, Deutsche Krebshilfe, AWMF): S3-Leitlinie Diagnostik und Therapie des Hepatozellulären Karzinoms und biliärer Karzinome, Langversion 5.0, 2024, AWMF-Registernummer: 032-053OL
- Florio AA, Ferlay J, Znaor A, Ruggieri D, Alvarez CS, Laversanne M et al (2020) Global trends in intrahepatic and extrahepatic cholangiocarcinoma incidence from 1993 to 2012. Cancer 126(11):2666–2678
- Yu TH, Chen X, Zhang XH, Zhang EC, Sun CX (2021) Clinicopathological characteristics and prognostic factors for intrahepatic cholangiocarcinoma: a population-based study. Sci Rep 11(1):3990
- Ziogas IA, Giannis D, Economopoulos KP, Hayat MH, Montenovo MI, Matsuoka LK et al (2021) Liver transplantation for intrahepatic cholangiocarcinoma: A Meta-analysis and Meta-regression of survival rates. Transplantation 105(10):2263–2271
- Chan KM, Tsai CY, Yeh CN, Yeh TS, Lee WC, Jan YY et al (2018) Characterization of intrahepatic cholangiocarcinoma after curative resection: outcome, prognostic factor, and recurrence. BMC Gastroenterol 18(1):180
- Wang W, Knovich MA, Coffman LG, Torti FM, Torti SV (2010) Serum ferritin: past, present and future. Biochim Biophys Acta 1800(8):760–769
- Hazard JT, Drysdale JW (1977) Ferritinaemia in cancer. Nature 265(5596):755–756
- Shi HB, Li XD, Jiang JT, Zhao WQ, Ji M, Wu CP (2014) Serum ferritin is elevated in advanced non-small cell lung cancer patients and is associated with efficacy of platinum-based chemotherapy. J Cancer Res Ther 10(3):681–685
- Kalousová M, Krechler T, Jáchymová M, Kuběna AA, Zák A, Zima T (2012) Ferritin as an independent mortality predictor in patients with pancreas cancer. Results of a pilot study. Tumour Biol 33(5):1695–1700
- Koyama S, Fujisawa S, Watanabe R, Itabashi M, Ishibashi D, Ishii Y et al (2017) Serum ferritin level is a prognostic marker in patients with peripheral T-cell lymphoma. Int J Lab Hematol 39(1):112–117

- Lee S, Song A, Eo W (2016) Serum ferritin as a prognostic biomarker for survival in relapsed or refractory metastatic colorectal Cancer. J Cancer 7(8):957–964
- Tingting H, Di S, Xiaoping C, Xiaohong W, Dong H (2017) High preoperative serum ferritin predicted poor prognosis in non-metastatic colorectal cancer. Saudi Med J 38(3):268–275
- Hann HW, Levy HM, Evans AE (1980) Serum ferritin as a guide to therapy in neuroblastoma. Cancer Res 40(5):1411–1413
- Sato Y, Honda Y, Asoh T, Oizumi K, Ohshima Y, Honda E (1998) Cerebrospinal fluid ferritin in glioblastoma: evidence for tumor synthesis. J Neurooncol 40(1):47–50
- Partin AW, Criley SR, Steiner MS, Hsieh K, Simons JW, Lumadue J et al (1995) Serum ferritin as a clinical marker for renal cell carcinoma: influence of tumor volume. Urology 45(2):211–217
- Luger TA, Linkesch W, Knobler R, Kokoschka EM (1983) Serial determination of serum ferritin levels in patients with malignant melanoma. Oncology 40(4):263–267
- Kukulj S, Jaganjac M, Boranic M, Krizanac S, Santic Z, Poljak-Blazi M (2010) Altered iron metabolism, inflammation, transferrin receptors, and ferritin expression in non-small-cell lung cancer. Med Oncol 27(2):268–277
- Hann HW, Lange B, Stahlhut MW, McGlynn KA (1990) Prognostic importance of serum transferrin and ferritin in childhood Hodgkin's disease. Cancer 66(2):313–316
- Nitti D, Fabris C, Del Favero G, Farini R, Grassi F, Farini A et al (1982) Serum ferritin in pancreatic disease. Accurate Test Malignancy?? Digestion 25(4):258–262
- Basso D, Fabris C, Del Favero G, Meggiato T, Panozzo MP, Vianello D et al (1991) Hepatic changes and serum ferritin in pancreatic cancer and other Gastrointestinal diseases: the role of cholestasis. Ann Clin Biochem 28(Pt 1):34–38
- Alkhateeb AA, Leitzel K, Ali SM, Campbell-Baird C, Evans M, Fuchs EM et al (2012) Elevation in inflammatory serum biomarkers predicts response to trastuzumab-containing therapy. PLoS ONE 7(12):e51379
- Facciorusso A, Del Prete V, Antonino M, Neve V, Crucinio N, Di Leo A et al (2014) Serum ferritin as a new prognostic factor in hepatocellular carcinoma patients treated with radiofrequency ablation. J Gastroenterol Hepatol 29(11):1905–1910
- Melia WM, Bullock S, Johnson PJ, Williams R (1983) Serum ferritin in hepatocellular carcinoma. A comparison with alphafetoprotein. Cancer 51(11):2112–2115
- Kew MC, Torrance JD, Derman D, Simon M, Macnab GM, Charlton RW et al (1978) Serum and tumour ferritins in primary liver cancer. Gut 19(4):294–299
- Wu SJ, Zhang ZZ, Cheng NS, Xiong XZ, Yang L (2019) Preoperative serum ferritin is an independent prognostic factor for liver cancer after hepatectomy. Surg Oncol 29:159–167
- Xun XD, Li Q (2016) Surgical treatment of intrahepatic cholangiocarcinoma: a retrospective study of 104 cases. Cancer Biol Med 13(4):469–473
- 27. Ma B, Meng H, Shen A, Ma Y, Zhao D, Liu G et al (2021) Prognostic value of inflammatory and tumour markers in Small-Duct subtype intrahepatic cholangiocarcinoma after Curative-Intent resection. Gastroenterol Res Pract 2021:6616062
- Bagante F, Spolverato G, Merath K, Weiss M, Alexandrescu S, Marques HP et al (2019) Intrahepatic cholangiocarcinoma tumor burden: A classification and regression tree model to define prognostic groups after resection. Surgery 166(6):983–990
- Kong J, Cao Y, Chai J, Liu X, Lin C, Wang J et al (2020) Effect of tumor size on Long-Term survival after resection for solitary intrahepatic cholangiocarcinoma. Front Oncol 10:559911
- 30. Sapisochin G, Rodríguez de Lope C, Gastaca M, Ortiz de Urbina J, Suarez MA, Santoyo J et al (2014) Very early intrahepatic cholangiocarcinoma in cirrhotic patients: should liver transplantation be reconsidered in these patients? Am J Transpl 14(3):660–667



- 31. De Martin E, Rayar M, Golse N, Dupeux M, Gelli M, Gnemmi V et al (2020) Analysis of liver resection versus liver transplantation on outcome of small intrahepatic cholangiocarcinoma and combined Hepatocellular-Cholangiocarcinoma in the setting of cirrhosis. Liver Transpl 26(6):785–798
- Jiang BG, Ge RL, Sun LL, Zong M, Wei GT, Zhang YJ (2011) Clinical parameters predicting survival duration after hepatectomy for intrahepatic cholangiocarcinoma. Can J Gastroenterol 25(11):603–608
- 33. Nuzzo G, Giuliante F, Ardito F, De Rose AM, Vellone M, Clemente G et al (2010) Intrahepatic cholangiocarcinoma: prognostic factors after liver resection. Updates Surg 62(1):11–19
- Hercberg S, Estaquio C, Czernichow S, Mennen L, Noisette N, Bertrais S et al (2005) Iron status and risk of cancers in the SU.VI. MAX cohort. J Nutr 135(11):2664–2668
- Tappin JA, George WD, Bellingham AJ (1979) Effect of surgery on serum ferritin concentration in patients with breast cancer. Br J Cancer 40(4):658–660

- Alkhateeb AA, Han B, Connor JR (2013) Ferritin stimulates breast cancer cells through an iron-independent mechanism and is localized within tumor-associated macrophages. Breast Cancer Res Treat 137(3):733–744
- Coffman LG, Parsonage D, D'Agostino R Jr., Torti FM, Torti SV (2009) Regulatory effects of ferritin on angiogenesis. Proc Natl Acad Sci U S A 106(2):570–575
- Matzner Y, Hershko C, Polliack A, Konijn AM, Izak G (1979) Suppressive effect of ferritin on in vitro lymphocyte function. Br J Haematol 42(3):345–353

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

