

RESEARCH ARTICLE

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The effect of an intracerebroventricular injection of metformin or AICAR on the plasma concentrations of melatonin in the ewe: potential involvement of AMPK?

Jean-Baptiste Menassol^{1,2,3,4*}, Claire Tautou^{1,2,3,4}, Armelle Collet^{2,3,4}, Didier Chesneau^{2,3,4}, Didier Lomet^{1,2,3,4}, Joëlle Dupont^{1,2,3,4}, Benoît Malpoux^{1,2,3,4} and Rex J Scaramuzzi^{1,2,3,4,5}

Abstract

Background: It is now widely accepted that AMP-activated protein kinase (AMPK) is a critical regulator of energy homeostasis. Recently, it has been shown to regulate circadian clocks. In seasonal breeding species such as sheep, the circadian clock controls the secretion of an endogenous rhythm of melatonin and, as a consequence, is probably involved in the generation of seasonal rhythms of reproduction. Considering this, we identified the presence of the subunits of AMPK in different hypothalamic nuclei involved in the pre- and post-pineal pathways that control seasonality of reproduction in the ewe and we investigated if the intracerebroventricular (i.c.v.) injection of two activators of AMPK, metformin and AICAR, affected the circadian rhythm of melatonin in ewes that were housed in constant darkness. In parallel the secretion of insulin was monitored as a peripheral metabolic marker. We also investigated the effects of i.c.v. AICAR on the phosphorylation of AMPK and acetyl-CoA carboxylase (ACC), a downstream target of AMPK, in brain structures along the photoneuroendocrine pathway to the pineal gland.

Results: All the subunits of AMPK that we studied were identified in all brain areas that were dissected but with some differences in their level of expression among structures. Metformin and AICAR both reduced ($p < 0.001$ and $p < 0.01$ respectively) the amplitude of the circadian rhythm of melatonin secretion independently of insulin secretion. The i.c.v. injection of AICAR only tended ($p = 0.1$) to increase the levels of phosphorylated AMPK in the paraventricular nucleus but significantly increased the levels of phosphorylated ACC in the paraventricular nucleus ($p < 0.001$) and in the pineal gland ($p < 0.05$).

Conclusions: Taken together, these results suggest a potential role for AMPK on the secretion of melatonin probably acting through the paraventricular nucleus and/or directly in the pineal gland. We conclude that AMPK may act as a metabolic cue to modulate the rhythm of melatonin secretion.

Background

The serine-threonine kinase, AMPK integrates energy homeostasis at both cellular [1] and whole-body levels [2]. It exists as a heterotrimeric complex of a catalytic α subunit and two regulatory β and γ subunits. In mammals there are two or three isoforms of each subunit ($\alpha 1$, $\alpha 2$, $\beta 1$, $\beta 2$, $\gamma 1$, $\gamma 2$, $\gamma 3$), encoded by different genes,

allowing for a variety of heterotrimeric combinations [3]. The treatment with pharmacological agents such as metformin and 5-aminoimidazole-4-carboxamide-1- β -D-ribofuranoside (AICAR) activates the AMPK complex both *in vitro* and *in vivo* [4-6]. This mechanism is central in the action of metformin when used as an insulin-sensitizing agent to treat type II diabetes [7]. Recent evidence points to an emerging role for AMPK as a critical element linking metabolism and reproduction [8]. Indeed, we demonstrated that treatment with either metformin or AICAR significantly inhibited the release

* Correspondence: jbmenassol@gmail.com

¹Physiologie de la Reproduction et des Comportements UMR85, INRA, Nouzilly F-37380, France

Full list of author information is available at the end of the article

of GnRH from immortalised GnRH neurones (GT1-7 cells) and *in vivo*, it shortened the inter-oestrus interval in mice [6]. In sheep as in all seasonal breeding species, the daily rhythm of melatonin secretion is the critical element mediating the synchronising effect of photoperiod on reproduction. This rhythm is endogenously controlled by the activity of the master circadian pacemaker located in the suprachiasmatic nucleus [9]. However, how this endogenous rhythm of melatonin affects the seasonal activity of the GnRH/LH/gonadal axis remains unclear.

Um *et al.* [10] demonstrated that the peripheral activation of AMPK with metformin induced phase-shifts in some of the core clock proteins thus suggesting a role for AMPK in the control of circadian rhythms. In support of these observations we have identified AMPK in the sheep brain and we have tested, the effects of i.c.v. injections of metformin or AICAR on the secretion of melatonin in ewes housed in constant darkness. In parallel, because of the functional interrelationships between melatonin and insulin [11,12], we also monitored the secretion of insulin as a potential metabolic marker of peripheral AMPK activation [13]. Our results suggest that i.c.v. injections of metformin and AICAR both affect the amplitude of the circadian rhythm of melatonin secretion without altering the patterns of plasma insulin secretion.

Methods

Hormones and reagents

The metformin and AICAR used for the i.c.v. injections were obtained from Sigma (Saint Quentin Fallavier, France). The dose injected by means of an intracerebroventricular catheter, was 128 mM for metformin (adapted to sheep from [14]) and 12 mM for AICAR (similar to [6]) in a total volume of 50 μ l. Untreated control ewes were injected with the same volume of an artificial cerebrospinal fluid (CSF) solution, prepared in the laboratory. All solutions had an equivalent osmolarity (\approx 305 mOsm).

Antibodies

Rabbit polyclonal antibodies to phospho-AMPK α Thr172, AMPK α 1/2, AMPK β 1/ β 2, AMPK γ 2, phospho-acetyl-CoA carboxylase (ACC) Ser79 and ACC were purchased from New England Biolabs Inc. (Beverly, MA, USA). A mouse monoclonal antibody to vinculin was obtained from Sigma (Saint Quentin Fallavier, France) and used as an internal standard.

Animals and experimental designs

Nineteen multiparous ewes were surgically fitted with cannulae in the anterodorsal region of their third ventricles [15]. Animals were housed in individual pens in a

light-sealed room under a lightning program mimicking a long-day photoperiod (LP) (16 hours of light/day). All ewes had free access to food, water and mineral licks.

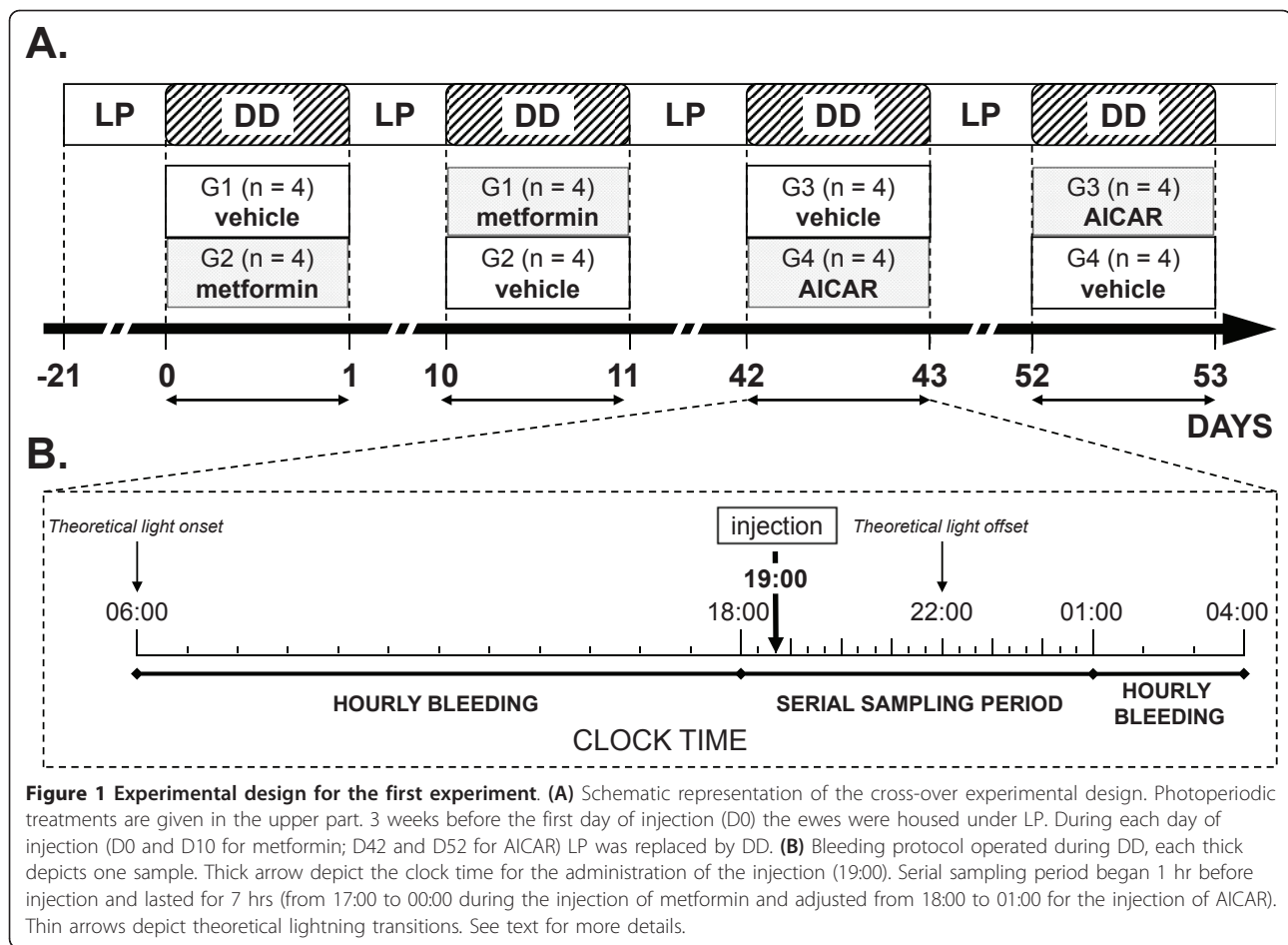
In the first experiment, two groups of ewes (G1, G2, n = 4) balanced for body condition score and live weight were used in a cross-over design so that each group received in random order 9 days apart (day 0 or D0 and D10), either artificial CSF or metformin. Forty days later, groups were re-balanced for their night-time plasma concentrations of melatonin (G3, G4, n = 4), and the experiment was repeated using AICAR with the same cross-over design (D42 and D52) (Figure 1A). For each i.c.v. injection, the ewes were fitted with an indwelling catheter in the left jugular vein and housed in constant darkness (DD) for one whole light/dark (L/D) cycle. The ewes were bled every 60 min for 22 hrs, from 06:00 to 04:00 the next day, with a period of more intensive blood sampling (every 20 min) around the time of the i.c.v. injection at 19:00 (Figure 1B). Blood (3 ml) was collected into heparinised tubes and the plasma was separated by centrifugation (3700 g, 20 min, 4°C) and stored at -20°C.

For the second experiment, two groups of ewes balanced as for experiment 1, were injected with either AICAR (n = 6) or artificial CSF (n = 5). One hour after injection the animals were slaughtered by decapitation, the brain was removed quickly and frozen in isopentane (SDS, Val De Rueil, France). The pituitary (PIT) and pineal (PIN) glands were immediately dissected, frozen in liquid nitrogen and stored at -80°C. Frontal sections (1 mm thick) were sliced from the diencephalon and other structures of interest were dissected using a sterile scalpel, all were frozen in liquid nitrogen and stored at -80°C. The hypothalamic structures of interest (suprachiasmatic nucleus [SCN], arcuate nucleus [ARC], median eminence [ME], paraventricular nucleus [PVN], dorsomedial hypothalamic nucleus [DMH], tanycytes [TAN] and premammillary hypothalamic nucleus [PMH]) were identified using sheep brain atlases [16,17].

All these procedures were approved by the French Agricultural and Scientific Research Agencies and conducted in accordance with the EU guidelines for Care and Use of Agricultural Animals in Agricultural Research and Teaching.

Western-blotting

Western-blotting procedures were conducted as previously described [6] using 60 μ g of protein per sample. The data for the phosphorylation of AMPK and ACC are expressed as the ratio of phosphorylated AMPK to total AMPK and phosphorylated ACC to total ACC respectively. The data for the AMPK subunits have been normalised against an internal standard (vinculin).



Assays

Melatonin was assayed using a radioimmunoassay (RIA) described by Fraser *et al.* [18], with an antibody raised by Tillet *et al.* [19]. The assay sensitivity was 4 pg/mL, the intra-assay coefficient of variation (CV) ranged between 9-14% and the inter-assay CV was 12%. Insulin was also assayed using a RIA that was developed in our laboratory. Plasma concentrations of insulin were determined in duplicate 100 μ L samples. The assay buffer was 0.025 M barbital buffer containing 3% bovine serum albumin (Sigma, Saint Quentin Fallavier, France). Plasma samples were incubated (24 hrs at 4°C) with 300 μ L of the primary antibody (anti-porcine insulin antibody raised in guinea pig, final dilution 1/80000; Sigma, Saint Quentin Fallavier, France) added with 0.6 μ L of Triton X-100 and 0.5 μ L of normal rabbit serum. Samples were then incubated (24 hrs) with 20,000 cpm of iodinated porcine insulin. The next day, samples were incubated for 48 hrs at 4°C with 2 mL of the secondary antibody (anti-guinea pig IgG raised in horses, final dilution 1/333), 4 g/L of CaCl₂ and 30 g/L of polyethylene glycol. Tubes were then centrifuged (3,000 g for 30 min at 4°C), the supernatant was removed, and the radioactivity in

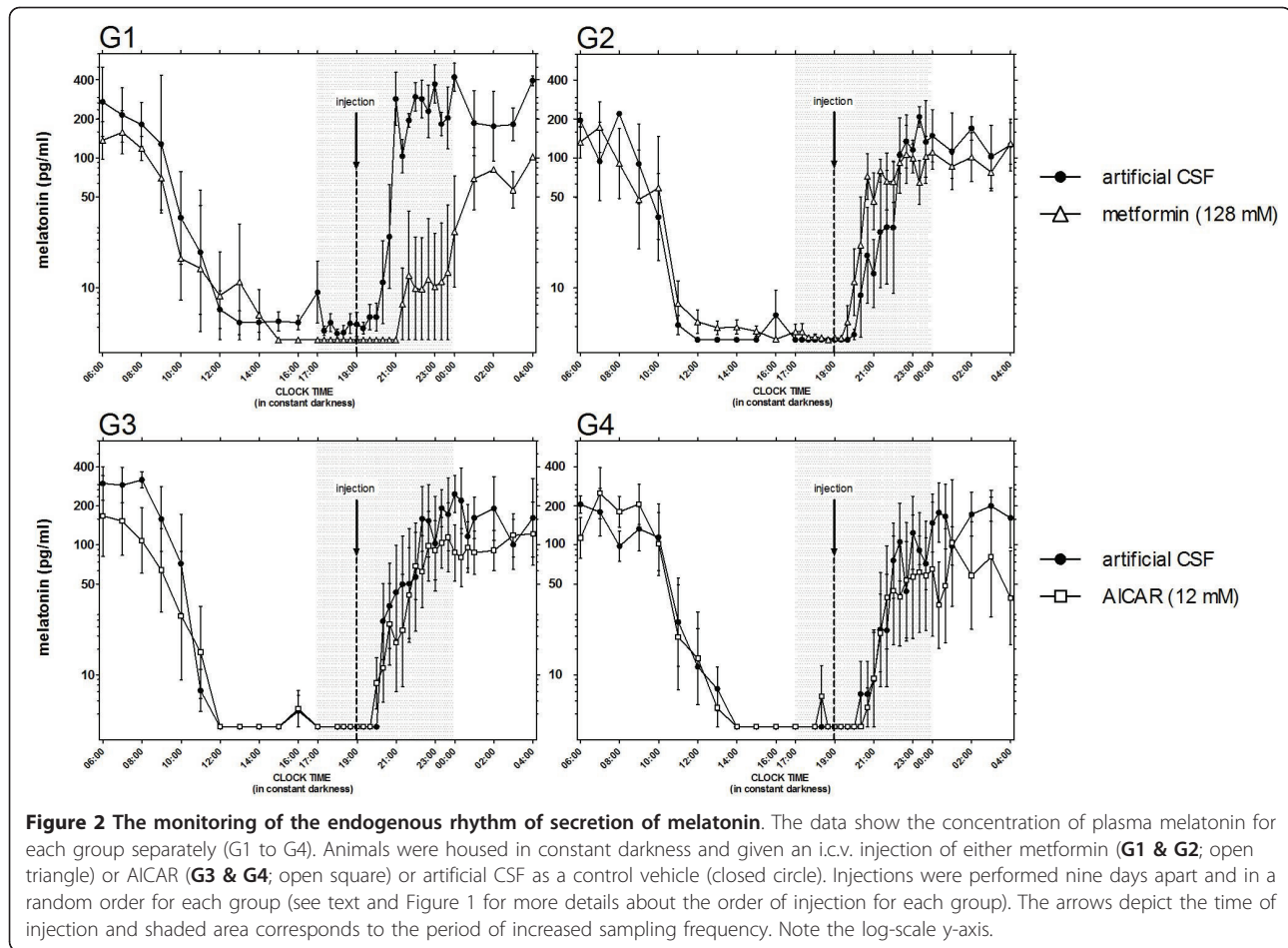
the precipitate was counted with a Wallac Wizard gamma counter (PerkinElmer Inc., Waltham, Massachusetts, USA) for 1 min. The sensitivity of the assay was 0.95 ± 0.06 ng/mL and the intra-assay CV was 12%.

Data and Statistical analysis

Melatonin profiles were analysed by comparing the timing of resumption and the amplitude, of the circadian rhythm of melatonin secretion. These criteria were determined using the method of Malpaux *et al.* [20]. Statistical analyses were performed using the statistical software package, R [21]. Data for insulin were analysed using a mixed-effects model and other data, after log transformation for normalization, were analysed using an analysis of variance. All results are presented as means \pm sem. Differences were considered statistically significant whenever $p < 0.05$.

Results

The amplitude of the endogenous rhythm of melatonin was reduced significantly by both metformin ($p < 0.001$) and AICAR ($p < 0.01$) (Figure 2) but the timing of resumption was not affected by either (data not shown).



After each i.c.v. injection of metformin at D0 and D10 (Figure 2A), the amplitude of the endogenous rhythm of melatonin was reduced when compared to controls ($p < 0.001$). This was also the case between D0 and D10 for the ewes in G1 ($p < 0.001$), that is those ewes injected first with the vehicle solution, but not for the ewes in G2 that is those ewes injected first with metformin. After injection of AICAR (Figure 2B), the amplitude of the endogenous rhythm of melatonin was reduced at D42 compared to controls ($p < 0.001$) but not at D52. However, when compared between D42 and D52, the amplitude of the endogenous rhythm of melatonin was consistently reduced after the injection of AICAR in both groups ($p < 0.05$). There was no significant effect of metformin or AICAR on the concentrations of insulin (Figure 3).

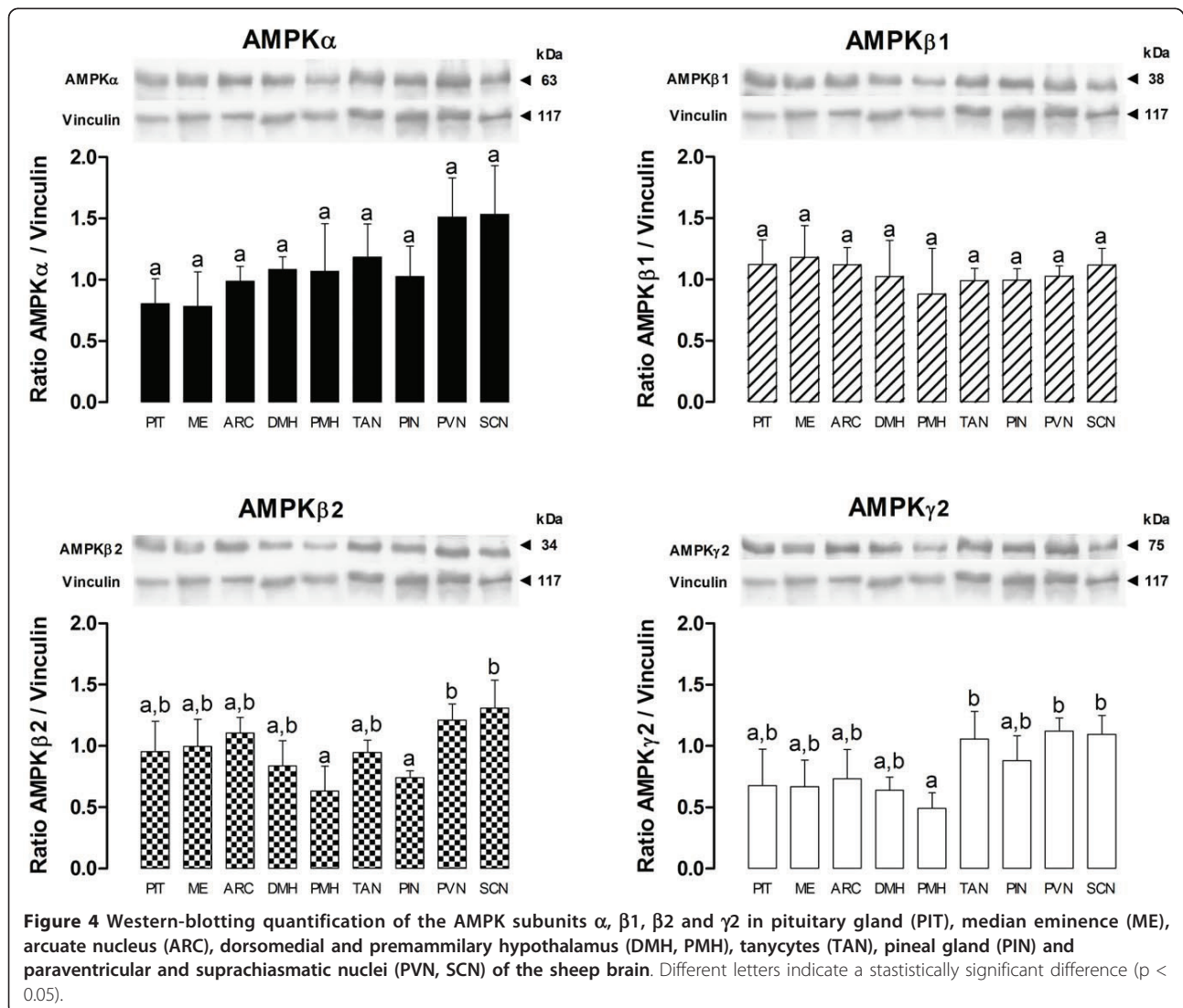
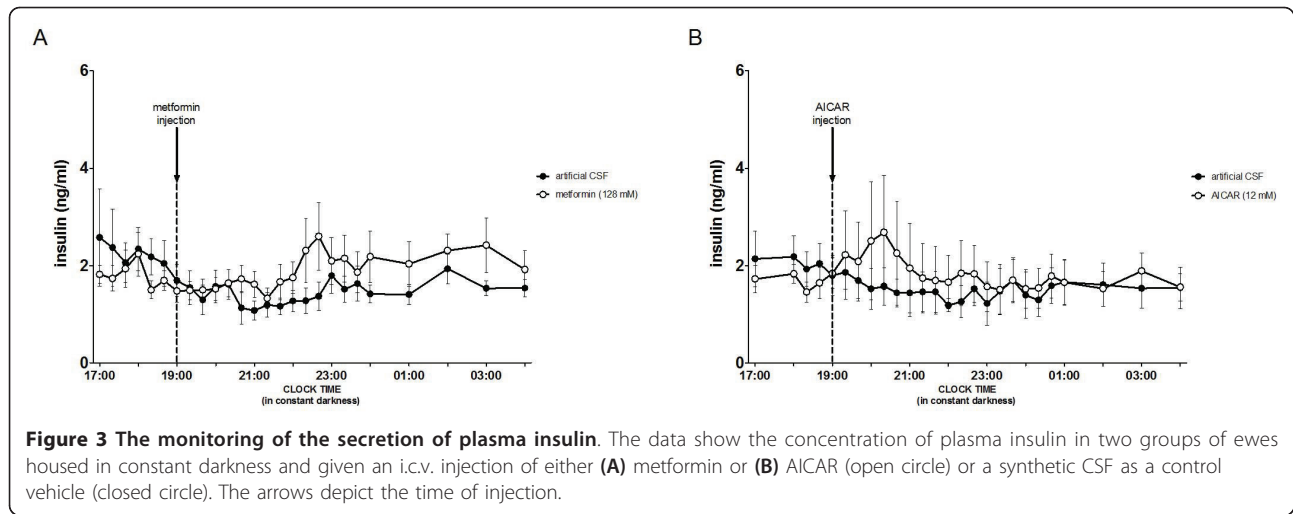
The results of experiment 2 show that the expression levels of the AMPK subunits (Figure 4) were different among hypothalamic structures. The expression of the $\beta 2$ subunit was lower in the PMH/PIN when compared to the PVN/SCN ($p < 0.05$) and the expression of the $\gamma 2$ subunit was lower in the PMH when compared to the

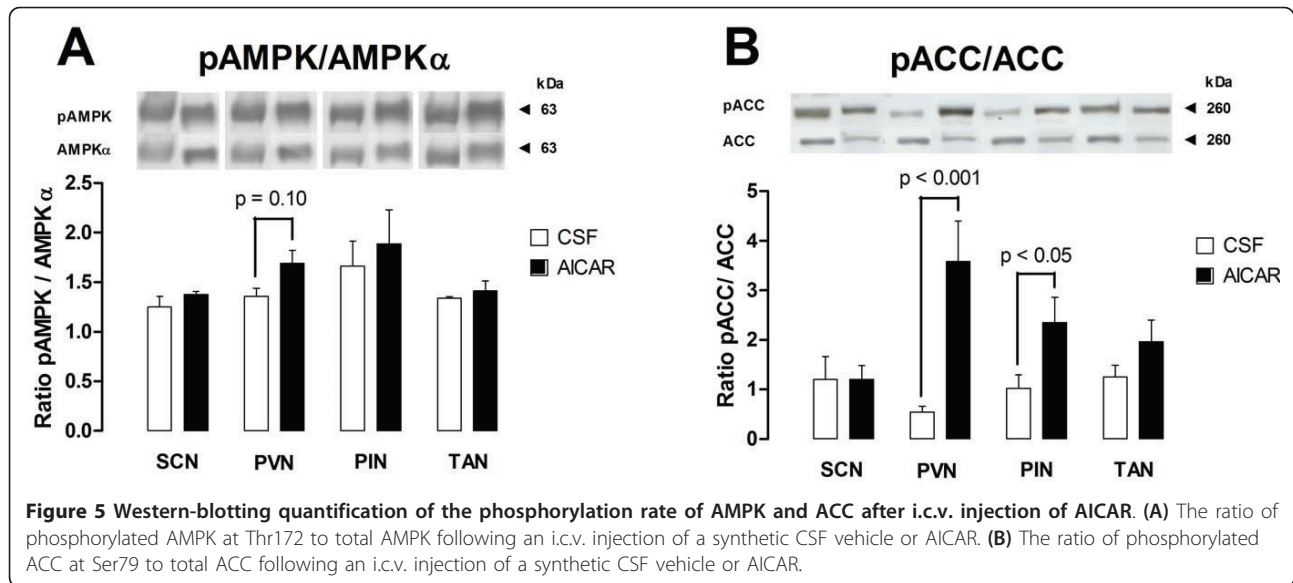
PVN/SCN/TAN ($p < 0.05$). After i.c.v. injection of AICAR, the phosphorylation rate of AMPK tended ($p = 0.1$) to be higher only in the PVN (Figure 5A) but the phosphorylation rate of ACC at Ser79 was significantly higher in both the PVN ($p < 0.001$) and PIN ($p < 0.05$) (Figure 5B).

Discussion

The hypothesis tested in this study, developed from the findings of Um et al. [10], was that central activation of AMPK could disrupt the circadian machinery located in the SCN and thus, alter the endogenous timing of the resumption of melatonin secretion by the pineal gland. In our study, such an effect was not observed after i.c.v. treatment with metformin or AICAR, both known activators of AMPK. Surprisingly we found that another component of the endogenous rhythm of melatonin, its amplitude, was altered by these compounds.

There are three main possibilities to account for the rejection of our hypothesis. First, the i.c.v. treatment with either metformin or AICAR was ineffective and did not stimulate AMPK in the hypothalamic structures we





examined. Indeed, the similarities of the effects between the i.c.v. injections of both drugs on the amplitude of the rhythm of melatonin secretion suggest that they might be mediated by the activation of AMPK, as a common element along their distinct signalling pathways [22]. However, we do not exclude possible AMPK-independent effects [23]. Second, the treatment effectively activated AMPK within the SCN, but in DD when the SCN is released from entrainment by the prevailing LP produced conflicting signals because of the persistence of entrainment by the preceding LP [24] and the expected SCN-driven phase shift in the resumption of melatonin secretion. Third, the treatment effectively activated AMPK but not within the SCN or without triggering phase-shifts in the master circadian machinery perhaps because of functional differences [25] between the master circadian clock and the peripheral-slaves clock that was studied by Um *et al.* [10].

The biological relevance of the lowering effects of both metformin and AICAR on the amplitude of the endogenous rhythm of melatonin is difficult to interpret. Indeed, in sheep as in most photoperiodic species, the duration of melatonin secretion is the critical feature of the rhythm of melatonin secretion that transmits photoperiodic information to the reproductive axis [26]; the role played by the amplitude, the other main feature of this rhythm, remains controversial [27-29]. Some authors have suggested that the amplitude of the melatonin rhythm acts as a potent integrator of environmental factors, other than photoperiod for example, temperature and the food supply (for review see [30]). In relation to this theory, a complementary study (Menassol *et al.*, unpublished data) has established that feed restriction can alter the seasonal reproductive patterns of the Île-de-France ewe as has been

reported in a previous study on sheep [31], and that this alteration is associated with lower concentrations of plasma melatonin. Therefore we speculate that the central activation of AMPK is involved in the integration of metabolic-related cues that alter the amplitude of the rhythm of melatonin secretion. We speculate further that this effect is mediated by neuropeptides or peptidergic hormones that regulate the amplitude of the rhythm of melatonin secretion such as Vasoactive Intestinal Peptide (VIP), Pituitary Adenylate Cyclase Activating Peptide (PACAP) or Vasopressin (VP) (for review see [32]) and particularly Neuropeptide Y (NPY) which is activated by AMPK [33].

The effect of metformin and AICAR on the amplitude of the rhythm of melatonin secretion was not associated with immediate changes in insulin secretion and thus, was not attributable to the interrelationship that exists between these two hormones [11,12,34]. Interestingly, some studies have reported a decrease in melatonin concentrations associated with the development of diabetes [35,36]. Thus, assuming that metformin crosses the blood-brain barrier, one can ask if therapeutic treatment with metformin could not have unreported side-effects on the secretion of melatonin.

The characterization of the expression of AMPK within the hypothalamus, pituitary and pineal glands of the ewes suggests a potentially wide range of action for metformin and AICAR since all subunits were identified in each of the structures that we examined. The differences among structures in the level of expression of the $\beta 2$ and $\gamma 2$ subunits may indicate structural specificities in the sensitivity or functionality of the AMPK complex [2]. Indeed, our investigation of the potential central sites of actions of the AMPK complex, point preferentially to the PVN as a

structure of particular interest because some of its neurons that express melatonin-regulating peptides project to the pineal gland [32]. In this particular structure, the direct assessment of the phosphorylation rate of AMPK only reveals a trend toward significance ($p = 0.1$) for a greater activation after i.c.v. injection of AICAR. However the indirect measurement of AMPK activity, through the assessment of the phosphorylation rate of its downstream target ACC [37,38], shows that AICAR treatment significantly inhibited ACC activity in both the paraventricular nucleus ($p < 0.001$) and the pineal gland ($p < 0.05$). Therefore the PVN is strongly suggested as a key structure that mediates metabolic influences on the rhythm of secretion of melatonin. Moreover, it seems that direct effects of AMPK on the pineal gland activity itself can not be excluded.

In this experiment, the failure to demonstrate significant direct activation of the AMPK complex following treatment with AICAR might be explained by the time elapsed between tissue fixation and injection (1 hr) and the possible action of endogenous phosphatases [39].

Conclusion

To conclude, the i.c.v. administration of AMPK-activators had an unexpected insulin-independent effect on the amplitude of the endogenous rhythm of secretion of melatonin that retrospectively, might have also been observed in natural photoperiodic conditions, on the daily pattern of melatonin secretion. This effect probably involves the AMPK system through the paraventricular nucleus and/or directly within the pineal gland. This mechanism remains to be precisely determined but we suggest that it involves the integration of metabolic-related cues with the photoperiodic signal that synchronizes the annual rhythm of reproduction in sheep.

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Author details

¹Physiologie de la Reproduction et des Comportements UMR85, INRA, Nouzilly F-37380, France. ²Physiologie de la Reproduction et des Comportements UMR6175, INRA, Nouzilly F-37380, France. ³Université François Rabelais de Tours, Tours F-37041, France. ⁴IFCE, Nouzilly F-37380, France. ⁵Department of Veterinary Basic Sciences, Royal Veterinary College, Herts, UK.

Authors' contributions

The authors contributed equally to this study. JBM conceived the study, carried out the experiment, collected data, and drafted the manuscript. CT collected data and performed the melatonin assays. AC and DC followed

the experiment and performed melatonin assays. DL performed the injection of drugs and blood sampling. JD, BM and RJS designed the study. JBM, CT, JD, BM and RJS were involved in the analysis and interpretation of the data. JBM and JD revised the manuscript. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

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References

1. Andersson U, Filipsson K, Abbott CR, Woods A, Smith K, Bloom SR, Carling D, Small CJ: **AMP-activated protein kinase plays a role in the control of food intake.** *J Biol Chem* 2004, **279**(13):12005-12008.
2. Kahn BB, Alquier T, Carling D, Hardie DG: **AMP-activated protein kinase: ancient energy gauge provides clues to modern understanding of metabolism.** *Cell Metab* 2005, **1**(1):15-25.
3. Hardie DG, Scott JW, Pan DA, Hudson ER: **Management of cellular energy by the AMP-activated protein kinase system.** *FEBS Lett* 2003, **546**(1):113-120.
4. Zhou G, Myers R, Li Y, Chen Y, Shen X, Fenyk-Melody J, Wu M, Ventre J, Doebber T, Fujii N, Musi N, Hirshman MF, Goodyear LJ, Moller DE: **Role of AMP-activated protein kinase in mechanism of metformin action.** *J Clin Invest* 2001, **108**(8):1167-1174.
5. Corton JM, Gillespie JG, Hawley SA, Hardie DG: **5-aminoimidazole-4-carboxamide ribonucleoside. A specific method for activating AMP-activated protein kinase in intact cells?** *Eur J Biochem* 1995, **229**(2):558-565.
6. Coyral-Castel S, Tosca L, Ferreira G, Jeanpierre E, Rame C, Lomet D, Caraty A, Monget P, Chabrolle C, Dupont J: **The effect of AMP-activated kinase activation on gonadotrophin-releasing hormone secretion in GT1-7 cells and its potential role in hypothalamic regulation of the oestrous cyclicity in rats.** *J Neuroendocrinol* 2008, **20**(3):335-346.
7. Long YC, Zierath JR: **AMP-activated protein kinase signaling in metabolic regulation.** *J Clin Invest* 2006, **116**:1776-1783.
8. Tosca L, Chabrolle C, Dupont J: **AMPK: a link between metabolism and reproduction?** *Med Sci (Paris)* 2008, **24**(3):297-300.
9. Malpaux B: **Seasonal regulation of reproduction in mammals.** In *Knobil and Neill's Physiology of Reproduction. Volume 1-2*. Third edition. Edited by: Neill JD. The University of Alabama, Birmingham, U.S.A.: Elsevier; 2006:2231-2281.
10. Um JH, Yang S, Yamazaki S, Kang H, Viollet B, Foretz M, Chung JH: **Activation of 5'-AMP-activated kinase with diabetes drug metformin induces casein kinase Iepsilon (CKIepsilon)-dependent degradation of clock protein mPer2.** *J Biol Chem* 2007, **282**(29):20794-20798.
11. Atkins TW, Bailey CJ, Matty AJ: **The effect of melatonin on insulin secretion in rat and mouse.** *J Endocrinol* 1973, **58**:XVII-XVIII.
12. Bailey CJ, Atkins TW, Matty AJ: **Melatonin inhibition of insulin secretion in the rat and mouse.** *Hormone Research* 1974, **5**:21-28.
13. Salt IP, Johnson G, Ashcroft SJ, Hardie DG: **AMP-activated protein kinase is activated by low glucose in cell lines derived from pancreatic beta cells, and may regulate insulin release.** *Biochem J* 1998, **335**(533-539).
14. Foretz M, Hebrard S, Leclerc J, Zarrinpashneh E, Soty M, Mithieux G, Sakamoto K, Andreelli F, Viollet B: **Metformin inhibits hepatic gluconeogenesis in mice independently of the LKB1/AMPK pathway via a decrease in hepatic energy state.** *J Clin Invest* 2010, **120**(7):2355-2369.
15. Thiery JC, Lomet D, Bougoin S, Malpaux B: **Turnover rate of cerebrospinal fluid in female sheep: changes related to different light-dark cycles.** *Cerebrospinal Fluid Res* 2009, **6**:9.
16. Richard P: *Atlas stéréotaxique du cerveau de brebis pré-alpes du sud* Paris: INRA; 1967.
17. Yoshikawa T: **The brain of the sheep.** *Atlas of the Brains of Domestic Animals* Tokyo: University of Tokyo Press; 1968.
18. Fraser S, Cowen P, Franklin M, Franey C, Arendt J: **Direct radioimmunoassay for melatonin in plasma.** *Clin Chem* 1983, **29**(2):396-397.
19. Tillet Y, Ravault JP, Selve C, Evin G, Castro B, Dubois MP: **[Conditions for the use of specific antibodies for immunohistochemical visualization of serotonin and melatonin in the pineal gland of sheep].** *C R Acad Sci III* 1986, **303**(3):77-82.

20. Malpoux B, Wayne NL, Karsch FJ: **Termination of the breeding season in the Suffolk ewe: involvement of an endogenous rhythm of reproduction.** *Biol Reprod* 1988, **39**(2):254-263.
21. R-Development-Core-Team: **R: A language and environment for statistical computing.** Vienna, Austria: R Foundation for Statistical Computing; 2010.
22. Fryer LG, Parbu-Patel A, Carling D: **The Anti-diabetic drugs rosiglitazone and metformin stimulate AMP-activated protein kinase through distinct signaling pathways.** *J Biol Chem* 2002, **277**(28):25226-25232.
23. Guigas B, Bertrand L, Taleux N, Foretz M, Wiernsperger N, Vertommen D, Andreelli F, Viollet B, Hue L: **5-Aminoimidazole-4-carboxamide-1-beta-D-ribofuranoside and metformin inhibit hepatic glucose phosphorylation by an AMP-activated protein kinase-independent effect on glucokinase translocation.** *Diabetes* 2006, **55**(4):865-874.
24. Matthews CD, Seamark RF, Guerin MV: **Plasma melatonin profiles of Romney Marsh sheep in natural photoperiod and in acutely extended darkness.** *J Reprod Fertil* 1992, **95**(3):869-875.
25. Challet E, Caldelas I, Graff C, Pevet P: **Synchronization of the molecular clockwork by light- and food-related cues in mammals.** *Biol Chem* 2003, **384**(5):711-719.
26. Wayne NL, Malpoux B, Karsch FJ: **How does melatonin code for day length in the ewe: duration of nocturnal melatonin release or coincidence of melatonin with a light-entrained sensitive period?** *Biol Reprod* 1988, **39**(1):66-75.
27. Vivien-Roels B, Pitrosky B, Zitouni M, Malan A, Canguilhem B, Bonn D, Pevet P: **Environmental control of the seasonal variations in the daily pattern of melatonin synthesis in the European hamster, *Cricetus cricetus*.** *Gen Comp Endocrinol* 1997, **106**(1):85-94.
28. Garidou ML, Vivien-Roels B, Pevet P, Miguez J, Simonneaux V: **Mechanisms regulating the marked seasonal variation in melatonin synthesis in the European hamster pineal gland.** *Am J Physiol Regul Integr Comp Physiol* 2003, **284**(4):R1043-1052.
29. Zarazaga LA, Malpoux B, Chemineau P: **Amplitude of the plasma melatonin nycthemeral rhythms is not associated with the dates of onset and offset of the seasonal ovulatory activity in the Ile-de-France ewe.** *Reprod Nutr Dev* 2003, **43**(2):167-177.
30. Vivien-Roels B: **Seasonal variations in the amplitude of the daily pattern of melatonin secretion in mammalian and non-mammalian vertebrates: Possible physiological consequences.** In *Comparative Endocrinology and Reproduction*. Edited by: Joy KP, Krishna A, Haldar C. New-Dehli: Narosa Publishing House; 1999:529-542.
31. Forcada F, Abecia JA, Sierra I: **Seasonal changes in oestrus activity and ovulation rate in Rasa Aragonesa ewes maintained at two different body condition levels.** *Small Ruminant Res* 1992, **8**(4):313-324.
32. Simonneaux V, Ribelayga C: **Generation of the melatonin endocrine message in mammals: a review of the complex regulation of melatonin synthesis by norepinephrine, peptides, and other pineal transmitters.** *Pharmacol Rev* 2003, **55**(2):325-395.
33. Claret M, Smith MA, Batterham RL, Selman C, Choudhury AI, Fryer LG, Clements M, Al-Qassab H, Heffron H, Xu AW, Speakman JR, Barsh GS, Viollet B, Vaulont S, Ashford ML, Carling D, Withers DJ: **AMPK is essential for energy homeostasis regulation and glucose sensing by POMC and AgRP neurons.** *J Clin Invest* 2007, **117**(8):2325-2336.
34. Conti A, Maestroni GJ: **Role of the pineal gland and melatonin in the development of autoimmune diabetes in non-obese diabetic mice.** *J Pineal Res* 1996, **20**:164-172.
35. Peschke E, Stumpf I, Bazwinsky I, Litvak L, Dralle H, Muhlbauer E: **Melatonin and type 2 diabetes - a possible link?** *J Pineal Res* 2007, **42**(4):350-358.
36. Peschke E, Frese T, Chankiewicz E, Peschke D, Preiss U, Shreyer U, Spessert R, Müllbauer E: **Diabetic Goto Kakizaki rats as well as type 2 diabetic patients show a decreased diurnal serum melatonin level and an increased pancreatic melatonin-receptor status.** *J Pineal Res* 2006, **40**(135-143).
37. Carling D, Zammit VA, Hardie DG: **A common bicyclic protein kinase cascade inactivates the regulatory enzymes of fatty acid and cholesterol biosynthesis.** *FEBS Lett* 1987, **223**(2):217-222.
38. Park SH, Gammon SR, Knippers JD, Paulsen SR, Rubink DS, Winder WW: **Phosphorylation-activity relationships of AMPK and acetyl-CoA carboxylase in muscle.** *J Appl Physiol* 2002, **92**(6):2475-2482.
39. Voss M, Paterson J, Kelsall IR, Martin-Granados C, Hastie CJ, Pegg MW, Cohen PTW: **Ppm1E is an *in cellulo* AMP-activated protein kinase phosphatase.** *Cell Signal* 2011, **23**:114-124.

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