

# Utility of GDF-15 as a diagnostic biomarker in gastric cancer: an investigation combining GEO, TCGA and meta-analysis

Jie-yu Liu<sup>1</sup>, Xing-xuan Dong<sup>1</sup>, Jia-nan Lu<sup>1</sup>, Yue Zhang<sup>1</sup>, Kai-fan Liu<sup>1</sup>, Ling-feng Liu<sup>2</sup>, Qing-zhi E<sup>2</sup>, Xiao-jing Lu<sup>2</sup>, Jie-yun Yin<sup>1</sup> and Yue-ping Shen<sup>1</sup>

1 Jiangsu Key Laboratory of Preventive and Translational Medicine for Geriatric Diseases, School of Public Health, Medical College of Soochow University, Suzhou, China

2 School of Basic Medicine, Medical College of Soochow University, Suzhou, China

#### Keywords

diagnostic; gastric cancer; GDF-15; metaanalysis

#### Correspondence

J. Yin, School of Public Health, Medical College of Soochow University, 199 Ren Ai Road, Suzhou 215123, China Tel: +86 0512 6588036 E-mail: jyyin@suda.edu.cn and Y. Shen, Jiangsu Key Laboratory of Preventive and Translational Medicine for Geriatric Diseases, Department of Epidemiology and Biostatistics, School of Public Health, Soochow University, 199 Ren-Ai Road, Suzhou, Jiangsu 215123, China E-mail: shenyueping@suda.edu.cn

Jie-yu Liu and Xing-xuan Dong contributed equally to this article.

(Received 26 July 2018, revised 20 September 2018, accepted 27 September 2018)

doi:10.1002/2211-5463.12537

It was recently suggested that growth differentiation factor-15 (GDF-15) is associated with gastric cancer (GC) carcinogenesis. However, the diagnostic potential of GDF-15 for GC remains unclear. To address this issue, we obtained RNA sequencing and microarray data from the Gene Expression Omnibus (GEO) and The Cancer Genome Atlas (TCGA) databases, and searched PubMed, Google Scholar and Web of Science for relevant literature. We then used STATA to perform a meta-analysis. In total, reports of 253 GC patients and 112 healthy controls who contributed peripheral blood samples were taken from the four literature sources, while information on 754 GC tumor and 263 gastric normal tissues was drawn from TCGA and seven GEO datasets. The expression level of GDF-15 mRNA was significantly higher in tumor tissues than in normal tissues, with a standard mean difference (SMD) of 0.79% and a 95% confidence interval (95% CI) of 0.63–0.95. Consistently, the GDF-15 protein in blood was significantly increased in GC patients as compared to controls (SMD = 3.74, 95% CI = 1.81-5.68). In addition, based on information from TCGA and GEO datasets, the expression level of *GDF-15* mRNA may be of use for the diagnosis of GC, with a combined sensitivity, specificity and odds ratio of 0.69 (95% CI = 0.58-0.79), 0.90 (95% CI = 0.84-0.93) and 6.32 (95%) CI = 4.22-9.49), respectively. The summary receiver operating characteristic curve demonstrated that the area under the curve was 0.90 (95% CI = 0.87-0.93). The results suggest higher levels of GDF-15 may be associated with GC tumorigenesis and may have the potential to be a diagnostic biomarker of GC.

Gastric Cancer (GC) is the third leading cause of cancer-related death worldwide [1–4]. By 2015, the total number of patients diagnosed with GC in China was approximately 485 000 and was growing at a speed of 2.63% per year [5]. According to Cancer Statistics in China in 2015, the incidence and mortality rates of GC were 679.1 and 498.0 per 100 000, respectively [5]. Similar to other cancers, GC is a multi-factorial disease, and several genetic and epigenetic factors are involved in its etiology. Environmental risk factors including smoking,

#### Abbreviations

GC, gastric cancer; GDF-15, growth differentiation factor-15; GEO, Gene Expression Omnibus; SMD, standard mean difference; SROC, summary receiver operating characteristic; TCGA, The Cancer Genome Atlas.

FEBS Open Bio **9** (2019) 35–42 © 2018 The Authors. Published by FEBS Press and John Wiley & Sons Ltd. This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.



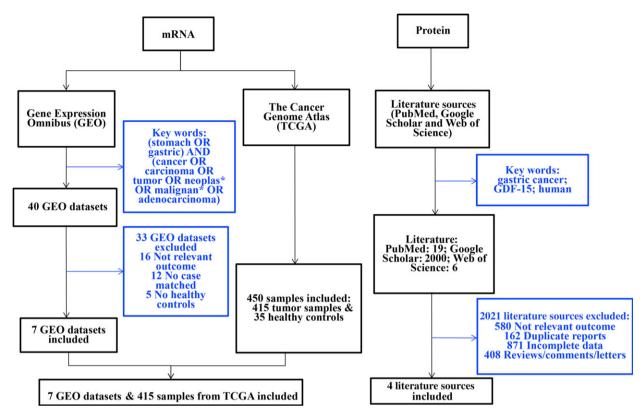


Fig. 1. Flow diagram of literature selection.

|                         |           |                          |          |                  | Tumo | or tissue/Ca | ise     | Norm | nal tissue/ | Control |
|-------------------------|-----------|--------------------------|----------|------------------|------|--------------|---------|------|-------------|---------|
| Dataset                 | Country   | Sample type <sup>a</sup> | Platform | Tested substance | No.  | Mean         | SD      | No.  | Mean        | SD      |
| GSE2685                 | Japan     | Tissues                  | GPL80    | mRNA             | 22   | 373.52       | 225.35  | 8    | 227.38      | 133.58  |
| GSE13911                | Italy     | Tissues                  | GPL570   | mRNA             | 38   | 3645.12      | 3353.66 | 31   | 785.59      | 448.62  |
| GSE19826                | China     | Tissues                  | GPL570   | mRNA             | 12   | 1426.26      | 815.20  | 12   | 525.28      | 156.66  |
| GSE79973                | China     | Tissues                  | GPL570   | mRNA             | 10   | 9.86         | 1.30    | 10   | 9.19        | 0.60    |
| GSE29272                | USA       | Tissues                  | GPL96    | mRNA             | 134  | 7.50         | 1.53    | 134  | 6.68        | 0.89    |
| GSE54129                | China     | Tissues                  | GPL570   | mRNA             | 111  | 7.55         | 1.37    | 21   | 6.48        | 0.43    |
| GSE38932                | Argentina | Tissues                  | GPL5936  | mRNA             | 12   | -0.06        | 0.36    | 12   | -0.28       | 0.24    |
| TCGA                    |           | Tissues                  |          | mRNA             | 415  | 1943.80      | 2137.17 | 35   | 277.69      | 267.26  |
| M. Blanco-Calvo [11]    | Spain     | Peripheral blood         |          | Protein          | 52   | 453.36       | 357.13  | 23   | 212.22      | 84.79   |
| T. Ishige [25]          | Japan     | Peripheral blood         |          | Protein          | 62   | 1159.00      | 579.00  | 22   | 383.00      | 110.00  |
| R. J. E. Skipworth [26] | UK        | Peripheral blood         |          | Protein          | 103  | 1592.00      | 2083.67 | 35   | 377.00      | 911.25  |
| L. Lu [20]              | China     | Peripheral blood         |          | Protein          | 36   | 14.28        | 1.03    | 32   | 1.05        | 0.21    |

SD, standard deviation.

<sup>a</sup>GDF-15 mRNA expression was compared between tumor tissues and normal tissues from gastric cancer patients, while GDF-15 protein levels in peripheral blood were compared between gastric cancer patients and healthy controls.

*Helicobacter pylori* infection and obesity are also suggested to contribute to GC carcinogenesis [1,6,7].

Most GC has no obvious symptoms at the early stage [8]; additionally, it is often mixed up with gastritis [9], gastric ulcer and gastric chronic disease symptoms [10]. Therefore, the majority of GC cases are diagnosed in an advanced stage, with poor prognosis and limited treatment options [3]. Endoscopic biopsy is the best way to find GC before clinical symptoms; however, few patients would like to undergo endoscopy due to potential offensive side effects, including aspiration, pneumonia, bleeding and perforation.

| Study                               |          |                               | %      |
|-------------------------------------|----------|-------------------------------|--------|
| ID                                  |          | SMD (95% CI)                  | Weight |
|                                     |          |                               |        |
| Tissue                              |          |                               |        |
| GSE2685                             |          | 0.71 (-0.12, 1.54)            | 8.06   |
| GSE13911                            | •        | 1.14 (0.63, 1.65)             | 9.22   |
| GSE19826                            | +        | 1.53 (0.62, 2.45)             | 7.71   |
| GSE79973                            |          | 0.66 (-0.24, 1.56)            | 7.78   |
| GSE29272                            | •        | 0.66 (0.41, 0.90)             | 9.88   |
| GSE54129                            | •        | 0.84 (0.36, 1.32)             | 9.32   |
| GSE38932                            | <u> </u> | 0.70 (-0.12, 1.53)            | 8.08   |
| TCGA                                | •        | 0.81 (0.46, 1.16)             | 9.67   |
| Subtotal $(I^2 = 0.0\%, P = 0.585)$ | 0        | 0.79 (0.63, 0.95)             | 69.72  |
| ,                                   |          |                               |        |
| Blood                               |          |                               |        |
| M. Blanco-Calvo                     | •        | 0.80 (0.29, 1.31)             | 9.23   |
| T. Ishige                           | •        | 1.54 (1.00, 2.09)             | 9.12   |
| R.J.E.Skipworth                     | •        | 0.65 (0.26, 1.04)             | 9.57   |
| L.Lu                                |          | <b>•</b> 17.32 (14.33, 20.32) | 2.36   |
|                                     |          |                               |        |

Fig. 2. Forest plot showing SMD of GDF-15 expression between tumor and normal tissues of gastric cancer patients, and between blood of gastric cancer patients and normal controls. Fixed-effects model was used for tissue group, and random-effects model was used for blood group.

0

Hence, it is indispensable to develop more acceptable. convenient and non-invasive diagnostic methods.

-20.3

Subtotal ( $I^2 = 97.5\%$ , P < 0.0001)

Overall  $(I^2 = 91.5\%, P < 0.0001)$ 

Growth differentiation factor-15 (GDF-15), also known as macrophage inhibitory cytokine 1 (MIC1) [11], is a dimeric cytokine belonging to TGF- $\beta$  superfamily involved in the regulation of macrophage activation [12,13]. Under normal physiological conditions, it is highly expressed in the placenta and the prostate, but not common in other organs [13,14], whereas in response to an unfavorable milieu, such as inflammation, oxidative stress, injury, ischemia, telomere erosion and oncogene activation, its production is potently upregulated in a broad range of tissues. For example, GDF-15 is reported to be highly expressed in various malignant cancers, and is associated with the proliferation, metastasis and prognosis of colon cancer, ovarian cancer, oral squamous cell carcinoma, melanoma and prostate cancer [15-18].

Recently, several studies revealed that the expression level of GDF-15 is higher in GC patients, compared with healthy controls [9,11,19-21]. Nevertheless, there are still inconsistent results [22–24]. Besides, the sample size in these studies was relatively small and brought concerns about the robustness of the results. Therefore, we aimed to explore the expression pattern and diagnostic role of GDF-15 in GC by utilizing public data and performing a meta-analysis.

3.74 (1.81, 5.68)

1.30 (0.77, 1.82)

20.3

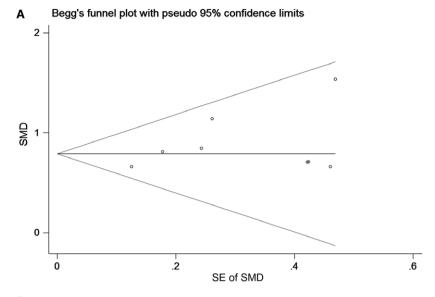
30.28

100.00

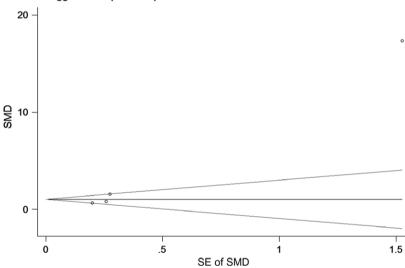
#### Materials and methods

## Search strategy and inclusion criteria

Initially, GC-related RNA-sequencing data were searched in the National Center of Biotechnology Information (NCBI) Gene Expression Omnibus (GEO; http://www.ncbi. nlm.nih.gov/geo/) up to 1 July 2018. The search strategy



B Begg's funnel plot with pseudo 95% confidence limits



**Fig. 3.** Begg's funnel plot for the assessment of potential publication bias in the tissue group (A) and the blood group (B).

was as follows: (stomach OR gastric) AND (cancer OR carcinoma OR tumor OR neoplas\* OR malignan\* OR adenocarcinoma). Afterwards, suitable literature was searched in PubMed, Google Scholar and Web of Science, using a combination of the following mesh words: 'gastric' AND 'cancer' AND 'GDF-15'.

The inclusion criteria of eligible data sets or literature were as follows. First, the study should evaluate the GDF-15 mRNA or protein expression levels. Because the expression pattern of GDF-15 in the body may vary with that in cell lines, which are cultured *in vitro*, we only collected the expression data from peripheral blood or tissues. Second, the expression of GDF-15 was compared between GC patients and healthy controls, or between GC tumor and normal tissues. Third, the expression data of GDF-15 and its mean and standard deviation should be available or

calculable. Fourth, only human samples were included. Figure 1 shows the flow diagram of the literature selection.

According to the inclusion criteria, seven GEO datasets (GSE2685, GSE13911, GSE19826, GSE79973, GSE29272, GSE54129, GSE38932) and four suitable literature sources [11,20,25,26] were finally included.

Finally, we extracted the mRNA expression data of *GDF-15* of GC tumor and normal tissues from The Cancer Genome Atlas Stomach Adenocarcinoma (TCGA-STAD) dataset.

#### **Statistical analysis**

Initially, the expression levels of GDF-15 mRNA or protein were extracted from datasets or articles; the mean and standard deviation were calculated. Later, a meta-analysis was

SPECIFICITY (95% CD)

0.94 [0.81 - 0.99]

0.95 [0.76 - 1.00]

0.80 [0.44 - 0.97]

0.91 [0.85 - 0.95]

0.83 [0.52 - 0.98]

0.92 [0.62 - 1.00

0.87 [0.70 - 0.96

0.71 [0.29 - 0.96]

0.90[0.84 - 0.93]

Q = 11.35, df = 7.00, P = 0.12

 $I^2 = 38.34 [0.00 - 88.74]$ 

1.0

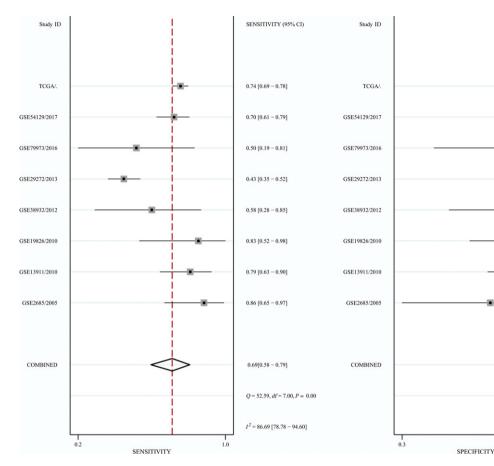


Fig. 4. Diagnostic analysis of tissue *GDF-15* mRNA in gastric cancer.

performed to get pooled standard mean difference (SMD) with 95% confidence interval (95% CI), which indicated the expression differences. Statistical heterogeneity was tested by using Cochran's Q statistic and  $I^2$  tests, and P < 0.05 and  $I^2 > 50\%$  were considered to be statistically heterogeneous [27]. Where this was so, a random-effects model was conducted for combination. Otherwise, a fixed-effects model was used. Additionally, funnel plots and Begg's test were used to check the potential publication bias. A one-way sensitivity analysis (one study excluded at the time) was also performed. Finally, as the original data of protein expression from articles were not available, we only included GDF-15 mRNA expression data from TCGA and GEO datasets, and then conducted diagnostic odds ratio analysis and assessed the diagnostic possibility of GDF-15 for GC patients. Another approach, meta-analysis with summary receiver operating characteristic (SROC), was further carried out to verify the expression level of GDF-15 mRNA in GC.

## Results

As shown in Table 1, 754 tumor and 263 normal tissues were derived from seven GEO (GSE2685, GSE13911, GSE19826, GSE79973, GSE29272, GSE54129, GSE38932) and TCGA datasets. At the same time, the protein expression levels of GDF-15 in peripheral blood of 253 GC patients and 112 controls were obtained from four literature sources.

Figure 2 shows that *GDF-15* mRNA expression was significantly increased in GC tumor tissues compared with normal gastric tissues, with a SMD of 0.79 (95% CI = 0.63-0.95). Also, the expression levels of GDF-15 protein were obviously higher in GC patients than in healthy controls (SMD = 3.74, 95% CI = 1.81-5.68). Besides, no publication bias existed for the tissue group (*P* for Begg's test = 0.536) and for the blood group (*P* for Begg's test = 0.089), as shown in Fig. 3. Additionally, the result remained stable according to the sensitivity analysis (Fig. S1).

Turning to the diagnostic analysis, 754 tumor and 263 normal tissues derived from the TCGA and GEO datasets were included. Figures 4 and 5 illustrate that the combined sensitivity, specificity, and odds ratio are 0.69 (95% CI = 0.58-0.79), 0.90 (95% CI = 0.84-0.93) and 6.32 (95% CI = 4.22-9.49), respectively. The SROC

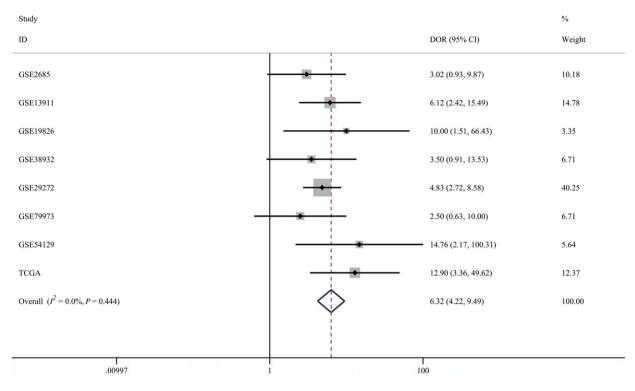


Fig. 5. Forest plot of the diagnostic value of tissue GDF-15 mRNA in gastric cancer.

curve represented in Fig. 6 demonstrated that the area under the curve was 0.90 (95% CI = 0.87-0.93).

#### Discussion

In the present study, it was found that *GDF-15* mRNA was significantly increased in GC tumor tissues compared with normal gastric tissues, while GDF-15 protein was over-expressed in the blood of GC patients compared with healthy controls. The additional diagnostic meta-analysis demonstrated that GDF-15 had a potential diagnostic value for GC. In our results, GDF-15 is highly expressed in GC and the area under the SROC curve was 0.90, which indicated a diagnostic value of GDF-15 in GC tumors compared with non-cancerous control. Hence, GDF-15 may be considered as an early diagnostic biomarker and even a candidate therapeutic target for GC.

In line with our study, other researchers reported that GDF-15 was over-expressed in gastric cancer cell lines [9,19]. In addition, GDF-15 was found to be correlated with progressive pathological parameters in GC [9]. The underlying mechanisms of GDF-15 in GC are not known in detail. Recent study has indicated that the circulating GDF-15 level correlates weakly with systemic inflammation in advanced gastric cancer and may also contribute to fibroblast activation as well

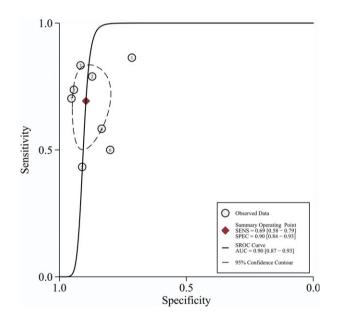


Fig. 6. Summary receiver operating characteristic curve of tissue *GDF-15* mRNA in gastric cancer. AUC, area under curve.

as TGF- $\beta$  [28]. It was also uncovered that the stimulation by GDF-15 of NIH3T3 fibroblasts could enhance proliferation and up-regulate expression of extracellular matrix genes, which were involved in malignant progression [9]. What is more, previous studies implied that GDF-15 could stimulate the urokinase-type plasminogen activator activation system [19] and induce ErbB2 transactivation [29], subsequently enhancing invasiveness of GC cells and eventually contributing to tumorigenesis.

It should be noted that this analysis has some limitations. The results obtained in different study datasets may vary depending on variant conditions. GDF-15 levels might be influenced by many factors, including age, gender, smoking status, diabetes mellitus and so on. Depending on the information we extracted, we were unable to exclude the influence of the above-mentioned variables. Besides, the present study is quite preliminary, and a study determining the pathophysiology of this relationship is urgently warranted.

# Conclusions

In conclusion, the present study suggests that a high level of GDF-15 is associated with GC. In addition, GDF-15 has the potential to serve as a biomarker for GC diagnosis. Further studies exploring the role of GDF-15 in GC carcinogenesis are urgently needed.

### Acknowledgements

This study was supported by the National Natural Science Foundation of China (grant: 81602911), Natural Science Foundation of Jiangsu Province-Youth Project (grant: BK20160337), and Top-notch Academic Programs Project of Jiangsu Higher Education Institutions (grant: SP12101615).

## **Author contributions**

YP-S and JY-Y contributed to the design and conception of the study. The others have contributed to writing and revision of the paper. JY-L and XX-D: acquisition of data, analysis and interpretation of data, and manuscript drafting. KF-L, YZ and JN-L: acquisition of data, and analysis and interpretation of data. XJ-L and QZ-E: analysis and interpretation of data, and critical revision of the manuscript for intellectual content.

## **Conflict of interest**

The authors declare no conflict of interest.

### References

1 Karimi P, Islami F, Anandasabapathy S, Freedman ND and Kamangar F (2014) Gastric cancer: descriptive epidemiology, risk factors, screening, and prevention. *Cancer Epidemiol Biomarkers Prev* 23, 700–713.

- 2 Jemal A, Bray F, Center MM, Ferlay J, Ward E and Forman D (2011) Global cancer statistics. *CA Cancer J Clin* **61**, 69.
- 3 Wils J (2014) Treatment of gastric cancer. *World J Gastroenterol* 56, 1635–1649.
- 4 Sitarz R, Skierucha M, Mielko J, Offerhaus J, Maciejewski R and Polkowski W (2018) Gastric cancer: epidemiology, prevention, classification, and treatment. *Cancer Manag Res* **10**, 239–248.
- 5 Chen W, Zheng R, Baade PD, Zhang S, Zeng H, Bray F, Jemal A, Yu XQ and He J (2016) Cancer statistics in China, 2015. *CA Cancer J Clin* **66**, 115–132.
- 6 Ping Y, Yong Z, Bo C, Wan HW, Jia GQ, Bai H-L and Wu X-T (2009) Overweight, obesity and gastric cancer risk: results from a meta-analysis of cohort studies. *Eur J Cancer* **45**, 2867.
- 7 Li Q, Zhang J, Zhou Y and Qiao L (2012) Obesity and gastric cancer. *Front Biosci* **17**, 2383.
- 8 Sun C, Yuan Q, Wu D, Meng X and Wang B (2017) Identification of core genes and outcome in gastric cancer using bioinformatics analysis. *Oncotarget* 8, 70271–70280.
- 9 Ishige T, Nishimura M, Satoh M, Mai F, Fukuyo M, Semba T, Kado S, Tsuchida S, Sawai S, Matsushita K *et al.* (2016) Combined secretomics and transcriptomics revealed cancer-derived GDF15 is involved in diffusetype gastric cancer progression and fibroblast activation. *Sci Rep* 6, 21681.
- 10 Huang WL, Li YG, Lv YC, Guan XH, Ji HF and Chi BR (2014) Use of lectin microarray to differentiate gastric cancer from gastric ulcer. *World J Gastroenterol* 20, 5474–5482.
- 11 Blanco-Calvo M, TarrãO N, Reboredo M, Haz-Conde M, Garcã J, Quindós M, Figueroa A, Antón-Aparicio L, Calvo L and Valladares-Ayerbes M (2014) Circulating levels of GDF15, MMP7 and miR-200c as a poor prognostic signature in gastric cancer. *Future Oncol* 10, 1187–1202.
- 12 Wang X, Baek SJ and Eling TE (2013) The diverse roles of nonsteroidal anti-inflammatory drug activated gene (NAG-1/GDF15) in cancer. *Biochem Pharmacol* 85, 597–606.
- 13 Ago T and Sadoshima J (2006) GDF15, a cardioprotective TGF-β superfamily protein. *Circ Res* 98, 294–297.
- 14 Bootcov MR, Bauskin AR, Valenzuela SM, Moore AG, Bansal M, He XY, Zhang HP, Donnellan M, Mahler S, Pryor K *et al.* (1997) MIC-1, a novel macrophage inhibitory cytokine, is a divergent member of the TGF-β superfamily. *Proc Natl Acad Sci U S A* 94, 11514–11519.
- 15 Brown DA, Ward RL, Buckhaults P, Liu T, Romans KE, Hawkins NJ, Bauskin AR, Kinzler KW,

Vogelstein B and Breit SN (2003) MIC-1 serum level and genotype: associations with progress and prognosis of colorectal carcinoma. *Clin Cancer Res* **9**, 2642–2650.

- 16 Joshi JP, Brown NE, Griner SE and Nahta R (2011) Growth differentiation factor 15 (GDF15)-mediated HER2 phosphorylation reduces trastuzumab sensitivity of HER2-overexpressing breast cancer cells. *Biochem Pharmacol* 82, 1090–1099.
- 17 Rasiah KK, Kench JG, Gardiner-Garden M, Biankin AV, Golovsky D, Brenner PC, Kooner R, O'neill GF, Turner JJ, Delprado W *et al.* (2006) Aberrant neuropeptide Y and macrophage inhibitory cytokine-1 expression are early events in prostate cancer development and are associated with poor prognosis. *Cancer Epidemiol Biomarkers Prev* 15, 711–716.
- 18 Boyle G, Pedley J, Martyn AC, Banducci K, Strutton G, Brown D, Breit SN and Parsons PG (2009) Macrophage inhibitory cytokine-1 is overexpressed in malignant melanoma and is associated with tumorigenicity. *J Investig Dermatol* **129**, 383.
- 19 Lee DH, Yang Y, Lee SJ, Kim KY, Koo TH, Shin SM, Song KS, Lee YH, Kim YJ, Lee JJ *et al.* (2003) Macrophage inhibitory cytokine-1 induces the invasiveness of gastric cancer cells by up-regulating the urokinase-type plasminogen activator system. *Can Res* 63, 4648–4655.
- 20 Lu L, Ma GQ, Liu XD, Sun RR, Wang Q, Liu M and Zhang PY (2017) Correlation between GDF15, MMP7 and gastric cancer and its prognosis. *Eur Rev Med Pharmacol Sci* 21, 535541.
- 21 Wang X, Li Y, Tian H, Qi J, Li M, Fu C, Wu F, Wang Y, Cheng D, Zhao W *et al.* (2014) Macrophage inhibitory cytokine 1 (MIC-1/GDF15) as a novel diagnostic serum biomarker in pancreatic ductal adenocarcinoma. *BMC Cancer* 14, 578.
- 22 Hippo Y, Taniguchi H, Tsutsumi S, Machida N, Chong JM, Fukayama M, Kodama T and Aburatani H (2002) Global gene expression analysis of gastric cancer by oligonucleotide microarrays. *Can Res* **62**, 233–240.
- 23 He J, Jin Y, Chen Y, Yao HB, Xia YJ, Ma YY, Wang W and Shao QS (2016) Downregulation of ALDOB is

associated with poor prognosis of patients with gastric cancer. *Onco Targets Ther* **9**, 6099–6109.

- 24 Guo J, Bian Y, Wang Y, Chen L, Yu A and Sun X (2016) S100A4 influences cancer stem cell-like properties of MGC803 gastric cancer cells by regulating GDF15 expression. *Int J Oncol* 49, 559–568.
- 25 Ishige T, Nishimura M, Satoh M, Fujimoto M, Fukuyo M, Semba T, Kado S, Tsuchida S, Sawai S, Matsushita K *et al.* (2016) Combined secretomics and transcriptomics revealed cancer-derived GDF15 is involved in diffuse-type gastric cancer progression and fibroblast activation. *Sci Rep* **6**, 21681.
- 26 Skipworth RJ, Deans DA, Tan BH, Sangster K, Paterson-Brown S, Brown DA, Hunter M, Breit SN, Ross JA and Fearon KC (2010) Plasma MIC-1 correlates with systemic inflammation but is not an independent determinant of nutritional status or survival in oesophago-gastric cancer. *Br J Cancer* 102, 665–672.
- 27 Allison KH and Sledge GW (2014) Heterogeneity and cancer. *Oncology* **28**, 772–778.
- 28 Chen YJ (2014) Change of body weight and macrophage inhibitory cytokine-1 during chemotherapy in advanced gastric cancer: what is their clinical significance? (vol 9, e88553, 2014). *PLoS One* 9, e88553.
- 29 Kim K-K, Lee JJ, Yang Y, You K-H and Lee J-H (2008) Macrophage inhibitory cytokine-1 activates AKT and ERK-1/2 via the transactivation of ErbB2 in human breast and gastric cancer cells. *Carcinogenesis* 29, 704–712.

# **Supporting information**

Additional supporting information may be found online in the Supporting Information section at the end of the article.

**Fig. S1.** Sensitivity analysis of the value of GDF-15 in the diagnosis of GC.