

Current Diagnosis and Management of Tangier Disease

Masahiro Koseki¹, Shizuya Yamashita², Masatsune Ogura³, Yasushi Ishigaki⁴, Koh Ono⁵,
Kazuhiisa Tsukamoto⁶, Mika Hori³, Kota Matsuki³, Shinji Yokoyama⁷ and Mariko Harada-Shiba⁸
on behalf of the Committee on Primary Dyslipidemia under the Research Program on Rare and Intractable
Disease of the Ministry of Health, Labour and Welfare of Japan[†]

¹Division of Cardiovascular Medicine, Department of Medicine, Osaka University Graduate School of Medicine, Osaka, Japan

²Department of Cardiology, Rinku General Medical Center, Osaka, Japan

³Department of Molecular Innovation in Lipidology, National Cerebral and Cardiovascular Center Research Institute, Osaka, Japan

⁴Division of Diabetes, Metabolism and Endocrinology, Department of Internal Medicine, Iwate Medical University, Iwate, Japan

⁵Department of Cardiovascular Medicine, Kyoto University Graduate School of Medicine, Kyoto, Japan

⁶Department of Internal Medicine, Teikyo University, Tokyo, Japan.

⁷Institute for Biological Functions, Chubu University, Aichi, Japan

⁸Department of Molecular Pathogenesis, National Cerebral and Cardiovascular Center Research Institute, Osaka, Japan

†The Committee on Primary Dyslipidemia under the Research Program on Rare and Intractable Disease of the Ministry of Health, Labour and Welfare of Japan: Mariko Harada-Shiba (Department of Molecular Pathogenesis, National Cerebral and Cardiovascular Center Research Institute, Osaka, Japan), Shun Ishibashi (Division of Endocrinology and Metabolism, Department of Internal Medicine, School of Medicine, Jichi Medical University, Tochigi, Japan), Shinji Yokoyama (Institute for Biological Functions, Chubu University, Aichi, Japan), Hitoshi Shimano (Department of Internal Medicine (Endocrinology and Metabolism), Faculty of Medicine University of Tsukuba, Tsukuba, Japan), Koutaro Yokote (Department of Endocrinology, Hematology and Gerontology, Chiba University Graduate School of Medicine, Chiba, Japan), Hideaki Bujo (Department of Clinical-Laboratory and Experimental-Research Medicine, Toho University Sakura Medical Center, Chiba, Japan), Shizuya Yamashita (Rinku General Medical Center, Osaka, Japan), Kazuhiisa Tsukamoto (Department of Internal Medicine, Teikyo University, Tokyo, Japan), Toshio Hayashi (School of Health Sciences, Nagoya University Graduate School of Medicine, Nagoya, Japan), Katsunori Ikewaki (Division of Neurology, Anti-Aging, and Vascular Medicine, Department of Internal Medicine, National Defense Medical College, Saitama, Japan), Takanari Gotoda (Department of Metabolic Biochemistry, Faculty of Medicine, Kyorin University, Tokyo, Japan), Kazushige Dobashi (Department of Pediatrics, School of Medicine, University of Yamanashi, Yamanashi, Japan), Yoshihiro Miyamoto (Open Innovation Center, National Cerebral and Cardiovascular Center, Osaka, Japan), Misa Takegami (Department of Preventive Medicine and Epidemiology, National Cerebral and Cardiovascular Center, Osaka, Japan), Yoshiki Sekijima (Department of Medicine (Neurology & Rheumatology), Shinshu University School of Medicine, Matsumoto, Japan), Yasushi Ishigaki (Division of Diabetes, Metabolism and Endocrinology, Department of Internal Medicine, Iwate Medical University, Iwate, Japan), Hiroaki Okazaki (Department of Diabetes and Metabolic Diseases, Graduate School of Medicine, The University of Tokyo, Tokyo, Japan), Atsushi Nohara (Ishikawa Prefectural Central Hospital, Kanazawa, Japan), Shingo Koyama (Division of Neurology and Clinical Neuroscience, Department of Internal Medicine III, Yamagata University Faculty of Medicine, Yamagata, Japan), Kyoko Inagaki (Division of Diabetes, Endocrinology, and Metabolism, Department of Medicine, Nippon Medical School, Tokyo, Japan), Koh Ono (Department of Cardiovascular Medicine, Kyoto University Graduate School of Medicine, Kyoto, Japan), Masahiro Koseki (Division of Cardiovascular Medicine, Department of Medicine, Osaka University Graduate School of Medicine, Osaka, Japan), Hiroyuki Daida (Faculty of Health Science, Juntendo University, Juntendo University Graduate School of Medicine, Tokyo, Japan), Manabu Takahashi (Division of Endocrinology and Metabolism, Department of Internal Medicine, Jichi Medical University, Tochigi, Japan), Kimitoshi Nakamura (Department of Pediatrics, Kumamoto University Graduate School of Medical Sciences, Kumamoto, Japan), Takashi Miida (Department of Clinical Laboratory Medicine, Juntendo University Graduate School of Medicine, Tokyo, Japan), Masa-aki Kawashiri (Department of Cardiovascular Medicine, Graduate School of Medical Sciences, Kanazawa University, Kanazawa, Japan), Tetsuo Minamino (Department of Cardiorenal and Cerebrovascular Medicine, Faculty of Medicine, Kagawa University, Kagawa, Japan), Sachiko Okazaki (Division for Health Service Promotion, The University of Tokyo, Tokyo, Japan), Hayato Tada (Department of Cardiovascular Medicine, Kanazawa University Graduate School of Medical Sciences, Kanazawa, Japan), Jun Wada (Department of Nephrology, Rheumatology, Endocrinology and Metabolism, Okayama University Graduate School of Medicine, Dentistry and Pharmaceutical Sciences, Okayama, Japan), Masatsune Ogura (Division of Lipid Metabolism, Department of Molecular Innovation in Lipidology, National Cerebral and Cardiovascular Center Research Institute, Osaka, Japan), Hirotoshi Ohmura (Department of Cardiovascular Medicine, School of Medicine, Juntendo University, Tokyo, Japan), Mika Hori (Department of Endocrinology, Research Institute of Environmental Medicine, Nagoya University, Nagoya, Japan), Kota Matsuki (Department of Endocrinology and Metabolism, Hirosaki University Graduate School of Medicine, Hirosaki, Japan), Masashi Yamamoto (Department of Endocrinology, Hematology and Gerontology, Chiba University Graduate School of

Medicine, Chiba, Japan), Yasuo Takeuchi (Division of Nephrology, Kitasato University School of Medicine, Kanagawa, Japan), Atsuko Nakatsuka (Department of Nephrology, Rheumatology, Endocrinology and Metabolism, Okayama University Graduate School of Medicine, Dentistry and Pharmaceutical Sciences, Okayama, Japan), Daisaku Masuda (Department of Cardiology, Health Care Center, Rinku Innovation Center for Wellness Care and Activities (RICWA), Rinku General Medical Center, Osaka, Japan), Satoshi Hirayama (Department of Clinical Laboratory Medicine, Juntendo University Graduate School of Medicine, Tokyo, Japan), Masayuki Kuroda (Center for Advanced Medicine, Chiba University Hospital, Chiba University, Chiba, Japan), Takashi Yamaguchi (Center of Diabetes, Endocrinology and Metabolism, Toho University Sakura Medical Center, Chiba, Japan), Takeyoshi Murano (Clinical Laboratory Program, Faculty of Science, Toho University, Chiba, Japan).

Tangier disease is a genetic disorder characterized by an absence or extremely low level of high-density lipoprotein (HDL)-cholesterol (HDL-C). It is caused by a dysfunctional mutation of the ATP-binding cassette transporter A1 (*ABCA1*) gene, the mandatory gene for generation of HDL particles from cellular cholesterol and phospholipids, and it appears in an autosomal recessive hereditary profile. To date, 35 cases have been reported in Japan and 109 cases outside Japan. With dysfunctional mutations in both alleles (homozygotes or compound heterozygotes), the HDL-C level is mostly less than 5 mg/dL and there is 10 mg/dL or less of apolipoprotein A-I (apoA-I), the major protein component of HDL. In patients with Tangier disease, major physical findings are orange-colored pharyngeal tonsils, hepatosplenomegaly, corneal opacity, lymphadenopathy, and peripheral neuropathy. Although patients tend to have decreased low-density lipoprotein (LDL)-cholesterol (LDL-C) levels, premature coronary artery disease is frequently observed. No specific curative treatment is currently available, so early identification of patients and preventing atherosclerosis development are crucial. Management of risk factors other than low HDL-C is also important, such as LDL-C levels, hypertension and smoking. Additionally, treatment for glucose intolerance might be required because impaired insulin secretion from pancreatic beta cells has occasionally been reported.

Key words: Tangier disease, HDL, Reverse cholesterol transport, *ABCA1*, Cholesterol efflux, Orange tonsil, Atherosclerosis

Introduction

Tangier disease is an autosomal recessive disease characterized by extremely low levels or absence of high-density lipoprotein (HDL)-cholesterol (HDL-C) and apolipoprotein A-I (apoA-I)¹⁾. The disease was named after Tangier Island in Chesapeake Bay, Virginia, USA, where the first case was discovered in 1960 and reported in 1961²⁾. In 1991, generation of HDL particles through the direct action of helical HDL apoproteins on cells was first reported³⁾, and this was found to be deficient in cells derived from a patient with Tangier disease in 1995⁴⁾. Eventually, ATP binding cassette transporter A1 (*ABCA1*) was identified as the gene responsible for this action and for Tangier disease in 1999⁵⁻⁷⁾. Sequential progress in the investigation of HDL biosynthesis showed that HDL particles generated through *ABCA1*-dependent interaction of apolipoproteins with cells are the main

source of plasma HDL. Patients with homozygous or compound heterozygous mutations in the *ABCA1* gene display the phenotype of Tangier disease and heterozygotes have decreases in HDL-cholesterol to various extents.

1. Disease Frequency

The number of cases of Tangier disease reported by 2020 was 35 in Japan and 109 in other countries (possibly including duplicates), indicating that it is a rather rare disease^{8, 9)}. However, the frequency of dysfunctional mutations in the *ABCA1* gene in the general population is not clear. A recent article using the Exome Aggregation Consortium database reported that 1 in 400 individuals in the general population is a heterozygote for a loss-of-function variant in the *ABCA1* gene on the basis of allele frequencies (frameshift, nonsense and splicing only; not missense), indicating a global prevalence of Tangier disease of at

Address for correspondence: Masahiro Koseki, Division of Cardiovascular Medicine, Department of Medicine, Osaka University Graduate School of Medicine, 2-2 Yamadaoka, Suita, Osaka 565-0871, Japan E-mail: koseki@cardiology.med.osaka-u.ac.jp

Received: March 18, 2021 Accepted for publication: March 21, 2021

Copyright©2021 Japan Atherosclerosis Society

This article is distributed under the terms of the latest version of CC BY-NC-SA defined by the Creative Commons Attribution License.

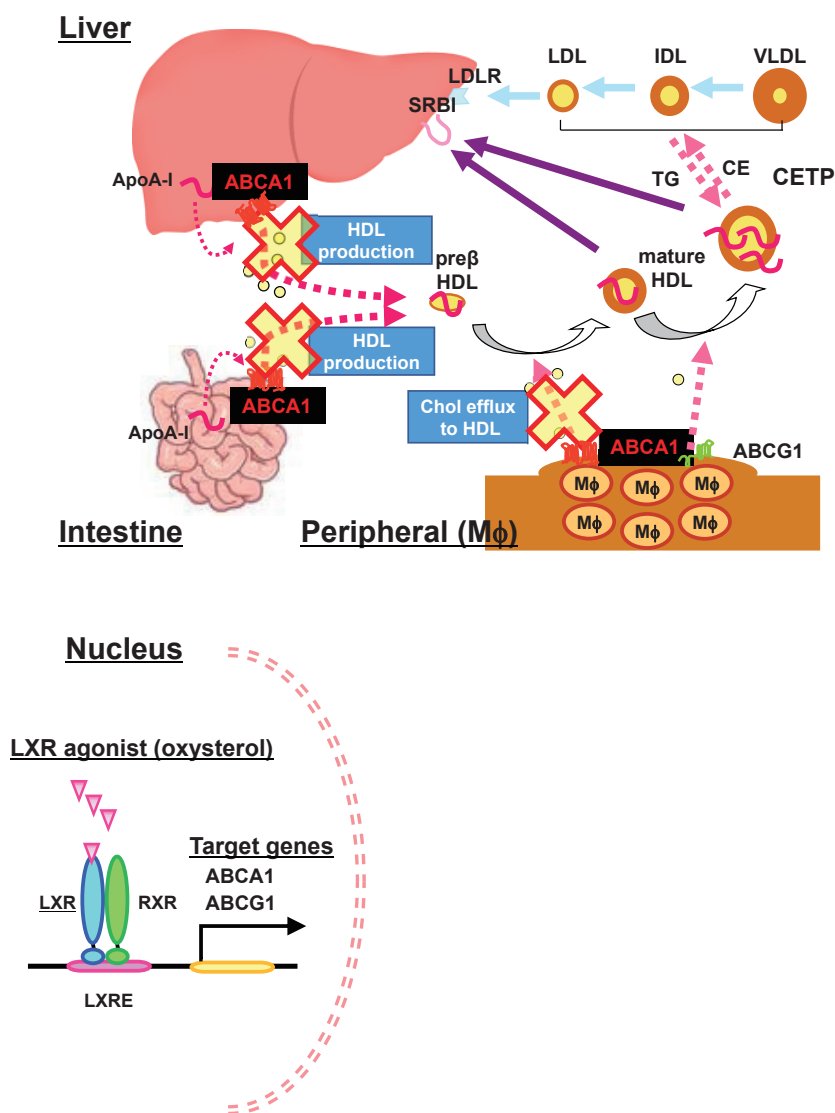


Fig. 1. Roles of ABCA1 in formation of HDL particles, reverse cholesterol transport and pathogenesis of Tangier disease

least 1 in 640,000¹⁰). It therefore seems that a substantial number of cases might go undiagnosed.

2. Genetic Backgrounds and Pathophysiology

ABCA1 is a member of the ATP-binding cassette transporter membrane protein family. It is an essential factor for generation of nascent HDL particles with extracellular helical apolipoproteins, through transport of cellular phospholipids and cholesterol (Fig. 1). This process is the major source of plasma HDL and one of the major mechanisms of cholesterol export from cells. It may be essential for final processing of cholesterol in mammalian somatic cells that are unable to catabolize cholesterol molecules. Peripheral cells sense intra-cellular cholesterol levels and increase ABCA1 expression for its excretion¹¹), while it undergoes

bidirectional control in hepatocytes to prevent cholesterol recovered from peripheral cells flowing back into blood plasma^{12, 13}). With functional deficiency in ABCA1, spherical HDL particles are not produced resulting in extremely low plasma HDL-C levels, which, in Tangier disease patients, are about one third of the normal level. The reason for this is unclear but it has been postulated that a substantial portion of cholesterol molecules in LDL in human plasma are those which have been acyl-esterified in HDL and transferred to VLDL/LDL and that a severe decrease in HDL cholesterol may lead to a decrease in LDL-C level.

In Tangier disease patients, cellular cholesterol export is impaired due to ABCA1 deficiency in peripheral cells, including macrophages and Schwann

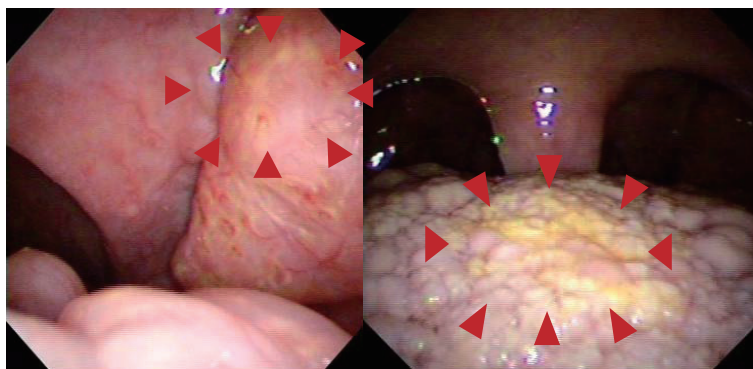


Fig. 2. Orange-colored tonsils observed in male patient with Tangier disease in his 50s
Arrow heads indicate palatine tonsil (left panel) and lingual tonsil (right panel). Reproduced from reference [8].

cells. Cholesterol therefore accumulates in these cells, causing orange-colored pharyngeal tonsillar swelling, corneal opacity, hepatosplenomegaly, lymphadenopathy and peripheral neuropathy. However, impairment of the initial stage of reverse cholesterol transport should be considered to be a risk for developing atherosclerotic diseases even though plasma LDL-C concentrations could be reduced.

ABCA1 appears to destabilize the raft structure, a cholesterol-rich domain of the plasma membrane^{14, 15}, its deficiency leads to an increase in “lipid” rafts and it has been suggested that this increases secretion of inflammatory cytokines¹⁶. It has also been reported that the insulinogenic index decreases due to cholesterol accumulation in pancreatic β -cells, which often accompanies glucose intolerance^{17, 18}. These metabolic disorders are collectively involved in the development of premature coronary artery disease⁸.

In early studies of Tangier disease, Schaefer et al. revealed the kinetics of plasma lipoprotein metabolism using externally labeled injected HDL and found that apoA-I was catabolized at a much greater fractional rate in patients¹⁹. Their data, however, should be reinterpreted in terms of external HDL catabolism on the basis of ABCA1 deficiency. A recent study using human pluripotent stem cell-derived hepatocytes has demonstrated that ABCA1 deficiency increases angiopoietin-like protein 3 secretion, which is consistent with increased triglyceride in the plasma of Tangier patients²⁰.

3. Clinical Manifestations

3.1. Abnormal Plasma Lipoproteins

Plasma HDL-C is mostly low, at 5 mg/dL or less (mean of identified cases 3 ± 3 mg/dL), and the apoA-I concentration is 10 mg/dL or less²¹. Plasma LDL-C is also reduced, to around 37% of the average normal level. The appearance of remnant lipoprotein particles

(intermediate products between VLDL and LDL) rich in triglycerides has been reported and this was found to result in abnormal small, dense LDL particles²¹. In subjects with heterozygous mutations of the *ABCA1* gene, plasma HDL-C and apoA-I levels are often reduced to about 50% of that in normal subjects, though the extent of HDL-C decline is not consistent.

3.2. Physical Findings

Impairment of HDL biogenesis results in reduced cholesterol export, which leads to lipid accumulation in cells. A typical finding of this disorder is orange-colored tonsils (Fig. 2)⁸. The tonsils of patients are lobulated and swollen and present a bright orange or yellow-grey surface. A history of recurrent tonsillitis or tonsillectomy is often noted. In addition, splenomegaly and associated thrombocytopenia and/or reticulocyte hyperplasia may be present (Fig. 3). Hepatomegaly is also observed in about one-third of cases, but liver dysfunction is not usually present²². There is also cholesterol accumulation in other organs, such as lymph nodes, thymus, intestinal mucosa, and skin. Its accumulation in the cornea causes corneal opacity.

3.3. Peripheral Neuropathy

Various peripheral neuropathies, ranging from mild to severe, have been reported. Sensory, motor or mixed disorders appear transiently or persistently. Reduced deep perception and tendon reflexes are rare. Peripheral neuropathy appears as a recurrent asymmetric disorder of peripheral nerves including cranial nerves, as neuropathy with symmetry in the lower limbs, or syringomyelia-like peripheral neuropathy^{23, 24}.

3.4. Cardiovascular Diseases

It has been reported that 12 out of 35 patients

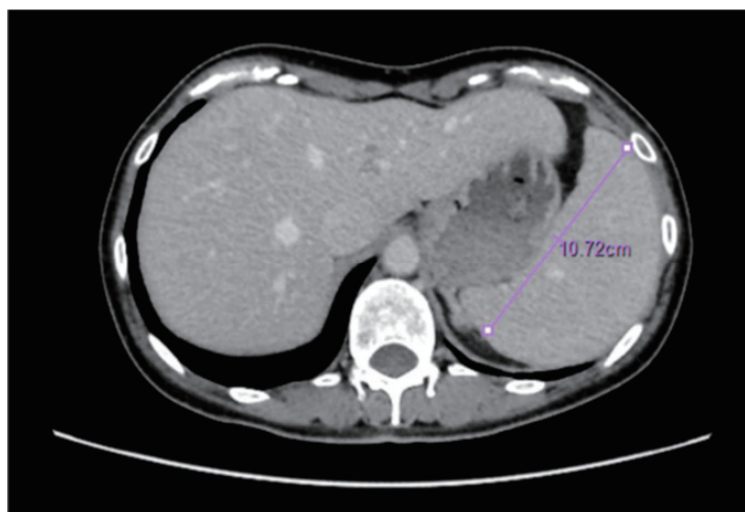


Fig. 3. Abdominal CT scan of splenomegaly in female patient with Tangier disease in her 40s
The photo was kindly provided by one of the co-authors, Prof. Yasushi Ishigaki (Iwate Medical University).

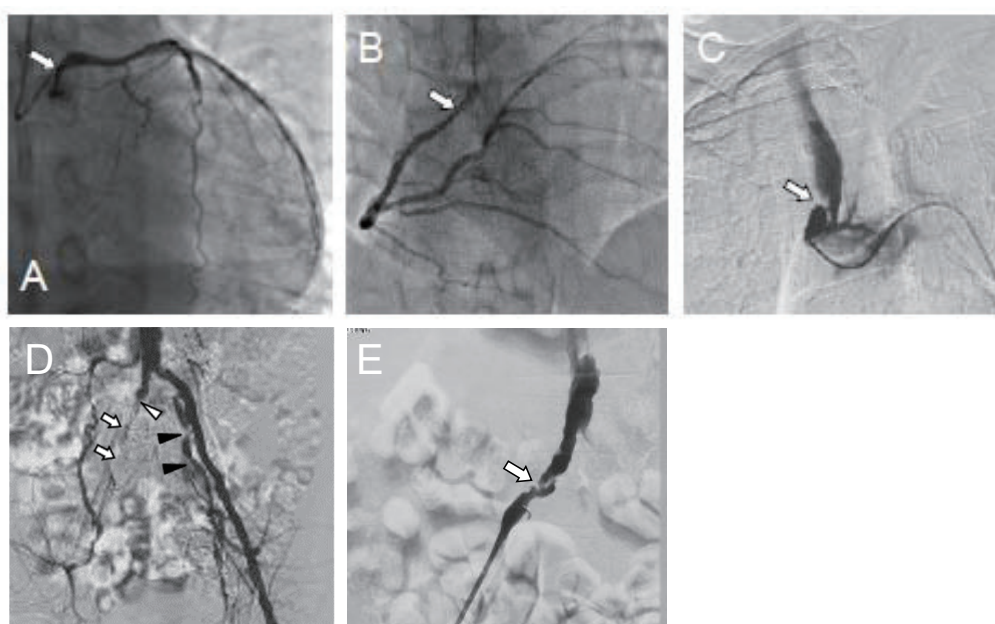


Fig. 4. Advanced systemic atherosclerotic lesions in male patient with Tangier disease in his 50s

Arrows indicate stenosis or occlusion of the artery.

A: left coronary artery, B: right coronary artery, C: brachiocephalic artery, D: left iliac artery, E: right external iliac artery [8].

(34.3%) in Japan and 34 out of 109 patients (31.2%) in other countries had some type of cardiovascular disease, suggesting accelerated atherogenicity in Tangier disease (Fig. 4)⁸⁾. A previous case study using intravascular ultrasound (IVUS) revealed diffuse calcified coronary artery lesions²⁵⁾, which might have been affected by HDL deficiency and glucose intolerance¹⁷⁾.

4. Diagnostic Criteria and Differential Diagnosis

Diagnostic criteria for Tangier disease are given in Table 1 and the flow chart for differential diagnosis of hypo-HDL-cholesterolemia is shown in Fig. 5, based on discussions by the Committee on Primary Dyslipidemia under the Research Program on Rare and Intractable Diseases of Japan's Ministry of Health, Labour and Welfare.

Inherited diseases that lead to hypo-HDL-

Table 1. Diagnostic criteria

<p>A. Required laboratory test results</p> <ol style="list-style-type: none"> 1. Plasma (serum) HDL-cholesterol less than 25 mg/dL 2. Plasma (serum) apoA-I concentration less than 20 mg/dL <p>B. Clinical symptoms</p> <ol style="list-style-type: none"> 1. Orange-colored tonsillar swelling 2. Hepatomegaly and/or splenomegaly 3. Corneal opacity 4. Peripheral neuropathy 5. Cardiovascular disease <p>C. Differential diagnosis</p> <p>The following diseases should be excluded; LCAT deficiency, apoA-I deficiency and secondary hypo-HDL-cholesterolemia*</p> <p>D. Genetic testing**</p> <p>Identification of pathogenic mutations in the <i>ABCA1</i> gene</p> <p>< Diagnostic category ></p> <p>Definite: Patients should satisfy both of required laboratory test results (A) AND at least one clinical symptom of (B) AND should be excluded for the diseases of differential diagnosis (C) AND should be positive for genetic testing (D).</p> <p>Probable: Patients should satisfy both of required laboratory test results (A) AND at least two clinical symptoms of (B) AND should be excluded for the diseases of differential diagnosis (C).</p> <p>Tangier disease can be diagnosed if patients are categorized “Definite” or “Probable”.</p>
--

*Secondary hypo-HDL-cholesterolemia: After surgery, liver disorders (especially liver cirrhosis and severe hepatitis, including convalescent stage), acute phase of systemic inflammatory disease, debilitating diseases such as cancer, history of oral probucol within the past 6 months, and combined probucol and fibrate (including fibrate administration after discontinuation of probucol).

**When differential diagnosis is difficult, genetic testing for *ABCA1* mutations should be performed. The diagnosis can be definite if pathogenic mutations in the *ABCA1* gene are identified.

cholesterolemia (Familial hypoalphalipoproteinemia) include classical lecithin: cholesterol acyltransferase (LCAT) deficiency, fish-eye disease, and familial apoA-I deficiency. Corneal opacity is commonly observed in these diseases, but tonsillar swelling and peripheral neuropathy are specific to Tangier disease, and xanthomas are found only in familial apoA-I deficiency^{24, 26}.

Care should be taken to exclude secondary hypo-HDL-cholesterolemia, such as in severe liver diseases, especially liver cirrhosis, and drug-induced hypo-HDL-cholesterolemia, most frequently due to combination of probucol and fibrates. It should be noted that probucol may continue to influence lipid profiles for several months after discontinuation, so HDL-C may be significantly reduced by switching from probucol to fibrates or permafibrate, a selective peroxisome proliferator-activated receptor alpha modulator (SPPARM α), in some cases.

5. Current Management of Tangier Disease

Based on the patients identified in Japan to date, there seem to be no distinct differences in clinical or genetic profiles with patients in other countries^{8, 9}. No curative treatment, such as gene therapy for the *ABCA1* gene, has yet been established. Since extremely

enhanced risk of atherosclerotic diseases is the major clinical problem, patients should be carefully monitored for presence of atherosclerotic lesions through regular testing including exercise electrocardiography, echocardiography and computed tomography coronary angiography⁸). The management of atherosclerotic risk factors, such as hypertension, smoking and diabetes mellitus, is crucial²⁷. Plasma LDL-C levels are generally low in patients with Tangier disease but if this is not the case, they should be reduced through administration of statins or other means. Impairment of the insulinogenic index should be estimated using a 75 g oral glucose tolerance test¹⁷.

Conclusions and Future Perspectives

Gene therapy for *ABCA1* gene may have the greatest potential for Tangier disease. There is the possibility that restoration of *ABCA1* expression in the liver would raise serum HDL-C levels but this might not be enough to recover cellular cholesterol efflux and suppress extra lipid accumulation in cells in atherogenic lesions such as macrophages, smooth muscle cells and endothelial cells. It may not be easy to develop a fundamental therapy for Tangier disease.

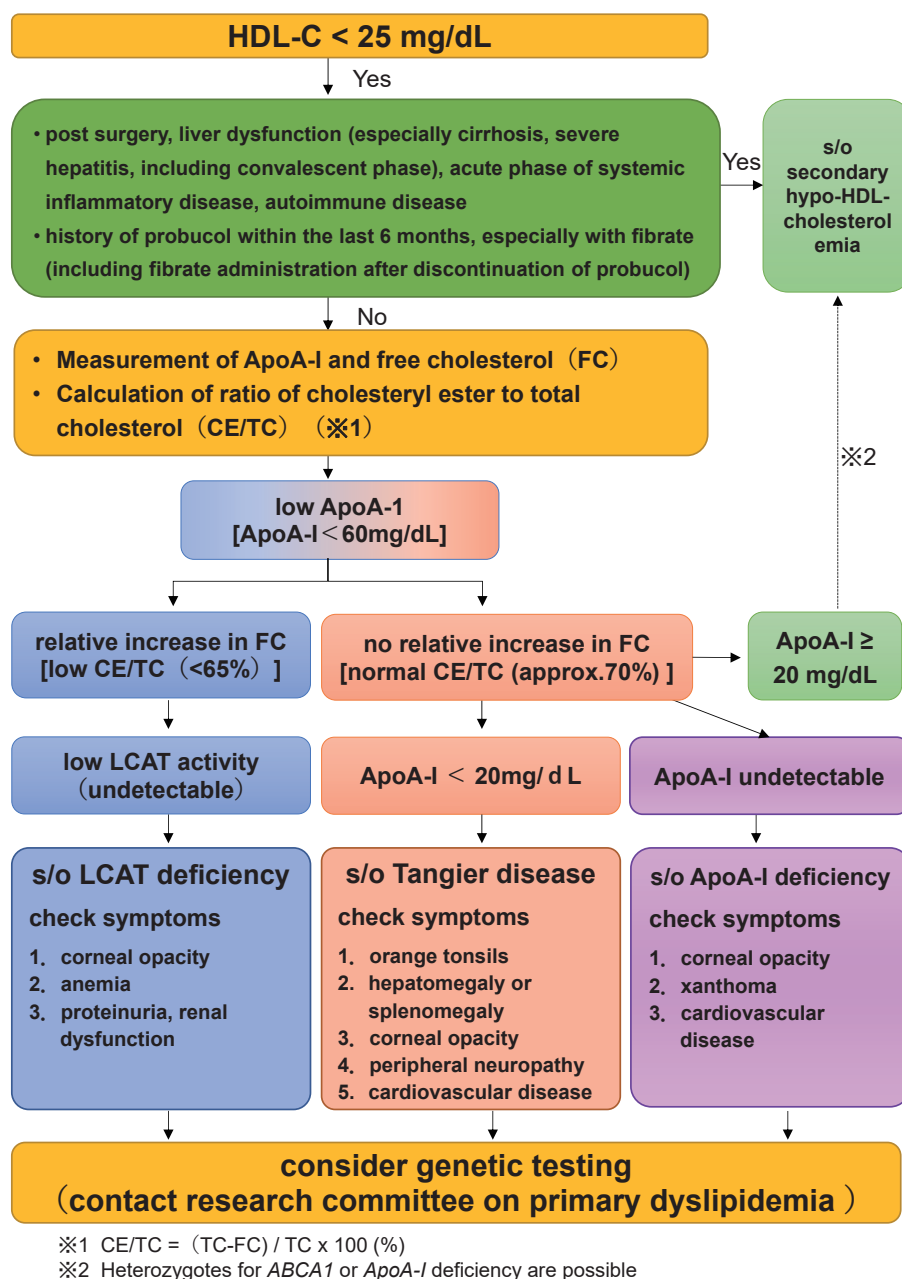


Fig. 5. Differential diagnosis flow chart for hypo-HDL-cholesterolemia

Acknowledgments and Notice of Grant Support

This work has been supported by Health, Labour and Welfare Sciences Research Grant for Research on Rare and Intractable Diseases (H30-nanji-ippan-003).

Conflicts of Interest

Atsushi Nohara has nothing to disclose. Hayato Tada has nothing to disclose. Masatsune Ogura has

received honoraria from Amgen Inc., Astellas Pharma Inc. Sachiko Okazaki has received scholarship grants from Minophagen Pharmaceutical Co., Ltd., Kowa Company, Ltd. Koh Ono has nothing to disclose. Hitoshi Shimano has nothing to disclose. Hiroyuki Daida has received honoraria from Amgen Inc., Daiichi-Sankyo Co., Ltd., Kowa Co., Ltd., and MSD K.K., Novartis Pharma K.K., Bayer Yakuhin, Ltd. and received clinical research funding from Canon Medical Systems Corporation, Philips Japan, Ltd., Toho Holdings Co., Ltd., Asahi Kasei Corporation, and

Inter Reha Co., Ltd. HD has also received scholarship grants from Nippon Boehringer Ingelheim Co., Ltd., Otsuka Pharmaceutical Co., Ltd., Sanofi K.K., MSD K.K., Daiichi-Sankyo Co., Ltd., Pfizer Co., Ltd., Mitsubishi Tanabe Pharma Corp., Astellas Pharma Inc., Takeda Pharmaceutical Co., Ltd., Teijin Pharma, Ltd., Shionogi & Co., Ltd., Actelion Pharmaceuticals, Ltd., Actelion Ltd., Kowa Co., Ltd., Bayer Yakuhin, Ltd. HD has also courses endowed by companies, including Philips Japan, Ltd., ResMed, Fukuda Denshi Co., Ltd., and Paramount Bed Co., Ltd. Kazushige Dobashi has nothing to disclose. Toshio Hayashi has nothing to disclose. Mika Horii has nothing to disclose. Kota Matsuki has nothing to disclose. Tetsuo Minamino has nothing to disclose. Shinji Yokoyama has nothing to disclose. Mariko Harada-Shiba has received stock holdings or options from Liid Pharma, honoraria from Amgen Inc., Astellas Pharma Inc., Sanofi, and scholarship grants from Aegerion Pharmaceuticals, Inc., Recordati Rare Diseases Japan, and Kaneka Corporation. Katsunori Ikewaki has nothing to disclose. Yasushi Ishigaki has nothing to disclose. Shun Ishibashi has received honoraria from Kowa Co., Ltd., and a scholarship grant from Ono Pharmaceutical Co., Ltd. Kyoko Inagaki has nothing to disclose. Hirotohi Ohmura has nothing to disclose. Hiroaki Okazaki has received scholarship grants from Minophagen Pharmaceutical Co., Ltd., Kowa Company, Ltd. Masa-aki Kawashiri has nothing to disclose. Masayuki Kuroda has nothing to disclose. Masahiro Koseki has received clinical research funding from Kowa Company, Ltd., Rohto Pharmaceutical Co., Ltd. Takanari Gotoda has nothing to disclose. Shingo Koyama has nothing to disclose. Yoshiki Sekijima has nothing to disclose. Manabu Takahashi has nothing to disclose. Yasuo Takeuchi has nothing to disclose. Misa Takegami has nothing to disclose. Kazuhisa Tsukamoto has received honoraria from Bayer Yakuhin, Ltd., MSD Ltd., Takeda Pharmaceutical Company Ltd., and scholarship grants from Mitsubishi Tanabe Pharma Corporation., Bayer Yakuhin, Ltd., Sanofi K.K. Atsuko Nakatsuka has nothing to disclose. Kimitoshi Nakamura has nothing to disclose. Satoshi Hirayama has nothing to disclose. Hideaki Bujo has nothing to disclose. Daisaku Masuda has received clinical research funding from MSD K.K., Ono Pharmaceutical Co., Ltd., Takeda Pharmaceutical Co., Ltd., Kowa Co., Ltd. Takashi Miida has nothing to disclose. Yoshihiro Miyamoto has nothing to disclose. Takeyoshi Murano has nothing to disclose. Takashi Yamaguchi has nothing to disclose. Shizuya Yamashita has received honoraria from Kowa Company, Ltd., MSD K.K. Masashi Yamamoto has nothing to disclose. Koutaro

Yokote has received honoraria from Kowa Company, Ltd., MSD K.K., Astellas Pharma Inc., Mitsubishi Tanabe Pharma Corp., Amgen K.K., Takeda Pharmaceutical Company Limited, Sanofi K.K., Ono Pharmaceutical Co., Ltd., AstraZeneca K.K., Daiichi-Sankyo Co., Ltd., Novartis Pharma K.K., Sumitomo Dainippon Pharma Co., Ltd., Kyowa Kirin Co., Ltd., Pfizer Japan Inc., Novo Nordisk Pharma Ltd., Nippon Boehringer Ingelheim Co., Ltd., Eli Lilly Japan K.K., Taisho Pharmaceutical Co., Ltd., Janssen Pharmaceutical K.K., and received clinical research funding from Taisho Pharmaceutical Co., Ltd. KY has also received scholarship grants from Mitsubishi Tanabe Pharma Corp., Takeda Pharmaceutical Co., Ltd., MSD K.K., Pfizer Japan Inc., Novo Nordisk Pharma Ltd., Taisho Pharmaceutical Co., Ltd., Kao Corporation, Ono Pharmaceutical Co., Ltd., Eli Lilly Japan K.K., Sumitomo Dainippon Pharma Co., Ltd., Nippon Boehringer Ingelheim Co., Ltd., Daiichi-Sankyo Co., Ltd., Teijin Pharma, Ltd., Shionogi Co., Ltd., Bayer Yakuhin, Ltd. Jun Wada has nothing to disclose.

References

- 1) Assmann G, Eckardstein A, Brewer HB. Familial anaphalipoproteinemia: Tangier disease. *The Metabolic and Molecular Bases of Inherited Diseases* (McGraw Hill), 2001; 8th ed. vol 2: 2937-2960
- 2) Fredrickson DS, Altrocchi PH, Avioli LV, Goodman DS, Goodman HC. Tangier disease—Combined clinical staff conference at the National Institutes of Health. *Ann Intern Med*, 1961; 55: 1016-1031
- 3) Hara H, Yokoyama S. Interaction of free apolipoproteins with macrophages. Formation of high-density lipoprotein-like lipoproteins and reduction of cellular cholesterol. *J Biol Chem*, 1991; 266: 3080-3086
- 4) Francis GA, Knopp RH, Oram JF. Defective removal of cellular cholesterol and phospholipids by apolipoprotein A-I in Tangier disease. *J Clin Invest*, 1995; 96: 78-87
- 5) Brooks-Wilson A, Marcil M, Clee SM, Zhang LH, Roomp K, van Dam M, Yu L, Brewer C, Collins JA, Molhuizen HO, Loubser O, Ouellette BF, Fichter K, Ashbourne-Excoffon KJ, Sensen CW, Scherer S, Mott S, Denis M, Martindale D, Frohlich J, Morgan K, Koop B, Pimstone S, Kastelein JJ, Genest J Jr, Hayden MR. Mutations in ABC1 in Tangier disease and familial high-density lipoprotein deficiency. *Nat Genet*, 1999; 22: 336-345
- 6) Bodzioch M, Orsó E, Klucken J, Langmann T, Böttcher A, Diederich W, Drobnik W, Barlage S, Büchler C, Porsch-Ozcürümez M, Kaminski WE, Hahmann HW, Oette K, Rothe G, Aslanidis C, Lackner KJ, Schmitz G. The gene encoding ATP-binding cassette transporter 1 is mutated in Tangier disease. *Nat Genet*, 1999; 22: 347-351
- 7) Rust S, Rosier M, Funke H, Real J, Amoura Z, Piette JC, Deleuze JF, Brewer HB, Duverger N, Denèfle P, Assmann

- G. Tangier disease is caused by mutations in the gene encoding ATP-binding cassette transporter 1. *Nat Genet*, 1999; 22: 352-355
- 8) Muratsu J, Koseki M, Masuda D, Yasuga Y, Tomoyama S, Ataka K, Yagi Y, Nakagawa A, Hamada H, Fujita S, Hattori H, Ohama T, Nishida M, Hiraoka H, Matsuzawa Y, Yamashita S. Accelerated atherogenicity in Tangier disease. *J Atheroscler Thromb*, 2018; 25: 1076-1085
 - 9) Maruyama T, Yamashita S, Matsuzawa Y, Bujo H, Takahashi K, Saito Y, Ishibashi S, Ohashi K, Shionoiri F, Gotoda T, Yamada N, Kita T; Research Committee on Primary Hyperlipidemia of the Ministry of Health and Welfare of Japan. Mutations in Japanese subjects with primary hyperlipidemia; Results from the Research Committee of the Ministry of Health and Welfare of Japan since 1996. *J Atheroscler Thromb*, 2004; 11: 131-145
 - 10) Hooper AJ, McCormick S, Hegele RA, Burnett JR. Clinical utility gene card for: Tangier disease. *Eur J Hum Genet*, 2017; 25: e1-e3
 - 11) Costet P, Luo Y, Wang N, Tall AR. Sterol-dependent transactivation of the ABC1 promoter by the liver X receptor/retinoid X receptor. *J Biol Chem*, 2000; 275: 28240-28245
 - 12) Tamehiro N, Shigemoto-Mogami Y, Kakeya T, Okuhira K, Suzuki K, Sato R, Nagao T, Nishimaki-Mogami T. Sterol regulatory element-binding protein-2- and liver X receptor-driven dual promoter regulation of hepatic ABC transporter A1 gene expression: mechanism underlying the unique response to cellular cholesterol status. *J Biol Chem*, 2007; 282: 21090-21099
 - 13) Ohoka N, Okuhira K, Cui H, Wu W, Sato R, Naito M, Nishimaki-Mogami T. HNF4 α increases liver-specific human ATP-binding cassette transporter A1 expression and cholesterol efflux to apolipoprotein A-I in response to cholesterol depletion. *Arterioscler Thromb Vasc Biol*, 2012; 32: 1005-1014
 - 14) Ito J, Nagayasu Y, Yokoyama S. Cholesterol-sphingomyelin interaction in membrane and apolipoprotein-mediated cellular cholesterol efflux. *J Lipid Res*, 2000; 41: 894-904
 - 15) Yamauchi Y, Iwamoto N, Rogers MA, Abe-Dohmae S, Fujimoto T, Chang CC, Ishigami M, Kishimoto T, Kobayashi T, Ueda K, Furukawa K, Chang TY, Yokoyama S. Deficiency in the lipid exporter ABCA1 impairs retrograde sterol movement and disrupts sterol sensing at the endoplasmic reticulum. *J Biol Chem*, 2015; 290: 23464-23477
 - 16) Koseki M, Hirano K, Masuda D, Ikegami C, Tanaka M, Ota A, Sandoval JC, Nakagawa-Toyama Y, Sato SB, Kobayashi T, Shimada Y, Ohno-Iwashita Y, Matsuura F, Shimomura I, Yamashita S. Increased lipid rafts and accelerated lipopolysaccharide-induced tumor necrosis factor- α secretion in Abca1-deficient macrophages. *J Lipid Res*, 2007; 48: 299-306
 - 17) Koseki M, Matsuyama A, Nakatani K, Inagaki M, Nakaoka H, Kawase R, Yuasa-Kawase M, Tsubakio-Yamamoto K, Masuda D, Sandoval JC, Ohama T, Nakagawa-Toyama Y, Matsuura F, Nishida M, Ishigami M, Hirano K, Sakane N, Kumon Y, Suehiro T, Nakamura T, Shimomura I, Yamashita S. Impaired insulin secretion in four Tangier disease patients with ABCA1 mutations. *J Atheroscler Thromb*, 2009; 16: 292-296
 - 18) Brunham LR, Kruit JK, Pape TD, Timmins JM, Reuwer AQ, Vasanji Z, Marsh BJ, Rodrigues B, Johnson JD, Parks JS, Verchere CB, Hayden MR. Beta-cell ABCA1 influences insulin secretion, glucose homeostasis and response to thiazolidinedione treatment. *Nat Med*, 2007; 13: 340-347
 - 19) Schaefer EJ, Blum CB, Levy RI, Jenkins LL, Alaupovic P, Foster DM, Brewer HB Jr. Metabolism of high-density lipoprotein apolipoproteins in Tangier disease. *N Engl J Med*, 1978; 299: 905-910
 - 20) Bi X, Pashos EE, Cuchel M, Lyssenko NN, Hernandez M, Picataggi A, McParland J, Yang W, Liu Y, Yan R, Yu C, DerOhannessian SL, Phillips MC, Morrissey EE, Duncan SA, Rader DJ. ATP-binding cassette transporter A1 deficiency in human induced pluripotent stem cell-derived hepatocytes abrogates HDL biogenesis and enhances triglyceride secretion. *EBioMedicine*, 2017; 18: 139-145
 - 21) Serfaty-Lacrosniere C, Civeira F, Lanzberg A, Isaia P, Berg J, Janus ED, Smith MP Jr, Pritchard PH, Frohlich J, Lees RS, Barnard FF, Ordovas JM, Schaefer EJ. Homozygous Tangier disease and cardiovascular disease. *Atherosclerosis*, 1994; 107: 85-98
 - 22) Puntoni M, Sbrana F, Bigazzi F, Sampietro T. Tangier disease: epidemiology, pathophysiology, and management. *Am J Cardiovasc Drugs*, 2012; 12: 303-311
 - 23) Zyss J, Béhin A, Couvert P, Bouhour F, Sassolas A, Kolev I, Denys V, Vial C, Lacour A, Carrié A, Stojkovic T. Clinical and electrophysiological characteristics of neuropathy associated with Tangier disease. *J Neurol*, 2012; 259, 1222-1226
 - 24) Amanda AJ, Hegele RA, Burnett JR. Tangier disease: update for 2020. *Curr Opin Lipidol*, 2020; 31: 80-84
 - 25) Komuro R, Yamashita S, Sumitsuji S, Hirano K, Maruyama T, Nishida M, Matsuura F, Matsuyama A, Sugimoto T, Ouchi N, Sakai N, Nakamura T, Funahashi T, Matsuzawa Y. Tangier disease with continuous massive and longitudinal diffuse calcification in the coronary arteries: demonstration by the sagittal images of intravascular ultrasonography. *Circulation*, 2000; 101: 2446-2448
 - 26) Zanon P, von Eckardstein A. Inborn errors of apolipoprotein A-I metabolism: implications for disease, research and development. *Curr Opin Lipidol*, 2020; 31: 62-70
 - 27) Kinoshita M, Yokote K, Arai H, Iida M, Ishigaki Y, Ishibashi S, Umemoto S, Egusa G, Ohmura H, Okamura T, Kihara S, Koba S, Saito I, Shoji T, Daida H, Tsukamoto K, Deguchi J, Dohi S, Dobashi K, Hamaguchi H, Hara M, Hiro T, Biro S, Fujioka Y, Maruyama C, Miyamoto Y, Murakami Y, Yokode M, Yoshida H, Rakugi H, Wakatsuki A, Yamashita S; Committee for Epidemiology and Clinical Management of Atherosclerosis. Japan Atherosclerosis Society (JAS) Guidelines for Prevention of Atherosclerotic Cardiovascular Diseases 2017. *J Atheroscler Thromb*, 2018; 25: 846-984