



Review

Neurological and Inflammatory Manifestations in Sjögren's Syndrome: The Role of the Kynurenine Metabolic Pathway

Fabiola Reis de Oliveira ¹, Marina Zilio Fantucci ¹, Leidiane Adriano ¹, Valéria Valim ²,
Thiago Mattar Cunha ¹, Paulo Louzada-Junior ¹ and Eduardo Melani Rocha ^{1,*} 

¹ Ribeirao Preto Medical School, Ribeirao Preto, University of Sao Paulo, Ribeirao Preto, SP 14049-900, Brazil; fabiolabi@hotmail.com (F.R.d.O.); marinazf@fmrp.usp.br (M.Z.F.); leidiadriano@hotmail.com (L.A.); thicunha@fmrp.usp.br (T.M.C.); plouzada@fmrp.usp.br (P.L.-J.)

² Espírito Santo Federal University, Vitoria, ES 29075-910 Brazil; val.valim@gmail.com

* Correspondence: emrocha@fmrp.usp.br; Tel./Fax: +55-16-3602-0593

Received: 20 November 2018; Accepted: 6 December 2018; Published: 8 December 2018



Abstract: For decades, neurological, psychological, and cognitive alterations, as well as other glandular manifestations (EGM), have been described and are being considered to be part of Sjögren's syndrome (SS). Dry eye and dry mouth are major findings in SS. The lacrimal glands (LG), ocular surface (OS), and salivary glands (SG) are linked to the central nervous system (CNS) at the brainstem and hippocampus. Once compromised, these CNS sites may be responsible for autonomic and functional disturbances that are related to major and EGM in SS. Recent studies have confirmed that the kynurenine metabolic pathway (KP) can be stimulated by interferon- γ (IFN- γ) and other cytokines, activating indoleamine 2,3-dioxygenase (IDO) in SS. This pathway interferes with serotonergic and glutamatergic neurotransmission, mostly in the hippocampus and other structures of the CNS. Therefore, it is plausible that KP induces neurological manifestations and contributes to the discrepancy between symptoms and signs, including manifestations of hyperalgesia and depression in SS patients with weaker signs of sicca, for example. Observations from clinical studies in acquired immune deficiency syndrome (AIDS), graft-versus-host disease, and lupus, as well as from experimental studies, support this hypothesis. However, the obtained results for SS are controversial, as discussed in this study. Therapeutic strategies have been reexamined and new options designed and tested to regulate the KP. In the future, the confirmation and application of this concept may help to elucidate the mosaic of SS manifestations.

Keywords: IDO; kynurenine; pain; Sjögren's syndrome; tryptophan

1. Introduction

Sjögren's syndrome (SS) is defined as an exocrinopathy of the salivary and lacrimal glands (SG and LG) mediated by autoimmune mechanisms that could manifest neurological dysfunctions, and those neurological dysfunctions may take part in the physiopathology of the disease [1–5]. However, the extraglandular manifestations (EGM) of neurological disorders are not considered in the definition or the diagnosis of SS, despite their presence during the disease progress evaluation and reported more frequent association with SS in recent years [6–9]. Of interest, 60–80% of patients develop neurological manifestations before or at SS diagnosis (early systemic presentation), indicating that neurological damage is precocious and it could play a role in the disease mechanism [10] (Figure 1).

The kynurenine metabolic pathway (KP) is the main pathway that is involved in the catabolism of tryptophan. There is evidence that KP participates in the inflammatory mechanisms of the neurogenic

manifestations of autoimmune diseases through the action of indoleamine 2,3-dioxygenase (IDO), the rate-limiting enzyme in tryptophan degradation [11–15].

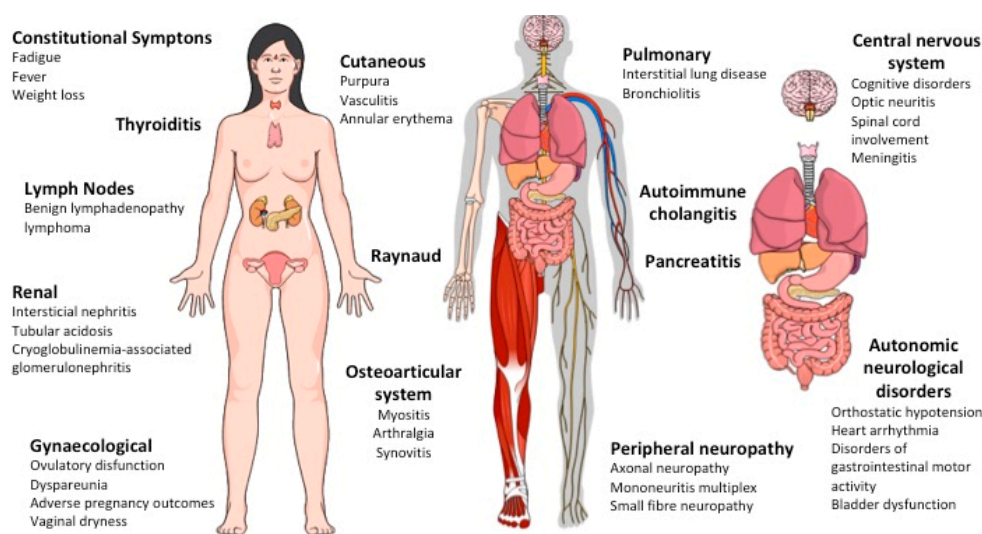


Figure 1. Extraglandular manifestations of Sjögren’s Syndrome. The pleiotropic features of systemic multiorgan involvement in SS are not well-understood. Some of the extraglandular manifestations may arise from immune-complex deposition in the context of cryoglobulinemia. Other symptoms and signs are related to lymphoproliferation, dendritic cell activation, and cytokine maintenance of the inflammatory process. Interferon- γ -inducible-Kynurenine Pathway could play a role in the neural manifestations, fatigue and chronic pain [16]. Figures were obtained from the free version of myndtheagraph.com.

This review summarizes the actual status of knowledge concerning the neurological manifestations in SS and it presents the hypothesis of the association between these neurological changes and the KP and their interactions to understand the unknown and paradoxical signs and symptoms of SS (Box 1).

Box 1. Summary of evidence linking Sjögren’s syndrome (SS) and the tryptophan/kynurenine signaling pathway (KP) in the central nervous system (CNS).

- Association among chronic inflammation, pain and neuropathic disorders in SS [16]
- Dryness and Indoleamine 2,3-dioxygenase (IDO) activity triggered by interferon [17,18]
- Clinical findings and inflammation modulated by sex hormones in SS [19]
- Tryptophan deprivation induces dry eye [20]
- Sjögren’s syndrome and salivary gland inflammation leads to increased expression of kynurenine, a metabolite of IDO [18,21]

2. Autoimmunity, Neuropathy and Chronic Pain

The constant basal and reflex wetting of the mouth and the ocular surface provided, respectively, by saliva and by tears are directly controlled by the autonomic nervous system [22–24]. The volume and content of fluids from several exocrine glands that are present in both locations (mouth and eye) are responsive to sensorial stimuli from the environment that is driven by sensitive nerves specialized in taste and vision but also general sense nerves related to touch, thermal and chemical changes [25–27]. Once detected by the brainstem, the feedback mechanisms are conducted in the parasympathetic and sympathetic systems, stimulating synapsis in α -adrenergic and muscarinic cholinergic receptors in the

many salivary and lacrimal gland subtypes distributed in the mouth and in the ocular surface [25,28]. This sensorial/autonomic feedback system that regulates the tear secretion in the ocular surface is called the lacrimal functional unit (LFU) (and a very similar system works in the mouth), whereas inflammation, trauma, or other damage in a segment of this integrative system can disrupt persistent dysfunction of secretion with variable sensorial manifestations, depending on the integrity of the sensorial loop [28,29].

Autoimmunity is linked to chronic neuropathy in at least three different domains in SS. First, the major clinical EGM observed in SS are fatigue and pain, with different manifestations, such as allodynia, dysesthesia, hypo or hyperesthesia, and hyperalgesia [9,30,31]. Second, chronic inflammation in the target organs (e.g., the ocular surface) generates a noxious stimulus and depression that may persist in a further phase in which the inflammatory process is already resolved [32,33]. Third, the central nervous system (CNS), mostly autonomic nervous system dysfunction, can induce or perpetuate an unbalanced inflammatory response in the target-innervated organs of autoimmune diseases [3,34]. Therefore, an unified theory of the relationship between the neural and immune mechanisms of SS manifestations take in account that lesions of the autonomic and peripheral neural system reduce the threshold for inflammatory and noxious events in their connected organs and disturb the balance between pro and anti-inflammatory mediators [35–37] (Figure 2).

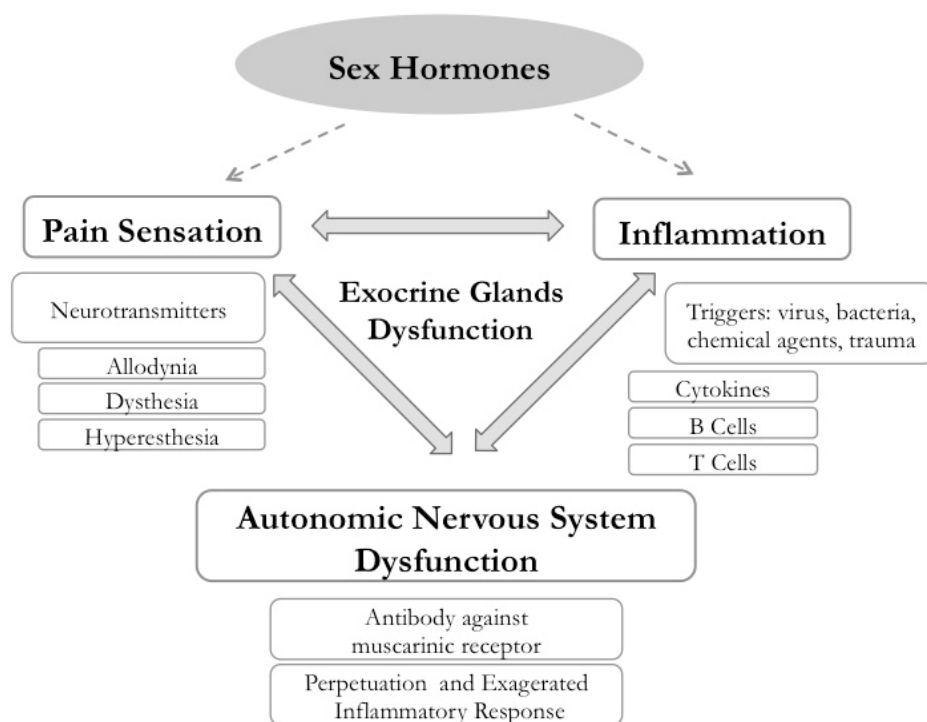


Figure 2. Schematic model showing the interrelationships among autonomic nervous system dysfunction, pathological pain and chronic inflammation in Sjögren's syndrome (SS) [19,29]. The dotted arrows indicate the effects of hormones with all their cyclic, indirect and also circadian influences. The gray two-way arrows indicate the interdependence of the three phenomena highlighted in the boxes: pain sensation, inflammation and autonomic nervous system dysfunction.

The chronic pain and persistent inflammation that are mediated by humoral factors and neurotransmitters associate autoimmunity and neuropathy in several target organs, including the hippocampus and lacrimal functional unit (LFU) in SS [27,34,38–41]. The dissociation between signs and symptoms in dry eye and dry mouth disease, including SS, delays and makes the diagnosis more difficult [5,42–48]. The lower intensity of signs that are associated with depression and diffusely

spread painful symptoms in the body suggest a mechanism involving integrative systems, such as the autonomic nervous system, immune, and endocrine systems (Figure 1) [42,45,49].

The pain, depression, and other neurological manifestations of SS can be associated with IFN- γ inducible KP activity. Therefore, our hypothesis considers three aspects: (a) the KP in the mechanism of SS neuropathy; (b) the possible roles of the KP in cross-talking pathways among the immune, endocrine, and neural systems to produce the SS signs and symptoms; and, (c) the possible role of the KP in the previously reported dissociation between those signs and symptoms in SS [50–55]. Therefore, IFN- γ -inducible KP could be the missing link between disease activity and neural manifestations in SS [56] (Figure 2).

In summary, this hypothesis implicates the KP in the spectrum of manifestations of SS, with two different poles of SS disease. At one pole, the major characteristic is chronic noninflammatory pain with neuropathic features. At the opposite pole is SS present with inflammation, including EGM inflammatory activity. In other words, we hypothesized that SS patients with Interferon- γ (IFN- γ)-inducible KP activation could develop chronic pain, depression, and low EGM disease activity, because the KP promotes an immunosuppressive and neurosensitive effect. However, these distinguished profiles of SS are more clearly documented in long-term cohort studies, as recently published [16].

2.1. SS and Neurological Manifestations

Neurological signs and symptoms have been described in SS. The prevalence of peripheral neuropathy ranges from 8–49%, depending on the selection bias related to different classification criteria, and whether neurological manifestations were diagnosed based on clinical marked symptoms versus asymptomatic, or detected by electrophysiological studies [2,57–61]. Distal sensory and sensorimotor neuropathies are the most common manifestations of peripheral nerve disease in primary SS (pSS). Sensory neuropathies include painful nonataxic sensory polyneuropathy, small fiber neuropathy, dorsal root ganglionitis, and trigeminal neuropathy. Other forms have also been described, including multiplex mononeuritis, acute or chronic inflammatory demyelinating polyradiculoneuropathy (CIDP), multiple cranial neuropathy, especially III, V, VI, VII, IX, X, and XII, and dysautonomia, which is very common in SS and can reach a frequency of 40% of SS patients, with or without different neurological manifestations [62–65]. Moreover, CNS manifestations of SS may be associated with focal syndromes (such as multiple-sclerosis-like epilepsy, movement disorders, neuromyelitis optica, and pseudotumor) and diffuse syndromes (encephalitis, meningitis, cognitive dysfunction, psychiatric disorders). Acute or chronic myelopathy and inferior motor neuron disease may also occur [57,58,62,66–89]. This high variability of EGM and responses to treatment in SS are not completely understood. The spectrum of the disease may be caused by environmental agents that lead to epigenetic phenomena working on diverse and complex mechanisms of organ injuries, such as vasculitis and lymphocytic infiltration [90–92]. In addition, the changes in SS patients in the CNS and PNS (such as in the dorsal spinal cord and the dorsal ganglion roots), including autonomic dysfunction, diffuse decreases in white matter, and loss of gray matter in the hippocampal area, may influence peripheral organ function and induce neurological symptoms [93–95].

2.2. SS and the Mechanisms of Neurological Manifestations

The mechanisms triggering the neurological manifestations in SS are unclear. They involve genetic predisposition, environmental agents, trauma and posttraumatic stress, autoimmunity against the CNS, and peripheral nervous systems (PNS), in addition to neuroimmunendocrine network disruption [60,90,96–100]. The events are attributed to DNA demethylation, microRNA abnormal expression, an imbalance of interferon I (α and β) and II (γ), and anti-neuron autoantibody production [68,90,101–103]. In SS, the 2-5 oligo-adenylate synthetase 1 (OAS1) gene defect leads to a reduced responsiveness to IFN- γ and higher production of IFN- γ , causing severe complications, such as lymphoma, neuropathy, and debilitating fatigue [100].

The sensorial, autonomic, cognitive neurological, or behavioral manifestations of SS are associated with structural changes, such as increases in corneal nerve thickness and a larger number of antigen-presenting cells in the cornea, nerve vasculitis, nonvascular encephalitis, neuromyelitis, the CNS, and axonal degeneration [38,60,97,104].

It is interesting to note that structures that are altered in the CNS, PNS, exocrine glands, and cornea (based on image exams) of SS patients are also sensitive to changes in the KP caused by trauma or ischemic damage (Table 1). These observations suggest that common inflammatory events are shared in autoimmune and non-immune inflammatory diseases, such as the synapses of the sensorial fibers, in the dorsal ganglion root, hippocampus, thalamus, and LFU [2,12,13,20,29,40,53,93,105–108] (Table 1). Those locations must be investigated in future studies addressing the anatomic correlations of clinical and biochemical changes in SS.

Table 1. Common locations of changes in the nervous system and in the exocrine glands caused by experimental traumatic modulation of the kynurenine metabolic pathway (KP) and Sjögren's Syndrome.

Structure	Kynurenine Metabolic Pathway	Sjögren's Syndrome
Dorsal ganglion root	Sciatic injury increases kynurenine monoxygenase (KMO) in the dorsal root ganglion and spinal cord of rats [105]	Dorsal root ganglion alterations in MRI, associated with increased intradermal nerve fiber density on skin biopsy [109]
Hippocampus	IDO and kynurenine-3-hydroxylase increase in the hippocampus after day 2 after CNS ischemia [110]	Hippocampal atrophy in SS patients [40]
Exocrine Glands and LFU	Increase in kynurenine in salivary gland after ductal ligation, LG atrophy due to tryptophan deprivation [20,21]	Changes in LG and SG in the MRI, nerve changes in the cornea of SS patients [107,111]

MRI: magnetic resonance image, SS: Sjögren's syndrome, LG: lacrimal gland, SG: salivary gland, CNS: central nervous system, IDO: indoleamine 2,3-dioxygenase.

2.3. Immune and Endocrine Modulation of Neurological Findings in SS

The concept that is implicit in the neuroimmunendocrine network predicts that cellular and molecular communication among those three systems are responsible for the homeostasis of the body organs, and a disruption in this network plays a role in the disease mechanism [112–115]. For example, acetyl-choline (Ach), dopamine, glutamatergic, and other neurotransmitters are secreted not only by neurons, but also by lymphocytes [116]. In contrast, the autonomic nervous system is capable of modulating lymphocyte proliferation in target organs, such as spleen, liver, kidney, and brain [37,117,118]. In addition, the sensory neurons are able to secrete peptides with immunomodulatory properties, such as galanin, netrin-1, and somatostatin and promote or attenuate inflammatory responses [119,120].

Hormones, in particular, sex hormones, can modulate inflammation and pain, sensitizing the ionic receptors expressed in neurons and epithelial cells, called the transient receptor of potential (TRP), and stimulating growth factor and cytokine expression in target tissues, such as the lacrimal and salivary gland (LG and SG), hippocampus, and trigeminal ganglion of the CNS, as well as other target tissues [19,121–127]. These mechanisms explain the sex hormone-mediated amplification and perpetuation of the inflammatory process, pain hypersensitivity, and exocrine gland dryness manifestations in SS, where estrogen potentiates the pain and pro-inflammatory mediators and androgens work in an opposite manner [19,96,128–130]. Although the trigger of the first event in SS and the steps leading to the chronic phase are poorly understood, there is a strong clinical association between female sex hormones and the inflammatory mediators that are involved in innate as well as adaptive immunity [4,19,57,124,131,132] (Figure 1).

Therefore, hormones and neural pathways interact with immune responses, acting on pain sensation, the inflammatory reaction, tissue integrity, and organ functional disruption [5,37]. These interactions certainly take part in the EGM and comorbidities in SS patients [2,106,133].

A loss or damage of nerve fibers, dorsal root ganglionitis, nerve vasculitis, and reduction of the CNS matter can be observed in imaging exams, such as magnetic resonance images (MRIs) or skin biopsies of SS patients; they are implicated in the underlying mechanisms and they could potentially function as diagnostic markers of neurological manifestations of SS [38,93,94,109]. For example, cognitive impairment in SS patients is associated with antibodies against the subtype 2 of the N-methyl aspartate receptor (NR2) (anti-NR2 antibodies) in the cerebrospinal fluid (CSF) mediating hippocampal gray matter atrophy, as observed by MRI [40].

In summary, the SS inflammatory activity in the PNS and CNS causes the above-described signs and symptoms, and the physiopathology implicates the hormones, neurotransmitters, and cytokines that are susceptible to the KP interference, as previously described [13,14,114]. The role of kynurenine and its metabolites in the neurological activities are detailed below.

3. Kynurenine Pathway (KP)

The metabolism of L-tryptophan (LTF) leads to the generation of several neuroactive compounds via the serotonergic and kynurenine pathways. In the serotonergic pathway, L-tryptophan is metabolized to serotonin (5-hydroxytryptamine or 5-HT), and, in some cells, to melatonin. Serotonin acts as a neurotransmitter with a variety of functions in behavior and psychiatric symptoms (depression and anxiety), platelet aggregation, gastrointestinal tract control (satiety, secretion, and peristalsis), and tumor resistance. Melatonin acts as a neurohormone in the circadian rhythm, inducing sleep, and it also has anti-inflammatory, anti-angiogenic, and anti-tumoral immunomodulatory effects [134] (Figure 3).

In the catabolic KP, tryptophan 2,3-dioxygenase (TDO) metabolizes tryptophan in the liver and responds to hormonal inputs, such as cortisol and glucagon, and to the tryptophan load in meals [135] (Figure 3).

The IDO enzyme regulates both innate and adaptive immune responses through degradation of the essential amino acid tryptophan into kynurenine and other metabolites, which suppress the effector T-cell function and promote the differentiation of regulatory T cells [136]. IDO also metabolizes serotonin, melatonin previously generated in the serotonergic pathway, and tryptophan to N-formyl kynurenine, and kynurenine, mostly in the lung, brain, and blood, producing quinolinic acid and nicotinamide adenine nucleotide (NAD⁺) [136–138]. The KP is responsible for 95–99% of tryptophan catabolism [139] (Figure 3).

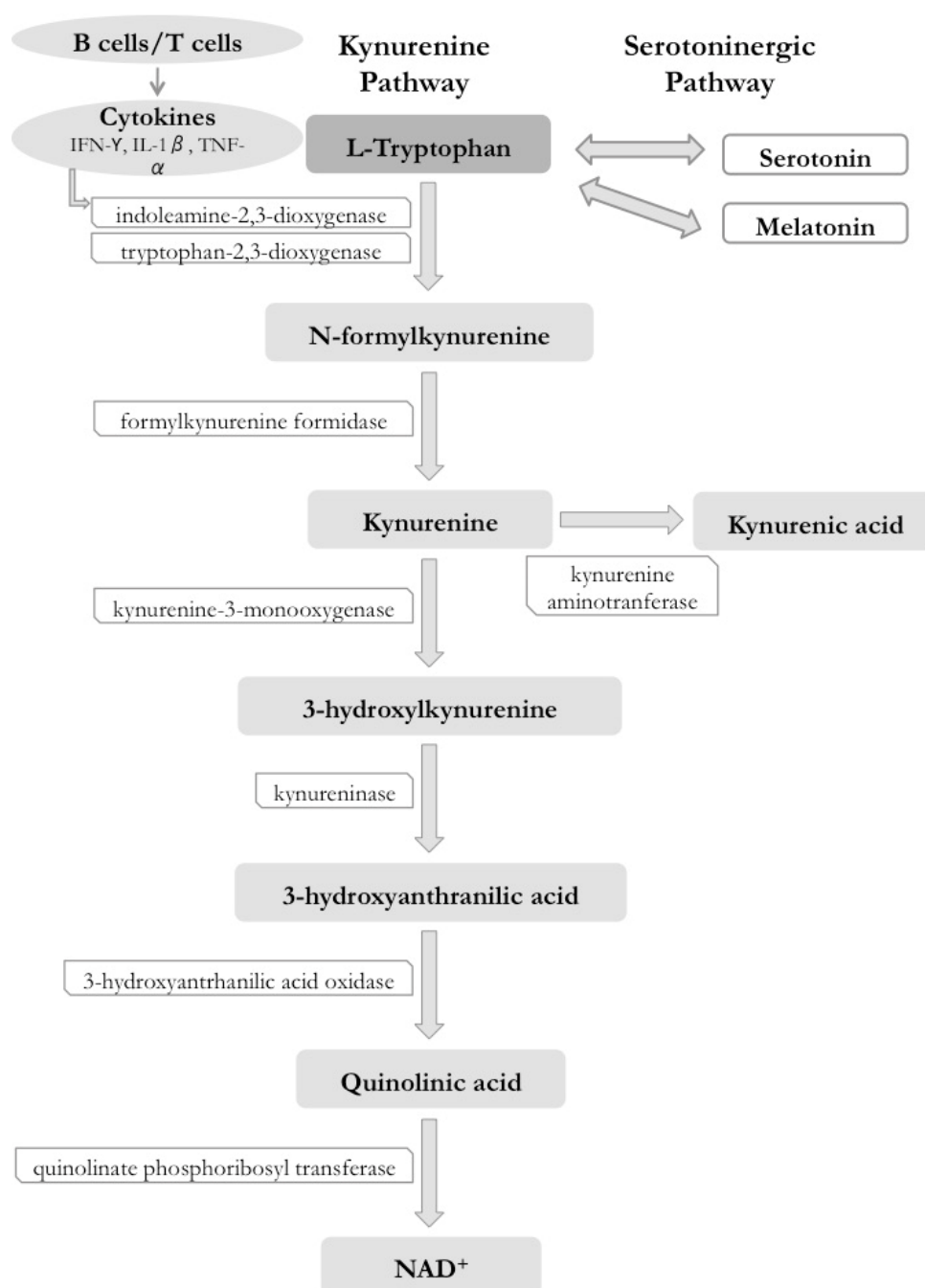


Figure 3. Kynurenine pathway (KP) and metabolites. Several of the metabolites have neuromodulatory effects [14,140–142]. The fine gray arrows indicate chemotactic and enzymatic effects, the thick gray arrows indicate the subsequent metabolite in the cascade, whereas the two-way gray arrows indicate the potential conversion of one molecule in the other, depending on demand and the substrate availability. IL-1 β is Interleukin-1beta and TNF- α is Tumor Necrosis Factor-alpha.

IDO is found mainly in immune cells, and it has enzymatic activity in the cytoplasm and transcriptional activity in the nucleus, playing a unique role as a signaling molecule modulating immune responses [143–145]. The nuclear effect contributes to enzymatic “self-amplification” in an IFN-dependent loop that may account for the tolerance phenotype, attenuating or preventing immune reactions and mediating persistent pain in several conditions, including SS [18,143,144,146,147]. IDO activity is induced in macrophages by cytokines, such as IFN- γ and TNF- α and by prostaglandins. During viral and bacterial infections, lipopolysaccharides can trigger IDO activity in dendritic cells

and enhance the subproducts of kynurenine [148–152]. IDO overexpression has been documented in patients with systemic lupus erythematosus (SLE) and SS, as well as in sepsis [15,50,153]. In patients who are positive for the IFN gene expression signature, Treg cell levels are elevated in combination with increased IDO activity, with tolerance and immune modulation [18]. These regulatory T lymphocytes represent a diverse subclass of T cells, a protagonist in the maintenance of self-tolerance and immune modulation [154].

The KP is catalyzed by the IDO enzyme and it can be induced by cytokine IFN- γ , as observed in studies in mouse, in which IFN- γ or IFN- γ receptor knockout prevents kynurenine production [155]. This phenomenon is dependent on the abovementioned antigen-presenting cells (APCs), as observed in mouse models of graft-versus host disease (GVHD), and the induction of this KP can extend the life-span and reduce inflammation in the gut of wild-type mice, but not IFN- γ receptor knockout, mice [155].

The balance between Interferon-alpha and beta at one side and gamma at the other (INF- α and β , and IFN- γ) activation in inflammatory processes can modulate the activation of KP and the resulting intensity of this inflammatory process, not just in SS but also in response to other stimuli, such as exogenous challenges [156–158]. IFN- α and β (type I INF) are associated with inflammatory activity (including higher levels of autoantibodies and inflammatory cell activity), and IFN- γ (type II INF) is associated with KP activation and the attenuation of inflammatory activity [91,159–161]. SS patients have higher levels of IFN- γ and INF- α and β mRNA in the peripheral blood and labial salivary glands in comparison to healthy individuals, and SS patients with lymphoma present lower levels of IFN- α and higher levels of INF- γ mRNA in labial salivary gland samples when compared with other SS and control individuals, supporting INF- α/γ as a predictor of lymphoma in SS [156]. Therefore, type I INF seems to have an antagonistic effect to type II INF, but an unclear, though potentially useful, combinatorial effect in SS associated with lymphoma.

The possibility that some cases of SS are disrupted by viral infection also raises the possibility of a subtype of SS that is triggered by viral disease and distinguished KP activity, with a particular set of signs and symptoms. In support of this possibility, 50% of pSS patients have been confirmed to have hepatitis delta virus, without hepatitis B virus, in the salivary gland in a recent report, and the affected patients presented elevated SG inflammation and autoantibody positivity [162].

The isoform IDO2 as well as IDO1 are enzymes that are involved in the catabolism of the amino acid tryptophan and operate in similar manners in immunomodulation, with variations in the target tissues and cells involved in the metabolic pathways. However, further studies are necessary to clarify their diversity in diseases, including SS [163,164].

4. KP and Neurological Manifestations

The relationship between tryptophan, serotonin, and depression has an extended history in psychiatry. The development of depressive symptoms is correlated with high levels of tryptophan metabolites in urine [51] and with a decrease in tryptophan in the blood and cerebrospinal fluid [165,166]. Tryptophan transport across the blood-brain barrier, specific inflammation, and damage that are caused by brain-reactive autoantibodies and immune complexes play a critical role in the regulation of tryptophan metabolism in the brain [134]. There is substantial evidence to suggest that, in addition to serotonergic neurons, other cells, such as astrocytes, dendritic cells, microglia, and macrophages also synthesize multiple neuroactive metabolites via the enzyme IDO and KP in the CNS [167]. Evidence for the underlying mechanisms have been obtained from clinical studies examining the effects of IFN- α on the mood of cancer and hepatitis C virus-infected patients. In both cases, the development of depressive symptoms was associated with decreased circulating tryptophan levels and the enhanced formation of kynurenine, indicating activity of the IDO pathway [168–170].

Tryptophan metabolites that are generated in the KP have been associated with neurodegenerative diseases, such as acquired immunodeficiency syndrome (AIDS)-related dementia, Alzheimer's and Huntington's diseases, and neuropsychiatric diseases, such as bipolar disorder and

schizophrenia [171,172]. The synthesis of kynurenine in the CNS is affected by dietary intake of tryptophan and by the gut microbiota [14,173]. Tryptophan deprivation induces depression and cognitive dysfunction (attention, memory, and execution), among other neurological dysfunctions, through the glutamatergic receptors [14,54,173–176].

Tryptophan metabolism induces a dual effect in astrocytes and microglial cells [177]. In astrocytes, it leads to the production of kynurenic acid, which has been reported to participate in neuroprotective effects. In contrast, microglial cells give rise to metabolites with reactive oxidative properties, including hydroxykynurenine and 3-hydroxyanthranilic acid and quinolinic acid, which also functions as an agonist of the glutamate N-methyl-D-aspartate (NMDA) receptor subtype and may contribute to excitotoxicity and neurotoxicity [177–179].

In healthy subjects, a well-adjusted system is established in the KP via the action of kynurenines aminotransferases (KATs) on kynurenic acid or quinolinic acid by (KMO) [141]. Kynurenic acid has been reported to promote neuroprotective and immunosuppressive actions in the CNS and plays a role as an NMDA antagonist, blocking this glutamatergic receptor. Enhanced IDO activity and a deviation to KMO downstream are observed by stimulation of cytokines, such as IFN- γ [141]. Additionally, SS1 patients with positive levels of IFN- γ in the blood have higher levels of CD25 FoxP3+ Treg cells, which correlate with higher plasma IDO activity, as measured by the tryptophan/kynurenine levels, in comparison to the controls [18]. KMO has a clear inflammatory and pro-apoptotic action. Quinolinic acid, its intermediate compound, acts as an agonist at the NMDA receptor, modulating excitatory amino acid transmission, and it may serve as a neurotoxic agent implicated in the pathogenesis of several neurological diseases [12,134,171,179]. Moreover, in SLE, the presence of the antibody against the NMDA receptor has several demonstrated pathological activities in the kidney and brain, particularly in the hippocampus, including neuron death, as observed in humans and in mouse models [180,181]. Likewise, 3-hydroxykynurenine also has a neurotoxic effect, which is probably associated with the conversion of reactive oxygen species and apoptosis [94,140,182–184].

The NR2 subtype NMDA receptor is ubiquitously distributed throughout the brain, with an unusually high density in the hippocampus [185]. The hippocampus is a brain structure that is linked to the autonomous nervous system, with critical importance for memory formation and learning, and it is also affected in mood disorders and in SS [93,94,186,187]. Likewise, the N-methyl aspartate receptor 2B (NR2B) subunit of the NMDA receptor is widespread in the dorsal root ganglion and it may mediate peripheral sensitization and visceral pain [188]. Those receptors are critically involved in the initiation and maintenance of neuronal hyperexcitability after noxious events and by C-fiber neuron stimulation, which consist of unmyelinated sensorial neurons [189].

In a rodent model of peripheral nerve injury subjected to tibial and peroneal nerve sectioning, leaving the sural branch nerve intact, with the aim to investigate the KP, the following changes were documented. After seven days, IDO1 was activated and kynurenine rose in the bloodstream, accompanied by depressive behavior (measured by an extended time of immobility in the forced swim test) and allodynia (tested by paw withdrawal in response to mechanical stimulation with von Frey hair). These findings were followed by an increase in the levels of KMO, quinolinic acid, and a reduction of kynurenic acid in the contralateral hippocampus [53]. These observations identify hippocampal neurons as the CNS site that is responsible for the perpetuation of pain and depressive symptoms. Injection of interleukin-1 β (IL-1 β) receptor antagonist in the CNS ventricular space reduced the depressive behavior and KMO mRNA levels, but did not change the allodynia, revealing the role of IL-1 β in depression but not in the pain mechanism [53].

Depression has also been associated with decreased levels of tryptophan in patients with cancer receiving IL-2 and IFN- α therapy, suggesting that these cytokines impact the levels of serotonin [169].

Studies investigating the triggers of inflammation of the macaque CNS with poliovirus inoculation have revealed that quinolinic acid, kynurenine, and other metabolites of the KP accumulate in the spinal cord and CSF (but not as much in the bloodstream) at levels at those sites that are associated with the neurological manifestations [12,148]. Moreover, the in vitro conversion of L-tryptophan to

kynurenine by fetal neuronal cells is dependent on IFN- γ stimulation in the presence of macrophages in culture [12]. Moreover, another study has shown that chronic pain in rats that are exposed to social stress or paw arthritis increases the levels of IDO and kynurenine and decreases the levels of serotonin in the hippocampus [190]. This situation is similar to that observed in human plasma levels of IDO, kynurenine/tryptophan and serotonin/tryptophan levels in patients with back pain and depression, in whom the first two rise and the third one decreases, as revealed in the same report [190]. Moreover, IDO1 knockout mice present reduced nociceptive and depressive behavior as compared with the wild type, and this behavior is not attenuated in the wild type that received an intraperitoneal injection of the NSAID acetaminophen, suggesting that this behavior is not dependent on the inflammatory mechanisms alone. Furthermore, the authors found that IL-6 is overexpressed in rats with arthritis and the Jak2/Stat3 signaling pathway is activated in the blood and hippocampus of rats with depressive and nociceptive behaviors. Injection of IL-6 anti-serum attenuated the allodynia and hyperesthesia in those animal models. In cultured Neuro2a cells (a mouse neuroblastoma cell line), incubation with IL-6 induced an increase in IDO1 mRNA and protein [190].

Therefore, persistent pain with allodynia and hyperalgesia are central components (spinal and supraspinal cord) supporting the involvement of glutamatergic neurotransmission associated with KP signaling in clinical manifestations and the role of the hippocampus as a critical organ in this process and the KP in the related physiopathology [9,30,41,190,191].

Role of the Hippocampus in the KP in Neurological Manifestations

Despite the agreement among clinical studies on changes in the KP and SS, it is admissible that the lack of an association between the symptoms of depression and fatigue in SS and the changes in the KP is due to difficulties in accessing and monitoring the changes in the CNS, more specifically in the hippocampus [18,56,192,193].

Not only SS patients, but also individuals exposed to chronic stress, present changes in the hippocampal structure and NMDA signaling [40,194]. Animal model studies have revealed that the hippocampus initially adapts to early, high, and frequent stress, but the persistence of aggression increases the levels of glutamate and disrupts hormonal and neurotransmitter control, leading to NMDA-driven neuronal death and hippocampal atrophy [195,196].

Moreover, hypothalamus-pituitary-adrenal activity has a modulatory effect on those events and the female sex hormone estrogen increases the synaptic connections and expression of nerve growth factor (NGF) in the hippocampus, supporting the conclusion that hormones influence and increase symptoms and pain sensitivity in females with SS [19,125,195,197].

Cytokines, such as IFN- γ , IL-1, and TNF- α , lead to increases in the expression of kynurenine and its metabolites, which are generated from tryptophan by the IDO enzyme, deviating this amino acid from the production of serotonin in the CNS throughout the KP (Figure 4) [13]. The resulting imbalance between the kynurenine metabolites and serotonin production in the hippocampus induces depression, slow reactions, and other cognitive disorders [198]. The target cells are microglia, astrocytes, and other inflammatory cells that are present in the hippocampus and other CNS areas [33]. Once impacted by those cytokines, the cells reduce glutamate reuptake, increase glutamatergic signaling, reduce the capacity to produce serotonin, trigger nociceptive and depressive behavior, and induce cell death (mostly the astrocytes), prolonging the inflammatory effect [13,33,41,53,199].

It is interesting to note that serotonin functions as a modulator of glutamate actions. PNS sensory transmission has silent glutamate synapses that are activated by serotonin. Once those silent synapses are activated by serotonin, they amplify the peripheral nociceptive glutamate signaling through the NMDA or α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptor (AMPA) receptors from the spinal dorsal horn to the CNS [200,201]. In the CNS, glutamate/serotonin co-neurotransmission has been extensively studied, including in the hippocampus. The five subtypes of serotonergic receptors are expressed in different combinations among several cells, antagonizing the glutamatergic

NMDA receptors at different levels, from preventing cell glutamate release to competing for the same intracellular signaling pathways in the hippocampus but not in other brain tissues [202–207].

Additionally, to demonstrate the differences in KP activity between the CNS and other parts of the body, systemic treatment with dexamethasone to reduce the inflammation that is induced by lipopolysaccharide (LPS) intraperitoneal injection was shown to promote a decrease in IDO enzymes in peripheral tissues (lung, spleen and liver) but an increase in brain microglial cells and astrocytes [199,208]. Moreover, the use of systemic subcutaneous slow-release corticosteroid pellets in rats increased the levels of NR2 NMDA glutamatergic receptors mRNA in the hippocampus [209]. These observations suggest that the use of corticosteroid treatment to reduce chronic inflammation may contribute to the nociceptive and depressive behavior over the long-term.

Additionally, in human immunodeficiency virus (HIV)-infected patients, quinolinic acid (a metabolite of kynurenine that mimics the glutamate in NMDA receptors) content is several-fold higher in the brain than in the cerebrospinal fluid or blood [210].

Taken together, these findings delineate the hippocampus as primarily responsible for nociception and mood control, as well as a site where inflammation, driven by cytokines (mostly IFN- γ), induces a rise in KP activity and its metabolites (e.g., quinolinic acid and glutamate) with neuroactive actions to induce pain and depressive behavior. Moreover, during chronic inflammation such as in SS, the levels of serotonin in the CNS are diminished by tryptophan consumption throughout the KP. Reversion of the inflammation with corticosteroids in the CNS has been unsuccessful as the inflammation is present in other target organs and leads to the death of microglia, astrocytes, and neurons, mostly in the hippocampus and dorsal ganglion root [199,208,211]. These observations support the possible mechanisms of SS neurological manifestations, in which symptoms of pain and depression (i.e., allodynia, hyperalgesia, and fatigue), manifestations of reduced tear and saliva secretion, and elevated expression of blood markers of inflammation present a dissociation or discrepancy in the affected patients. Additionally, activation of the KP provides metabolites that mimic neurotransmitters and could be the cause of this dissociation and corroborate our hypothesis, as stated at the beginning of this review (Figure 4).

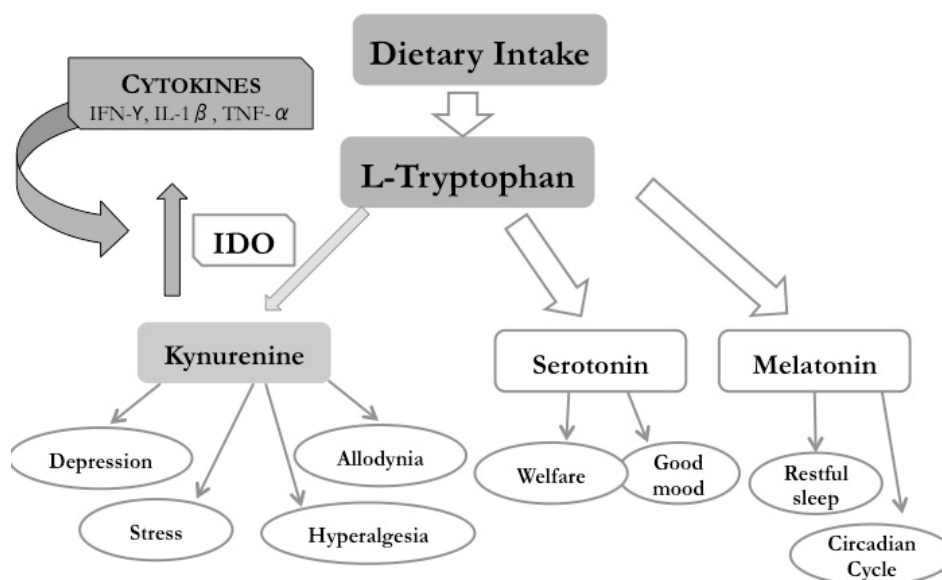


Figure 4. Sjögren's syndrome physiopathology, considering the elevated cytokine expression activation of the kynurenine pathway (KP) and its implications in the pain and mood symptoms dissociated with the signs of SS disease. The light thick gray arrows indicate the pathway to obtain L-tryptophan to produce serotonin and melatonin and the dark gray arrow indicate the catabolic pathway of the L-tryptophan and its relationship with inflammatory cytokines. The thin gray arrows indicate the clinical manifestation of different L-tryptophan metabolites.

5. KP and Neuropathy in SS

Decades ago, reports have revealed that the ingestion of L-5-hydroxytryptophan induces signs and symptoms that are similar to scleroderma, with high plasma levels of kynurenine [212]. Moreover, excessive doses of tryptophan (greater than 1.2 g/day) trigger eosinophilia, severe muscular weakness and pain, and oral ulcers, with a rise in the hepatic enzymes aspartate and alanine aminotransferase (ASA and ALA), in addition to an inflammatory infiltrate in various organs [175]. Those events were associated with the upper plasma levels of kynurenine, as observed in cases with scleroderma and SS [153,212].

Salivary gland (SG) ductal ligation in rats induces tissue damage and atrophy, increases in systemic levels of kynurenine (as measured in the hair), and is associated not only with salivary hypofunction, but also body weight loss, over the six months of the experimental period. These findings indicate that higher levels of plasma kynurenine can reflect peripheral organ damage but also that SG damage is sufficient to impact whole-body metabolism, as demonstrated by the lower body weight as compared with the controls [21].

As mentioned above, the availability of the major substrate of the KP, tryptophan, is dependent on dietary intake, intake, but it is also influenced by environmental conditions and metabolism based on an individual's genetic background [51,173–175,213–215]. For example, when female C57 ovariectomized mice ingest bisphenol A (BPA), an environmental contaminant with endocrine disruption capacity, it causes bowel inflammation and reduced levels of tryptophan and serotonin, indicating that environmental contaminants and the intestinal microbiota affect the KP in chronic inflammatory diseases [174].

Flow cytometric analysis of the peripheral blood has revealed a higher expression of IDO in the dendritic cells of pSS patients and in each of the subgroups, classified either by the presence of clinical or serological activity, as compared to the dendritic cells of healthy controls [216].

Measurements of IDO in antigen-presenting cells (APCs) and in T cells have demonstrated higher levels in the peripheral blood cells of pSS patients than controls matched by age and sex, also while using specific antibodies and flow cytometry, despite the heterogeneity of the groups and the high internal variability of the results [217]. Therefore, the T cell-mediated autoimmune activity present in autoimmune diseases, including SLE and SS, has been associated with elevated activity of the KP; however, the effects on autoantigen stimulation and IFN- γ activity remain unknown [216,217].

Different profiles regarding IFN- γ activity have been identified in the pSS population, and 55 genes and 19 metabolic pathways have been distinctly identified in a subset of pSS patients with fatigue [218,219]. Elevated IDO activity has been detected in IFN- γ -positive pSS patients, with higher levels of IDO mRNA and IFN- γ mRNA in circulating monocytes, and those observations were associated with the upregulation of apoptotic and neurotoxic downstream steps in the KP [18]. The levels of serum tryptophan are higher in healthy than in primary SS women (pSS). However, the levels of kynurenine and the kynurenine/tryptophan ratio are more elevated in pSS than in healthy women and patients with non-SS *sicca*. The same observations were found in pSS men, confirming the elevated activity of the IDO enzyme in the KP [153,220]. Moreover, the higher levels of kynurenine were associated with higher levels of inflammatory markers in the serum, such as the erythrocyte sedimentation rate, C-reactive protein, creatinine, Immunoglobulin A (IgA), β -2 microglobulin, and anti-nuclear antibody positivity [153,220]. Higher levels of kynurenine have also been associated with a lower proportion of individuals on corticosteroids but not with the frequency of neurological manifestations in the pSS group [153,220]. In another recent study, polyneuropathy showed more frequent positivity for the autoantibodies anti-Ro (SSa) and anti-La (SSb) in pSS [16]. Taken together, these studies suggest that the higher activity of the KP is related to clinical and laboratory signs of systemic inflammation, which may conflict with our hypothesis, but also indicates that those studies documented a midway point between the pain/neuropathic and the inflammatory poles of the disease [153,220,221]. Although the association of neurological or laboratory findings and kynurenine

metabolites is evident in those studies, the cause/effect relationship between the metabolites of the KP and these manifestations remains unclear.

In another study, an association of fibromyalgia and of other psychological symptoms, such as anxiety, depression, insomnia, psychoticism, and neuroticism, with fatigue being observed in a large series of pSS patients comprising 106 cases, among which 32 were fatigued and 74 non-fatigued, as identified by the Functional Assessment of Chronic Illness Therapy-Fatigue (FACIT-F) scale, with a cut-off of 30 on a scale ranging from 0 to 52. However, the levels of IDO mRNA in peripheral blood leukocytes did not differ between pSS patients with and without fatigue [192]. In addition, no other clinical or laboratory association was identified, excluding the number of individuals using hydrochloroquine, making up 50% of the pSS fatigue group and 28% of the pSS nonfatigue group. In contrast, the higher expression of IDO-1 mRNA levels has been associated with plasma levels of IFN- γ [192].

Fatigue has been associated with high KP activity levels in SLE patients, but only in those with clinical activity of the disease, as measured by the Systemic Lupus Erythematosus Disease Activity Index (SLEDAI) with a score above five, which may confirm the possibility that the KP is overexpressed in the presence of elevated inflammatory activity [50]. Moreover, the levels of serum tryptophan are lower in SLE than in controls, systemic sclerosis (SSc), and pSS patients, which may reflect the higher activity and broad manifestations of the disease as compared to the other two conditions (i.e., pSS and SSc), which are more tissue-specific than SLE [193]. Interestingly, a modular IFN signature, including INF- α , INF- β , and INF- γ , has been identified in the majority of cases in a series of consecutive SLE patients [222]. Moreover, a metabolomic analysis comparing serum samples of SLE, pSS, and SSc patients and healthy volunteers allowed a specificity classification of 67% of the SLE group when compared with the other three groups, in which the most discriminatory metabolite was tryptophan, with lower levels in SLE than in the other groups [193]. The authors interpreted this reduction of tryptophan as a response to KP activation [193]. Experimental studies mimic the clinical findings of cognitive impairment, but not of depression, associated with the activation of microglia and astrocyte in the hippocampus in SLE mouse models induced by the injection of anti-ribosomal antibodies in the CNS as compared to controls [223]. Unfortunately, the KP was not investigated in that study.

The evidence collected thus far from the medical literature does not confirm that the KP is the one and sole pathway responsible for the neurological manifestations of SS. In fact, two studies have presented opposing evidence, with higher KP metabolites being associated with higher levels of inflammatory markers [153,220]. This result may be due to (a) the heterogeneity of the SS cases recruited for clinical studies in terms of demography and disease duration; (b) the absence of a control group with healthy individuals in some studies; (c) alterations in the levels of metabolites with neuroactive properties in CNS tissues by KP activity-induced neurogenic changes, especially the hippocampus and dorsal ganglion root, but not in the blood, where they are measured in the actual studies; and, (d) observations collected in the middle of the inflammatory process and thus not representative of the anti-inflammatory effect of KP stated in the hypothesis. Such pitfalls must be taken into consideration in future studies addressing the present hypothesis of the association between SS neurological features and the KP.

6. Therapy to Modulate the KP

The overexpression of IFN- γ , induction of pro-inflammatory genes, such as TNF- α , interleukins, and B-cell activating factor (BAFF), the promotion of B cell activation and rise in autoantibodies in the blood are involved in the pathophysiology of SS [99,156,224–228]. Therefore, therapeutic strategies to treat SS include immune modulators and biological therapy to inhibit B and T cells activity and proliferation [229–232]. The limitations of such strategies open opportunities for new procedures and complementary therapies. Among several possibilities, the potential modulation of KP has been explored in SS, as in other autoimmune diseases, including rheumatoid arthritis, SLE, and systemic sclerosis [15,233]. Experimental studies showing the effects of inflammatory challenges on the levels of

KP metabolites and subsequent functional responses of the therapeutic modulation of this pathway in pain and neurological parameters are not always synergic and positive, which may be a consequence of observations that were collected at different time points and rebound effects [105,142,234–243].

Considering the broad spectrum of substrates, target tissues, and alternative effects of KP, therapeutic strategies to interfere with different steps have resulted in one or more of the following outcomes: attenuation of inflammation, reduction of chronic pain or improvement of fatigue, and depressive feelings in SS [18,217,233].

The traditional nonsteroidal anti-inflammatory drugs (NSAID), acetylsalicylic acid (ASA or aspirin), and sodium diclofenac have been investigated [244,245]. In rats, systemic intraperitoneal injection of tryptophan alone was able to increase kynurenine levels between 20 and 120 min. However, after combined injection of subcutaneous diclofenac with intraperitoneal tryptophan, the concentration of kynurenine increased, not just in the plasma and liver but also in the spinal cord and brain, with a remarkable increase in the kidney, after 60 and 120 min. Therefore, diclofenac disrupts the renal clearance of KP metabolites, which may amplify anti-inflammatory and excitatory stimuli on nociceptive NMDA receptors, independently of the analgesic effects of prostaglandins [245]. In an opposite manner, a study using human peripheral blood mononuclear cells (PBMCs) has revealed that aspirin at the dose of 5 mM, incubated for three days or 2 h, reduces tryptophan metabolism and kynurenine production in those PBMCs stimulated by concanavalin A and pokeweed mitogen (PKM), suggesting an inhibitory effect on IFN- γ based on the triggered mechanisms of action [244]. These observations indicate that NSAIDs have functions in addition to their known effects on the cyclooxygenase/prostaglandin pathway, and the impact on KP can be diverse, depending on the cell type and the specific NSAID [15]. It is also interesting to note that aspirin, a longer and broadly used analgesic drug in the NSAID group, has well-known positive effects on dry eye symptoms and LG dysfunction, which are critical elements in the manifestation of SS [246–248].

In an *in vitro* study using samples of T cells from 68 SS patients, coculture with mesenchymal stem umbilical cells revealed the suppression of proliferation and activation of these circulating follicular T helper cells, in association with the enhanced expression and enzymatic activity of IDO, as measured by reversal transcription polymerase chain reaction (RT-PCR) and high performance liquid chromatography (HPLC), respectively [249]. Another study has shown that human complementarity determining region 1 (hCDR1), a tolerogenic peptide that is complementary to the human anti-DNA monoclonal antibody, reduces the expression of inflammatory cytokines, down regulates the proliferation and activity of B cells, and increases the expression of anti-inflammatory cytokines in a rodent model of SLE [233]. In cultured mature leukocytes from 16 SS individuals, hCDR1 has been shown to reduce the expression of inflammatory cytokines, including IFN- γ , and increase the expression of anti-inflammatory cytokines, up regulating IDO gene expression. However, in the presence of 1mT, an IDO inhibitor, the effect of hCDR1 on the gene expression of the T cell regulator cytokine fork head box protein-3 (FOXP3) is reduced, suggesting that the immunomodulatory effect of hCDR1 is partially associated with its effect on IDO [233].

Suppression of the KP by inhibiting the KMO can reduce the pain that is triggered by LPS injection in the dorsal ganglion root in rodents. Systemic administration of the antibiotic minocycline or local administration of inhibitors of KMO [105] reduces the local levels of pro-inflammatory cytokines in the dorsal ganglion root and spinal cord and decreases pain and the protein expression of the following inflammatory mediators: ionized calcium-binding adapter molecule 1 (IBA-1), IL-6, IL-1 β , and Nitric oxide synthase 2 (NOS2) [105].

When considering the strategy to overload KP to modulate the pain sensation, rats were subject to systemic administration of L-4-chlorokynurenine [250]. The experiments revealed that L-4-chlorokynurenine, a NMDA/glutamate receptor antagonist, administered by intraperitoneal injection reached the CNS and attenuated the hyperalgesia in four models of pain and behavioral response (general behavioral, formalin plantar injection, Carrageenan model, and Chung neuropathy) as compared with the controls, Dizocilpine (MK-801), and gabapentin [250].

Twelve of 20 GVHD individuals, who were non-responders to corticosteroids, presented clinical improvement in skin inflammation with human chorionic gonadotropin (hCG) treatment [251]. They also showed a significant increase in IDO mRNA expression in PBMCs and in IL-10 expression in the blood serum [251]. The underlying mechanism of action was thought to occur via stimulation of IDO-mediated immunotolerance, similar to the mother/fetus coexistence [144].

Although useful in its conception and to revert immune-mediated diseases, KP may be deleterious in neoplastic diseases, where it can allow tumor growth by inducing IFN secretion, and, after activating IDO, suppress the immune response toward the tumor [252]. According to this concept, the first human clinical trial aiming to block KP as an anti-cancer therapy was recently published [253]. The study investigated whether orally administered epacadostat, an IDO inhibitor, would be well tolerated and capable of slowing the growth of tumors in 52 refractory cancer patients by removing the immune tolerance to those tumors. The drug, at doses of 200 mg/day, reduced kynurenine plasma levels, indicating a reduction of tryptophan degradation. The mean treatment duration was 52 (from 7 to 284) days, and the daily doses ranged from 43 to 1400 mg. The side effects included fatigue, nausea, and pain, among others. However, no plasma changes in C-reactive protein or in the levels of the tested interleukins were observed [253]. The data confirmed the expected manifestations of KP inhibition and a safe strategy overall. Further conclusions are limited due to the small number and heterogeneity of the clinical cases.

Therefore, the present data on interventions in the KP reveal dual direction activities, in which inhibition at specific steps, such as KMO activity and quinolinic acid formation, has beneficial effects on neuropathic pain and neurodegenerative disorders, whereas enhancing the activity of IDO ultimately leads to an inhibition of pro-inflammatory cytokines and the reduction of inflammatory processes. How and at which steps those events can reach a conciliatory mechanism to diminish chronic pain and neurological symptoms in SS patients, but also prevent chronic inflammatory reactions will be the subject of further investigations.

7. Future Perspectives

IFN- γ triggers the deviation of the tryptophan to the KP pathway, possibly contributing to depression and pain through its action on particular organs, such as the hippocampus. This signaling pathway modulates the inflammation and chronic damage of the PNS and CNS, with potential repercussions on the SG, joints, and LFU. The suppression of APCs and production of anti-inflammatory cytokines in exocrine glands and other target tissues are the potential benefits of KP actions. Therefore, the elevated expression of IDO and kynurenine metabolites in SS suggests, but to date does not clearly indicate, a reactive process to modulate the mechanism of inflammation induced by other pathways. Improved strategies to access the CNS and PNS organs by imaging analysis and to monitor the local activity of the KP in the involved organs in SS, glandular or not, would facilitate insights regarding the physiopathology of this signaling pathway in SS, its interference in exocrine secretion impairment and strategies for improvement. More effective treatments and an enhanced quality of life for SS patients will be potential benefits from this knowledge.

8. Conclusions

Glandular and EGM of SS are not exclusively inflammatory but also involve a neuroimmunendocrine network, in which the KP plays a role. The involvement of the KP is difficult to track and confirm because of the delicate methods for tracing, the timing of the response, and the location of the metabolites of this pathway in the CNS. A better understanding of the relationship between the physiopathology of SS and the KP in the CNS and target tissues may help to clarify the discrepancies among the signs and symptoms and the neurological manifestations. This knowledge could improve therapy for SS.

Author Contributions: Conceptualization: F.R.d.O., V.V., T.M.C., P.L.-J. and E.M.R.; Methodology: F.R.d.O., V.V., P.L.-J. and E.M.R.; Formal Analysis: M.Z.F., F.R.d.O., T.M.C. and E.M.R.; Investigation: L.A., M.Z.F., F.R.d.O.,

T.M.C. and E.M.R.; Resources: T.M.C., E.M.R.; Data Curation: F.R.d.O., T.M.C., V.V. and E.M.R.; Writing—Original Draft Preparation: F.R.d.O. and E.M.R.; Writing—Review & Editing: F.R.d.O., V.V., L.A., M.Z.F., T.M.C., P.L.-J. and E.M.R.; Project Administration, P.L.-J. and E.M.R.; Funding Acquisition: T.M.C., P.L.-J. and E.M.R.

Funding: This research was funded by the following Brazilian public agencies: Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP) (2014-23211-0), Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) (32355-2017-0), Coordenação de Pessoal de Nível Superior (CAPES) (Finance code 001) e Fundação de Apoio ao Ensino, Pesquisa e Assistência do Hospital das Clínicas da Faculdade de Medicina de Ribeirão Preto, Universidade de São Paulo (FAEPA).

Acknowledgments: The authors would like to acknowledge Rogerio Aparecido Mazzucato Castania, Adriana de Andrade Batista Murashima and Rita Amâncio Diegues for technical assistance with this work.

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

5-HT	serotonin
APCs	antigen-presenting cells
ASA	acetyl salicylic acid
CIDP	chronic inflammatory demyelinating polyradiculoneuropathy
CNS	central nervous system
EGM	extraglandular manifestations
GVHD	graft-versus host disease
hCDR1	human complementarity determining region 1
HCG	human chorionic gonadotropin
IDO	indoleamine 2,3-dioxygenase
IFN- γ	Interferon- γ
KAT	kynurenine aminotransferase
KMO	Kynurenine 3
LFU	lacrimal functional unit
LG	lacrimal gland
LTF	L-tryptophan
NAD+	nicotinamide adenine nucleotide
NAISD	nonsteroidal anti-inflammatory drugs
NMDA	N-methyl-D-aspartate
OAS1	2-5 oligo-adenylate synthetase 1 PBC
PBMCs	peripheral blood mononuclear cells
PKM	pokeweed mitogen
PNS	peripheral nervous system
SLE	systemic lupus erythematosus
SLEDAI	Systemic Lupus Erythematosus Disease Activity Index
SS	Sjögren's syndrome
TDO	tryptophan 2,3-dioxygenase
TG	trigeminal ganglion
KP	kynurenine pathway

References

1. Vitali, C.; Bombardieri, S.; Jonsson, R.; Moutsopoulos, H.; Alexander, E.; Carsons, S.; Daniels, T.; Fox, P.; Fox, R.; Kassan, S.; et al. Classification criteria for Sjögren's syndrome: A revised version of the European criteria proposed by the American-European Consensus Group. *Ann. Rheum. Dis.* **2002**, *61*, 554–558. [[CrossRef](#)] [[PubMed](#)]
2. Delalande, S.; de Seze, J.; Fauchais, A.L.; Hachulla, E.; Stojkovic, T.; Ferriby, D.; Dubucquoi, S.; Pruvo, J.P.; Vermersch, P.; Hatron, P.Y. Neurologic manifestations in primary Sjogren syndrome: A study of 82 patients. *Medicine* **2004**, *83*, 280–291. [[CrossRef](#)] [[PubMed](#)]

3. Humphreys-Beher, M.G.; Brayer, J.; Yamachika, S.; Peck, A.B.; Jonsson, R. An alternative perspective to the immune response in autoimmune exocrinopathy: Induction of functional quiescence rather than destructive autoaggression. *Scand. J. Immunol.* **1999**, *49*, 7–10. [[CrossRef](#)] [[PubMed](#)]
4. Hayashi, T. Dysfunction of lacrimal and salivary glands in Sjögren's syndrome: Nonimmunologic injury in preinflammatory phase and mouse model. *J. Biomed. Biotechnol.* **2011**, *2011*, 407031. [[CrossRef](#)] [[PubMed](#)]
5. Van Bijsterveld, O.P.; Kruijze, A.A.; Bleyers, R.L. Central nervous system mechanisms in Sjogren's syndrome. *Br. J. Ophthalmol.* **2003**, *87*, 128–130. [[CrossRef](#)] [[PubMed](#)]
6. Seror, R.; Ravaud, P.; Bowman, S.J.; Baron, G.; Tzioufas, A.; Theander, E.; Gottenberg, J.E.; Bootsma, H.; Mariette, X.; Vitali, C.; et al. EULAR Sjogren's syndrome disease activity index: Development of a consensus systemic disease activity index for primary Sjogren's syndrome. *Ann. Rheum. Dis.* **2010**, *69*, 1103–1109. [[CrossRef](#)] [[PubMed](#)]
7. Murube, J. The first definition of Sjogren's syndrome. *Ocul. Surf.* **2010**, *8*, 101–110. [[CrossRef](#)]
8. Ramos-Casals, M.; Brito-Zeron, P.; Seror, R.; Bootsma, H.; Bowman, S.J.; Dorner, T.; Gottenberg, J.E.; Mariette, X.; Theander, E.; Bombardieri, S.; et al. Characterization of systemic disease in primary Sjogren's syndrome: EULAR-SS Task Force recommendations for articular, cutaneous, pulmonary and renal involvements. *Rheumatology* **2015**, *54*, 2230–2238. [[CrossRef](#)]
9. Koh, J.H.; Kwok, S.K.; Lee, J.; Son, C.N.; Kim, J.M.; Kim, H.O.; Park, S.H.; Sung, Y.K.; Choe, J.Y.; Lee, S.S.; et al. Pain, xerostomia, and younger age are major determinants of fatigue in Korean patients with primary Sjogren's syndrome: A cohort study. *Scand. J. Rheumatol.* **2017**, *46*, 49–55. [[CrossRef](#)]
10. Brito-Zeron, P.; Theander, E.; Baldini, C.; Seror, R.; Retamozo, S.; Quartuccio, L.; Bootsma, H.; Bowman, S.J.; Dorner, T.; Gottenberg, J.E.; et al. Early diagnosis of primary Sjogren's syndrome: EULAR-SS task force clinical recommendations. *Expert Rev. Clin. Immunol.* **2016**, *12*, 137–156. [[CrossRef](#)]
11. Seara, F.A.C.; Maciel, L.; Barbosa, R.A.Q.; Rodrigues, N.C.; Silveira, A.L.B.; Marassi, M.P.; Carvalho, A.B.; Nascimento, J.H.M.; Olivares, E.L. Cardiac ischemia/reperfusion injury is inversely affected by thyroid hormones excess or deficiency in male Wistar rats. *PLoS ONE* **2018**, *13*, e0190355. [[CrossRef](#)]
12. Saito, K.; Nowak, T.S., Jr.; Markey, S.P.; Heyes, M.P. Mechanism of delayed increases in kynurenine pathway metabolism in damaged brain regions following transient cerebral ischemia. *J. Neurochem.* **1993**, *60*, 180–192. [[CrossRef](#)] [[PubMed](#)]
13. Muller, N.; Schwarz, M.J. The immune-mediated alteration of serotonin and glutamate: Towards an integrated view of depression. *Mol. Psychiatry* **2007**, *12*, 988–1000. [[CrossRef](#)]
14. Vecsei, L.; Szalardy, L.; Fulop, F.; Toldi, J. Kynurenines in the CNS: Recent advances and new questions. *Nat. Rev.* **2013**, *12*, 64–82. [[CrossRef](#)]
15. Filippini, P.; Del Papa, N.; Sambataro, D.; Del Bufalo, A.; Locatelli, F.; Rutella, S. Emerging concepts on inhibitors of indoleamine 2,3-dioxygenase in rheumatic diseases. *Curr. Med. Chem.* **2012**, *19*, 5381–5393. [[CrossRef](#)] [[PubMed](#)]
16. Ter Borg, E.J.; Kelder, J.C. Development of new extra-glandular manifestations or associated auto-immune diseases after establishing the diagnosis of primary Sjogren's syndrome: A long-term study of the Antonius Nieuwegein Sjogren (ANS) cohort. *Rheumatol. Int.* **2017**, *37*, 1153–1158. [[CrossRef](#)] [[PubMed](#)]
17. Zhang, X.; Chen, W.; De Paiva, C.S.; Corrales, R.M.; Volpe, E.A.; McClellan, A.J.; Farley, W.J.; Li, D.Q.; Pflugfelder, S.C. Interferon-gamma exacerbates dry eye-induced apoptosis in conjunctiva through dual apoptotic pathways. *Investig. Ophthalmol. Vis. Sci.* **2011**, *52*, 6279–6285. [[CrossRef](#)]
18. Maria, N.I.; van Helden-Meeuwse, C.G.; Brkic, Z.; Paulissen, S.M.; Steenwijk, E.C.; Dalm, V.A.; van Daele, P.L.; Martin van Hagen, P.; Kroese, F.G.; van Roon, J.A.; et al. Association of Increased Treg Cell Levels With Elevated Indoleamine 2,3-Dioxygenase Activity and an Imbalanced Kynurenine Pathway in Interferon-Positive Primary Sjogren's Syndrome. *Arthritis Rheumatol.* **2016**, *68*, 1688–1699. [[CrossRef](#)]
19. Sullivan, D.A.; Rocha, E.M.; Aragona, P.; Clayton, J.A.; Ding, J.; Golebiowski, B.; Hampel, U.; McDermott, A.M.; Schaumberg, D.A.; Srinivasan, S.; et al. TFOS DEWS II Sex, Gender, and Hormones Report. *Ocul. Surf.* **2017**, *15*, 284–333. [[CrossRef](#)]
20. Imada, T.; Nakamura, S.; Hisamura, R.; Izuta, Y.; Jin, K.; Ito, M.; Kitamura, N.; Tanaka, K.F.; Mimura, M.; Shibuya, I.; et al. Serotonin hormonally regulates lacrimal gland secretory function via the serotonin type 3a receptor. *Sci. Rep.* **2017**, *7*, 6965. [[CrossRef](#)]
21. Ikeno, K.; Saikatsu, S.; Uno, T.; Ikeno, T. Effects of prolonged duct ligation of the rat salivary glands on the activity of trypsin-like protease. *Arch. Oral Biol.* **1988**, *33*, 613–615. [[CrossRef](#)]

22. Dartt, D.A. Neural regulation of lacrimal gland secretory processes: Relevance in dry eye diseases. *Prog. Retin. Eye Res.* **2009**, *28*, 155–177. [[CrossRef](#)] [[PubMed](#)]
23. Perez, P.; Rowzee, A.M.; Zheng, C.; Adriaansen, J.; Baum, B.J. Salivary epithelial cells: An unassuming target site for gene therapeutics. *Int. J. Biochem. Cell Biol.* **2010**, *42*, 773–777. [[CrossRef](#)]
24. Lemp, M.A.; Wolfley, D.E. The Lacrimal Apparatus. In *Adler's Physiology of the Eye*, 9th ed.; Hart, W.M., Jr., Ed.; MosbyYear Book, Inc.: St. Louis, MO, USA, 1992.
25. Willcox, M.D.P.; Argueso, P.; Georgiev, G.A.; Holopainen, J.M.; Laurie, G.W.; Millar, T.J.; Papas, E.B.; Rolland, J.P.; Schmidt, T.A.; Stahl, U.; et al. TFOS DEWS II Tear Film Report. *Ocul. Surf.* **2017**, *15*, 366–403. [[CrossRef](#)] [[PubMed](#)]
26. Meng, I.D.; Kurose, M. The role of corneal afferent neurons in regulating tears under normal and dry eye conditions. *Exp. Eye Res.* **2013**, *117*, 79–87. [[CrossRef](#)] [[PubMed](#)]
27. Stern, M.E.; Gao, J.; Siemasko, K.F.; Beuerman, R.W.; Pflugfelder, S.C. The role of the lacrimal functional unit in the pathophysiology of dry eye. *Exp. Eye Res.* **2004**, *78*, 409–416. [[CrossRef](#)] [[PubMed](#)]
28. Bron, A.J.; de Paiva, C.S.; Chauhan, S.K.; Bonini, S.; Gabison, E.E.; Jain, S.; Knop, E.; Markoulli, M.; Ogawa, Y.; Perez, V.; et al. TFOS DEWS II pathophysiology report. *Ocul. Surf.* **2017**, *15*, 438–510. [[CrossRef](#)] [[PubMed](#)]
29. Belmonte, C.; Nichols, J.J.; Cox, S.M.; Brock, J.A.; Begley, C.G.; Bereiter, D.A.; Dartt, D.A.; Galor, A.; Hamrah, P.; Ivanusic, J.J.; et al. TFOS DEWS II pain and sensation report. *Ocul. Surf.* **2017**, *15*, 404–437. [[CrossRef](#)]
30. Rosenthal, P.; Borsook, D. The corneal pain system. Part I: The missing piece of the dry eye puzzle. *Ocul. Surf.* **2012**, *10*, 2–14. [[CrossRef](#)]
31. Gur, A.; Oktayoglu, P. Central nervous system abnormalities in fibromyalgia and chronic fatigue syndrome: New concepts in treatment. *Curr. Pharm. Des.* **2008**, *14*, 1274–1294. [[CrossRef](#)]
32. Rosenthal, P.; Baran, I.; Jacobs, D.S. Corneal pain without stain: Is it real? *Ocul. Surf.* **2009**, *7*, 28–40. [[CrossRef](#)]
33. Fasick, V.; Spengler, R.N.; Samankan, S.; Nader, N.D.; Ignatowski, T.A. The hippocampus and TNF: Common links between chronic pain and depression. *NeuroSci. Biobehav. Rev.* **2015**, *53*, 139–159. [[CrossRef](#)]
34. Tracey, K.J. Physiology and immunology of the cholinergic antiinflammatory pathway. *J. Clin. Investig.* **2007**, *117*, 289–296. [[CrossRef](#)] [[PubMed](#)]
35. Lockwood, A.; Hope-Ross, M.; Chell, P. Neurotrophic keratopathy and diabetes mellitus. *Eye* **2006**, *20*, 837–839. [[CrossRef](#)]
36. Partanen, J.; Niskanen, L.; Lehtinen, J.; Mervaala, E.; Siitonen, O.; Uusitupa, M. Natural history of peripheral neuropathy in patients with non-insulin-dependent diabetes mellitus. *N. Engl. J. Med.* **1995**, *333*, 89–94. [[CrossRef](#)] [[PubMed](#)]
37. Pavlov, V.A.; Tracey, K.J. Neural regulation of immunity: Molecular mechanisms and clinical translation. *Nat. Neurosci.* **2017**, *20*, 156–166. [[CrossRef](#)] [[PubMed](#)]
38. Tuisku, I.S.; Konttinen, Y.T.; Konttinen, L.M.; Tervo, T.M. Alterations in corneal sensitivity and nerve morphology in patients with primary Sjogren's syndrome. *Exp. Eye Res.* **2008**, *86*, 879–885. [[CrossRef](#)] [[PubMed](#)]
39. Nguyen, D.H.; Vadlamudi, V.; Toshida, H.; Beuerman, R.W. Loss of parasympathetic innervation leads to sustained expression of pro-inflammatory genes in the rat lacrimal gland. *Auton. NeuroSci.* **2006**, *124*, 81–89. [[CrossRef](#)]
40. Lauvsnes, M.B.; Beyer, M.K.; Kvaloy, J.T.; Greve, O.J.; Appenzeller, S.; Kvivik, I.; Harboe, E.; Tjensvoll, A.B.; Goransson, L.G.; Omdal, R. Association of hippocampal atrophy with cerebrospinal fluid antibodies against the NR2 subtype of the N-methyl-D-aspartate receptor in patients with systemic lupus erythematosus and patients with primary Sjogren's syndrome. *Arthritis Rheumatol.* **2014**, *66*, 3387–3394. [[CrossRef](#)]
41. Levite, M. Glutamate receptor antibodies in neurological diseases: Anti-AMPA-GluR3 antibodies, anti-NMDA-NR1 antibodies, anti-NMDA-NR2A/B antibodies, anti-mGluR1 antibodies or anti-mGluR5 antibodies are present in subpopulations of patients with either: Epilepsy, encephalitis, cerebellar ataxia, systemic lupus erythematosus (SLE) and neuropsychiatric SLE, Sjogren's syndrome, schizophrenia, mania or stroke. These autoimmune anti-glutamate receptor antibodies can bind neurons in few brain regions, activate glutamate receptors, decrease glutamate receptor's expression, impair glutamate-induced signaling and function, activate blood brain barrier endothelial cells, kill neurons, damage the brain, induce behavioral/psychiatric/cognitive abnormalities and ataxia in animal models, and can be removed or silenced in some patients by immunotherapy. *J. Neural Transm.* **2014**, *121*, 1029–1075.

42. Hay, E.M.; Thomas, E.; Pal, B.; Hajeer, A.; Chambers, H.; Silman, A.J. Weak association between subjective symptoms or and objective testing for dry eyes and dry mouth: Results from a population based study. *Ann. Rheum. Dis.* **1998**, *57*, 20–24. [[CrossRef](#)] [[PubMed](#)]
43. Nichols, K.K.; Nichols, J.J.; Mitchell, G.L. The lack of association between signs and symptoms in patients with dry eye disease. *Cornea* **2004**, *23*, 762–770. [[CrossRef](#)]
44. Vehof, J.; Kozareva, D.; Hysi, P.G.; Harris, J.; Nessa, A.; Williams, F.K.; Bennett, D.L.; McMahon, S.B.; Fahy, S.J.; Direk, K.; et al. Relationship between dry eye symptoms and pain sensitivity. *JAMA Ophthalmol.* **2013**, *131*, 1304–1308. [[CrossRef](#)] [[PubMed](#)]
45. Galor, A.; Covington, D.; Levitt, A.E.; McManus, K.T.; Seiden, B.; Felix, E.R.; Kalangara, J.; Feuer, W.; Patin, D.J.; Martin, E.R.; et al. Neuropathic Ocular Pain due to Dry Eye Is Associated With Multiple Comorbid Chronic Pain Syndromes. *J. Pain* **2016**, *17*, 310–318. [[CrossRef](#)] [[PubMed](#)]
46. Alves, M.; Reinach, P.S.; Paula, J.S.; Vellasco e Cruz, A.A.; Bachette, L.; Faustino, J.; Aranha, F.P.; Vigorito, A.; de Souza, C.A.; Rocha, E.M. Comparison of diagnostic tests in distinct well-defined conditions related to dry eye disease. *PLoS ONE* **2014**, *9*, e97921. [[CrossRef](#)] [[PubMed](#)]
47. Barboza, M.N.; Barboza, G.N.; de Melo, G.M.; Sato, E.; Dantas, M.C.; Dantas, P.E.; Felberg, S. Correlation between signals and symptoms of dry eye in Sjögren’s syndrome patients. *Arq. Bras. Oftalmol.* **2008**, *71*, 547–552. [[CrossRef](#)] [[PubMed](#)]
48. Seror, R.; Gottenberg, J.E.; Devauchelle-Pensec, V.; Dubost, J.J.; Le Guern, V.; Hayem, G.; Fauchais, A.L.; Goeb, V.; Hachulla, E.; Hatron, P.Y.; et al. European League Against Rheumatism Sjogren’s Syndrome Disease Activity Index and European League Against Rheumatism Sjogren’s Syndrome Patient-Reported Index: A complete picture of primary Sjogren’s syndrome patients. *Arthritis Care Res.* **2013**, *65*, 1358–1364. [[CrossRef](#)] [[PubMed](#)]
49. Vehof, J.; Sillevs Smitt-Kamminga, N.; Kozareva, D.; Nibourg, S.A.; Hammond, C.J. Clinical Characteristics of Dry Eye Patients With Chronic Pain Syndromes. *Am. J. Ophthalmol.* **2016**, *162*, 59–65. [[CrossRef](#)]
50. Akesson, K.; Pettersson, S.; Stahl, S.; Surowiec, I.; Hedenstrom, M.; Eketjall, S.; Trygg, J.; Jakobsson, P.J.; Gunnarsson, I.; Svenungsson, E.; et al. Kynurenine pathway is altered in patients with SLE and associated with severe fatigue. *Lupus Sci. Med.* **2018**, *5*, e000254. [[CrossRef](#)]
51. Curzon, G.; Bridges, P.K. Tryptophan metabolism in depression. *J. Neurol. Neurosurg. Psychiatry* **1970**, *33*, 698–704. [[CrossRef](#)]
52. Heyes, M.P.; Saito, K.; Crowley, J.S.; Davis, L.E.; Demitrack, M.A.; Der, M.; Dilling, L.A.; Elia, J.; Kruesi, M.J.; Lackner, A.; et al. Quinolinic acid and kynurenine pathway metabolism in inflammatory and non-inflammatory neurological disease. *Brain* **1992**, *115 Pt 5*, 1249–1273. [[CrossRef](#)]
53. Laumet, G.; Zhou, W.; Dantzer, R.; Edralin, J.D.; Huo, X.; Budac, D.P.; O’Connor, J.C.; Lee, A.W.; Heijnen, C.J.; Kavelaars, A. Upregulation of neuronal kynurenine 3-monooxygenase mediates depression-like behavior in a mouse model of neuropathic pain. *Brain Behav. Immun.* **2017**, *66*, 94–102. [[CrossRef](#)] [[PubMed](#)]
54. Remus, J.L.; Dantzer, R. Inflammation Models of Depression in Rodents: Relevance to Psychotropic Drug Discovery. *Int. J. Neuropsychopharmacol.* **2016**, *19*. [[CrossRef](#)] [[PubMed](#)]
55. Bortolato, B.; Berk, M.; Maes, M.; McIntyre, R.S.; Carvalho, A.F. Fibromyalgia and Bipolar Disorder: Emerging Epidemiological Associations and Shared Pathophysiology. *Curr. Mol. Med.* **2016**, *16*, 119–136. [[CrossRef](#)] [[PubMed](#)]
56. Valim, V.; Sardemberg, W.M.; Brun, J.G.; Zandonade, E.; Balarini, G.M.; Tanure, L.V.; Ferreira, G.V.; Serrano, E.V.; Tonini, J.F.V.; Brokstad, K.A.; et al. Interferon-gamma-inducible kynurenines inflammation pathway: The missing link between disease activity and symptoms in Sjögren’s syndrome. *Ann. Rheum. Dis.* **2017**, *76*, 1102.
57. Mori, K.; Iijima, M.; Koike, H.; Hattori, N.; Tanaka, F.; Watanabe, H.; Katsuno, M.; Fujita, A.; Aiba, I.; Ogata, A.; et al. The wide spectrum of clinical manifestations in Sjogren’s syndrome-associated neuropathy. *Brain* **2005**, *128*, 2518–2534. [[CrossRef](#)] [[PubMed](#)]
58. Alexander, G.E.; Provost, T.T.; Stevens, M.B.; Alexander, E.L. Sjogren syndrome: Central nervous system manifestations. *Neurology* **1981**, *31*, 1391–1396. [[CrossRef](#)]
59. Barendregt, P.J.; van den Bent, M.J.; van Raaij-van den Aarssen, V.J.; van den Meiracker, A.H.; Vecht, C.J.; van der Heijde, G.L.; Markusse, H.M. Involvement of the peripheral nervous system in primary Sjogren’s syndrome. *Ann. Rheum. Dis.* **2001**, *60*, 876–881.

60. Chai, J.; Logigian, E.L. Neurological manifestations of primary Sjogren's syndrome. *Curr. Opin. Neurol.* **2010**, *23*, 509–513. [[CrossRef](#)]
61. Goransson, L.G.; Herigstad, A.; Tjensvoll, A.B.; Harboe, E.; Mellgren, S.I.; Omdal, R. Peripheral neuropathy in primary sjogren syndrome: A population-based study. *Arch. Neurol.* **2006**, *63*, 1612–1615. [[CrossRef](#)]
62. Indart, S.; Hugon, J.; Guillausseau, P.J.; Gilbert, A.; Dumurgier, J.; Paquet, C.; Sene, D. Impact of pain on cognitive functions in primary Sjogren syndrome with small fiber neuropathy: 10 cases and a literature review. *Medicine* **2017**, *96*, e6384. [[CrossRef](#)] [[PubMed](#)]
63. Terkelsen, A.J.; Karlsson, P.; Lauria, G.; Freeman, R.; Finnerup, N.B.; Jensen, T.S. The diagnostic challenge of small fibre neuropathy: Clinical presentations, evaluations, and causes. *Lancet Neurol.* **2017**, *16*, 934–944. [[CrossRef](#)]
64. Kocer, B.; Tezcan, M.E.; Batur, H.Z.; Haznedaroglu, S.; Goker, B.; Irkec, C.; Cetinkaya, R. Cognition, depression, fatigue, and quality of life in primary Sjogren's syndrome: Correlations. *Brain Behav.* **2016**, *6*, e00586. [[CrossRef](#)] [[PubMed](#)]
65. Tezcan, M.E.; Kocer, E.B.; Haznedaroglu, S.; Sonmez, C.; Mercan, R.; Yucel, A.A.; Irkec, C.; Bitik, B.; Goker, B. Primary Sjogren's syndrome is associated with significant cognitive dysfunction. *Int. J. Rheum. Dis.* **2016**, *19*, 981–988. [[CrossRef](#)] [[PubMed](#)]
66. Milin, M.; Cornec, D.; Chastaing, M.; Griner, V.; Berrouguet, S.; Nowak, E.; Marhadour, T.; Saraux, A.; Devauchelle-Pensec, V. Sicca symptoms are associated with similar fatigue, anxiety, depression, and quality-of-life impairments in patients with and without primary Sjogren's syndrome. *Jt. Bone Spine* **2016**, *83*, 681–685. [[CrossRef](#)] [[PubMed](#)]
67. Imrich, R.; Alevizos, I.; Bebris, L.; Goldstein, D.S.; Holmes, C.S.; Illei, G.G.; Nikolov, N.P. Predominant Glandular Cholinergic Dysautonomia in Patients With Primary Sjogren's Syndrome. *Arthritis Rheumatol.* **2015**, *67*, 1345–1352. [[CrossRef](#)] [[PubMed](#)]
68. Qiao, L.; Wang, Q.; Fei, Y.; Zhang, W.; Xu, Y.; Zhang, Y.; Zhao, Y.; Zeng, X.; Zhang, F. The Clinical Characteristics of Primary Sjogren's Syndrome with Neuromyelitis Optica Spectrum Disorder in China: A STROBE-Compliant Article. *Medicine* **2015**, *94*, e1145. [[CrossRef](#)]
69. Abu-Amero, K.K.; Helwa, I.; Al-Muammar, A.; Strickland, S.; Hauser, M.A.; Allingham, R.R.; Liu, Y. Screening of the Seed Region of MIR184 in Keratoconus Patients from Saudi Arabia. *BioMed Res. Int.* **2015**, *2015*, 604508.
70. Brito, G.N.; Araujo, G.R.; Papi, J.A. Neuropsychological, neuroimage and psychiatric aspects of primary Sjogren's syndrome. *Arq. Neuro-Psiquiatr.* **2002**, *60*, 28–31. [[CrossRef](#)]
71. Carvalho, D.C.; Tironi, T.S.; Freitas, D.S.; Kleinpaul, R.; Talim, N.C.; Lana-Peixoto, M.A. Sjogren syndrome and neuromyelitis optica spectrum disorder co-exist in a common autoimmune milieu. *Arq. Neuro-Psiquiatr.* **2014**, *72*, 619–624. [[CrossRef](#)]
72. Pelizza, L.; Bonacini, F.; Ferrari, A. Psychiatric disorder as clinical presentation of primary sjögren's syndrome: Two case reports. *Ann. Gen. Psychiatry* **2010**, *9*, 12. [[CrossRef](#)] [[PubMed](#)]
73. Wong, J.K.; Nortley, R.; Andrews, T.; D'Cruz, D. Case report: Psychiatric manifestations of primary sjögren's syndrome: A case report and literature review. *BMJ Case Rep.* **2014**. [[CrossRef](#)] [[PubMed](#)]
74. Caselli, R.J.; Scheithauer, B.W.; Bowles, C.A.; Trenerry, M.R.; Meyer, F.B.; Smigielski, J.S.; Rodriguez, M. The treatable dementia of sjögren's syndrome. *Ann. Neurol.* **1991**, *30*, 98–101. [[CrossRef](#)]
75. Escudero, D.; Latorre, P.; Codina, M.; Coll-Canti, J.; Coll, J. *Central Nervous System Disease in Sjögren's Syndrome*; Annales de Médecine Interne; Masson: Paris, France, 1995; pp. 239–242.
76. Alexander, E.L.; Provost, T.T.; Stevens, M.B.; Alexander, G.E. Neurologic complications of primary sjögren's syndrome. *Medicine* **1982**, *61*, 247–257. [[CrossRef](#)]
77. Lafitte, C.; Amoura, Z.; Cacoub, P.; Pradat-Diehl, P.; Picq, C.; Salachas, F.; Léger, J.-M.; Piette, J.C.; Delattre, J.Y. Neurological complications of primary sjögren's syndrome. *J. Neurol.* **2001**, *248*, 577–584. [[CrossRef](#)] [[PubMed](#)]
78. Gemignani, F.; Marbini, A.; Pavesi, G.; Di Vittorio, S.; Manganeli, P.; Cenacchi, G.; Mancina, D. Peripheral neuropathy associated with primary sjögren's syndrome. *J. Neurol. Neurosurg. Psychiatry* **1994**, *57*, 983–986. [[CrossRef](#)]
79. Govoni, M.; Bajocchi, G.; Rizzo, N.; Tola, M.; Caniatti, L.; Tugnoli, V.; Colamussi, P.; Trotta, F. Neurological involvement in primary sjögren's syndrome: Clinical and instrumental evaluation in a cohort of italian patients. *Clin. Rheumatol.* **1999**, *18*, 299–303. [[CrossRef](#)] [[PubMed](#)]

80. Lopate, G.; Pestronk, A.; Al-Lozi, M.; Lynch, T.; Florence, J.; Miller, T.; Levine, T.; Rampy, T.; Beson, B.; Ramneantu, I. Peripheral neuropathy in an outpatient cohort of patients with sjögren's syndrome. *Muscle Nerve* **2006**, *33*, 672–676. [[CrossRef](#)]
81. Kawashima, N.; Shindo, R.; Kohno, M. Primary sjögren's syndrome with subcortical dementia. *Intern. Med.* **1993**, *32*, 561–564. [[CrossRef](#)]
82. Teixeira, F.; Moreira, I.; Martins-Silva, A.; Vasconcelos, C.; Farinha, F.; Santos, E. Neurological involvement in primary sjögren's syndrome. *Acta Reumatol. Port.* **2013**, *38*, 29–36.
83. Michel, L.; Toulgoat, F.; Desal, H.; Laplaud, D.A.; Magot, A.; Hamidou, M.; Wiertlewski, S. Atypical Neurologic Complications in Patients with Primary Sjögren's Syndrome: Report of 4 Cases. *Semin. Arthritis Rheum.* **2011**, *40*, 338–342. [[CrossRef](#)] [[PubMed](#)]
84. Nițescu, D.; Nicolau, A.; Caraiola, S.; Predețeanu, D.; Ionescu, R.; Tănăsescu, C. Neuromyelitis optica—complication or comorbidity in primary sjögren's syndrome? *Rom. J. Intern. Med.* **2011**, *49*, 295–300. [[PubMed](#)]
85. Brito-Zeron, P.; Akasbi, M.; Bosch, X.; Bove, A.; Perez-De-Lis, M.; Diaz-Lagares, C.; Retamozo, S.; Gandia, M.; Perez-Alvarez, R.; Soto-Cardenas, M. Classification and characterisation of peripheral neuropathies in 102 patients with primary sjögren's syndrome. *Clin. Exp. Rheumatol.* **2013**, *31*, 103–110. [[PubMed](#)]
86. Liu, J.-Y.; Zhao, T.; Zhou, C.-K. Central nervous system involvement in primary sjogrens syndrome manifesting as multiple sclerosis. *Neurosciences* **2014**, *19*, 134–137. [[PubMed](#)]
87. Pavlakis, P.P.; Alexopoulos, H.; Kosmidis, M.L.; Stamboulis, E.; Routsias, J.G.; Tzartos, S.J.; Tzioufas, A.G.; Moutsopoulos, H.M.; Dalakas, M.C. Peripheral neuropathies in sjögren syndrome: A new reappraisal. *J. Neurol. Neurosurg. Psychiatry* **2011**, *82*, 798–802. [[CrossRef](#)] [[PubMed](#)]
88. Hasiloglu, Z.I.; Albayram, S.; Tasmali, K.; Erer, B.; Selcuk, H.; Islak, C. A case of primary sjögren's syndrome presenting primarily with central nervous system vasculitic involvement. *Rheumatol. Int.* **2012**, *32*, 805–807. [[CrossRef](#)]
89. Koh, J.H.; Kwok, S.-K.; Lee, J.; Park, S.-H. Autonomic dysfunction in primary sjogren's syndrome: A prospective cohort analysis of 154 korean patients. *Korean J. Intern. Med.* **2017**, *32*, 165. [[CrossRef](#)] [[PubMed](#)]
90. Konsta, O.D.; Thabet, Y.; Le Dantec, C.; Brooks, W.H.; Tzioufas, A.G.; Pers, J.O.; Renaudineau, Y. The contribution of epigenetics in Sjogren's Syndrome. *Front. Genet.* **2014**, *5*, 71. [[CrossRef](#)]
91. Thorlacius, G.E.; Wahren-Herlenius, M.; Ronnblom, L. An update on the role of type I interferons in systemic lupus erythematosus and Sjogren's syndrome. *Curr. Opin. Rheumatol.* **2018**, *30*, 471–481.
92. Shoenfeld, Y.; Agmon-Levin, N. 'ASIA'—Autoimmune/inflammatory syndrome induced by adjuvants. *J. Autoimmun.* **2011**, *36*, 4–8. [[CrossRef](#)]
93. Yoshida, T.; Sueyoshi, T.; Suwazono, S.; Kinjo, M.; Nodera, H. Detection of atrophy of dorsal root ganglion with 3-T magnetic resonance neurography in sensory ataxic neuropathy associated with Sjogren's syndrome. *Eur. J. Neurol.* **2018**, *25*, e78–e79. [[CrossRef](#)] [[PubMed](#)]
94. Lauvsnes, M.B.; Beyers, M.K.; Appenzeller, S.; Greve, O.J.; Harboe, E.; Goransson, L.G.; Tjensvoll, A.B.; Omdal, R. Loss of cerebral white matter in primary Sjogren's syndrome: A controlled volumetric magnetic resonance imaging study. *Eur. J. Neurol.* **2014**, *21*, 1324–1329. [[CrossRef](#)] [[PubMed](#)]
95. Mori, K.; Koike, H.; Misu, K.; Hattori, N.; Ichimura, M.; Sobue, G. Spinal cord magnetic resonance imaging demonstrates sensory neuronal involvement and clinical severity in neuronopathy associated with Sjogren's syndrome. *J. Neurol. Neurosurg. Psychiatry* **2001**, *71*, 488–492. [[CrossRef](#)] [[PubMed](#)]
96. Sullivan, D.A.; Wickham, L.A.; Rocha, E.M.; Krenzer, K.L.; Sullivan, B.D.; Steagall, R.; Cermak, J.M.; Dana, M.R.; Ullman, M.D.; Sato, E.H.; et al. Androgens and dry eye in Sjogren's syndrome. *Ann. N. Y. Acad. Sci.* **1999**, *876*, 312–324. [[CrossRef](#)]
97. Pavlakis, P.P.; Alexopoulos, H.; Kosmidis, M.L.; Mamali, I.; Moutsopoulos, H.M.; Tzioufas, A.G.; Dalakas, M.C. Peripheral neuropathies in Sjogren's syndrome: A critical update on clinical features and pathogenetic mechanisms. *J. Autoimmun.* **2012**, *39*, 27–33. [[CrossRef](#)]
98. Roberts, A.L.; Malspeis, S.; Kubzansky, L.D.; Feldman, C.H.; Chang, S.C.; Koenen, K.C.; Costenbader, K.H. Association of Trauma and Posttraumatic Stress Disorder With Incident Systemic Lupus Erythematosus in a Longitudinal Cohort of Women. *Arthritis Rheumatol.* **2017**, *69*, 2162–2169. [[CrossRef](#)]
99. Garcia-Carrasco, M.; Fuentes-Alexandro, S.; Escarcega, R.O.; Salgado, G.; Riebeling, C.; Cervera, R. Pathophysiology of Sjogren's syndrome. *Arch. Med. Res.* **2006**, *37*, 921–932. [[CrossRef](#)]

100. Li, H.; Reksten, T.R.; Ice, J.A.; Kelly, J.A.; Adrianto, I.; Rasmussen, A.; Wang, S.; He, B.; Grundahl, K.M.; Glenn, S.B.; et al. Identification of a Sjogren's syndrome susceptibility locus at OAS1 that influences isoform switching, protein expression, and responsiveness to type I interferons. *PLoS Genet.* **2017**, *13*, e1006820. [[CrossRef](#)]
101. Owada, K.; Uchihara, T.; Ishida, K.; Mizusawa, H.; Watabiki, S.; Tsuchiya, K. Motor weakness and cerebellar ataxia in Sjogren syndrome—identification of antineuronal antibody: A case report. *J. Neurol. Sci.* **2002**, *197*, 79–84. [[CrossRef](#)]
102. Rosler, D.H.; Conway, M.D.; Anaya, J.M.; Molina, J.F.; Carr, R.F.; Gharavi, A.E.; Wilson, W.A. Ischemic optic neuropathy and high-level anticardiolipin antibodies in primary Sjogren's syndrome. *Lupus* **1995**, *4*, 155–157. [[CrossRef](#)]
103. Rassi, D.M.; De Paiva, C.S.; Dias, L.C.; Modulo, C.M.; Adriano, L.; Fantucci, M.Z.; Rocha, E.M. MicroRNAs in ocular surface and dry eye diseases: Good, bad and healing situations. *Ocul. Surf.* **2017**. [[CrossRef](#)]
104. Josephs, K.A.; Rubino, F.A.; Dickson, D.W. Nonvasculitic autoimmune inflammatory meningoencephalitis. *Neuropathology* **2004**, *24*, 149–152. [[CrossRef](#)] [[PubMed](#)]
105. Rojewska, E.; Piotrowska, A.; Makuch, W.; Przewlocka, B.; Mika, J. Pharmacological kynurenine 3-monooxygenase enzyme inhibition significantly reduces neuropathic pain in a rat model. *Neuropharmacology* **2016**, *102*, 80–91. [[CrossRef](#)] [[PubMed](#)]
106. Akpek, E.K.; Mathews, P.; Hahn, S.; Hessen, M.; Kim, J.; Grader-Beck, T.; Birnbaum, J.; Baer, A.N. Ocular and systemic morbidity in a longitudinal cohort of Sjögren's syndrome. *Ophthalmology* **2015**, *122*, 56–61. [[CrossRef](#)] [[PubMed](#)]
107. Kawai, Y.; Sumi, M.; Kitamori, H.; Takagi, Y.; Nakamura, T. Diffusion-weighted MR microimaging of the lacrimal glands in patients with Sjogren's syndrome. *AJR Am. J. Roentgenol.* **2005**, *184*, 1320–1325. [[CrossRef](#)]
108. Demeter, I.; Nagy, K.; Farkas, T.; Kis, Z.; Kocsis, K.; Knapp, L.; Gellert, L.; Fulop, F.; Vecsei, L.; Toldi, J. Paradox effects of kynurenines on LTP induction in the Wistar rat. An in vivo study. *Neurosci. Lett.* **2013**, *553*, 138–141. [[CrossRef](#)] [[PubMed](#)]
109. Birnbaum, J.; Duncan, T.; Owoyemi, K.; Wang, K.C.; Carrino, J.; Chhabra, A. Use of a novel high-resolution magnetic resonance neurography protocol to detect abnormal dorsal root Ganglia in Sjogren patients with neuropathic pain: Case series of 10 patients and review of the literature. *Medicine* **2014**, *93*, 121–134. [[CrossRef](#)]
110. Saito, K.; Quearry, B.J.; Saito, M.; Nowak, T.S., Jr.; Markey, S.P.; Heyes, M.P. Kynurenine 3-hydroxylase in brain: Species activity differences and effect of gerbil cerebral ischemia. *Arch. Biochem. Biophys.* **1993**, *307*, 104–109. [[CrossRef](#)]
111. Kojima, I.; Sakamoto, M.; Iikubo, M.; Shimada, Y.; Nishioka, T.; Sasano, T. Relationship of MR imaging of submandibular glands to hyposalivation in Sjogren's syndrome. *Oral Dis.* **2018**. [[CrossRef](#)]
112. Kelleher, R.S.; Hann, L.E.; Edwards, J.A.; Sullivan, D.A. Endocrine, neural, and immune control of secretory component output by lacrimal gland acinar cells. *J. Immunol.* **1991**, *146*, 3405–3412.
113. Bacman, S.; Berra, A.; Sterin-Borda, L.; Borda, E. Muscarinic acetylcholine receptor antibodies as a new marker of dry eye Sjögren syndrome. *Investig. Ophthalmol. Vis. Sci.* **2001**, *42*, 321–327.
114. Tzioufas, A.G.; Tsonis, J.; Moutsopoulos, H.M. Neuroendocrine dysfunction in Sjogren's syndrome. *Neuroimmunomodulation* **2008**, *15*, 37–45. [[CrossRef](#)] [[PubMed](#)]
115. Wilder, R.L. Neuroendocrine-immune system interactions and autoimmunity. *Annu. Rev. Immunol.* **1995**, *13*, 307–338. [[CrossRef](#)] [[PubMed](#)]
116. Rosas-Ballina, M.; Olofsson, P.S.; Ochani, M.; Valdes-Ferrer, S.I.; Levine, Y.A.; Reardon, C.; Tusche, M.W.; Pavlov, V.A.; Andersson, U.; Chavan, S.; et al. Acetylcholine-synthesizing T cells relay neural signals in a vagus nerve circuit. *Science* **2011**, *334*, 98–101. [[CrossRef](#)] [[PubMed](#)]
117. Ueno, M.; Ueno-Nakamura, Y.; Niehaus, J.; Popovich, P.G.; Yoshida, Y. Silencing spinal interneurons inhibits immune suppressive autonomic reflexes caused by spinal cord injury. *Nat. Neurosci.* **2016**, *19*, 784–787. [[CrossRef](#)] [[PubMed](#)]
118. Mina-Osorio, P.; Rosas-Ballina, M.; Valdes-Ferrer, S.I.; Al-Abed, Y.; Tracey, K.J.; Diamond, B. Neural signaling in the spleen controls B-cell responses to blood-borne antigen. *Mol. Med.* **2012**, *18*, 618–627. [[CrossRef](#)] [[PubMed](#)]
119. Mirakaj, V.; Dalli, J.; Granja, T.; Rosenberger, P.; Serhan, C.N. Vagus nerve controls resolution and pro-resolving mediators of inflammation. *J. Exp. Med.* **2014**, *211*, 1037–1048. [[CrossRef](#)]

120. Chiu, I.M.; Heesters, B.A.; Ghasemlou, N.; Von Hehn, C.A.; Zhao, F.; Tran, J.; Wainger, B.; Strominger, A.; Muralidharan, S.; Horswill, A.R.; et al. Bacteria activate sensory neurons that modulate pain and inflammation. *Nature* **2013**, *501*, 52–57. [[CrossRef](#)]
121. Kumar, V.; Sur, V.P.; Guha, R.; Konar, A.; Hazra, S. Estrogen Modulates Corneal Nociception and Maintains Corneal Homeostasis in Rat Eye. *Cornea* **2017**. [[CrossRef](#)]
122. Rocha, E.; Wickham, L.; Huang, Z.; Toda, I.; Gao, J.; da Silveira, L.; Sullivan, D.; Dartt, D.; Meneray, M. Presence and testosterone influence on the levels of anti- and pro-inflammatory cytokines in lacrimal tissues of a mouse model of Sjogren's syndrome. In *Lacrimal Gland, Tear Film, and Dry Eye Syndromes 2*; Advances in Experimental Medicine and Biology; Springer: Boston, MA, USA, 1998; Volume 438, pp. 485–491.
123. Payrits, M.; Saghy, E.; Cseko, K.; Pohoczky, K.; Bolcskei, K.; Ernszt, D.; Barabas, K.; Szolcsanyi, J.; Abraham, I.M.; Helyes, Z.; et al. Estradiol Sensitizes the Transient Receptor Potential Vanilloid 1 Receptor in Pain Responses. *Endocrinology* **2017**, *158*, 3249–3258. [[CrossRef](#)]
124. Flake, N.M.; Bonebreak, D.B.; Gold, M.S. Estrogen and inflammation increase the excitability of rat temporomandibular joint afferent neurons. *J. Neurophysiol.* **2005**, *93*, 1585–1597. [[CrossRef](#)] [[PubMed](#)]
125. Wu, Y.W.; Kou, X.X.; Bi, R.Y.; Xu, W.; Wang, K.W.; Gan, Y.H.; Ma, X.C. Hippocampal nerve growth factor potentiated by 17beta-estradiol and involved in allodynia of inflamed TMJ in rat. *J. Pain* **2012**, *13*, 555–563. [[CrossRef](#)] [[PubMed](#)]
126. Bi, R.Y.; Meng, Z.; Zhang, P.; Wang, X.D.; Ding, Y.; Gan, Y.H. Estradiol upregulates voltage-gated sodium channel 1.7 in trigeminal ganglion contributing to hyperalgesia of inflamed TMJ. *PLoS ONE* **2017**, *12*, e0178589. [[CrossRef](#)] [[PubMed](#)]
127. Jin, X.; Wang, B.H.; Wang, X.; Antony, B.; Zhu, Z.; Han, W.; Cicuttini, F.; Wluka, A.E.; Winzenberg, T.; Blizzard, L.; et al. Associations between endogenous sex hormones and MRI structural changes in patients with symptomatic knee osteoarthritis. *Osteoarthr. Cartil.* **2017**, *25*, 1100–1106. [[CrossRef](#)] [[PubMed](#)]
128. Sullivan, D.A.; Belanger, A.; Cermak, J.M.; Berube, R.; Papas, A.S.; Sullivan, R.M.; Yamagami, H.; Dana, M.R.; Labrie, F. Are women with Sjogren's syndrome androgen-deficient? *J. Rheumatol.* **2003**, *30*, 2413–2419.
129. Sullivan, D.A. Sex hormones and Sjogren's syndrome. *J. Rheumatol. Suppl.* **1997**, *50*, 17–32.
130. Taiym, S.; Haghghat, N.; Al-Hashimi, I. A comparison of the hormone levels in patients with Sjogren's syndrome and healthy controls. *Oral Surg. Oral Med. Oral Pathol. Oral Radiol. Endod.* **2004**, *97*, 579–583. [[CrossRef](#)]
131. Woller, S.A.; Eddinger, K.A.; Corr, M.; Yaksh, T.L. An overview of pathways encoding nociception. *Clin. Exp. Rheumatol.* **2017**, *35*, 40–46.
132. Rosenthal, P.; Borsook, D. Ocular neuropathic pain. *Br. J. Ophthalmol.* **2016**, *100*, 128–134. [[CrossRef](#)]
133. Sandhya, P.; Jeyaseelan, L.; Scofield, R.H.; Danda, D. Clinical Characteristics and Outcome of Primary Sjogren's Syndrome: A Large Asian Indian Cohort. *Open Rheumatol. J.* **2015**, *9*, 36–45. [[CrossRef](#)]
134. Ruddick, J.P.; Evans, A.K.; Nutt, D.J.; Lightman, S.L.; Rook, G.A.; Lowry, C.A. Tryptophan metabolism in the central nervous system: Medical implications. *Expert Rev. Mol. Med.* **2006**, *8*, 1–27. [[CrossRef](#)] [[PubMed](#)]
135. Nakamura, T.; Shinno, H.; Ichihara, A. Insulin and glucagon as a new regulator system for tryptophan oxygenase activity demonstrated in primary cultured rat hepatocytes. *J. Biol. Chem.* **1980**, *255*, 7533–7535. [[PubMed](#)]
136. Hayaishi, O.; Yoshida, R. Specific induction of pulmonary indoleamine 2,3-dioxygenase by bacterial lipopolysaccharide. *Ciba Found. Symp.* **1978**, 199–203.
137. Heyes, M.P.; Saito, K.; Milstien, S.; Schiff, S.J. Quinolinic acid in tumors, hemorrhage and bacterial infections of the central nervous system in children. *J. Neurol. Sci.* **1995**, *133*, 112–118. [[CrossRef](#)]
138. Hayaishi, O. Properties and function of indoleamine 2,3-dioxygenase. *J. Biochem.* **1976**, *79*, 13P–21P. [[CrossRef](#)]
139. Wolf, H. The effect of hormones and vitamin B6 on urinary excretion of metabolites of the kynurenine pathway. *Scand. J. Clin. Lab. Investig. Suppl.* **1974**, *136*, 1–186.
140. Colin-Gonzalez, A.L.; Maldonado, P.D.; Santamaria, A. 3-Hydroxykynurenine: An intriguing molecule exerting dual actions in the central nervous system. *Neurotoxicology* **2013**, *34*, 189–204. [[CrossRef](#)] [[PubMed](#)]
141. O'Farrell, K.; Fagan, E.; Connor, T.J.; Harkin, A. Inhibition of the kynurenine pathway protects against reactive microglial-associated reductions in the complexity of primary cortical neurons. *Eur. J. Pharm.* **2017**, *810*, 163–173. [[CrossRef](#)]

142. Chiarugi, A.; Cozzi, A.; Ballerini, C.; Massacesi, L.; Moroni, F. Kynurenine 3-mono-oxygenase activity and neurotoxic kynurenine metabolites increase in the spinal cord of rats with experimental allergic encephalomyelitis. *Neuroscience* **2001**, *102*, 687–695. [[CrossRef](#)]
143. Munn, D.H.; Shafizadeh, E.; Attwood, J.T.; Bondarev, I.; Pashine, A.; Mellor, A.L. Inhibition of T cell proliferation by macrophage tryptophan catabolism. *J. Exp. Med.* **1999**, *189*, 1363–1372. [[CrossRef](#)]
144. Munn, D.H.; Zhou, M.; Attwood, J.T.; Bondarev, I.; Conway, S.J.; Marshall, B.; Brown, C.; Mellor, A.L. Prevention of allogeneic fetal rejection by tryptophan catabolism. *Science* **1998**, *281*, 1191–1193. [[CrossRef](#)] [[PubMed](#)]
145. Puccetti, P.; Grohmann, U. IDO and regulatory T cells: A role for reverse signalling and non-canonical NF-kappaB activation. *Nat. Rev. Immunol.* **2007**, *7*, 817–823. [[CrossRef](#)] [[PubMed](#)]
146. Mellor, A.L.; Munn, D.H. IDO expression by dendritic cells: Tolerance and tryptophan catabolism. *Nat. Rev. Immunol.* **2004**, *4*, 762–774. [[CrossRef](#)] [[PubMed](#)]
147. Munn, D.H.; Armstrong, E. Cytokine regulation of human monocyte differentiation in vitro: The tumor-cytotoxic phenotype induced by macrophage colony-stimulating factor is developmentally regulated by gamma-interferon. *Cancer Res.* **1993**, *53*, 2603–2613. [[PubMed](#)]
148. Heyes, M.P.; Saito, K.; Jacobowitz, D.; Markey, S.P.; Takikawa, O.; Vickers, J.H. Poliovirus induces indoleamine-2,3-dioxygenase and quinolinic acid synthesis in macaque brain. *FASEB J.* **1992**, *6*, 2977–2989. [[CrossRef](#)] [[PubMed](#)]
149. Saito, K.; Lackner, A.; Markey, S.P.; Heyes, M.P. Cerebral cortex and lung indoleamine-2,3-dioxygenase activity is increased in type-D retrovirus infected macaques. *Brain Res.* **1991**, *540*, 353–356. [[CrossRef](#)]
150. Saito, K.; Markey, S.P.; Heyes, M.P. Chronic effects of gamma-interferon on quinolinic acid and indoleamine-2,3-dioxygenase in brain of C57BL6 mice. *Brain Res.* **1991**, *546*, 151–154. [[CrossRef](#)]
151. Sayama, S.; Yoshida, R.; Oku, T.; Imanishi, J.; Kishida, T.; Hayaishi, O. Inhibition of interferon-mediated induction of indoleamine 2,3-dioxygenase in mouse lung by inhibitors of prostaglandin biosynthesis. *Proc. Natl. Acad. Sci. USA* **1981**, *78*, 7327–7330. [[CrossRef](#)]
152. Yoshida, R.; Urade, Y.; Nakata, K.; Watanabe, Y.; Hayaishi, O. Specific induction of indoleamine 2,3-dioxygenase by bacterial lipopolysaccharide in the mouse lung. *Arch. Biochem. Biophys.* **1981**, *212*, 629–637. [[CrossRef](#)]
153. Pertovaara, M.; Raitala, A.; Uusitalo, H.; Pukander, J.; Helin, H.; Oja, S.S.; Hurme, M. Mechanisms dependent on tryptophan catabolism regulate immune responses in primary Sjogren’s syndrome. *Clin. Exp. Immunol.* **2005**, *142*, 155–161. [[CrossRef](#)]
154. Campbell, D.J.; Koch, M.A. Treg cells: Patrolling a dangerous neighborhood. *Nat. Med.* **2011**, *17*, 929–930. [[CrossRef](#)] [[PubMed](#)]
155. Jaspersen, L.K.; Bucher, C.; Panoskaltis-Mortari, A.; Mellor, A.L.; Munn, D.H.; Blazar, B.R. Inducing the tryptophan catabolic pathway, indoleamine 2,3-dioxygenase (IDO), for suppression of graft-versus-host disease (GVHD) lethality. *Blood* **2009**, *114*, 5062–5070. [[CrossRef](#)] [[PubMed](#)]
156. Nezos, A.; Gravani, F.; Tassidou, A.; Kapsogeorgou, E.K.; Voulgarelis, M.; Koutsilieris, M.; Crow, M.K.; Mavragani, C.P. Type I and II interferon signatures in Sjogren’s syndrome pathogenesis: Contributions in distinct clinical phenotypes and Sjogren’s related lymphomagenesis. *J. Autoimmun.* **2015**, *63*, 47–58. [[CrossRef](#)] [[PubMed](#)]
157. Raitala, A.; Pertovaara, M.; Karjalainen, J.; Oja, S.S.; Hurme, M. Association of interferon-gamma +874(T/A) single nucleotide polymorphism with the rate of tryptophan catabolism in healthy individuals. *Scand. J. Immunol.* **2005**, *61*, 387–390. [[CrossRef](#)] [[PubMed](#)]
158. Silver, R.M.; McKinley, K.; Smith, E.A.; Quearry, B.; Harati, Y.; Sternberg, E.M.; Heyes, M.P. Tryptophan metabolism via the kynurenine pathway in patients with the eosinophilia-myalgia syndrome. *Arthritis Rheum.* **1992**, *35*, 1097–1105. [[CrossRef](#)] [[PubMed](#)]
159. Barbosa, F.L.; Xiao, Y.; Bian, F.; Coursey, T.G.; Ko, B.Y.; Clevers, H.; de Paiva, C.S.; Pflugfelder, S.C. Goblet Cells Contribute to Ocular Surface Immune Tolerance-Implications for Dry Eye Disease. *Int. J. Mol. Sci.* **2017**, *18*, 978. [[CrossRef](#)]
160. Baboonian, C.; Venables, P.J.; Booth, J.; Williams, D.G.; Roffe, L.M.; Maini, R.N. Virus infection induces redistribution and membrane localization of the nuclear antigen La (SS-B): A possible mechanism for autoimmunity. *Clin. Exp. Immunol.* **1989**, *78*, 454–459.

161. Brkic, Z.; Maria, N.I.; van Helden-Meeuwsen, C.G.; van de Merwe, J.P.; van Daele, P.L.; Dalm, V.A.; Wildenberg, M.E.; Beumer, W.; Drexhage, H.A.; Versnel, M.A. Prevalence of interferon type I signature in CD14 monocytes of patients with Sjogren's syndrome and association with disease activity and BAFF gene expression. *Ann. Rheum. Dis.* **2013**, *72*, 728–735. [[CrossRef](#)]
162. Weller, M.L.; Gardener, M.R.; Bogus, Z.C.; Smith, M.A.; Astorri, E.; Michael, D.G.; Michael, D.A.; Zheng, C.; Burbelo, P.D.; Lai, Z.; et al. Hepatitis Delta Virus Detected in Salivary Glands of Sjogren's Syndrome Patients and Recapitulates a Sjogren's Syndrome-Like Phenotype in Vivo. *Pathog. Immun.* **2016**, *1*, 12–40. [[CrossRef](#)]
163. Prendergast, G.C.; Metz, R.; Muller, A.J.; Merlo, L.M.; Mandik-Nayak, L. IDO2 in Immunomodulation and Autoimmune Disease. *Front. Immunol.* **2014**, *5*, 585. [[CrossRef](#)]
164. Murakami, Y.; Saito, K. Species and cell types difference in tryptophan metabolism. *Int. J. Tryptophan Res.* **2013**, *6*, 47–54. [[CrossRef](#)] [[PubMed](#)]
165. Maes, M.; De Ruyter, M.; Hobin, P.; Suy, E. Relationship between the dexamethasone suppression test and the L-tryptophan/competing amino acids ratio in depression. *Psychiatry Res.* **1987**, *21*, 323–335. [[CrossRef](#)]
166. Maes, M.; Meltzer, H.Y.; Scharpe, S.; Bosmans, E.; Suy, E.; De Meester, I.; Calabrese, J.; Cosyns, P. Relationships between lower plasma L-tryptophan levels and immune-inflammatory variables in depression. *Psychiatry Res.* **1993**, *49*, 151–165. [[CrossRef](#)]
167. Schwarcz, R.; Pellicciari, R. Manipulation of brain kynurenes: Glial targets, neuronal effects, and clinical opportunities. *J. Pharm. Exp.* **2002**, *303*, 1–10. [[CrossRef](#)] [[PubMed](#)]
168. Bonaccorso, S.; Marino, V.; Puzella, A.; Pasquini, M.; Biondi, M.; Artini, M.; Almerighi, C.; Verkerk, R.; Meltzer, H.; Maes, M. Increased depressive ratings in patients with hepatitis C receiving interferon-alpha-based immunotherapy are related to interferon-alpha-induced changes in the serotonergic system. *J. Clin. Psychopharmacol.* **2002**, *22*, 86–90. [[CrossRef](#)] [[PubMed](#)]
169. Capuron, L.; Ravaud, A.; Neveu, P.J.; Miller, A.H.; Maes, M.; Dantzer, R. Association between decreased serum tryptophan concentrations and depressive symptoms in cancer patients undergoing cytokine therapy. *Mol. Psychiatry* **2002**, *7*, 468–473. [[CrossRef](#)] [[PubMed](#)]
170. Myint, A.M.; Bondy, B.; Baghai, T.C.; Eser, D.; Nothdurfter, C.; Schule, C.; Zill, P.; Muller, N.; Rupprecht, R.; Schwarz, M.J. Tryptophan metabolism and immunogenetics in major depression: A role for interferon-gamma gene. *Brain Behav. Immun.* **2013**, *31*, 128–133. [[CrossRef](#)] [[PubMed](#)]
171. Myint, A.M.; Kim, Y.K. Network beyond IDO in psychiatric disorders: Revisiting neurodegeneration hypothesis. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **2014**, *48*, 304–313. [[CrossRef](#)] [[PubMed](#)]
172. O'Farrell, K.; Harkin, A. Stress-related regulation of the kynurenine pathway: Relevance to neuropsychiatric and degenerative disorders. *Neuropharmacology* **2017**, *112*, 307–323. [[CrossRef](#)]
173. Kennedy, P.J.; Cryan, J.F.; Dinan, T.G.; Clarke, G. Kynurenine pathway metabolism and the microbiota-gut-brain axis. *Neuropharmacology* **2017**, *112*, 399–412. [[CrossRef](#)]
174. DeLuca, J.A.; Allred, K.F.; Menon, R.; Riordan, R.; Weeks, B.R.; Jayaraman, A.; Allred, C.D. Bisphenol-A alters microbiota metabolites derived from aromatic amino acids and worsens disease activity during colitis. *Exp. Biol. Med.* **2018**, *243*, 864–875. [[CrossRef](#)] [[PubMed](#)]
175. Hertzman, P.A.; Blevins, W.L.; Mayer, J.; Greenfield, B.; Ting, M.; Gleich, G.J. Association of the eosinophilia-myalgia syndrome with the ingestion of tryptophan. *N. Engl. J. Med.* **1990**, *322*, 869–873. [[CrossRef](#)] [[PubMed](#)]
176. Young, S.N.; Smith, S.E.; Pihl, R.O.; Ervin, F.R. Tryptophan depletion causes a rapid lowering of mood in normal males. *Psychopharmacology* **1985**, *87*, 173–177. [[CrossRef](#)] [[PubMed](#)]
177. Guillemain, G.J.; Kerr, S.J.; Smythe, G.A.; Smith, D.G.; Kapoor, V.; Armati, P.J.; Croitoru, J.; Brew, B.J. Kynurenine pathway metabolism in human astrocytes: A paradox for neuronal protection. *J. Neurochem.* **2001**, *78*, 842–853. [[CrossRef](#)] [[PubMed](#)]
178. Guillemain, G.J.; Williams, K.R.; Smith, D.G.; Smythe, G.A.; Croitoru-Lamoury, J.; Brew, B.J. Quinolinic acid in the pathogenesis of Alzheimer's disease. *Adv. Exp. Med. Biol.* **2003**, *527*, 167–176. [[PubMed](#)]
179. Guillemain, G.J. Quinolinic acid, the inescapable neurotoxin. *FEBS J.* **2012**, *279*, 1356–1365. [[CrossRef](#)] [[PubMed](#)]
180. Diamond, B. Antibodies and the Brain: Lessons from Lupus. *J. Immunol.* **2010**, *185*, 2637–2640. [[CrossRef](#)] [[PubMed](#)]

181. Faust, T.W.; Chang, E.H.; Kowal, C.; Berlin, R.; Gazaryan, I.G.; Bertini, E.; Zhang, J.; Sanchez-Guerrero, J.; Fragoso-Loyo, H.E.; Volpe, B.T.; et al. Neurotoxic lupus autoantibodies alter brain function through two distinct mechanisms. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 18569–18574. [[CrossRef](#)]
182. Eastman, C.L.; Guilarte, T.R. Cytotoxicity of 3-hydroxykynurenine in a neuronal hybrid cell line. *Brain Res.* **1989**, *495*, 225–231. [[CrossRef](#)]
183. Eastman, C.L.; Guilarte, T.R.; Lever, J.R. Uptake of 3-hydroxykynurenine measured in rat brain slices and in a neuronal cell line. *Brain Res.* **1992**, *584*, 110–116. [[CrossRef](#)]
184. Schwarcz, R.; Stone, T.W. The kynurenine pathway and the brain: Challenges, controversies and promises. *Neuropharmacology* **2017**, *112*, 237–247. [[CrossRef](#)] [[PubMed](#)]
185. Shigemoto, R.; Nakanishi, S.; Mizuno, N. Distribution of the mRNA for a metabotropic glutamate receptor (mGluR1) in the central nervous system: An in situ hybridization study in adult and developing rat. *J. Comp. Neurol.* **1992**, *322*, 121–135. [[CrossRef](#)] [[PubMed](#)]
186. Arnone, D.; Job, D.; Selvaraj, S.; Abe, O.; Amico, F.; Cheng, Y.; Colloby, S.J.; O'Brien, J.T.; Frodl, T.; Gotlib, I.H.; et al. Computational meta-analysis of statistical parametric maps in major depression. *Hum. Brain Mapp.* **2016**, *37*, 1393–1404. [[CrossRef](#)] [[PubMed](#)]
187. Castle, M.; Comoli, E.; Loewy, A.D. Autonomic brainstem nuclei are linked to the hippocampus. *Neuroscience* **2005**, *134*, 657–669. [[CrossRef](#)] [[PubMed](#)]
188. Petralia, R.S.; Wang, Y.X.; Wenthold, R.J. The NMDA receptor subunits NR2A and NR2B show histological and ultrastructural localization patterns similar to those of NR1. *J. Neurosci.* **1994**, *14*, 6102–6120. [[CrossRef](#)] [[PubMed](#)]
189. Petrenko, A.B.; Yamakura, T.; Baba, H.; Sakimura, K. Unaltered pain-related behavior in mice lacking NMDA receptor GluRepsilon 1 subunit. *NeuroSci. Res.* **2003**, *46*, 199–204. [[CrossRef](#)]
190. Kim, H.; Chen, L.; Lim, G.; Sung, B.; Wang, S.; McCabe, M.F.; Rusanescu, G.; Yang, L.; Tian, Y.; Mao, J. Brain indoleamine 2,3-dioxygenase contributes to the comorbidity of pain and depression. *J. Clin. Investig.* **2012**, *122*, 2940–2954. [[CrossRef](#)]
191. Petrenko, A.B.; Yamakura, T.; Baba, H.; Shimoji, K. The role of N-methyl-D-aspartate (NMDA) receptors in pain: A review. *Anesth. Analg.* **2003**, *97*, 1108–1116. [[CrossRef](#)]
192. Karageorgas, T.; Fragioudaki, S.; Nezos, A.; Karaiskos, D.; Moutsopoulos, H.M.; Mavragani, C.P. Fatigue in Primary Sjogren's Syndrome: Clinical, Laboratory, Psychometric, and Biologic Associations. *Arthritis Care Res.* **2016**, *68*, 123–131. [[CrossRef](#)]
193. Bengtsson, A.A.; Trygg, J.; Wuttge, D.M.; Sturfelt, G.; Theander, E.; Donten, M.; Moritz, T.; Sennbro, C.J.; Torell, F.; Lood, C.; et al. Metabolic Profiling of Systemic Lupus Erythematosus and Comparison with Primary Sjogren's Syndrome and Systemic Sclerosis. *PLoS ONE* **2016**, *11*, e0159384. [[CrossRef](#)]
194. McEwen, B.S. Plasticity of the hippocampus: Adaptation to chronic stress and allostatic load. *Ann. N. Y. Acad. Sci.* **2001**, *933*, 265–277. [[CrossRef](#)] [[PubMed](#)]
195. Brake, W.G.; Alves, S.E.; Dunlop, J.C.; Lee, S.J.; Bulloch, K.; Allen, P.B.; Greengard, P.; McEwen, B.S. Novel target sites for estrogen action in the dorsal hippocampus: An examination of synaptic proteins. *Endocrinology* **2001**, *142*, 1284–1289. [[CrossRef](#)] [[PubMed](#)]
196. McEwen, B.S. Sex, stress and the hippocampus: Allostasis, allostatic load and the aging process. *Neurobiol. Aging* **2002**, *23*, 921–939. [[CrossRef](#)]
197. Woolley, C.S.; Weiland, N.G.; McEwen, B.S.; Schwartzkroin, P.A. Estradiol increases the sensitivity of hippocampal CA1 pyramidal cells to NMDA receptor-mediated synaptic input: Correlation with dendritic spine density. *J. Neurosci.* **1997**, *17*, 1848–1859. [[CrossRef](#)] [[PubMed](#)]
198. Arnone, D.; Saraykar, S.; Salem, H.; Teixeira, A.L.; Dantzer, R.; Selvaraj, S. Role of Kynurenine pathway and its metabolites in mood disorders: A systematic review and meta-analysis of clinical studies. *NeuroSci. Biobehav. Rev.* **2018**, *92*, 477–485. [[CrossRef](#)] [[PubMed](#)]
199. Dostal, C.R.; Gamsby, N.S.; Lawson, M.A.; McCusker, R.H. Glia- and tissue-specific changes in the Kynurenine Pathway after treatment of mice with lipopolysaccharide and dexamethasone. *Brain Behav. Immun.* **2018**, *69*, 321–335. [[CrossRef](#)] [[PubMed](#)]
200. Zhuo, M. Ionotropic glutamate receptors contribute to pain transmission and chronic pain. *Neuropharmacology* **2017**, *112*, 228–234. [[CrossRef](#)]
201. Zhuo, M. Silent glutamatergic synapses and long-term facilitation in spinal dorsal horn neurons. *Prog. Brain Res.* **2000**, *129*, 101–113.

202. Ciranna, L. Serotonin as a modulator of glutamate- and GABA-mediated neurotransmission: Implications in physiological functions and in pathology. *Curr. Neuropharmacol.* **2006**, *4*, 101–114. [[CrossRef](#)]
203. Gandolfi, O.; Gaggi, R.; Voltattorni, M.; Dall'Olio, R. The activation of serotonin receptors prevents glutamate-induced neurotoxicity and NMDA-stimulated cGMP accumulation in primary cortical cell cultures. *Pharm. Res.* **2002**, *46*, 409–414. [[CrossRef](#)]
204. Gandolfi, O.; Dall'Olio, R.; Roncada, P.; Montanaro, N. NMDA antagonists interact with 5-HT-stimulated phosphatidylinositol metabolism and impair passive avoidance retention in the rat. *Neurosci. Lett.* **1990**, *113*, 304–308. [[CrossRef](#)]
205. Maura, G.; Marcoli, M.; Pepicelli, O.; Rosu, C.; Viola, C.; Raiteri, M. Serotonin inhibition of the NMDA receptor/nitric oxide/cyclic GMP pathway in human neocortex slices: Involvement of 5-HT(2C) and 5-HT(1A) receptors. *Br. J. Pharm.* **2000**, *130*, 1853–1858. [[CrossRef](#)] [[PubMed](#)]
206. Maura, G.; Raiteri, M. Serotonin 5-HT1D and 5-HT1A receptors respectively mediate inhibition of glutamate release and inhibition of cyclic GMP production in rat cerebellum in vitro. *J. Neurochem.* **1996**, *66*, 203–209. [[CrossRef](#)] [[PubMed](#)]
207. Schmitz, D.; Gloveli, T.; Empson, R.M.; Heinemann, U. Comparison of the effects of serotonin in the hippocampus and the entorhinal cortex. *Mol. Neurobiol.* **1998**, *17*, 59–72. [[CrossRef](#)]
208. Dostal, C.R.; Carson Sulzer, M.; Kelley, K.W.; Freund, G.G.; McCusker, R.H. Glial and tissue-specific regulation of Kynurenine Pathway dioxygenases by acute stress of mice. *Neurobiol. Stress* **2017**, *7*, 1–15. [[CrossRef](#)] [[PubMed](#)]
209. Weiland, N.G.; Orchinik, M.; Tanapat, P. Chronic corticosterone treatment induces parallel changes in N-methyl-D-aspartate receptor subunit messenger RNA levels and antagonist binding sites in the hippocampus. *Neuroscience* **1997**, *78*, 653–662. [[CrossRef](#)]
210. Heyes, M.P.; Saito, K.; Lackner, A.; Wiley, C.A.; Achim, C.L.; Markey, S.P. Sources of the neurotoxin quinolinic acid in the brain of HIV-1-infected patients and retrovirus-infected macaques. *FASEB J.* **1998**, *12*, 881–896. [[CrossRef](#)] [[PubMed](#)]
211. Parrott, J.M.; Redus, L.; O'Connor, J.C. Kynurenine metabolic balance is disrupted in the hippocampus following peripheral lipopolysaccharide challenge. *J. Neuroinflamm.* **2016**, *13*, 124. [[CrossRef](#)]
212. Sternberg, E.M.; Van Woert, M.H.; Young, S.N.; Magnussen, I.; Baker, H.; Gauthier, S.; Osterland, C.K. Development of a scleroderma-like illness during therapy with L-5-hydroxytryptophan and carbidopa. *N. Engl. J. Med.* **1980**, *303*, 782–787. [[CrossRef](#)]
213. Christensen, L.; Redig, C. Effect of meal composition on mood. *Behav. Neurosci.* **1993**, *107*, 346–353. [[CrossRef](#)]
214. Fernstrom, M.H.; Fernstrom, J.D. Brain tryptophan concentrations and serotonin synthesis remain responsive to food consumption after the ingestion of sequential meals. *Am. J. Clin. Nutr.* **1995**, *61*, 312–319. [[CrossRef](#)] [[PubMed](#)]
215. Kan, H.; London, S.J.; Chen, G.; Zhang, Y.; Song, G.; Zhao, N.; Jiang, L.; Chen, B. Season, sex, age, and education as modifiers of the effects of outdoor air pollution on daily mortality in Shanghai, China: The Public Health and Air Pollution in Asia (PAPA) Study. *Environ. Health Perspect.* **2008**, *116*, 1183–1188. [[CrossRef](#)] [[PubMed](#)]
216. Furuzawa-Carballeda, J.; Hernandez-Molina, G.; Lima, G.; Rivera-Vicencio, Y.; Ferez-Blando, K.; Llorente, L. Peripheral regulatory cells immunophenotyping in primary Sjogren's syndrome: A cross-sectional study. *Arthritis Res. Ther.* **2013**, *15*, R68. [[CrossRef](#)] [[PubMed](#)]
217. Legany, N.; Berta, L.; Kovacs, L.; Balog, A.; Toldi, G. The role of B7 family costimulatory molecules and indoleamine 2,3-dioxygenase in primary Sjogren's syndrome and systemic sclerosis. *Immunol. Res.* **2017**, *65*, 622–629. [[CrossRef](#)] [[PubMed](#)]
218. James, K.; Al-Ali, S.; Tarn, J.; Cockell, S.J.; Gillespie, C.S.; Hindmarsh, V.; Locke, J.; Mitchell, S.; Lendrem, D.; Bowman, S.; et al. A Transcriptional Signature of Fatigue Derived from Patients with Primary Sjogren's Syndrome. *PLoS ONE* **2015**, *10*, e0143970. [[CrossRef](#)] [[PubMed](#)]
219. Hall, J.C.; Baer, A.N.; Shah, A.A.; Criswell, L.A.; Shiboski, C.H.; Rosen, A.; Casciola-Rosen, L. Molecular Subsetting of Interferon Pathways in Sjogren's Syndrome. *Arthritis Rheumatol.* **2015**, *67*, 2437–2446. [[CrossRef](#)] [[PubMed](#)]
220. Valim, V.; Zandonade, E.; Brun, J.G.; Jonsson, R.; Ueland, P.; Mydel, P.M. Kynurenines pathway biomarkers for primary Sjögren's syndrome. *Clin. Exp. Rheumatol.* **2018**, *36*, S-290.

221. Ter Borg, E.J.; Kelder, J.C. Is extra-glandular organ damage in primary Sjogren's syndrome related to the presence of systemic auto-antibodies and/or hypergammaglobulinemia? A long-term cohort study with 110 patients from the Netherlands. *Int. J. Rheum. Dis.* **2017**, *20*, 875–881. [[CrossRef](#)]
222. Chiche, L.; Jourde-Chiche, N.; Whalen, E.; Presnell, S.; Gersuk, V.; Dang, K.; Anguiano, E.; Quinn, C.; Burtey, S.; Berland, Y.; et al. Modular Transcriptional Repertoire Analyses of Adults With Systemic Lupus Erythematosus Reveal Distinct Type I and Type II Interferon Signatures. *Arthritis Rheumatol.* **2014**, *66*, 1583–1595. [[CrossRef](#)]
223. Kivity, S.; Katzav, A.; Arango, M.T.; Landau-Rabi, M.; Zafir, Y.; Agmon-Levin, N.; Blank, M.; Anaya, J.M.; Mozes, E.; Chapman, J.; et al. 16/6-idiotype expressing antibodies induce brain inflammation and cognitive impairment in mice: The mosaic of central nervous system involvement in lupus. *BMC Med.* **2013**, *11*, 90. [[CrossRef](#)]
224. Mariette, X.; Roux, S.; Zhang, J.; Bengoufa, D.; Lavie, F.; Zhou, T.; Kimberly, R. The level of BLYS (BAFF) correlates with the titre of autoantibodies in human Sjogren's syndrome. *Ann. Rheum. Dis.* **2003**, *62*, 168–171. [[CrossRef](#)] [[PubMed](#)]
225. Gottenberg, J.E.; Cagnard, N.; Lucchesi, C.; Letourneur, F.; Mistou, S.; Lazure, T.; Jacques, S.; Ba, N.; Ittah, M.; Lepajolec, C.; et al. Activation of IFN pathways and plasmacytoid dendritic cell recruitment in target organs of primary Sjogren's syndrome. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 2770–2775. [[CrossRef](#)] [[PubMed](#)]
226. Grisius, M.M.; Bermudez, D.K.; Fox, P.C. Salivary and serum interleukin 6 in primary Sjogren's syndrome. *J. Rheumatol.* **1997**, *24*, 1089–1091. [[PubMed](#)]
227. Der, S.D.; Zhou, A.; Williams, B.R.; Silverman, R.H. Identification of genes differentially regulated by interferon alpha, beta, or gamma using oligonucleotide arrays. *Proc. Natl. Acad. Sci. USA* **1998**, *95*, 15623–15628. [[CrossRef](#)] [[PubMed](#)]
228. Quartuccio, L.; Salvin, S.; Fabris, M.; Maset, M.; Pontarini, E.; Isola, M.; De Vita, S. BLYS upregulation in Sjogren's syndrome associated with lymphoproliferative disorders, higher ESSDAI score and B-cell clonal expansion in the salivary glands. *Rheumatology* **2013**, *52*, 276–281. [[CrossRef](#)] [[PubMed](#)]
229. Akpek, E.K.; Lindsley, K.B.; Adyanthaya, R.S.; Swamy, R.; Baer, A.N.; McDonnell, P.J. Treatment of Sjögren's syndrome-associated dry eye an evidence-based review. *Ophthalmology* **2011**, *118*, 1242–1252. [[PubMed](#)]
230. Bowman, S.J. Biologic therapies in primary Sjögren's syndrome. *Curr. Pharm. Biotechnol* **2012**, *13*, 1997–2008. [[CrossRef](#)]
231. Ramos-Casals, M.; Brito-Zerón, P.; Sisó-Almirall, A.; Bosch, X.; Tzioufas, A.G. Topical and systemic medications for the treatment of primary Sjögren's syndrome. *Nat. Rev. Rheumatol.* **2012**, *8*, 399–411. [[CrossRef](#)]
232. Both, T.; Dalm, V.A.; van Hagen, P.M.; van Daele, P.L. Reviewing primary Sjogren's syndrome: Beyond the dryness—From pathophysiology to diagnosis and treatment. *Int. J. Med. Sci.* **2017**, *14*, 191–200. [[CrossRef](#)]
233. Sthoeger, Z.; Sharabi, A.; Asher, I.; Zinger, H.; Segal, R.; Shearer, G.; Elkayam, O.; Mozes, E. The tolerogenic peptide hCDR1 immunomodulates cytokine and regulatory molecule gene expression in blood mononuclear cells of primary Sjogren's syndrome patients. *Clin. Immunol.* **2018**, *192*, 85–91. [[CrossRef](#)]
234. Chiarugi, A.; Carpenedo, R.; Molina, M.T.; Mattoli, L.; Pellicciari, R.; Moroni, F. Comparison of the neurochemical and behavioral effects resulting from the inhibition of kynurenine hydroxylase and/or kynureninase. *J. Neurochem.* **1995**, *65*, 1176–1183. [[CrossRef](#)] [[PubMed](#)]
235. Carpenedo, R.; Chiarugi, A.; Russi, P.; Lombardi, G.; Carla, V.; Pellicciari, R.; Mattoli, L.; Moroni, F. Inhibitors of kynurenine hydroxylase and kynureninase increase cerebral formation of kynurenate and have sedative and anticonvulsant activities. *Neuroscience* **1994**, *61*, 237–244. [[CrossRef](#)]
236. Pellicciari, R.; Natalini, B.; Costantino, G.; Mahmoud, M.R.; Mattoli, L.; Sadeghpour, B.M.; Moroni, F.; Chiarugi, A.; Carpenedo, R. Modulation of the kynurenine pathway in search for new neuroprotective agents. Synthesis and preliminary evaluation of (m-nitrobenzoyl) alanine, a potent inhibitor of kynurenine-3-hydroxylase. *J. Med. Chem.* **1994**, *37*, 647–655.
237. Kocki, T.; Luchowski, P.; Luchowska, E.; Wielosz, M.; Turski, W.A.; Urbanska, E.M. L-cysteine sulphinate, endogenous sulphur-containing amino acid, inhibits rat brain kynurenic acid production via selective interference with kynurenine aminotransferase ii. *Neurosci. Lett.* **2003**, *346*, 97–100. [[CrossRef](#)]
238. Luchowski, P.; Kocki, T.; Urbanska, E.M. N^g-nitro-l-arginine and its methyl ester inhibit brain synthesis of kynurenic acid possibly via nitric oxide-independent mechanism. *Pol. J. Pharmacol.* **2001**, *53*, 597–604. [[PubMed](#)]

239. Agudelo, L.Z.; Femenía, T.; Orhan, F.; Porsmyr-Palmertz, M.; Goiny, M.; Martinez-Redondo, V.; Correia, J.C.; Izadi, M.; Bhat, M.; Schuppe-Koistinen, I. Skeletal muscle pgc-1 α 1 modulates kynurenine metabolism and mediates resilience to stress-induced depression. *Cell* **2014**, *159*, 33–45. [[CrossRef](#)] [[PubMed](#)]
240. Barth, M.C.; Ahluwalia, N.; Anderson, T.J.; Hardy, G.J.; Sinha, S.; Alvarez-Cardona, J.A.; Pruitt, I.E.; Rhee, E.P.; Colvin, R.A.; Gerszten, R.E. Kynurenine acid triggers firm arrest of leukocytes to vascular endothelium under flow conditions. *J. Biol. Chem.* **2009**. [[CrossRef](#)]
241. Kwidzinski, E.; Bunse, J.r.; Aktas, O.; Richter, D.; Mutlu, L.; Zipp, F.; Nitsch, R.; Bechmann, I. Indolamine 2, 3-dioxygenase is expressed in the cns and down-regulates autoimmune inflammation. *FASEB J.* **2005**, *19*, 1347–1349. [[CrossRef](#)] [[PubMed](#)]
242. Flanagan, E.M.; Erickson, J.B.; Viveros, O.H.; Chang, S.Y.; Reinhard, J.F., Jr. Neurotoxin quinolinic acid is selectively elevated in spinal cords of rats with experimental allergic encephalomyelitis. *J. Neurochem.* **1995**, *64*, 1192–1196. [[CrossRef](#)] [[PubMed](#)]
243. Paul, C.; Bolton, C. Modulation of blood-brain barrier dysfunction and neurological deficits during acute experimental allergic encephalomyelitis by then-methyl-d-aspartate receptor antagonist memantine. *J. Pharmacol. Exp. Ther.* **2002**, *302*, 50–57. [[CrossRef](#)] [[PubMed](#)]
244. Schroecksadel, K.; Winkler, C.; Wirleitner, B.; Schennach, H.; Fuchs, D. Aspirin down-regulates tryptophan degradation in stimulated human peripheral blood mononuclear cells in vitro. *Clin. Exp. Immunol.* **2005**, *140*, 41–45. [[CrossRef](#)]
245. Edwards, S.R.; Mather, L.E. Diclofenac increases the accumulation of kynurenate following tryptophan pretreatment in the rat: A possible factor contributing to its antihyperalgesic effect. *Inflammopharmacology* **2003**, *11*, 277–292. [[CrossRef](#)] [[PubMed](#)]
246. Jorge, A.G.; Modulo, C.M.; Dias, A.C.; Braz, A.M.; Filho, R.B.; Jordao, A.A., Jr.; de Paula, J.S.; Rocha, E.M. Aspirin prevents diabetic oxidative changes in rat lacrimal gland structure and function. *Endocrine* **2009**, *35*, 189–197. [[CrossRef](#)] [[PubMed](#)]
247. Tong, L.; Wong, T.Y. Aspirin and dry eye? *Ophthalmology* **2009**, *116*, 167. [[CrossRef](#)]
248. Yazici, A.; Sari, E.; Ayhan, E.; Sahin, G.; Tiskaoglu, N.S.; Gurbuzer, T.; Kurt, H.; Ermis, S.S. The Effect of Low-Dose Aspirin on Dry Eye Parameters and Ocular Surface Disease Index Questionnaire. *J. Ocul. Pharm.* **2018**, *34*, 256–259. [[CrossRef](#)]
249. Liu, R.; Su, D.; Zhou, M.; Feng, X.; Li, X.; Sun, L. Umbilical cord mesenchymal stem cells inhibit the differentiation of circulating T follicular helper cells in patients with primary Sjogren’s syndrome through the secretion of indoleamine 2,3-dioxygenase. *Rheumatology* **2015**, *54*, 332–342. [[CrossRef](#)] [[PubMed](#)]
250. Yaksh, T.L.; Schwarcz, R.; Snodgrass, H.R. Characterization of the Effects of L-4-Chlorokynurenine on Nociception in Rodents. *J. Pain* **2017**, *18*, 1184–1196. [[CrossRef](#)] [[PubMed](#)]
251. Elmaagacli, A.H.; Ditschkowski, M.; Steckel, N.K.; Gromke, T.; Ottinger, H.; Hillen, U.; Baba, H.A.; Trenchel, R.; Beelen, D.W.; Koldehoff, M. Human chorionic gonadotropin and indolamine 2,3-dioxygenase in patients with GVHD. *Bone Marrow Transpl.* **2014**, *49*, 800–805. [[CrossRef](#)]
252. Gajewski, T.F.; Schreiber, H.; Fu, Y.X. Innate and adaptive immune cells in the tumor microenvironment. *Nat. Immunol.* **2013**, *14*, 1014–1022. [[CrossRef](#)]
253. Beatty, G.L.; O’Dwyer, P.J.; Clark, J.; Shi, J.G.; Bowman, K.J.; Scherle, P.A.; Newton, R.C.; Schaub, R.; Maleski, J.; Leopold, L.; et al. First-in-Human Phase I Study of the Oral Inhibitor of Indoleamine 2,3-Dioxygenase-1 Epacadostat (INCB024360) in Patients with Advanced Solid Malignancies. *Clin. Cancer Res.* **2017**, *23*, 3269–3276. [[CrossRef](#)]

