



Congenital Heart Disease: An Immunological Perspective

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Singampalli KL, Jui E, Shani K, Ning Y, Connell JP, Birla RK, Bollyky PL, Caldarone CA, Keswani SG and Grande-Allen KJ (2021) Congenital Heart Disease: An Immunological Perspective. Front. Cardiovasc. Med. 8:701375. doi: 10.3389/fcvm.2021.701375 Congenital heart disease (CHD) poses a significant global health and economic burden—despite advances in treating CHD reducing the mortality risk, globally CHD accounts for approximately 300,000 deaths yearly. Children with CHD experience both acute and chronic cardiac complications, and though treatment options have improved, some remain extremely invasive. A challenge in addressing these morbidity and mortality risks is that little is known regarding the cause of many CHDs and current evidence suggests a multifactorial etiology. Some studies implicate an immune contribution to CHD development; however, the role of the immune system is not well-understood. Defining the role of the immune and inflammatory responses in CHD therefore holds promise in elucidating mechanisms underlying these disorders and improving upon current diagnostic and treatment options. In this review, we address the current knowledge coinciding CHDs with immune and inflammatory associations, emphasizing conditions where this understanding would provide clinical benefit, and challenges in studying these mechanisms.

Keywords: congenital heart disease, immune response, inflammation, primary immunodeficiency, models of CHD

INTRODUCTION

Pediatric heart disease, though often overshadowed by its adult counterpart, is a significant health burden, affecting over 15 million children globally (1). Additionally, congenital malformations are a major source of mortality, accounting for accounting for an annual 300,000 deaths worldwide (2). Congenital heart diseases (CHDs) have been reported in anywhere from 3.7 to 75 per 1,000 live births depending on the population, methods of diagnosis, and severity of the disease (3, 4). These heart defects form in utero due to abnormalities in the formation of the cardiac structures and conduction system (5). In many cases, CHDs alter the blood flow pattern throughout the heart as they generate lower pressure pathways that disrupt typical flow; this can have significant morbidity risks as it affects oxygenation and the systemic/pulmonary volume status, which, in turn, can lead to reactionary inflammation (6, 7).

Although vast improvements have been made in CHD treatment and survival (8), they are not without complications. Even in mild cases, children with CHDs report a lower quality of

life, poorer performance in school and an inability to physically keep up with their peers (9). Children with more severe CHDs often require serial invasive heart surgeries or transplants (10). Furthermore, these diseases can cause fatal cardiac complications even throughout adulthood (11) and are estimated to reduce life expectancy by nearly 5 years (12).

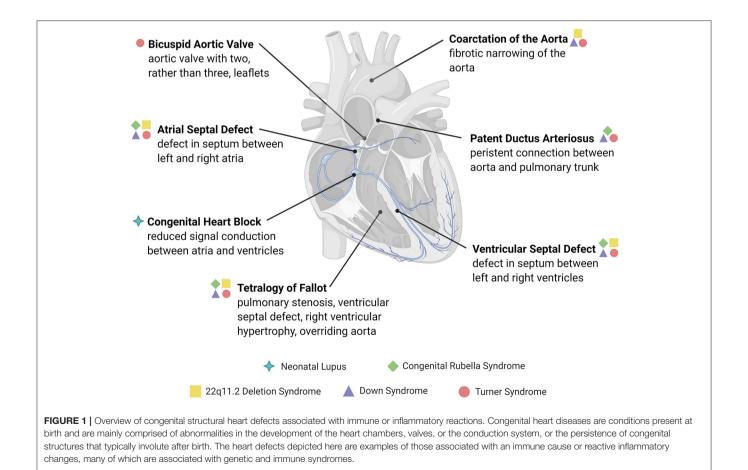
Despite these complications, there is little conclusive evidence on the specific cause of many structural CHDs, and collectively they are quite heterogeneous. Current research implicates a combination of genetic, epigenetic, and environmental factors as causative mechanisms underlying CHDs (13). A link to the immune system has not been well-defined; however, there is a clear association, as evidenced by an increased risk of these children contracting and experiencing severe complications from common infections. Clinical studies have shown a reduced cellular immune response to infection and increased proinflammatory cytokine levels among children with structural CHDs (14-16), which indicates that the immune system may be a dynamic partner in the development of complications from CHDs. Furthermore, immune cells, such as macrophages, have been shown to play a critical role in cardiac development (17). Given how changes in the immune response can be seen in children with CHDs, a potentially fruitful avenue to study pediatric heart disease can be through investigating the immune mechanisms at play.

To this end, we discuss the clinical motivation for studying the intersection of the immune system and CHDs, discuss specific CHDs associated with immune or inflammatory complications (**Figure 1**), and identify challenges in studying the connection between the two. Building on these data, we highlight areas where modulating the immune response may show benefit in preventing the development of disease or its sequelae.

CLINICAL IMMUNOLOGIC IMPLICATIONS OF CHDs

A motivation for studying the link between immunology and heart disease is that structural CHDs are associated with reduced immune cell counts and maturity. Specifically, children with CHDs have reduced granulocyte activity against bacterial infections (18), T and B lymphocyte levels (14, 19), naïve Tcell production and T-Cell Receptor Excision Circles (TREC) levels (20), IgA and IgG levels, and complement levels, as well as increased suppressor T-cell function (14).

This immune profile becomes clinically relevant, as children with structural CHDs have increased morbidity when exposed to common pathogens, such as respiratory syncytial virus (RSV), and an increased risk of developing bronchopneumonia or other infectious complications. In RSV bronchiolitis, the presence of a CHD can lead to lower oxygen saturations, increased length



of hospital stays, likelihood of ICU admission, and up to a $25 \times$ increased risk of mortality (21, 22). Furthermore, in premature infants, CHD is associated with an increased risk of sepsis (23) with up to 35% of late-onset neonatal sepsis presenting in children with Patent Ductus Arteriosus (PDA). These infants are more likely to develop recurrent sepsis, thus requiring longer ventilation times and hospital stays (24). One explanation for the immune complications is the altered anatomy of children with CHDs, specifically the changes in pulmonary circulation. For example, in children with left to right shunts, where increased right-sided flow can cause pulmonary edema or cyanotic heart disease, RSV can be more severe at baseline (25). However, the increased severity of respiratory diseases can also be caused by a shift in immune responses. This is seen in children with CHD who develop bronchopneumonia (BP), who then exhibit an exaggerated increase in both B- and T- cell subsets in addition to the expected increase of $CD3^+$ and $CD8^+$ cells (26).

The increased susceptibility to infectious complications among children with CHDs may be due in part to the inflammatory response. Although the inflammatory response is elevated at baseline in children with CHDs (6, 15, 27), it is particularly exaggerated in infections, when children present with elevated cytokine and bacterial endotoxin levels (15). Though these inflammatory changes have mainly been observed in small scale clinical studies and there is no consensus on the specific inflammatory profiles, alterations in inflammatory responses have been seen across multiple CHDs. For example, changes in the cytokine profile have been studied as part of a compensatory remodeling mechanism in a variety of structural diseases, including septal defects and shunts. Children born with an atrial septal defect (ASD) undergo cardiac remodeling to compensate for the flow of blood from the left atrium to right atrium. This leads to the presence of markers for mechanical stress, inflammation, and remodeling (28), including tumor necrosis factor (TNF)- α (27). Similar changes in inflammatory cytokines and acute-phase reactants are seen with ventricular septal defects (VSDs) and other left to right shunts (6, 16). In some untreated septal defects or left to right shunts, the pressures in the right heart can become greater than that of the left heart, leading to the reversal of flow, known as Eisenmenger syndrome. This also creates a systemic proinflammatory response, with increased C-reactive protein (CRP) and Interferon (IFN)- γ levels (29). Furthermore, in children with coarctation of the aorta, increased inflammatory and apoptotic mediators, including Interleukin (IL)-6, IL-10, TNF-a, and soluble Fas (sFas), may contribute to vascular disease as these children age (30).

Better understanding this inflammatory response can aid in developing prophylactic treatments to prevent complications from CHDs. This has been studied in adult cardiac diseases, where combatting inflammatory markers has been beneficial in therapeutics. For example, heart failure or remodeling after myocardial infarction are associated with a pro-inflammatory cytokine-induced immune response, which, in turn, can cause irreversible fibrosis or systolic dysfunction (31, 32). Specifically, inhibiting molecules such as IL-6 and transforming growth factor (TGF)- β reduces the fibrotic response, making them potential targets for treatment (33, 34). Furthermore, cardiac remodeling in adults is associated with similar cytokine profiles to CHD, including elevated IL-6 and TNF levels (35–37). Though the parallel studies have not been conducted in pediatrics, future large-scale studies can better predict the cytokine profile in CHD, accomplish similar therapeutic success and prevent long-term complications in children.

Since an understanding of the immune and inflammatory contributions to CHD is limited, we have chosen to highlight conditions that establish the range of CHDs with an immune association (**Table 1**), and the long-term implications of these diseases. In the following sections, we will address examples of CHDs with an immune cause, followed by genetic syndromes involving both CHDs and immune dysregulation. For each, we will discuss the current mechanistic understanding linking the two as well as areas for future research to successfully translate this understanding to clinical practice.

EXAMPLES OF CHDs ASSOCIATED WITH IMMUNE CHANGES

CHDs With an Immune Etiology Autoimmune-Mediated CHDs: Congenital Heart Block

The prototypical example of autoimmune-mediated CHD is congenital heart block (CHB). CHB is a blockage of the appropriate contractile signaling traveling between the atria and ventricles and is part of a larger constellation of symptoms identified as neonatal lupus, which is correlated to the presence of autoimmune disease in the mother. CHB first can be detected using fetal echocardiography (54); however, at the time of birth, most CHB has progressed to third-degree heart block (55), leading to a 60% pacemaker rate among these children (56, 57). Long-term consequences of CHB include cardiomyopathy, valve disease, endothelial fibroelastosis (58, 59), and a 16% mortality risk (60), all of which necessitate a better understanding of this condition.

A majority of cases are associated with maternal antibodies against Ro (SSA)–52, Ro-60 or La (SSB)–48, as seen in systemic lupus erythematosus (SLE) (61), or autoantibodies associated with Sjogren's syndrome (39). Specifically, the presence of anti-Ro antibodies in maternal serum is associated with a greater risk of progressive CHB and complications, including irregular rhythms, neonatal pacemaker insertion, and spontaneous abortion (62). Newer data show a correlation between antibodies to Ro-52 amino acid residues 1–135 and 200–239, correlated to zinc finger coding regions, and the development of CHB (63).

Although an autoimmune component has been established, specific mechanisms connecting autoantibodies to cardiac damage are still being investigated. One mechanism focuses on anti-Ro/anti-La autoantibody targeting of apoptotic cardiac cells (61), as shown in **Figure 2A**. During apoptosis, Ro

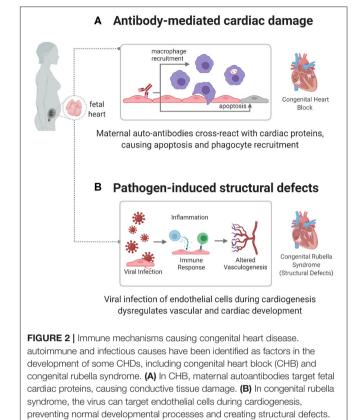
TABLE 1 Immune and inflammatory markers associated with congenital hear	rt
disease.	

Disorder	Mechanisms	References
Structural CHDs	↑ Suppressor T cell function	Parikh et al. (18)
	↓ T and B lymphocytes	Khalil et al. (14)
	↓ IgA and IgG	Rhoden et al. (19)
	↓ T cell maturity	Davey et al. (20)
	↓ Granulocyte activity against bacterial infections	
	↓ Complement levels	
Congenital Heart Block	↑ Maternal antibodies against Ro (SSA)-52, Ro-60 or La (SSB)-48	Llanos et al. (38), Rischmuller et al. (39)
	↑ Maternal autoantibodies from Sjogren's syndrome	
Congenital Rubella Syndrome	↑ Rubella antigen	Dhiman et al. (40),
	↑ Cytokine (IL-10, IFN-γ) production	Geyer et al. (41), Lazar et al. (42)
22q11.2 Deletion Syndrome	\downarrow T and B lymphocytes	Sullivan et al. (43)
	↓ Antibody production	Human et al. (44), Kuo et al. (45), Lambert et al. (46)
Down Syndrome	↑ Toll-Like Receptor 2 signaling	Ram et al. (47)
	↓ T and B lymphocytes	Zampierei et al. (48)
	↓ Effector T cell responsiveness	Farroni et al. (49)
	↓ IgG production	Huggard et al. (50)
Turner Syndrome	↑ IgA production	Stenberg et al. (51)
	↑ Cytokine (IL-6, TGF-β) production	Su et al. (52)
	↓ IgG production	Gawlik et al. (53)
	↓ IgM production	
	\downarrow T and B lymphocytes	
	↓ CD4:CD8 ratio	
	↓ Cytokine (IL-10) production	

↑ Means increased levels/activity. ↓ Means decreased levels/activity.

and La antigens are translocated to the cell membrane, where they can be targeted by autoantibodies. This leads to opsonization, which attracts macrophages, leading to myofibroblast differentiation, TGF- β release, and fibrosis and calcification of the atrioventricular node and conduction system (64–67). Another mechanism focuses on the cross-reactivity of autoantibodies with L-type calcium channels, present in pacemaker cardiac cells (68), or with laminin (69). However, these pathways need to be better defined to act as potential diagnostic or therapeutic targets in children with CHB.

Due to the combination of challenges in early diagnosis and the irreversible nature of CHB, effective therapies have been difficult to develop. Since the immune origins of CHB have been established, therapies have focused on immunosuppression. However, none of the current treatment options have shown widespread efficacy. Though some research suggests that dexamethasone may prevent the progression of heart block (70, 71), other studies have shown that it is not sufficient to improve outcomes (72). Dexamethasone may show some benefit



in combination with a β -adrenergic medication to maintain heart rate (70), but both steroids and β -adrenergic drugs have adverse effects, making their use controversial (73, 74). Intravenous immunoglobulin (IVIG) has also been studied as a potential treatment and is believed to work by outcompeting the anti-Ro and anti-La antibodies for transfer across the placenta (74); unfortunately, IVIG has not been shown to prevent heart block or improve survival (75, 76). Furthermore, hydroxychloroquine, one of the treatments typically used in adult autoimmune disease, has been shown to reduce the development of CHB (77), but there is not sufficient evidence for its efficacy. This lack of adequate treatment options places children with CHB at risk of high pacemaker rates and subsequent complications.

Since CHB is one of the better understood connections between the immune system and congenital disease, it depicts the potential of targeted immunomodulatory therapies for heart disease. A comprehensive understanding of autoimmune mediated pathways can therefore help advance the prevention of maternal autoimmune complications in the fetus and the treatment of CHB in children.

Pathogen-Induced CHDs: Congenital Rubella Syndrome

Congenital rubella is a viral infection that postnatally causes rash, lymphadenopathy, fever, and sore throat. Intrauterine infection with the rubella virus, however, can lead to intrauterine growth restriction, cataracts, abnormal brain

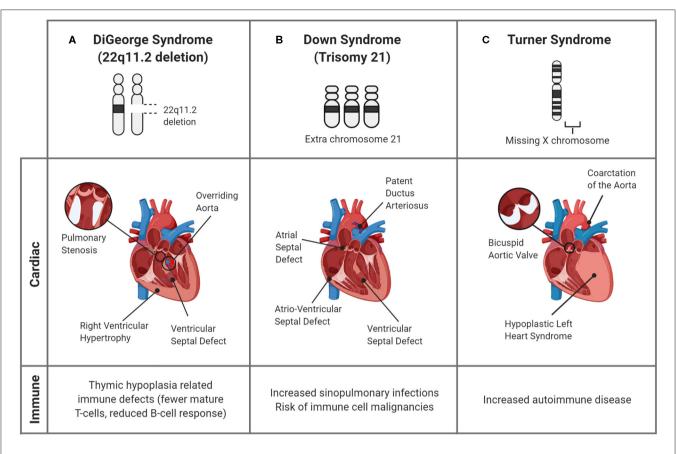


FIGURE 3 | Genetic syndromes associated with CHDs and altered immune responses. Though many genetic syndromes are associated with both immune deficiencies and CHDs, they are not linked directly. (A) In DiGeorge Syndrome, the abnormal development of the pharyngeal arches, the precursor structure for both the heart and thymus, leads to mature T-cell deficiency and cardiac defects. (B) Down Syndrome is commonly associated with cardiac septal defects and an increased propensity for infections in childhood. Though the associations are unclear, the alterations in immune response are evidenced by the greater prevalence of immune cell malignancies. (C) In Turner Syndrome, children can have CHDs, including bicuspid aortic valve and aortic damage, as well as immune complications, such as autoimmune disease. A connection between the cardiac and immune systems is seen in the inflammatory complications of the bicuspid valves, which undergo premature immune-mediated calcification, and induce a systemic pro-inflammatory state.

development, and CHDs, most commonly septal defects, PDA, pulmonary artery stenosis, and Tetralogy of Fallot (ToF) (13, 78, 79).

The mechanism by which the infection causes cardiac defects has not been clearly identified. However, current leading theories include viral damage to vasculature or endothelial cells, affecting the development of tissue early in gestation (Figure 2B) (78). Lazar et al. identified the rubella antigen in cardiac and aortic fibroblasts, placental capillary endothelial cells, and cerebral progenitor cells (42), which match the structures associated with disease phenotypes. Furthermore, the rubella virus has been shown to replicate and persist in endothelial cells in vitro, without affecting their shape, structure, or ability to proliferate (80). This infection induces gene expression changes in endothelial cells. Furthermore, induced Pluripotent Stem Cells (iPSCs) infected with rubella and differentiated to an endodermal lineage show an altered expression of endodermal markers and genes related to interferon responsiveness, chromatin remodeling, vasculogenesis, and cell adhesion (81), which induce the

recruitment of inflammatory cytokines (IL-10, IFN- γ) and effector lymphocyte responses (40, 41). This connection between altered gene expression and cardiac maldevelopment can help identify specific pathways affected by an immune response in the development of disease.

Syndromic Associations With Altered Immune Responses and CHDs

Many genetic conditions present with both immunologic and cardiac disease manifestations (44). In fact, some CHDs are associated with concomitant genetic immune disorders, collectively termed primary immune deficiencies. Though a comprehensive discussion of these heterogeneous conditions is beyond the scope of this review, there are several relatively common syndromic diseases that present with cardiac and immunologic features as part of a single genetic defect. These conditions, discussed below, include 22q11.2 deletion (DiGeorge) Syndrome, Down Syndrome, and Turner Syndrome (**Figure 3**).

22q11.2 Deletion Syndrome

A syndrome commonly associated with both immunodeficiency and cardiac disease is 22q11.2 deletion syndrome. In this condition, the third and fourth pharyngeal arches, and their subsequent structures, including the nasopharynx, thyroid, parathyroid, thymus, and cardiac outflow tract can be maldeveloped, leading to palatal defects, hypothyroidism, hypocalcemia, T-cell immunodeficiency, and conotruncal cardiac defects among characteristic facial features (44-46). Since the thymus selects for T-cells that recognize foreign antigens and not self-antigens, thymic hypoplasia causes a T-cell deficiency in up to 75% of these children (43), leading to chronic sinopulmonary infections (82). This reduced thymic T-cell maturation is also evidenced by reduced numbers of TRECs, a byproduct of the selection process (44). The body can attempt to compensate for T-cell lymphopenia through homeostatic expansion, where naive T-cells, often against self-antigens, replicate and mature to restore normal T-cell counts, introducing an avenue for autoimmunity (83).

Additionally, due to the abnormal development of the cardiac outflow tract, up to 80% of children with 22q11.2 deletion syndrome can have congenital heart defects, including VSDs and ToF (84), as depicted in **Figure 3A**. Studies suggest that a unifying factor between immune and cardiac defects may be altered arterial formation, specifically due to endothelial and neural crest cells. This can lead to improper formation of the cardiac outflow tract and changes in the blood supply feeding the thymus and parathyroid glands (85). Further research to identify the mechanisms that link these deficiencies is crucial to determine deleted genes that are responsible for both immune and cardiac developmental anomalies, which could act as a prognostic or therapeutic target.

Down Syndrome

Down Syndrome (DS), also known as trisomy 21, is a condition where children are born with a third copy of chromosome 21 causing a characteristic appearance and clinical comorbidities including developmental delay, a risk of obstructive sleep apnea, gastrointestinal atresias, intellectual disability, and CHD (50). These children are susceptible to frequent and/or prolonged respiratory infections and otitis media, which can be attributed to both altered anatomy and immune changes (47).

Immune complications are focused on the adaptive immune system. Altered myelopoiesis and incomplete fate switching to a memory subtype results in lower numbers of B- and T-cells in up to 90% of children with DS, complicating their responsiveness to vaccines and increasing their risk of autoimmunity (47, 49, 86). Additionally, effector and regulatory T-cell responsiveness and IgG production are reduced in DS (47, 87). Although there is no consensus, potential mechanisms include altered miRNA expression within immune cells, systemic expression of immune-related genes, and increased Toll-Like Receptor (TLR)–2 signaling leading to chronic inflammation (48–50). A potentially fatal complication that supports immune dysregulation in DS is the risk of developing childhood megakaryoblastic or lymphoblastic leukemias. However, these children may have an immune profile that is protective from solid tumors later in life

(88). In this case, not only can understanding the immune profile inform cancer diagnosis and treatment in children with DS, but can also provide insight into immune mediators of solid tumors.

Although a direct connection to the immune response has not been elucidated, many of these children have conotruncal cardiac defects, such as atrioventricular septal defects (AVSD) and PDA (**Figure 3B**), leading to congestive heart failure or valvular insufficiency. Furthermore, these changes expose endothelial cells to altered shear stresses, causing vascular changes including persistent pulmonary hypertension of the newborn (89, 90). The exact mechanism is unknown, however the similarities in defects among children with DS indicate that chromosome 21 encodes genes important for cardiac development (90). The identification of the specific causes would be beneficial since the greatest risk of infant mortality in DS is associated with CHDs (89).

Interestingly, patients with DS have a reduced incidence of coronary artery disease and atherosclerosis, which may be attributed to concomitant increases in protective factors (90). Similar to the immune complications, understanding these mechanisms can allow for the treatment of a major cause of morbidity in the general population.

Turner Syndrome

Turner Syndrome (TS) is a female chromosomal disorder that results in a missing or partially missing X chromosome. With an incidence of 1 in 2,000–2,500 births, TS is one of the most common chromosomal defects (53). Though there is often a delay in diagnosis (91), many TS patients have HLHS, bicuspid aortic valves (BAV), aortic stenosis, or coarctation of the aorta (CoA) (92), as illustrated in **Figure 3C**. As adults, patients also are at risk of left ventricular hypertrophy, coronary artery disease, valve dysfunction (91) and most acutely, the fatal rupture of the thoracic aorta following chronic enlargement (92, 93). The development of these complications may have an inflammatory etiology, as, patients with thoracic aortic aneurysms have an increased number of natural killer, B, and T-cells (94) and inflammatory markers such as TNF- α , IL-1, IL-6, and IL-17 (95).

The risk of autoimmune disease in patients with TS is two times greater than the general female population, suggesting that the immune system plays a clear role in the development of TS complications. Approximately 20-50% of patients with TS are diagnosed with an autoimmune disease (53), with the most common being thyroid disease and diabetes (91). Furthermore, several studies determined that TS patients with an increased risk of autoimmune disorders exhibit a lower percentage of CD4⁺ cells and a lower CD4⁺:CD8⁺ ratio (51-53). Immunological disturbances, such as low levels of IgG, IgM, T- and B-lymphocytes, increased IgA, and pro-inflammatory changes (increased IL-6 and TGF- β , decreased IL-10), have also been associated with TS (51, 91), all of which could serve as predictive factors of complications. It is currently unclear whether the cardiovascular aberrations provoke these immunological deficiencies, or vice versa. However, further studies are warranted given the multitude of evidence linking the two in multiple genetic syndromes.

MODELS OF THE IMMUNE ASPECTS OF CHDS

Despite the advances in studying CHDs, one of the challenges in understanding the immune component is the paucity of *in vitro* or animal models for these diseases. Though syndromic CHDs have been studied in genetic-based animal models and computational models, these do not always lend themselves easily to the study of molecular and immune mechanisms. For example, most mice used to study 22q11.2 deletion syndrome are typically created using single gene mutations (96), which may not encompass the complexity of the disease. Therefore, much of the research in this field is focused on extrapolating data from other diseases or studying clinical manifestations.

In vitro models have shown promise in recapitulating mechanisms behind some CHDs. Human induced pluripotent stem cell (hiPSC)-derived cardiomyocytes can be used to model gene regulatory interactions, cell-cell interactions, and tissue interactions contributing to CHD. Specifically, the use of hiPSCs allows for a multitude of benefits such as the direct application to human phenotypes and disease, the ability to study singlecell genomics and epigenetics, scalability to increase detection of low level or transient signaling molecules, the ability to perturb and study developmental interactions that occur at the molecular and cellular levels, and the ability to study tissue organization and interactions using hiPSCs on bioengineered matrices (97). This is especially promising in studying the interactions of immune cells and cardiac cells under specific environmental cues. Furthermore, patient-specific cell lines can be used to investigate multiple pediatric cardiovascular disorders to provide insight to the mechanisms involved and usher in novel personalized therapies for patients.

Computational fluid dynamic models can also provide insight to varied flow patterns and areas of maximum stress in aortic coarctation resulting in patient-specific treatment options (98). These models can predict changes in shear stress, pressure, cardiac energy losses, and PV loops caused by structural differences in these patients (99), which can then be correlated to immune or inflammatory changes seen physiologically with the altered hemodynamics. Computational models are beneficial because they can be individualized to the anatomy of a specific patient. This allows for personalized models of pediatric cardiovascular disorders which can then be compared with immune responses to elucidate trends. Especially since the extent of structural changes in many of these diseases is variable, trends from these computational models can be beneficial in predicting the severity and progression of disease when correlated with clinical data. For example, patient data including inflammatory cytokine levels, CRP, and other immunological profiles can be used as input parameters for the prediction of disease pathogenesis (100, 101). However, these tools still fail to elucidate molecular mechanisms leading to CHDs.

In vivo models that have been used to study cardiac development include chicken embryos, zebrafish, and mice. Studying the development of microscopic cardiac structural changes or macroscopic defects in chick embryos involves

altering blood flow using drugs or surgical changes, such as banding arteries (102, 103). These models can then be used to study downstream immune and inflammatory changes due to alterations in cardiac flow patterns. Similarly, mouse models of CHDs associated with genetic mutations show a promising avenue for the study of the related immune mechanisms (104). For example, mutations associated with septal defects, such as in NKX2-5 (105) and GATA4 (106), have caused both ASDs and VSDs in mice, whereas mutations in the NOTCH (107) and GATA5 (108) pathways have recapitulated BAVs in mice. These models are beneficial to understand the downstream immune and inflammatory changes of CHDs. However, creating these requires a significant understanding of the genetic causes of CHD, imposes a lack of control on the severity of the defect, and are associated with a long time and high cost for development (104).

Although animal models do not yet exist for many types of structural CHDs, one method of studying their effects is with animal models of the hemodynamic changes associated with CHD. For instance, to model the high cardiac work conditions causing hypertrophy, a pump can infuse isoproterenol, a β agonist, into pediatric mice (109). Similarly, in diseases where forward flow is impaired, such as coarctation of the aorta or valve stenosis, models of volume or pressure overload can be used as a surrogate. An example would be an aortic coarctation rat model used by Shang et al. (110), which led to increased pressure within the left heart. This allows for the study of inflammatory markers associated with CHDs, including an increase in reactive oxygen species (ROS) and macrophage chemoattractant protein (MCP)-1. Models like this can help in studying the consequences of a particular CHD; however, the specific immune causes of the defects are likely not clearly elucidated from these downstream pathways. This also holds true for larger animal models, such as porcine (111, 112) and ovine (113) models, where the structural heart defect needs to be created by fetal surgery and would therefore not mimic immune mechanisms that cause the initial defects in humans, but could model reactionary inflammatory and immune changes.

The lack of a model to comprehensively study the relationship between the immune system and CHD imposes a challenge to thoroughly understand these conditions. Furthermore, the variability seen in phenotypes of animal models highlights the complexity of the mechanisms underlying these diseases. Despite these limitations, a combination of techniques can be used to study the immune and inflammatory causes of and responses to CHDs. Further refining the models used to study CHDs can expand our current insight into these conditions, which in turn can improve the care of affected children.

CONCLUSION

Despite our improved understanding of CHDs and treatment modalities, the role of the immune system in their development or complications is relatively poorly understood. Studies have shown that children with CHDs have altered inflammatory and immune profiles which may contribute to the severe reactions to common infectious agents. However, the specific interaction between the immune system and cardiac defects is not well-understood. Some CHDs, such as defects resulting from congenial rubella or neonatal lupus, have immune mechanisms involved in their development. However, many syndromic associations between CHDs and the immune response have yet to be linked. Additionally, the contribution of the immune system to genetic and environmental factors that cause CHDs has not been well-understood.

A challenge to better defining this relationship is the limits of *in vitro* and *in vivo* models in recapitulating the interactions between the various factors. Much of the current understanding of the immune and inflammatory response in CHD is typically from clinical research and is focused on complications of altered blood flow patterns caused by CHDs. However, by expanding research to a more mechanistic understanding, we can identify specific targets in the development of CHDs or the progression of their complications, thereby expanding our potential to clinically

REFERENCES

- Musa NL, Hjortdal V, Zheleva B, Murni IK, Sano S, Schwartz S, et al. The global burden of paediatric heart disease. *Cardiol Young*. (2017) 27:S3– S8. doi: 10.1017/S1047951117002530
- Global, regional, and national life expectancy, all-cause mortality, and causespecific mortality for 249 causes of death, 1980–2015: a systematic analysis for the Global Burden of Disease Study 2015. *Lancet.* (2016) 388:1459– 544. doi: 10.1016/S0140-6736(16)31012-1
- Hoffman JIE, Kaplan S. The incidence of congenital heart disease. J Am Coll Cardiol. (2002) 39:1890–900. doi: 10.1016/S0735-1097(02)01886-7
- Ferencz C, Rubin JD, McCarter RJ, Brenner JI, Neill CA, Perry LW, et al. Congenital heart disease: prevalence at livebirth. The Baltimore-Washington Infant Study. *Am J Epidemiol.* (1985) 121:31– 6. doi: 10.1093/oxfordjournals.aje.a113979
- Wenink AC, Gittenberger-de Groot AC. Pathogenesis of congenital cardiac malformations and mechanisms of cardiac remodelling. *Cardiol Young.* (2005) 15 (Suppl 3):3–6. doi: 10.1017/S1047951105 001551
- Buchhorn R, Wessel A, Hulpke-Wette M, Bürsch J, Werdan K, Loppnow H. Endogenous nitric oxide and soluble tumor necrosis factor receptor levels are enhanced in infants with congenital heart disease. *Crit Care Med.* (2001) 29:2208–10. doi: 10.1097/00003246-200111000-00026
- Holst KA, Said SM, Nelson TJ, Cannon BC, Dearani JA. Current interventional and surgical management of congenital heart disease: specific focus on valvular disease and cardiac arrhythmias. *Circ Res.* (2017) 120:1027– 44. doi: 10.1161/CIRCRESAHA.117.309186
- Gilboa SM, Salemi JL, Nembhard WN, Fixler DE, Correa A. Mortality Resulting From Congenital Heart Disease Among Children and Adults in the United States, 1999 to 2006. *Circulation*. (2010) 122:2254– 63. doi: 10.1161/CIRCULATIONAHA.110.947002
- Uzark K, Jones K, Slusher J, Limbers CA, Burwinkle TM, Varni JW. Quality of life in children with heart disease as perceived by children and parents. *Pediatrics*. (2008) 121:e1060–7. doi: 10.1542/peds.2006-3778
- 10. Thrush PT, Hoffman TM. Pediatric heart transplantation-indications and outcomes in the current era. *J Thorac Dis.* (2014) 6:1080–96.
- Wu M, Lu C, Chen H, Kao F and Huang S. Adult congenital heart disease in a nationwide population 2000–2014: epidemiological trends, arrhythmia, and standardized mortality ratio. J Am Heart Assoc. (2018) 7:e007907. doi: 10.1161/JAHA.117.007907
- Gong CL, Zhao H, Wei Y, Tysinger B, Goldman DP, Williams RG. Lifetime burden of adult congenital heart disease in the USA using a microsimulation model. *Pediatr Cardiol.* (2020) 41:1515–25. doi: 10.1007/s00246-020-02409-9

manage both acute and long-term complications of CHDs and reducing the global morbidity and mortality burden imposed on children with these conditions.

AUTHOR CONTRIBUTIONS

KG-A conceptualized the article. EJ and KS conducted the literature review, drafted the manuscript, and created the figures. All authors critically edited the article.

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- Blue GM, Kirk EP, Sholler GF, Harvey RP, Winlaw DS. Congenital heart disease: current knowledge about causes and inheritance. *Med J Aust.* (2012) 197:155–9. doi: 10.5694/mja12.10811
- Khalil A, Trehan R, Tiwari A, Malik R and Arora R. Immunological profile in congenital heart disease. *Indian Pediatr.* (1994) 31:295–300.
- Sharma R, Bolger AP, Li W, Davlouros PA, Volk H-D, Poole-Wilson PA, et al. Elevated circulating levels of inflammatory cytokines and bacterial endotoxin in adults with congenital heart disease. *Am J Cardiol.* (2003) 92:188–93. doi: 10.1016/S0002-9149(03)00536-8
- Zhang X, Wang K, Yang Q, Wang J, Xuan C, Liu XC, et al. Acute phase proteins altered in the plasma of patients with congenital ventricular septal defect. *Proteomics Clin Appl.* (2015) 9:1087–96. doi: 10.1002/prca.201400166
- Swirski FK, Nahrendorf M. Cardioimmunology: the immune system in cardiac homeostasis and disease. *Nat Rev Immunol.* (2018) 18:733– 44. doi: 10.1038/s41577-018-0065-8
- Parikh S, Bharucha B, Kamdar S and Kshirsagar N. Polymorphonuclear leukocyte functions in children with cyanotic and acyanotic congenital heart disease. *Indian Pediatr.* (1993) 30:883–90.
- Rhoden DK, Leatherbury L, Helman S, Gaffney M, Strong WB, Guill MF. Abnormalities in lymphocyte populations in infants with neural crest cardiovascular defects. *Pediatr Cardiol.* (1996) 17:143– 9. doi: 10.1007/BF02505203
- Davey BT, Elder RW, Cloutier MM, Bennett N, Lee JH, Wang Z, et al. T-cell receptor excision circles in newborns with congenital heart disease. *J Pediatr.* (2019) 213:96–102.e2. doi: 10.1016/j.jpeds.2019.05.061
- La Via WV, Grant SW, Stutman HR, Marks MI. Clinical profile of pediatric patients hospitalized with respiratory syncytial virus infection. *Clin Pediatr.* (1993) 32:450–4. doi: 10.1177/000992289303200801
- MacDonald NE, Hall CB, Suffin SC, Alexson C, Harris PJ, Manning JA. Respiratory syncytial viral infection in infants with congenital heart disease. *N Engl J Med.* (1982) 307:397–400. doi: 10.1056/NEJM198208123070702
- Hadzimuratovic E, Dinarevic SM, Hadzimuratovic A. Sepsis in premature newborns with congenital heart disease. *Congenit Heart Dis.* (2010) 5:435– 8. doi: 10.1111/j.1747-0803.2010.00406.x
- Chiang PJ, Hsu JF, Tsai MH, Lien R, Chiang MC, Huang HR, et al. The impact of patent ductus arteriosus in neonates with late onset sepsis: a retrospective matched-case control study. *Pediatr Neonatol.* (2012) 53:309– 14. doi: 10.1016/j.pedneo.2012.07.006
- Geskey JM, Cyran SE. Managing the morbidity associated with respiratory viral infections in children with congenital heart disease. *Int J Pediatr.* (2012) 2012:646780. doi: 10.1155/2012/646780
- 26. Huang R, Zhu L, Guo H, Wang L, Zhang J, Li W, et al. Cellular immunity profile in children with congenital heart disease and bronchopneumonia:

evaluation of lymphocyte subsets and regulatory T cells. *Cent Eur J Immunol.* (2014) 39:488–92. doi: 10.5114/ceji.2014.47734

- Takaya J, Ikemoto Y, Teraguchi M, Nogi S, Kobayashi Y. Plasma nitric oxide products correlate with cardiac index of congenital heart disease. *Pediatr Cardiol.* (2000) 21:378–81. doi: 10.1007/s002460010086
- Rouatbi H, Farhat N, Heying R, Gerard A, Vazquez-Jimenez JF, Seghaye MC. Right atrial myocardial remodeling in children with atrial septal defect involves inflammation, growth, fibrosis, and apoptosis. *Front Pediatr.* (2020) 8:40. doi: 10.3389/fped.2020.00040
- Ramakrishnan S, Kukreti BB, Ramakrishnan L, Salahuddin S, Pendharkar A, Karthikeyan G, et al. Inflammatory markers are elevated in Eisenmenger syndrome. *Pediatr Cardiol.* (2013) 34:1791–6. doi: 10.1007/s00246-013-0715-3
- Moutafi AC, Alissafi T, Chryssanthopoulos S, Thanopoulos V, Tousoulis D, Stefanadis C, et al. Neurohormones, cytokines, and aortic function in children with repaired coarctation of the aorta. *Int J Cardiol.* (2014) 172:e26– 7. doi: 10.1016/j.ijcard.2013.12.091
- Segiet OA, Piecuch A, Mielanczyk L, Michalski M, Nowalany-Kozielska E. Role of interleukins in heart failure with reduced ejection fraction. *Anatol J Cardiol.* (2019) 22:287–99. doi: 10.14744/AnatolJCardiol.2019.32748
- Westman PC, Lipinski MJ, Luger D, Waksman R, Bonow RO, Wu E, et al. Inflammation as a driver of adverse left ventricular remodeling after acute myocardial infarction. J Am Coll Cardiol. (2016) 67:2050– 60. doi: 10.1016/j.jacc.2016.01.073
- Fredj S, Bescond J, Louault C, Delwail A, Lecron JC, Potreau D. Role of interleukin-6 in cardiomyocyte/cardiac fibroblast interactions during myocyte hypertrophy and fibroblast proliferation. J Cell Physiol. (2005) 204:428–36. doi: 10.1002/jcp.20307
- 34. Okada H, Takemura G, Kosai K, Li Y, Takahashi T, Esaki M, et al. Postinfarction gene therapy against transforming growth factor-beta signal modulates infarct tissue dynamics and attenuates left ventricular remodeling and heart failure. *Circulation.* (2005) 111:2430–7. doi: 10.1161/01.CIR.0000165066.71481.8E
- 35. Deten A, Volz HC, Briest W, Zimmer HG. Cardiac cytokine expression is upregulated in the acute phase after myocardial infarction. Experimental studies in rats. *Cardiovasc Res.* (2002) 55:329–40. doi: 10.1016/S0008-6363(02)00413-3
- Nian M, Lee P, Khaper N, Liu P. Inflammatory cytokines and postmyocardial infarction remodeling. *Circ Res.* (2004) 94:1543–53. doi: 10.1161/01.RES.0000130526.20854.fa
- Sun M, Chen M, Dawood F, Zurawska U, Li JY, Parker T, et al. Tumor necrosis factor-alpha mediates cardiac remodeling and ventricular dysfunction after pressure overload state. *Circulation*. (2007) 115:1398– 407. doi: 10.1161/CIRCULATIONAHA.106.643585
- Llanos C, Izmirly PM, Katholi M, Clancy RM, Friedman DM, Kim MY, et al. Recurrence rates of cardiac manifestations associated with neonatal lupus and maternal/fetal risk factors. *Arthritis Rheum.* (2009) 60:3091– 7. doi: 10.1002/art.24768
- 39. Rischmueller M, Tieu J, Lester S. Primary Sjogren's syndrome. *Best Pract Res Clin Rheumatol.* (2016) 30:189–220. doi: 10.1016/j.berh.2016.04.003
- Dhiman N, Haralambieva IH, Vierkant RA, Pankratz VS, Ryan JE, Jacobson RM, et al. Predominant inflammatory cytokine secretion pattern in response to two doses of live rubella vaccine in healthy vaccinees. *Cytokine*. (2010) 50:24–9. doi: 10.1016/j.cyto.2009.12.002
- 41. Geyer H, Bauer M, Neumann J, Ludde A, Rennert P, Friedrich N, et al. Gene expression profiling of rubella virus infected primary endothelial cells of fetal and adult origin. *Virol J.* (2016) 13:21. doi: 10.1186/s12985-016-0475-9
- Lazar M, Perelygina L, Martines R, Greer P, Paddock CD, Peltecu G, et al. Immunolocalization and distribution of rubella antigen in fatal congenital rubella syndrome. *EBioMedicine*. (2016) 3:86–92. doi: 10.1016/j.ebiom.2015.11.050
- Sullivan KE. Chromosome 22q11.2 deletion syndrome: diGeorge syndrome/velocardiofacial syndrome. *Immunol Allergy Clin North Am.* (2008) 28:353–66. doi: 10.1016/j.iac.2008.01.003
- Human A, Murguia-Favela L, Benson L, Roifman I and Grunebaum E. Cardiovascular abnormalities in primary immunodeficiency diseases. *LymphoSign J.* (2015) 2:107–34. doi: 10.14785/lpsn-2014-0013

- Kuo CY, Signer R, Saitta SC. Immune and genetic features of the chromosome 22q11.2 deletion (DiGeorge Syndrome). Curr Allergy Asthma Rep. (2018) 18:75. doi: 10.1007/s11882-018-0823-5
- 46. Lambert MP, Arulselvan A, Schott A, Markham SJ, Crowley TB, Zackai EH, et al. The 22q11.2 deletion syndrome: cancer predisposition, platelet abnormalities and cytopenias. Am J Med Genet A. (2018) 176:2121–7. doi: 10.1002/ajmg.a.38474
- Ram G and Chinen J. Infections and immunodeficiency in Down syndrome. *Clin Exp Immunol.* (2011) 164:9–16. doi: 10.1111/j.1365-2249.2011.04335.x
- Zampieri BL, Biselli-Perico JM, de Souza JE, Burger MC, Silva Junior WA, Goloni-Bertollo EM, et al. Altered expression of immunerelated genes in children with Down syndrome. *PLoS ONE.* (2014) 9:e107218. doi: 10.1371/journal.pone.0107218
- Farroni C, Marasco E, Marcellini V, Giorda E, Valentini D, Petrini S, et al. Dysregulated miR-155 and miR-125b Are Related to Impaired B-cell Responses in Down Syndrome. *Front Immunol.* (2018) 9:2683. doi: 10.3389/fimmu.2018.02683
- Huggard D, Koay WJ, Kelly L, McGrane F, Ryan E, Lagan N, et al. Altered Toll-Like Receptor Signalling in Children with Down Syndrome. *Mediators Inflamm*. (2019) 2019:4068734. doi: 10.1155/2019/4068734
- Stenberg AE, Sylven L, Magnusson CG, Hultcrantz M. Immunological parameters in girls with Turner syndrome. J Negat Results Biomed. (2004) 3:6. doi: 10.1186/1477-5751-3-6
- Su MA, Stenerson M, Liu W, Putnam A, Conte F, Bluestone JA, et al. The role of X-linked FOXP3 in the autoimmune susceptibility of Turner Syndrome patients. *Clin Immunol.* (2009) 131:139–44. doi: 10.1016/j.clim.2008.11.007
- Gawlik AM, Berdej-Szczot E, Blat D, Klekotka R, Gawlik T, Blaszczyk E, et al. Immunological profile and predisposition to autoimmunity in girls with turner syndrome. *Front Endocrinol.* (2018) 9:307. doi: 10.3389/fendo.2018.00307
- Sonesson SE, Salomonsson S, Jacobsson LA, Bremme K, Wahren-Herlenius M. Signs of first-degree heart block occur in one-third of fetuses of pregnant women with anti-SSA/Ro 52-kd antibodies. *Arthritis Rheum.* (2004) 50:1253–61. doi: 10.1002/art.20126
- Brito-Zeron P, Izmirly PM, Ramos-Casals M, Buyon JP, Khamashta MA. The clinical spectrum of autoimmune congenital heart block. *Nat Rev Rheumatol.* (2015) 11:301–12. doi: 10.1038/nrrheum.2015.29
- 56. Buyon JP, Hiebert R, Copel J, Craft J, Friedman D, Katholi M, et al. Autoimmune-associated congenital heart block: demographics, mortality, morbidity and recurrence rates obtained from a national neonatal lupus registry. J Am Coll Cardiol. (1998) 31:1658–66. doi: 10.1016/S0735-1097(98)00161-2
- 57. Izmirly PM, Saxena A, Kim MY, Wang D, Sahl SK, Llanos C, et al. Maternal and fetal factors associated with mortality and morbidity in a multi-racial/ethnic registry of anti-SSA/Ro-associated cardiac neonatal lupus. *Circulation.* (2011) 124:1927–35. doi: 10.1161/CIRCULATIONAHA.111.033894
- Klein-Gitelman MS. Neonatal Lupus: what we have learned and current approaches to care. *Curr Rheumatol Rep.* (2016) 18:60. doi: 10.1007/s11926-016-0610-z
- Moak JP, Barron KS, Hougen TJ, Wiles HB, Balaji S, Sreeram N, et al. Congenital heart block: development of late-onset cardiomyopathy, a previously underappreciated sequela. J Am Coll Cardiol. (2001) 37:238– 42. doi: 10.1016/S0735-1097(00)01048-2
- Eronen M, Sirèn MK, Ekblad H, Tikanoja T, Julkunen H, Paavilainen T. Short- and long-term outcome of children with congenital complete heart block diagnosed in utero or as a newborn. *Pediatrics*. (2000) 106(1 Pt 1):86– 91. doi: 10.1542/peds.106.1.86
- Izmirly PM, Rivera TL, Buyon JP. Neonatal lupus syndromes. *Rheum Dis Clin North Am.* (2007) 33:267–85, vi. doi: 10.1016/j.rdc.2007.02.005
- Frohn-Mulder IM, Meilof JF, Szatmari A, Stewart PA, Swaak TJ, Hess J. Clinical significance of maternal anti-Ro/SS-A antibodies in children with isolated heart block. J Am Coll Cardiol. (1994) 23:1677– 81. doi: 10.1016/0735-1097(94)90674-2
- 63. Salomonsson S, Dorner T, Theander E, Bremme K, Larsson P, Wahren-Herlenius M. A serologic marker for fetal risk of congenital heart block. *Arthritis Rheum.* (2002) 46:1233–41. doi: 10.1002/art.10232

- Brito-Zerón P, Izmirly PM, Ramos-Casals M, Buyon JP, Khamashta MA. Autoimmune congenital heart block: complex and unusual situations. *Lupus*. (2016) 25:116–28. doi: 10.1177/0961203315624024
- Buyon JP, Clancy RM. From antibody insult to fibrosis in neonatal lupus—the heart of the matter. *Arthritis Res Ther.* (2003) 5:266– 70. doi: 10.1186/ar763
- Buyon JP, Clancy RM. Dying right to live longer: positing apoptosis as a link between maternal autoantibodies and congenital heart block. *Lupus*. (2008) 17:86–90. doi: 10.1177/0961203307085115
- Clancy RM, Buyon JP. Autoimmune-associated congenital heart block: dissecting the cascade from immunologic insult to relentless fibrosis. *Anat Rec A Discov Mol Cell Evol Biol.* (2004) 280:1027–35. doi: 10.1002/ar.a.20072
- Xiao GQ, Hu K, Boutjdir M. Direct inhibition of expressed cardiac l- and t-type calcium channels by IgG from mothers whose children have congenital heart block. *Circulation*. (2001) 103:1599–604. doi: 10.1161/01.CIR.103.11.1599
- 69. Horsfall AC, Li JM, Maini RN. Placental and fetal cardiac laminin are targets for cross-reacting autoantibodies from mothers of children with congenital heart block. *J Autoimmun*. (1996) 9:561–8. doi: 10.1006/jaut.1996.0075
- Jaeggi ET, Fouron JC, Silverman ED, Ryan G, Smallhorn J, Hornberger LK. Transplacental fetal treatment improves the outcome of prenatally diagnosed complete atrioventricular block without structural heart disease. *Circulation*. (2004) 110:1542–8. doi: 10.1161/01.CIR.0000142046.58632.3A
- Saleeb S, Copel J, Friedman D, Buyon JP. Comparison of treatment with fluorinated glucocorticoids to the natural history of autoantibody-associated congenital heart block: retrospective review of the research registry for neonatal lupus. *Arthritis Rheum.* (1999) 42:2335–45.
- 72. Izmirly PM, Saxena A, Sahl SK, Shah U, Friedman DM, Kim MY, et al. Assessment of fluorinated steroids to avert progression and mortality in anti-SSA/Ro-associated cardiac injury limited to the fetal conduction system. *Ann Rheum Dis.* (2016) 75:1161–5. doi: 10.1136/annrheumdis-2015-208311
- Breur JM, Visser GH, Kruize AA, Stoutenbeek P, Meijboom EJ. Treatment of fetal heart block with maternal steroid therapy: case report and review of the literature. *Ultrasound Obstet Gynecol.* (2004) 24:467– 72. doi: 10.1002/uog.1713
- Saxena A, Izmirly PM, Mendez B, Buyon JP, Friedman DM. Prevention and treatment in utero of autoimmune-associated congenital heart block. *Cardiol Rev.* (2014) 22:263–7. doi: 10.1097/CRD.00000000000026
- 75. Friedman DM, Llanos C, Izmirly PM, Brock B, Byron J, Copel J, et al. Evaluation of fetuses in a study of intravenous immunoglobulin as preventive therapy for congenital heart block: results of a multicenter, prospective, open-label clinical trial. *Arthritis Rheum.* (2010) 62:1138– 46. doi: 10.1002/art.27308
- Pisoni CN, Brucato A, Ruffatti A, Espinosa G, Cervera R, Belmonte-Serrano M, et al. Failure of intravenous immunoglobulin to prevent congenital heart block: findings of a multicenter, prospective, observational study. *Arthritis Rheum.* (2010) 62:1147–52. doi: 10.1002/art.27350
- 77. Izmirly PM, Costedoat-Chalumeau N, Pisoni CN, Khamashta MA, Kim MY, Saxena A, et al. Maternal use of hydroxychloroquine is associated with a reduced risk of recurrent anti-SSA/Ro-antibody-associated cardiac manifestations of neonatal lupus. *Circulation*. (2012) 126:76– 82. doi: 10.1161/CIRCULATIONAHA.111.089268
- Overall JC Jr. Intrauterine virus infections and congenital heart disease. Am Heart J. (1972) 84:823–33. doi: 10.1016/0002-8703(72)90077-4
- Yazigi A, De Pecoulas AE, Vauloup-Fellous C, Grangeot-Keros L, Ayoubi JM, Picone O. Fetal and neonatal abnormalities due to congenital rubella syndrome: a review of literature. *J Matern Fetal Neonatal Med Heart*. (2017) 30:274–8. doi: 10.3109/14767058.2016.1169526
- Perelygina L, Zheng Q, Metcalfe M, Icenogle J. Persistent infection of human fetal endothelial cells with rubella virus. *PLoS ONE*. (2013) 8:e73014. doi: 10.1371/journal.pone.0073014
- Bilz NC, Willscher E, Binder H, Böhnke J, Stanifer ML, Hübner D, et al. Teratogenic rubella virus alters the endodermal differentiation capacity of human induced pluripotent stem cells. *Cells.* (2019) 8:870. doi: 10.3390/cells8080870
- Gennery AR. Immunological features of 22q11 deletion syndrome. *Curr Opin Pediatr.* (2013) 25:730–5. doi: 10.1097/MOP.0000000000 000027

- Zemble R, Luning Prak E, McDonald K, McDonald-McGinn D, Zackai E and Sullivan K. Secondary immunologic consequences in chromosome 22q11.2 deletion syndrome (DiGeorge syndrome/velocardiofacial syndrome). *Clin Immunol.* (2010) 136:409–18. doi: 10.1016/j.clim.2010.04.011
- Momma K. Cardiovascular anomalies associated with chromosome 22q11.2 deletion syndrome. *Am J Cardiol.* (2010) 105:1617– 24. doi: 10.1016/j.amjcard.2010.01.333
- McDonald-McGinn DM, Sullivan KE, Marino B, Philip N, Swillen A, Vorstman JA, et al. 22q11.2 deletion syndrome. *Nat Rev Dis Primers*. (2015) 1:15071. doi: 10.1038/nrdp.2015.71
- Huggard D, Kelly L, Ryan E, McGrane F, Lagan N, Roche E, et al. Increased systemic inflammation in children with Down syndrome. *Cytokine*. (2020) 127:154938. doi: 10.1016/j.cyto.2019.154938
- Pellegrini FP, Marinoni M, Frangione V, Tedeschi A, Gandini V, Ciglia F, et al. Down syndrome, autoimmunity and T regulatory cells. *Clin Exp Immunol.* (2012) 169:238–43. doi: 10.1111/j.1365-2249.2012.04610.x
- Satge D, Seidel MG. The pattern of malignancies in down syndrome and its potential context with the immune system. *Front Immunol.* (2018) 9:3058. doi: 10.3389/fimmu.2018.03058
- Versacci P, Di Carlo D, Digilio MC, Marino B. Cardiovascular disease in Down syndrome. *Curr Opin Pediatr.* (2018) 30:616– 22. doi: 10.1097/MOP.00000000000661
- Vis JC, Duffels MG, Winter MM, Weijerman ME, Cobben JM, Huisman SA, et al. Down syndrome: a cardiovascular perspective. J Intellect Disabil Res. (2009) 53:419–25. doi: 10.1111/j.1365-2788.2009.01158.x
- Gravholt CH, Viuff MH, Brun S, Stochholm K, Andersen NH. Turner syndrome: mechanisms and management. *Nat Rev Endocrinol.* (2019) 15:601–14. doi: 10.1038/s41574-019-0224-4
- Silberbach M, Roos-Hesselink JW, Andersen NH, Braverman AC, Brown N, Collins RT, et al. Cardiovascular health in turner syndrome: a scientific statement from the american heart association. *Circ Genom Precis Med.* (2018) 11:e000048. doi: 10.1161/HCG.000000000000048
- Volkl TM, Degenhardt K, Koch A, Simm D, Dorr HG, Singer H. Cardiovascular anomalies in children and young adults with Ullrich-Turner syndrome the Erlangen experience. *Clin Cardiol.* (2005) 28:88– 92. doi: 10.1002/clc.4960280209
- 94. Balistreri CR, Buffa S, Allegra A, Pisano C, Ruvolo G, Colonna-Romano G, et al. A typical immune T/B subset profile characterizes bicuspid aortic valve: in an old status? Oxid Med Cell Longev. (2018) 2018:5879281. doi: 10.1155/2018/5879281
- 95. Balistreri CR, Marullo AGM, Madonna M, Cavarretta E, Allegra A, Cesarini V, et al. Deregulation of TLR4 signaling pathway characterizes Bicuspid Aortic valve syndrome. *Sci Rep.* (2019) 9:11028. doi: 10.1038/s41598-019-47412-0
- Jerome LA, Papaioannou VE. DiGeorge syndrome phenotype in mice mutant for the T-box gene, Tbx1. Nature Genetics. (2001) 27:286– 91. doi: 10.1038/85845
- Doyle MJ, Lohr JL, Chapman CS, Koyano-Nakagawa N, Garry MG, Garry DJ. Human induced pluripotent stem cell-derived cardiomyocytes as a model for heart development and congenital heart disease. *Stem Cell Rev Rep.* (2015) 11:710–27. doi: 10.1007/s12015-015-9596-6
- Olivieri LJ, de Zelicourt DA, Haggerty CM, Ratnayaka K, Cross RR, Yoganathan AP. Hemodynamic modeling of surgically repaired coarctation of the Aorta. *Cardiovasc Eng Technol.* (2011) 2:288–95. doi: 10.1007/s13239-011-0059-1
- Marsden AL, Feinstein JA. Computational modeling and engineering in pediatric and congenital heart disease. *Curr Opin Pediatr.* (2015) 27:587– 96. doi: 10.1097/MOP.00000000000269
- 100. Gustafson MP, Lin Y, LaPlant B, Liwski CJ, Maas ML, League SC, et al. Immune monitoring using the predictive power of immune profiles. J Immuno Ther Cancer. (2013) 1:7. doi: 10.1186/2051-1426-1-7
- 101. Robinson GA, Peng J, Donnes P, Coelewij L, Naja M, Radziszewska A, et al. Disease-associated and patient-specific immune cell signatures in juvenile-onset systemic lupus erythematosus: patient stratification using a machine-learning approach. *Lancet Rheumatol.* (2020) 2:e485–96. doi: 10.1016/S2665-9913(20)30168-5
- 102. Midgett M, Lopez CS, David L, Maloyan A, Rugonyi S. Increased hemodynamic load in early embryonic stages alters myofibril and

mitochondrial organization in the myocardium. Front Physiol. (2017) 8:631. doi: 10.3389/fphys.2017.00631

- 103. Oosterbaan AM, Bon E, Steegers-Theunissen RP, Van Der Steen AF, Ursem NT. Homocysteine exposure affects early hemodynamic parameters of embryonic chicken heart function. *Anat Rec.* (2012) 295:961–7. doi: 10.1002/ar.22477
- 104. Majumdar U, Yasuhara J and Garg V. In vivo and in vitro genetic models of congenital heart disease. *Cold Spring Harb Perspect Biol.* (2019) 13:a036764. doi: 10.1101/cshperspect.a036764
- 105. Winston JB, Erlich JM, Green CA, Aluko A, Kaiser KA, Takematsu M, et al. Heterogeneity of genetic modifiers ensures normal cardiac development. *Circulation*. (2010) 121:1313– 21. doi: 10.1161/CIRCULATIONAHA.109.887687
- 106. Rajagopal SK, Ma Q, Obler D, Shen J, Manichaikul A, Tomita-Mitchell A, et al. Spectrum of heart disease associated with murine and human GATA4 mutation. J Mol Cell Cardiol. (2007) 43:677– 85. doi: 10.1016/j.yjmcc.2007.06.004
- 107. MacGrogan D, D'Amato G, Travisano S, Martinez-Poveda B, Luxan G, Del Monte-Nieto G, et al. Sequential Ligand-Dependent Notch Signaling Activation Regulates Valve Primordium Formation and Morphogenesis. *Circ Res.* (2016) 118:1480–97. doi: 10.1161/CIRCRESAHA.115. 308077
- Laforest B, Andelfinger G and Nemer M. Loss of Gata5 in mice leads to bicuspid aortic valve. J Clin Invest. (2011) 121:2876–87. doi: 10.1172/JCI44555
- 109. Sucharov CC, Hijmans JG, Sobus RD, Melhado WF, Miyamoto SD, Stauffer BL. Beta-Adrenergic receptor antagonism in mice: a model for pediatric heart disease. J Appl Physiol (1985). (2013) 115:979– 87. doi: 10.1152/japplphysiol.00627.2013
- 110. Shang F, Wang J, Liu X, Li J, Zheng Q, Xue Y, et al. Involvement of reactive oxygen species and JNK in increased expression of MCP-1 and infiltration of inflammatory cells in pressure-overloaded rat hearts. *Mol Med Rep.* (2012) 5:1491–6. doi: 10.3892/mmr.2012.852

- 111. Ghorbel MT, Jia H, Swim MM, Iacobazzi D, Albertario A, Zebele C, et al. Reconstruction of the pulmonary artery by a novel biodegradable conduit engineered with perinatal stem cellderived vascular smooth muscle cells enables physiological vascular growth in a large animal model of congenital heart disease. *Biomaterials.* (2019) 217:119284. doi: 10.1016/j.biomaterials.2019.11 9284
- 112. Xu M, Jiang Y, Wang J, Liu D, Wang S, Yi H, et al. Distribution of distinct subsets of circulating T follicular helper cells in Kawasaki disease. BMC Pediatr. (2019) 19:43. doi: 10.1186/s12887-019-1412-z
- 113. Johnson Kameny R, Datar SA, Boehme JB, Morris C, Zhu T, Goudy BD, et al. Ovine Models of Congenital Heart Disease and the Consequences of Hemodynamic Alterations for Pulmonary Artery Remodeling. Am J Respir Cell Mol Biol. (2019) 60:503–14. doi: 10.1165/rcmb.2018-0305MA

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