



Article

# Enhancement of the Water Affinity of Histidine by Zinc and Copper Ions

Yongshun Song<sup>1</sup> , Jing Zhan<sup>2</sup>, Minyue Li<sup>2</sup>, Hongwei Zhao<sup>3,4</sup>, Guosheng Shi<sup>2</sup>, Minghong Wu<sup>2,\*</sup> and Haiping Fang<sup>1,5,\*</sup>

<sup>1</sup> School of Physics, East China University of Science and Technology, Shanghai 200237, China; songyongshun11@mailsucas.ac.cn

<sup>2</sup> Shanghai Applied Radiation Institute, Shanghai University, Shanghai 200444, China; zhanjing0215@163.com (J.Z.); 18800313926@163.com (M.L.); gssshi@shu.edu.cn (G.S.)

<sup>3</sup> Zhangjiang Laboratory, Shanghai Advanced Research Institute, Chinese Academy of Sciences, Shanghai 201210, China; zhaohongwei@zjlab.org.cn

<sup>4</sup> Shanghai Institute of Applied Physics, Chinese Academy of Sciences, Shanghai 201800, China

<sup>5</sup> Wenzhou Institute, University of Chinese Academy of Sciences, Wenzhou 325000, China

\* Correspondence: mhwu@shu.edu.cn (M.W.); fanghaiping@sinap.ac.cn (H.F.)

**Abstract:** Histidine (His) is widely involved in the structure and function of biomolecules. Transition-metal ions, such as Zn<sup>2+</sup> and Cu<sup>2+</sup>, widely exist in biological environments, and they are crucial to many life-sustaining physiological processes. Herein, by employing density function calculations, we theoretically show that the water affinity of His can be enhanced by the strong cation- $\pi$  interaction between His and Zn<sup>2+</sup> and Cu<sup>2+</sup>. Further, the solubility of His is experimentally demonstrated to be greatly enhanced in ZnCl<sub>2</sub> and CuCl<sub>2</sub> solutions. The existence of cation- $\pi$  interaction is demonstrated by fluorescence, ultraviolet (UV) spectroscopy and nuclear magnetic resonance (NMR) experiments. These findings are of great importance for the bioavailability of aromatic drugs and provide new insight for understanding the physiological functions of transition metal ions.

**Keywords:** solubility; aromatic amino acids; cation- $\pi$  interaction; transition-metal ions



**Citation:** Song, Y.; Zhan, J.; Li, M.; Zhao, H.; Shi, G.; Wu, M.; Fang, H. Enhancement of the Water Affinity of Histidine by Zinc and Copper Ions. *Int. J. Mol. Sci.* **2022**, *23*, 3957. <https://doi.org/10.3390/ijms23073957>

Academic Editors: Attila Bende and Mihai V. Putz

Received: 16 February 2022

Accepted: 30 March 2022

Published: 2 April 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

Histidine (His) is one of the most essential and naturally occurring aromatic amino acids. As one of the building blocks of proteins, it is commonly involved in the structure and function of proteins, since its side-chain contains an imidazole ring which can easily participate in  $\pi$ - $\pi$  [1,2], hydrogen bond [3,4], coordinate bond [5–8] and cation- $\pi$  interactions [9–11]. For example, the coordination of ligand to the imidazole ring of His can inhibit the enzymic activity of tyrosine hydroxylase [12] and  $\pi$ - $\pi$  stacking between His and Tyr is associated with the mitogenicity function of lectin [13]. Besides, histidine-rich peptides can be incorporated into polymers, liposomes, and proteins, including virus-like particles [14–16].

In many proteins, His is coordinated with transition metal ions, among which zinc and copper are the two with the most occurrence. This coordination is critical to the structures and functions of biomolecules [17–21]. For instance, the carboxylate-histidine-zinc triad is frequently found in zinc proteins and is important in the function of these metalloproteins [22]. The riboflavin binding protein (RBP) can bind copper and form a distinct well-ordered type II site under dialysis conditions [19]. In addition, the zinc-histidine complex had been used as a zinc supplement and was shown better absorption than zinc sulfate in humans [23,24]. The copper-histidine complex has been found in human blood, and has been applied to treat Menkes disease [25].

Most biochemical processes occur in aqueous solution. The dispersion behavior of biomolecules in water is important for their participation in physical, chemical, and

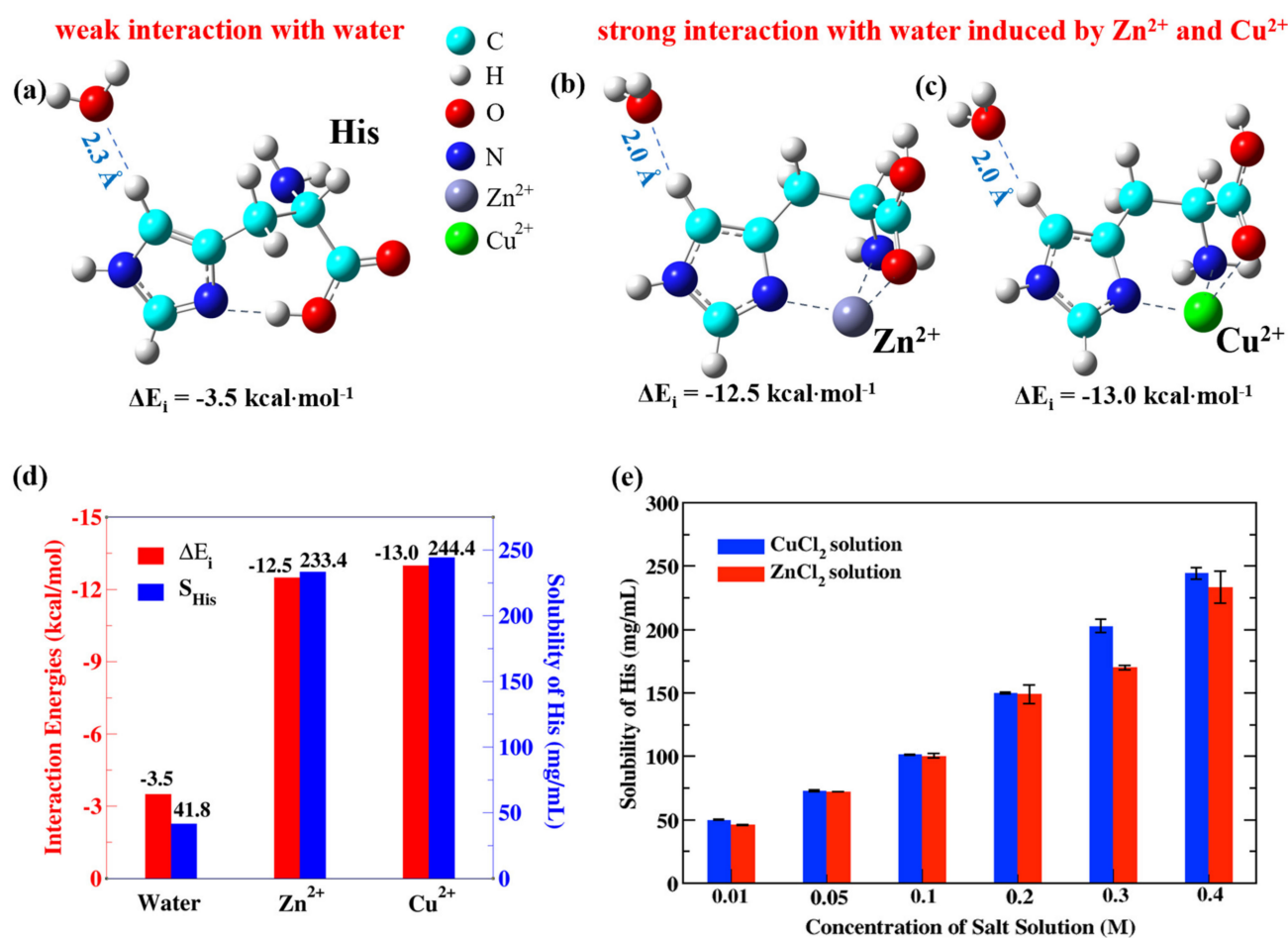
biological processes [26–30]. Previous studies on the coordination of His with transition metal ions are mainly focused on the specific coordination modes and related biological functions. Whereas, studies on some general features of this coordination, such as the hydrophilicity of His, are fewer. By the binding of zinc or copper, the hydrophilicity of His will be modified, which will have significance for a wide range of problems, such as protein folding [31,32], structure stabilization [33,34]. The effect on the hydrophilicity of His by the binding of zinc/copper can be studied through the solubility change for His and complex  $\text{Zn}^{2+}/\text{Cu}^{2+}$ -His. Shi et al. studied the solubility of tryptophan (Trp) in transition metal ion salt solutions and found that the solubility of Trp is dramatically increased [35].

In this paper, by employing DFT calculations, we investigate the interaction between His and transition metal ions  $\text{Zn}^{2+}$  and  $\text{Cu}^{2+}$ . Experiments on the solubility of His in  $\text{ZnCl}_2$  and  $\text{CuCl}_2$  solutions were then performed to verify the results of theoretical calculations. First principles calculations and solubility experiments strongly support that the strong cation– $\pi$  interaction between His and transition metal ions  $\text{Zn}^{2+}$  and  $\text{Cu}^{2+}$  greatly affects the water affinity of His. Finally, the cation– $\pi$  interaction between His and  $\text{Zn}^{2+}/\text{Cu}^{2+}$  was demonstrated by fluorescence, ultraviolet (UV) spectroscopy and nuclear magnetic resonance (NMR) experiments.

## 2. Results and Discussion

To investigate the interaction between His and transition metal ions  $\text{Zn}^{2+}$  and  $\text{Cu}^{2+}$ , we first calculated the interaction energies between the imidazole ring in His with and without  $\text{Zn}^{2+}$  binding and the nearest neighbor water molecule by density functional theory (DFT). As shown in Figure 1b, when  $\text{Zn}^{2+}$  interacts with His, the interaction energy between the imidazole ring in His and the nearest water molecule is about  $-12.5 \text{ kcal}\cdot\text{mol}^{-1}$ , which is much larger than the corresponding interaction energy without  $\text{Zn}^{2+}$  binding ( $-3.5 \text{ kcal}\cdot\text{mol}^{-1}$ , Figure 1a). In addition, we also calculated the interaction energy between the imidazole ring in His and the nearest water molecule with  $\text{Cu}^{2+}$  binding, which is about  $-13.0 \text{ kcal}\cdot\text{mol}^{-1}$  (Figure 1c). These results show that both  $\text{Zn}^{2+}$  and  $\text{Cu}^{2+}$  greatly increase the water affinity of His to about the same degree.

It should be pointed out that in the most stable structures of  $\text{Zn}^{2+}$ -His and  $\text{Cu}^{2+}$ -His,  $\text{Zn}^{2+}$  and  $\text{Cu}^{2+}$  interact with the side of the imidazole ring together with the amino N atom and carbonyl O atom rather than at the top of the imidazole ring. The interaction energy of this ring/N/O tridentate coordination is  $20.3 \text{ kcal}\cdot\text{mol}^{-1}$  larger than that of the structure in which  $\text{Zn}^{2+}$  stays at the top of the imidazole ring (Figure S1). Rimola et al. also pointed out that the configuration of  $\text{Cu}^{2+}$ -His with  $\text{Cu}^{2+}$  at the top of the imidazole ring was not obtainable by first principle calculations [36]. Besides, Sastry et al. calculated the interaction between multiple cations and imidazole, finding that cations show affinity only toward the heteroatom N atom instead of the whole  $\pi$ -face of the imidazole ring [9]. Thus, the configuration of  $\text{Zn}^{2+}/\text{Cu}^{2+}$  stays at the top of the imidazole ring, which interacts with a traditional cation– $\pi$  interaction way, could not be the real complex in the experiments. For the configuration of ring/N/O tridentate coordination, it is expected that there is also a cation– $\pi$  interaction between  $\text{Zn}^{2+}/\text{Cu}^{2+}$  and His, because the  $\pi$  electron of imidazole ring and valence electron of  $\text{Zn}^{2+}/\text{Cu}^{2+}$  are both involved in the HOMO-1 orbital (Figure S2) and the electron distribution of the whole imidazole ring is affected by the  $\text{Zn}^{2+}$  binding (Figure S3). To further analyze the cation– $\pi$  interactions, we provide EDA calculations on the  $\text{Zn}^{2+}/\text{Cu}^{2+}$ -His complex. EDA can divide the total interaction energy into several interaction terms with physical meaning and is widely used for analyzing various intermolecular interactions [37,38]. We divided the optimized structure of  $\text{Zn}^{2+}/\text{Cu}^{2+}$ -His into two parts, i.e., the  $\text{Zn}^{2+}/\text{Cu}^{2+}$  ion and His, and then performed single-point calculations at the B3LYP/TZP level of theory in the framework of DFT by using the ADF program [39]. The total interaction energies between  $\text{Zn}^{2+}/\text{Cu}^{2+}$  and His were decomposed into Pauli repulsion, orbital interaction and electrostatic interaction (Table S1). The EDA results show that the electrostatic interactions and orbital interactions approximately equally contribute to the interaction energy, and the Pauli repulsion affords the main repulsive energy.



**Figure 1.** Interaction energies between His,  $\text{Zn}^{2+}$ -His,  $\text{Cu}^{2+}$ -His and the nearest water and the solubilities of His in  $\text{ZnCl}_2$  and  $\text{CuCl}_2$  solutions. (a–c) Optimized geometric structures of the His–water,  $\text{Zn}^{2+}$ -His–water, and  $\text{Cu}^{2+}$ -His–water systems and the interaction energies between the imidazole rings and the nearest water molecules. (a) Optimized distance between the imidazole ring in His and the nearest water molecule is 2.3 Å, and the interaction energy between them is  $-3.5 \text{ kcal}\cdot\text{mol}^{-1}$ . (b) Optimized distance between the imidazole ring and the nearest water molecule after  $\text{Zn}^{2+}$  binding decreases to 2.0 Å, and the interaction energy increases to  $-12.5 \text{ kcal}\cdot\text{mol}^{-1}$ . (c) Optimized distance between the imidazole ring in His and the nearest water molecule after  $\text{Cu}^{2+}$  binding decreases to 2.0 Å, and the interaction energy increases to  $-13.0 \text{ kcal}\cdot\text{mol}^{-1}$ . (d) Interaction energies ( $\Delta E_i$ ) between the imidazole ring in His and the nearest water molecule, without binding to any metal ion, with  $\text{Zn}^{2+}$  binding, and with  $\text{Cu}^{2+}$  binding and the solubilities of His ( $S_{\text{His}}$ ) in pure water, 0.4 M  $\text{ZnCl}_2$  solution, and 0.4 M  $\text{CuCl}_2$  solution. (e) Solubilities of His in  $\text{CuCl}_2$  and  $\text{ZnCl}_2$  solutions.

We also would like to note that the geometric structures of the  $\text{Zn}^{2+}$ -His–water and  $\text{Cu}^{2+}$ -His–water systems were optimized based on the water molecule forming a hydrogen bond with the imidazole ring at the C–H site (Figure 1b,c). In fact, water can interact with the imidazole ring of  $\text{Zn}^{2+}$ -His and  $\text{Cu}^{2+}$ -His complexes at other sites, for example, N–H of the imidazole ring (shown in Figure S4), which also indicate that the water affinity of imidazole ring is increased. Since N–H is not as hydrophobic as C–H in His, the increase of water affinity of N–H is not as critical as C–H for the water affinity enhancement of the whole complex.

The water affinity is well related to solubility. A weak interaction with water indicates that hydration of the compound is unfavorable, contributing to poor water solubility. Because the water affinities of  $\text{Zn}^{2+}$ -His and  $\text{Cu}^{2+}$ -His are much larger than that of His, we expected that the solubilities of His in  $\text{ZnCl}_2$  and  $\text{CuCl}_2$  solutions would both dramatically

increase. We then performed experiments on the solubilities of His ( $S_{\text{His}}$ ) in  $\text{ZnCl}_2$  and  $\text{CuCl}_2$  aqueous solutions. The solubility of His in  $\text{ZnCl}_2$  solution increases as much as that in  $\text{CuCl}_2$  solution (Figure 1d). In 0.4 M  $\text{ZnCl}_2$  and  $\text{CuCl}_2$  solutions, the solubility of His can increase to more than 5 times that in pure water. This is consistent with the first principles calculations of the interaction energy between the aromatic ring structure in His and the nearest water molecule. The results of His were then compared with previous work on Trp to show the enhanced water affinity of His by  $\text{Zn}^{2+}$  is nontrivial. Previous work on Trp [35] showed that the solubility of Trp in  $\text{ZnCl}_2$  solution slightly increases compared with the solubility in water ( $17.1 \text{ mg}\cdot\text{mL}^{-1}$  in 0.5 M  $\text{ZnCl}_2$  solution vs.  $11.4 \text{ mg}\cdot\text{mL}^{-1}$  in pure water, Table 1). These results indicate that the aromatic amino acid His has a binding preference for  $\text{Zn}^{2+}$  compared with other amino acids and the interaction between them can greatly enhance the water affinity of His.

**Table 1.** Solubilities of His and Trp in pure water,  $\text{ZnCl}_2$  and  $\text{CuCl}_2$ .

		Pure Water	$\text{ZnCl}_2$	$\text{CuCl}_2$
Solubility ( $\text{mg}\cdot\text{mL}^{-1}$ )	His	41.9	233.4 <sup>1</sup>	244.4 <sup>1</sup>
	Trp	11.4	17.1 <sup>2</sup>	57.6 <sup>2</sup>

<sup>1</sup> Salt concentration 0.4 M. <sup>2</sup> Salt concentration 0.5 M.

We would like to point out that there is no obvious difference when different orders of adding cation ions and His are employed. Both orders will lead to the same result that the solubility of His is enhanced significantly. This is a major difference between His in this work and Trp in previous work [35], where the order of adding cations and Trp show totally different results for the solubility enhancement of Trp.

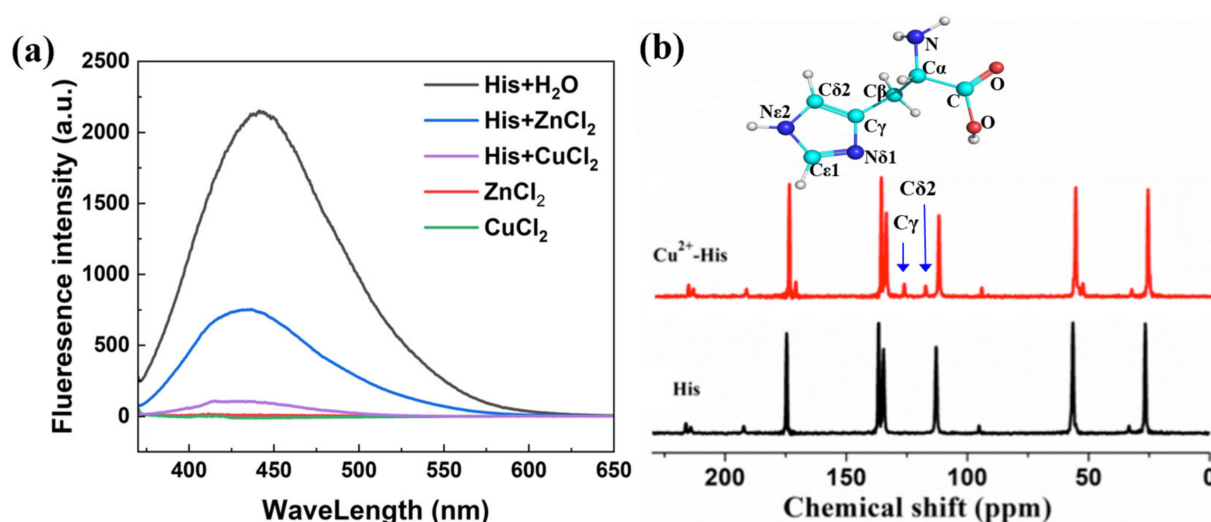
The solubility behavior of His with respect to the concentrations of  $\text{ZnCl}_2$  and  $\text{CuCl}_2$  solutions was then investigated. As shown in Figure 1e and Figure S5, the solubility behavior can be well fitted by  $S_A = A_M C_M + S_A^0$ , where  $S_A$  and  $S_A^0$  are the solubilities of amino acid  $A$  in the salt solution and pure water,  $A_M$  is the water affinity factor of amino acid  $A$  induced by metal ion  $M^{2+}$ , and  $C_M$  is the concentration of  $M^{2+}$ .  $A_M$  has a distinct physical meaning that for every  $M$  salt concentration increase, the solubility of the amino acid increases by  $A_M M$ . For His,  $A_{\text{Cu}} = 4.86$  and  $A_{\text{Zn}} = 2.80$ , which are both much greater than the water affinity factor of Trp induced by  $\text{Cu}^{2+}$  ( $A_{\text{Cu}} = 0.46$ ) [35].

It should be noted that different  $\text{Zn}^{2+}$ -His and  $\text{Cu}^{2+}$ -His complexes exist at different  $p\text{H}$  values in experiments [40]. The structure that we used for the DFT calculations was referred to as the  $\text{M}(\text{HL})^{2+}$  species in refs [40,41]. Under weak acid conditions, this structure of tridentate coordination is the most accepted one [42,43]. Indeed, the  $p\text{H}$  values of the  $\text{ZnCl}_2$  and  $\text{CuCl}_2$  solutions used in the experiments correspond to the weak acid condition (5.24–5.75 for the  $\text{ZnCl}_2$  solutions and 3.04–5.75 for the  $\text{CuCl}_2$  solutions) (Tables S2 and S3).

To verify that the increase of the solubility mainly comes from the cation- $\pi$  interaction between the transition-metal ion and the imidazole ring in His, the solubility of the non-aromatic amino acid glycine (Gly) was measured under the same conditions. In Figure S5, it can be found that the solubilities of Trp and His both dramatically increased in  $\text{CuCl}_2$  solutions, while the solubility of Gly only showed a slight increase. This result strongly supports that the enhanced solubility of His can be mainly attributed to the imidazole ring in His.  $\text{Zn}^{2+}$  is expected to show the same behavior.

Fluorescence and ultraviolet (UV) absorption spectral experiments were then performed to provide evidence for the cation- $\pi$  interaction between the metal ion and the imidazole ring in His. In Figure 2a, the fluorescence spectrum of His excited at 360 nm has an emission peak at 448 nm, which is assigned to the conjugated double bonds of the imidazole ring that can easily generate the  $\pi$ - $\pi^*$  transition [44]. Compared with the fluorescence intensity of His in water, the intensity of His in 50 mM  $\text{CuCl}_2$  solution markedly decreased, indicating that the conjugated double bonds of the imidazole ring in His were greatly affected in  $\text{CuCl}_2$  solution. The fluorescence spectrum of His in  $\text{ZnCl}_2$  solution is also shown in Figure 2a. Again, the fluorescence intensities of His and  $\text{ZnCl}_2$  were quenched

when the  $\text{Zn}^{2+}$ -His complex was formed, but to a smaller degree. The reason for the difference of the quenching degree in fluorescence is beyond the scope of this study. These results indicate that in  $\text{ZnCl}_2$  and  $\text{CuCl}_2$  solutions,  $\text{Zn}^{2+}$  and  $\text{Cu}^{2+}$  directly interact with the imidazole ring in His and decrease the fluorescence intensity of His. The imidazole ring of His exhibits absorption in the lower UV region (220 nm) [45]. Here, we observed that the UV absorption spectrum of His was also affected by the cation- $\pi$  interaction between the imidazole ring in His and  $\text{Cu}^{2+}$  in solution (Figure S6), which is important evidence for the existence of the cation- $\pi$  interaction. Overall, the fluorescence and UV absorption spectral experiments show the existence of the cation- $\pi$  interaction between the imidazole ring in His and the metal ion, which is consistent with the theoretical prediction.



**Figure 2.** Fluorescence and NMR spectra on His in different solutions. (a) Fluorescence spectra of His in pure water, His in  $\text{ZnCl}_2$  solution, His in  $\text{CuCl}_2$  solutions (50 mM),  $\text{ZnCl}_2$  solution and  $\text{CuCl}_2$  solution. The concentrations of His in all the solutions are  $30 \text{ mg}\cdot\text{mL}^{-1}$ . The concentrations of  $\text{ZnCl}_2$  and  $\text{CuCl}_2$  are 50 mM in all the samples. (b) Cross-polarization/magic angle spinning (CP/MAS) NMR C12 spectra of  $\text{Cu}^{2+}$ -His and the control group (His in pure water). The C $\delta$ 2 and C $\gamma$  peaks are labeled.

Nuclear magnetic resonance (NMR) experiments on C12 also showed that  $\text{Cu}^{2+}$ -His had a clear chemical shift at 130 ppm, which is attributed to the imidazole ring in His (Figure 2b) [46].  $\text{Zn}^{2+}$ -His also showed a chemical shift at 130 ppm, although with a weaker signal (data not shown).

The structure of His did not change much after the binding of the transition metal ions  $\text{Zn}^{2+}$  and  $\text{Cu}^{2+}$ . Infrared (IR) spectra of the dried samples of His in  $\text{ZnCl}_2$  and  $\text{CuCl}_2$  solutions were similar to that in pure water (Figure S7) [47,48].

The enhancement of  $S_{\text{His}}$  in  $\text{ZnCl}_2$  and  $\text{CuCl}_2$  solutions does not come from the  $p\text{H}$  effect induced by hydrolysis of  $\text{Zn}^{2+}$  or  $\text{Cu}^{2+}$ . As shown in Tables S2 and S3, the  $p\text{H}$  values of the  $\text{ZnCl}_2$  and  $\text{CuCl}_2$  solutions both slightly decreased (from 6.0 to 5.2 for  $\text{Zn}^{2+}$  and from 4.5 to 3.1 for  $\text{Cu}^{2+}$ ), indicating that the  $p\text{H}$  effect does not greatly contribute to the increased solubility of His.

### 3. Materials and Methods

#### 3.1. Computational Methods

The B3LYP functional [49] in the framework of DFT is used to calculate the  $\text{Zn}^{2+}$ -His- $\text{H}_2\text{O}$  and  $\text{Cu}^{2+}$ -His- $\text{H}_2\text{O}$  systems with Gaussian 09 package [50]. Geometric structures were first optimized by Berny algorithm [51] with the convergence criteria of a maximum step size of 0.0018 au and a root mean square force of 0.0003 a.u. A hybrid pseudo potential



LanL2DZ is employed to calculate  $\text{Cu}^{2+}$  and  $\text{Zn}^{2+}$ , while other atoms are calculated at the 6-31+G(d,p) basis set level (see detailed methods in the Supplementary Material (SM)).

Interaction energies between water molecule and aromatic ring of amino acid (AA) with and without metal ion (M) binding are represented as  $\Delta E_i$ , which can be calculated as,

$$\Delta E_i = E_{\text{Total}} - E_{\text{complex}} - E_w, \quad (1)$$

where,  $E_{\text{Total}}$ ,  $E_{\text{complex}}$ , and  $E_w$  are the single-point energies of His- $\text{M}^{2+}$ - $\text{H}_2\text{O}$  (or His- $\text{H}_2\text{O}$ ), complex His- $\text{M}^{2+}$  (or His), and  $\text{H}_2\text{O}$ , respectively.

All the calculations are performed in vacuum condition. There is no other water molecule and ion except the nearest water and the bound ion we considered explicitly.

### 3.2. Experiments Materials

L-Histidine (His, 99%) and Glycine (Gly, 99%) were purchased from Sigma-Aldrich, Shanghai, China. Zinc(II) chloride ( $\text{ZnCl}_2$ , 98%) was purchased from J&K Scientific Ltd, Beijing, China. Copper(II) chloride dihydrate ( $\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$ ) (99%) was purchased from Sinopharm Chemical Reagent Co., Ltd, Beijing, China. All samples were used without purification and preprocessing. All salt solutions and other aqueous solutions were prepared with 18.2 M $\Omega$ , 3 ppb TOC Milli-Q water (Millipore, Burlington, MA, USA).

### 3.3. Solubility Measurement

Histidine (or Glycine) powder was added into pure water (or into the given 0.01–0.4 M  $\text{CuCl}_2$  and  $\text{ZnCl}_2$  solutions) with constant shaking until apparent saturation (with some insoluble His (or Gly) powder appeared), and then the solution was kept continuously stirring for 24 h in a thermostat at 25 °C. Afterward, the solution was centrifuged at 25 °C to remove insoluble His (or Gly) powder. Then, approximately 1 mL of solution was withdrawn by a pipette from the supernatant phase and transferred to a clean and weighed centrifuge tube. These centrifuge tubes were then transferred to liquid nitrogen for freezing and lyophilized overnight in a vacuum flask at 0.125 mbar and –58 °C in a freeze-dryer (Virtis Freezer Dryer, New York, NY, USA). The drying process was repeated until a constant mass reading was achieved. The solubility of His (or Gly) was calculated by the mass value difference of the centrifuge tube after removing salt mass. The data reported in this work were ensured by measurement of solubility for at least three replicate experiments at all compositions (Tables S2 and S3). Based on our measurement strategy, the solubility of His in pure water is 41.9 mg·mL<sup>-1</sup>, consistent with previous reports [52].

### 3.4. Measurement of pH

The pH values were measured by SevenCompact™ S220 pH meter (METTLER TOLEDO, Zurich, Switzerland) (pH = 0–14).

### 3.5. UV Spectroscopy

UV absorption spectra of His,  $\text{CuCl}_2$ , and His ( $\text{CuCl}_2$ ) solutions were recorded on a U-3100 spectrophotometer (Hitachi, Tokyo, Japan). The concentration for His used in this experiment is 30 mg·mL<sup>-1</sup>, for  $\text{CuCl}_2$  is 50 mM.

### 3.6. Fluorescence Spectrofluorophotometer

Excitation and photoluminescence (PL) spectra were measured with a Hitachi 7000 fluorescence spectrophotometer (Tokyo, Japan) and emission slit width of 10 nm was used to record fluorescence spectra, and the fluorescence spectra of the work were recorded with  $\lambda_{\text{ex}}/\lambda_{\text{em}} = 360 \text{ nm}/440 \text{ nm}$ . The thickness of all liquid sample cells is 10 mm.

### 3.7. IR Spectra

IR spectra from 4000 cm<sup>-1</sup> to 500 cm<sup>-1</sup> of His,  $\text{Zn}^{2+}$ -His, and  $\text{Cu}^{2+}$ -His powder were measured using a Bio-Rad FTIR spectrometer FTS165 (Benton, ME, USA) equipped with

resolution of  $4\text{ cm}^{-1}$ . The drying His,  $\text{Zn}^{2+}$ -His, and  $\text{Cu}^{2+}$ -His powder were obtained by freezing and lyophilized from corresponding saturated solution.

### 3.8. Solid-State NMR Spectroscopy

Solid-state NMR experiments were carried out on a wide-bore Bruker AVANCE-600 spectrometer (14.1 T) and a Bruker DSX-400 spectrometer (Karlsruhe, Germany) on 4-mm triple-resonance MAS probes. The drying His,  $\text{Zn}^{2+}$ -His, and  $\text{Cu}^{2+}$ -His powders were obtained by freezing and lyophilized from corresponding saturated solutions.

## 4. Conclusions

In summary, by first principles calculations, we have shown that the cation- $\pi$  interaction between  $\text{Zn}^{2+}$  and His is very strong, which can enhance the water affinity of His to a comparable extent to  $\text{Cu}^{2+}$ . Theoretical studies showed that the strong cation- $\pi$  interaction between  $\text{Zn}^{2+}/\text{Cu}^{2+}$  and His is the key. This cation- $\pi$  interaction modifies the electronic distribution of the imidazole ring in His and significantly enhances the water affinity of His. Further EDA analysis showed that this cation- $\pi$  interaction is approximately contributed equally by electrostatic interactions and orbital interactions. We also performed solubility experiments, which showed that the solubilities of His in  $\text{ZnCl}_2$  and  $\text{CuCl}_2$  solutions can reach more than 5 times that of His in pure water.

The results highlight that the solubility enhancement of many imidazole derivatives by  $\text{Zn}^{2+}$  and  $\text{Cu}^{2+}$  may be a general phenomenon and needs to attract more attention in different research fields, such as drug chemistry and colloid chemistry. These findings will enrich the understanding of the interactions between metal ions and biomolecules, and provide new insight into the physiological functions of multivalent metal ions.

We also would like to note that, our previous study on the solubility of Trp showed that  $\text{Zn}^{2+}$  is relatively weak in affecting the hydrophobicity of Trp by cation- $\pi$  interaction, which is about one third of the solubility enhancement of  $\text{Cu}^{2+}$  [35]. Combined with this study, we found that the cation- $\pi$  interaction is sensitive to the specific interaction environment, providing a possible scheme for the selectivity mechanism of biomolecules. Actually, Tu et al. recently showed that the cation- $\pi$  interaction is the origin of the selectivity mechanism of calcium and magnesium in phosphotyrosine, demonstrating the sensitivity of cation- $\pi$  interaction to different ion species with identical charges [53]. Considering the common binding mode of zinc-histidine in biology, it is expected histidine will participate in the selectivity mechanism of transition metal ions.

**Supplementary Materials:** The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/ijms23073957/s1>. Reference [54] is cited in Supplementary Materials.

**Author Contributions:** Conceptualization, H.F. and G.S.; methodology, J.Z. and M.L.; Data curation and formal analysis, Y.S. and J.Z.; investigation, Y.S. and J.Z.; resources, H.F. and M.W.; writing—original draft preparation, Y.S.; writing—review and editing, H.F., G.S. and H.Z.; project administration, H.F. and M.W. All authors have read and agreed to the published version of the manuscript.

**Funding:** This work was financially supported by the National Natural Science Foundation of China (Grant Nos. 11974366 and 62075225), the Fundamental Research Funds for the Central Universities, China, the National Science Fund for Outstanding Young Scholars (Grant No. 11722548), the Key Research Program of Frontier Sciences of the Chinese Academy of Sciences (No. QYZDJ-SSW-SLH053), and the National Defense Science and Technology Innovation Special Zone Project.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Not applicable.

**Acknowledgments:** We thank Xiaoling Lei, Shiqi Sheng, Shanshan Liang for their constructive suggestions and helpful discussions. We are also thankful for the discussions with the staff from the BL06B beamline of the Shanghai Synchrotron Radiation Facilities(SSRF). We thank the Shanghai

Supercomputer Center of China. We also thank Tim Cooper for editing the English text of a draft of this manuscript.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Sundberg, R.J.; Martin, R.B. Interactions of histidine and other imidazole derivatives with transition metal ions in chemical and biological systems. *Chem. Rev.* **1974**, *74*, 471–517. [\[CrossRef\]](#)
2. Wang, L.; Sun, N.; Terzyan, S.; Zhang, X.; Benson, D.R. A histidine/tryptophan  $\pi$ -stacking interaction stabilizes the heme-independent folding core of microsomal apocytochrome b5 relative to that of mitochondrial apocytochrome b5. *Biochemistry* **2006**, *45*, 13750–13759. [\[CrossRef\]](#) [\[PubMed\]](#)
3. Henry, M. Thermodynamics of hydrogen bond patterns in supramolecular assemblies of water molecules. *ChemPhysChem* **2002**, *3*, 607–616. [\[CrossRef\]](#)
4. Graham, B.; Comba, P.; Hearn, M.T.W.; Spiccia, L. An examination of the binding behavior of histidine-containing peptides with immobilized metal complexes derived from the macrocyclic ligand, 1,4,7-triazacyclononane. *J. Biol. Inorg. Chem.* **2007**, *12*, 11–21. [\[CrossRef\]](#)
5. Miessler, G.L.; Tarr, D.A. *Inorganic Chemistry*, 3rd ed.; Pearson Prentice Hall: Upper Saddle River, NJ, USA, 2003.
6. Ginotra, Y.P.; Kulkarni, P.P. Solution structure of physiological Cu(His)<sub>2</sub>: Novel considerations into imidazole coordination. *Inorg. Chem.* **2009**, *48*, 7000–7002. [\[CrossRef\]](#)
7. Annual Review of Biophysics; Watly, J.; Simonovsky, E.; Wieczorek, R.; Barbosa, N.; Miller, Y.; Kozlowski, H. Insight into the coordination and the binding sites of Cu<sup>2+</sup> by the histidyl-6-tag using experimental and computational tools. *Inorg. Chem.* **2014**, *53*, 6675–6683.
8. Milorey, B.; Malyshka, D.; Schweitzer-Stenner, R. pH dependence of ferricytochrome c conformational transitions during binding to cardiolipin membranes: Evidence for histidine as the distal ligand at neutral pH. *J. Phys. Chem. Lett.* **2017**, *8*, 1993–1998. [\[CrossRef\]](#)
9. Reddy, A.S.; Sastry, G.N. Cation [M = H<sup>+</sup>, Li<sup>+</sup>, Na<sup>+</sup>, K<sup>+</sup>, Ca<sup>2+</sup>, Mg<sup>2+</sup>, NH<sub>4</sub><sup>+</sup>, and NMe<sub>4</sub><sup>+</sup>] interactions with the aromatic motifs of naturally occurring amino acids: A theoretical study. *J. Phys. Chem. A* **2005**, *109*, 8893–8903. [\[CrossRef\]](#)
10. Dougherty, D.A. Cation- $\pi$  interactions in chemistry and biology: A new view of benzene, Phe, Tyr, and Trp. *Science* **1996**, *271*, 163–168. [\[CrossRef\]](#)
11. Liao, S.; Du, Q.; Meng, J.; Pang, Z.; Huang, R. The multiple roles of histidine in protein interactions. *Chem. Cent. J.* **2013**, *7*, 44. [\[CrossRef\]](#)
12. Martínez, A. Evidence for a functionally important histidine residue in human tyrosine hydroxylase. *Amino Acids* **1995**, *9*, 285–292. [\[CrossRef\]](#)
13. Covés-Datson, E.M.; King, S.R.; Legendre, M.; Swanson, M.D.; Gupta, A.; Claes, S.; Meagher, J.L.; Boonen, A.; Zhang, L.; Kalveram, B.; et al. Targeted disruption of  $\pi$ - $\pi$ stacking in Malaysian banana lectin reduces mitogenicity while preserving antiviral activity. *Sci. Rep.* **2021**, *11*, 656. [\[CrossRef\]](#)
14. Chang, K.-L.; Higuchi, Y.; Kawakami, S.; Yamashita, F.; Hashida, M. Development of lysine-histidine dendron modified chitosan for improving transfection efficiency in HEK293 cells. *J. Control. Release* **2011**, *156*, 195–202. [\[CrossRef\]](#)
15. Gu, J.; Wang, X.; Jiang, X.; Chen, Y.; Chen, L.; Fang, X.-L.; Sha, X. Self-assembled carboxymethyl poly (L-histidine) coated poly (+<sup>2</sup>-amino ester)/DNA complexes for gene transfection. *Biomaterials* **2012**, *33*, 644–658. [\[CrossRef\]](#)
16. Ferrer-Miralles, N.; Corchero, J.L.; Kumar, P.; Cedano, J.A.; Gupta, K.C.; Villaverde, A.; Vazquez, E. Biological activities of histidine-rich peptides; merging biotechnology and nanomedicine. *Microb. Cell Fact.* **2011**, *10*, 101. [\[CrossRef\]](#)
17. Berg, J.M.; Shi, Y. The galvanization of biology: A growing appreciation for the roles of zinc. *Science* **1996**, *271*, 1081–1085. [\[CrossRef\]](#)
18. Hoch, E.; Lin, W.; Chai, J.; Hershinkel, M.; Fu, D.; Sekler, I. Histidine pairing at the metal transport site of mammalian ZnT transporters controls Zn<sup>2+</sup> over Cd<sup>2+</sup> selectivity. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 7202. [\[CrossRef\]](#)
19. Smith, S.R.; Bencze, K.Z.; Russ, K.A.; Wasiukanis, K.; Benore-Parsons, M.; Stemmler, T.L. Investigation of the Copper Binding Site and the Role of Histidine as a Ligand in Riboflavin Binding Protein. *Inorg. Chem.* **2008**, *47*, 6867–6872. [\[CrossRef\]](#)
20. Higuchi, Y.; Suzuki, T.; Arimori, T.; Ikemura, N.; Mihara, E.; Kirita, Y.; Ohgitani, E.; Mazda, O.; Motooka, D.; Nakamura, S.; et al. Engineered ACE2 receptor therapy overcomes mutational escape of SARS-CoV-2. *Nat. Commun.* **2021**, *12*, 3802. [\[CrossRef\]](#)
21. Thompsett, A.R.; Abdelraheim, S.R.; Daniels, M.; Brown, D.R. High Affinity Binding between Copper and Full-length Prion Protein Identified by Two Different Techniques\*. *J. Biol. Chem.* **2005**, *280*, 42750–42758. [\[CrossRef\]](#)
22. Christianson, D.W.; Alexander, R.S. Carboxylate-histidine-zinc interactions in protein structure and function. *J. Am. Chem. Soc.* **1989**, *111*, 6412–6419. [\[CrossRef\]](#)
23. Schölmerich, J.; Krauss, E.; Wietholtz, H.; Köttgen, E.; Löhle, E.; Gerok, W. Bioavailability of zinc from zinc-histidine complexes. II Studies on patients with liver cirrhosis and the influence of the time of application. *Am. J. Clin. Nutr.* **1987**, *45*, 1487–1491. [\[CrossRef\]](#)
24. Schölmerich, J.; Freudemann, A.; Köttgen, E.; Wietholtz, H.; Steiert, B.; Löhle, E.; Häussinger, D.; Gerok, W. Bioavailability of zinc from zinc-histidine complexes. I. Comparison with zinc sulfate in healthy men. *Am. J. Clin. Nutr.* **1987**, *45*, 1480–1486. [\[CrossRef\]](#)



25. Sarkar, B. Treatment of Wilson and Menkes Diseases. *Chem. Rev.* **1999**, *99*, 2535–2544. [[CrossRef](#)]
26. Bahng, J.H.; Yeom, B.; Wang, Y.C.; Tung, S.O.; Hoff, J.D.; Kotov, N. Anomalous dispersions of ‘hedgehog’ particles. *Nature* **2015**, *517*, 596–599. [[CrossRef](#)]
27. Babu, N.J.; Nangia, A. Solubility advantage of amorphous drugs and pharmaceutical cocrystals. *Cryst. Growth Des.* **2011**, *11*, 2662–2679. [[CrossRef](#)]
28. Ashbaugh, H.S.; Pratt, L.R. Colloquium: Scaled particle theory and the length scales of hydrophobicity. *Rev. Mod. Phys.* **2006**, *78*, 159–178. [[CrossRef](#)]
29. Li, Z.; Loh, X.J. Water soluble polyhydroxyalkanoates: Future materials for therapeutic applications. *Chem. Soc. Rev.* **2015**, *44*, 2865–2879. [[CrossRef](#)]
30. Schaper, L.-A.; Hock, S.J.; Herrmann, W.A.; Kühn, F.E. Synthesis and application of water-soluble NHC transition-metal complexes. *Angew. Chem. Int. Ed.* **2013**, *52*, 270–289. [[CrossRef](#)]
31. Dill, K.A.; MacCallum, J.L. The protein-folding problem, 50 years on. *Science* **2012**, *338*, 1042–1046. [[CrossRef](#)]
32. Wei, G.; Xi, W.; Nussinov, R.; Ma, B. Protein ensembles: How does nature harness thermodynamic fluctuations for life? The diverse functional roles of conformational ensembles in the cell. *Chem. Rev.* **2016**, *116*, 6516–6551. [[CrossRef](#)] [[PubMed](#)]
33. Berne, B.J.; Weeks, J.D.; Zhou, R. Dewetting and hydrophobic interaction in physical and biological systems. *Annu. Rev. Phys. Chem.* **2009**, *60*, 85–103. [[CrossRef](#)] [[PubMed](#)]
34. Bonn, D.; Eggers, J.; Indekeu, J.; Meunier, J.; Rolley, E. Wetting and spreading. *Rev. Mod. Phys.* **2009**, *81*, 739–805. [[CrossRef](#)]
35. Shi, G.; Dang, Y.; Pan, T.; Liu, X.; Liu, H.; Li, S.; Zhang, L.; Zhao, H.; Li, S.; Han, J.; et al. Unexpectedly enhanced solubility of aromatic amino acids and peptides in an aqueous solution of divalent transition-metal cations. *Phys. Rev. Lett.* **2016**, *117*, 238102. [[CrossRef](#)] [[PubMed](#)]
36. Rimola, A.; Rodríguez-Santiago, L.; Sodupe, M. Cation- $\pi$  interactions and oxidative effects on Cu<sup>+</sup> and Cu<sup>2+</sup> binding to Phe, Tyr, Trp, and His amino acids in the gas phase. Insights from first-principles calculations. *J. Phys. Chem. B* **2006**, *110*, 24189–24199. [[CrossRef](#)]
37. Su, P.; Li, H. Energy decomposition analysis of covalent bonds and intermolecular interactions. *J. Chem. Phys.* **2009**, *131*, 014102. [[CrossRef](#)]
38. Zhang, Y.; Chen, S.; Ying, F.; Su, P.; Wu, W. Valence Bond Based Energy Decomposition Analysis Scheme and Its Application to Cation- $\pi$  Interactions. *J. Phys. Chem. A* **2018**, *122*, 5886–5894. [[CrossRef](#)]
39. Te Velde, G.; Bickelhaupt, F.M.; Baerends, E.J.; Fonseca Guerra, C.; van Gisbergen, S.J.A.; Snijders, J.G.; Ziegler, T. Chemistry with ADF. *J. Comput. Chem* **2001**, *22*, 931–967. [[CrossRef](#)]
40. Deschamps, P.; Kulkarni, P.P.; Gautam-Basak, M.; Sarkar, B. The saga of copper(II)-L-histidine. *Coord. Chem. Rev.* **2005**, *249*, 895–909. [[CrossRef](#)]
41. Perrin, D.D.; Sharma, V.S. Histidine complexes with some bivalent cations. *J. Chem. Soc. A* **1967**, 724–728. [[CrossRef](#)]
42. Sigel, H.; Griesser, R.; McCormick, D.B. On the structure of manganese (II)- and copper (II)-histidine complexes. *Arch. Biochem. Biophys.* **1969**, *134*, 217–227. [[CrossRef](#)]
43. Mesu, J.G.; Visser, T.; Soulimani, F.; van Faassen, E.E.; de Peinder, P.; Beale, A.M.; Weckhuysen, B.M. New insights into the coordination chemistry and molecular structure of copper(II) histidine complexes in aqueous solutions. *Inorg. Chem.* **2006**, *45*, 1960–1971. [[CrossRef](#)]
44. Isaac, M.; Denisov, S.A.; Roux, A.; Imbert, D.; Jonusauskas, G.; McClenaghan, N.D.; Seneque, O. Lanthanide luminescence modulation by cation- $\pi$  interaction in a bioinspired scaffold: Selective detection of copper(I). *Angew. Chem. Int. Ed.* **2015**, *54*, 11453–11456. [[CrossRef](#)]
45. Takeuchi, H. UV Raman markers for structural analysis of aromatic side chains in proteins. *Anal. Sci.* **2011**, *27*, 1077. [[CrossRef](#)]
46. Li, S.; Hong, M. Protonation, tautomerization, and rotameric structure of histidine: A comprehensive study by magic-angle-spinning solid-state NMR. *J. Am. Chem. Soc.* **2011**, *133*, 1534–1544. [[CrossRef](#)]
47. Polfer, N.C.; Oomens, J.; Moore, D.T.; von Helden, G.; Meijer, G.; Dunbar, R.C. Infrared spectroscopy of phenylalanine Ag(I) and Zn(II) complexes in the gas phase. *J. Am. Chem. Soc.* **2006**, *128*, 517–525. [[CrossRef](#)]
48. Kruck, T.P.A.; Sarkar, B. Structure of the species in the copper (II)-L-histidine system. *Can. J. Chem.* **1973**, *51*, 3563–3571. [[CrossRef](#)]
49. Becke, A.D. Density-functional thermochemistry. III. The role of exact exchange. *J. Chem. Phys.* **1993**, *98*, 5648–5652. [[CrossRef](#)]
50. Frisch, M.; Trucks, G.; Schlegel, H.B.; Scuseria, G.; Robb, M.; Cheeseman, J.; Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G. *Gaussian 09, Revision, A. 02*; Gaussian, Inc.: Wallingford, CT, USA, 2009; p. 200.
51. Peng, C.Y.; Ayala, P.Y.; Schlegel, H.B.; Frisch, M.J. Using redundant internal coordinates to optimize equilibrium geometries and transition states. *J. Comput. Chem.* **1996**, *17*, 49–56. [[CrossRef](#)]
52. Rumble, J.R. *CRC Handbook of Chemistry and Physics*, 98th ed.; CRC Press: Boca Raton, FL, USA, 2017.
53. Tu, Y.; Liu, H.; Shi, G.; Zhang, F.; Su, T.; Wu, Y.; Sun, J.; Zhang, L.; Zhang, S.; Fang, H. Selectivity mechanism of magnesium and calcium in cation-binding pocket structures of phosphotyrosine. *Phys. Rev. E* **2020**, *101*, 022410. [[CrossRef](#)]
54. Hirshfeld, F.L. Bonded-atom fragments for describing molecular charge densities. *Theor. Chim. Acta* **1977**, *44*, 129–138. [[CrossRef](#)]