COMMENTARY

Advances in Therapeutic Approaches to Extend Healthspan: a perspective from the 2nd Scripps Symposium on the Biology of Aging

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Summary

The 2nd Scripps Florida Symposium on The Biology of Aging entitled 'Advances in Therapeutic Approaches to Extend Healthspan' was held on January 22nd–25th, 2017 at The Scripps Research Institute in Jupiter, Florida. The meeting highlighted a variety of therapeutic approaches in animal models of aging that either are or soon will be in clinic trials. For example, drugs targeting senescent cells, metformin, rapalogs, NAD precursors, young plasma, mitochondrial-targeted free radical scavengers, stem cells, and stem cell factors all have shown significant preclinical efficacy. This perspective, based on presentations and discussions at the symposium, outlines the current and future state of development of therapeutic approaches to extend human healthspan.

The 2nd Scripps Florida Symposium on The Biology of Aging entitled 'Advances in Therapeutic Approaches to Extend Healthspan' was held on January 22nd–25th, 2017 at The Scripps Research Institute in Jupiter, Florida. The goal of the symposium was to bring together leaders in the fields of aging and drug development to discuss strategies for identifying and developing therapeutic approaches to extend human healthspan. This symposium made it highly evident that the biology of aging field is moving quickly toward translational research. At the symposium, there were numerous reports of successful drug screens and drug testing in a variety of model systems. There was also an overall sense of excitement, given that multiple therapeutic modalities, including young plasma, recombinant proteins, and small molecules, extend healthspan and lifespan in model organisms and that clinical trials to test the efficacy of these treatment modalities on healthspan and resilience have been initiated.

The concept of Geroscience, defined as the understanding of the relationship between aging and age-related diseases and preventing/ delaying disease by targeting fundamental mechanisms of aging (Kennedy *et al.*, 2014), was an underlying theme of the symposium.

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As reiterated by the keynote speakers, Jay Olshansky, Felipe Sierra and Steve Austad, aging is the main risk factor for most chronic diseases. Thus, developing approaches to therapeutically target aging should be a funding priority for the majority of institutes at the National Institutes of Health, as well as other funding agencies, philanthropists, and foundations (Kaeberlein *et al.*, 2015). The socioeconomic need to extend human healthspan also was made clear. As a consequence of the advances in prevention and treatment of infectious diseases, there will be an unprecedented increase in the number of persons over 65 over the next decades. By 2035, the cost of treating Americans 65 years and older is expected to be over \$2 trillion annually. Thus, finding ways to prevent all age-related diseases is one of the most imperative biomedical pursuits.

A common theme arising from the symposium was the need for appropriate model organisms to study aging and age-related disease including models carrying reporters of senescence, mitochondrial function, autophagy and reactive oxygen species (ROS). The use of these reporters or testing of therapeutics needs to be performed in aged model organisms, a problem that has also plagued the cancer field because of the cost and time involved, at least in rodent models. Here, the National Institute on Aging (NIA) Interventions Testing Program in mice (ITP) and in Caenorhabditis elegans (C. elegans) (CITP) have made significant contributions to the identification of drugs/compounds able to extend lifespan (Nadon et al., 2008; Strong et al., 2008; Harrison et al., 2009, 2014; Miller et al., 2011). For example, aspirin, rapamycin, acarbose, 17a-estradiol, Protandim and nordihydroguaiaretic acid (NDGA) extend the lifespan of at least one sex of genetically heterogeneous mice. With this number of current 'hits' and likely future 'hits', it is possible that shared mechanism(s) of action will emerge, illuminating novel biology regarding the pathways that drives aging. Unfortunately, the ITP is not incorporating reporters into their heterogeneous mouse strains. The development of novel model organisms of aging, including the African Killifish (Nothobranchius furzeri) (Harel et al., 2015; Valenzano et al., 2015) and different strains of nematodes that can be easily modified to carrying specific reporters should help not only identify pathways of aging, but also in the screening for therapeutics.

Despite the ITP, CITP, and new models of aging, there still is a need for an expansion of efforts to measure the effects of drugs/compounds on healthspan or resilience, which has greater translational relevance (Niedernhofer *et al.*, 2017). Thus, many investigators are beginning to incorporate functional analysis of aged mice undergoing therapeutic interventions, for example, echocardiography (Chiao & Rabinovitch, 2015). Also, mouse models of accelerated aging afford rapid testing of interventions and measurement of the impact on age-related symptoms and functional decline. These models include those of Hutchinson– Guilford Progeria Syndrome (HGPS) and XFE progeroid syndrome, which mimic many aspects of natural aging, including senescence, stem cell dysfunction, mitochondrial dysfunction, metabolic changes, and predisposition to common age-related diseases (Harkema *et al.*, 2016). Murine strains with tissue-specific accelerated aging also have been developed

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to aid in the discovery of which organ(s) are sentinel for driving organismal aging. Preliminary data point toward aging of the immune system as being critical for systemic aging (Laura Niedernhofer). Consistent with this is the key role of hemocytes in tissue repair of invertebrates (Neves *et al.*, 2016) and the observation that transplantation of healthy, young immune cells into mice after total body irradiation reduced the burden of senescent cells (Andrei Gudkov).

mTOR continues to be a favored molecular target for aging, given the ability of rapamycin and rapalogs to extend lifespan in multiple model systems (Harrison *et al.*, 2009; Liao *et al.*, 2016). Remarkably, there is still no consensus on its mechanism of action. Upstream of mTOR, an age-related decline in proteolysis by the lysosome may limit circulating amino acids, driving mTOR activation (Wolfson *et al.*, 2017). Downstream of mTOR, 4EBP1 is a critical effector that regulates protein translation. 4EBP1 appears to be regulated by inflammation (a key feature of aging) in a sex-specific manner in mice (Brian Kennedy). However, much work remains to define the key age-related activator of mTOR and downstream effector(s).

Biologics represent another exciting therapeutic approach to improve healthspan. Heterochronic parabiosis or repeated injection of young plasma into old mice results in improvement in the function of some tissues, including the central nervous system (Conboy et al., 2005, 2013; Conboy & Rando, 2012; Loffredo et al., 2013; Villeda et al., 2014; Smith et al., 2015). This suggests that serum factors can be identified that slow or even reverse certain aspects of aging. This is true not only for naturally aged mice, but also for murine models of progeria such as Zmpste24^{-/-} and $Ercc 1^{-/\Delta}$ mice where parabiosis with young wild-type mice improved some aging phenotypes (Johnny Huard). There have been several factors identified in the serum of young animals that have been proposed to act as antigeronic factors, including oxytocin, TIMP2, mesencephalic astrocyte-derived neurotrophic factor (MANF) and growth differentiation factor 11 (GDF-11) (Loffredo et al., 2013; Sinha et al., 2014; Neves et al., 2016). GDF-11 has offered a potent reminder that accurate detection of serum proteins is tremendously difficult due to interference by isoforms, receptors, multiple activity states (pre-, pro- forms) and serum binding partners (Walker et al., 2016). Levels of other proteins, such as GM-CSF and TIMP-2, also appear change significantly with aging (Bonnema et al., 2007; Kim et al., 2011). Remarkably, injection of mice with human fetal cord blood significantly improved the cognitive function of mice. This was mimicked by injecting recombinant TIMP-2 whereas, in contrast, depletion of TIMP-2 from fetal cord blood abrogated its positive effects on cognition (Tony Wyss-Coray). These observations demonstrate that tissue rejuvenation and improved cognitive function could be attainable by treatment of the elderly with young plasma and ultimately recombinant proteins. Clearly, more effort needs to be placed on identifying and understanding how young serum slows aging. Results from an ongoing clinical trial to assess the effect of young plasma on Alzheimer's disease will provide significant impetus to identify antigeronic serum factors (Middeldorp et al., 2016).

An emerging paradigm in the field of aging is that the burden of senescent cells increases with age in multiple tissues and reducing this senescent cell burden improves healthspan. The development and characterization of two transgenic mouse models, p16-INK-ATTAC and p16-3MR mice, clearly demonstrated that senescent cells play a role in driving aging and age-related diseases (Baker *et al.*, 2011, 2016; Childs *et al.*, 2016). The reduction of senescent cells in the p16-3MR mice reduced atherosclerosis, improved metabolism, prevented tumor metastasis and reduced osteoarthritis in an injury model (Demaria *et al.*, 2017). Thus, the hunt is on for senolytics or drugs that specifically kill senescent cells (Zhu *et al.*, 2015, 2016, 2017; Chang *et al.*, 2016; Wang *et al.*,

2016). Screening of compounds in senescent human and mouse cultures revealed a number of classes of drugs/compounds that can induce apoptosis of senescent cells. These senolytics include the tyrosine kinase inhibitor Dasatinib (D), the flavonoid Quercetin (Q), the Bcl-2 family inhibitor Navitoclax, piperlongumine and Hsp90 inhibitors (Zhu *et al.*, 2015; Chang *et al.*, 2016; Wang *et al.*, 2016; Zhu *et al.*, 2016; Schafer *et al.*, 2017; Zhu *et al.*, 2017; Paul Robbins). Treatment of a mouse model of accelerated aging with D+Q or the Hsp90 inhibitor 17-DMAG extended their healthspan (Zhu *et al.*, 2015). Similarly, treatment of naturally aged mice with D+Q improved heart function and improved metabolism, especially in mice on a high-fat diet (Roos *et al.*, 2016). Whether these or more optimized senolytics will have similar positive effects on human healthspan is still unclear, but clinical trials are being planned to determine their effectiveness.

It is important to note that it is likely that no one senolytic will be effective in eliminating all types of senescent cells. Individual senolytic compounds are apt to have tissue-specific and even cell type-specific effects. Furthermore, there is increasing evidence that different drivers of senescence can lead to differences in how senescence manifests, which in turn could have a variable impact on the senescent cell's environment. Another complexity is that some drugs/compounds affect expression of senescent-associated secretory phenotype (SASP) factors without eliminating the senescent cells (Tilstra et al., 2012; Laberge et al., 2015; Xu et al., 2015). For example, rapamycin and NF-κB and JAK inhibitors reduce SASP and other markers of senescence without actually inducing cell death. This class of compounds, termed senomorphics, also may affect healthspan through reducing the cell nonautonomous effects of senescence on aging. Clearly documenting whether a compound functions as a senolytic or senomorphic in vivo will require the development of new reporters and methodologies.

Adult stem cell function is known to decline with aging. However, it has taken longer to demonstrate that the loss of stem cell function contributes to aging and is not simply a consequence of it. Treatment of a mouse model of accelerated aging with two types of young stem cells, muscle derived stem/progenitor cells (MDSPCs) and bone marrowderived mesenchymal stem cells (MSCs) extends healthspan and lifespan (Lavasani et al., 2012). This was also recapitulated in aged rats using adipose-derived mesenchymal stem cells (Kim et al., 2015). Although the exact mechanism for how these stem cell populations affect aging is unknown, preliminary data suggest that their effect is mediated by factors secreted by young, but not old stem cells. These factors appear to reduce cellular senescence and improve the function of endogenous, aged stem cells. Whether these stem cell-derived soluble factors are the same as those found in young plasma is currently unknown. In addition to loss of stem cell function with age, there is also evidence that progenitor cell populations in muscle contribute to driving aging (Johnny Huard). Importantly, this age-related stem cell dysfunction appears reversible. New data was presented pointing toward the existence of cardiac stem cells necessary for tissue regeneration (Nadal-Ginard et al., 2014; Smith et al., 2014). Currently, senotherapeutics are being tested on these cardiac stem cells to determine whether senescence is an underlying factor in cardiac aging. This will undoubtedly remain an area of intense research spanning from continued investigations into fundamental mechanisms of aging to clinical trials.

The occurrence of genetic variants in humans that affect longevity and health in old age offers an ideal starting point for the systematic identification of targets for interventions that affect healthspan. Examples of these 'natural mutants' are human centenarians and supercentenarians where rare variants in the coding sequence of IGF-1R, FOXO3a, and SIRT6 have been identified (Suh *et al.*, 2008). An alternative approach to identifying gene variants important for healthy aging is to link a human phenotype, such as presence or absence of agerelated disease, to a genotype. The extensive exome sequencing being performed by companies such as 23andMe and Regeneron will facilitate the discovery of genes linked to extended healthspan defined as the avoidance of age-related disease (Dewey *et al.*, 2016). However, many rare variants lie in noncoding regions or transcriptional regulatory regions located at significant distances from their effector gene, making their identification and characterization of their mechanisms of action quite difficult.

Clear evidence that the field of aging is moving forward quickly is the number of ongoing or soon-to-be-initiated clinical trials. Importantly, the use of specific short-term clinical endpoints to determine if resilience or function of a specific tissue could be improved is employed to reduce study size, duration, and cost. For example, short-term treatment of a cohort of elderly people with a rapamycin analogue (rapalog) was tested for its ability to improve immune function (Mannick et al., 2014). The inclusion of additional endpoints provides further information about not only the effect of the intervention on immune function, but also on other aspects of aging that might be modulated and measured in future clinical trials. Similarly, short-term clinical trials with the mitochondrialtargeted SS-31 peptide are in progress for heart disease based on very promising preclinical data (Dai et al., 2011; Siegel et al., 2013). In contrast, a well-controlled clinical study to examine the effect of high dose resveratrol on diabetes showed no positive effect, despite promising preclinical data (Jill Crandell). Unfortunately, a negative result might only mean that the wrong dose or dosing regimen was used or the wrong patient population enrolled.

These short-term treatment trials with well-defined, disease-specific endpoints are in contrast to the highly anticipated Treating Aging with Metformin (TAME) trial. The TAME trial is designed to enable evaluating whether metformin extends the healthspan of humans albeit in a rapid 3-5 year format (Barzilai *et al.*, 2016; Newman *et al.*, 2016). It is hoped that the TAME trial will serve as a template for pharmaceutical companies to do future testing of drugs aimed at targeting fundamental mechanisms of aging.

In addition to the exceptional science presented, several important issues were raised during the panel discussion. One of these issues is that it is impossible to stop the world's population from getting older, but it is possible to make this older population healthier. A second issue is emphasizing that the push is for therapies that ablate age-related disease, not extend lifespan. A small lifespan extension may be a 'side effect' of this therapeutic approach. A third issue is that we already have therapeutics that extend health like statins and insulin, but patient compliance is not perfect. How compliant will people be with taking additional preventative therapies? Finally, dialogue about how society should adapt, both socially and economically, to a healthier older population is needed.

The Glenn Foundation and the American Federation for Aging Research (AFAR) are doing an admirable job in funding top quality aging research, but with limited budgets. However, it also is clear that support from the private sector will be essential for moving clinical trials forward as there is a huge need for funding from sources other than the NIH to expedite aging research. The successful completion of the first clinical trial demonstrating that human healthspan can be extended is anticipated to instigate tremendous interest in the field by biotech investors and potentially philanthropists. Thus, this first proof-of-principle clinical trial and funding support for it is considered a significant hurdle that must be crossed to accelerate funding and progress in the field.

Another hurdle is the lack of biomarkers for assessing the efficacy of a compound in slowing aging, which can be translated from animal models to human trials. A defined biomarker of biological aging would facilitate not only predicting who should be enrolled in clinical trials for aging therapeutics, but also in determining the efficacy of a treatment regimen. A third hurdle is the need for infrastructure for running largescale, aging clinical trials, analogous to NCI Comprehensive Cancer Centers. Here, leveraging and expanding the existing infrastructure within the NIH-funded Clinical and Translational Science Award (CTSA) program, Claude D. Pepper Older Americans Independence Centers (OAIC), Nathan Shock Centers of Excellence and Glenn Centers for Research in Aging could help facilitate successful clinical trials. There also is a lack of geriatricians with appropriate training in clinical trial design. This combination of lack of biomarkers, geriatricians experienced in clinical trials, and a successful trial template creates a unique challenge that will be difficult, but critical to overcome. Despite these barriers to the field, all of which can be overcome with research funding and appropriate planning, there likely will be a number of significant preclinical and clinical advances by the 3rd Scripps Symposium on the Biology of Aging in January of 2019.

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Conflict of interest

None declared.

References

- Baker DJ, Wijshake T, Tchkonia T, LeBrasseur NK, Childs BG, van de Sluis B, Kirkland JL, van Deursen JM (2011) Clearance of p16lnk4a-positive senescent cells delays ageing-associated disorders. *Nature* **479**, 232–236.
- Baker DJ, Childs BG, Durik M, Wijers ME, Sieben CJ, Zhong J, Saltness RA, Jeganathan KB, Verzosa GC, Pezeshki A, Khazaie K, Miller JD, van Deursen JM (2016) Naturally occurring p16(Ink4a)-positive cells shorten healthy lifespan. *Nature* 530, 184–189.
- Barzilai N, Crandall JP, Kritchevsky SB, Espeland MA (2016) Metformin as a tool to target aging. *Cell Metab.* **23**, 1060–1065.
- Bonnema DD, Webb CS, Pennington WR, Stroud RE, Leonardi AE, Clark LL, McClure CD, Finklea L, Spinale FG, Zile MR (2007) Effects of age on plasma matrix metalloproteinases (MMPs) and tissue inhibitor of metalloproteinases (TIMPs). J. Card. Fail. **13**, 530–540.
- Chang J, Wang Y, Shao L, Laberge RM, Demaria M, Campisi J, Janakiraman K, Sharpless NE, Ding S, Feng W, Luo Y, Wang X, Aykin-Burns N, Krager K, Ponnappan U, Hauer-Jensen M, Meng A, Zhou D (2016) Clearance of senescent cells by ABT263 rejuvenates aged hematopoietic stem cells in mice. *Nat. Med.* 22, 78–83.
- Chiao YA, Rabinovitch PS (2015) The aging heart. Cold Spring Harb. Perspect Med. 5, a025148.

- Childs BG, Baker DJ, Wijshake T, Conover CA, Campisi J, van Deursen JM (2016) Senescent intimal foam cells are deleterious at all stages of atherosclerosis. *Science* **354**, 472–477.
- Conboy IM, Rando TA (2012) Heterochronic parabiosis for the study of the effects of aging on stem cells and their niches. *Cell Cycle* **11**, 2260–2267.
- Conboy IM, Conboy MJ, Wagers AJ, Girma ER, Weissman IL, Rando TA (2005) Rejuvenation of aged progenitor cells by exposure to a young systemic environment. *Nature* **433**, 760–764.
- Conboy MJ, Conboy IM, Rando TA (2013) Heterochronic parabiosis: historical perspective and methodological considerations for studies of aging and longevity. *Aging Cell* **12**, 525–530.
- Dai DF, Chen T, Szeto H, Nieves-Cintron M, Kutyavin V, Santana LF, Rabinovitch PS (2011) Mitochondrial targeted antioxidant peptide ameliorates hypertensive cardiomyopathy. J. Am. Coll. Cardiol. 58, 73–82.
- Demaria M, O'Leary MN, Chang J, Shao L, Liu S, Alimirah F, Koenig K, Le C, Mitin N, Deal AM, Alston S, Academia EC, Kilmarx S, Valdovinos A, Wang B, de Bruin A, Kennedy BK, Melov S, Zhou D, Sharpless NE, Muss H, Campisi J (2017) Cellular senescence promotes adverse effects of chemotherapy and cancer relapse. *Cancer Discov.* 7, 165–176.
- Dewey FE, Murray MF, Overton JD, Habegger L, Leader JB, Fetterolf SN, O'Dushlaine C, Van Hout CV, Staples J, Gonzaga-Jauregui C, Metpally R, Pendergrass SA, Giovanni MA, Kirchner HL, Balasubramanian S, Abul-Husn NS, Hartzel DN, Lavage DR, Kost KA, Packer JS, Lopez AE, Penn J, Mukherjee S, Gosalia N, Kanagaraj M, Li AH, Mitnaul LJ, Adams LJ, Person TN, Praveen K, Marcketta A, Lebo MS, Austin-Tse CA, Mason-Suares HM, Bruse S, Mellis S, Phillips R, Stahl N, Murphy A, Economides A, Skelding KA, Still CD, Elmore JR, Borecki IB, Yancopoulos GD, Davis FD, Faucett WA, Gottesman O, Ritchie MD, Shuldiner AR, Reid JG, Ledbetter DH, Baras A, Carey DJ (2016) Distribution and clinical impact of functional variants in 50,726 whole-exome sequences from the DiscovEHR study. *Science* **354**, aaf6814.
- Harel I, Benayoun BA, Machado B, Singh PP, Hu CK, Pech MF, Valenzano DR, Zhang E, Sharp SC, Artandi SE, Brunet A (2015) A platform for rapid exploration of aging and diseases in a naturally short-lived vertebrate. *Cell* **160**, 1013–1026.
- Harkema L, Youssef SA, de Bruin A (2016) Pathology of mouse models of accelerated aging. *Vet. Pathol.* **53**, 366–389.
- Harrison DE, Strong R, Sharp ZD, Nelson JF, Astle CM, Flurkey K, Nadon NL, Wilkinson JE, Frenkel K, Carter CS, Pahor M, Javors MA, Fernandez E, Miller RA (2009) Rapamycin fed late in life extends lifespan in genetically heterogeneous mice. *Nature* **460**, 392–395.
- Harrison DE, Strong R, Allison DB, Ames BN, Astle CM, Atamna H, Fernandez E, Flurkey K, Javors MA, Nadon NL, Nelson JF, Pletcher S, Simpkins JW, Smith D, Wilkinson JE, Miller RA (2014) Acarbose, 17-alpha-estradiol, and nordihydroguaiaretic acid extend mouse lifespan preferentially in males. *Aging Cell* 13, 273–282.
- Kaeberlein M, Rabinovitch PS, Martin GM (2015) Healthy aging: the ultimate preventative medicine. *Science* 350, 1191–1193.
- Kennedy BK, Berger SL, Brunet A, Campisi J, Cuervo AM, Epel ES, Franceschi C, Lithgow GJ, Morimoto RI, Pessin JE, Rando TA, Richardson A, Schadt EE, Wyss-Coray T, Sierra F (2014) Geroscience: linking aging to chronic disease. *Cell* **159**, 709–713.
- Kim HO, Kim HS, Youn JC, Shin EC, Park S (2011) Serum cytokine profiles in healthy young and elderly population assessed using multiplexed bead-based immunoassays. J. Transl. Med. 9, 113.
- Kim D, Kyung J, Park D, Choi EK, Kim KS, Shin K, Lee H, Shin IS, Kang SK, Ra JC, Kim YB (2015) Health span-extending activity of human amniotic membraneand adipose tissue-derived stem cells in F344 rats. *Stem Cells Transl. Med.* 4, 1144–1154.
- Laberge RM, Sun Y, Orjalo AV, Patil CK, Freund A, Zhou L, Curran SC, Davalos AR, Wilson-Edell KA, Liu S, Limbad C, Demaria M, Li P, Hubbard GB, Ikeno Y, Javors M, Desprez PY, Benz CC, Kapahi P, Nelson PS, Campisi J (2015) MTOR regulates the pro-tumorigenic senescence-associated secretory phenotype by promoting IL1A translation. *Nat. Cell Biol.* **17**, 1049–1061.
- Lavasani M, Robinson AR, Lu A, Song M, Feduska JM, Ahani B, Tilstra JS, Feldman CH, Robbins PD, Niedernhofer LJ, Huard J (2012) Muscle-derived stem/ progenitor cell dysfunction limits healthspan and lifespan in a murine progeria model. *Nat. Commun.* **3**, 608.
- Liao CY, Anderson SS, Chicoine NH, Mayfield JR, Academia EC, Wilson JA, Pongkietisak C, Thompson MA, Lagmay EP, Miller DM, Hsu YM, McCormick MA, O'Leary MN, Kennedy BK (2016) Rapamycin reverses metabolic deficits in Lamin A/C-Deficient mice. *Cell Rep.* **17**, 2542–2552.
- Loffredo FS, Steinhauser ML, Jay SM, Gannon J, Pancoast JR, Yalamanchi P, Sinha M, Dall'Osso C, Khong D, Shadrach JL, Miller CM, Singer BS, Stewart A, Psychogios N, Gerszten RE, Hartigan AJ, Kim MJ, Serwold T, Wagers AJ, Lee RT

(2013) Growth differentiation factor 11 is a circulating factor that reverses agerelated cardiac hypertrophy. *Cell* **153**, 828–839.

- Mannick JB, Del Giudice G, Lattanzi M, Valiante NM, Praestgaard J, Huang B, Lonetto MA, Maecker HT, Kovarik J, Carson S, Glass DJ, Klickstein LB (2014) mTOR inhibition improves immune function in the elderly. *Sci. Transl. Med.* **6**, 268ra179.
- Middeldorp J, Lehallier B, Villeda SA, Miedema SS, Evans E, Czirr E, Zhang H, Luo J, Stan T, Mosher KI, Masliah E, Wyss-Coray T (2016) Preclinical assessment of young blood plasma for alzheimer disease. *JAMA Neurol.* **73**, 1325–1333.
- Miller RA, Harrison DE, Astle CM, Baur JA, Boyd AR, de Cabo R, Fernandez E, Flurkey K, Javors MA, Nelson JF, Orihuela CJ, Pletcher S, Sharp ZD, Sinclair D, Starnes JW, Wilkinson JE, Nadon NL, Strong R (2011) Rapamycin, but not resveratrol or simvastatin, extends life span of genetically heterogeneous mice. J. Gerontol. A Biol. Sci. Med. Sci. 66, 191–201.
- Nadal-Ginard B, Ellison GM, Torella D (2014) The cardiac stem cell compartment is indispensable for myocardial cell homeostasis, repair and regeneration in the adult. Stem Cell Res. 13, 615–630.
- Nadon NL, Strong R, Miller RA, Nelson J, Javors M, Sharp ZD, Peralba JM, Harrison DE (2008) Design of aging intervention studies: the NIA interventions testing program. Age 30, 187–199.
- Neves J, Zhu J, Sousa-Victor P, Konjikusic M, Riley R, Chew S, Qi Y, Jasper H, Lamba DA (2016) Immune modulation by MANF promotes tissue repair and regenerative success in the retina. *Science* **353**, aaf3646.
- Newman JC, Milman S, Hashmi SK, Austad SN, Kirkland JL, Halter JB, Barzilai N (2016) Strategies and challenges in clinical trials targeting human aging. *J. Gerontol. A Biol. Sci. Med. Sci.* **71**, 1424–1434.
- Niedernhofer LJ, Kirkland JL, Ladiges W (2017) Molecular pathology endpoints useful for aging studies. *Ageing Res. Rev.* **35**, 241–249.
- Roos CM, Zhang B, Palmer AK, Ogrodnik MB, Pirtskhalava T, Thalji NM, Hagler M, Jurk D, Smith LA, Casaclang-Verzosa G, Zhu Y, Schafer MJ, Tchkonia T, Kirkland JL, Miller JD (2016) Chronic senolytic treatment alleviates established vasomotor dysfunction in aged or atherosclerotic mice. *Aging Cell* **15**, 973–977.
- Schafer MJ, White TA, Iijima K, Haak AJ, Ligresti G, Atkinson EJ, Oberg AL, Birch J, Salmonowicz H, Zhu Y, Mazula DL, Brooks RW, Fuhrmann-Stroissnigg H, Pirtskhalava T, Prakash YS, Tchkonia T, Robbins PD, Aubry MC, Passos JF, Kirkland JL, Tschumperlin DJ, Kita H, LeBrasseur NK (2017) Cellular senescence mediates fibrotic pulmonary disease. *Nat. Commun.* 8, 14532.
- Siegel MP, Kruse SE, Percival JM, Goh J, White CC, Hopkins HC, Kavanagh TJ, Szeto HH, Rabinovitch PS, Marcinek DJ (2013) Mitochondrial-targeted peptide rapidly improves mitochondrial energetics and skeletal muscle performance in aged mice. *Aging Cell* **12**, 763–771.
- Sinha M, Jang YC, Oh J, Khong D, Wu EY, Manohar R, Miller C, Regalado SG, Loffredo FS, Pancoast JR, Hirshman MF, Lebowitz J, Shadrach JL, Cerletti M, Kim MJ, Serwold T, Goodyear LJ, Rosner B, Lee RT, Wagers AJ (2014) Restoring systemic GDF11 levels reverses age-related dysfunction in mouse skeletal muscle. *Science* **344**, 649–652.
- Smith AJ, Lewis FC, Aquila I, Waring CD, Nocera A, Agosti V, Nadal-Ginard B, Torella D, Ellison GM (2014) Isolation and characterization of resident endogenous c-Kit+ cardiac stem cells from the adult mouse and rat heart. *Nat. Protoc.* 9, 1662–1681.
- Smith LK, He Y, Park JS, Bieri G, Snethlage CE, Lin K, Gontier G, Wabl R, Plambeck KE, Udeochu J, Wheatley EG, Bouchard J, Eggel A, Narasimha R, Grant JL, Luo J, Wyss-Coray T, Villeda SA (2015) beta2-microglobulin is a systemic pro-aging factor that impairs cognitive function and neurogenesis. *Nat. Med.* **21**, 932–937.
- Strong R, Miller RA, Astle CM, Floyd RA, Flurkey K, Hensley KL, Javors MA, Leeuwenburgh C, Nelson JF, Ongini E, Nadon NL, Warner HR, Harrison DE (2008) Nordihydroguaiaretic acid and aspirin increase lifespan of genetically heterogeneous male mice. *Aging Cell* **7**, 641–650.
- Suh Y, Atzmon G, Cho MO, Hwang D, Liu B, Leahy DJ, Barzilai N, Cohen P (2008) Functionally significant insulin-like growth factor I receptor mutations in centenarians. *Proc. Natl. Acad. Sci. U.S.A.* **105**, 3438–3442.
- Tilstra JS, Robinson AR, Wang J, Gregg SQ, Clauson CL, Reay DP, Nasto LA, St Croix CM, Usas A, Vo N, Huard J, Clemens PR, Stolz DB, Guttridge DC, Watkins SC, Garinis GA, Wang Y, Niedernhofer LJ, Robbins PD (2012) NF-kappaB inhibition delays DNA damage-induced senescence and aging in mice. J. Clin. Invest. **122**, 2601–2612.
- Valenzano DR, Benayoun BA, Singh PP, Zhang E, Etter PD, Hu CK, Clement-Ziza M, Willemsen D, Cui R, Harel I, Machado BE, Yee MC, Sharp SC, Bustamante CD, Beyer A, Johnson EA, Brunet A (2015) The African turquoise killifish genome provides insights into evolution and genetic architecture of lifespan. *Cell* 163, 1539–1554.
- Villeda SA, Plambeck KE, Middeldorp J, Castellano JM, Mosher KI, Luo J, Smith LK, Bieri G, Lin K, Berdnik D, Wabl R, Udeochu J, Wheatley EG, Zou B, Simmons DA,

Xie XS, Longo FM, Wyss-Coray T (2014) Young blood reverses age-related impairments in cognitive function and synaptic plasticity in mice. *Nat. Med.* **20**, 659–663.

- Walker RG, Poggioli T, Katsimpardi L, Buchanan SM, Oh J, Wattrus S, Heidecker B, Fong YW, Rubin LL, Ganz P, Thompson TB, Wagers AJ, Lee RT (2016) Biochemistry and biology of GDF11 and myostatin: similarities, differences, and questions for future investigation. *Circ. Res.* **118**, 1125–1141. discussion 1142.
- Wang Y, Chang J, Liu X, Zhang X, Zhang S, Zhang X, Zhou D, Zheng G (2016) Discovery of piperlongumine as a potential novel lead for the development of senolytic agents. *Aging* 8, 2915–2926.
- Wolfson RL, Chantranupong L, Wyant GA, Gu X, Orozco JM, Shen K, Condon KJ, Petri S, Kedir J, Scaria SM, Abu-Remaileh M, Frankel WN, Sabatini DM (2017) KICSTOR recruits GATOR1 to the lysosome and is necessary for nutrients to regulate mTORC1. *Nature* **543**, 438–442.
- Xu M, Tchkonia T, Ding H, Ogrodnik M, Lubbers ER, Pirtskhalava T, White TA, Johnson KO, Stout MB, Mezera V, Giorgadze N, Jensen MD, LeBrasseur NK, Kirkland JL (2015) JAK inhibition alleviates the cellular senescence-associated

secretory phenotype and frailty in old age. Proc. Natl. Acad. Sci. U.S.A. 112, E6301–E6310.

- Zhu Y, Tchkonia T, Pirtskhalava T, Gower AC, Ding H, Giorgadze N, Palmer AK, Ikeno Y, Hubbard GB, Lenburg M, O'Hara SP, LaRusso NF, Miller JD, Roos CM, Verzosa GC, LeBrasseur NK, Wren JD, Farr JN, Khosla S, Stout MB, McGowan SJ, Fuhrmann-Stroissnigg H, Gurkar AU, Zhao J, Colangelo D, Dorronsoro A, Ling YY, Barghouthy AS, Navarro DC, Sano T, Robbins PD, Niedernhofer LJ, Kirkland JL (2015) The Achilles' heel of senescent cells: from transcriptome to senolytic drugs. Aging Cell 14, 644–658.
- Zhu Y, Tchkonia T, Fuhrmann-Stroissnigg H, Dai HM, Ling YY, Stout MB, Pirtskhalava T, Giorgadze N, Johnson KO, Giles CB, Wren JD, Niedernhofer LJ, Robbins PD, Kirkland JL (2016) Identification of a novel senolytic agent, navitoclax, targeting the Bcl-2 family of anti-apoptotic factors. *Aging Cell* **15**, 428–435.
- Zhu Y, Doornebal EJ, Pirtskhalava T, Giorgadze N, Wentworth M, Fuhrmann-Stroissnigg H, Niedernhofer LJ, Robbins PD, Tchkonia T, Kirkland JL (2017) New agents that target senescent cells: the flavone, fisetin, and the BCL-XL inhibitors, A1331852 and A1155463. *Aging* **9**, 955–963.