Extensive metabolic consequences of human glycosyltransferase gene knockouts in prostate cancer

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Supplementary Information:

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quantitative data

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in UGT2B17 KO and UGT2B28 KO

Table S1 - Overview of metabolomics data

	UGT2B17 KO					UGT2B28 KO				
MS-based analysis	metabolites changed <i>vs</i> gene proficient cases			n (% of changed) ⁶		metabolites changed <i>vs</i> gene proficient cases			n (% of changed) ⁶	
n=measured metabolites1		n	% ⁵	up	down		n	%	up	down
Global (untargeted) ²	P≤0.05	48	7.4	26 (54%)	22 (46%)	P≤0.05	56	8.6	11 (20%)	45 (80%)
n=651	P<0.1	39	6	14 (36%)	25 (64%)	P<0.1	47	7.2	9 (19%)	38 (81%)
Lipidomics ²	P≤0.05	37	4.6	32 (86%)	5 (14%)	P≤0.05	19	2.4	1 (5%)	18 (95%)
n=804	P<0.1	49	6	36 (73%)	13 (17%)	P<0.1	17	2.1	4 (24%)	13 (76%)
Oxylipins ³	P≤0.05	2	2.9	0	2 (100%)	P≤0.05	11	16.2	0	11 (100%)
n=68	P<0.1	1	1.5	0	1 (100%)	P<0.1	5	7.3	0	5 (100%)
Steroids (assay #1)4	P≤0.05	1	6.7	0	1 (100%)	P≤0.05	1	6.7	0	1 (100%)
n=15	P<0.1	0	0	0	0	P<0.1	2	13.3	0	2 (100%)
Steroids (assay #2)4	P≤0.05	1	16.7	1 (100%)	0	P≤0.05	1	16.7	1 (100%)	0
n=7	P<0.1	0	0	0	0	P<0.1	0	0	0	0
Total	P≤0.05	89	5.8	59 (66%)	30 (34%)	P≤0.05	88	5.7	13 (15%)	75 (85%)
n=1545	P<0.1	89	5.8	50	39	P<0.1	71	4.6	13	58

¹Measured in plasma from male prostate cancer patients (matched for characteristics presented in Table 1).

²Untargeted global metabolomics and lipidomics were performed with the Metabolon plateform.

³Oxylipins were measured by the West Coast Metabolomics Center.

⁴Steroids were measured in-house using two multiplex assays measuring adrenal precursors, androgens and estrogens (assay #1) and 11-oxygenated steroids (assay #2).

^{5%} of measured metabolites.

^{6%} of changed metabolites.

Table S2 – Most changed metabolites in *UGT2B17* KO and *UGT2B28* KO

UGT2B17 KO		UGT2B28 KO	UGT2B28 KO			
5α -androstan- 3β , 17α -diol disulfate	2.07	adipoylcarnitine	2.70			
adipate	1.69	glycerol	1.91			
androsterone sulfate	1.61	sphinganine	1.55			
androstenediol $(3\alpha,17\alpha)$ monosulfate	1.61	2-keto-3-deoxy-gluconate	1.55			
nervonoylcarnitine (C24:1)	1.50	pyruvate	1.53			
5α -androstan- 3α ,17 β -diol monosulfate	1.49	mannonate	1.52			
hydroxy-CMPF ¹	1.46	sphingosine	1.47			
gamma-glutamyl-2-aminobutyrate	1.45	succinoyltaurine	1.31			
epiandrosterone sulfate	1.41	5-(galactosylhydroxy)-L-lysine	1.30			
N-acetylcarnosine	1.39	N-acetylglucosamine/N-acetylgalactosamine	1.29			
deoxycholic acid glucuronide	-8.33	eicosenedioate	-1.82			
trigonelline (N'-methylnicotinate)	-2.33	1-oleoyl-GPG (18:1) ²	-1.72			
isoursodeoxycholate	-2.33	N,N-dimethyl-5-aminovalerate	-1.61			
2,3-dihydroxyisovalerate	-2.22	N6-methyllysine	-1.59			
5-acetylamino-6-amino-3-methyluracil	-1.96	N,N,N-trimethyl-5-aminovalerate	-1.59			
pipecolate	-1.89	isobutyrylcarnitine	-1.49			
N2-acetyl,N6,N6-dimethyllysine	-1.82	HWESASLLR	-1.49			
homocitrulline	-1.61	propionylcarnitine	-1.47			
cysteinylglycine disulfide	-1.56	3β,17α-dihydroxy-5-cholestenoate	-1.45			
5α-androstan-3α,17β-diol 17-glucuronide	-1.56	glutarate	-1.45			
		laurylcarnitine	-1.45			

¹CMPF: 3-carboxy-4-methyl-5-propyl-2-furanpropanoate ²GPG: glycerophosphoglycerol

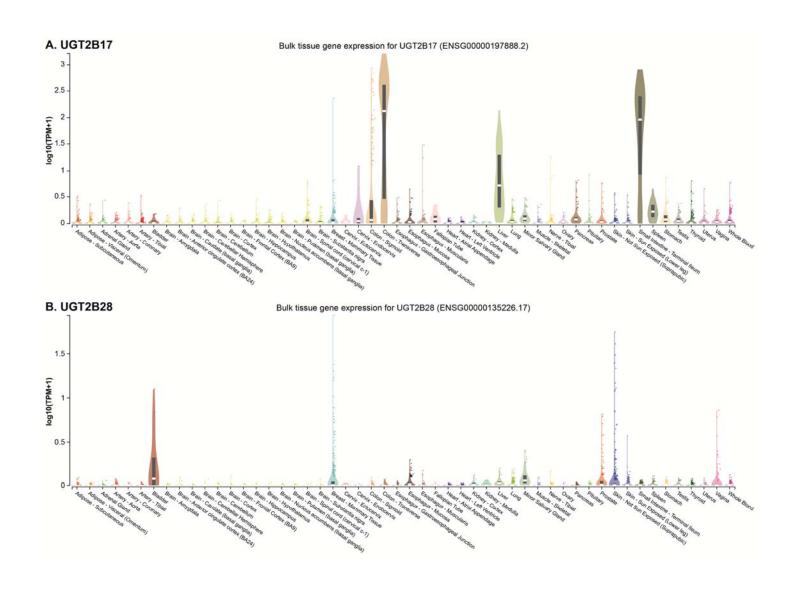


Fig.S1. Tissue expression profiles of UGT2B17 and UGT2B28. Gene expression levels were reported according to the GTEx portal (accessed on June 20, 2022; https://gtexportal.org/home/)

TPM: transcripts per million read.

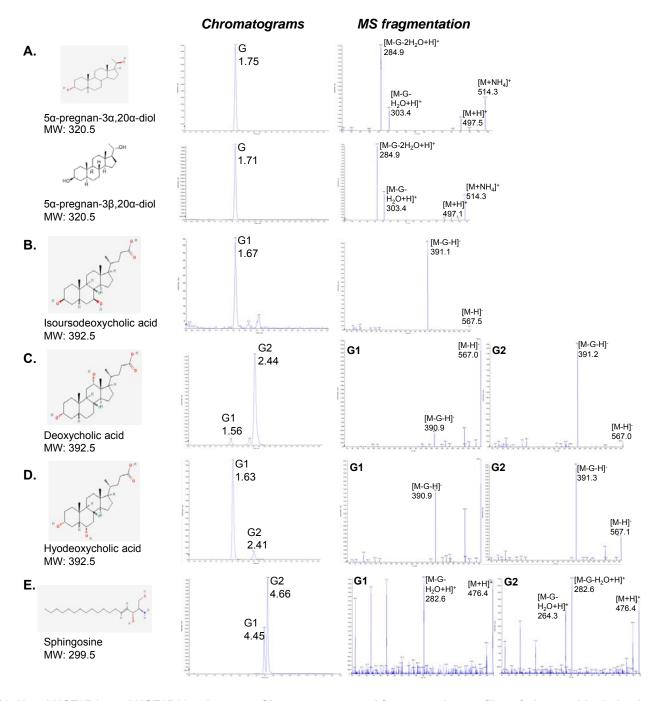
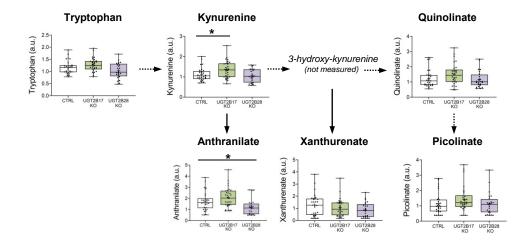


Fig.S2. Novel UGT2B17 and UGT2B28 substrates: Chromatograms and fragmentation profiles of glucuronide derivatives by mass spectrometry analysis (MS). MS analysis indicated the formation of one or two glucuronidated products named G1 and G2 according to their order of elution. A. 5α -pregnandiols glucuronide (G) conjugation by LNCaP prostate cancer cells overexpressing UGT2B28; B-D. Glucuronide of bile acids formed by LAPC4 prostate cancer cells overexpressing UGT2B17. E. Sphingosine glucuronide derivatives formed by LAPC4 prostate cancer cells overexpressing UGT2B28. Glucuronides were also formed by microsomes of human liver, expressing both UGT2B17 and UGT2B28. Chromatograms indicate elution time of each glucuronide. Fragmentation patterns assessed by MS show the loss of the GlcA moiety (M-G) corresponding to a m/z shift of 176 Da relative to the glucuronidated compound (M+H).

A.



В.

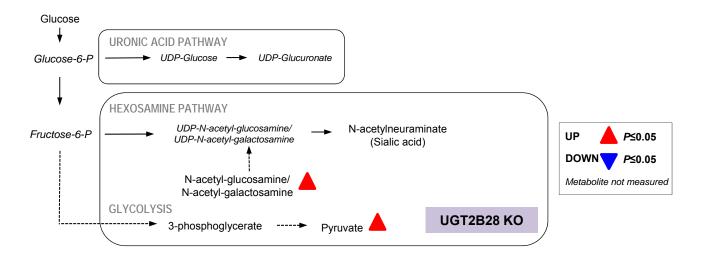


Fig.S3. Distinctive changes in the tryptophan-kynurenine and glucose-derived pathways in *UGT2B17* KO and *UGT2B28* KO. A. Kynurenine is elevated in *UGT2B17* KO individuals. Kynurenine and several other tryptophan-derived metabolites are higher than controls in *UGT2B17* KO. In *UGT2B28* KO, fewer metabolites are changed, with only anthranilate significantly reduced relative to controls. Boxes represent the interquartile range (25-75%), whiskers indicate maximum and minimum values, the median and mean values are indicated by a solid line and a + sign, respectively. Significant changes are indicated. Tryptophan-derived metabolites that were unchanged are not shown. **B.** Metabolic perturbations of the glucose-derived uronic acid and hexosamine pathways in *UGT2B17* KO and *UGT2B28* KO cases. Quantitative data are provided in Table S3A.