



## Immune Crosstalk Between Lymph Nodes and Breast Carcinomas, With a Focus on B Cells

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Lymph nodes (LNs) are highly organized secondary lymphoid organs, and reflective of immune responses to infection, injuries, or the presence of cancer. Extensive molecular and morphological analyses of immune and stromal features in tumors and LNs of breast cancer patients have revealed novel patterns indicative of disease progression. Within LNs, there are dynamic structures called germinal centers (GCs), that act as the immunological hubs for B cell development and generation of affinity matured memory B and antibody-producing plasma cells. Acting as a bridge between systemic and local immunity, associations are observed between the frequency of GCs within cancerfree LNs, the levels of stromal tumor infiltrating lymphocytes, and cancer progression. Scattered throughout the tumor microenvironment (TME) or aggregated in clusters forming tertiary lymphoid structures (TLS), the occurrence of tumor infiltrating B cells (TIL-Bs) has been linked mostly to superior disease trajectories in solid cancers. Recent TIL-Bs profiling studies have revealed a plethora of different TIL-B populations, their functional roles, and whether they are derived from GC reactions in the LN, and/or locally from GC-like structures within the TME remains to be investigated. However, parallels between the immunogenic nature of LNs as a pre-metastatic niche, TIL-B populations within the TME, and the presence of TLS will help to decipher local and widespread TL-Bs responses and their influence on cancer progression to the lymphatics. Therapies that enhance TIL-Bs responses in the LN GC and/or in GC-like structures in the TME are thus emerging management strategies for breast and other cancer patients.

Keywords: lymph node, TILs, tertiary lymphoid structures, germinal centers, breast cancer

## HIGHLIGHTS

- Morphological alterations in cancer-free and cancer-involved axillary lymph nodes hold predictive prognostic information for breast cancer patients.
- Tumor infiltrating B cells (TIL-Bs) can form tertiary lymphoid structures that bear morphological and cellular similarities to germinal centers in lymph nodes.
- TIL-Bs at the primary tumor site and the formation of tertiary lymphoid structures are mostly associated with superior disease trajectories in breast and other solid cancers.

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1

- Deep single-cell profiling has revealed a plethora of markers and new TIL-B populations, and for many their functional role remains to be determined.
- Exploring the responses of TIL-Bs at the primary tumor, in peritumoral germinal center-like structures and in lymph nodes will inform the development of novel therapies.

## INTRODUCTION

Lymph nodes (LNs) are secondary lymphoid organs that act as a platform to facilitate antigen dispersal and promote interactions between immune cell subsets. In response to disease, inflammatory chemokines and cytokines mediate the recruitment of lymphocytes and antigen-presenting cells that access the LNs via lymphatic vessel-mediated lymph drainage. Passing through the subcapsular sinus, these lymph-borne solutes disseminate into the cortex where B cells, intrafollicular T cells, and dendritic cells are arranged in a highly size-restricted manner, before moving through conduits to reach the T cell zone of the LN paracortex. Depending on the nature of the stimuli, these compartments can expand or diminish to generate an optimal B cell response. Eventually, lymphocytes exit the LNs via efferent lymphatic vessels, and the LNs return to a naïve, resting state (Willard-Mack, 2006).

Despite serving as transportation channels essential to an effective immune response, the lymphatics also act as corridors for cancerous cells to pass through into the LNs (Stacker et al., 2014; Ji, 2016). Typically presenting as the initial seeding site outside of the primary tumor, the presence of metastatic growth within LNs has been associated with both shortened disease-free survival and a heightened risk of developing metastases in distant organs. Therefore, the incidence of cancer cells within the LNs, the number of metastatic LNs, and the occurrence of extra nodal extension have formed essential assessment parameters for routine pathological diagnosis of several cancers, including breast cancer.

Historically, axillary LN clearance was undertaken for all patients with invasive breast cancer. Today, the standard treatment of care for patients with clinically and radiologically negative nodes prior to surgery is surgical resection of only the nodes adjacent to and draining from the tumor bed, the socalled sentinel LNs. It is also becoming more common for breast cancer patients to receive neoadjuvant chemotherapy (NACT), which presents new challenges for the assessment of the LNs. Treatment-induced fibrosis and reactive changes can obscure the local environment and prevent an accurate diagnosis of LN metastasis. Thus, exploring the spatial, cellular, and molecular alterations in LNs occurring due to immune surveillance of nearby tumor growth, as well as from NACT or immunotherapy will expand our understanding of the immune crosstalk between LNs and breast carcinomas.

Germinal centers (GC) are immunological sites in the LN within which B cell receptor (BCR) affinity maturation occurs to generate long-lived memory B and plasma cells (Victora and Nussenzweig, 2012; Nakagawa et al., 2021). Tightly regulated mechanisms within the GC promote targeted responses to

pathogens whilst ensuring the elimination of autoreactive clones. B cells retrieve antigen via their BCR from follicular dendritic cells (FDCs) and present that antigen to follicular T helper (Tfh) cells (Victora and Nussenzweig, 2012). GC B cells compete for Tfh derived signals critical for positive selection, which culminates in the upregulation of the transcription factor MYC (Calado et al., 2012; Dominguez-Sola et al., 2012). BCR affinity maturation in GC B cells occurs through iterative rounds of clonal expansion, somatic hypermutation and selection to generate fine-tuned humoral responses (Victora and Nussenzweig, 2012). Owing to their highly organized structures, morphological alterations in GCs indicates fluctuations in their molecular mechanisms and polarization, which ultimately has an impact on the quality and quantity of the resulting memory B and plasma cell populations (Victora et al., 2010; Bannard et al., 2013; Toboso-Navasa et al., 2020). Early autochthonous models of breast cancer showed enlarged GCs in regional LNs with increased levels of lymphoblasts, a sign of active lymphopoieses, and clusters of dividing plasma cells in the vicinity of the GC (Ciocca, 1980).

Based on extensive H&E assessment of immune and stromal features at primary breast carcinomas and patient-matched axillary LNs, we were the first to report on histological changes in cancer-free LNs carrying additive risk predictive value for developing distant metastasis (Grigoriadis et al., 2018). By incorporating the number and size of GCs in cancer-free LNs, combined with the level of stromal tumor-infiltrating lymphocytes (sTILs) at the primary tumor, the presence of lymphocytic lobulitis, and location of GCs in the involved LN, an immune-stroma-histological (ISH)-risk score was implemented indicative for the risk of developing distant metastasis for breast cancer patients (Figure 1). Amongst the 309 breast cancer patients enriched of triple-negative phenotype and of high histological grade, a group of LN-positive patients with the lowest quartile ISH scores showed a superior outcome even when compared to LN-negative breast cancer patients. In particular, the number of GC formations in cancer-free LNs added valuable information to TILs in triple-negative breast cancer prognostication (Liu et al., 2021). Multiple questions emerge from this clinically relevant association, and further investigations are needed to dissect the mechanisms driving the formation of LN GCs in breast cancer patients and to understand the function of GC derived memory B and plasma cells in this context.

## TUMOR INFILTRATING B CELL RESPONSES AT THE PRIMARY BREAST CARCINOMAS

Triple-negative breast cancers (TNBCs), defined by their lack of expression of estrogen receptor, progesterone receptor, and human epidermal growth factor receptor 2 (HER2), are associated with a poorer prognosis and higher rates of distant recurrence compared to receptor-positive breast cancers (Reddy et al., 2018; Yin et al., 2020). Crucially, lymphatic involvement is more prevalent in TNBC and contributes to both local and



distant metastasis (Shen et al., 2014; van Roozendaal et al., 2016; Yao et al., 2019). Both TNBC and HER2-positive breast cancers often present with more immune infiltrated tumors compared to other subtypes, and sTIL assessment of these breast carcinomas have been shown to be superior to classical TNM staging when predicting outcome and response to anti-HER2 therapy, chemotherapy, and immunotherapy (Salgado et al., 2015; Ignatiadis et al., 2019; Loi et al., 2019, 2020). In contrast to the longstanding characterization and manipulation of tumor infiltrating T cells (TIL-Ts) for clinical research, the potential of tumor infiltrating B cells, denoted (TIL-Bs), has only recently sparked interest amongst breast cancer researchers. The majority of breast carcinomas present with relatively low levels of TIL-Bs ( $\sim$ 2–3%), yet this is heightened compared to healthy breast tissue (Garaud et al., 2019). A variety of TIL-Bs at multiple stages of differentiation, including naïve, GC-like, memory-like, and plasma cells (Chung et al., 2017; Garaud et al., 2019) have been reported within and around the tumor microenvironment (TME) of breast tumors, with a high percentage of TIL-B exhibiting a memory-like phenotype (Buisseret et al., 2017). Notably, the occurrence of GC-like B cells consistently correlates with numbers of Tfh cells, signifying an active and constantly evolving humoral response (Garaud et al., 2019). Using multiplexed ion beam imaging to analyze spatial information, TIL-B populations were found to be consistently depleted along the tumor border of TNBCs (Keren et al., 2018), however the cause of this relative reduction is unknown. The spatial patterns and colocalizations with other cell types further indicate the involvement of TIL-Bs

in humoral immunity, with possible roles in antigen presentation and modulation of other immune populations with relevance to tumor progression. Particularly in TNBC patients, increased levels of TIL in residual disease post-NACT has been associated with a better prognosis (Dieci et al., 2014). Specifically, the presence of TIL-B at primary tumor lesions is shown to be an independent predictor of NACT response (García-Martínez et al., 2014), and the coexistence of TIL-B and PD-L1-positive carcinoma cells is significantly associated with a high pathological complete response rate and overall survival (Arias-Pulido et al., 2018). Treatment-induced B cell lymphopenia, often seen to predominantly deplete class-switched memory B cells, may impact B cell orientated protection against tumor progression (Mellios et al., 2010; Verma et al., 2016; Gustafson et al., 2020). It is important to consider that TIL-Bs are shown to re-populate at a much slower rate than other immune cells after chemotherapy (Verma et al., 2016; Gustafson et al., 2020), as post-treatment profiles of patients with high TIL-B levels identifies a highly responsive group of breast cancer patients (Song et al., 2017).

## TUMOR INFILTRATING B CELLS IN TERTIARY LYMPHOID STRUCTURES

In addition to antibody production and antigen presentation, B cells may contribute to peritumoral immunity by associating with T cells to form organized structures known as tertiary lymphoid structures (TLS) (Germain et al., 2015; Shen et al., 2018). In

comparison with GCs in LNs, TLSs are non-encapsulated, transient structures induced and maintained in chronically inflamed tissues. The presence of TLS has been correlated with higher TIL-B levels in the peritumoral area and is frequently associated with superior disease free and overall survival (Figenschau et al., 2015; Lee et al., 2016; Sautès-Fridman et al., 2019). Mature TLS bear similar morphological and molecular characteristics to secondary lymphoid follicles, forming a definitive marginal zone, mantle zone and a central GC-like structure in which B cell centroblast, centrocyte subsets, and Tfh cells are found (Garaud et al., 2019). Somatic mutations of the Ig variable domains of TIL-Bs isolated from T/B clusters in ductal carcinomas revealed evidence of local oligoclonal expansion of cells that have previously undergone antigen-driven hypermutation, proliferation, and affinity maturation, much like within a GC (Nzula et al., 2003). To facilitate recruitment, positioning and interactions within GCs, Tfh cells can express the checkpoint molecules ICOS and PD-1 that engage with cognate ICOSL<sup>+</sup>PD-L1<sup>+</sup> centrocytes (Shi et al., 2018). Although the proportion of PD-1<sup>+</sup> and PD-L1<sup>+</sup> TILs is low in breast cancers, much of the PD-1 is primarily expressed on T cells that can also localize within the TLS. This brings to light the impact of PD-1 as a marker of immune activation in contrast to its connotations with immune exhaustion in the context of cancer. Moreover, expression of PD-1 and PD-L1 in TILs is significantly associated with improved clinical outcomes in TNBC and HER2-enriched breast cancers (Solinas et al., 2017; Schmid et al., 2018).

Yet to be explored in breast cancer, detailed phenotypic characterizations of TLS within other solid cancers such as hepatocellular carcinoma or colorectal cancers identified those of an "early stage" and "immature" phenotype lacking a central GC-like structure (Meylan et al., 2020). In patients with nonmetastatic colorectal cancer, tumors presenting with more than one mature TLS that harbors active GC-like interactions were associated with a significantly reduced risk of recurrence compared to patients with solely immature TLS (Posch et al., 2017). In contrast, in oral squamous cell carcinoma patients, no differences in overall or recurrence free survival was observed when the density of immature to mature TLS was compared (Li et al., 2020). In lung squamous cell carcinoma, the existence of TLS was the strongest independent factor when patients were untreated, whilst in patients treated with NACT and corticosteroid therapy, the development of mature TLS seemed to be impaired and were not informative of disease progression (Siliòa et al., 2018). In immature TLS, TIL-B might interact more with cancerous cells than with T cells, and one hypothesis is that those TIL-B release factors that dampen the response of other immune cells, in turn hindering the targeting and destruction of tumor cells. Three recent studies provide indirect evidence that immature TLS are associated with low activity of T cells in tumors, whilst mature TLS nurture B cell development (Cabrita et al., 2020; Helmink et al., 2020; Petitprez et al., 2020). The abovelisted disease-specific findings highlight the need to explore in depth the formation and the development of immature and mature TLS responses within the context of breast carcinomas, to establish their drivers, their pro/anti-tumor properties, and the potential impact of neoadjuvant treatment on their formation.

## TRANSCRIPTIONAL PROFILING OF THE GC REACTION AND OF GC B CELL DERIVED SUBSETS

Tightly controlled transcriptional profiles govern the formation and development of GCs, and potentially by extension TLS. In the LN, GC B cells transit between two functionally distinct compartments, the dark zone (DZ) and light zone (LZ) which represent polarized areas in which gene expression patterns drive somatic hypermutation and selection, respectively (Calado et al., 2012; Dominguez-Sola et al., 2012; Victora and Nussenzweig, 2012). By utilizing single-cell RNA-sequencing (scRNA-seq) on tissue derived from human and mouse, a plethora of transcriptional changes occurring in GCs has been revealed, and with it novel cell populations have been defined (McHeyzer-Williams et al., 2015; Holmes et al., 2020; Kennedy et al., 2020; King et al., 2021; Nakagawa et al., 2021). Whilst the functional roles for some of these populations remain to be determined, the identification of defining markers provides the opportunity to determine their spatial distribution and possible isolation for further study. scRNA-seq experiments have also tentatively identified the gene expression profiles of memory B and plasma cell precursors in the LZ of GCs. The differentiation of LZ B cells toward the plasma cell fate is associated with increased Tfh help that enhances NF-KB signaling, IRF4, XBP1, FKBP11, and PRDM1 (BLIMP1) expression (Klein et al., 2006; Heise et al., 2014; Nutt et al., 2015; Ruer-Laventie et al., 2015; Ise et al., 2018; Holmes et al., 2020). In contrast, memory B cell differentiation from LZ B cells is restricted to positively selected cells, seemingly requires minimal Tfh help and associates with increased BACH2 and HHEX1 expression levels (Sidwell and Kallies, 2016; Laidlaw and Cyster, 2020; Laidlaw et al., 2020; Toboso-Navasa et al., 2020; Nakagawa et al., 2021). The transposition of these datasets to TLS in breast cancer, together with the ability to record the temporal, spatial and transcriptional profiles of GCs and TLS may further our understanding of the TIL-B populations within the TME and provide a rationale for their contribution to disease progression.

## ANTITUMOR AND AUTOANTIBODY PRODUCTION IN BREAST CANCER

As a product of the GC response, plasma cells that have undergone somatic hypermutation and affinity maturation are typically long lived and capable of evoking a humoral response for many years (Brynjolfsson et al., 2018). By contrast, those that develop in extrafollicular foci do not undergo somatic hypermutation, are typically short-lived, and secrete a combination of switched or unswitched antibodies (Paus et al., 2006). Comprehensive gene expression studies of TIL-B populations in breast cancer identified IgG-associated gene sets in primary carcinomas indicative of pathological complete response to trastuzumab combination therapies and superior overall survival in TNBC (Perou et al., 1999; Carey et al., 2014; Iglesia et al., 2014). Spatial analysis of such antibody responses revealed that breast lesions with high levels of tumor infiltrating



**FIGURE 2** Graphical depiction of GC regulation. Within the lymph nodes (LNs), B cells primed by T cells at the T/B border move into the GC and proliferate rapidly in the dark zone (DZ), before transitioning into the light zone. Here, engagement with T follicular helper cells through checkpoint inhibitor mechanism and cytokines such as IL-21 ensures affinity-based maturation of GC B cells. Upon upregulation of transcriptionally distinct profiles, the B cells differentiate into either a memory B cell, antibody secreting plasma cell or return to the DZ for additional proliferative rounds and hypermutation. Within the context of breast cancer, antibody production has been linked to both the inhibition and promotion of tumor progression. Regulatory B cell subsets can regulate this mechanism by suppressing Tfh mediated activation of B cells and promote an immune tolerant environment through secretion of IL-10. The transition of cancerous cells from the primary tumor to the lymphatics may be accompanied by a multitude of immune cell subsets, including antigen presenting cells. Further communication between LN and tumor and potential migration of B cell subsets, cytokines, and chemokines may be informative of how the GC responses can influence the tumor microenvironment. As the morphology of tertiary lymphoid structures is comparable to that of a B cell follicle, there may be mechanistic parallels that can be drawn between the two structures (created with BioRender.com).

plasma cells present with antibodies in their tumor core, at the invasive margin and within the stromal compartments (Seow et al., 2020). Some of these antibodies bind tumor cells and display a clonal relationship with those present in the axillary LNs, indicative of a systemic response beyond the local TME (Novinger et al., 2015). Supporting a functional role for antibodies in breast cancer, mice deficient for antibody production display a more aggressive disease progression, and the adoptive transfer of IgG secreting plasma cells present in tumor draining LNs limits metastatic spread (Li et al., 2011; Tao et al., 2013; Brynjolfsson et al., 2018; Hollern et al., 2019). However, the antigen specificity of these functionally relevant antibodies is not completely understood. Conversely, the analysis of the IgG and IgA autoantibody repertoire in breast cancer patients revealed that autoantibodies to one or more tumor-associated antigens occurred in most patients. Notably, patients with a higher level of IgG reactivity to breast cancer-associated antigens have significantly shorter recurrence free survival (Garaud et al., 2018). These findings align with studies of spontaneous LN metastasis breast cancer mouse models. Here, the presence of IgG antibodies to a breast cancer antigen promoted tumor progression through the lymphatics (Gu et al., 2019). It remains unclear whether GC reactions contribute to the production of protective and/or tumor promoting antibodies, and the extent to which GC reactions in the context of breast cancer follow the canonical checkpoints that curb self-reactivity in physiology. This knowledge is clinically

relevant as it may provide insight for strategies that selectively inhibit the development of tumor promoting antibodies and enhance cancer-protective humoral immunity.

# IMMUNE TOLERANCE AND REGULATION

An appropriate immune response relies on a "goldilocks window" of immune checkpoint control; too little regulation promotes the expansion of autoreactive cells, whereas exacerbated expression leads to anergy and exhaustion. Tolerance of GC responses is in part maintained by T follicular regulatory (Tfr) cells, that dampen immune responses by preventing CD28-B7 co-stimulatory interactions through CTLA-4 engagement (Miles and Connick, 2018). Circulating Tfr and Treg cells are enriched in breast cancer patients, particularly in more aggressive cancers (Kohrt et al., 2005; Song et al., 2019; Núñez et al., 2020). This may correlate with the knowledge that Tfr cells potently inhibit antigenspecific antibody responses (Linterman et al., 2011). Tfr and Treg cells can induce an immunosuppressive microenvironment, often through IL-10 production, and promote the expansion of immunosuppressive B cells, so-called Bregs. In a cyclical fashion, IL-10 secreting Bregs support the Treg pool and impede Tfh responses within the GC (Liu et al., 2016; Achour et al., 2017; Song et al., 2019). The number and localization of Bregs in breast

carcinomas strongly fluctuates with levels of Tregs, notably in and around B and T cell TIL aggregates (Guan et al., 2016; Ishigami et al., 2019). Supporting a relationship between Bregs and Tregs, a B cell deficient breast cancer model displayed a reduced fraction of Tregs in tumor draining LNs and peritumoral areas (Tadmor et al., 2011). Accumulation of Tregs and Bregs within the cancer-free LNs of breast cancer patients correlates with fewer class-switched B cells in adjacent LNs with cancerous growth, indicative of a possible role in GC suppression (Mehdipour et al., 2016). This synergistic relationship between Bregs and Tregs, their ability to promote a tolerant environment and suppress Tfh cells may influence the efficacy of GC reactions within breast cancer patients (Figure 2). Supporting this hypothesis, IL-10 blockade in vivo stimulates IgG production and enhances immune infiltration within the primary breast carcinoma (Zheng et al., 2012; Li and Xia, 2015; Tao et al., 2015).

Bregs express the immune checkpoint molecules PD-1 and PD-L1 which have immunomodulatory functions (Sun et al., 2019). The emerging role of Bregs in promoting immunosuppression in breast cancer may indicate these cells as candidate targets in immune-checkpoint (ICP) blockade therapy. ICP blockades have been shown to promote the proliferation of class-switched memory B cells and enhance antibody production *in vitro* and within the context of chronic infection (Pioli et al., 2000; Velu et al., 2009; Intlekofer and Thompson, 2013). However, a potential effect of immune checkpoint inhibitors on the GC reactions must be considered given that GC B cells themselves express PD-L1, and Tfh cells require many of these immune checkpoint interactions including the PD-1/PD-L1 axis and CTLA-4 for B cell affinity maturation (Good-Jacobson et al., 2010; Hams et al., 2011).

## CONCLUSION AND FUTURE DIRECTIONS

During the past decade, molecular and cellular parallels between GCs in LNs and TLS formation at the primary tumor suggest

#### REFERENCES

- Achour, A., Simon, Q., Mohr, A., Séité, J. F., Youinou, P., Bendaoud, B., et al. (2017). Human regulatory B cells control the TFH cell response. J. Allergy Clin. Immunol. 140, 215–222. doi: 10.1016/j.jaci.2016.09.042
- Arias-Pulido, H., Cimino-Mathews, A., Chaher, N., Qualls, C., Joste, N., Colpaert, C., et al. (2018). The combined presence of CD20 + B cells and PD-L1 + tumorinfiltrating lymphocytes in inflammatory breast cancer is prognostic of improved patient outcome. *Breast Cancer Res. Treat.* 171, 273–282. doi: 10. 1007/s10549-018-4834-7
- Bannard, O., Horton, R. M., Allen, C. D., An, J., Nagasawa, T., and Cyster, J. G. (2013). Germinal center centroblasts transition to a centrocyte phenotype according to a timed program and depend on the dark zone for effective selection. *Immunity* 39, 912–924. doi: 10.1016/j.immuni.2013.08. 038
- Brynjolfsson, S. F., Persson Berg, L., Olsen Ekerhult, T., Rimkute, I., Wick, M. J., Mårtensson, I. L., et al. (2018). Long-lived plasma cells in mice and men. *Front. Immunol.* 9:2673. doi: 10.3389/fimmu.2018.02673
- Buisseret, L., Garaud, S., de Wind, A., van den Eynden, G., Boisson, A., Solinas, C., et al. (2017). Tumor-infiltrating lymphocyte composition, organization and PD-1/ PD-L1 expression are linked in breast cancer. *Oncoimmunology* 6:e1257452. doi: 10.1080/2162402X.2016.1257452

a degree of communication via the lymphatics between the sites and revealed a key role for B cells in breast and other cancers. However, we do not understand the processes that render B cell activation able to eradicate neoplastic cells through immunoglobulin-mediated mechanisms versus those leading to chronic B cell activation that potentiates tumor progression. By facilitating an immune crosstalk between LNs, TLS and primary carcinomas, B cells may modulate the dynamic interplay between immune responses and tumor progression. Uncovering the mechanistic drivers that influence specific B cell environments at the tumor side, peritumoral and in the LNs may provide attractive targets, many of which could be incorporated into current immunotherapies to treat both breast and other cancers.

#### **AUTHOR CONTRIBUTIONS**

EA, IW, DC, and AG wrote a complete draft and first version of the manuscript. All authors contributed to the manuscript revision, read, and approved final version and contributed to the principal layout of the article.

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- Cabrita, R., Lauss, M., Sanna, A., Donia, M., Skaarup Larsen, M., Mitra, S., et al. (2020). Tertiary lymphoid structures improve immunotherapy and survival in melanoma. *Nature* 577, 561–565. doi: 10.1038/s41586-019-1914-8
- Calado, D., Sasaki, Y., Godinho, S., Pellerin, A., Köchert, K., Sleckman, B. P., et al. (2012). MYC is essential for the formation and maintenance of germinal centers. *Nat. Immunol.* 13, 1092–1100. doi: 10.1038/ni.2418
- Carey, L. A., Barry, W. T., Pitcher, B., Hoadley, K. A., Cheang, M. C. U., AndersN, C. K., et al. (2014). Gene expression signatures in pre- and post-therapy (Rx) specimens from CALGB 40601 (Alliance), a neoadjuvant phase III trial of weekly paclitaxel and trastuzumab with or without lapatinib for HER2-positive breast cancer (BrCa). J. Clin.Oncol. 32(Suppl. 15), 506–506. doi: 10.1200/jco. 2014.32.15\_suppl.506
- Chung, W., Eum, H. H., Lee, H. O., Lee, K. M., Lee, H. B., Kim, K. T., et al. (2017). Single-cell RNA-seq enables comprehensive tumour and immune cell profiling in primary breast cancer. *Nat. Commun.* 8:15081. doi: 10.1038/ncomms15081
- Ciocca, D. R. (1980). Immunomorphologic lymph node changes in rats bearing experimental breast tumors. *Am. J. Pathol.* 99, 193–206.
- Dieci, M. V., Criscitiello, C., Goubar, A., Viale, G., Conte, P., Guarneri, V., et al. (2014). Prognostic value of tumor-infiltrating lymphocytes on residual disease after primary chemotherapy for triple-negative breast cancer: a retrospective multicenter study. Ann. Oncol. 25, 611–618. doi: 10.1093/annonc/mdt556

- Dominguez-Sola, D., Victora, G., Ying, Y. C., Phan, R. T., Saito, M., Nussenzweig, M. C., et al. (2012). The proto-oncogene MYC is required for selection in the germinal center and cyclic reentry. *Nat. Immunol.* 13, 1083–1091. doi: 10.1038/ ni.2428
- Figenschau, S. L., Fismen, S., Fenton, K. A., Fenton, C., and Mortensen, E. S. (2015). Tertiary lymphoid structures are associated with higher tumor grade in primary operable breast cancer patients. *BMC Cancer* 15:101. doi: 10.1186/s12885-015-1116-1
- Garaud, S., Buisseret, L., Solinas, C., Gu-Trantien, C., de Wind, A., van den Eynden, G., et al. (2019). Tumor infiltrating B-cells signal functional humoral immune responses in breast cancer. *JCI Insight* 5:e129641. doi: 10.1172/jci. insight.129641
- Garaud, S., Zayakin, P., Buisseret, L., Rulle, U., Silina, K., de Wind, A., et al. (2018). Antigen specificity and clinical significance of IgG and IgA autoantibodies produced in situ by tumor-infiltrating B cells in breast cancer. *Front. Immunol.* 9:2660. doi: 10.3389/fimmu.2018.02660
- García-Martínez, E., Gil, G. L., Benito, A. C., González-Billalabeitia, E., Vicente Conesa, M. A., García, T. G., et al. (2014). Tumor-infiltrating immune cell profiles and their change after neoadjuvant chemotherapy predict response and prognosis of breast cancer. *Breast Cancer Res.* 16:488. doi: 10.1186/s13058-014-0488-5
- Germain, C., Gnjatic, S., and Dieu-Nosjean, M. C. (2015). Tertiary lymphoid structure-associated B cells are key players in anti-tumor immunity. *Front. Immunol.* 6:67. doi: 10.3389/fimmu.2015.00067
- Good-Jacobson, K. L., Szumilas, C. G., Chen, L., Sharpe, A. H., Tomayko, M. M., and Shlomchik, M. J. (2010). PD-1 regulates germinal center B cell survival and the formation and affinity of long-lived plasma cells. *Nat. Immunol.* 11, 535–542. doi: 10.1038/ni.1877
- Grigoriadis, A., Gazinska, P., Pai, T., Irhsad, S., Wu, Y., Millis, R., et al. (2018). Histological scoring of immune and stromal features in breast and axillary lymph nodes is prognostic for distant metastasis in lymph node-positive breast cancers. J. Pathol. Clin. Res. 4, 39–54. doi: 10.1002/cjp2.87
- Gu, Y., Liu, Y., Fu, L., Zhai, L., Zhu, J., Han, Y., et al. (2019). Tumor-educated B cells selectively promote breast cancer lymph node metastasis by HSPA4-targeting IgG. Nat. Med. 25, 312–322. doi: 10.1038/s41591-018-0309-y
- Guan, H., Wan, Y., Lan, J., Wang, Q., Wang, Z., Li, Y., et al. (2016). PD-L1 is a critical mediator of regulatory B cells and T cells in invasive breast cancer. *Sci. Rep.* 6:35651. doi: 10.1038/srep35651
- Gustafson, C. E., Jadhav, R., Cao, W., Qi, Q., Pegram, M., Tian, L., et al. (2020). Immune cell repertoires in breast cancer patients after adjuvant chemotherapy. *JCI Insight* 5:e134569. doi: 10.1172/jci.insight.134569
- Hams, E., McCarron, M. J., Amu, S., Yagita, H., Azuma, M., Chen, L., et al. (2011). Blockade of B7-H1 (Programmed Death Ligand 1) enhances humoral immunity by positively regulating the generation of T follicular helper cells. *J. Immunol.* 186, 5648–5655. doi: 10.4049/jimmunol.1003161
- Heise, N., de Silva, N. S., Silva, K., Carette, A., Simonetti, G., Pasparakis, M., et al. (2014). Germinal center B cell maintenance and differentiation are controlled by distinct NF-KB transcription factor subunits. *J. Exp. Med.* 211, 2103–2118. doi: 10.1084/jem.20132613
- Helmink, B. A., Reddy, S. M., Gao, J., Zhang, S., Basar, R., Thakur, R., et al. (2020).
  B cells and tertiary lymphoid structures promote immunotherapy response. *Nature* 577, 549–555. doi: 10.1038/s41586-019-1922-8
- Hollern, D. P., Xu, N., Thennavan, A., Glodowski, C., Garcia-Recio, S., Mott, K. R., et al. (2019). B cells and T follicular helper cells mediate response to checkpoint inhibitors in high mutation burden mouse models of breast cancer. *Cell* 179, 1191–1206.e21. doi: 10.1016/j.cell.2019.10.028
- Holmes, A. B., Corinaldesi, C., Shen, Q., Kumar, R., Compagno, N., Wang, Z., et al. (2020). Single-cell analysis of germinal-center B cells informs on lymphoma cell of origin and outcome. *J. Exp. Med.* 217:e20200483. doi: 10.1084/jem.2020 0483
- Iglesia, M. D., Vincent, B. G., Parker, J. S., Hoadley, K. A., Carey, L. A., Perou, C. M., et al. (2014). Prognostic B-cell signatures using MRNA-seq in patients with subtype-specific breast and ovarian cancer. *Clin. Cancer Res.* 20, 3818–3829. doi: 10.1158/1078-0432.CCR-13-3368
- Ignatiadis, M., van den Eynden, G., Roberto, S., Fornili, M., Bareche, Y., Desmedt, C., et al. (2019). Tumor-infiltrating lymphocytes in patients receiving trastuzumab/pertuzumab-based chemotherapy: a TRYPHAENA substudy. *J. Natl. Cancer Inst.* 111, 69–77. doi: 10.1093/jnci/djy076

- Intlekofer, A. M., and Thompson, C. B. (2013). At the bench: preclinical rationale for CTLA-4 and PD-1 blockade as cancer immunotherapy. J. Leukoc. Biol. 94, 25–39. doi: 10.1189/jlb.1212621
- Ise, W., Fujii, K., Shiroguchi, K., Ito, A., Kometani, K., Takeda, K., et al. (2018). T follicular helper cell-germinal center B cell interaction strength regulates entry into plasma cell or recycling germinal center cell fate. *Immunity* 48, 702–715.e4. doi: 10.1016/j.immuni.2018.03.027
- Ishigami, E., Sakakibara, M., Sakakibara, J., Masuda, T., Fujimoto, H., Hayama, S., et al. (2019). Coexistence of regulatory B cells and regulatory T cells in tumorinfiltrating lymphocyte aggregates is a prognostic factor in patients with breast cancer. *Breast Cancer* 26, 180–189. doi: 10.1007/s12282-018-0910-4
- Ji, R.-C. (2016). Lymph nodes and cancer metastasis: new perspectives on the role of intranodal lymphatic sinuses. Int. J. Mol. Sci. 18:51. doi: 10.3390/ ijms18010051
- Kennedy, D. E., Okoreeh, M. K., Maienschein-Cline, M., Ai, J., Veselits, M., McLean, K. C., et al. (2020). Novel specialized cell state and spatial compartments within the germinal center. *Nat. Immunol.* 21, 660–670. doi: 10.1038/s41590-020-0660-2
- Keren, L., Bosse, M., Marquez, D., Angoshtari, R., Jain, S., Varma, S., et al. (2018). A structured tumor-immune microenvironment in triple negative breast cancer revealed by multiplexed ion beam imaging. *Cell* 174, 1373–1387.e19. doi: 10. 1016/j.cell.2018.08.039
- King, H. W., Orban, N., Riches, J. C., Clear, A. J., Warnes, G., Teichmann, S. A., et al. (2021). Single-cell analysis of human B cell maturation predicts how antibody class switching shapes selection dynamics. *Sci. Immunol.* 6:eabe6291. doi: 10.1126/sciimmunol.abe6291
- Klein, U., Casola, S., Cattoretti, G., Shen, Q., Lia, M., Mo, T., et al. (2006). Transcription factor IRF4 controls plasma cell differentiation and class-switch recombination. *Nat. Immunol.* 7, 773–782. doi: 10.1038/ni1357
- Kohrt, H. E., Nouri, N., Nowels, K., Johnson, D., Holmes, S., and Lee, P. P. (2005). Profile of immune cells in axillary lymph nodes predicts disease-free survival in breast cancer. *PLoS Med.* 2:e284. doi: 10.1371/journal.pmed.0020284
- Laidlaw, B. J., and Cyster, J. G. (2020). Transcriptional regulation of memory B cell differentiation. *Nat. Rev. Immunol.* 21, 209–220. doi: 10.1038/s41577-020-00446-2
- Laidlaw, B. J., Duan, L., Xu, Y., Vazquez, S. E., and Cyster, J. G. (2020). The transcription factor Hhex cooperates with the corepressor Tle3 to promote memory B cell development. *Nat. Immunol.* 21, 1082–1093. doi: 10.1038/ s41590-020-0713-6
- Lee, H. J., Park, I. A., Song, I. H., Shin, S. J., Kim, J. Y., Yu, J. H., et al. (2016). Tertiary lymphoid structures: prognostic significance and relationship with tumour-infiltrating lymphocytes in triple-negative breast cancer. *J. Clin. Pathol.* 69, 422–430. doi: 10.1136/jclinpath-2015-203089
- Li, Q., and Xia, Y. (2015). "Antitumor effector B cells directly kill tumor cells involving the CXCL12/CXCR4 pathway and their therapeutic efficacy is enhanced by IL-2. Journal for immunotherapy of cancer," in *Proceedings of the 30th Annual Scientific Meeting of the Society for Immunotherapy of Cancer, SITC 2015*, Vol. 3(Suppl. 2), National Harbor, MD, 27.
- Li, Q., Lao, X., Pan, Q., Ning, N., Yet, J., Xu, Y., et al. (2011). Adoptive transfer of tumor reactive B cells confers host T-cell immunity and tumor regression. *Clin. Cancer Res.* 17, 4987–4995. doi: 10.1158/1078-0432.CCR-11-0207
- Li, Q., Liu, X., Wang, D., Wang, Y., Lu, H., Wen, S., et al. (2020). Prognostic value of tertiary lymphoid structure and tumour infiltrating lymphocytes in oral squamous cell carcinoma. *Int. J. Oral Sci.* 12:24. doi: 10.1038/s41368-020-00092-3
- Linterman, M. A., Pierson, W., Lee, S. K., Kallies, A., Kawamoto, S., Rayner, T. F., et al. (2011). Foxp3+ follicular regulatory T cells control the germinal center response. *Nat. Med.* 17, 975–982. doi: 10.1038/nm.2425
- Liu, F., Hardiman, T., Gazinska, P., Quist, J., Pinder, S., and Grigoriadis, A. (2021). Abstract PS6-36: the formation of GCs in cancer-free ALNs, a non-monotonic prognostic factor in HR-negative invasive breast cancer patients. *Cancer Res.* 81(Suppl. 4):PS6-36. doi: 10.1158/1538-7445.SABCS20-PS6-36
- Liu, Y., Cheng, L. S., Wu, S. D., Wang, S. Q., Li, L., She, W. M., et al. (2016). IL-10-producing regulatory B-cells suppressed effector T-cells but enhanced regulatory T-cells in chronic HBV infection. *Clin. Sci.* 130, 907–919. doi: 10. 1042/CS20160069
- Loi, S., Drubay, D., Adams, S., Pruneri, G., Francis, P. A., Lacroix-Triki, M., et al. (2019). Tumor-infiltrating lymphocytes and prognosis: a pooled individual

patient analysis of early-stage triple-negative breast cancers. J. Clin. Oncol. 37, 559–569. doi: 10.1200/JCO.18.01010

- Loi, S., Winer, E., Lipatov, O., Im, S.-A., Goncalves, A., Cortes, J., et al. (2020). Abstract PD5-03: relationship between tumor-infiltrating lymphocytes (TILs) and outcomes in the KEYNOTE-119 study of pembrolizumab vs chemotherapy for previously treated metastatic triple-negative breast cancer (MTNBC). *Cancer Res.* 80(Suppl. 4):PD5-03. doi: 10.1158/1538-7445.SABCS19-PD5-03
- McHeyzer-Williams, L. J., Milpied, P. J., Okitsu, S. L., and McHeyzer-Williams, M. G. (2015). Class-switched memory B cells remodel BCRs within secondary germinal centers. *Nat. Immunol.* 16, 296–305. doi: 10.1038/ni.3095
- Mehdipour, F., Razmkhah, M., Hosseini, A., Bagheri, M., Safaei, A., Talei, A. R., et al. (2016). Increased B regulatory phenotype in non-metastatic lymph nodes of node-positive breast cancer patients. *Scand. J. Immunol.* 83, 195–202. doi: 10.1111/sji.12407
- Mellios, T., Ko, H. L., and Beuth, J. (2010). Impact of adjuvant chemo- and radiotherapy on the cellular immune system of breast cancer patients. *In Vivo* 24, 227–230.
- Meylan, M., Petitprez, F., Lacroix, L., Di Tommaso, L., Roncalli, M., Bougoüin, A., et al. (2020). Early hepatic lesions display immature tertiary lymphoid structures and show elevated expression of immune inhibitory and immunosuppressive molecules. *Clin. Cancer Res.* 26, 4381–4389. doi: 10.1158/ 1078-0432.CCR-19-2929
- Miles, B., and Connick, E. (2018). Control of the germinal center by follicular regulatory T cells during infection. *Front. Immunol.* 9:2704. doi: 10.3389/ fimmu.2018.02704
- Nakagawa, R., Toboso-Navasa, A., Schips, M., Young, G., Bhaw-Rosun, L., Llorian-Sopena, M., et al. (2021). Permissive selection followed by affinity-based proliferation of GC light zone B cells dictates cell fate and ensures clonal breadth. *Proc. Natl. Acad. Sci. U.S.A.* 118:e2016425118. doi: 10.1073/pnas. 2016425118
- Novinger, L. J., Ashikaga, T., and Krag, D. N. (2015). Identification of tumorbinding ScFv derived from clonally related B cells in tumor and lymph node of a patient with breast cancer. *Cancer Immunol. Immunother.* 64, 29–39. doi: 10.1007/s00262-014-1612-1
- Núñez, N. G., Tosello Boari, J., Ramos, R. N., Richer, W., Cagnard, N., Anderfuhren, C. D., et al. (2020). Tumor invasion in draining lymph nodes is associated with Treg accumulation in breast cancer patients. *Nat. Commun.* 11:3272. doi: 10.1038/s41467-020-17046-2
- Nutt, S. L., Hodgkin, P. D., Tarlinton, D. M., and Corcoran, L. M. (2015). The generation of antibody-secreting plasma cells. *Nat. Rev. Immunol.* 15, 160–171. doi: 10.1038/nri3795
- Nzula, S., Going, J. J., and Stott, D. I. (2003). Antigen-driven clonal proliferation, somatic hypermutation, and selection of B lymphocytes infiltrating human ductal breast carcinomas. *Cancer Res.* 63, 3275–3280.
- Paus, D., Phan, T. G., Chan, T. D., Gardam, S., Basten, A., and Brink, R. (2006). Antigen recognition strength regulates the choice between extrafollicular plasma cell and germinal center B cell differentiation. J. Exp. Med. 203, 1081– 1091. doi: 10.1084/jem.20060087
- Perou, C. M., Jeffrey, S. S., van de Rijn, M., Rees, C. A., Eisen, M. B., Ross, D. T., et al. (1999). Distinctive gene expression patterns in human mammary epithelial cells and breast cancers. *Proc. Natl. Acad. Sci. U.S.A.* 96, 9212–9217.
- Petitprez, F., de Reyniès, A., Keung, E. Z., Chen, T. W., Sun, C. M., Calderaro, J., et al. (2020). B cells are associated with survival and immunotherapy response in sarcoma. *Nature* 577, 556–560. doi: 10.1038/s41586-019-1906-8
- Pioli, C., Gatta, L., Ubaldi, V., and Doria, G. (2000). Inhibition of IgG1 and IgE production by stimulation of the B Cell CTLA-4 receptor. J. Immunol. 165, 5530–5536. doi: 10.4049/jimmunol.165.10.5530
- Posch, F., Silina, K., Leibl, S., Mündlein, A., Moch, H., Siebenhüner, A., et al. (2017). Maturation of tertiary lymphoid structures and recurrence of stage II and III colorectal cancer. *Oncoimmunology* 7:e1378844. doi: 10.1080/2162402X.2017. 1378844
- Reddy, S. M., Barcenas, C. H., Sinha, A. K., Hsu, L., Moulder, S. L., Tripathy, D., et al. (2018). Long-term survival outcomes of triple-receptor negative breast cancer survivors who are disease free at 5 years and relationship with low hormone receptor positivity. *Br. J. Cancer* 118, 17–23. doi: 10.1038/bjc.20 17.379
- Ruer-Laventie, J., Simoni, L., Schickel, J. N., Soley, A., Duval, M., Knapp, A. M., et al. (2015). Overexpression of Fkbp11, a feature of lupus B cells, leads to B cell

tolerance breakdown and initiates plasma cell differentiation. *Immun. Inflamm. Dis.* 3, 265–279. doi: 10.1002/iid3.65

- Salgado, R., Denkert, C., Demaria, S., Sirtaine, N., Klauschen, F., Pruneri, G., et al. (2015). The evaluation of tumor-infiltrating lymphocytes (TILs) in breast cancer: recommendations by an international TILs working group 2014. Ann. Oncol. 26, 259–271. doi: 10.1093/annonc/mdu450
- Sautès-Fridman, C., Petitprez, F., Calderaro, J., and Fridman, W. H. (2019). Tertiary lymphoid structures in the era of cancer immunotherapy. *Nat. Rev. Cancer* 19, 307–325. doi: 10.1038/s41568-019-0144-6
- Schmid, P., Adams, S., Rugo, H. S., Schneeweiss, A., Barrios, C. H., Iwata, H., et al. (2018). Atezolizumab and Nab-paclitaxel in advanced triple-negative breast cancer. N. Engl. J. Med. 379, 2108–2121. doi: 10.1056/NEJMoa1809615
- Seow, D. Y. B., Yeong, J. P. S., Lim, J. X., Chia, N., Lim, J. C. T., Ong, C. C. H., et al. (2020). Tertiary lymphoid structures and associated plasma cells play an important role in the biology of triple-negative breast cancers. *Breast Cancer Res. Treat.* 180, 369–377. doi: 10.1007/s10549-020-05548-y
- Shen, M., Wang, J., and Ren, X. (2018). New insights into tumor-infiltrating B lymphocytes in breast cancer: clinical impacts and regulatory mechanisms. *Front. Immunol.* 9:470. doi: 10.3389/fimmu.2018.00470
- Shen, S., Sun, Q., Liang, Z., Cui, X., Ren, X., Chen, H., et al. (2014). A prognostic model of triple-negative breast cancer based on MiR-27b-3p and node status. *PLoS One* 9:e100664. doi: 10.1371/journal.pone.0100664
- Shi, J., Hou, S., Fang, Q., Liu, X., Liu, X., and Qi, H. (2018). PD-1 controls follicular T helper cell positioning and function. *Immunity* 49, 264–274.e4. doi: 10.1016/j.immuni.2018.06.012
- Sidwell, T., and Kallies, A. (2016). Bach2 is required for B cell and T cell memory differentiation. *Nat. Immunol.* 17, 744–745. doi: 10.1038/ni.3493
- Siliòa, K., Soltermann, A., Attar, F. M., Casanova, R., Uckeley, Z. M., Thut, H., et al. (2018). Germinal centers determine the prognostic relevance of tertiary lymphoid structures and are impaired by corticosteroids in lung squamous cell carcinoma. *Cancer Res.* 78, 1308–1320. doi: 10.1158/0008-5472.CAN-17-1987
- Solinas, C., Garaud, S., de Silva, P., Boisson, A., van den Eynden, G., de Wind, A., et al. (2017). Immune checkpoint molecules on tumor-infiltrating lymphocytes and their association with tertiary lymphoid structures in human breast cancer. *Front. Immunol.* 8:1412. doi: 10.3389/fimmu.2017.01412
- Song, H., Liu, A., Liu, G., Wu, F., and Li, Z. (2019). T follicular regulatory cells suppress Tfh-mediated B cell help and synergistically increase IL-10-producing B cells in breast carcinoma. *Immunol. Res.* 67, 416–423. doi: 10.1007/s12026-019-09090-y
- Song, I. H., Heo, S. H., Bang, W. S., Park, H. S., Park, I. A., Kim, Y. A., et al. (2017). Predictive value of tertiary lymphoid structures assessed by high endothelial venule counts in the neoadjuvant setting of triple-negative breast cancer. *Cancer Res. Treat.* 49, 399–407. doi: 10.4143/crt.2016.215
- Stacker, S. A., Williams, S. P., Karnezis, T., Shayan, R., Fox, S. B., and Achen, M. G. (2014). Lymphangiogenesis and lymphatic vessel remodelling in cancer. *Nat. Rev. Cancer* 14, 159–172. doi: 10.1038/nrc3677
- Sun, X., Zhang, T., Li, M., Yin, L., and Xue, J. (2019). Immunosuppressive B cells expressing PD-1/PD-L1 in solid tumors: a mini review. QJM doi: 10.1093/ qjmed/hcz162 Online ahead of print.
- Tadmor, T., Zhang, Y., Cho, H. M., Podack, E. R., and Rosenblatt, J. D. (2011). The absence of B lymphocytes reduces the number and function of T-regulatory cells and enhances the anti-tumor response in a murine tumor model. *Cancer Immunol. Immunother.* 60, 609–619. doi: 10.1007/s00262-011-0972-z
- Tao, H., Lu, L., Egenti, M., Chang, A. E., and Li, Q. (2013). "Effector B cells in cancer adoptive immunotherapy," in *Proceedings of the 104th Annual Meeting* of the American Association for Cancer Research, AACR 2013, Vol. 73(8 Suppl. 1), Washington, DC. doi: 10.1158/1538-7445.AM2013-487
- Tao, H., Lu, L., Xia, Y., Dai, F., Wang, Y., Bao, Y., et al. (2015). Antitumor effector B cells directly kill tumor cells via the Fas/FasL pathway and are regulated by IL-10. *Eur. J. Immunol.* 45, 999–1009. doi: 10.1002/ejj.201444625
- Toboso-Navasa, A., Gunawan, A., Morlino, G., Nakagawa, R., Taddei, A., Damry, D., et al. (2020). Restriction of memory B cell differentiation at the germinal center B cell positive selection stage. J. Exp. Med. 217:e20191933. doi: 10.1084/ jem.20191933
- van Roozendaal, L. M., Smit, L. H. M., Duijsens, G. H. N. M., de Vries, B., Siesling, S., Lobbes, M. B. I., et al. (2016). Risk of regional recurrence in triple-negative

breast cancer patients: a dutch cohort study.". Breast Cancer Res. Treat. 156, 465-472. doi: 10.1007/s10549-016-3757-4

- Verma, R., Foster, R. E., Horgan, K., Mounsey, K., Nixon, H., Smalle, N., et al. (2016). Lymphocyte Depletion and Repopulation after Chemotherapy for Primary Breast Cancer.". *Breast Cancer Res.* 18:10. doi: 10.1186/s13058-015-0669-x
- Velu, V., Titanji, K., Zhu, B., Husain, S., Pladevega, A., Lai, L., et al. (2009). Enhancing SIV-specific immunity *in vivo* by PD-1 blockade. *Nature* 458, 206–210. doi: 10.1038/nature07662
- Victora, G. D., and Nussenzweig, M. C. (2012). Germinal centers. Annu. Rev. Immunol. 30, 429–457. doi: 10.1146/annurev-immunol-020711-075032
- Victora, G. D., Schwickert, T. A., Fooksman, D. R., Kamphorst, A. O., Meyer-Hermann, M., Dustin, M. L., et al. (2010). Germinal center dynamics revealed by multiphoton microscopy with a photoactivatable fluorescent reporter. *Cell* 143, 592–605. doi: 10.1016/j.cell.2010.10.032
- Willard-Mack, C. L. (2006). Normal structure, function, and histology of lymph nodes. *Toxicol. Pathol.* 34, 409–424. doi: 10.1080/0192623060086 7727
- Yao, Y., Chu, Y., Xu, B., Hu, Q., and Song, Q. (2019). Risk factors for distant metastasis of patients with primary triple-negative breast cancer. *Biosci. Rep.* 39:BSR20190288. doi: 10.1042/BSR20190288

- Yin, L., Duan, J. J., Bian, X. W., and Yu, S. C. (2020). Triple-negative breast cancer molecular subtyping and treatment progress.". *Breast Cancer Res.* 22:61. doi: 10.1186/s13058-020-01296-5
- Zheng, F., Pan, Q., Egenti, M., Chang, A. E., and Li, Q. (2012). "Blockade of IL-10 production in effector B cells significantly increases their therapeutic efficacy in cancer adoptive immunotherapy," in *Proceeings of the 103rd Annual Meeting of the American Association for Cancer Research, AACR* 2012, Vol. 72(8 Suppl. 1), Chicago, IL. doi: 10.1158/1538-7445.AM2012-4840

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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