



# Thallium - poisoner's poison: An overview and review of current knowledge on the toxicological effects and mechanisms

Junko Fujihara<sup>a,\*</sup>, Naoki Nishimoto<sup>b</sup>

<sup>a</sup> Department of Legal Medicine, Shimane University Faculty of Medicine, 89-1 Enya, Izumo, Shimane 693-8501, Japan

<sup>b</sup> Shimane Institute for Industrial Technology, 1 Hokuryo, Matsue, Shimane 690-0816, Japan

## ARTICLE INFO

### Keywords:

Thallium  
Concentrations in biological samples  
Toxicological effects  
Toxicological mechanism  
Toxicological symptoms  
Antidote

## ABSTRACT

Thallium (Tl) is one of the most toxic metals and its historic use in homicides has led it to be known as “the poisoner’s poison.” This review summarizes the methods for identifying Tl and determining its concentrations in biological samples in recently reported poisoning cases, as well as the toxicokinetics, toxicological effects, toxicity mechanisms, and detoxication methods of Tl. Recent findings regarding Tl neurotoxicological pathways and toxicological effects of Tl during pregnancy are also presented. Confirmation of elevated Tl concentrations in blood, urine, or hair is indispensable for diagnosing Tl poisoning. The kidneys show the highest Tl concentration within 24 h after ingestion, while the brain shows the highest concentration thereafter. Tl has a very slow excretion rate due to its large distribution volume. Following acute exposure, gastrointestinal symptoms are observed at an early stage, and neurological dysfunction is observed later: Tl causes the most severe damage in the central nervous system. Alopecia and Mees’ lines in the nails are observed within 1 month after Tl poisoning. The toxicological mechanism of Tl is considered to be interference of vital potassium-dependent processes with  $Tl^+$  because its ionic radius is similar to that of  $K^+$ , as well as inhibition of enzyme reactions by the binding of Tl to -SH groups, which disturbs vital metabolic processes. Tl toxicity is also related to reactive oxygen species generation and mitochondrial dysfunction. Prussian blue is the most effective antidote, and metallothionein alone or in combination with Prussian blue was recently reported to have cytoprotective effects after Tl exposure. Because Tl poisoning cases are still reported, early determination of Tl in biological samples and treatment with an antidote are essential.

## 1. Introduction

Thallium (Tl) is a nonessential element that is homologous for aluminum, gallium, and indium in group IIIA of the periodic table. Tl, which discovered in 1861, is one of the most toxic metals (Goyer and Clarkson, 2001), with higher toxicity than Hg, Cd, Pb, Cu, and Zn (Cvjetko et al., 2010). Tl is listed by the US Environmental Protection agency as one of the major pollutants, along with Hg, Cd, and Pb (ATSDR, 2015). The lethal Tl dose is 10–15 mg/kg in humans and death can even occur at a lower dose of 8 mg/kg (Al Hammouri et al., 2011; Riyaz et al., 2013). Compared with adults, children are more sensitive to Tl exposure: one-tenth of the adult lethal dose can cause death in children (Duan et al., 2020).

Tl exists in two oxidation states:  $Tl^+$  and  $Tl^{3+}$ . Both forms are toxic to living organisms (Rickwood et al., 2015), although the toxicity of  $Tl^{3+}$  is about 50,000 times higher than that of  $Tl^+$  on the basis of the free ion

concentration of each Tl redox state (Ralph and Twiss, 2002). However, the bioavailable concentration of  $Tl^{3+}$  is much lower than that of  $Tl^+$ . In contrast to other metals ( $Al^{3+}$ ,  $Ga^{3+}$ , and  $In^{3+}$ ) in group IIIA,  $Tl^+$  is more stable than  $Tl^{3+}$  (Rodríguez-Mercado and Altamirano-Lozano, 2013). The properties of  $Tl^+$  and potassium ion ( $K^+$ ) are similar due to their comparable ionic radii, and  $Tl^+$  generally forms the most stable Tl salts (John Peter and Viraraghavan, 2005).  $Tl^{3+}$  is similar to aluminum, which has strong oxidizing properties and rarely exists in nature (Zhuang and Song, 2021). Tl salts with high solubility are uncolored, inodorous, and tasteless compounds. At low doses, symptoms are slow to manifest and can easily be misdiagnosed as other ailments. The English writer Agatha Christie made Tl poisoning a central part of her 1961 novel *The Pale Horse*. Due to their properties, high toxicity, and high recognition as a poison, Tl-based salts have been used as “the poisoner’s poison” to the present (Lennartson, 2015). This review summarizes the analytical methods for Tl determination in biological samples, Tl

\* Corresponding author.

E-mail address: [fujihar@med.shimane-u.ac.jp](mailto:fujihar@med.shimane-u.ac.jp) (J. Fujihara).

<https://doi.org/10.1016/j.crttox.2024.100157>

Received 30 October 2023; Received in revised form 13 February 2024; Accepted 15 February 2024

Available online 18 February 2024

2666-027X/© 2024 The Author(s). Published by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

concentrations in biological samples from recent criminal cases of Tl poisoning, and the toxicokinetics, toxicological effects, toxicological mechanisms, and recent detoxication methods of Tl. The neurotoxicity of Tl exposure and the toxicological effects of Tl during pregnancy have not been fully elucidated, although current studies are investigating these topics. This review also covers the findings of a recent study on Tl neuro-toxicological pathways and the toxicological effects of Tl during pregnancy.

## 2. Tl in the environment

Tl is widely distributed in the environment at very low concentrations (Fig. 1a). It is naturally found as sulfide minerals (e.g., lorandite and crookesite). The average Tl concentrations in the Earth's crust are between 0.3 and 0.5 mg/kg and it is present mainly in the sulfide ores of Zn, Cu, Pb, Cd, and Fe (Galván-Arzate and Santamaría, 1998; John Peter and Viraraghavan, 2005). Tl concentrations in uncontaminated sediment range from 0.22 to 0.55 mg/kg (Liu et al., 2019), while those in soil range from 0.1 to 1 mg/kg (Rodríguez-Mercado and Altamirano-Lozano, 2013). Tl is released into the environment as  $Tl_2O$  by air emissions from coal burning and by Zn, Cu, Pb, and Cd smelters. Tl concentrations are less than  $1 \text{ ng/m}^3$  in unpolluted air, approximately 10–20 ng/L in sea water and less than 1 µg/L in fresh water. Elevated Tl concentrations (1–88 µg/L) have been reported in river waters near metal mining areas (Zitko et al., 1975). The maximum permissible Tl contaminant level in drinking water is suggested to be 2 µg/L (USEPA, 2022). Tl concentrations in food are less than 100 µg/kg dry weight, and higher concentrations have been found in seafood (IPCS, 1996). A recent study reported that  $Tl^+$ ,  $TlSO_4^{4-}$ ,  $TlCl$ , and  $TlNO_3$  are the main forms of Tl in

ground water (Liu et al., 2021). In most natural environments,  $Tl^+$  ions are the predominant species because they are thermodynamically stable (Lan and Lin, 2005).

## 3. Production and use of Tl

Tl is obtained from Fe, Cd, and Zn refining as a by-product (Galván-Arzate and Santamaría, 1998) and is separated by electrolysis (Blain and Kazantzis, 2015). The most common commercial forms of Tl are Tl acetate ( $CH_3COOTl$ ), Tl carbonate ( $Tl_2CO_3$ ), and Tl sulfate ( $Tl_2SO_4$ ), which have oxidation states of I, while compounds with an oxidation state of III include Tl trichloride ( $TlCl_3$ ) and Tl sesquioxide ( $Tl_2O_3$ ) (Rodríguez-Mercado and Altamirano-Lozano, 2013). With the exception of Tl sesquioxide, Tl compounds are stable in aqueous solutions and soluble in water (Rodríguez-Mercado and Altamirano-Lozano, 2013).

The uses of Tl are shown in Fig. 1b.  $Tl_2SO_4$  and  $CH_3COOTl$  have been used as rodenticides and insecticides. They were banned in 1965 in the United States due to concerns about environmental pollution and high toxicity (Riyaz et al., 2013). However, they are still used in many developing countries (Sánchez-Chapul et al., 2023).  $^{201}Tl$ -thallous chloride has been used in myocardial imaging to evaluate ischemic heart disease; this radioactive Tl emits X-rays and gamma-rays and is intravenously administered (Genchi et al., 2021).  $Tl^+$  shows good nuclear magnetic resonance properties and can be used as a probe to emulate the biological functions of  $K^+$  and  $Na^+$  (Blain and Kazantzis, 2015). Tl is used to produce imitation jewelry and artificial diamonds, to create the green color in fireworks, and as a catalyst of organic compound synthesis (Rodríguez-Mercado and Altamirano-Lozano, 2013). Contemporary demand for Tl is increasing in advanced industrial technologies. Tl is

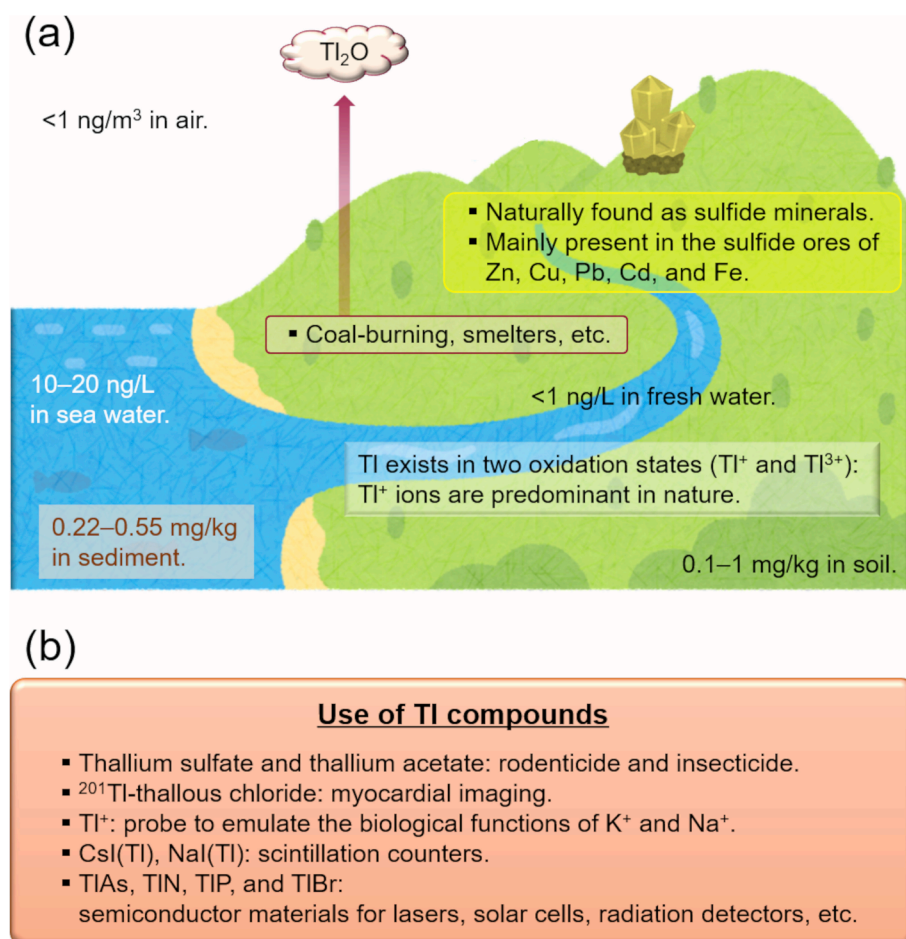


Fig. 1. Thallium in the environment and its usage.

utilized in camera lenses, scintillation counters (CsI(Tl)), and low-temperature thermometers in alloys with mercury (Genchi et al., 2021). In addition, Tl-containing semiconductor materials have been expected to apply for various electronic devices. The theoretical band gap energies of TlAs, TlN, and TlP that have zinc blende structure are  $\sim 0$  eV (Ferreira da Silva et al., 2005; Gulebaglan et al., 2013). Accordingly, Tl-containing III–V compound semiconductors are attractive materials for optoelectronic devices operating at infrared region. For instance, TlGaAs could be used to fabricate higher-quality and more efficient devices (e.g., laser diodes, solar cells) (Nishimoto et al., 2003; Zayan and Vandervelde, 2019; Nikoo et al., 2022). TlBr with cesium chloride structure has high density (7.56 g/cm<sup>3</sup>) and a comparatively wide band gap energy (2.68 eV) (Dönmez et al., 2010). Due to these properties, the gamma-ray absorption efficiency is high. Hence, TlBr is suitable material for radiation detector (Churilov et al., 2009; Park and Lee, 2023).

#### 4. Methods for determining Tl in biological samples

Atomic absorption spectrometry (AAS) is most frequently used method for determining Tl in biological samples and includes flame AAS (Richelmi et al., 1980; Chandler et al., 1990), flameless AAS, graphite furnace AAS (Tanaka et al., 1978; Ríos et al., 1989), and electrothermal AAS (Yang and Smeyers-Verbeke, 1991). The detection range of flame AAS is in the order of ppb to ppm, whereas that of flameless AAS is in ppb. Atomic fluorescence spectrometry is rarely used for Tl analysis in biological samples (Mori et al., 1994). Inductively coupled plasma-mass spectrometry (ICP-MS) has recently become the most commonly used method for Tl determination in biological specimens due to its high sensitivity (dynamic range: ppq to ppb) (Das et al., 2006; Li et al., 2015; Di Candia et al., 2020; Pragst and Hartwig, 2022). In addition, laser ablation-inductively coupled plasma-sector field-mass spectrometry (LA-ICP-SF-MS) was used to determine the Tl concentration in the bone of someone who was killed by Tl poisoning 38 years ago (Hann et al., 2005). Inductively coupled plasma-optical emission spectrometry (ICP-OES) is also used for detecting Tl in biological samples, with a dynamic range from ppb to ppm (Lech and Lachowicz, 2009; Gupta et al., 2023). Microwave plasma-atomic emission spectrometry, which is a recently introduced technique for multi-element analysis with a low running cost (Fujihara and Nishimoto, 2020; Fujihara and Nishimoto, 2023), is also expected to be applied to Tl analysis in biological samples.

To determine Tl in biological samples (body fluids and tissues) by using the above methods, pretreatment is required. In the case of urine and serum analysis, pretreatment can be achieved by HNO<sub>3</sub> dilution. In Tl analysis using GF-AAS, Duan et al. (2020) performed a 10-time dilution of urine with a diluent containing 0.03 % HNO<sub>3</sub>, 0.02 % Triton, and 0.66 g/L palladium chloride. In other studies, urine samples were diluted 1/10 (v/v) in a solution containing 1 % HNO<sub>3</sub>, and blood and serum were diluted by an alkaline solution containing 0.02 % Triton X-100 and 3 % ammonia solution before ICP-MS analysis (Heitland and Köster, 2021; Pragst and Hartwig, 2022). For tissue analysis, acid digestion by concentrated HNO<sub>3</sub> (65 %) with or without H<sub>2</sub>O<sub>2</sub> (30 %) is performed by heating, using a microwave, heat block, and so on (Tanaka et al., 1978; Hann et al., 2005; Das et al., 2006; Di Candia et al., 2020). For example, Lech and Lachowicz (2009) performed acid digestion with a microwave by adding 4 mL of HNO<sub>3</sub> and 1 mL of H<sub>2</sub>O<sub>2</sub> to blood (1 mL), urine (2–8 mL), and tissue (1–2 g) and then heating at 250 W (1 min), 0 W (2 min), and 250, 400, and 600 W (5 min each). Following digestion, the samples were diluted to 10 or 25 mL. Hair samples should be washed with detergent such as Triton X-100 before acid digestion to remove exogenous contaminants.

In contrast to the above destructive methods, nondestructive methods are also available to determine Tl levels in biological specimens. Goldman et al. (1966) performed Tl determination using X-ray emission spectroscopy in urine and feces at ppm levels. Neutron activation analysis (NAA) is another nondestructive analytical technique for

trace element determination with an ultralow detection limit and simple preparation of samples (Das et al., 2023). Minoia et al. (1990) used NAA to perform Tl determination in urine and blood from healthy Italians. Similarly, Henke (1991) performed Tl analysis in bovine liver and bone with NAA.

#### 5. Tl concentrations in biological samples

Equipment for Tl analysis is present in a minority of reference laboratories and evidence for Tl exposure is not rapidly available to clinicians (Ghannoum et al., 2012). However, confirmation of an elevated Tl concentration in blood, urine, or hair is necessary for the definitive diagnosis of Tl poisoning. Tl concentrations in biological specimens in Tl poisoning cases are summarized in Table 1.

A 24-h urine sample is considered necessary to identify Tl poisoning. Tl cannot be detected in most persons, and levels up to 20 µg/specimen are considered to be normal for occupational and environmental exposure (Rusyniak et al., 2010). The normal urine Tl concentration is less than 5 µg/L. The typical clinical symptoms are observed when the urinary Tl concentration exceeds 500 µg/L (Huang et al., 2014), and a concentration greater than 200 µg/L can be used as a basis for diagnosing acute Tl poisoning (Moore et al., 1993). Renal excretion reflects the total Tl load (Moore et al., 1993). However, urinary Tl concentrations do not correlate with blood concentrations or symptoms (Ghannoum et al., 2012). Serum Tl levels are much lower than in other tissues and poorly representative of the toxicokinetic profile of target organs (Richelmi et al., 1980). The normal total blood Tl concentration is less than 2 µg/L and levels greater than 100 µg/L are toxic (Moore et al., 1993).

Hair is an important specimen for understanding the historical internal exposure to heavy metals: the keratin in hair, due to its high cysteine content, has high affinity for Tl, similar to other heavy metals. Tl levels in hair less than 15 ng/g are considered normal (Mulkey and Oehme, 1993). Hirata et al. (1998) reported elevated Tl concentrations (0.02–0.58 ng/mg) in the hair of workers at a glass factory, based on ICP-MS results. Ash and He (2018) evaluated the Tl concentration in the hair of a “cold case” victim of Tl poisoning that occurred 24 years ago using LA-ICP-MS and revealed that the victim had been repeatedly exposed to Tl for 4 months.

#### 6. Toxicokinetics of Tl

##### 6.1. Absorption

The toxicokinetics of Tl follow a three-compartment model. The first compartment with rapid exchange comprises the intravascular system and highly perfused organs (within 4 h of exposure). The second compartment with slow exchange includes the central nervous system, which lasts 4–48 h (Riyaz et al., 2013). The third compartment comprises the large and small intestines, which are responsible for wide enteroenteric circulation and the long half-life of Tl (De Groot and van Heijst, 1988). The toxicokinetics of Tl are summarized in Fig. 2a.

Both Tl<sup>+</sup> and Tl<sup>3+</sup> can exist under biological conditions (Harris and Messori, 2002). Tl<sup>+</sup> salts, which are water-soluble, can enter the body through the K<sup>+</sup> uptake mechanism (Mulkey and Oehme, 1993; Galván-Arzate and Santamaría, 1998). Tl is absorbed through almost all routes: the skin, inhalation, and the gastrointestinal tract (Goyer and Clarkson, 2001; Rodríguez-Mercado and Altamirano-Lozano, 2013). Respiration is the main absorption route (in the form of fumes) in the industrial environment, and significant amounts of Tl can also be absorbed via the skin (Rodríguez-Mercado and Altamirano-Lozano, 2013). When ingested, Tl is rapidly and completely absorbed through the gastrointestinal tract (Achenbach et al., 1980; Rodríguez-Mercado and Altamirano-Lozano, 2013). The oral bioavailability of hydrophilic Tl salts reaches 90 %–100 % (De Groot and van Heijst, 1988; Mulkey and Oehme, 1993).

**Table 1**  
Summary of thallium poisoning cases and thallium concentrations in biological samples.

Survival time after Tl ingestion	Case	Time after ingestion	Tl concentrations at hospital	Postmortem Tl Concentrations	Detection method	Reference
Survivor	A 28-year-old woman ingested about 1 g of thallium sulphate and was discharged 28 days after admission	4 days	Urine: 3 mg/L Gastric content: 10.8 mg/L	–	AAS	<a href="#">Richelmi et al., 1980</a>
		7 days	Saliva: 9 mg/L			
	A 43-year-old man developed acute abdominal colic and diarrhea a few hours after drinking a beverage contaminated with thallium sulphate	21 months	Blood: 336.5 µg/L Urine: 252.3 µg/L	–	AAS	<a href="#">Liu and Liao, 2021</a>
	Five adult patients who attended a banquet ate a Tl-contaminated meal (33–49 years old)	9–12 days	Blood: 40–280 µg/L Urine: 250–7200 µg/L	–	AAS	<a href="#">Wang et al., 2021</a>
	A 40-year-old man ate a thallium nitrate-contaminated supper	3 days	Blood: 3764 ng/mL	–	ICP-MS	<a href="#">Huang et al., 2014</a>
	A 44-year-old man ate a meal contaminated with thallium sulphate	49 days	Blood: 175 µg/L	–	ICP-MS	<a href="#">Pragst and Hartwig, 2022</a>
	A 23-year-old woman drank tea contaminated with thallium sulphate	10 days	Blood: 223 µg/L Urine: 351 µg/L	–	Unknown	<a href="#">Yumoto et al., 2017</a>
	A 28-year-old woman lapsed into a coma due to severe thallium poisoning (source unknown) and was discharged 137 days after admission	35 days	Blood: 950 ng/mL Urine: 7600 ng/mL	–	Unknown	<a href="#">Lin et al., 2019</a>
	A 24-year-old woman ingested food deliberately mixed with Tl	22 days	Blood: 180 ng/µL Urine: 930 ng/µL	–	Unknown	
	A 31-year-old man ingested food deliberately mixed with Tl	23 days	Blood: 90.9 ng/µL Urine: 1825 ng/µL	–	Unknown	<a href="#">Zhao et al., 2008</a>
	A 37-year-old man drank water deliberately mixed with Tl	21 days	Blood: 288.4 ng/µL Urine: 2359 ng/µL	–	Unknown	
	Eleven members of two families (2–42 years old) ingested Tl	5–7 days	Blood: 323.5 µg/L Urine: 1959 µg/L	–	Unknown	<a href="#">Al Hammouri et al., 2011</a>
	Nine patients (3 adults and 6 children) (2–73 years old) from a family ingested Tl-contaminated rice	9–15 days	Serum: 506.3–985.2 µg/L Urine: 4345–9998 µg/L	–	Unknown	<a href="#">Zhang et al., 2014</a>
A 35-year-old woman exposed to rodenticide at her workplace at 13 weeks of pregnancy experienced a spontaneous abortion at 18 weeks	Unknown	Urine (24 h): 3400 µg/L	–	Unknown	<a href="#">Hoffman, 2000</a>	
30 days	A 22-year-old man ingested 50 mL of 2 % thallium sulphate and died 30 days later	30 days	–	Brain: 0.02 µg/g Liver: 0.015 µg/g Kidney: 0.01 µg/g	GF-AAS	<a href="#">Tanaka et al., 1978</a>
23 days	A 49-year-old woman ingested thallium sulphate mixed with table salt	About a week	Urine: 8.8 µg/mL	Hair: 10.20 µg/mL Blood: 0.019 µg/mL Urine: 3.15 µg/mL Brain: 2.03 µg/mL Liver: 0.98 µg/mL Kidney: 0.98 µg/mL Heart: 0.57 µg/mL Lung: 0.46 µg/mL Stomach: 0.38 µg/mL	ICP-MS	<a href="#">Li et al., 2015</a>
3 days	A 50-year-old man ingested thallium sulphate mixed with table salt	About 2 weeks	Urine: 4.3 µg/mL	Hair: 13.75 µg/mL Blood 0.15 µg/mL Urine: 3.60 µg/mL Brain: 2.58 µg/mL Liver: 5.08 µg/mL Kidney: 2.21 µg/mL Heart: 1.83 µg/mL Lung: 0.88 µg/mL Stomach: 4.02 µg/mL	ICP-MS	
2 days	Thallium sulfate poisoning of 8 members of a family (3 of whom died)	Father Unknown	Blood: 3.4 µg/mL Urine: 22.7 mg/L	Blood: 2.75 µg/mL Urine: 1.49 mg/L Hair: 10.11 ng/mg	ICP-MS	<a href="#">Di Candia et al., 2020</a>

(continued on next page)

Table 1 (continued)

Survival time after Tl ingestion	Case	Time after ingestion	Tl concentrations at hospital	Postmortem Tl Concentrations	Detection method	Reference
2 days	Mother	Unknown	Blood: 10 µg/mL Urine: 42.0 mg/L	Blood: 1.15 µg/mL Gastric content: 1.11 mg/L Hair: 10.38 ng/mg	ICP-MS	
14 days	Daughter	Unknown	Blood: 5.7 µg/mL Urine: 16.3 mg/L	Blood: 6.01 µg/mL Gastric content: 3.43 mg/L Hair: 5.72 ng/mg	ICP-MS	
–	A man poisoned with rodenticide by his daughter-in-law and buried for 29 years	Unknown	–	Bone: 1.07–2.63 µg/g	LA-ICP-MS	Hann et al., 2005
4 days	A 36-year-old man ingested an unknown amount of thallium sulphate from rodenticide and died after 4 days	1 days	Blood: >1000 µg/L Urine: >2000 µg/L	Blood: 5349 µg/L	Unknown	Riyaz et al., 2013

AAS, atomic absorption spectrometry

ICP-MS, inductively coupled plasma-mass spectrometry

GF-AAS, graphite furnace-atomic absorption spectrometry

LA-ICP-MS, laser ablation-inductively coupled plasma-mass spectrometry

## 6.2. Tissue distribution

Following its rapid absorption, Tl<sup>+</sup> is widely distributed to organs and tissues (Ríos et al., 1989; Mulkey and Oehme, 1993) through the systemic circulation after binding to serum transferrin (Harris and Messori, 2002). A large apparent volume of distribution of Tl (3–10 L/kg) has been reported (De Groot and van Heijst, 1988). The highest concentration of Tl is found in the kidney after acute poisoning (Goyer and Clarkson, 2001). Ríos et al. (1989) reported that the Tl concentration in the kidney was about three times higher than that in other tissues and that the whole brain had the lowest concentration following 24 h of intraperitoneal administration of Tl<sub>2</sub>SO<sub>4</sub> in rats. They reported differential Tl distribution within the brain: the hypothalamus showed the highest concentration, while the cortex showed the lowest concentration. Achenbach et al. (1980) reported the organ-specific uptake of Tl by the heart, liver, kidney, and stomach during the first 2–3 h, and the Tl concentration in the brain was relatively low and constant during the first 12 h; all organs showed increased Tl concentrations after 24 h following oral administration to mice. Galván-Arzate et al. (2005) showed higher Tl concentrations in the brain following subchronic intraperitoneal administration (30 days) of a sublethal dose (0.8 and 1.6 mg/kg) of CH<sub>3</sub>COOTl to adult rats. In a human case (Table 1), the Tl concentration was highest in the brain (0.02 µg/g), followed by the liver (0.015 µg/g) and kidney (0.01 µg/g) in a man who died 30 days after the ingestion of 50 mL of 2 % Tl<sub>2</sub>SO<sub>4</sub> (Tanaka et al., 1978) (Table 1). Li et al. (2015) also reported that the brain showed the highest Tl concentration among tissues in a man and woman who died about a week after Tl<sub>2</sub>SO<sub>4</sub> ingestion (Table 1). Tl can easily cross the blood–brain barrier (BBB) (Galván-Arzate et al., 2000) and placenta (Hoffman, 2000). The placenta is another selective transport interface through which a number of environmental metals have been documented to exert toxicological effects. Heavy metals can pass through the placenta (Caserta et al., 2013) and Tl is one of those metals (Hoffman, 2003).

## 6.3. Metabolism

Metallothionein (MT) is a cysteine-rich (20 residues) metal-binding protein comprising 61–68 amino acids that is synthesized in hepatocytes following heavy metal exposure (Nordberg and Nordberg, 2000). Its possible biological role lies in metabolizing and neutralizing heavy metals (Klaassen and Liu, 1998; Sugiura and Yamashita, 2000) and scavenging free radicals. Kiliç and Kutlu (2010) suggested that exogenous MT acts as a scavenger of Tl and defends against reactive oxygen species (ROS) in rat liver intraperitoneally exposed to CH<sub>3</sub>COOTl.

## 6.4. Excretion

Large amounts of Tl are excreted in the urine approximately 24 h after exposure; thereafter, excretion from the urine slows and the feces may then become an important route of excretion (Goyer and Clarkson, 2001). Overall, 51 % of Tl is eliminated through the bile and feces, while 26 % is excreted in the urine (Mulkey and Oehme, 1993). In humans, Tl is also excreted in sweat, saliva, tears, and breast milk (Rodríguez-Mercado and Altamirano-Lozano, 2013). Richelmi et al. (1980) reported that salivary Tl levels were 5–15 times higher than those of urine during the first 2 weeks and that the time-course change in Tl concentrations was quite similar to that in urine in a 28-year-old woman who ingested Tl<sub>2</sub>SO<sub>4</sub>. Deposition in the nails and hair is also a key route of slow Tl elimination because the keratin in nails and hair has a high cysteine content and cysteine has a high affinity for Tl. Tl excreted slowly from the body explains its large distribution volume (a term used for quantifying the distribution of Tl between plasma and the rest of the body). Accordingly, Tl is retained in tissues for a long period and can be detected months or even years after exposure. Liu and Liao (2021) detected Tl in the blood and urine of a patient as for up to 21 months after the ingestion of Tl<sub>2</sub>SO<sub>4</sub> (Table 1). The slow excretion rate enables the accumulation of Tl even at low exposure levels (Cvjetko et al., 2010). The half-life of Tl depends on the dose and the nature of exposure (route, acute, or chronic). The elimination half-life of Tl in humans has been reported to be as long as 30 days (Goyer and Clarkson, 2001), and Tl remains in the body for days or even months, leading to prolonged neurological insufficiency or permanent damage.

## 7. Toxicological effects of Tl

### 7.1. Short-term effects following acute poisoning

Acute Tl poisoning, that is, single ingestion of large amount of Tl in a short time, will usually cause gastrointestinal symptoms, which emerge within the first few hours (John Peter and Viraraghavan, 2005; Cvjetko et al., 2010). When a lower amount of Tl is ingested, symptoms usually develop within 1–2 days. Severe abdominal pain is observed, with vomiting, nausea, and diarrhea. Tl specifically affects the nervous system (sensory and motor changes) (ATSDR, 2015). In humans, acute exposure to Tl leads to critical damage in the central, peripheral, and autonomic nervous systems (Galván-Arzate et al., 2005; John Peter and Viraraghavan, 2005). Rapid progressive and painful peripheral neuropathy development is observed within 2–3 days of exposure (Reed et al., 1963; Malbrain et al., 1997; Rusyniak et al., 2002). Symptoms

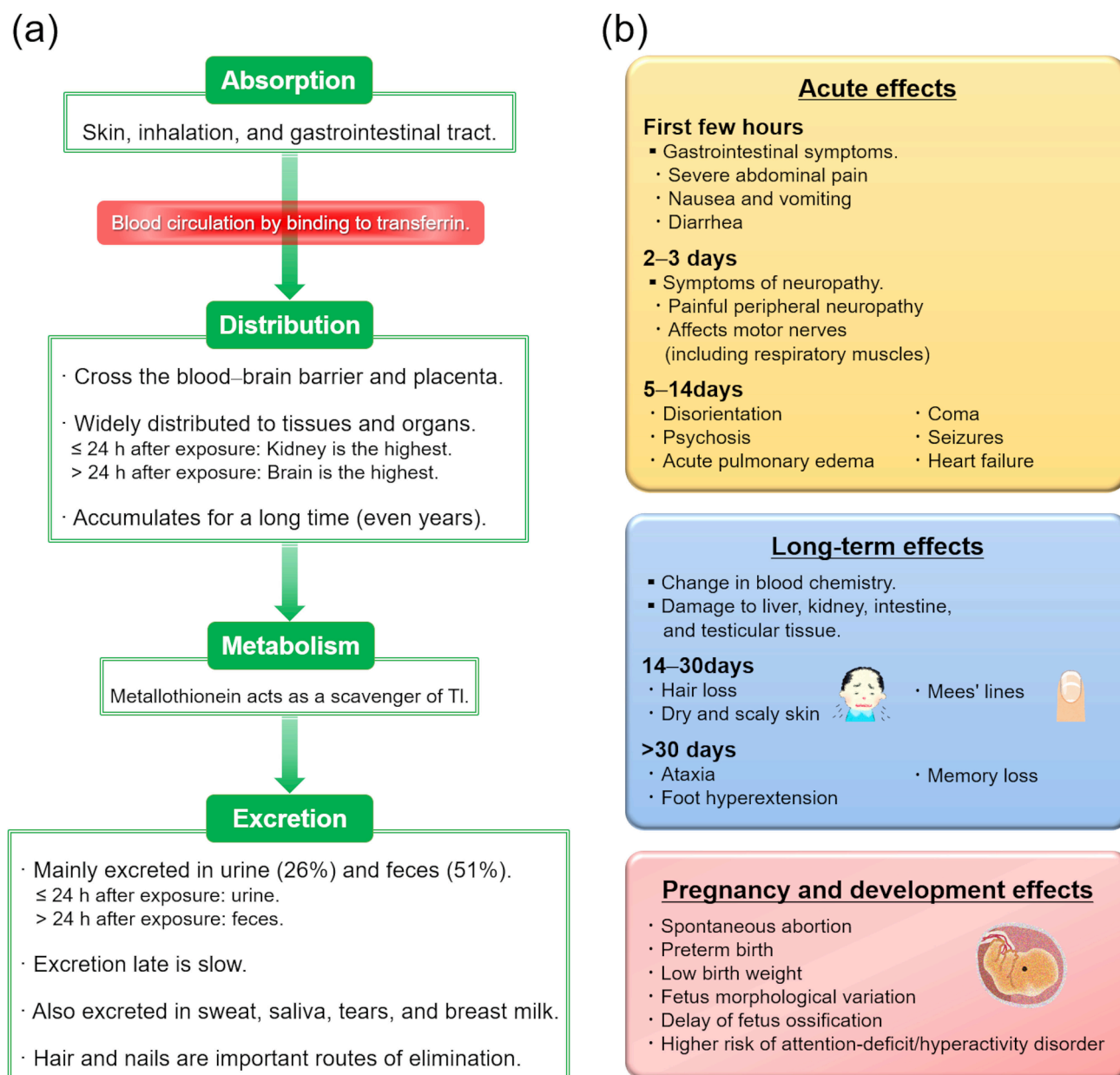


Fig. 2. Biological fate (a) and toxicological effects (b) of thallium.

begin in the feet and legs and extend to the hands (Rusyniak et al., 2010). Motor nerves are affected, including those innervating respiratory muscles (Hologgita et al., 1980; Rusyniak et al., 2002). Tl poisoning can sometimes be misdiagnosed as Guillain–Barré syndrome due to similar symptoms of neuropathy (Mulkey and Oehme, 1993). Moreover, disorientation, coma, psychosis, seizures, acute pulmonary edema, and heart failure are observed 5 h to 14 days after poisoning (Meggs et al., 1994; Hoffman et al., 1999; Rusyniak et al., 2002). Psychiatric disorders such as psychosis, depression, aggressiveness, hallucinations, cognitive disorders, and emotional disorders have been reported in patients with acute Tl intoxication (Cavanagh et al., 1974; Zavalij et al., 2021). Disorientation and generalized slowing on electroencephalography is observed with acute Tl poisoning (McMillan et al., 1997). This acute cardiovascular effect may result from competition of  $Tl^+$  with  $K^+$  for membrane transport systems, which leads to inhibited oxidative phosphorylation in mitochondria and disrupted protein synthesis (Goyer and Clarkson, 2001).

## 7.2. Long-term effects following acute poisoning

In the long term, Tl alters the blood chemistry, damages the liver, kidney, intestine, and tissue in the testes, and causes hair loss (Das et al., 2006). Elevated levels of aspartate transaminase and alanine transaminase have been reported (Zhao et al., 2008; Al Hammouri et al., 2011; Riyaz et al., 2013; Li et al., 2015; Lin et al., 2019; Wang et al., 2021), as well as elevated CK-MB (Wang et al., 2021) and lactate dehydrogenase (Al Hammouri et al., 2011). Alopecia is the best-known symptom of Tl poisoning (Galván-Arzate and Santamaría, 1998; Rusyniak et al., 2002). Tl poisoning specifically induces active hair loss >100 hairs/day in the 2–4 week period after poisoning and often presents as diffuse alopecia 2–3 weeks after Tl exposure (Cvjetko et al., 2010; Yu et al., 2018). Mees' lines (transverse white lines in the nails) are observed about 1 month after Tl poisoning due to impaired fingernail growth (Zhao et al., 2008). Additionally, dry and scaly skin is observed 2–4 weeks after the poisoning, and ataxia, foot hyperextension, and memory loss have been reported more than 30 days after exposure to Tl

(Meggs et al., 1994; Hoffman et al., 1999; Rusyniak et al., 2002). Chronic exposure to Tl results in alterations to the brain, spinal cord, and peripheral nerves (Galván-Arzate et al., 2005). The toxicological effects of Tl are summarized in Fig. 2b.

### 7.3. Toxicological effects of prenatal and postnatal exposure

As mentioned above, Tl can easily be transported via the placenta to the fetus. Rapid uptake and retention of Tl occurred in both the maternal and fetal organs following oral exposure to  $Tl_2SO_4$  in pregnant rats (Sabbioni et al., 1982). Prenatal exposure increases the risk of spontaneous abortion (Hoffman, 2000) as well as preterm birth and low birth weight (Qi et al., 2019; Wu et al., 2019; Zhou et al., 2021; Chen et al., 2022). Alopecia has been observed in children exposed to Tl during the fetal stage, although some children were born grossly normal or with limited poisoning manifestations even though their mothers exhibited symptoms of severe Tl poisoning (Hoffman, 2000). Álvarez-Barrera et al. (2019) demonstrated that intraperitoneal injection of  $CH_3COOTl$  to pregnant mice induced morphological variations in the fetus and a delay in fetal ossification. Moreover, recent studies have reported that high levels of Tl exposure during pregnancy are related to a higher risk of attention-deficit/hyperactivity disorder and lower full-scale intelligence (Tong et al., 2020).

Infants are also at risk of Tl exposure via breastfeeding. Previous quantitative evidence has confirmed that Tl is eliminated in breast milk (Hoffman, 2000). Johnston et al. (1996) reported  $^{201}Tl$  concentrations in the breastmilk of a female brain-tumor patient administered  $^{201}Tl$  for a brain scan at 2–500 h after administration. To our knowledge, no studies

have investigated the effects of Tl on infants exposed through breast milk. The toxicological effects of Tl during pregnancy are summarized in Fig. 2b.

### 8. Toxicological mechanism

The toxicological mechanisms of Tl are shown in Fig. 3. One proposed mechanism of Tl toxicity is the similarity of the ionic radius of  $Tl^+$  to that of  $K^+$  (Rusznayk et al., 1968). In addition, they are both univalent ions (John Peter and Viraraghavan, 2005). In the reduced state, the ionic radius for  $Tl^+$  is 1.76 Å while that of  $K^+$  is 1.60 Å (Rader et al., 2019; Rinklebe et al., 2020).  $K^+$  can easily be replaced with  $Tl^+$  and  $Tl^+$  can mimic the biological behavior of  $K^+$  (Diwan and Lehrer, 1977) and interfere with vital potassium-dependent processes. A low level of serum potassium (3.3 mEq/L) was reported in a Tl poisoning case (Tanaka et al., 1978).  $Tl^+$  may replace  $K^+$  in  $Na^+/K^+$  ATPase and thereby inhibit  $Na^+/K^+$  ATPase activity (John Peter and Viraraghavan, 2005; Cvjetko et al., 2010). In the rabbit kidney, the affinity of  $Tl^+$  for activating  $Na^+/K^+$  ATPase has been reported to be 10 times greater than that of  $K^+$  (Britten and Blank, 1968). In addition to ATPase,  $Tl^+$  replaces physiological  $K^+$  in monovalent cation-activated enzymes such as pyruvate kinase and aldehyde dehydrogenase (John Peter and Viraraghavan, 2005).

Another mechanism of Tl toxicity is its ability to react with thiol (-SH) groups of proteins and mitochondrial membranes, similar to other heavy metals (John Peter and Viraraghavan, 2005; Cvjetko et al., 2010). Tl has a high affinity for S ligands because it has empty d-orbitals in its electronic configuration (Cvjetko et al., 2010). Tl poisoning may be

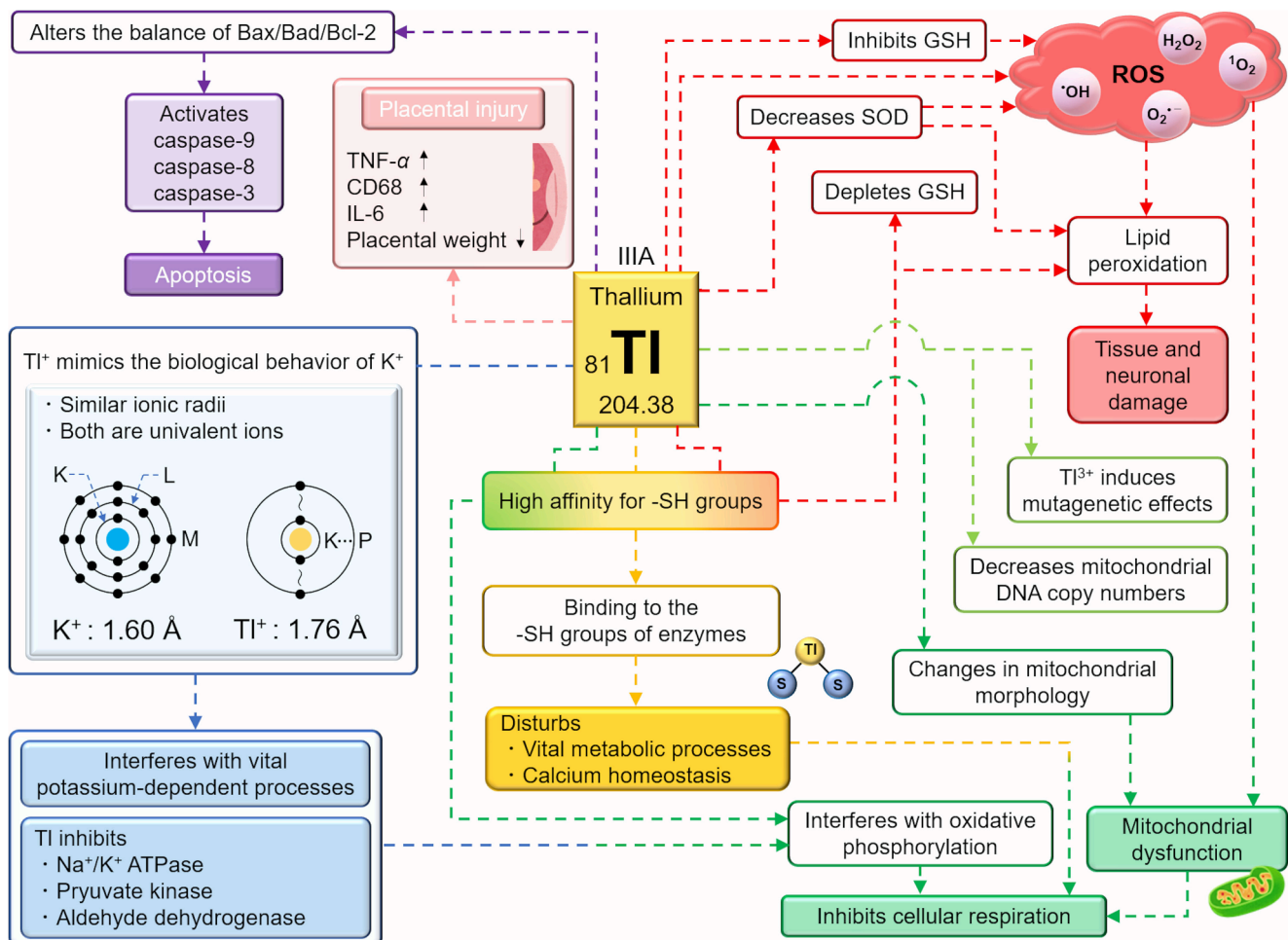


Fig. 3. Schematic of the toxicity mechanism of thallium. GSH, glutathione; ROS, reactive oxygen species; SOD, superoxide dismutase.

induced by inhibition of a variety of enzyme reactions as a result of -SH group binding, which disturbs vital metabolic processes (Ramsden, 2002). Tl may exert toxicity by inhibiting cellular respiration, interacting with riboflavin and riboflavin-based cofactors, and disrupting calcium homeostasis (Mulkey and Oehme, 1993). In addition to competing with  $K^+$ , Tl binds to -SH groups in mitochondria and interferes with oxidative phosphorylation. The affinity of Tl for -SH groups may also lead to lipid peroxidation and intracellular glutathione depletion (Goyer and Clarkson, 2001) and disruption of calcium homeostasis (Kiliç and Kutlu, 2010). Intra-subunit disulfide bonds of Cu-Zn superoxide dismutase (SOD), which remove superoxide anions from living organisms, may also be a target of Tl. In the rat, Cu-Zn SOD activity is significantly decreased by Tl administration (Galván-Arzate et al., 2005). In addition, hair loss, the main symptom of Tl poisoning, is caused by its binding to the -SH groups of hair keratins and its disruption of hair shaft formation (Kanwar and Narang, 2013). Moreover, Tl toxicity is related to the effect of Tl on glutathione (GSH) activity: GSH inhibits heavy metal toxicity by binding heavy metals through -SH groups (Genchi et al., 2021). Oxidative stress may be increased by the inhibition of enzymes containing cysteine residues in the active site as a result of GSH modification (Mulkey and Oehme, 1993).

Tl toxicity is also related to the generation of ROS, which cause tissue damage and dysfunction (Villaverde et al., 2004; Galván-Arzate et al., 2005; Hanzel et al., 2005; Eskandari et al., 2015; Kiliç and Kutlu, 2010; Anaya-Ramos et al., 2021). Eskandari et al. (2015) demonstrated that  $Tl^+$  increases ROS production by impairing the electron transfer chain in isolated rat liver mitochondria, activating the cell death signaling pathway. Lipid peroxidation and hydrogen peroxide ( $H_2O_2$ ) have been shown to be suitable markers of ROS involvement in Tl toxicity (Galván-Arzate et al., 2005; Hanzel and Verstraeten, 2006). Lipid peroxidation by ROS formation due to Tl exposure induces tissue damage and organ dysfunction in the brain and liver tissue (Galván-Arzate et al., 2000; Maya-López et al., 2018). Furthermore, Tl increases neural lipid peroxidation, which damages neurons (Hasan and Ali, 1981). A recent study found that Tl induces ROS generation and mitochondrial dysfunction in primary hippocampal neurons from Wistar rat embryos (Lin et al., 2020). It has also been reported that the Tl-mediated disruption of mitochondrial function is related to Tl toxicity. Changes in the morphology of mitochondria, such as swelling, have been reported (Herman and Bensch, 1967; Spencer et al., 1973). Tl affects isolated mitochondrial function by opening transition pores and uncoupling the respiratory chain (Bragadin et al., 2003; Korotkov and Lapin, 2003).

Tl induces apoptosis (Bragadin et al., 2003). Tl also alters the balance of Bax/Bad/Bcl-2 proteins and activates caspase-9, caspase-8, and caspase-3 in mitochondria, leading to apoptotic death (Osorio-Rico et al., 2017). It has been suggested that the toxicity is due to DNA damage (Nishioka, 1975). However, the genetic effects of Tl have not been fully elucidated.  $Tl^+$  is known to be molecularly more stable than  $Tl^{3+}$ , which has strong oxidizing capacity (Harris and Messori, 2002). Sánchez-Chapul et al. (2023) demonstrated that the DNA double helix and its oxidization were not altered by  $Tl^+$ . In contrast, Nowicka et al. (2013) showed that  $Tl^+$  is oxidized to  $Tl^{3+}$  by experimental UV irradiation and the presence of oxygen and that  $Tl^{3+}$  oxidizes guanine residues in DNA to 8-oxoguanine, inducing DNA breaks and exerting mutagenic effects. They also demonstrated that Tl interacts with oligonucleotide gene sequences in human 8-oxoguanine DNA glycosylase (*hOGG1*), which is responsible for repairing DNA damage. Recently, Wu et al. (2019) reported decreased amounts of mitochondrial DNA copy numbers and shortened neonatal telomere length due to epigenetic changes in proteins of H3 and H4 histones in the urine of pregnant women exposed to environmental Tl. They also revealed that prenatal Tl exposure shortened neonatal telomere length (Wu et al., 2021).

Few studies have investigated the toxicological mechanisms of prenatal thallium exposure and whether the effect of Tl on fetal development is direct or indirect. Zhou et al. (2021) suggested that prenatal

exposure to Tl is negatively associated with birth weight and that this association may be mediated by decreased placental weight. Zhu et al. (2020) reported that prenatal Tl exposure in a Chinese population induced an inflammatory response in the placenta and found that serum Tl levels in pregnant women were positively associated with CD68, TNF- $\alpha$ , and IL-6. Based on the findings of the above studies, the effect of Tl on the fetus is considered to be indirect, via placental injury.

As previously mentioned, Tl causes the most severe damage in the central nervous system. ROS formation, disruption of  $K^+$ -regulated homeostasis, mitochondrial dysfunction, excitotoxicity, and apoptosis are all considered to be mechanisms of Tl neurotoxicity. In particular, the brain is sensitive to oxidative damage because it contains high levels of unsaturated lipids and it has a high rate of oxidative metabolism (Chevalier et al., 1994; Goering et al., 2002). The inhibition of  $Na^+/K^+$  ATPase activity may disturb neurotransmission, while that of  $Na^+/K^+$  ATPase and pyruvate kinase activities induces excitotoxicity due to ATP depletion (Maya-López et al., 2018). Maya-López et al. (2018) suggested that energy depletion (mitochondrial dysfunction), inhibition of  $Na^+/K^+$  ATPase activity, and lipid peroxidation (oxidative damage) might account for the toxic pattern elicited by  $Tl^+$  in the nerve terminals of rat brains. Aldehyde dehydrogenase catalyzes the production of retinoic acid from retinol (vitamin A), and endogenous retinoic acid is essential for embryonic development and adult physiological processes (Wang et al., 2023). Retinoic acid also plays an important role in the development of the BBB in humans and mice (Mizee et al., 2013), and its signaling is a critical process for neurodevelopment (Cho et al., 2021; Menegola et al., 2021). The inhibition of aldehyde dehydrogenase by  $Tl^+$  exposure may prevent the development of the BBB as well as neurodevelopment by decreasing retinoic acid levels. The toxicological mechanisms of neuro-damage by Tl are shown in Fig. 4.

## 9. Medical treatment of Tl poisoning

Treatment for Tl poisoning patients is performed to eliminate Tl and to prevent further absorption. The most effective antidote is ferric hexacyanoferrate ( $Fe_4[Fe(CN)_6]_3$ ), which is known as Prussian blue (Cvjetko et al., 2010). Prussian blue is orally administered (Galván-Arzate and Santamaría, 1998) and exchanges K for Tl in the gut to increase the fecal excretion of a complex of Tl and Prussian blue (Rusyniak et al., 2010). A recent study demonstrated that MT alone or in combination with Prussian blue plays a cytoprotective role after Tl exposure (Anaya-Ramos et al., 2021). In addition, activated charcoal can be used (Cvjetko et al., 2010; Rusyniak et al., 2010). Hemodialysis and hemoperfusion are also recommended for removing Tl from the blood stream (Malbrain et al., 1997; Thompson, 1981). However, a single antidote is not effective in severe Tl poisoning cases and combinations of different administration have been reported to be effective in a number of cases (Riyaz et al., 2013).

## 10. Conclusions

Tl is one of the most toxic metals. Because Tl salts are colorless, odorless, and tasteless, it has been used historically in homicides, leading it to be known as “the poisoner’s poison.” Tl is found in the natural environment at low levels, and it has been used as, for example, a rodenticide, semiconductor material, and for myocardial imaging. To diagnose Tl poisoning, confirmation of elevated Tl concentrations in the blood, urine, or hair is indispensable. However, evidence for Tl exposure cannot be rapidly obtained by clinicians because Tl analysis can be performed only in limited laboratories. AAS is the most frequently used method for Tl determination in biological samples, and ICP-MS is the most recently introduced method.  $Tl^+$  is more stable than  $Tl^{3+}$ , and  $Tl^+$  salts enter the body through the  $K^+$  uptake mechanism.  $Tl^+$  is rapidly absorbed and is widely distributed to organs. The kidneys show the highest Tl concentration about 24 h after ingestion while the concentration is highest in the brain thereafter. Tl is excreted in the feces and



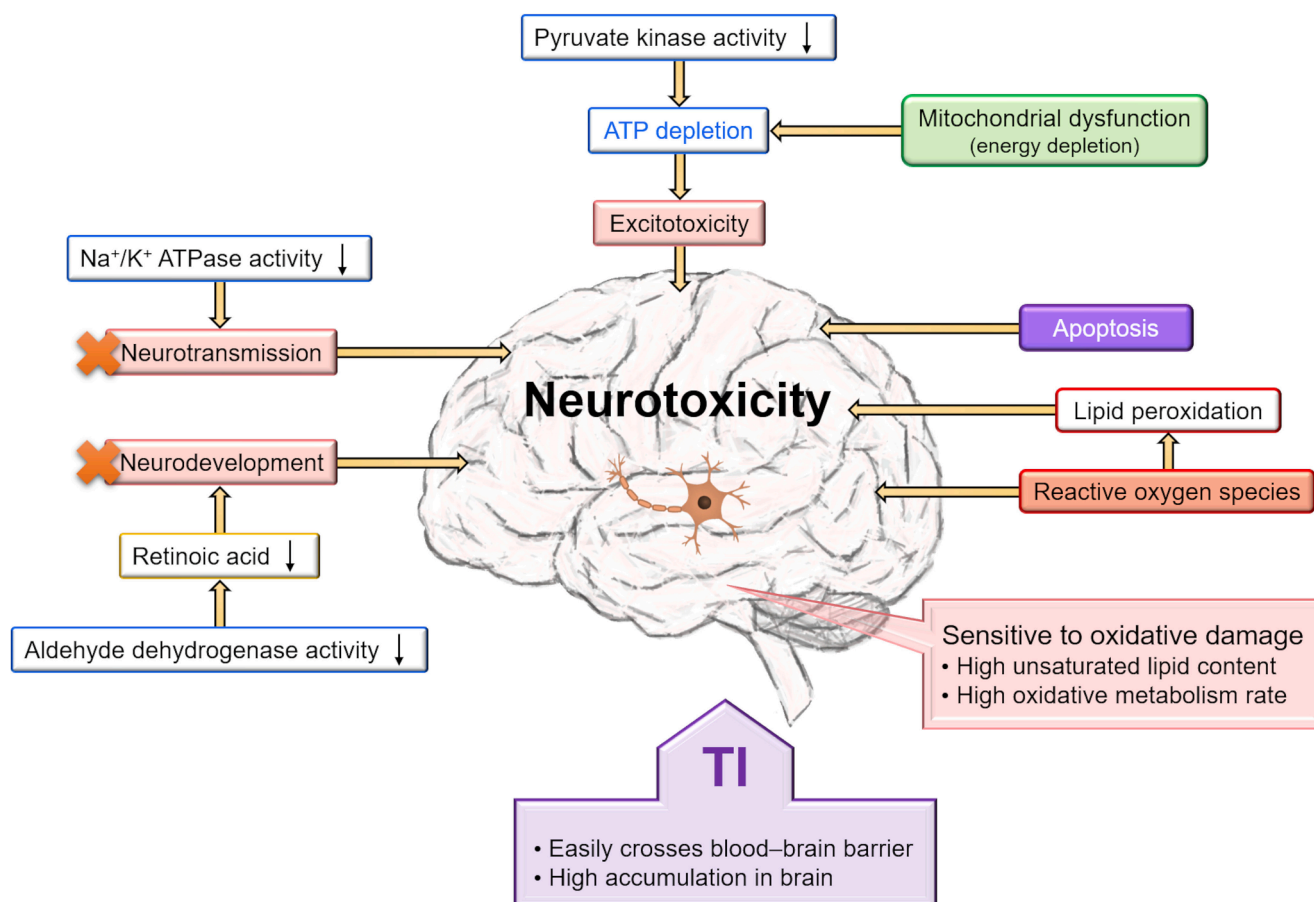


Fig. 4. Schematic of the neurotoxicity mechanism of thallium.

urine. The excretion rate for Tl is quite slow due to the large distribution volume of Tl and it persists for as long as 21 months in untreated Tl poisoning cases. Gastrointestinal symptoms are observed when acute poisoning by ingestion of large amount of Tl. Tl induces neurological dysfunction with similar symptoms to Guillain-Barré syndrome after 2–3 days of exposure. Following 2–4 weeks after exposure, alopecia is observed. Mees' lines, transverse white lines in the nails, are observed about 1 month after Tl poisoning. The toxicological mechanism of Tl is considered to be the ionic radius similarity of  $Tl^+$  to  $K^+$  and the interference of  $Tl^+$  with vital potassium-dependent processes. Another possible mechanism of Tl toxicity is inhibition of enzyme reactions by the binding of Tl to -SH groups, which disturbs vital metabolic processes. Tl toxicity is also related to ROS generation and mitochondrial dysfunction. Tl causes the most severe damage in the central nervous system. The brain is sensitive to oxidative damage and Tl neuro-damage is caused by ROS formation in addition to mitochondrial dysfunction, excitotoxicity, and apoptosis. Placental exposure to Tl can lead to spontaneous abortion, preterm birth, low birth weight, and teratogenicity. The most effective antidote is the ion exchanger Prussian blue, which exchanges K for Tl in the gut to increase the fecal excretion of a complex of Tl and Prussian blue. Recently, it was reported that MT alone or in combination with Prussian blue has cytoprotective effects after Tl exposure. Tl poisoning cases are still reported, and the early detection of Tl in biological samples and its treatment with an antidote are essential.

#### Funding

This work was supported by a JSPS KAKENHI Grant-in-Aid for Scientific Research (B) [grant number 21H03212] to JF.

#### CRediT authorship contribution statement

**Junko Fujihara:** Writing – original draft, Visualization. **Naoki Nishimoto:** Writing – review & editing, Visualization.

#### Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

#### Data availability

No data was used for the research described in the article.

#### References

- Achenbach, C., Hauswirth, O., Heindrichs, C., et al., 1980. Quantitative measurement of time-dependent thallium distribution in organs of mice by field desorption mass spectrometry. *J. Toxicol. Environ. Health* 6, 519–528. <https://doi.org/10.1080/15287398009529870>.
- Agency for Toxic Substances and Disease Registry (ATSDR), 2015. *Toxicological Profile for Thallium*. U.S. Department of Health and Human Services, Public Health Service, Atlanta, GA <https://www.cdc.gov/TSP/ToxProfiles/ToxProfiles.aspx?id=309&tid=49>.
- Al Hammouri, F., Darwazeh, G., Said, A., Ghosh, R.A., 2011. Acute thallium poisoning: series of ten cases. *J. Med. Toxicol.* 7, 306–311. <https://doi.org/10.1007/s13181-011-0165-3>.
- Álvarez-Barrera, L., Rodríguez-Mercado, J.J., Mateos-Nava, R.A., Vázquez-Martínez, Y., Altamirano-Lozano, M.A., 2019. Effect on the offspring of pregnant females CD-1 mice treated with a single thallium(I) application. *Reprod. Toxicol.* 90, 1–7. <https://doi.org/10.1016/j.reprotox.2019.07.022>.
- Anaya-Ramos, L., Díaz-Ruiz, A., Ríos, C., et al., 2021. The acute systemic toxicity of thallium in rats produces oxidative stress: attenuation by metallothionein and

- Prussian blue. *Biometals* 34, 1295–1311. <https://doi.org/10.1007/s10534-021-00343-8>.
- Ash, R.D., He, M., 2018. Details of a thallium poisoning case revealed by single hair analysis using laser ablation inductively coupled plasma mass spectrometry. *Forensic Sci. Int.* 292, 224–231. <https://doi.org/10.1016/j.forsciint.2018.10.002>.
- Blain, R., Kazantzis, G., 2015. Chapter 55 -Thallium. In: Nordberg GF, Costa M (ed) *Handbook on the toxicology of metals*, 4th edn. Volume II: Specific Metals. Academic Press, Cambridge, pp 1229–1240 <https://doi.org/10.1016/B978-0-444-59453-2.00055-X>.
- Bragadin, M., Toninello, A., Bindoli, A., Rigobello, M.P., Canton, M., 2003. Thallium induces apoptosis in Jurkat cells. *Ann. N. Y. Acad. Sci.* 1010, 283–291. <https://doi.org/10.1196/annals.1299.049>.
- Britten, J.S., Blank, M., 1968. Thallium activation of the (Na<sup>+</sup>-K<sup>+</sup>)-activated ATPase of rabbit kidney. *Biochim. Biophys. Acta* 159, 160–166. [https://doi.org/10.1016/0005-2744\(68\)90254-4](https://doi.org/10.1016/0005-2744(68)90254-4).
- Caserta, D., Graziano, A., Lo Monte, G., Bordini, G., Moscarini, M., 2013. Heavy metals and placental fetal-maternal barrier: a mini-review on the major concerns. *Eur. Rev. Med. Pharmacol. Sci.* 17, 2198–2206.
- Cavanagh, J.B., Fuller, N.H., Johnson, H.R.M., Rudge, P., 1974. The effects of thallium salts, with particular reference to the nervous system changes: a report of three cases. *Q. J. Med.* 43, 293–319.
- Chandler, H.A., Archbold, G.P., Gibson, J.M., O'Callaghan, P., Marks, J.N., Pethybridge, R.J., 1990. Excretion of a toxic dose of thallium. *Clin. Chem.* 36, 1506–1509. <https://doi.org/10.1093/clinchem/36.8.1506>.
- Chen, X., Huang, L., Li, Q., et al., 2022. Effect of maternal thallium exposure in early pregnancy on the risk of preterm birth. *Environ. Sci. Pollut. Res. Int.* 29, 49966–49975. <https://doi.org/10.1007/s11356-022-19332-6>.
- Chevalier, G., Ricard, A.C., Manca, D., 1994. Age-related variations of lipid peroxidation in cadmium-treated rats. *Toxicol. Ind. Health* 10, 43–51. <https://doi.org/10.1177/074823799401000103>.
- Cho, K., Lee, S.M., Heo, J., et al., 2021. Retinaldehyde dehydrogenase inhibition-related adverse outcome pathway: Potential risk of retinoic acid synthesis inhibition during embryogenesis. *Toxins* 13, 739. <https://doi.org/10.3390/toxins13110739>.
- Churilov, A.V., Ciampi, G., Kim, H., et al., 2009. Thallium bromide nuclear radiation detector development. *IEEE Trans. Nucl. Sci.* 56, 1875–1881.
- Cvjetko, P., Cvjetko, I., Pavlica, M., 2010. Thallium toxicity in humans. *Arh. Hig. Rada Toksikol.* 61, 111–119. <https://doi.org/10.2478/10004-1254-61-2010-1976>.
- Das, A.K., Chakraborty, R., Cervera, M.L., de la Guardia, M., 2006. Determination of thallium in biological samples. *Anal. Bioanal. Chem.* 385, 665–670. <https://doi.org/10.1007/s00216-006-0411-8>.
- Das, D.D., Sharma, N., Chawla, P.A., 2023. Neutron activation analysis: an excellent nondestructive analytical technique for trace metal analysis. *Crit. Rev. Anal. Chem.* 27, 1–17. <https://doi.org/10.1080/10408347.2023.2178841>.
- De Groot, G., van Heijst, A.N.P., 1988. Toxicokinetic aspects of thallium poisoning. Methods of treatment by toxin elimination. *Sci. Total Environ.* 71, 411–418. [https://doi.org/10.1016/0048-9697\(88\)90213-6](https://doi.org/10.1016/0048-9697(88)90213-6).
- Di Candia, D., Muccino, E., Battistini, A., Boracchi, M., Gentile, G., Zoja, R., 2020. Thallium toxicity due to adulterated infusion with thallium sulfate in eight members belonging to the same family nucleus: Autopsy findings and ICP-MS analysis (inductively coupled plasma mass spectrometry) in a triple homicide. *Leg. Med. (Tokyo)* 42, 101661. <https://doi.org/10.1016/j.legalmed.2019.101661>.
- Diwan, J.J., Lehrer, P.H., 1977. Inhibition of mitochondrial potassium ion flux by thallous ions. *Biochem. Soc. Trans.* 5, 203–205. <https://doi.org/10.1042/bst0050203>.
- Dönmez, B., He, Z., Kim, H., Cirignano, L.J., Shah, K.S., 2010. The stability of TlBr detectors at low temperature. *Nucl. Instrum. Methods Phys. Res. A* 623, 1024–1029. <https://doi.org/10.1016/j.nima.2010.08.024>.
- Duan, W., Wang, Y., Li, Z., et al., 2020. Thallium exposure at low concentration leads to early damage on multiple organs in children: A case study followed-up for four years. *Environ. Pollut.* 258, 113319. <https://doi.org/10.1016/j.envpol.2019.113319>.
- Eskandari, M.R., Mashayekhi, V., Aslani, M., Hosseini, M.J., 2015. Toxicity of thallium on isolated rat liver mitochondria: the role of oxidative stress and MPT pore opening. *Environ. Toxicol.* 30, 232–241. <https://doi.org/10.1002/tox.21900>.
- Ferreira da Silva, A., Souza Dantas, N., de Almeida, J.S., Ahtuja, R., Persson, C., 2005. Electronic and optical properties of wurtzite and zinc-blende TiN and AlN. *J. Cryst. Growth* 281, 151–160. <https://doi.org/10.1016/j.jcrysgro.2005.03.021>.
- Fujihara, J., Nishimoto, N., 2020. Total antimony analysis by hydride generation-microwave plasma-atomic emission spectroscopy with applications. *Microchem. J.* 157, 104992. <https://doi.org/10.1016/j.microc.2020.104992>.
- Fujihara, J., Nishimoto, N., 2023. Speciation analysis of inorganic Sb leached from InSb thin films by hydride generation-microwave plasma-atomic emission spectroscopy. *J. Iran. Chem. Soc.* 20, 2555–2560. <https://doi.org/10.1007/s13738-023-02853-3>.
- Galván-Arzate, S., Martínez, A., Medina, E., Santamaría, A., Ríos, C., 2000. Subchronic administration of sublethal doses of thallium to rats: effects on distribution and lipid peroxidation in brain regions. *Toxicol. Lett.* 116, 37–43. [https://doi.org/10.1016/s0378-4274\(00\)00200-9](https://doi.org/10.1016/s0378-4274(00)00200-9).
- Galván-Arzate, S., Pedraza-Chaverrí, J., Medina-Campos, O.N., et al., 2005. Delayed effects of thallium in the rat brain: regional changes in lipid peroxidation and behavioral markers, but moderate alterations in antioxidants, after a single administration. *Food Chem. Toxicol.* 43, 1037–1045. <https://doi.org/10.1016/j.fct.2005.02.006>.
- Galván-Arzate, S., Santamaría, A., 1998. Thallium toxicity. *Toxicol. Lett.* 99, 1–13. [https://doi.org/10.1016/s0378-4274\(98\)00126-x](https://doi.org/10.1016/s0378-4274(98)00126-x).
- Genchi, G., Carocci, A., Lauria, G., Sinicropi, M.S., Catalano, A., 2021. Thallium use, toxicity, and detoxification therapy: An overview. *Appl. Sci.* 11, 8322. <https://doi.org/10.3390/app11188322>.
- Ghannoum, M., Nolin, T.D., Goldfarb, D.S., et al., 2012. Extracorporeal treatment for thallium poisoning: recommendations from the EXTRIP Workgroup. *Clin. J. Am. Soc. Nephrol.* 7, 1682–1690. <https://doi.org/10.2215/CJN.01940212>.
- Goering, P.L., Morgan, D.L., Ali, S.F., 2002. Effects of mercury vapor inhalation on reactive oxygen species and antioxidant enzymes in rat brain and kidney are minimal. *J. Appl. Toxicol.* 22, 167–172. <https://doi.org/10.1002/jat.844>.
- Goldman, M., Anderson, R.P., Henry, J.P., Peoples, S.A., 1966. X-ray emission spectrographic determination of thallium in biologic materials. *J. Agric. Food Chem.* 14, 367–369. <https://doi.org/10.1021/jf60146a009>.
- Goyer, R.A., Clarkson, T.W., 2001. Toxic effects of metals. In: Klaassen, C.D. (Ed.), *Casarett and Doull's Toxicology: the Basic Science of Poisons*, 6th ed. McGraw-Hill, New York, pp. 855–856.
- Gulebaglan, S.E., Dogan, E.K., Aycibin, M., Secuk, M.N., Erdinc, B., Akkus, H., 2013. Structural and electronic properties of zincblende phase of Tl x Ga1-x As y P1-y quaternary alloys: First-principles study. *Open Phys.* 11 (12), 1680–1685. <https://doi.org/10.2478/s11534-013-0314-1>.
- Gupta, V., Kumar, D., Dwivedi, A., et al., 2023. Heavy metal contamination in river water, sediment, groundwater and human blood, from Kanpur, Uttar Pradesh, India. *Environ. Geochem. Health* 45, 1807–1818. <https://doi.org/10.1007/s10653-022-01290-0>.
- Hann, S., Latkoczy, C., Bereuter, T.L., Prohaska, T., Stinger, G., Reiter, C., 2005. Reconstruction of a case of thallium poisoning using LA-ICP-SFMS. *Int. J. Leg. Med.* 119, 35–39. <https://doi.org/10.1007/s00414-004-0465-0>.
- Hanzel, C.E., Verstraeten, S.V., 2006. Thallium induces hydrogen peroxide generation by impairing mitochondrial function. *Toxicol. Appl. Pharmacol.* 216, 485–492. <https://doi.org/10.1016/j.taap.2006.07.003>.
- Hanzel, C.E., Villaverde, M.S., Verstraeten, S.V., 2005. Glutathione metabolism is impaired in vitro by thallium(III) hydroxide. *Toxicology* 207, 501–510. <https://doi.org/10.1016/j.tox.2004.11.002>.
- Harris, W.R., Messori, L., 2002. A comparative study of aluminum (III), gallium (III), indium (III), and thallium (III) binding to human serum transferrin. *Coord. Chem. Rev.* 228, 237–262. [https://doi.org/10.1016/S0010-8545\(02\)00037-1](https://doi.org/10.1016/S0010-8545(02)00037-1).
- Hasan, M., Ali, S.F., 1981. Effects of thallium, nickel, and cobalt administration of the lipid peroxidation in different regions of the rat brain. *Toxicol. Appl. Pharmacol.* 57, 8–13. [https://doi.org/10.1016/0041-008x\(81\)90019-3](https://doi.org/10.1016/0041-008x(81)90019-3).
- Heitland, P., Köster, H.D., 2021. Human biomonitoring of 73 elements in blood, serum, erythrocytes and urine. *J. Trace Elem. Med. Biol.* 64, 126706. <https://doi.org/10.1016/j.jtemb.2020.126706>.
- Henke, G., 1991. Thallium determination in biological materials by radiochemical neutron activation analysis. *Presenius J. Anal. Chem.* 339, 245–248. <https://doi.org/10.1007/BF00325746>.
- Herman, M.M., Bensch, K.G., 1967. Light and electron microscopic studies of acute and chronic thallium intoxication in rats. *Toxicol. Appl. Pharmacol.* 10, 199–222. [https://doi.org/10.1016/0041-008x\(67\)90104-4](https://doi.org/10.1016/0041-008x(67)90104-4).
- Hirata, M., Taoda, K., Ono-Ogasawara, M., Takaya, M., Hisanaga, N., 1998. A probable case of chronic occupational thallium poisoning in a glass factory. *Ind. Health* 36, 300–303. <https://doi.org/10.2486/indhealth.36.300>.
- Hoffman, R.S., 2000. Thallium poisoning during pregnancy: a case report and comprehensive literature review. *J. Toxicol. Clin. Toxicol.* 38, 767–775. <https://doi.org/10.1081/clt-100102390>.
- Hoffman, R.S., 2003. Thallium toxicity and the role of Prussian blue in therapy. *Toxicol. Rev.* 22, 29–40. <https://doi.org/10.2165/00139709-200322010-00004>.
- Hoffman, R.S., Stringer, J.A., Feinberg, R.S., Goldfrank, L.R., 1999. Comparative efficacy of thallium adsorption by activated charcoal, Prussian blue, and sodium polystyrene sulfonate. *J. Toxicol. Clin. Toxicol.* 37, 833–837. <https://doi.org/10.1081/CLT-100102462>.
- Hologgatis, J., Ullucci, P., Driscoll, J., Grauerholz, J., Martin, H., 1980. Thallium elimination kinetics in acute thallotoxicosis. *J. Anal. Toxicol.* 4, 68–75. <https://doi.org/10.1093/jat/4.2.68>.
- Huang, C., Zhang, X., Li, G., Jiang, Y., Wang, Q., Tian, R., 2014. A case of severe thallium poisoning successfully treated with hemoperfusion and continuous veno-venous hemofiltration. *Hum. Exp. Toxicol.* 33, 554–558. <https://doi.org/10.1177/0960327113499039>.
- Ipcs, 1996. Thallium. *Environmental Health Criteria*, Vol. 182. World Health Organization, Geneva <https://www.inchem.org/documents/ehc/ehc/ehc182.htm>.
- John Peter, A.L., Viraraghavan, T., 2005. Thallium: a review of public health and environmental concerns. *Environ. Int.* 31:493–501. <https://doi.org/10.1016/j.envint.2004.09.003>.
- Johnston, R.E., Mukherji, S.K., Perry, R.J., Stabin, M.G., 1996. Radiation dose from breastfeeding following administration of thallium-201. *J. Nucl. Med.* 37, 2079–2082.
- Kanwar, A.J., Narang, T., 2013. Anagen effluvium. *Indian J. Dermatol. Venereol. Leprol.* 79, 604–612. <https://doi.org/10.4103/0378-6323.116728>.
- Kiliç, G.A., Kutlu, M., 2010. Effects of exogenous metallothionein against thallium-induced oxidative stress in rat liver. *Food Chem. Toxicol.* 48, 980–987. <https://doi.org/10.1016/j.fct.2010.01.013>.
- Klaassen, C.D., Liu, J., 1998. Induction of metallothionein as an adaptive mechanism affecting the magnitude and progression of toxicological injury. *Environ. Health Perspect.* 106, 297–300. <https://doi.org/10.1289/ehp.98106s1297>.
- Korotkov, S.M., Lapin, A.V., 2003. Thallium induces opening of the mitochondrial permeability transition pore in the inner membrane of rat liver mitochondria. *Dokl. Biochem. Biophys.* 392, 247–252. <https://doi.org/10.1023/a:1026182511897>.
- Lan, C.H., Lin, T.S., 2005. Acute toxicity of trivalent thallium compounds to *Daphnia magna*. *Ecotoxicol. Environ. Saf.* 61, 432–435. <https://doi.org/10.1016/j.ecoenv.2004.12.021>.

- Lech, T., Lachowicz, T., 2009. Application of ICP-OES to multielement analysis of biological material in forensic inorganic toxicology. *Prob. Forensic Sci.* 77, 64–78.
- Lennartson, A., 2015. Toxic Thallium. *Nature Chem.* 7, 610. <https://doi.org/10.1038/nchem.2286>.
- Li, S., Huang, W., Duan, Y., Xing, J., Zhou, Y., 2015. Human fatality due to thallium poisoning: autopsy, microscopy, and mass spectrometry assays. *J. Forensic Sci.* 60, 247–251. <https://doi.org/10.1111/1556-4029.12623>.
- Lin, G., Yuan, L., Bai, L., Liu, Y., Wang, Y., Qiu, Z., 2019. Successful treatment of a patient with severe thallium poisoning in a coma using Prussian blue and plasma exchange: A case report. *Medicine (Baltimore)* 98, e14629. <https://doi.org/10.1097/MD.00000000000014629>.
- Lin, G., Sun, Y., Long, J., et al., 2020. Involvement of the Nrf2-Keap1 signaling pathway in protection against thallium-induced oxidative stress and mitochondrial dysfunction in primary hippocampal neurons. *Toxicol. Lett.* 319, 66–73. <https://doi.org/10.1016/j.toxlet.2019.11.008>.
- Liu, H., Liao, G., 2021. Long-term misdiagnosis and neurologic outcomes of thallium poisoning: A case report and literature review. *Brain Behav.* 11, e02032 <https://doi.org/10.1002/brb3.2032>.
- Liu, J., Luo, X., Sun, Y., et al., 2019. Thallium pollution in China and removal technologies for waters: A review. *Environ. Int.* 126, 771–790. <https://doi.org/10.1016/j.envint.2019.01.076>.
- Liu, Y., Wei, L., Luo, D., et al., 2021. Geochemical distribution and speciation of thallium in groundwater impacted by acid mine drainage (Southern China). *Chemosphere* 280, 130743. <https://doi.org/10.1016/j.chemosphere.2021.130743>.
- Malbrain, M.L., Lambrecht, G.L., Zandijk, E., et al., 1997. Treatment of severe thallium intoxication. *J. Toxicol. Clin. Toxicol.* 35, 97–100. <https://doi.org/10.3109/15563659709001173>.
- Maya-López, M., Mireles-García, M.V., Ramírez-Toledo, M., et al., 2018. Thallium-induced toxicity in rat brain crude synaptosomal/mitochondrial fractions is sensitive to anti-excitatory and antioxidant agents. *Neurotox. Res.* 33, 634–640. <https://doi.org/10.1007/s12640-017-9863-1>.
- McMillan, T.M., Jacobson, R.R., Gross, M., 1997. Neuropsychology of thallium poisoning. *J. Neurol. Neurosurg. Psychiatry* 63, 247–250. <https://doi.org/10.1136/jnnp.63.2.247>.
- Meggs, W.J., Hoffman, R.S., Shih, R.D., Weisman, R.S., Goldfrank, L.R., 1994. Thallium poisoning from maliciously contaminated food. *J. Toxicol. Clin. Toxicol.* 32, 723–730. <https://doi.org/10.3109/15563659409017979>.
- Menegola, E., Veltman, C.H.J., Battistoni, M., et al., 2021. An adverse outcome pathway on the disruption of retinoic acid metabolism leading to developmental craniofacial defects. *Toxicology* 458, 152843. <https://doi.org/10.1016/j.tox.2021.152843>.
- Minoia, C., Sabbioni, E., Apostoli, P.I.E.T.R.A., et al., 1990. Trace element reference values in tissues from inhabitants of the European community I. A study of 46 elements in urine, blood and serum of Italian subjects. *Sci. Total Environ.* 95, 89–105. [https://doi.org/10.1016/0048-9697\(90\)90055-Y](https://doi.org/10.1016/0048-9697(90)90055-Y).
- Mizee, M.R., Wooldrik, D., Lakeman, K.A., et al., 2013. Retinoic acid induces blood-brain barrier development. *J. Neurosci.* 33, 1660–1671. <https://doi.org/10.1523/JNEUROSCI.1338-12.2013>.
- Moore, D., House, I., Dixon, A., 1993. Thallium poisoning. Diagnosis may be elusive but alopecia is the clue. *BMJ* 306, 1527–1529. <https://doi.org/10.1136/bmj.306.6891.1527>.
- Mori, I., Matsuo, T., Fujita, Y., et al., 1994. Spectrofluorometric determination of thallium(III) with pyrogallol red and 3,4,5,6-tetrachlorofluorescein. *Presenius J. Anal. Chem.* 348, 346–349. <https://doi.org/10.1007/BF00323132>.
- Mulkey, J.P., Oehme, F.W., 1993. A review of thallium toxicity. *Vet. Hum. Toxicol.* 35, 445–453.
- Nikoo, A.M., Arab, A., Sadeghi, H., 2022. First-principle investigation of TiGaAs Alloys for band detection in SWIR region. *Ind. J. Phys.* 96, 3527–3533. <https://doi.org/10.1007/s12648-022-02295-2>.
- Nishimoto, N., Kobayashi, N., Kawasaki, N., et al., 2003. Low-temperature MBE growth of a TiGaAs/GaAs multiple quantum-well structure. *IEICE Trans. Electron.* E86-C, 2082–2084.
- Nishioka, H., 1975. Mutagenic activities of metal compounds in bacteria. *Mutat. Res.* 31, 185–189. [https://doi.org/10.1016/0165-1161\(75\)90088-6](https://doi.org/10.1016/0165-1161(75)90088-6).
- Nordberg, M., Nordberg, G.F., 2000. Toxicological aspects of metallothionein. *Cell. Mol. Biol. (Noisy-le-Grand)* 46, 451–463.
- Nowicka, A.M., Mackiewicz, M., Matysiak, E., et al., 2013. Voltammetric and electrochemical gravimetric selective detection of interactions between Tl(I) and guanine and the influence on activity of DNA drug-intercalators. *Talanta* 106, 85–91. <https://doi.org/10.1016/j.talanta.2012.12.018>.
- Osorio-Rico, L., Santamaria, A., Galván-Arzate, S., 2017. Thallium toxicity: general issues, neurological symptoms, and neurotoxic mechanisms. *Adv. Neurobiol.* 18, 345–353. [https://doi.org/10.1007/978-3-319-60189-2\\_17](https://doi.org/10.1007/978-3-319-60189-2_17).
- Park, C., Lee, Y., 2023. Monte Carlo simulation study of performance evaluation for sensitivity and scatter fraction in gamma camera scintigraphy with TlBr pixelated semiconductor detector with different parallel-hole collimator designs. *Nucl. Instrum. Methods Phys. Res. A* 1048, 167884. <https://doi.org/10.1016/j.nima.2022.167884>.
- Pragst, F., Hartwig, S., 2022. Repeated poisoning of the life partner by thallium - a case of questionable Munchausen by adult proxy syndrome with ensuing attempted murder. *Int. J. Leg. Med.* 136, 695–704. <https://doi.org/10.1007/s00414-022-02791-4>.
- Qi, J., Lai, Y., Liang, C., et al., 2019. Prenatal thallium exposure and poor growth in early childhood: A prospective birth cohort study. *Environ. Int.* 123, 224–230. <https://doi.org/10.1016/j.envint.2018.12.005>.
- Rader, S.T., Maier, R.M., Barton, M.D., Mazdab, F.K., 2019. Uptake and fractionation of thallium by *Brassica Juncea* in a geogenic thallium-amended substrate. *Environ. Sci. Technol.* 53, 2441–2449. <https://doi.org/10.1021/acs.est.8b06222>.
- Ralph, L., Twiss, M., 2002. Comparative toxicity of thallium(I), thallium(III), and cadmium(II) to the unicellular alga *Chlorella* isolated from Lake Erie. *Bull. Environ. Contam. Toxicol.* 68, 261–268. <https://doi.org/10.1007/s001280247>.
- Ramsden, D., 2002. Thallium. In: Waring, R.H., Steventon, G.B., Mitchell, S.C. (Eds.), *Molecules of Death*. Imperial College Press, London, pp. 304–311.
- Reed, D., Crawley, J., Faro, S.N., Pieper, S.J., Kurland, L.T., 1963. Thallotoxicosis: Acute manifestations and sequelae. *J. Am. Med. Assoc.* 183, 516–522. <https://doi.org/10.1001/jama.1963.03700070044007>.
- Richelmi, P., Bono, F., Guardia, L., Ferrini, B., Manzo, L., 1980. Salivary levels of thallium in acute human poisoning. *Arch. Toxicol.* 43, 321–325. <https://doi.org/10.1007/BF00366188>.
- Rickwood, C.J., King, M., Huntsman-Mapila, P., 2015. Assessing the fate and toxicity of Thallium I and Thallium III to three aquatic organisms. *Ecotoxicol. Environ. Saf.* 115, 300–308. <https://doi.org/10.1016/j.ecoenv.2014.12.024>.
- Rinklebe, J., Shaheen, S.M., El-Naggar, A., et al., 2020. Redox-induced mobilization of Ag, Sb, Sn, and Tl in the dissolved, colloidal and solid phase of a biochar-treated and un-treated mining soil. *Environ. Int.* 140, 105754 <https://doi.org/10.1016/j.envint.2020.105754>.
- Ríos, C., Galván-Arzate, S., Tapia, R., 1989. Brain regional thallium distribution in rats acutely intoxicated with Tl<sub>2</sub>SO<sub>4</sub>. *Arch. Toxicol.* 63, 34–37. <https://doi.org/10.1007/BF00334631>.
- Riyaz, R., Pandalai, S.L., Schwartz, M., Kazzi, Z.N., 2013. A fatal case of thallium toxicity: challenges in management. *J. Med. Toxicol.* 9, 75–78. <https://doi.org/10.1007/s13181-012-0251-1>.
- Rodríguez-Mercado, J.J., Altamirano-Lozano, M.A., 2013. Genetic toxicology of thallium: a review. *Drug Chem. Toxicol.* 36, 369–383. <https://doi.org/10.3109/01480545.2012.710633>.
- Rusyniak, D.E., Furbee, R.B., Kirk, M.A., 2002. Thallium and arsenic poisoning in a small midwestern town. *Ann. Emerg. Med.* 39, 307–311. <https://doi.org/10.1067/mem.2002.122008>.
- Rusyniak, D.E., Arroyo, A., Acciani, J., Froberg, B., Kao, L., Furbee, B., 2010. Heavy metal poisoning: management of intoxication and antidotes. *EXS* 100, 365–396. [https://doi.org/10.1007/978-3-7643-8338-1\\_11](https://doi.org/10.1007/978-3-7643-8338-1_11).
- Rusznayk, I., György, L., Ormai, S., Millner, T., 1968. On some potassium-like qualities of the thallium ion. *Experientia* 24, 809–810. <https://doi.org/10.1007/BF02144884>.
- Sabbioni, E., Gregotti, C., Edel, J., Marafante, E., Di Nucci, A., Manzo, L., 1982. Organ/tissue disposition of thallium in pregnant rats. *Arch. Toxicol. Suppl.* 5, 225–230. [https://doi.org/10.1007/978-3-642-68511-8\\_41](https://doi.org/10.1007/978-3-642-68511-8_41).
- Sánchez-Chapul, L., Santamaria, A., Aschner, M., et al., 2023. Thallium-induced DNA damage, genetic, and epigenetic alterations. *Front. Genet.* 14, 1168713. <https://doi.org/10.3389/fgene.2023.1168713>.
- Spencer, P.S., Peterson, E.R., Madrid, R., Raine, C.S., 1973. Effects of thallium salts on neuronal mitochondria in organotypic cord-ganglia-muscle combination cultures. *J. Cell Biol.* 58, 79–95. <https://doi.org/10.1083/jcb.58.1.79>.
- Sugiura, T., Yamashita, U., 2000. B cell stimulating activity of metallothionein in vitro. *Int. J. Immunopharmacol.* 22, 113–122. [https://doi.org/10.1016/s0192-0561\(99\)00065-x](https://doi.org/10.1016/s0192-0561(99)00065-x).
- Tanaka, J., Yonezawa, T., Ueyama, M., 1978. Acute thallotoxicosis: neuropathological and spectrophotometric studies on an autopsy case. *J. Toxicol. Sci.* 3, 325–334. <https://doi.org/10.2131/jts.3.325>.
- Thompson, D.F., 1981. Management of thallium poisoning. *Clin. Toxicol.* 18, 979–990. <https://doi.org/10.3109/15563658108990328>.
- Tong, J., Liang, C.M., Huang, K., et al., 2020. Prenatal serum thallium exposure and 36-month-old children's attention-deficit/hyperactivity disorder symptoms: Ma'anshan birth cohort study. *Chemosphere* 244, 125499. <https://doi.org/10.1016/j.chemosphere.2019.125499>.
- Tong, J., Liang, C., Wu, X., et al., 2022. Prenatal serum thallium exposure and cognitive development among preschool-aged children: A prospective cohort study in China. *Environ. Pollut.* 293, 118545 <https://doi.org/10.1016/j.envpol.2021.118545>.
- US EPA, 2022. Basic Information about Thallium in Drinking Water. <http://water.epa.gov/drink/contaminants/basicinformation/thallium.cfm>.
- Villaverde, M.S., Hanzel, C.E., Verstraeten, S.V., 2004. In vitro interactions of thallium with components of the glutathione-dependent antioxidant defence system. *Free Radic. Res.* 38, 977–984. <https://doi.org/10.1080/10715760400000950>.
- Wang, X., Ma, T., Wei, C., et al., 2023. Toxic effects of exogenous retinoic acid on the neurodevelopment of zebrafish (*Danio rerio*) embryos. *Neurotoxicol. Teratol.* 100, 107291. <https://doi.org/10.1016/j.ntt.2023.107291>.
- Wang, T.T., Wen, B., Yu, X.N., et al., 2021. Early diagnosis, treatment, and outcomes of five patients with acute thallium poisoning. *World J. Clin. Cases* 9, 5082. <https://doi.org/10.12998/wjcc.v9.i19.5082>.
- Wu, M., Shu, Y., Song, L., et al., 2019. Prenatal exposure to thallium is associated with decreased mitochondrial DNA copy number in newborns: Evidence from a birth cohort study. *Environ. Int.* 129, 470–477. <https://doi.org/10.1016/j.envint.2019.05.053>.
- Wu, M., Wang, L., Song, L., et al., 2021. The association between prenatal exposure to thallium and shortened telomere length of newborns. *Chemosphere* 265, 129025. <https://doi.org/10.1016/j.chemosphere.2020.129025>.
- Yang, Q., Smeyers-Verbeke, J., 1991. Effectiveness of palladium matrix modification for the determination of thallium by graphite furnace atomic absorption spectrometry. *Clin. Chim. Acta* 204, 23–35. [https://doi.org/10.1016/0009-8981\(91\)90213-v](https://doi.org/10.1016/0009-8981(91)90213-v).
- Yu, V., Juhász, M., Chiang, A., Atanaskova Mesinkovska, N., 2018. Alopecia and associated toxic agents: A systematic review. *Skin Appendage Disord.* 4, 245–260. <https://doi.org/10.1159/000485749>.
- Yumoto, T., Tsukahara, K., Naito, H., Iida, A., Nakao, A., 2017. A successfully treated case of criminal thallium poisoning. *J. Clin. Diagn. Res.* 11, OD01–OD02. <https://doi.org/10.7860/JCDR/2017/24286.9494>.

- Zavaliy, L.B., Petrikov, S.S., Simonova, A.Y., et al., 2021. Diagnosis and treatment of persons with acute thallium poisoning. *Toxicol. Rep.* 8, 277–281. <https://doi.org/10.1016/j.toxrep.2021.01.013>.
- Zayan, A., Vandervelde, T.E., 2019. GaTLAs quantum well solar cells for sub-band gap absorption. *MRS Adv.* 4, 2015–2021. <https://doi.org/10.1557/adv.2019.334>.
- Zhang, H.T., Qiao, B.P., Liu, B.P., Zhao, X.G., 2014. Study on the treatment of acute thallium poisoning. *Am. J. Med. Sci.* 347, 377–381. <https://doi.org/10.1097/MAJ.0b013e318298de9c>.
- Zhao, G., Ding, M., Zhang, B., et al., 2008. Clinical manifestations and management of acute thallium poisoning. *Eur. Neurol.* 60, 292–297. <https://doi.org/10.1159/000157883>.
- Zhou, H., Sun, X., Wang, Y., et al., 2021. The mediating role of placental weight change in the association between prenatal exposure to thallium and birth weight: A prospective birth cohort study. *Front. Public Health* 9, 679406. <https://doi.org/10.3389/fpubh.2021.679406>.
- Zhu, Y.D., Liang, C.M., Hu, Y.B., et al., 2020. Repeated measures of prenatal thallium exposure and placental inflammatory cytokine mRNA expression: The Ma'anshan birth cohort (MABC) study. *Chemosphere* 246, 125721. <https://doi.org/10.1016/j.chemosphere.2019.125721>.
- Zhuang, W., Song, J., 2021. Thallium in aquatic environments and the factors controlling TI behavior. *Environ. Sci. Pollut. Res. Int.* 28, 35472–35487. <https://doi.org/10.1007/s11356-021-14388-2>.
- Zitko, V., Carson, W.V., Carson, W.G., 1975. Thallium: occurrence in the environment and toxicity to fish. *Bull. Environ. Contam. Toxicol.* 13, 23–30. <https://doi.org/10.1007/BF01684859>.