Supplementary Material

Screening bisphenols in complex samples via a planar *Arxula*adeninivorans bioluminescence bioassay

Max Jaber^{1,#}, Martin Jähne², Michaela Oberle³, Gertrud E. Morlock^{1,#,*}

¹Institute of Nutritional Science, Chair of Food Science, and TransMIT Center for Effect-Directed Analysis, Justus Liebig University Giessen, Heinrich-Buff-Ring 26-32, 35392 Giessen, Germany

²QuoData GmbH, Prellerstrasse 14, 01309 Dresden, Germany

³Merck KGaA, Frankfurter Str. 250, 64293 Darmstadt, Germany

*Authors contributed equally.

*Corresponding Author: Prof. Dr. Gertrud Morlock, Justus Liebig University Giessen, Heinrich-Buff-Ring 26-32, 35392 Giessen, Germany, phone: +49-641-9939141; fax +49-641-99-39149; gertrud.morlock@uni-giessen.de

Table S1 Origin of the (a) six tin cans, (b) five thermal papers with concentration, and (c) eleven botanicals

a) S	ix differently co	oated R&D tin cans from Ceritec SRL,	, Metlac Group, Italy	7
ID	34	36 38 39	64	65
b) F	ive thermal pap	oers collected from local retailers in G	iessen, Germany, in	January 2022
ID	Distributor, C	City, Country	c [g/mL]	
A	Aldi Süd, Mühlheim an der Ruhr, Germany		0.13	
Е	Esso, Echo Tar	nkstellen, Hamburg, Germany	0.17	
О	Obi, Wermelsk	kirchen, Germany	0.17	
R	Rewe, Köln, Germany		0.20	
V	Vision Augenoptik & Kontaktlinsen, Giessen, Germany		y 0.20	
ID	Common	Botanical name	Family	Plant part
1	Acerola	Malpighia glabra L.	Malphighiaceae	fruit
2	Galangal	Alpinia officinarum Hance	Zingiberaceae	root
3	Hops	Humulus lupulus L.	Cannabaceae	blossom
4	Chamomile	Matricaria chamomilla L.	Asteraceae	blossom
5	Orange	Citrus × aurantium L.	Rutaceae	peel
6	Oregano	Origanum vulgare L.	Lamiaceae	herb
7	Rooibos	Aspalathus linearis (Burm. f.) Dahlgren	R. Fabaceae	leaf
8	Licorice	Glycyrrhiza glabra L.	Fabaceae	root
9	Thyme	Thymus vulgaris L.	Lamiaceae	herb
10	Hawthorn	Crataegus sp.	Rosaceae	leaf
11	Lemon	Citrus × limon (L.) Osbeck	Rutaceae	peel

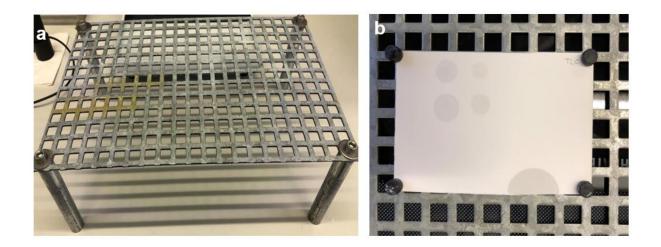


Fig. S1 Setup for plate incubation: metal perforated plate (a) and TLC plate with magnets attached to the underside of the metal plate. The construction was placed in a closed water bath, so that the distance between the TLC plate and the water was 3 cm.

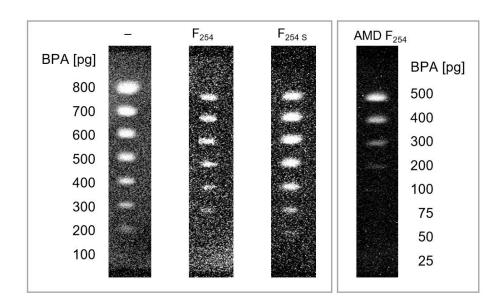


Fig. S2 Study of the bioluminescent signal response and dose-response dependency of BPA (100–800 or 25–500 pg/band) on four different types of HPTLC plates via the pA-YBS bioluminescence bioassay with a 2-h incubation; bioluminescence depicted as greyscale image with a 10-min exposure time.

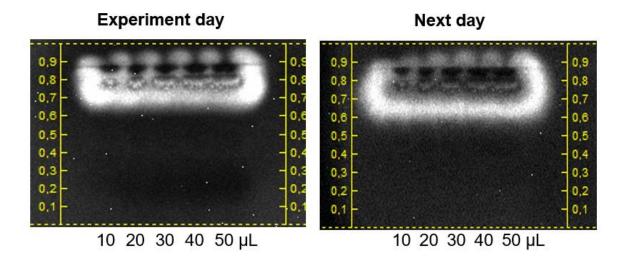


Fig. S3 Bioautogram of thermal paper V (Table 1) at different amounts (10–50 μ L) on HPTLC plate silica gel 60. After the bioluminescence measurement (experiment day), the plate was dried and stored overnight. To reactivate the luciferase reaction on the next day, the plate was incubated for 20 min in the water bath (Figure S1), and then the bioluminescence was recorded with a 10-min exposure time.