SYSTEMATIC REVIEW Open Access

BMC Pulmonary Medicine

Effect of emodin on acute lung injury: a meta-analysis of preclinical trials

Lei Liu^{1,2†}, Yu Zhang^{3†}, Xiao-Ren Tang⁴, Guo-Bing Jia¹, Shan Zhou⁵, Guo-Long Yue² and Cheng-Shi He^{1*}

Abstract

Background Emodin has protective effects on acute lung injury (ALI) or acute respiratory distress syndrome (ARDS). This meta-analysis intended to illustrate the efficacy of emodin on ALI/ARDS animal models.

Methods Relevant preclinical studies were searched on PubMed, EMBASE, and Web of Science. Standardized mean diferences (SMDs) with corresponding confdence intervals (CIs) were used to compare lung injury scores, lung wetto-dry weight ratios (W/D), myeloperoxidase (MPO), tumor necrosis factor-α (TNF-α), interleukin-1β (IL-1β), IL-6, IL-18, $PaO₂$, and PaCO₂ between the treatment and control groups. The article quality was appraised using the SYRCLE tool.

Results Twenty one studies published between 2014 and 2023 were enrolled. Compared with the control group, emodin signifcantly reduced lung injury scores (SMD: -3.63; 95% CI: -4.36, -2.90; *p*<0.00001), W/D ratios (SMD: -3.23; 95% CI: -4.29, -2.16; *p*<0.00001), and MPO levels (SMD: -2.96; 95% CI: -3.92, -1.99; *p*<0.00001). Furthermore, emodin downregulated TNF-α (SMD: -3.04; 95% CI: -3.62, -2.47; *p*<0.00001), IL-1β (SMD: -3.76; 95% CI: -4.65, -2.87; *p*<0.00001), IL-6 (SMD: -3.19; 95% CI: -3.95, -2.43; *p*<0.00001), and IL-18 levels (SMD: -4.83; 95% CI: -6.10, -3.57; *p*<0.00001). Emodin improved gas exchange dysfunction, increased PaO₂ (SMD: 3.76; 95% CI: 2.41, 5.11; $p < 0.00001$), and decreased PaCO₂ (SMD: -3.83; 95% CI: -4.90, -2.76; *p*<0.00001). Sensitivity analyses and stratifed analyses were conducted for outcome measures with heterogeneity.

Conclusions Emodin treatment can efectively reduce the severity of ALI in animal models. Additional animal investigations and clinical trials involving human subjects are imperative.

Keywords Emodin, Acute lung injury, Acute respiratory distress syndrome, Animal model, Meta-analysis

† Lei Liu and Yu Zhang have contributed equally this work.

*Correspondence:

Cheng-Shi He

18708401072@163.com

¹ Department of Respiratory Medicine, Hospital of Chengdu University of Traditional Chinese Medicine, Chengdu Sichuan 610000, China

² Department of Respiratory and Critical Care Medicine, Chongqing

Hospital of Traditional Chinese Medicine, Jiangbei Chongqing 400000, China

³ Department of Respiratory Medicine, Traditional Chinese Medicine Hospital of Renshou County, Meishan Sichuan 620500, China

4 Department of Traditional Chinese Medicine, Bishan Hospital

of Chongqing, Bishan Hospital of Chongqing Medical University, Bishan Chongqing 404000, China

⁵ Department of Endocrinology, Hospital of Chengdu University of Traditional Chinese Medicine, Chengdu Sichuan 610000, China

Introduction

ALI/ARDS is a heterogeneous disease attributed to various pulmonary and extra-pulmonary factors, such as severe sepsis caused by endotoxins, trauma, and burns [[1\]](#page-10-0). Its clinical presentations encompass alveolar damage, acute difuse infammation, interstitial edema, enhanced pulmonary capillary permeability, and infammatory infltration [[2\]](#page-10-1). Acute and progressive hypoxemia and ARDS are common in ALI patients [[3\]](#page-10-2). According to the international epidemiological survey of "Lung Safe", the mortality rate of ARDS is 34.9%-46.1%, and its prevalence accounts for 10.4% of all ICU admissions globally [\[4](#page-10-3)]. Despite extensive research efforts on exploring new treatment modalities for ARDS, efective therapeutic interventions are still lacking [[1](#page-10-0)]. In common mammals such

© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License, which permits any non-commercial use, sharing, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if you modifed the licensed material. You do not have permission under this licence to share adapted material derived from this article or parts of it. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit<http://creativecommons.org/licenses/by-nc-nd/4.0/>.

as rats and mice, ALI/ARDS can be simulated by various methods, including cecal ligation and puncture (CLP) [[5\]](#page-10-4), intratracheal instillation [[6\]](#page-10-5), intraperitoneal injection [[7\]](#page-10-6) or tail vein injection [\[8\]](#page-10-7) of lipopolysaccharide (LPS), intraperitoneal injection of taurocholate sodium [\[9\]](#page-10-8), and pulmonary artery occlusion and reperfusion $[10]$ $[10]$. Therefore, systematic reviews of preclinical trials may enhance our understanding of the characteristics and treatment of human ALI/ARDS.

Emodin is a natural anthraquinone derivative, chemically known as 1, 3, 8 – trihydroxy – 6-methylanthraquinone, with a molecular weight of 270.23 [\[8](#page-10-7)]. It is extracted from various plants, such as rhubarb [\[11](#page-10-10)], Polygonum cuspidatum [[12](#page-10-11)], Polygonum multiflorum [[13\]](#page-10-12), and Cassia obtusifolia [\[14](#page-10-13)]. Numerous prior investigations have established that emodin has a wide range of pharmacological efects against infammation [\[6](#page-10-5)], oxidant stress [\[15](#page-10-14)], tumor [\[16](#page-10-15)], bacteria [[17\]](#page-10-16), viral infections [[18\]](#page-10-17), as well as immune regulation [\[13](#page-10-12)]. Preclinical studies have confrmed the protective efect of emodin on ALI [\[10](#page-10-9), [19](#page-10-18), [20\]](#page-10-19), suggesting emodin may be a potential valuable therapeutic option for ALI. However, it is hard to assess the overall therapeutic efect because of diferent research methods. Currently, there is no systematic review of the role of emodin in ALI. This study was conducted to systematically evaluate the quality and comprehensive evidence of preclinical articles on the functions of emodin on diferent animal models of ALI.

Materials and methods

Search strategy

Systematic reviews and meta-analyses were conducted according to the PRISMA guidelines [[21](#page-10-20)]. PubMed, EMBASE, and Web of Science were searched up to October 23, 2023 to flter preclinical articles related to the effect of emodin on animal models of ALI. The search terms involved "[emodin (title/abstract)] and [acute lung injury (title/abstract) or acute respiratory distress syndrome (title/abstract)]" (Appendix 1).

Eligibility criteria

The inclusion criteria were established as per the PICOS protocol: 1) animal model of ALI; 2) the emodin experimental group; 3) with a placebo control group; 4) exploring the efficacy of emodin on the animal model of ALI; 5) results including lung injury score, MPO, W/D ratio, TNF- α , IL-1 β , IL-6, IL-18, PaO₂, and PaCO₂; 6) results expressed or converted to mean and standard deviation; 7) published in English.

Exclusion criteria covered: 1) review articles, letters, and case reports, repeatedly published articles and abstracts without full text; 2) observational studies and non-randomized controlled studies; 3) ex vivo studies, in vitro studies, human studies, and in silico studies; 4) animals treated with other drugs or surgical procedures; 5) studies testing the efects of emodin in combination with other chemicals or drugs.

Data collection

All data were independently extracted by two reviewers (Liu and Zhang). Any discrepancies were addressed through discussion with a third reviewer (Tang). The collected information encompassed: 1) author and publication year; 2) animal model: species, age, gender, and modeling method); 3) emodin treatment: intervention dose (initial and total dose), treatment time, and administration route; 4) evaluation time; 5) outcome measurement; 6) detection specimen; 7) quality score. If a study contained more than one experimental group, the experimental groups would be diferentiated according to animal species, emodin doses, treatment times, evaluation time points, and detection specimen sources, and compared with the control group. For continuous variables, statistical calculations were performed using mean and standard deviation. For graphical data, Getdata Graph Digitizer was used to check the mean and standard deviation values of high magnifcation. Standard deviations not reported directly were calculated by multiplying the reported mean standard error (SEM) by the square root of the group size.

Quality assessment

Quality assessment was undertaken by the same two reviewers independently using the SYRCLE (adapted from the Cochrane tool) [\[22](#page-10-21)]. Any disputes were addressed through discussion with a third reviewer (Tang).

Statistical analysis

Review Manager (RevMan) 5.4 (Cochrane Library, London, UK) and STATA 17 (StataCorp, College Station, TX, USA) were adopted for statistical analysis. As the primary outcome was continuous variables, standard mean diference (SMD) and 95% confdence interval (95% CI) were utilized to present the effect size. I^2 was used to test the heterogeneity between studies. I^2 < 50% denoted low heterogeneity and the fxed-efect model was used, otherwise, for high heterogeneity, the random-efect model was used. Sensitivity analysis was implemented using the leave-one-out method, and stratifed meta-analysis was

performed to confrm the infuence of methodological diferences. Egger's linear regression test was utilized to determine publication bias. *p*<0.05 implied statistically significant.

Results

Study selection

The flowchart of literature search process is displayed in Fig. [1](#page-2-0). 35 articles from PubMed, 69 from EMBASE, and 60 from Web of Science were searched. After eliminating duplicates, 131 articles were left. Based on the abstract and/or title, we excluded 94 irrelevant reports and selected 37 articles. Then, we read the full texts, of which 17 studies were excluded due to cell experiments, reporting diferent outcome indicators, and irrelevant or unclear interventions. Finally, 20 articles published from 2014 to 2023 were enrolled [[5–](#page-10-4)[10,](#page-10-9) [19](#page-10-18), [20,](#page-10-19) [23](#page-10-22)[–34](#page-11-0)].

Study characteristics

We included 20 articles, of which one [[25\]](#page-10-23) contained two studies, so there were 21 studies. Among the included animal models, there were 3 BALB/c mice, 11 SD rats, 4 Wistar rats, 1 Lewis rat, 1 C57/black mouse, and 1 C57BL/6J mouse. 14 studies reported animal age, ranging from 5 to 8 weeks. Among these studies, there were several methods of ALI modeling: 2 CLP, 4 abdominal injections of LPS, 3 intratracheal injections of LPS, 2 intravenous injections of LPS, 9 severe acute pancreatitis, and 1 pulmonary ischemia–reperfusion injury. 9 studies initiated emodin treatment before ALI and 12 studies after ALI. For the routes of emodin administration, 5 studies adopted abdominal injections and 16 used gavages. 13 studies adopted single-dose administration and 8 studies used multiple-dose administration. Detailed characteristics are exhibited in Table [1](#page-3-0).

Fig. 1 PRISMA fow diagram for review and selection process of studies included in meta-analysis of Emodin in rodent models of ALI

Quality assessment

The quality assessment results are detailed in Table [2](#page-4-0). A high score indicates high methodological quality. In our study, the research scores ranged from 4 to 6. The reported baseline characteristics between the two groups were similar in all studies, without incomplete outcome data, nor biases in the selection of outcome recording or other sources. All studies used randomization methods for animal grouping but no specifc method was mentioned. No study described allocation concealment or blinding for outcome evaluators, animal breeders, and researchers.

*Meta***‑analysis**

Emodin signifcantly reduced lung injury scores (SMD: -3.63; 95% CI: -4.36, -2.90; *p*<0.00001, 11 articles, 18 comparisons, Fig. [2A](#page-5-0)), with high heterogeneity $(I^2=62\%;$ $p=0.0002$). Emodin diminished the W/D ratio (SMD: -3.23; 95% CI: -4.29, -2.16; *p*<0.00001, 8 articles, 13 com-parisons, Fig. [2](#page-5-0)B), with notable heterogeneity $(I^2=86\%;$ *p*<0.00001). Emodin downregulated MPO (SMD: -2.96; 95% CI: -3.92, -1.99; *p*<0.00001, 9 articles, 13 com-parisons, Fig. [2C](#page-5-0)), with marked heterogeneity $(I^2=79\%;$ *p*<0.00001). Emodin downregulated TNF-α (SMD: -3.04;

95% CI: -3.62, -2.47; *p*<0.00001, 16 articles, 32 compari-sons, Fig. [2D](#page-5-0)), with significant heterogeneity $(I^2=73\%;$ *p*<0.00001). Emodin downregulated IL-1β (SMD: -3.76; 95% CI: -4.65, -2.87; *p*<0.00001, 12 articles, 26 compari-sons, Fig. [2](#page-5-0)E) with significant heterogeneity ($I^2 = 83\%$; p <0.00001). Emodin downregulated IL-6 (SMD = -3.19; 95% CI: -3.95, -2.43; *p*<0.00001, 10 studies, 21 com-parisons, Fig. [2](#page-5-0)F) with notable heterogeneity $(I^2=76\%;$ *p*<0.00001). Emodin downregulated IL-18 (SMD: -4.83; 95% CI: -6.10, -3.57; *p*<0.00001, 4 articles, 8 compari-sons, Fig. [2](#page-5-0)G) with marked heterogeneity $(I^2=64\%;$ $p = 0.007$). Emodin increased PaO₂ (SMD = 3.76; 95% CI: 2.41, 5.11; *p*<0.00001, 4 articles, 8 comparisons, Fig. [2H](#page-5-0)) with notable heterogeneity $(I^2=75\%, p=0.0002)$. Emodin reduced PaCO₂ (SMD = -3.83; 95% CI: -4.90, -2.76; *p*<0.00001, 4 articles, 8 comparisons, F [ig](#page-5-0). [2I](#page-5-0)) with notable heterogeneity (I^2 =58%, *p*=0.02).

Stratifed analyses

Stratifed analyses were performed based on animal species, ALI modeling methods, emodin treatment time points, administration routes, dosing frequency, total emodin dose, and detection specimen sources. For lung injury scores, there were signifcant diferences

1-stochastic distribution sequence; 2-analogous baseline traits; 3-distribution concealment; 4-stochastic housing; 5-blinded intervening; 6-random collection for outcome measurement; 7-blinded evaluation of result; 8-unfnished outcome data; 9-selecting outcome recording; 10-else sources of bias

+: yes

no: ※: unclear

← Fig. 2 A Forest plot analyzing the effect of emodin treatment of lung injury score. **B** Forest plot analyzing the effect of emodin treatment of W/D ratio. **C** Forest plot analyzing the efect of emodin treatment of MPO. **D** Forest plot analyzing the effect of emodin treatment of TNF−α. **E** Forest plot analyzing the efect of emodin treatment of IL−1β. **E** Forest plot analyzing the efect of emodin treatment of IL−6. **G** Forest plot analyzing the efect of emodin treatment of IL−18. **H** Forest plot analyzing the efect of emodin treatment of PaO₂. **I** Forest plot analyzing the effect of emodin treatment of $PaCO₂$

in estimated efects on animal species, ALI modeling methods, emodin treatment time points, administration routes, dosing frequency, and total dose $(p<0.05)$ (Table S1). For the W/D ratio, there were signifcant diferences in estimated efects on animal species, ALI modeling methods, emodin treatment time points, administration routes, dosing frequency, and total dose $(p<0.05)$ (Table S2). For MPO, there were significant diferences in estimated efects on animal species, ALI modeling methods, emodin treatment time points, administration routes, dosing frequency, and detection specimen $(p<0.05)$; there was significant estimated efect only in the 10–50 mg/kg and>200 mg/kg groups of total emodin dose $(p<0.05)$ (Table S3). For TNF- α , there were signifcant diferences in estimated efects on ALI modeling methods, emodin treatment time points, administration routes, dosing frequency, total emodin dose, and detection specimen $(p<0.05)$; there was marked estimated efect except for the C57BL/6J mice group within animal species group $(p<0.05)$ (Table S4). For IL-1β, there were signifcant diferences in estimated efects on animal species, ALI modeling methods, emodin treatment time points, administration routes, dosing frequency, and detection specimen $(p<0.05)$; there were notable estimated efects only in the 10–50 mg/kg and 50–200 mg/kg groups of total emodin dose $(p < 0.05)$ (Table S5). For IL-6, there were signifcant diferences in estimated efects on animal species, emodin treatment time points, administration routes, dosing frequency, total emodin dose, and detection specimen $(p < 0.05)$; there was visible estimated efect except for the tail vein LPS injection group within ALI modeling methods group $(p<0.05)$ (Table S6). For IL-18, there were significant differences in estimated efects on animal species, ALI modeling methods, emodin treatment time points, dosing frequency, and detection specimen $(p < 0.05)$ (Table S7). There were significant differences in estimated effects on animal species, dosing frequency, and total emodin dose for PaO₂ ($p < 0.05$) (Table S8) and PaCO₂ ($p < 0.05$) (Table S9).

Fig. 3 A Sensitivity analysis of emodin treatment of lung injury score. **B** Sensitivity analysis of emodin treatment of W/D ratio. **C** Sensitivity analysis of emodin treatment of MPO. **D** Sensitivity analysis of emodin treatment of TNF−α. **E** Sensitivity analysis of emodin treatment of IL−1β. **F** Sensitivity analysis of emodin treatment of IL−6. **G** Sensitivity analysis of emodin treatment of IL−18. **H** Sensitivity analysis of emodin treatment of PaO2. **I** Sensitivity analysis of emodin treatment of PaCO₂

Sensitivity analysis

Sensitivity analyses were implemented to judge the result stability. Lung injury score (Fig. [3](#page-6-0)A), W/D ratio (Fig. [3](#page-6-0)B), MPO (Fig. [3C](#page-6-0)), TNF-α (Fig. [3](#page-6-0)D), IL-1β (Fig. [3E](#page-6-0)), IL-6 (Fig. [3](#page-6-0)F), IL-18 (Fig. 3G), PaO₂ (Fig. 3H), and PaCO₂ (Fig. [3I](#page-6-0)) were not signifcantly afected by any combined SMD studies.

Publication *bias*

According to the Egger test, there was a publication bias in the lung injury score ($p < 0.001$), W/D ratio ($p = 0.002$), MPO (*p*<0.001), TNF-α (*p*<0.001), IL-1β (*p*<0.001), and IL-6 $(p < 0.001)$. In this case, to estimate the missing studies and recalculate the efect estimates, we conducted a trim-and-fll analysis on the lung injury score, W/D, MPO, TNF-α, IL-1β, and IL-6, which showed no small sample efect (Figure S1A-F and Table S10).

Time‑dose interval analysis

Lung injury score is a microscopic pathological scoring scale extensively adopted in preclinical trials to assess the severity of lung injury $[35]$ $[35]$. The time-dose effect diagram showed that the minimum dose of emodin to improve the lung injury score was 5 mg/kg $(p<0.05)$, and the maximum dose was 700 mg/kg $(p<0.05)$. The shortest expected duration of emodin administration to improve the lung injury score $(p<0.01)$ was 2 h, and the longest expected duration was 72 h $(p<0.05)$. The overall results disclosed that emodin at doses of 5–700 mg/kg and expected durations of 2–72 h showed relatively superior efficacy (Fig. 4).

Discussion

Summary of evidence

Emodin can alleviate tissue damage in various diseases. Studies have shown that emodin can efectively repress the infammatory response and tissue damage in animal models of keratitis, nephritis, and hepatitis [[36–](#page-11-5)[38\]](#page-11-6). In terms of lung injury, emodin is also a promising protective agent in animal models of ALI [[7](#page-10-6), [24](#page-10-28), [25\]](#page-10-23). To date, there has been no meta-analysis on the efficacy of emodin in animal models of ALI. Our research in animal

Fig. 4 Time-dose interval analysis scatter plot for lung injury score

models of ALI confrmed the therapeutic efect of emodin. The pooled results indicated that emodin could markedly reduce the lung injury score, directly proving the alleviation of emodin on lung injury $[39]$ $[39]$. The lung W/D ratio is widely used in animal experiments to evaluate pulmonary vascular permeability. Our analysis of the studies suggested a decrease in pulmonary vascular permeability, indicating that emodin could improve lung water clearance [[35\]](#page-11-4). TNF-α, IL-1β, and IL-6 are major initiators of lung injury. These prominent cytokines actively recruit leukocytes to the lungs and accelerate lung injury by inducing the production of monocytes and macrophages $[40, 41]$ $[40, 41]$ $[40, 41]$ $[40, 41]$ $[40, 41]$. Additionally, an excessive release of IL-1β enhances the permeability of alveolar epithelium and vascular endothelial cells, leading to pulmonary edema [[42\]](#page-11-10). Our study results indicated that emodin downregulated TNF-α, IL-1β, and IL-6 levels. Neutrophils-released MPO acts as a marker for neutrophil accumulation [\[43](#page-11-11)]. During infammation, it is released into the extracellular fuid and participates in lipid oxidation. Increased MPO activity is recognized as a sign of neutrophil activation and accumulation, potentially resulting in tissue damage [[44,](#page-11-12) [45\]](#page-11-13). Our study confrmed the downregulating efect of emodin on MPO. As a product of apoptosis, IL-18 triggers a cascade of infammatory reactions to exacerbate the infammatory response in ALI [[46\]](#page-11-14). IL-18 expression is conversely correlated with long-term survival in patients with ARDS [[47\]](#page-11-15). Our results unraveled that emodin downregulated IL-18 levels. Gas exchange dysfunction is an important pathological feature of ARDS, which manifests as hypoxemia and hypercapnia [\[48](#page-11-16), [49](#page-11-17)]. Our analysis of the studies found that emodin increased Pa O_2 level, decreased PaCO₂ level, and improved gas exchange. The metaanalysis of preclinical articles suggests that emodin could potentially be utilized to protect patients with ALI.

Possible mechanisms

Over the past several years, extensive research has been conducted to examine the efectiveness and mechanism of emodin in ALI. Emodin has shown great potential as a therapeutic drug for lung injury. Emodin treatment has demonstrated its potential to ameliorate the detrimental efects of LPS-induced ALI. It efectively alleviates pulmonary pathological damage, pulmonary edema, oxidative stress, the hypercoagulable state of alveoli, and fbrinolysis [\[6](#page-10-5)]. For ischemia–reperfusion-induced ALI,

emodin can mitigate alveolar damage and pulmonary edema and increase arterial oxygenation and dynamic compliance [[10\]](#page-10-9). For CLP-induced ALI, emodin can reduce pulmonary cell apoptosis [\[5\]](#page-10-4). Emodin treatment also alleviates pulmonary edema and tissue damage and inhibits inflammation in SAP-ALL $[23]$ $[23]$ $[23]$. The therapeutic efects of emodin treatment may achieved by inhibiting the activation of NLRP3 infammasome [[9\]](#page-10-8), the activation of NF-κB $[6]$ $[6]$, the MAPK inflammatory pathway [[27\]](#page-10-29), the aggregation of infammatory cells, the release of infammatory cytokines [[6\]](#page-10-5), PMN infltration, and promoting PMN apoptosis [\[29](#page-10-26)], tight junction proteins and aquaporin $[5]$ $[5]$, and autophagy pathways $[28]$ $[28]$. These mechanisms will guide future exploration of the therapeutic targets of ALI. Therefore, we believe that emodin is a promising drug for ALI and deserves more research.

Interpretation of stratifed analysis

Emodin signifcantly improved lung injury scores, W/D ratio, MPO, TNF-α, IL-1β, IL-6, IL-18, PaO₂, and PaCO₂. However, there was significant heterogeneity. Therefore, we conducted stratifed analyses based on animal species, ALI modeling methods, emodin treatment time points, administration routes, dosing frequency, total doses, and detection specimens.

Animal species

The stratified analysis of animal species demonstrated that emodin treatment efectively improved lung injury scores, W/D ratio, MPO, IL-1 β , IL-6, IL-18, PaO₂, and PaCO₂ in all animal species. Except for C57/black mice, the other animal species showed signifcantly improved TNF-α. Among all animal species, BALB/c mice exhibited the largest efect size. Due to fewer studies on $BALB/c$ mice, the efficacy of emodin may be overestimated. Most studies selected SD rats and Wistar rats as research subjects. We cannot confrm which animal species is more suitable for ALI models. Therefore, the choice of animal species ought to be standardized in future investigations.

ALI modeling methods

The stratified analysis of the methods of inducing ALI modeling methods displayed that emodin treatment efectively improved lung injury scores, W/D ratio, MPO, TNF-α, IL-1β, and IL-18 in all induction method groups. Except for the LPS tail vein injection, the other groups showed signifcantly improved IL-6. We further found diferences in the total dose of emodin within the LPS tail vein injection group [\[34](#page-11-0)], which were 20mg/ kg and 50mg/kg. Emodin of 20mg/kg did not signifcantly improve IL-6, while 50mg/kg emodin signifcantly improved IL-6. Therefore, the difference in the total dose of emodin may explain the insignifcant combined results. Among all ALI induction methods, most studies selected the construction of SAP-induced ALI, followed by LPS-induced ALI. The induction methods included LPS intratracheal perfusion, LPS intraperitoneal injection, and LPS tail vein injection. The LPS doses varied across studies; few studies selected CLP for modeling. However, CLP had the largest effect size. Previous studies have considered LPS as one of the most compatible methods for inducing animal models of ALI [[50\]](#page-11-18). Furthermore, compared with CLP-induced ALI models, ALI models induced by LPS intratracheal perfusion are more similar to human ARDS in the acute stage [\[51](#page-11-19)]. However, regardless of the induction method, it can only simulate a part of the basic diseases of human ALI. Therefore, more studies are needed in the future to determine which method is more suitable for ALI research.

Time, frequency, route, and dose of emodin treatment

Firstly, the stratifed analysis of the time points denoted that emodin treatment efectively improved lung injury scores, W/D ratio, MPO, TNF-α, IL-1β, IL-6, and IL-18 whether administered before or after ALL modeling. Therefore, emodin treatment had both preventive and therapeutic efects on ALI.

Secondly, the stratifed analysis of emodin dosing frequency found that both single-dose and multiple-dose administration displayed efective improvement in all outcomes. Besides, multiple doses of emodin administration were superior to single-dose administration in improving lung injury scores, W/D ratio, MPO, PaO₂, and PaCO₂; single-dose administration was superior to multiple-dose administration in improving TNF-α, IL-1β, IL-6, and IL-18. More studies are needed to explore the optimal dosing frequency.

Thirdly, the stratified analysis of administration routes found that both intragastric and intraperitoneal injections of emodin showed efective improvement. Most studies used intragastric administration and had a larger effect size. Therefore, intragastric administration may be more suitable than intraperitoneal injection, and more studies are required to determine more appropriate administration routes.

Finally, our stratifed analysis of the total dose of emodin found that emodin at all four doses efectively improved the lung injury score and TNF-α. At doses of 10–50 mg/ kg, 50–200 mg/kg, and>200 mg/kg, emodin efectively improved the W/D ratio and IL-6. At doses of 10–50 mg/ kg and>200 mg/kg, emodin efectively improved MPO. However, there was no signifcant improvement in MPO at the dose of 50–200 mg/kg. Additionally, MPO was markedly improved in two studies within the group of 50–200 mg/kg [\[20](#page-10-19), [24\]](#page-10-28), but the detection specimens were

lavage fuid and lung tissue, with large heterogeneity, which may lead to insignifcant combined results. Emodin at>200 mg/kg had no signifcant improvement in IL-1β. However, two studies within the group [\[26,](#page-10-24) [34\]](#page-11-0) showed signifcant improvement efects on IL-1β. Moreover, there were diferences in animal species, ALI modeling method, emodin treatment time point, and total emodin dose. High heterogeneity may lead to insignifcant combined results. This conclusion should be interpreted with caution. At doses of 10–50 mg/kg and 50–200 mg/kg, emodin effectively improved $PaO₂$ and $PaCO₂$. In addition, emodin at<10 mg/kg had no signifcant improvement in IL-1β and MPO. The above studies suggest a certain correlation between the efect size of emodin treatment and the total dose of emodin. Previous studies have shown that within a certain dose range, emodin dose-dependently inhibits lung infammation in LPS-stimulated ARDS mice to improve lung injury $[6]$. The total dose of emodin was $5-700$ mg/kg in the included studies. The groups with 50–200 mg/kg and>200 mg/kg of emodin showed signifcant results. However, fewer studies chose these doses, and most studies chose $10-50$ mg/kg. Therefore, we currently cannot determine the more appropriate emodin dose. To date, there is a lack of clinical trials on the efect of emodin on ALI/ARDS. Adequate time and research are essential for the clinical application of emodin in ALI/ ARDS. Consequently, additional studies are warranted to ascertain the time, frequency, administration route, and dose of emodin.

Animal detection specimens

Stratifed analysis based on lung tissue, serum, and lavage fluid specimens all confirmed the efficacy of emodin in acute lung injury. The degree of improvement in MPO activity in lung tissue specimens was superior to that in lavage fuid; the serum specimens showed a greater improvement in TNF-α levels compared to lavage fuid and lung tissue; in terms of the detection of IL-1β, IL-6, IL-18, serum specimens outperformed lung tissue, which in turn was better than lavage fluid. Lung tissue and serum may more sensitively refect changes in these biomarkers, suggesting the need for further research to explore which type of specimen exhibits higher sensitivity for the detection of specifc indicators.

Advantages and limitations

First, we endeavored to gather an extensive range of research in this feld to obtain complete data; second, data extraction and evaluation were undertaken by two reviewers independently. Any disputes were addressed through consultation with a third reviewer. The metaanalysis of the diferences in relevant articles confrmed the therapeutic efect of emodin in animal models of ALI. Sensitivity analysis confrmed stable results on lung injury score, W/D ratio, TNF-α, MPO, IL-1β, IL-6, IL-18, PaO₂ and PaCO₂. These findings signal that emodin has a certain positive efect on the treatment of ALI and is expected to provide a new treatment strategy for clinical ALI patients.

However, we only included studies published in English and already published studies, which omitted studies published in other languages and unpublished studies with negative results, which may exaggerate the effect. In addition, the published studies included were limited and presented with high heterogeneity, possibly due to insufficient sample size. Thereby, sufficient evidence is needed in future studies with large sample sizes.

Conclusion

Our meta-analysis confrmed that emodin treatment could improve the pathological conditions, pulmonary edema, and inflammation in animal models of ALI. These results offer valuable insights for future preclinical and clinical research to a certain extent and hold signifcant implications for human health.

Abbreviations

Supplementary Information

The online version contains supplementary material available at [https://doi.](https://doi.org/10.1186/s12890-024-03406-x) [org/10.1186/s12890-024-03406-x.](https://doi.org/10.1186/s12890-024-03406-x)

Supplementary Material 1: Table S1 Stratifed meta-analysis of lung injury score. Table S2 Stratifed meta-analysis of lung wet-dry weight ratio. Table S3 Stratifed meta-analysis of MPO. Table S4 Stratifed meta-analysis of TNF-α. Table S5 Stratifed meta-analysis of IL−1β. Table S6 Stratifed meta-analysis of IL−6. Table S7 Stratifed meta-analysis of IL−18. Table S8 Stratified meta-analysis of PaO₂. Table S9 Stratified meta-analysis of PaCO₂. Table S10 Publication bias evaluated used by Egger's.

Supplementary Material 2: Figure S1 (A) Trim-and-fll analysis of emodin treatment of lung injury score. (B)Trim-and-fll analysis of emodin treatment of W/D ratio. (C) Trim-and-fll analysis of emodin treatment of MPO. (D) Trim-and-fll analysis of emodin treatment of TNF−α. (E) Trim-and-fll analysis of emodin treatment of IL−1β. (F) Trim-and-fll analysis of emodin treatment of IL−6.

Supplementary Material 3.

Acknowledgements

Not applicable.

Authors' contributions

Writing—original draft preparation: [Yu Zhang]; Writing—review and editing: [Lei Liu]; Conceptualization: [Shan Zhou]; Methodology: [Xiao-Ren Tang]; Formal analysis and investigation: [Guo-Bing Jia and Guo-Long Yue]; Funding acquisition: [Cheng-Shi He and Lei Liu]; Resources: [Lei Liu]; Supervision: [Cheng-Shi He],and all authors commented on previous versions of the manuscript. All authors read and approved the fnal manuscript.

Funding

This work was supported by (1) Chengdu Traditional Chinese Medicine Administration Project (H2023086); (2) Chengdu University of Traditional Chinese Medicine "Xinglin Scholar" Hospital Special (YYZX2021052).

Data availability

All data generated or analyzed during this study are included in this published article (and its Supplementary Information fles).

Declarations

Ethics approval and consent to participate

All analyses were based on previous published studies, thus no ethical approval and patient consent are required.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

Received: 3 April 2024 Accepted: 21 November 2024

References

- 1. Meyer NJ, Gattinoni L, Calfee CS. Acute respiratory distress syndrome. Lancet (London, England). 2021;398(10300):622–37. [https://doi.org/10.](https://doi.org/10.1016/s0140-6736(21)00439-6) [1016/s0140-6736\(21\)00439-6.](https://doi.org/10.1016/s0140-6736(21)00439-6)
- 2. Rice TW. Obesity in acute lung injury: The "weight" is over. Chest. 2007;131(2):333–4. <https://doi.org/10.1378/chest.06-2584>.
- 3. Mowery NT, Terzian WTH, Nelson AC. Acute lung injury. Curr Probl Surg. 2020;57(5):100777. <https://doi.org/10.1016/j.cpsurg.2020.100777>.
- 4. Bellani G, Lafey JG, Pham T, Fan E, Brochard L, Esteban A, Gattinoni L, van Haren F, Larsson A, McAuley DF, et al. Epidemiology, Patterns of Care, and Mortality for Patients With Acute Respiratory Distress Syndrome in Intensive Care Units in 50 Countries. JAMA. 2016;315(8):788–800. [https://](https://doi.org/10.1001/jama.2016.0291) [doi.org/10.1001/jama.2016.0291.](https://doi.org/10.1001/jama.2016.0291)
- 5. Guo R, Li Y, Han M, Liu J, Sun Y. Emodin attenuates acute lung injury in Cecal-ligation and puncture rats. Int Immunopharmacol. 2020;85:106626. <https://doi.org/10.1016/j.intimp.2020.106626>.
- 6. Liu B, Cheng Y, Wu Y, Zheng X, Li X, Yang G, He T, Li S, Shen F. Emodin improves alveolar hypercoagulation and inhibits pulmonary infammation in LPS-provoked ARDS in mice via NF-κB inactivation. Int Immunopharmacol. 2020;88:107020. [https://doi.org/10.1016/j.intimp.2020.107020.](https://doi.org/10.1016/j.intimp.2020.107020)
- 7. Wang WB, Li JT, Hui Y, Shi J, Wang XY, Yan SG. Combination of pseudoephedrine and emodin ameliorates LPS-induced acute lung injury by regulating macrophage M1/M2 polarization through the VIP/cAMP/ PKA pathway. Chinese medicine. 2022;17(1):19. [https://doi.org/10.1186/](https://doi.org/10.1186/s13020-021-00562-8) [s13020-021-00562-8](https://doi.org/10.1186/s13020-021-00562-8).
- 8. Mei H, Tao Y, Zhang T, Qi F. Emodin alleviates LPS-induced inflammatory response in lung injury rat by afecting the function of granulocytes. Journal of infammation (London, England). 2020;17:26. [https://doi.org/](https://doi.org/10.1186/s12950-020-00252-6) [10.1186/s12950-020-00252-6](https://doi.org/10.1186/s12950-020-00252-6).
- 9. Gao Z, Sui J, Fan R, Qu W, Dong X, Sun D. Emodin Protects Against Acute Pancreatitis-Associated Lung Injury by Inhibiting NLPR3 Infammasome Activation via Nrf2/HO-1 Signaling. Drug Des Dev Ther. 2020;14:1971–82. [https://doi.org/10.2147/dddt.S247103.](https://doi.org/10.2147/dddt.S247103)
- 10. Jin T, Ai F, Zhou J, Kong L, Xiong Z, Wang D, Lu R, Chen Z, Zhang M. Emodin alleviates lung ischemia-reperfusion injury by suppressing gasdermin D-mediated pyroptosis in rats. Clin Respir J. 2023;17(3):241–50. [https://](https://doi.org/10.1111/crj.13582) doi.org/10.1111/crj.13582.
- 11. Huang Q, Lu G, Shen HM, Chung MC, Ong CN. Anti-cancer properties of anthraquinones from rhubarb. Med Res Rev. 2007;27(5):609–30. [https://](https://doi.org/10.1002/med.20094) [doi.org/10.1002/med.20094.](https://doi.org/10.1002/med.20094)
- 12. Li L, Song X, Yin Z, Jia R, Li Z, Zhou X, Zou Y, Li L, Yin L, Yue G, et al. The antibacterial activity and action mechanism of emodin from Polygonum

cuspidatum against Haemophilus parasuis in vitro. Microbiol Res. 2016;186–187:139–45. [https://doi.org/10.1016/j.micres.2016.03.008.](https://doi.org/10.1016/j.micres.2016.03.008)

- 13. Dong X, Fu J, Yin X, Cao S, Li X, Lin L, Ni J. Emodin: A Review of its Pharmacology, Toxicity and Pharmacokinetics. Phytotherapy research : PTR. 2016;30(8):1207–18. [https://doi.org/10.1002/ptr.5631.](https://doi.org/10.1002/ptr.5631)
- 14. Tu C, Gao D, Li XF, Li CY, Li RS, Zhao YL, Li N, Jia GL, Pang JY, Cui HR, et al. Infammatory stress potentiates emodin-induced liver injury in rats. Front Pharmacol. 2015;6:233.<https://doi.org/10.3389/fphar.2015.00233>.
- 15. Tian SL, Yang Y, Liu XL, Xu QB. Emodin Attenuates Bleomycin-Induced Pulmonary Fibrosis via Anti-Infammatory and Anti-Oxidative Activities in Rats. Medical science monitor : international medical journal of experimental and clinical research. 2018;24:1–10. [https://doi.org/10.12659/](https://doi.org/10.12659/msm.905496) [msm.905496](https://doi.org/10.12659/msm.905496).
- 16. Wei WT, Lin SZ, Liu DL, Wang ZH. The distinct mechanisms of the antitumor activity of emodin in diferent types of cancer (Review). Oncol Rep. 2013;30(6):2555–62. <https://doi.org/10.3892/or.2013.2741>.
- 17. Dey D, Ray R, Hazra B. Antitubercular and antibacterial activity of quinonoid natural products against multi-drug resistant clinical isolates. Phytotherapy research : PTR. 2014;28(7):1014–21. <https://doi.org/10.1002/ptr.5090>.
- 18. Schwarz S, Wang K, Yu W, Sun B, Schwarz W. Emodin inhibits current through SARS-associated coronavirus 3a protein. Antiviral Res. 2011;90(1):64–9. [https://doi.org/10.1016/j.antiviral.2011.02.008.](https://doi.org/10.1016/j.antiviral.2011.02.008)
- 19. Hu Q, Yao J, Wu X, Li J, Li G, Tang W, Liu J, Wan M. Emodin attenuates severe acute pancreatitis-associated acute lung injury by suppressing pancreatic exosome-mediated alveolar macrophage activation. Acta pharmaceutica Sinica B. 2022;12(10):3986–4003. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.apsb.2021.10.008) apsh.2021.10.008
- 20. Liu FJ, Gu TJ, Wei DY. Emodin alleviates sepsis-mediated lung injury via inhibition and reduction of NF-kB and HMGB1 pathways mediated by SIRT1. Kaohsiung J Med Sci. 2022;38(3):253–60. [https://doi.org/10.1002/](https://doi.org/10.1002/kjm2.12476) kim2.12476
- 21. Moher D, Liberati A, Tetzlaff J, Altman DG. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. PLoS Med. 2009;6(7):e1000097.<https://doi.org/10.1371/journal.pmed.1000097>.
- 22. Hooijmans CR, Rovers MM, de Vries RB, Leenaars M, Ritskes-Hoitinga M, Langendam MW. SYRCLE's risk of bias tool for animal studies. BMC Med Res Methodol. 2014;14:43. <https://doi.org/10.1186/1471-2288-14-43>.
- 23. Jiang N, Li Z, Luo Y, Jiang L, Zhang G, Yang Q, Chen H. Emodin ameliorates acute pancreatitis-induced lung injury by suppressing NLRP3 infammasome-mediated neutrophil recruitment. Exp Ther Med. 2021;22(2):857. <https://doi.org/10.3892/etm.2021.10289>.
- 24. Liu Y, Shang L, Zhou J, Pan G, Zhou F, Yang S. Emodin Attenuates LPS-Induced Acute Lung Injury by Inhibiting NLRP3 Infammasome-Dependent Pyroptosis Signaling Pathway In vitro and In vivo. Infammation. 2022;45(2):753–67. [https://doi.org/10.1007/s10753-021-01581-1.](https://doi.org/10.1007/s10753-021-01581-1)
- 25. Wu X, Yao J, Hu Q, Kang H, Miao Y, Zhu L, Li C, Zhao X, Li J, Wan M, et al. Emodin Ameliorates Acute Pancreatitis-Associated Lung Injury Through Inhibiting the Alveolar Macrophages Pyroptosis. Front Pharmacol. 2022;13:873053. [https://doi.org/10.3389/fphar.2022.873053.](https://doi.org/10.3389/fphar.2022.873053)
- 26. Xiao M, Zhu T, Zhang W, Wang T, Shen YC, Wan QF, Wen FQ. Emodin ameliorates LPS-induced acute lung injury, involving the inactivation of NF-κB in mice. Int J Mol Sci. 2014;15(11):19355–68. [https://doi.org/10.3390/ijms1](https://doi.org/10.3390/ijms151119355) [51119355](https://doi.org/10.3390/ijms151119355).
- 27. Xie P, Yan LJ, Zhou HL, Cao HH, Zheng YR, Lu ZB, Yang HY, Ma JM, Chen YY, Huo C, et al. Emodin Protects Against Lipopolysaccharide-Induced Acute Lung Injury via the JNK/Nur77/c-Jun Signaling Pathway. Front Pharmacol. 2022;13:717271. [https://doi.org/10.3389/fphar.2022.717271.](https://doi.org/10.3389/fphar.2022.717271)
- 28. Dong Y, Zhang L, Jiang Y, Dai J, Tang L, Liu G. Emodin reactivated autophagy and alleviated infammatory lung injury in mice with lethal endotoxemia. Exp Anim. 2019;68(4):559–68. [https://doi.org/10.1538/](https://doi.org/10.1538/expanim.19-0004) [expanim.19-0004](https://doi.org/10.1538/expanim.19-0004).
- 29. Cui H, Li S, Xu C, Zhang J, Sun Z, Chen H. Emodin alleviates severe acute pancreatitis-associated acute lung injury by decreasing pre-B-cell colony-enhancing factor expression and promoting polymorphonuclear neutrophil apoptosis. Mol Med Rep. 2017;16(4):5121–8. [https://doi.org/](https://doi.org/10.3892/mmr.2017.7259) [10.3892/mmr.2017.7259.](https://doi.org/10.3892/mmr.2017.7259)
- 30. Xu J, Huang B, Wang Y, Tong C, Xie P, Fan R, Gao Z. Emodin ameliorates acute lung injury induced by severe acute pancreatitis through the up-regulated expressions of AQP1 and AQP5 in lung. Clin Exp Pharmacol Physiol. 2016;43(11):1071–9.<https://doi.org/10.1111/1440-1681.12627>.
- 31. Xu C, Zhang J, Liu J, Li Z, Liu Z, Luo Y, Xu Q, Wang M, Zhang G, Wang F, et al. Proteomic analysis reveals the protective effects of emodin on severe acute pancreatitis induced lung injury by inhibiting neutrophil proteases activity. J Proteomics. 2020;220:103760. [https://doi.org/10.](https://doi.org/10.1016/j.jprot.2020.103760) [1016/j.jprot.2020.103760](https://doi.org/10.1016/j.jprot.2020.103760).
- 32. Xu C, Luo Y, Ntim M, Quan W, Li Z, Xu Q, Jiang L, Zhang J, Shang D, Li L, et al. Efect of emodin on long non-coding RNA-mRNA networks in rats with severe acute pancreatitis-induced acute lung injury. J Cell Mol Med. 2021;25(4):1851–66. <https://doi.org/10.1111/jcmm.15525>.
- 33. Xu Q, Wang M, Guo H, Liu H, Zhang G, Xu C, Chen H. Emodin Alleviates Severe Acute Pancreatitis-Associated Acute Lung Injury by Inhibiting the Cold-Inducible RNA-Binding Protein (CIRP)-Mediated Activation of the NLRP3/IL-1β/CXCL1 Signaling. Front Pharmacol. 2021;12:655372. [https://](https://doi.org/10.3389/fphar.2021.655372) [doi.org/10.3389/fphar.2021.655372.](https://doi.org/10.3389/fphar.2021.655372)
- 34. Li X, Shan C, Wu Z, Yu H, Yang A, Tan B. Emodin alleviated pulmonary infammation in rats with LPS-induced acute lung injury through inhibiting the mTOR/HIF-1α/VEGF signaling pathway. Infammation research : official journal of the European Histamine Research Society [et al]. 2020;69(4):365–73. <https://doi.org/10.1007/s00011-020-01331-3>.
- 35. Wang F, Fang B, Qiang X, Shao J, Zhou L. The efficacy of mesenchymal stromal cell-derived therapies for acute respiratory distress syndrome-a meta-analysis of preclinical trials. Respir Res. 2020;21(1):307. [https://doi.](https://doi.org/10.1186/s12931-020-01574-y) [org/10.1186/s12931-020-01574-y.](https://doi.org/10.1186/s12931-020-01574-y)
- 36. Chen G, Zhang J, Zhang H, Xiao Y, Kao X, Liu Y, Liu Z. Anti-infammatory efect of emodin on lipopolysaccharide-induced keratitis in Wistar rats. Int J Clin Exp Med. 2015;8(8):12382–9.
- 37. Li Y, Xiong W, Yang J, Zhong J, Zhang L, Zheng J, Liu H, Zhang Q, Ouyang X, Lei L, et al. Attenuation of Infammation by Emodin in Lipopolysaccharide-induced Acute Kidney Injury via Inhibition of Toll-like Receptor 2 Signal Pathway. Iran J Kidney Dis. 2015;9(3):202–8.
- 38. Yin X, Gong X, Jiang R, Kuang G, Wang B, Zhang L, Xu G, Wan J. Emodin ameliorated lipopolysaccharide-induced fulminant hepatic failure by blockade of TLR4/MD2 complex expression in D-galactosamine-sensitized mice. Int Immunopharmacol. 2014;23(1):66–72. [https://doi.org/10.](https://doi.org/10.1016/j.intimp.2014.08.018) [1016/j.intimp.2014.08.018](https://doi.org/10.1016/j.intimp.2014.08.018).
- 39. Kulkarni HS, Lee JS, Bastarache JA, Kuebler WM, Downey GP, Albaiceta GM, Altemeier WA, Artigas A, Bates JHT, Calfee CS, et al. Update on the Features and Measurements of Experimental Acute Lung Injury in Animals: An Official American Thoracic Society Workshop Report. Am J Respir Cell Mol Biol. 2022;66(2):e1–14. [https://doi.org/10.1165/rcmb.](https://doi.org/10.1165/rcmb.2021-0531ST) [2021-0531ST.](https://doi.org/10.1165/rcmb.2021-0531ST)
- 40. Xiping Z, Jun F, Jie Z, Bingyan Y, Jing M, Wei Z, Jing Y, Penghui J, Wenqin Y, Ninnin Z, et al. Infuence of dexamethasone on the expression levels of P-selectin protein in multiple organs of rats with severe acute pancreatitis. Inflammation research : official journal of the European Histamine Research Society [et al]. 2010;59(1):31–9. [https://doi.org/10.1007/](https://doi.org/10.1007/s00011-009-0067-x) [s00011-009-0067-x](https://doi.org/10.1007/s00011-009-0067-x).
- 41. Ye W, Zheng C, Yu D, Zhang F, Pan R, Ni X, Shi Z, Zhang Z, Xiang Y, Sun H, et al. Lipoxin A4 Ameliorates Acute Pancreatitis-Associated Acute Lung Injury through the Antioxidative and Anti-Infammatory Efects of the Nrf2 Pathway. Oxid Med Cell Longev. 2019;2019:2197017. [https://doi.org/](https://doi.org/10.1155/2019/2197017) [10.1155/2019/2197017](https://doi.org/10.1155/2019/2197017).
- 42. Hybertson BM, Lee YM, Cho HG, Cho OJ, Repine JE. Alveolar type II cell abnormalities and peroxide formation in lungs of rats given IL-1 intratracheally. Infammation. 2000;24(4):289–303. [https://doi.org/10.1023/a:](https://doi.org/10.1023/a:1007092529261) [1007092529261](https://doi.org/10.1023/a:1007092529261).
- 43. Klebanoff SJ. Myeloperoxidase: friend and foe. J Leukoc Biol. 2005;77(5):598–625. [https://doi.org/10.1189/jlb.1204697.](https://doi.org/10.1189/jlb.1204697)
- 44. Kisic B, Miric D, Dragojevic I, Rasic J, Popovic L. Role of Myeloperoxidase in Patients with Chronic Kidney Disease. Oxid Med Cell Longev. 2016;2016:1069743. [https://doi.org/10.1155/2016/1069743.](https://doi.org/10.1155/2016/1069743)
- 45. Zhu T, Wang DX, Zhang W, Liao XQ, Guan X, Bo H, Sun JY, Huang NW, He J, Zhang YK, et al. Andrographolide protects against LPS-induced acute lung injury by inactivation of NF-κB. PLoS ONE. 2013;8(2):e56407. [https://](https://doi.org/10.1371/journal.pone.0056407) doi.org/10.1371/journal.pone.0056407.
- 46. Goodman RB, Pugin J, Lee JS, Matthay MA. Cytokine-mediated infammation in acute lung injury. Cytokine Growth Factor Rev. 2003;14(6):523–35. [https://doi.org/10.1016/s1359-6101\(03\)00059-5](https://doi.org/10.1016/s1359-6101(03)00059-5).
- 47. Makabe H, Kojika M, Takahashi G, Matsumoto N, Shibata S, Suzuki Y, Inoue Y, Endo S. Interleukin-18 levels refect the long-term prognosis

of acute lung injury and acute respiratory distress syndrome. J Anesth. 2012;26(5):658–63. [https://doi.org/10.1007/s00540-012-1409-3.](https://doi.org/10.1007/s00540-012-1409-3)

- 48. Ferguson ND, Fan E, Camporota L, Antonelli M, Anzueto A, Beale R, Brochard L, Brower R, Esteban A, Gattinoni L, et al. The Berlin defnition of ARDS: an expanded rationale, justifcation, and supplementary material. Intensive Care Med. 2012;38(10):1573–82. [https://doi.org/10.1007/](https://doi.org/10.1007/s00134-012-2682-1) [s00134-012-2682-1.](https://doi.org/10.1007/s00134-012-2682-1)
- 49. Grasselli G, Calfee CS, Camporota L, Poole D, Amato MBP, Antonelli M, Arabi YM, Baroncelli F, Beitler JR, Bellani G, et al. ESICM guidelines on acute respiratory distress syndrome: defnition, phenotyping and respiratory support strategies. Intensive Care Med. 2023;49(7):727–59. [https://](https://doi.org/10.1007/s00134-023-07050-7) doi.org/10.1007/s00134-023-07050-7.
- 50. Matute-Bello G, Frevert CW, Martin TR. Animal models of acute lung injury. Am J Physiol Lung Cell Mol Physiol. 2008;295(3):L379-399. [https://](https://doi.org/10.1152/ajplung.00010.2008) [doi.org/10.1152/ajplung.00010.2008.](https://doi.org/10.1152/ajplung.00010.2008)
- 51. Chimenti L, Morales-Quinteros L, Puig F, Camprubi-Rimblas M, Guillamat-Prats R, Gómez MN, Tijero J, Blanch L, Matute-Bello G, Artigas A. Comparison of direct and indirect models of early induced acute lung injury. Intensive Care Med Exp. 2020;8(Suppl 1):62. [https://doi.org/10.1186/](https://doi.org/10.1186/s40635-020-00350-y) [s40635-020-00350-y.](https://doi.org/10.1186/s40635-020-00350-y)

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional afliations.