

Cerebro-Cerebellar Networks in Migraine Symptoms and Headache

Rodrigo Noseda 1,2*

¹ Department of Anesthesia, Critical Care and Pain Medicine, Beth Israel Deaconess Medical Center, Boston, MA, United States, ² Harvard Medical School, Boston, MA, United States

The cerebellum is associated with the biology of migraine in a variety of ways. Clinically, symptoms such as fatigue, motor weakness, vertigo, dizziness, difficulty concentrating and finding words, nausea, and visual disturbances are common in different types of migraine. The neural basis of these symptoms is complex, not completely known, and likely involve activation of both specific and shared circuits throughout the brain. Posterior circulation stroke, or neurosurgical removal of posterior fossa tumors, as well as anatomical tract tracing in animals, provided the first insights to theorize about cerebellar functions. Nowadays, with the addition of functional imaging, much progress has been done on cerebellar structure and function in health and disease, and, as a consequence, the theories refined. Accordingly, the cerebellum may be useful but not necessary for the execution of motor, sensory or cognitive tasks, but, rather, would participate as an efficiency facilitator of neurologic functions by improving speed and skill in performance of tasks produced by the cerebral area to which it is reciprocally connected. At the subcortical level, critical regions in these processes are the basal ganglia and thalamic nuclei. Altogether, a modulatory role of the cerebellum over multiple brain regions appears compelling, mainly by considering the complexity of its reciprocal connections to common neural networks involved in motor, vestibular, cognitive, affective, sensory, and autonomic processing-all functions affected at different phases and degrees across the migraine spectrum. Despite the many associations between cerebellum and migraine, it is not known whether this structure contributes to migraine initiation, symptoms generation or headache. Specific cerebellar dysfunction via genetically driven excitatory/inhibitory imbalances, oligemia and/or increased risk to white matter lesions has been proposed as a critical contributor to migraine pathogenesis. Therefore, given that neural projections and functions of many brainstem, midbrain and forebrain areas are shared between the cerebellum and migraine trigeminovascular pathways, this review will provide a synopsis on cerebellar structure and function, its role in trigeminal pain, and an updated overview of relevant clinical and preclinical literature on the potential role of cerebellar networks in migraine pathophysiology.

Keywords: pain, cognition, hypothalamus, basal ganglia, thalamus

OPEN ACCESS

Edited by:

Simon Akerman, University of Maryland, Baltimore, United States

Reviewed by:

Roberta Messina, Vita-Salute San Raffaele University, Italy Kuan-Lin Lai, Taipei Veterans General Hospital, Taiwan

*Correspondence: Rodrigo Noseda rnoseda@bidmc.harvard.edu

Specialty section:

This article was submitted to Headache, a section of the journal Frontiers in Pain Research

Received: 10 May 2022 **Accepted:** 13 June 2022 **Published:** 13 July 2022

Citation:

Noseda R (2022) Cerebro-Cerebellar Networks in Migraine Symptoms and Headache. Front. Pain Res. 3:940923. doi: 10.3389/fpain.2022.940923

INTRODUCTION

Migraine is a complex, cyclic, and multi-symptomatic chronic disorder characterized by episodic manifestations, "the attack." During a migraine attack (i.e., the ictal period), headache is the most burdensome symptom, ranging from a moderate to severe unilateral throbbing pain, usually in the periocular area of the face (1). As part of the diagnostic criteria, migraine headache is usually accompanied by nausea and/or vomiting, increased sensitivity to light, sound and smell, and intensification by routine physical activity (2). This phase is thought to originate in either extra- and/or intra-cranial tissues, but particularly within the dura and its vasculature, known today to be densely innervated by the maxillary and ophthalmic branches of the trigeminal nerve, as well as considerable innervation from upper cervical spinal nerves C2-C3 (3-7) (Figure 1A). Neuronal cell bodies in the trigeminal (TG) and upper cervical ganglia (C2/3DRG) and axonal fibers of this innervation express CGRP and its receptors (8, 9), for which current therapies blocking their pathways are proving effective at significantly reducing migraine and allodynia in nearly half of patients (10, 11). Indeed, monoclonal antibodies against CGRP inhibit meningeal nociception in rodents and appear to act mainly in the peripheral nervous system (12, 13), suggesting, in most cases, a peripheral origin for head pain (5).

The nociceptive information from the intracranial meninges is transmitted to second-order neurons in the brainstem trigeminocervical complex, where it is further processed and sent upstream to a multitude of brain areas, collectively known as the ascending central trigeminovascular pathway, where information reaches many subcortical regions and thalamic nuclei, and then relayed to a variety of cortical areas for processing and perception of multi-symptoms and headache (1, 3, 7, 14-16) (Figure 1B). However, the fact that patients can usually identify the signs of an upcoming attack suggests the presence of an internal process slowly recruiting neuronal/glial signaling cascades from a multitude of brain regions, which may eventually lead to a moderate or severe headache. Accordingly, the headache may often be preceded or accompanied by visual or sensory auras in a third of patients-events widely accepted to originate in the cerebral cortex due to excitability imbalances and neurovascular changes, generally referred to as spreading depolarizations (17, 18). Migraine may also be preceded by premonitory symptoms, or prodrome, in nearly half of patients. Among the most frequent, fatigue, difficulty at concentrating, and neck pain or tenderness are the most significant (19-22). There is, however, a whole array of mild manifestations in addition to sensory allodynias that have been identified between attacks (i.e., the interictal period) throughout the duration of the migraine cycle (23), all of which can be attributed to neural networks connecting specific regions in the brain, such as the thalamus and cerebral cortex, basal ganglia, hypothalamus, brainstem, and cerebellum. Along this line, migraine disorder can be better understood as a network disorder potentially leading to neuro-inflammation and meningeal pain (1, 18, 24–26).

In this context, the prodrome phase starts about a day before the headache and may be characterized by a wealth

of clinical signs and symptoms, suggesting a central origin of the disorder (19, 22). Along this line, a unique and highly significant series of imaging studies in the human, aiming at deciphering functional interactions in the brain across the migraine cycle, have shown widespread alterations in excitability behavior of brain networks. The most widely recognized is the altered excitability balance in the cerebral cortex and pain processing across the trigeminal system (27-29). However, dysexcitability is not exclusive to migraine, but other chronic pain conditions, such as fibromyalgia and low back pain, may share similar electrophysiological features (28, 30, 31). Given that neural excitability is dynamic and associated with a variety of factors, current evidence suggests that some of these variations might be associated with hypothalamic activity, in which alterations in functional coupling to spinal trigeminal nuclei and dorsal rostral pons are significantly increased in the prodromal phase. During the headache, the hypothalamus increases functional connectivity with the dorsal rostral ponscontaining the locus coeruleus, parabrachial, cuneiform, and vestibular nuclei, among others (32). These findings have led to the interesting proposal that the functional changes in hypothalamic-brainstem connectivity may be the real driver or generator of attacks (33, 34). More specifically, these data showed that variations in the anterior hypothalamic areas are linked to initiation and chronification of migraine, whereas those in more posterior parts of the hypothalamus seem to be involved in the headache phase. The latest report using longitudinal fMRI, showed that cyclic changes of brain perfusion in hypothalamic and limbic regions (insula, accumbens, hippocampus), with the highest perfusion during the headache attack, may be linked to specific neural dynamics throughout the migraine cycle (35). Interestingly, there was a progressive increase of functional connectivity among these structures, reaching a peak before collapsing at or the near headache onset, in a sort of sudden meltdown between the hypothalamus and the limbic brain (36). Similarly, a progressive increase before headache and a sudden drop of functional connectivity of the hypothalamus at the headache onset was found for sensory and frontal cortices, basal ganglia, and cerebellum (36). In another recent study, interictal assessment of episodic migraineurs has found alterations in dynamic functional coupling between the hypothalamus and brain regions, processing pain and vision, as well as high-order sensations, representing clinical features, such as disease duration and disability in the orbitofrontal gyrus of the prefrontal cortex (37). In a resting-state analysis, decreased functional connectivity between the hypothalamus and medial prefrontal cortex was also observed, suggesting hypothalamic network involvement for pain perception and severity during attacks in chronic migraine (38, 39). These aversive sensations (i.e., pain, photophobia, nausea, etc.) can be exacerbated by ambient illumination through hypothalamic regulation of cranial and systemic autonomic outflow during both ictal and interictal migraine (40). These studies provide compelling evidence, adding to existing research on hypothalamic functional connectivity with autonomic, limbic, and cerebellar regions. Altogether, these studies seem to support the "reset mode" theory of an overloaded brain in migraine. Main



output pathways from hypothalamic nuclei are represented in Figure 2.

on these brain dynamics, migraine varied Based symptomatology and emerging subclinical evidence in patients; the role of the cerebellum has become highly relevant in the last two decades (41, 42). Moreover, the view of the cerebellum has significantly evolved and is now considered a highly organized modular structure strongly associated with pain-related cognitive, autonomic, and somato-visceral regions in the brain, including specific reciprocal connections to trigeminal pain pathways, while also sharing multiple global neural networks involved in motor, vestibular, cognitive, affective, sensory, and autonomic functions. Additional functional ties are clear when we look at imaging studies, showing cerebellar activity during acute and chronic pain conditions, and in association with the emotional, cognitive, and motor responses to pain (43, 44). In addition, multiple types of sensory information going to the cerebellum-as recorded in the mouse (45) and mapped in the human (46, 47)-demonstrate its involvement in multisensory processing, likely for efficient control of visuomotor tasks, auditory, and visual sensory acquisition, visually guided eye movements and motion perception, among other sensory-associated functions (48-51).

On the pathological side, there is a wealth of clinical and subclinical signs that link the cerebellum with different migraine

phenotypes. The most characteristic is Familial Hemiplegic Migraine (FHM)-a rare monogenic migraine disorder with aura, and with a set of severe motor, speech, cognitive, sensory, and affective symptoms that may occur spontaneously (52-55). Across the spectrum, vestibular migraine (VM) is the most closely associated with the vestibulo-cerebellar system, usually presenting with vertigo, dizziness, motor clumsiness, fatigue, neck tenderness, lack of concentration, nausea, and vomiting (56). Despite appearing milder in intensity than in VM, cerebellar manifestations in migraine with or without aura are not uncommon (57-60). Indeed, imaging data in interictal migraine showed altered functional connectivity between the hypothalamus and subcortical areas, such as the locus coeruleus, pontine nuclei, caudate, cerebellar Crus I and II, temporal lobe, and hippocampal regions, suggesting a stronger influence on the autonomic nervous system in patients with migraine (61). Another study also found ictal vs. interictal differences in the activity of brainstem/pons, cerebellum, thalamus, insular, and cingulate cortices in association with allodynia (62). Interestingly, imaging studies have also shown increased prevalence of white mater lesions (63-65), as well as functional and structural alterations in the cerebellum of patients with migraine (66) in association with a prognosis and chronification of migraine (67).

Despite the progress, many questions still remain on how an overload may occur, under what circumstances, and the



role of the other brain structures in the development of 'reset mode' conditions observed in the above studies, as well as its relationship with neurovascular events and spreading depolarization phenomena in the cerebral cortex. In addition, the cerebellum can also experience similar hypoperfusion and vascular alterations in migraine (68); however, there is, virtually, no information about SD-like events occurring in the cerebellum, and how these events could be related to migraine headache or its preceding phases. Certainly, the cerebellum is connected to a variety of neural systems (69), and is not foreign to the genetics of migraine. Thus, it would be critical to understand why expression of a variety of receptors and neuropeptides is higher in the cerebellum and how this pattern differs across the CNS in health and disease. Also relevant is how biomarkers' expression is related to the diversity in migraine phenotype and how the modulatory role of the cerebellum could be playing in migraine-related circuits. The potential of the cerebellum for therapeutical neurostimulation has also emerged and is rapidly growing (70). Therefore, given the renewed clinical and scientific interest in understanding the cerebellum, this review focuses on conceptual advances in cerebellar connectome and migraine neurobiology.

FUNCTIONAL ARCHITECTURE OF THE CEREBELLUM

A plethora of new information and literature reviews on cerebellar structure and function has emerged in the last decade, reporting extensively on the underpinnings of the cerebellum and its relationship with the rest of the brain (46, 71–73). Integrating this information into the migraine lexicon is critical to improve our understanding of cerebellar structure and function, both in health and in this complex disorder. Here, I provide with a brief overview on the functional anatomy of this highly organized local circuitry, the neuronal input it receives from many regions of the brain and spinal cord *via* brainstem inferior olive (IO), vestibular (VN) and pontine nuclei (PN), and the output it sends back to the brain *via* Purkinje cells (PC) projections to deep cerebellar nuclei (DCN) and VN in the so-called cortico-nuclear connection.

Initial gross anatomical studies and current functional imaging of activity and connectivity of the human cerebellum describe an anterior sensorimotor and a posterior cognitive region (72, 74–77). It is also divided longitudinally into the midline vermis, a thick longitudinal band flanked in both sides by the paravermis, lateral hemispheres, and flocculus, each folded into lobules in lower mammals and smaller folia in humans. Functionally, the anterior lobe (hemispheric lobules I–V) is activated during sensorimotor tasks, including limb movement and visuomotor control (78). The posterior lobe (Crus I and II, and hemispheric lobules VI–IX) is involved in higher-level tasks, including learning, language, verbal, and auditory working memory, as well as spatial and emotional processing, with a clear lateralization for language in the right and spatial tasks in the left (79, 80). Along the cerebellar midline, the vermis has been associated with emotional processing (81); and, on the most caudal part, the flocculonodular lobe (Lobule X) for balance or equilibrium, as well as oculomotor-associated functions, through direct reciprocal connections to vestibular nuclei (72, 82–84).

Intrinsic Organization of the Cerebellum

The intrinsic organization of the cerebellar cortex has a unique hierarchical neuronal structure formed by three cytoarchitectonic and clearly defined layers, namely, (a) Granular cell (GC), the densest excitatory cellular components of the cerebellum, with nearly 99% of the total, and receiving most input from abroad; (b) Purkinje cell (PC), the exclusive output to deep cerebellar nuclei and from there to the rest of the brain (85); and (c) the Molecular, the most superficial layer (ML) containing all the dendritic tree of PC, parallel fibers from GC's running orthogonally in this layer, and the inhibitory interneurons Stellate (SC), Golgi (GoC), and Basket cells (BC). The most characteristic and dense intra-connection is about 100,000 GCs sending excitatory synaptic input via parallel fibers to each PC's dendritic tree (GC-PC) across many millimeters (86). In contrast, only one climbing fiber from IO contacts a single PC (87). Of the many more cells found in the cerebellum, the Bergman glia (BG), Lugaro (LgC), and Unipolar Brush Cells (UBC) have been characterized (88-91), and perform inhibitory actions to help maintain homeostasis of synaptic neurotransmission at multiple levels of the cerebellar cortex.

Altogether, neural computations in the cerebellum are based on four principles: feedforward processing, divergence and convergence, modularity, and plasticity. Accordingly, signal processing in the cerebellum is almost entirely feed forward, meaning signals move through the circuit from input to output, with minimal internal transmission, and using a restricted number of cells. Incoming signals diverge heavily to contact a large population of GCs (92), which, in turn, converge into PCs distal dendrites in the molecular layer *via* parallel fibers.

Each of these input/output circuits will form modules across the cerebellum, known as an olivo-cortico-nuclear module, and it is organized in longitudinal zones defined by strict somatotopic rules for both input from IO and output to DCN from Purkinje cells (93–95), in which the excitatory cerebellar output system and the inhibitory feedback to the inferior olive are controlled simultaneously (96) in a way that each olivary sub-region is under the influence of the cerebellar zone upon which it projects.

Such strict, repetitive, and topographically organized network of cerebellar computations is also subject to neural plasticity as a fundamental physiological property, governed by genetics and shaped by the environment. Indeed, following the same rules, input and output belonging to longitudinal zones (also known as parasagittal bands) of the cerebellar cortex, belong to either zebrin II (+) (or aldolase C positive) or zebrin II (-) bands. Each band is a longitudinally oriented stripe of PCs, expressing similar phenotype and contain other molecular markers along with zebrin I and II, including many from excitatory glutamatergic and inhibitory GABAergic pathways. In addition, other neuropeptides, channels, and receptors are located across the cerebellum (see below).

Axonal Input to the Cerebellum

Generally, the primary input to the cerebellum is via Mossy Fibers (MF) from different parts of the brain and spinal cord directly to pontine nuclei and/or the GC layer. The cortico-ponto-cerebellar pathway is part of mossy fibers' input to GC's, which, in turn, send massive excitatory connections onto a single PC in an extreme case of convergence. Vestibular and spinal information is also transmitted along mossy fibers to the cerebellar cortex via vestibulo-cerebellar and spino-cerebellar (dorsal and ventral) tracts. Incoming signal then propagates in a restricted matter across multiple zones of PCs within the same lobule via long parallel fibers running orthogonally within the cerebellar folium (92). Another, second but equally important direct input, comes from Climbing Fibers (CF), originating exclusively from the IO, and runs through the olivo-cerebellar tract to make direct excitatory connections across the entire dendritic tree of a single PC, in addition to sending collaterals to DCN (46, 94, 97) (Figure 3). These brainstem nuclei, including reticular formation areas and the trigemino-cervical complex processing meningeal nociceptive input, receive dense and somatotopically organized mono-synaptic input from the cerebral cortex across the corticoponto (olivar) and cortico-trigeminal pathways for regulation of motor and trigeminal sensory functions, respectively (98-103) (Figure 4).

Electrophysiological responses of receptive fields have been recorded in microzones of PC activated by CF, and patches of granule cells activated by MF input, respectively. Given PC's morphology and diverse pre-synaptic input, these electrophysiological properties have been identified as "complex" spikes triggered by CF and "simple" spikes driven, in part, by MF-GC inputs, as well as robust intrinsic activity (46, 104, 105). Both of these physiological Purkinje spike types can be modulated by direct vestibular influence through CF (106).

Similar to the thalamus and the cerebral cortex, the cerebellum receives great influence from hypothalamic and brainstem neurochemical systems involved in stress, sleep, feeding, emotion, pain and sensory processing (107–109). Some of these neuropeptides may be contained in common cerebellar afferent MF and CF, but others are contained in *Beaded Fibers* (BF), or third cerebellar afferent fibers, which form diffuse plexuses to contact PCs and other neurons in molecular and granular layers (110–112). Of relevance, identified neurotransmitter and neuropeptidergic systems arise from widespread areas of the hypothalamus but primarily from the lateral, posterior, and dorsal areas; tuberomammillary (113), periventricular zone, dorsomedial, and ventromedial nuclei (114). Accordingly, the neurochemical identity, as well as anatomical and physiological



data, suggests a strong interplay between glutamate and GABA/glycine, and significant influence from extra-cerebellar neuronal systems transporting noradrenaline (NA), dopamine (DA), serotonin (5HT), acetylcholine (Ach), histamine (Hist), and other neuropeptides directly from hypothalamic and brainstem nuclei into the cerebellum (114). Highly relevant to migraine is CGRP expression in the cerebellum, which has been identified in the cytoplasm of olivocerebellar climbing fibers, contacting Purkinje cell bodies and dendrites, whereas CGRP receptor components, calcitonin receptor-like and activity-modifying protein type 1, have been reported on the surface of Purkinje cell bodies, Purkinje cell dendrites and afferent fibers (115-118). Also relevant to migraine is the presence of PACAP in granule cells (119) and the high proportion of calcium channels associated with mutations in FHM (120, 121).

Given that these neuropeptides can be released from MF, GC, or BF sources, which converge onto granule cells for further processing, each molecule released in the cerebellum likely has specific functional consequences. For example, NA input to cerebellar nuclei and cortex regulates PC GABAergic transmission and appears to participate in motor coordination (122, 123). Cholinergic influence is exerted at the level of CF and MF input from brainstem sources, which are different from cerebral cortex sources in the basal ganglia, and are thought to play a role in non-motor cognitive tasks, whereas dysfunctions are involved in cognitive impairments and other neurological disorders, such as cerebellar ataxia, autism, and Parkinson's (124, 125). Dopamine in the cerebellum has also

been implicated in neurological and psychiatric disorders, such as Parkinson's disease, schizophrenia, autism spectrum disorders, and drug addiction, likely through intrinsic reciprocal connections between DCN and cerebellar cortex, as well as extrinsic connectivity to traditional dopaminergic areas of the brain, including the brainstem, midbrain, and striatum (126). Nevertheless, as a general rule, these diverse neuropeptidergic inputs are considered to convey modulatory signals to the cerebellum to set the baseline activity level (110, 127) in cerebellar circuits contributing to motor and cognitive functions.

Similar to spinocerebellar tract carrying somatosensory information from axial and proximal parts of the body to the vermis, trigeminal signals to the cerebellum travel through shared pathways and have been described in detail in both humans and animals (128-132). In the cats and rats, anatomical studies suggest a direct and indirect connection of the trigeminal system with the cerebellum. Terminal axonal labeling from tracing studies described bi-synaptic connections from trigeminal afferents to mesencephalic trigeminal nucleus, which, in turn, projects to cerebellar nuclei (133, 134). Also, anatomical studies on rodents have shown that cerebellar input from the trigeminal nucleus originates more densely from the contralateral subnuclei Interpolaris (Sp5I), moderately from Oralis (Sp5O), and scarcely with fewer neurons in Principalis (Sp5Pr) and Caudalis (Sp5C). These ascending trigeminal pathways contact neurons in the contralateral rostral dorsal accessory (DAO) and adjacent principal olive, while sending few axon collaterals to the PAG and superior colliculus (135–139). This sensory information appears to be tactile for proprioception purposes as well as



nociceptive and is terminally mapped in Crus I and II of the cerebellar cortex, where a representation of oral and facial regions dominates their somatosensory maps (74, 84, 140, 141).

Finally, yet importantly, vestibular input plays arguably the most important functional role in association with the cerebellum. Vestibular nuclei in the dorsal pons are reciprocally connected with the cerebellar nuclei to form the vestibulocerebellar system, which controls balance by influencing the discharge of motor and pre-motor neurons for regulation of postural reflexes (142). Not only vestibular but also visual, face and neck proprioceptive and other central signals are processed in the VN, allowing for rapid adaptation (error correction) in a changing spatial environment by integrating vestibular input with visuo-motor, cutaneous, and proprioceptive signals from extrinsic cerebral cortex and cerebellar sources (142). Similar to sensory neurons in the TG, neurons in the vestibular ganglion (VG) emit a peripheral branch to the vestibular apparatus in the inner ear and central afferents onto unipolar brush cells (UBC) in the GC layer of the cerebellar cortex (primary afferents) or into vestibular nuclei in the brainstem (secondary afferents). Both primary and secondary vestibular afferents belong to the mossy fiber system whose signals reach UBCs and then are distributed to GC parallel fibers across each folium of the uvula-nodulus (lobules IX and X) in the posterior cerebellum (143). Interestingly, vestibular nuclei also engage the oculomotor areas of the vermis (lobules V–VII) (82) while processing direct input from the mesencephalic trigeminal nucleus that processes proprioceptive information from extraocular and neck muscles to control eye-head coordination (134) and from periodontal ligaments to coordinate mastication (133). As expected, damage to flocculo-nodular and adjacent portions of the vermis, such as the uvula, can cause vertigo and extreme disturbance of equilibrium, head and body posture, and ocular movements (144, 145).

Neuronal Output From the Cerebellum

After processing of multiple incoming signals across the dendritic tree, PCs send this information mainly to deep cerebellar nuclei (DCN) as a feedforward node of cerebellar modulation to the rest of the brain. Information across this cortico-nuclear system reaches the brain through the fastigial (FN), interposed (IPN; formed by Globose and Emboliform nuclei), and dentate (DN) nuclei in the deep cerebellum. Axonal projections from these nuclei are vast, and terminal labeling has been historically observed in mainly five regions: the IO, PN, and VN in the brainstem and the red nucleus (RN) and ventrolateral thalamus



the brain, including reticular, vestibular, inferior olive, and pontine nuclei in the brainstem; the RN, PAG, SC, and VTA in the midbrain; and thalamic nuclei in the forebrain. Note the poly-synaptic connection with the cerebral cortex and the basal ganglia as part of a main functional network (see text). Output to the hypothalamus is not shown.

(VL) in mid- and forebrain, respectively (146, 147). Axon collaterals from main pathways also reach the PAG, superior colliculus (SC), ventral tegmental area (VTA), as well as reticular medullary regions and trigeminal spinal nucleus in the lower brainstem (95, 148–150) (**Figure 5**).

More recently, anatomical studies reevaluating both excitatory and inhibitory outputs from cerebellar nuclei show extensive projections to a variety of regions. Purkinje projections aim at inhibiting the mostly excitatory (glutamatergic) neurons in DCN, which will, in turn, influence its extra-cerebellar targets. Inhibitory influence of DCN has also been described, in which IPN and DN neurons targeting brainstem and midbrain structures are GABAergic and glycinergic (151–156). For example, there is GABAergic input to the IO through the nucleoolivary circuit (DCN-IO); there is another important inhibitory input to the PN (DCN-PN)—a major source of mossy fiber inputs to the cerebellum, suggesting that cerebellar inhibition of PN is a parallel regulatory feedback pathway similar to that looping through the IO (94, 138, 151). Concerning cerebellar output to VN, areas of the DCN under control of the flocculonodular cerebellar PCs connect to VN for oculomotor control during voluntary eye movements, as well as for optokinetic reflex (156– 158).

Another cerebellar target is the RN (DCN-RN), a midbrain premotor nucleus that receives projections originating from DN and IPN, but not FN (159, 160), and is also part of the modular cerebellar output to IO (RN-IO, or rubroolivar), as a source of climbing fibers, and to PN (RN-PN, or rubro-pontine), as a source of mossy fibers (95). In addition to the thalamus, the RN also projects densely to the spinal cord through the rubro-spinal tract for control of motor functions (159, 161). Its projection to the thalamus is

Cerebellum and Migraine

part of a topographically organized network to the cerebral cortex, and descending projections from the cortex via the cortico-rubral pathway converging with the cerebello-rubral pathway into the RN in a topographically organized manner (159). Similarly, the ventral tegmental area (VTA) is controlled by Crus I via DN (DCN-VTA), whose activity has recently been shown to control depression-like behavior induced by stress in mice (162). Projections to the PAG and superior colliculus are thought to play a role in pain modulation (see below), and oculo-motor functions, respectively (160, 163). Moreover, all DCN sends dense projections to the pontomedullary reticular formation (95, 149), a brainstem area acting as a relay for cortico/cerebello-reticulospinal control of posture and locomotion (164, 165), as well as many other fundamental brain functions, including somatic motor control and pain (43).

Thalamic projections from the cerebellum deserve special attention, given the complexity and the critical role it plays in both cerebellar function and migraine pathophysiology (166, 167). Dense and diverse, thalamic projections controlled by specific sets of PCs in the hemispheres are those from the DN to the thalamus via the dento-thalamic tract. In anatomical tract-tracing studies, DN sends axonal projections to central (CM, Pc, CL), ventral (VMl, VL, VPL, VPM, VA) and posterior (Po, Pf) thalamic nuclei (168-172). Although without the same level of structural resolution, the projection pattern has been confirmed by tractography and functional connectivity studies in the humans (173). In this case, dento-thalamic projections were nearly equally divided into 2 separate networks: the dorsal part of DN, for motor output associated with movement generation and control, and the ventral part of DN, for non-motor output associated with cognition and visuospatial function (174).

Direct cerebellar communication with the thalamus is also achieved from the FN. For instance, using single-cell genetic dissection of circuits in mice, different projection patterns emerged from five distinct group of excitatory glutamatergic neurons in the FN. Each group was controlled by a specific set of PCs in the vermis and IO neurons in the brainstem and sends topographically organized axonal projections to motor and non-motor areas in the brainstem, midbrain, and diencephalon. Interestingly, diencephalic thalamic projections were not limited to motor VL and VMl, but terminal labeling was also seen within other thalamic intralaminar areas, such as parafascicular (Pf), zona incerta (ZI), centrolateral (CL), and mediodorsal (MD) nuclei (156, 175). This pattern of projection suggests actions contributing to control of motor functions (i.e., axial and proximal parts of the body and ocular motion), and regulation of autonomic and affective reactions (i.e., cardiovascular, respiratory, and emotional responses) (171).

Cerebellar Di-Synaptic Connection to the Basal Ganglia and Cerebral Cortex

Altogether, cerebello-thalamic connections are extensive, and the thalamus plays a fundamental role in relaying this information to

the basal ganglia and cerebral cortex to form an interconnected neural network operating across multiple functional domains (69, 176). From this integrated network perspective, the basal ganglia role in the network is supported by recent data, showing that the subthalamic nucleus in the basal ganglia is the source of a dense di-synaptic projection to the cerebellar cortex, and the DCN is the source of a dense di-synaptic projection to the striatum. Basically, these two subcortical systems communicate to motor and non-motor areas of the cerebral cortex via the thalamus in a close-loop arrangement (176-180), which led to a new functional perspective that the basal ganglia, cerebellum, thalamus, and the cerebral cortex form an interconnected and topographically organized network for motor, sensory, cognitive, and affective functions (69, 76, 176, 181-183). Supporting this functional connectome, anatomical studies on primates and rodents using transsynaptic tracing have shown extensive di-synaptic connections from cerebellar nuclei to cognitive, affective, and motor forebrain circuits in many cortical areas. For instance, a widespread pattern of axonal terminals in Layer 1 of the cortex from the thalamus was observed throughout much of the cortex, from frontal to association and visual areas. A second, more restricted pattern of axonal terminals in Layers 3-5 was observed in sensorimotor (i.e., S1/S2, M1/M2), frontal association, and orbitofrontal cortices. Altogether, this complex circuit was suggested to participate in adaptive control of somatomotor, autonomic, and arousal functions during postural changes and locomotion (77, 160).

As expected, all the cerebral cortex areas that are targeted by cerebellar output (or ascending trigeminal) send reciprocal feedback from cortical Layer 5 pyramidal neurons via axonal projections to PN, IO, and VN, but also medullary reticular formation and Sp5C in the trigeminocervical complex (46, 49, 98, 100-102, 184-190). The functional implications of these cortical descending pathways are intricately associated with the ascending input they receive, and are somatotopically organized for the regulation of motor, cognitive, autonomic, and sensory functions. For instance, anatomical studies have shown that descending cortical pathways include the cortico-pontocerebellar and cortico-olivo-cerebellar pathways. In return, information from the cerebellum flows within the cerebellothalamo-cortical pathway, supporting the hypothesis of a loop as a substrate for cerebro-cerebellar communication during sensorimotor and cognitive processing. In this loop, motor, prefrontal, and temporal cortices appear to be the most relevant (173, 191-194). A classic demonstration in which these pathways are activated is when performing visually guided motor tasks, in which along with specific regions in the pons and midbrain to integrate visual information for the cerebellum to control motor output (51, 69, 176). Given the extensive and organized association of the cerebellum with the forebrain across cerebro-cerebellar loops, Schmahmann noted that "motor, cognitive and neuropsychiatric disorders in patients with lesions restricted to basal ganglia, thalamus, or cerebellum mimic deficits resulting from cortical lesions, with qualitative differences between the manifestations of lesions in functionally related areas of cortical and subcortical nodes" (183, 195).

TRIGEMINAL PAIN AND THE CEREBELLUM

The cerebellum is not exempt from pain processing duties, including that of migraine. Accordingly, sensory afferents from trigeminal territories have been shown to send direct axonal projections to the cerebellum (128, 129). In addition, 2ndorder neurons in spinal trigeminal nuclei also send axonal fibers to the cerebellum from all subnuclei (Sp5I, Sp5O, and Sp5Pr), except from Sp5C (131), where most dural nociceptive processing occurs (3, 15, 196-199). Indeed, anatomical studies on rodents showed that Sp5C areas receiving innocuous and noxious input from trigeminal periocular and dural areas do not project directly to the cerebellum (200), suggesting that trigeminovascular input to the cerebellum is poly-synaptic. There is, however, a strong functional connectivity from Sp5C to rostral areas of trigeminal nuclei, particularly in somatosensation from oral and extra-oral regions of the face and the head (201-205). Tracing studies on non-human primates and rodents have shown that this ascending pathway includes trigeminal input to reticular areas, such as the subnucleus reticularis dorsalis (206-208); nucleus of the solitary tract (NTS), superior salivatory nucleus (SSN), and PAG, among others (200). Indirect input from trigeminovascular neurons to the cerebellum is likely through other trigeminal nuclei connecting via brainstem IO, VN, and/or PN (128-131, 134, 138). Other than nociceptive information, these areas can integrate autonomic and sensory-motor signals from trigeminal territories and send them to the cerebellum, thalamus, and cerebral cortex. Accordingly, electrophysiological studies on animals and imaging studies on humans suggest the existence of two pathways carrying cutaneous and visceral nociceptive information to the cerebellum, one from the IO, mainly through the spino-olivo-cerebellar pathway to Purkinje cells in the anterior lobe; and another from the PN via spino-pontocerebellar pathway to the vermis (140, 209-211). For instance, C-fiber nociceptor signals from visceral organs (i.e., colorectal distension) can affect cerebellar PC activity in the vermis through MF originating in the medullary reticular formation (44, 212, 213).

Also important in the pain-cerebellum relationship is the many supra-medullary structures, such as the PAG, dorsal raphe nuclei (DRN), and LC-all critical nodes in nociceptive processing, which are in close association with the cerebellum either directly or through poly-synaptic communication. Functionally, activity recorded from the IO as input to the cerebellum is reduced by PAG stimulation, suggesting descending modulation of motor nodes, extending the role of this midbrain structure to modulation of ascending CF inputs to the cerebellum. Functionally, this circuit may play a role in regulating motor activity in different behavioral states during acute and chronic pain conditions (163). In animals, FN and DN projections to glutamatergic, GABA, and dopamine neurons in the ventrolateral PAG are thought to participate in freezing behavior and fear memory formation (81, 163, 214-217). Similarly, cerebellar projections to the DRN, a major source of serotonin, may cooperate with the DN-VTA pathway to regulate mood-, reward-, and stress-associated behaviors, as evidenced by recovery from depressive symptoms due to increased firing of DRN serotonergic neurons in rodents (156, 218). Lastly, DCN and Purkinje projections to the LC, the main source of NA, may participate in functions related to arousal, attention, motivation, and stress responses (219–221). On the many functions of the LC, this nucleus has been recently shown to simultaneously inhibit trigeminovascular neurons in the Sp5C and increase cortical SD susceptibility (222).

It is not completely understood, however, how direct or indirect cerebellar connections interact with trigeminovascular nodes or migraine-related structures. In the context of experimental trigeminal pain, a recent notable work using fMRI in the humans has mapped the activations in the cerebellum from trigeminal nociception in the nostril of healthy individuals to the ipsilateral posterior lobe of the cerebellum, namely, lobules VI, VIIIa, and Crus I, where intensity and unpleasantness were mostly processed in the face area of lobule VI. During stimulation, trigeminal pain induced strong functional connectivity with the rostral pons, PAG, and thalamus, suggesting descending antinociceptive and motor reactions, in addition to higher processing in the insular cortex, the operculum and the putamen, and the face areas in the precentral gyrus (35). Another study applying noxious stimuli in the thumb and the toe found cerebellar activations consistent with somatotopic organization of direct input from the spinal cord, and non-somatotopic activation consistent with indirect information, flowing from cortical and subcortical connections, possibly involved in processing contextual emotional states, like expectation of pain (223).

On the pathological side, functional and structural alterations have been observed in the Crus I and II of the posterior cerebellum in patients with migraine, in addition to gray matter volume abnormalities. Moreover, cerebellar activity in response to trigeminal pain was modulated by migraine severity and the migraine phase, along with decreased connectivity to the thalamus and higher cortical areas (66). Also, and based on the neural associations described above, where the cerebellum is connected to thalamic and basal ganglia nuclei, fMRI studies on patients with migraine detected decreased responses to pain in the caudate, the putamen, and the pallidum of the basal ganglia, suggesting neural plasticity changes, particularly in highfrequency migraineurs, leading to altered pain processing and chronification (224). As described above, increased functional connectivity was detected between the hypothalamus and brain regions involved in regulation of autonomic functions, including the locus coeruleus, the basal ganglia, and the cerebellum, in which some of these responses were synchronized across autonomic, sensory, and cerebellar networks (61). In addition, areas of the brain involved in cognition and nociception, including the cerebellum, respond stronger in migraineurs to expectation and awareness of painful events (225). As main players in this network, the cerebellum and the anterior insular cortex were implicated in the affective component of pain perception in association with tumors in the posterior cerebellum

of children (226), an idea that is also supported by cerebellar infarction cases with altered pain perception (227).

CLINICAL AND SUBCLINICAL SYMPTOMS ASSOCIATED WITH MIGRAINE AND CEREBELLAR FUNCTION

There is a wide range of acute and chronic neurological conditions associated with cerebellar abnormalities, in which symptomatology can fully express itself as ataxia or tremor, nystagmus or visuomotor alterations, and even obsessivecompulsive disorder or depression, to name a few (157, 174, 228, 229). Such apparent global implication of the cerebellum in brain functions appears more difficult to track in subtle or subclinical presentations, particularly in common types of migraine, in which specific involvement in migraine may be difficult to identify during clinical evaluation, probably because standard clinical tests are not sensitive enough to capture subtle alterations in sensorimotor, affective or cognitive control, requiring a more scientific approach for their exploration. Another challenge is that migraine symptoms may emerge or accentuate at different phases of the disorder, particularly at the preictal and ictal phases. For example, in clear support of cerebellar involvement in migraine, Sandor and colleagues reported subclinical alteration of reaching movements in common types of migraine, where patients show a hypermetric component accentuated by increasing the inertial load of the moving arm, which is more pronounced in migraine with aura, followed by migraine without aura and healthy control (57). Such ballistic movements are thought to be processed by the lateral (hemispheres) cerebellum, so the hypermetria could be attributed to abnormal functioning at subclinical levels. Another study unveiling subclinical features in migraine also showed vestibulocerebellar alterations. In this case, patients with migraine with and without aura had altered results in a neurotologic test, evaluating oculomotor function and posture, and with only 25% of them experiencing vertigo and/or dizziness (230).

Historically, motor processing is the most studied function, and like in many other diseases, cerebellar symptoms linked to motor dysfunctions are seen across the migraine spectrum, particularly in patients carrying monogenic hereditary mutations, leading to the most severe phenotypes of migraine (53, 231). Infarcts affecting the posterior cerebellar lobe are associated with symptoms such as dizziness, headaches, nausea and vomiting, unilateral limb weakness, nystagmus, and dysarthria (232, 233). Although less common, infarcts affecting the anterior lobe may lead to the cerebellar motor syndrome characterized by a lack of coordination in voluntary movements, speech deficits, impairments of limb movements, and abnormalities of posture/gait (76, 83, 234).

In episodic migraine, but, more frequently, in its chronic form, *fatigue, tiredness and/or motor weakness* are common complaints in anticipation of, during or after an attack (235–238). Increased fatigue, generally defined as difficulty initiating or sustaining voluntary motor activity, has been reported in both episodic and chronic migraine, being more severe and complex

in the latter, as well as associated with other comorbidities, such as depression (239, 240). Fatigue and tiredness may be conceived as a psychological sensation associated with or without effective decrease in muscular force (241). The latter is more clearly illustrated in stroke survivors experiencing fatigue and heavy limbs but not muscle weakness (241). In migraine induced with nitroglycerin, premonitory symptoms also include tiredness and concentration difficulty (242), supporting a dual cognitive and physical dimension of fatigue. The physical dimension is associated with the need to rest, sleepiness, drowsiness, a lack of energy, and the feeling of less strength in muscles or motor weakness. The cognitive dimension of fatigue, however, may express as difficulty concentrating and finding words, and a lack of clarity or memory problems (239). More robust motor manifestations have been reported as common in migraine with sensory disturbances and unilateral weakness (243), and, recently, in novel forms of Familial Hemiplegic Migraine (FHM) of CACNA1A mutations with reversible unilateral or bilateral motor weakness (244). In other extreme examples, patients with mutations in CACNA1A, ATP1A2, or SCN1A had attacks more often, may be triggered by mild head trauma and were associated with extensive motor weakness, ataxia, confusion, and brain edema. Patients with FHM without these mutations display a milder phenotype similar to that of common migraine (231). Interestingly, the subclinical hypermetria originating in the lateral cerebellum in patients with migraine with aura (57) was correlated with alterations in neuromuscular transmission measured with single-fiber EMG, suggesting a common molecular alteration in these two systems (245).

This is highly relevant, given the role of the cerebellum in regulating muscle tone and strength throughout the body (234). Accordingly, alteration in muscle strength and endurance has been linked to migraine in the past. More specifically, cervical muscles were weaker in migraine concurrent with neck pain and were associated with the severity of cutaneous allodynia and chronification (246-248). In this context, migraine is more common in patients with cervical dystonia, a disease involving one or more muscles from the neck and the shoulders, with or without pain, and characterized by involuntary muscle contractions, twisting and abnormal postures in the body (249). It is thought to originate from abnormal processing in the basal ganglia and cerebellum, likely due to genetic mutations affecting pumps and ion channels in the cerebellum (i.e., some of the same pumps and channels that are affected in migraine), and may be precipitated by activity or exercise (249-254).

Additional support for the participation of the cerebellum in these processes originates from patients with lesions produced by superior cerebellar artery infarction who develop gait and trunk ataxia and dysarthria (72). *Cerebellar ataxia* is probably the most recognized symptom originating from cerebellar malfunctioning. Widely recognized in a variety of pathologies, such as spinocerebellar ataxia, its presence in many types of migraine has also been reported (231, 255). In FHM, patients present symptoms commonly associated with hemiparesis during the aura phase. So far, three FHM genes have been identified corresponding to FHM-1,-2 and-3, with mutations in CACNA1A—encoding subunits of voltage-gated calcium

channels; in ATP1A2-encoding for Na-K+ ATPases; and in SCN1A genes-that encode for voltage-gated sodium channels (120, 256, 257). All these channels are widely expressed across the CNS, but with a significantly higher expression in the cerebellum (110, 121), which led to suggestions that the cerebellum is critically involved in migraine (41). Each of the three mutations has a dominant phenotype, including severe migraine and contralateral hemiplegia; however, due to natural mutations and epigenetic changes, phenotypes from a single mutation may vary substantially (53, 231, 258, 259). Two recent reports, indeed, have shown an increase in new variants or mutations for a single gene, the CACNA1A, which could produce alleviation of episodic ataxia type 2 by sleep, and fully reversible bilateral motor weakness or diplegia, suggesting that multiple variants of a single, critical gene for neuronal excitability can produce divergent phenotypes (244, 260). Contrary to the relatively small number of patients with FHM (<.1% in population) or those with a family-linked genetic condition, producing migraine-type phenotypes, GWAS studies on common migraine show a complex picture on genetic association for primary cerebellar dysfunction. The study found three lead variants associated with migraine with aura, in which one of them, CACNA1A-related to both monogenic and polygenic forms of migraine-provides a gene-based connection between migraine sub-types (261). Also interesting was the identification of risk loci, containing genes that encode targets for migraine-specific therapeutics, such as CALCA and CALCB genes encoding for CGRP. Mechanistically, neuronal expression of CACNA1A across the CNS, with significantly higher expression of calcium channels in the cerebellum, points to altered brain excitability and susceptibility to SD-like events, in which CGRP plays an unknown but likely significant role (262-265).

Undoubtedly, the most significant advance from the scientific point of view has emerged from the creation of the first genetically modified mice carrying the human CACNA1A gene mutations (266, 267). Since then, genetic, electrophysiological and behavioral studies on knock-in mice have increased exponentially, and provided a large flow of data on the effects of these mutations on synaptic transmission, excitability, and other basic neurophysiological functions, critical to mechanismbased disease understanding (256, 265, 266, 268-272). For instance, in mice carrying FHM-1/2 mutations, cerebellar ataxia is a common symptom, particularly in FHM-2 and some variants of FHM-1. Some phenotypes also present with increased susceptibility to seizures and cerebral edema after minor head trauma (120). Nevertheless, despite the fact that the phenotypic spectrum of migraine is wide and certainly associated with complex interactions between genetics and the environment, the functional characterization of this genetic model is critical. The creation of more viable heterozygous mutant mice or rats will certainly provide new guidance on the pathologies associated with cortical and cerebellar dys-excitability conditions, including migraine and its subtypes.

Among other motor signs associated with migraine and cerebellar function is *Tremor*. Essential tremor (ET) is one of the most common movement disorders, characterized by high-amplitude tremor during posture or intentional movement. It

has been shown that the risk and the prevalence of ET are substantially higher in migraineurs, as well as higher prevalence of migraine in patients with ET (273-275). Intention tremor occurs mostly in upper limbs and speech muscles and may be related to alterations across the olivo-cerebellar pathways and their thalamic targets (276, 277). Indeed, dysarthria, a speech disorder characterized by a dysfunction in initiation or coordination of motor structures involved in speech (278), where there is a reduced lingual (tongue elevation) and facial strength producing a transient loss of speech output (279), has also been associated with migraine (280-282). In the extreme cases, reports of cerebellar mutism syndrome-a rare condition in children with developmental conditions-a bilateral disruption in cerebellar hemispheres and abnormal signaling across the dento-thalamo-cortical pathway has been proposed as a neural substrate (283-285). Nevertheless, and despite having a narrow understanding of language processing in the migraine brain, it is hard to conceive dysarthria or other motor or cognitive language disorder as a pure emanation of local cerebellar neurovascular conflict, given the loop type of communication it maintains with the cerebral cortex (286).

A second complex function of the cerebellum, in coordination with the recently reviewed motor and cognitive functions, is the integration of vestibular signals from the inner ear for balance and equilibrium. As described above, Purkinje cells in the cerebellar cortex send heavy projection to vestibular nuclei in the dorsal pons area (287), and VN projects back to DCN in a strong reciprocal anatomical and functional connection (142). Infarct lesions from anterior and inferior cerebellar artery may present with vestibular syndrome, and auditory symptoms in some cases (288, 289). Given the above, alterations of cerebellarvestibular pathways can lead to vertigo, dizziness, motion sickness, and altered balance. Such symptomatology has been associated with migraine, particularly with the specific clinical entity vestibular migraine. The exact pathophysiology of VM is not known, and symptoms-based hypothetical proposals are likely describing a simpler picture of what is actually occurring at the neurobiological level. Studies looking at associating lesions with function have mostly reported headache and vertigo in patients with posterior inferior cerebellar artery lesions, and in greater proportion than those with superior cerebellar artery compromised (289).

Vertigo or dizziness can occur occasionally in about 50% of patients with migraine (290–292), with lifetime prevalence of 16% for migraine and 7% for vertigo in the general population, and 1% for VM (58). In addition, given that thalamic nuclei receive vestibular signals from both vestibular and cerebellar nuclei (293), it has been proposed that abnormal functional response to vestibular stimulation in the thalamus may contribute to VM pathophysiology (294). In episodic migraine, however, evidence of subclinical symptoms in patients without a history of vertigo, dizziness, or postural imbalance has recently emerged, supporting a vestibulo-cerebellar dysfunction in common migraine (60). Indeed, a recent study has shown that up to 85% of women with migraine with aura and chronic migraine had vestibular symptoms, which are exacerbated during the headache attacks (59). In another study using

MRI, Messina and colleagues compared healthy individuals with VM and patients with migraine with (MwA) or without aura (MwO), and found a distinctive pattern of regional gray matter abnormalities, including increased volume of the red nucleus, thalamus, and occipital regions, and decreased volume of the cerebellum compared to controls, suggesting a critical involvement of cerebellar networks in these clinically separated migraine phenotypes (295, 296). Within the migraine spectrum, clinical entities with similar symptomatology are referred to as migraine-associated vertigo, vertiginous migraine, migraine-associated balance disturbance, and benign paroxysmal vertigo (56).

Despite cerebellar infarctions in the superior cerebellar artery triggering nausea and vomiting associated with gait ataxia, it is not known how these symptoms are generated. Very likely, sensation of nausea may emerge through activation of the nucleus of the solitary tract via vestibular signaling (56, 142). These direct projections from VN to NTS have been traced in the cats, rabbits, and rats (297-299). In the cases with nausea and/or *vomiting*, they most frequently occur during the headache phase but may be also present interictally (300). Nausea and vomiting are triggered by visceral inputs to vagal areas in the brainstem, including those from the visceral meninges, where nausea can be produced by nociceptive input from the viscera. This suggests that, in migraine, nausea can be an anticipatory or premonitory symptom, signaling an increase in trigeminovascular traffic from the periphery to the brainstem, at a subclinical level without pain to be consciously detected yet. In nitroglycerin-induced headaches, PET imaging in migraineurs shows activation in the nucleus of the solitary tract, dorsal motor vagus, and ambiguous nuclei, all neural nodes that have been singled out as pivotal structures for the generation of nausea sensation in migraine (300).

Although the cerebellum has not been associated with the sensory perceptual creation itself, modulation of sensory acquisition and related processes are considered one of its integral functions (48, 301). Accordingly, in an interesting association between visual perception, migraine and the cerebellum, Puledda and colleagues reported a case of visual snow syndrome (VSS), evolving from episodicto chronic without headache in a patient after infarction affecting the cerebellar Crus I and Lobule VI (302). This is an educational demonstration of cerebellar influence on high-level sensory functions, where altered functional connectivity at rest or during visual stimulation was observed in several pre-cortical and cortical networks involved in visual motion, attention, and salience (303), including the occipital lingual gyrus, which has been previously associated with migrainous photophobia (304, 305). Photophobia has actually been quantified and reported highly prevalent in patients with VSS, in a level similar to chronic migraine, which represents an interesting association, given the shared functional networks involved in both conditions (167, 306-309).

As described above, the cerebellar role in cognitive functions is very active, and *cognition* is affected in migraine as well. For example, in the premonitory phase, common cognitionassociated complaints include difficulty concentrating, finding words or having a diminished capacity for tasks requiring working memory (19, 310, 311). Transient global amnesia has also been associated with cerebellar ischemia (312). These disabilities can be attributed, at least in part, to network dysfunction that includes the cerebellum and its reciprocal functional connectivity with prefrontal, limbic, and autonomic cortical areas. In addition, alterations in cognitive processing are more pronounced in the preictal, ictal, and postictal phases (311). This cognitive component may be associated with the modular connections of DCN neurons with brainstem, autonomic, and thalamic nuclei, providing a neural substrate for the cerebellar cognitive affective syndrome (313) and could account for the therapeutic effect of vermis intervention on neuropsychiatric disorders (195, 314–316).

CLINICAL AND SCIENTIFIC EVIDENCE LINKING CEREBELLAR NETWORKS TO MIGRAINE PATHOGENESIS

A large body of clinical evidence and, more recently, imaging findings in the brain of patients with migraine have identified a number of pathological events connecting directly the cerebellum with migraine pathogenesis (66). Clinically, an arguable association can be established between the cerebellum and premonitory symptoms, such as feeling tired, having difficulty concentrating, and a stiff neck (19, 22, 317). These symptoms complement those presented above in association with migraine; however, the cerebellum has not been directly linked to premonitory symptoms. Functional connectivity of the hypothalamus is altered during the premonitory phase (22, 33, 318), where areas of the dorsolateral pons become activated at the headache onset as part of the descending pain modulatory response, in addition to activations across the cortical and subcortical pain matrix and cerebellum (319-321). Interestingly, an fMRI study of visually triggered migraines replicated these findings (i.e., activation of dorsolateral pons and PAG) and also found activation across nodes connected with the cerebellum, such as RN and PN, before the onset of migraine symptoms (322), suggesting an early involvement of the cerebellum.

Additional abnormalities in microstructure and/or function of the cerebellum in migraine have been described lately (323-325). Also, many connectivity alterations of the cerebellum with its targets, particularly the thalamus, pons, and cerebral cortex, in addition to the hypothalamus in the premonitory phase described extensively above, appear to emerge in the headache phase. For example, a resting-state fMRI study showed altered thalamic connectivity during spontaneous attacks of migraine without aura (326). Moreover, during migraine with aura, there was an increased connectivity between the pons and somatosensory cortex (327), reflecting cortico-pontine communication. Another interesting finding is the increased perfusion in the brainstem during attacks preceded by aura (328). Altogether, these functional imaging studies support a framework of altered dynamic connectivity in the different phases of migraine and across neural structures involved in motor, sensory, autonomic, affective, and cognitive functions.

Another line of evidence linking the cerebellum to migraine pathogenesis comes from population-based imaging studies

on the humans, where migraineurs had significantly higher prevalence of white matter hyperintense and infarct-like lesions in the deep cerebellum (329). A follow-up description of these findings showed small border zone infarct-like lesions in patients with migraine, particularly in those with aura, probably due to hypoperfusion and embolism within the small vasculature (330). It was then determined that patients with migraine are at increased risk of developing these subclinical white matter lesions in the brain, and, particularly, in the cerebellum supplied by the posterior circulation (331). Despite the risk, a more recent study has shown that a cohort of migraineurs displayed no or minimal cerebellar dysfunction, despite having higher prevalence of these clinically silent infarctions (64). Therefore, whether symptomatic or subclinical, it seems that vascular pathology is at play. Accordingly, the apparent cerebellar predilection of ischemic lesions in migraine with aura might be due to a combination of altered autoregulation and artery cerebellar angioarchitecture (332). Additional evidence using perfusion MRI supports a benign presentation of these lesions as well as cerebellar hypoperfusion associated with cortical oligemia in migraine with aura (68).

A recent renewed interest has emerged to better understand energy metabolism in the brain as a potential pathogenetic mechanism for migraine predisposition, which has been reviewed extensively (333, 334). There, Gross and colleagues provide diverse evidence supporting the role of energy metabolism and mitochondrial dysfunction in the migraine brain, and propose that "migraine is a response to cerebral energy deficiency or oxidative stress levels that exceed antioxidant capacity and that the attack itself helps to restore brain energy homeostasis and reduces harmful oxidative stress levels" (333). Accordingly, among the many mitochondrial disorders linked to migraine, there is one called MELAS-for mitochondrial encephalomyopathy, lactic acidosis, and stroke-like episodesthat develop symptoms of recurrent headaches along with muscle weakness and exercise intolerance in normal psychomotor individuals (335). Lucchesi and colleagues reviewed another interesting association between energy metabolism, fatigue, and migraine (240). The authors reported evidence on impairment of oxidative mitochondrial metabolism in the brain, and concluded that fatigue is related to migraine, probably caused by systemic impaired energy metabolism, and that the effect of antioxidant molecules, such as riboflavin and coenzyme Q10, may improve migraine frequency, severity, and fatigue levels (240, 336). Although none of these studies localized these abnormalities to a specific brain region, metabolic dysfunction appears widespread to all brain regions, and it seems particularly relevant in the cerebral cortex, the cerebellum, and the brainstem. Actually, emerging technologies, such as proton magnetic resonance spectroscopy (1H-MRS), have identified regional metabolic activity in the brain by quantifying the levels of glutamate/glutamine (Glx), lactate, total creatine, and N-acetylaspartate in migraine without aura interictally. Interestingly, creatine was the only one significantly increased in the pons, an area that includes the PAG, LC, DRN, and pre-cerebellar nodes, and was proposed as a biological marker for migraine without aura (337). In another study, systemic lactate levels were found to be increased in migraine and fibromyalgia as compared

to healthy controls (338). These studies suggest that abnormal energy metabolism at the most basic cellular level in the PN and other brain areas may contribute to migraine pathogenesis. In any case, given the above and given the fundamental role of the hypothalamo-brainstem in regulating feeding, temperature, stress response, and energy homeostasis (339, 340), the cerebellum is well positioned to play a relevant role in migraine pathogenesis via energy metabolism. Accordingly, the cerebellum is a highly populated and metabolically demanding neural structure. Its computational power is involved in virtually all brain functions, and, despite being overshadowed in mass by the cerebral cortex (10%), particularly in humans, its energy demand is thought to be around 75% of the whole brain, which would be necessary to maintain the 2/3 of total brain cells that are exclusively contained in the cerebellum (341). Also, it contains large neural populations, firing at more than 50 Hz to keep the connected systems in proper balance at all times, adding an extra load in energy requirement.

Although the consequences of these metabolic alterations are not completely understood, animal studies have led the way in using spreading depolarization (SD)-like phenomena as an experimental paradigm to test the effects of this neurovascular event on electrophysiological, vascular, and metabolic responses across the cerebral cortex (17, 342). Triggering this event in animals can activate meningeal nociceptive pathways through impairment of the paravascular space and glymphatic flow in the cerebral cortex, as well as activation of immune cells near pial and dural tissues, signaling potential mechanisms of the headache onset (343-345). Undisputable evidence shows the presence of CSD in the human occipital cortex (346), and, possibly, a mechanism of activation through inflammation in the adjacent bone marrow (347); however, other than hypoperfusion (68), a homologous event in the cerebellum has not been described in the humans. In rodents, however, SD-like phenomena can be triggered in the cerebellum, but it is difficult to evoke experimentally (348), and thus to find a pathophysiological correlate. A possible link refers to short-lived changes in pH and ionic concentration in the cerebellar parenchyma during experimental induction of the spreading acidification and depression (SAD) phenomenon (349). How these changes in the cerebellar microenvironment may lead to headache is unknown, but anatomical and physiological data from both humans and animals suggest a possible connection of events in the posterior fossa, with activation of afferent sensory information from the posterior dura to the brainstem trigeminocervical complex via C2/3 DRG cervical innervation (4, 350, 351). Lastly, given that pain in occipital areas of head, including neck muscle tenderness, is experienced by a large number of migraineurs, and that patients with vestibular migraine appear more likely to experience headaches in the occipital region of the head (352), it is possible that the cerebellum is playing a larger role in migraine pathophysiology than previously thought.

CONCLUSIONS

There is an increasing amount of clinical and scientific evidence linking cerebellar networks with migraine pathophysiological mechanisms, likely originating in altered structure and function

across shared network connections. In migraine, genetically driven dysfunction in cerebellar activity has been consistently shown, and, based on preclinical research from animal models, it may be associated with intrinsic metabolic alterations driving excitatory/inhibitory imbalances at different phases of the migraine cycle. These neural dynamics may be relevant when looking at some fundamental theories about the cerebellum role within the brain. For instance, cerebellar motor functions are diverse, ranging from oculomotor and speech to grip force, voluntary limb movement, and sensorimotor coordination, and, despite a lack of consensus, the motor control theory proposes a core concept on representation of time to perform behaviors requiring real-time prediction. Here, the GC layer of the cerebellar cortex is critical to such temporal encoding, where parallel CF input to PC carries error signals predicting movement through trial-and-error practice (72, 79). In Schmahmann's dysmetria of thought theory-a sophisticated conceptual proposal-the cerebellum plays the fundamental function of performance optimization across the brain. It requires the integration and coordination of multiple incoming signals that, in turn, are continuously modulating the output behavior according to cognitive, emotional, and sensory contexts (83). The modular structure of the cerebellum allows the presence of simple processing units, repeated across the cerebellar cortex, which, when disturbed by damage in the form of altered excitability, blood supply or injury, a variety of different symptoms associated with their specific function allocation may emerge (43, 183). This view is complemented by Bower's control of sensory data acquisition theory, which describes a cerebellum as an efficiency facilitator for other brain regions directly involved in executing those functions or tasks. Basically, it does not participate directly in the production of behavioral expressions (i.e., in the context of this review, the experience of pain and learned behaviors, headache, and related

REFERENCES

- Ferrari MD, Goadsby PJ, Burstein R, Kurth T, Ayata C, Charles A, et al. Migraine. Nat Rev Dis Primers. (2022) 8:2. doi: 10.1038/s41572-021-00328-4
- IHS. Headache classification committee of the international headache society (IHS) the international classification of headache disorders, 3rd edition. *Cephalalgia*. (2018) 38:1–211. doi: 10.1177/0333102417738202
- Noseda R, Burstein R. Migraine pathophysiology: anatomy of the trigeminovascular pathway and associated neurological symptoms, cortical spreading depression, sensitization, and modulation of pain. *Pain.* (2013) 154 (Supplement1):S44–53. doi: 10.1016/j.pain.2013.07.021
- Noseda R, Melo-Carrillo A, Nir RR, Strassman AM, Burstein R. Non-trigeminal nociceptive innervation of the posterior dura: implications to occipital headache. J Neurosci. (2019) 39:1867–80. doi: 10.1523/JNEUROSCI.2153-18.2018
- Olesen J, Burstein R, Ashina M, Tfelt-Hansen P. Origin of pain in migraine: evidence for peripheral sensitisation. *Lancet Neurol.* (2009) 8:679– 90. doi: 10.1016/S1474-4422(09)70090-0
- Ashina M, Hansen JM, Do TP, Melo-Carrillo A, Burstein R, Moskowitz MA. Migraine and the trigeminovascular system-40 years and counting. *Lancet Neurol.* (2019) 18:795–804. doi: 10.1016/S1474-4422(19)30185-1
- Goadsby PJ, Holland PR, Martins-Oliveira M, Hoffmann J, Schankin C, Akerman S. Pathophysiology of migraine: a disorder of sensory processing. *Physiol Rev.* (2017) 97:553–622. doi: 10.1152/physrev.00034.2015

protective reactions) but, rather, is involved in monitoring and adjusting the acquisition of most sensory data on which the rest of CNS depends (301). This type of modulatory role is exerted through communication across the cerebrocerebellar loops formed by the cerebral cortex, basal ganglia, thalamus, and hypothalamus, which requires fast and robust processing capabilities for stabilizing brain areas that fall out from homeostasis (182). Accordingly, homeostatic challenges become triggers that may drive migraine attacks in susceptible individuals, probably to reset network abnormalities (236, 353). Nevertheless, we have a limited understanding of the role of the cerebellum in triggering migraine and in initiating headache. Therefore, given the diverse clinical and subclinical neurological symptoms outlined in this review, understanding the role of the cerebro-cerebellar networks, particularly the underpinnings of hypothalamo-medullary connectivity in handling external and internal stressors in migraine, becomes more relevant than ever (340, 354).

AUTHOR CONTRIBUTIONS

RN performed bibliographic research, prepared artwork, and wrote the manuscript.

FUNDING

This work was funded by NIH/NINDS Grant R01 NS104296.

ACKNOWLEDGMENTS

The author would like to acknowledge Laura V. Cantera for proofreading the final version of the manuscript and my family for the unconditional support.

- Edvinsson L, Grell AS, Warfvinge K. Expression of the CGRP Family of Neuropeptides and their Receptors in the Trigeminal Ganglion. J Mol Neurosci. (2020) 70:930–44. doi: 10.1007/s12031-020-01493-z
- Russo AF. Calcitonin gene-related peptide (CGRP): a new target for migraine. Annu Rev Pharmacol Toxicol. (2015) 55:533-52. doi: 10.1146/annurev-pharmtox-010814-124701
- Silberstein SD, Dodick DW, Bigal ME, Yeung PP, Goadsby PJ, Blankenbiller T, et al. Fremanezumab for the preventive treatment of chronic migraine. N Engl J Med. (2017) 377:2113–22. doi: 10.1056/NEJMoa1709038
- Diener HC. CGRP antibodies for migraine prevention new kids on the block. Nat Rev Neurol. (2019) 15:129–30. doi: 10.1038/s41582-019-0139-7
- Melo-Carrillo A, Noseda R, Nir RR, Schain AJ, Stratton J, Strassman AM, et al. Selective inhibition of trigeminovascular neurons by fremanezumab: a humanized monoclonal anti-CGRP antibody. *J Neurosci.* (2017) 37:7149– 63. doi: 10.1523/JNEUROSCI.0576-17.2017
- Noseda R, Schain AJ, Melo-Carrillo A, Tien J, Stratton J, Mai F, et al. Fluorescently-labeled fremanezumab is distributed to sensory and autonomic ganglia and the dura but not to the brain of rats with uncompromised blood brain barrier. *Cephalalgia.* (2020) 40:229– 40. doi: 10.1177/0333102419896760
- Noseda R, Jakubowski M, Kainz V, Borsook D, Burstein R. Cortical projections of functionally identified thalamic trigeminovascular neurons: implications for migraine headache and its associated symptoms. *J Neurosci.* (2011) 31:14204–17. doi: 10.1523/JNEUROSCI.3285-11.2011

- Akerman S, Holland PR, Goadsby PJ. Diencephalic and brainstem mechanisms in migraine. Nat Rev Neurosci. (2011) 12:570–84. doi: 10.1038/nrn3057
- Ashina M, Terwindt GM, Al-Karagholi MAM, Boer I de, Lee MJ, Hay DL, et al. Migraine: disease characterisation, biomarkers, and precision medicine. *Lancet.* (2021) 397:1496–504. doi: 10.1016/S0140-6736(20)32162-0
- Ayata C, Lauritzen M. Spreading depression, spreading depolarizations, and the cerebral vasculature. *Physiol Rev.* (2015) 95:953–93. doi: 10.1152/physrev.00027.2014
- Brennan KC, Pietrobon D. A systems neuroscience approach to migraine. Neuron. (2018) 97:1004–21. doi: 10.1016/j.neuron.2018.01.029
- Karsan N, Goadsby PJ. Biological insights from the premonitory symptoms of migraine. Nat Rev Neurol. (2018) 14:699– 710. doi: 10.1038/s41582-018-0098-4
- Dodick DW. A phase-by-phase review of migraine pathophysiology. Headache J Head Face Pain. (2018) 58:4–16. doi: 10.1111/head.13300
- Maniyar FH, Sprenger T, Monteith T, Schankin CJ, Goadsby PJ. The premonitory phase of migraine - what can we learn from it? *Headache J Head Face Pain*. (2015) 55:609–20. doi: 10.1111/head.12572
- Schulte LH, Peng KP. Current understanding of premonitory networks in migraine: a window to attack generation. *Cephalalgia*. (2019) 39:1720– 7. doi: 10.1177/0333102419883375
- Schulte LH, Jürgens TP, May A. Photo-, osmo- and phonophobia in the premonitory phase of migraine: mistaking symptoms for triggers? J Headache Pain. (2015) 16:14. doi: 10.1186/s10194-015-0495-7
- 24. Karatas H, Erdener SE, Gursoy-Ozdemir Y, Lule S, Eren-Kocak E, Sen ZD, et al. Spreading depression triggers headache by activating neuronal Panx1 channels. *Science*. (2013) 339:1092–5. doi: 10.1126/science.1231897
- Zhao J, Blaeser AS, Levy D. Astrocytes mediate migraine-related intracranial meningeal mechanical hypersensitivity. *Pain.* (2021) 162:2386–96. doi: 10.1097/j.pain.00000000002229
- Vecchia D, Pietrobon D. Migraine: a disorder of brain excitatory-inhibitory balance? *Trends Neurosci.* (2012) 35:507–20. doi: 10.1016/j.tins.2012.04.007
- 27. Coppola G, Schoenen J. Cortical excitability in chronic migraine. *Curr Pain Headache Rep.* (2011) 16:93–100. doi: 10.1007/s11916-011-0231-1
- Stankewitz A, May A. The phenomenon of changes in cortical excitability in migraine is not migraine-specific-a unifying thesis. *Pain.* (2009) 145:14– 7. doi: 10.1016/j.pain.2009.03.010
- Stankewitz A, Aderjan D, Eippert F, May A. Trigeminal nociceptive transmission in migraineurs predicts migraine attacks. J Neurosci. (2011) 31:1937–43. doi: 10.1523/JNEUROSCI.4496-10.2011
- Baliki MN, Baria AT, Apkarian AV. The cortical rhythms of chronic back pain. J Neurosci. (2011) 31:13981– 90. doi: 10.1523/JNEUROSCI.1984-11.2011
- Zhuo M. Cortical excitation and chronic pain. Trends Neurosci. (2008) 31:199–207. doi: 10.1016/j.tins.2008.01.003
- 32. Borsook D, Burstein R. The enigma of the dorsolateral pons as a migraine generator. *Cephalalgia*. (2012) 32:803-12. doi: 10.1177/0333102412453952
- Schulte LH, May A. The migraine generator revisited: continuous scanning of the migraine cycle over 30 days and three spontaneous attacks. *Brain.* (2016) 139:1987–93. doi: 10.1093/brain/aww097
- Schulte LH, Allers A, May A. Hypothalamus as a mediator of chronic migraine. *Neurology*. (2017) 88:2011– 6. doi: 10.1212/WNL.00000000003963
- Mehnert J, Schulte L, Timmann D, May A. Activity and connectivity of the cerebellum in trigeminal nociception. *Neuroimage*. (2017) 150:112– 8. doi: 10.1016/j.neuroimage.2017.02.023
- Stankewitz A, Keidel L, Rehm M, Irving S, Kaczmarz S, Preibisch C, et al. Migraine attacks as a result of hypothalamic loss of control. *Neuroimage Clin.* (2021) 32:102784. doi: 10.1016/j.nicl.2021.102784
- Messina R, Rocca MA, Valsasina P, Misci P, Filippi M. Clinical correlates of hypothalamic functional changes in migraine patients. *Cephalalgia*. (2022) 42:279–90. doi: 10.1177/03331024211046618
- Porcaro C, Renzo AD, Tinelli E, Lorenzo GD, Seri S, Lorenzo CD, et al. Hypothalamic structural integrity and temporal complexity of cortical information processing at rest in migraine without aura patients between attacks. *Sci Rep UK*. (2021) 11:18701. doi: 10.1038/s41598-021-98213-3

- Coppola G, Renzo AD, Petolicchio B, Tinelli E, Lorenzo CD, Serrao M, et al. Increased neural connectivity between the hypothalamus and cortical resting-state functional networks in chronic migraine. *J Neurol.* (2020) 267:185–91. doi: 10.1007/s00415-019-09571-y
- Noseda R, Lee AJ, Nir RR, Bernstein CA, Kainz VM, Bertisch SM, et al. Neural mechanism for hypothalamic-mediated autonomic responses to light during migraine. *Proc National Acad Sci USA*. (2017) 114:E5683– 92. doi: 10.1073/pnas.1708361114
- Vincent M, Hadjikhani N. The cerebellum and migraine. *Headache*. (2007) 47:820–33. doi: 10.1111/j.1526-4610.2006.00715.x
- 42. Kros L, Aristizábal CAA, Khodakhah K. Cerebellar involvement in migraine. *Cephalalgia.* (2017) 38:1782–91. doi: 10.1177/0333102417752120
- Moulton EA, Schmahmann JD, Becerra L, Borsook D. The cerebellum and pain: passive integrator or active participator? *Brain Res Rev.* (2010) 65:14–27. doi: 10.1016/j.brainresrev.2010.05.005
- Saab CY, Willis WD. The cerebellum: organization, functions and its role in nociception. *Brain Res Rev.* (2003) 42:85– 95. doi: 10.1016/S0165-0173(03)00151-6
- 45. Ju C, Bosman LWJ, Hoogland TM, Velauthapillai A, Murugesan P, Warnaar P, et al. Neurons of the inferior olive respond to broad classes of sensory input while subject to homeostatic control. *J Physiol.* (2019) 597:2483–514. doi: 10.1113/JP277413
- Apps R, Hawkes R. Cerebellar cortical organization: a one-map hypothesis. Nat Rev Neurosci. (2009) 10:670–81. doi: 10.1038/nrn2698
- Ashida R, Walsh P, Brooks JCW, Cerminara NL, Apps R, Edwards RJ. Sensory and motor electrophysiological mapping of the cerebellum in humans. *Sci Rep UK*. (2022) 12:177. doi: 10.1038/s41598-021-04220-9
- Baumann O, Borra RJ, Bower JM, Cullen KE, Habas C, Ivry RB, et al. Consensus paper: the role of the cerebellum in perceptual processes. *Cerebellum.* (2014) 14:197–220. doi: 10.1007/s12311-014-0627-7
- Ren J, Hubbard CS, Ahveninen J, Cui W, Li M, Peng X, et al. Dissociable auditory cortico-cerebellar pathways in the human brain estimated by intrinsic functional connectivity. *Cereb Cortex.* (2021) 31:bhaa398. doi: 10.1093/cercor/bhaa398
- Thier P, Markanday A. Role of the vermal cerebellum in visually guided eye movements and visual motion perception. *Annu Rev Vision Sci.* (2019) 5:247–68. doi: 10.1146/annurev-vision-091718-015000
- Chu WT, Mitchell T, Foote KD, Coombes SA, Vaillancourt DE. Functional imaging of the brainstem during visually-guided motor control reveals visuomotor regions in the pons and midbrain. *Neuroimage*. (2021) 226:117627. doi: 10.1016/j.neuroimage.2020.117627
- Costa C, Prontera P, Sarchielli P, Tonelli A, Bassi MT, Cupini LM, et al. A novel ATP1A2 gene mutation in familial hemiplegic migraine and epilepsy. *Cephalalgia*. (2013) 34:68–72. doi: 10.1177/0333102413498941
- Thomsen LL, Kirchmann M, Bjornsson A, Stefansson H, Jensen RM, Fasquel AC, et al. The genetic spectrum of a population-based sample of familial hemiplegic migraine. *Brain.* (2007) 130:346–56. doi: 10.1093/brain/ awl334
- Ducros A, Denier C, Joutel A, Cecillon M, Lescoat C, Vahedi K, et al. The clinical spectrum of familial hemiplegic migraine associated with mutations in a neuronal calcium channel. *New Engl J Med.* (2001) 345:17– 24. doi: 10.1056/NEJM200107053450103
- Pietrobon D. Familial hemiplegic migraine. *Neurotherapeutics*. (2007) 4:274– 84. doi: 10.1016/j.nurt.2007.01.008
- Obermann M, Wurthmann S, Steinberg BS, Theysohn N, Diener HC, Naegel S. Central vestibular system modulation in vestibular migraine. *Cephalalgia*. (2014) 34:1053–61. doi: 10.1177/0333102414527650
- Sandor PS, Mascia A, Seidel L, Pasqua VD, Schoenen J. Subclinical cerebellar impairment in the common types of migraine: a threedimensional analysis of reaching movements. *Ann Neurol.* (2001) 49:668– 72. doi: 10.1002/ana.1019
- Lempert T, Neuhauser H, Daroff RB. Vertigo as a symptom of migraine. *Ann* N Y Acad Sci. (2009) 1164:242–51. doi: 10.1111/j.1749-6632.2009.03852.x
- Carvalho GF, Vianna-Bell FH, Florencio LL, Pinheiro CF, Dach F, Bigal ME, et al. Presence of vestibular symptoms and related disability in migraine with and without aura and chronic migraine. *Cephalalgia*. (2018) 39:29– 37. doi: 10.1177/0333102418769948

- Bernetti L, Pellegrino C, Corbelli I, Caproni S, Eusebi P, Faralli M, et al. Subclinical vestibular dysfunction in migraineurs without vertigo: a Clinical study. Acta Neurol Scand. (2018) 138:270–7. doi: 10.1111/ane.12941
- Moulton EA, Becerra L, Johnson A, Burstein R, Borsook D. Altered hypothalamic functional connectivity with autonomic circuits and the locus coeruleus in migraine. *PLoS ONE.* (2014) 9:e95508. doi: 10.1371/journal.pone.0095508
- Maleki N, Szabo E, Becerra L, Moulton E, Scrivani SJ, Burstein R, et al. Ictal and interictal brain activation in episodic migraine: Neural basis for extent of allodynia. *PLoS ONE.* (2021) 16:e0244320. doi: 10.1371/journal.pone.0244320
- Kruit MC, Launer LJ, Ferrari MD, Buchem MA van. Brain stem and cerebellar hyperintense lesions in migraine. *Stroke*. (2006) 37:1109– 12. doi: 10.1161/01.STR.0000206446.26702.e9
- Koppen H, Boele HJ, Palm-Meinders IH, Koutstaal BJ, Horlings CG, Koekkoek BK, et al. Cerebellar function and ischemic brain lesions in migraine patients from the general population. *Cephalalgia*. (2016) 37:177– 90. doi: 10.1177/0333102416643527
- Arkink EB, Palm-Meinders IH, Koppen H, Milles J, Lew B van, Launer LJ, et al. Microstructural white matter changes preceding white matter hyperintensities in migraine. *Neurology*. (2019) 93:e688– 94. doi: 10.1212/WNL.000000000007940
- Mehnert J, May A. Functional and structural alterations in the migraine cerebellum. J Cereb Blood Flow Metab. (2019) 39:730–9. doi: 10.1177/0271678X17722109
- Liu HY, Lee PL, Chou KH, Lai KL, Wang YF, Chen SP, et al. The cerebellum is associated with 2-year prognosis in patients with high-frequency migraine. *J Headache Pain*. (2020) 21:1–10. doi: 10.1186/s10194-020-01096-4
- Kellner-Weldon F, El-Koussy M, Jung S, Jossen M, Klinger-Gratz PP, Wiest R. Cerebellar hypoperfusion in migraine attack: incidence and significance. *Am J Neuroradiol.* (2018) 39:435–40. doi: 10.3174/ajnr.A5508
- Caligiore D, Pezzulo G, Baldassarre G, Bostan AC, Strick PL, Doya K, et al. Consensus paper: towards a systems-level view of cerebellar function: the interplay between cerebellum, Basal Ganglia, and Cortex. *Cerebellum*. (2017) 16:203–29. doi: 10.1007/s12311-016-0763-3
- Miterko LN, Baker KB, Beckinghausen J, Bradnam LV, Cheng MY, Cooperrider J, et al. Consensus paper: experimental neurostimulation of the cerebellum. *Cerebellum.* (2019) 18:1064– 97. doi: 10.1007/s12311-019-01041-5
- Habas C, Manto M. Chapter 14 Probing the neuroanatomy of the cerebellum using tractography. *Handb Clin Neurol.* (2018) 154:235– 49. doi: 10.1016/B978-0-444-63956-1.00014-X
- Ashida R, Cerminara NL, Brooks J, Apps R. Principles of organization of the human cerebellum: macro- and microanatomy. In: Manto M, Huisman T, editors. *Handbook of Clinical Neurology: The Cerebellum: From Embryology* to Diagnostic Investigations, Vol. 154. (2018). p. 45–58.
- Cerminara NL, Lang EJ, Sillitoe RV, Apps R. Redefining the cerebellar cortex as an assembly of non-uniform Purkinje cell microcircuits. *Nat Rev Neurosci*. (2015) 16:79–93. doi: 10.1038/nrn3886
- Buckner RL, Krienen FM, Castellanos A, Diaz JC, Yeo BTT. The organization of the human cerebellum estimated by intrinsic functional connectivity. J Neurophysiol. (2011) 106:2322–45. doi: 10.1152/jn.00339.2011
- Cocker LJLD, Veluw SJ van, Fowkes M, Luijten PR, Mali WPThM, Hendrikse J. Very small cerebellar infarcts: integration of recent insights into a functional topographic classification. *Cerebrovasc Dis.* (2013) 36:81– 7. doi: 10.1159/000353668
- Schmahmann JD, MacMore J, Vangel M. Cerebellar stroke without motor deficit: clinical evidence for motor and non-motor domains within the human cerebellum. *Neuroscience.* (2009) 162:852–61. doi: 10.1016/j.neuroscience.2009.06.023
- Stoodley CJ, Schmahmann JD. Evidence for topographic organization in the cerebellum of motor control versus cognitive and affective processing. *Cortex.* (2010) 46:831–44. doi: 10.1016/j.cortex.2009.11.008
- Stoodley CJ, Valera EM, Schmahmann JD. Functional topography of the cerebellum for motor and cognitive tasks: an fMRI study. *NeuroImage.* (2012) 59:1560–70. doi: 10.1016/j.neuroimage.2011. 08.065

- Manto M, Bower JM, Conforto AB, Delgado-García JM, Guarda SNF da, Gerwig M, et al. Consensus paper: roles of the cerebellum in motor controlthe diversity of ideas on cerebellar involvement in movement. *Cerebellum*. (2012) 11:457–87. doi: 10.1007/s12311-011-0331-9
- Moulton EA, Elman I, Pendse G, Schmahmann J, Becerra L, Borsook D. Aversion-related circuitry in the cerebellum: responses to noxious heat and unpleasant images. *J Neurosci.* (2011) 31:3795–804. doi: 10.1523/JNEUROSCI.6709-10.2011
- Sacchetti B, Scelfo B, Strata P. Cerebellum and emotional behavior. Neuroscience. (2009) 162:756–62. doi: 10.1016/j.neuroscience.2009.01.064
- Voogd J, Schraa-Tam CKL, Geest JN van der, Zeeuw CID. Visuomotor cerebellum in human and nonhuman primates. *Cerebellum Lond Engl.* (2012) 11:392–410. doi: 10.1007/s12311-010-0204-7
- Schmahmann JD. Disorders of the cerebellum: ataxia, dysmetria of thought, and the cerebellar cognitive affective syndrome. J Neuropsychiatry Clin Neurosci. (2004) 16:367–78. doi: 10.1176/jnp.16.3.367
- Manni E, Petrosini L. A century of cerebellar somatotopy: a debated representation. Nat Rev Neurosci. (2004) 5:241–9. doi: 10.1038/nrn1347
- Bower JM. The 40-year history of modeling active dendrites in cerebellar Purkinje cells: emergence of the first single cell "community model." Front Comput Neurosci. (2015). 9:129. doi: 10.3389/fncom.2015.00129
- Chadderton P, Margrie TW, Häusser M. Integration of quanta in cerebellar granule cells during sensory processing. *Nature*. (2004) 428:856– 60. doi: 10.1038/nature02442
- Zeeuw CID, Hoogenraad CC, Koekkoek SKE, Ruigrok TJH, Galjart N, Simpson JI. Microcircuitry and function of the inferior olive. *Trends Neurosci.* (1998) 21:391–400. doi: 10.1016/S0166-2236(98)01310-1
- Zeeuw CID, Hoogland TM. Reappraisal of Bergmann glial cells as modulators of cerebellar circuit function. *Front Cell Neurosci.* (2015) 9:246. doi: 10.3389/fncel.2015.00246
- Kalinichenko SG, Pushchin II. The modular architecture and neurochemical patterns in the cerebellar cortex. J Chem Neuroanat. (2018) 92:16– 24. doi: 10.1016/j.jchemneu.2018.05.001
- Balmer TS, Trussell L. Selective targeting of unipolar brush cell subtypes by cerebellar mossy fibers. *eLife*. (2019) 8:e44964. doi: 10.1101/523266
- Buffo A, Rossi F. Origin, lineage and function of cerebellar glia. Prog Neurobiol. (2013) 109:42–63. doi: 10.1016/j.pneurobio.2013.08.001
- Consalez GG, Goldowitz D, Casoni F, Hawkes R. Origins, development, and compartmentation of the granule cells of the cerebellum. *Front Neural Circuit.* (2021) 14:611841. doi: 10.3389/fncir.2020.611841
- Ruigrok TJH, Voogd J. Organization of projections from the inferior olive to the cerebellar nuclei in the rat. J Comp Neurol. (2000) 426:209– 28. doi: 10.1002/1096-9861(20001016)426:2<209::AID-CNE4>3.0.CO;2-0
- Ruigrok TJH, Voogd J. Cerebellar nucleo-olivary projections in the rat: an anterograde tracing study with Phaseolus vulgaris-leucoagglutinin (PHA-L). *J Comp Neurol.* (1990) 298:315–33. doi: 10.1002/cne.902980305
- Ruigrok TJH, Teune TM. Collateralization of cerebellar output to functionally distinct brainstem areas. A retrograde, nonfluorescent tracing study in the rat. *Front Syst Neurosci.* (2014) 8:23. doi: 10.3389/fnsys.2014.00023
- 96. Zeeuw CID, Berrebi AS. Postsynaptic targets of purkinje cell terminals in the cerebellar and vestibular nuclei of the rat. *Eur J Neurosci.* (1995) 7:2322–33. doi: 10.1111/j.1460-9568.1995.tb0 0653.x
- Reeber SL, White JJ, George-Jones NA, Sillitoe RV. Architecture and development of olivocerebellar circuit topography. *Front Neural Circuit*. (2013) 6:115. doi: 10.3389/fncir.2012.00115
- Noseda R, Constandil L, Bourgeais L, Chalus M, Villanueva L. Changes of meningeal excitability mediated by corticotrigeminal networks: a link for the endogenous modulation of migraine pain. *J Neurosci.* (2010) 30:14420– 9. doi: 10.1523/JNEUROSCI.3025-10.2010
- 99. Furuta T, Urbain N, Kaneko T, Deschênes M. Corticofugal control of vibrissa-sensitive neurons in the interpolaris nucleus of the trigeminal complex. J Neurosci. (2010) 30:1832– 8. doi: 10.1523/JNEUROSCI.4274-09.2010
- 100. Desbois C, Bars DL, Villanueva L. Organization of cortical projections to the medullary subnucleus reticularis dorsalis: a retrograde and

anterograde tracing study in the rat. J Compar Neurol. (1999) 410:178– 96. doi: 10.1002/(SICI)1096-9861(19990726)410:2<178::AID-CNE2>3.0. CO;2-F

- 101. Kratochwil CF, Maheshwari U, Rijli FM. The long journey of pontine nuclei neurons: from rhombic lip to cortico-ponto-cerebellar circuitry. *Front Neural Circuit.* (2017) 11:33. doi: 10.3389/fncir.2017.00033
- Swenson RS, Sievert CF, Terreberry RR, Neafsey EJ, Castro AJ. Organization of cerebral cortico-olivary projections in the rat. *Neurosci Res.* (1989) 7:43– 54. doi: 10.1016/0168-0102(89)90036-9
- Saint-Cyr JA. The projection from the motor cortex to the inferior olive in the cat. An experimental study using axonal transport techniques. *Neuroscience*. (1983) 10:667–84. doi: 10.1016/0306-4522(83)90209-9
- Nguyen-Vu TDB, Kimpo RR, Rinaldi JM, Kohli A, Zeng H, Deisseroth K, et al. Cerebellar Purkinje cell activity drives motor learning. *Nat Neurosci.* (2013) 16:1734–6. doi: 10.1038/nn.3576
- 105. Shimuta M, Sugihara I, Ishikawa T. Multiple signals evoked by unisensory stimulation converge onto cerebellar granule and Purkinje cells in mice. *Commun Biol.* (2020) 3:381. doi: 10.1038/s42003-020-1110-2
- Barmack NH, Yakhnitsa V. Climbing fibers mediate vestibular modulation of both "complex" and "simple spikes" in purkinje cells. *Cerebellum.* (2015) 14:597–612. doi: 10.1007/s12311-015-0725-1
- 107. Kagan R, Kainz V, Burstein R, Noseda R. Hypothalamic and basal ganglia projections to the posterior thalamus: possible role in modulation of migraine headache and photophobia. *Neuroscience.* (2013) 248:359– 68. doi: 10.1016/j.neuroscience.2013.06.014
- 108. Noseda R, Kainz V, Borsook D, Burstein R. Neurochemical pathways that converge on thalamic trigeminovascular neurons: potential substrate for modulation of migraine by sleep, food intake, stress and anxiety. *PLoS ONE.* (2014) 9:e103929. doi: 10.1371/journal.pone.0103929
- Noseda R, Borsook D, Burstein R. Neuropeptides and neurotransmitters that modulate thalamo-cortical pathways relevant to migraine headache. *Headache.* (2017) 57:97–111. doi: 10.1111/head.13083
- Ito M. Functional roles of neuropeptides in cerebellar circuits. *Neuroscience*. (2009) 162:666–72. doi: 10.1016/j.neuroscience.2009.01.019
- 111. Onat F, Çavdar S. Cerebellar connections: hypothalamus. *Cerebellum*. (2003) 2:263–9. doi: 10.1080/14734220310016187
- Hirono M, Karube F, Yanagawa Y. Modulatory effects of monoamines and perineuronal nets on output of cerebellar purkinje cells. *Front Neural Circuit.* (2021) 15:661899. doi: 10.3389/fncir.2021.661899
- 113. Li B, Zhu JN, Wang JJ. Histaminergic afferent system in the cerebellum: structure and function. *Cerebellum Ataxias*. (2014) 1:5. doi: 10.1186/2053-8871-1-5
- 114. Haines DE, Dietrichs E, Mihailoff GA, McDonald EF. The cerebellarhypothalamic axis: basic circuits and clinical observations. *Int Rev Neurobiol.* (1997) 41:83–107. doi: 10.1016/S0074-7742(08)60348-7
- 115. Zagrebelsky M, Rossi F. Postnatal development and adult organisation of the olivocerebellar projection map in the hypogranular cerebellum of the rat. *J Comp Neurol.* (1999) 407:527–42. doi: 10.1002/(SICI)1096-9861(19990517)407:4<527::AID-CNE5>3.0. CO;2-H
- 116. Morara S, Want JJL van der, Weerd H de, Provini L, Rosina A. Ultrastructural analysis of climbing fiber-Purkinje cell synaptogenesis in the rat cerebellum. *Neuroscience*. (2001) 108:655–71. doi: 10.1016/S0306-4522(01)0 0433-X
- 117. Edvinsson L, Eftekhari S, Salvatore CA, Warfvinge K. Cerebellar distribution of calcitonin gene-related peptide (CGRP) and its receptor components calcitonin receptor-like receptor (CLR) and receptor activity modifying protein 1 (RAMP1) in rat. *Mol Cell Neurosci.* (2011) 46:333–9. doi: 10.1016/j.mcn.2010. 10.005
- 118. Moore EL, Burgey CS, Paone DV, Shaw AW, Tang YS, Kane SA, et al. Examining the binding properties of MK-0974: A CGRP receptor antagonist for the acute treatment of migraine. *Eur J Pharmacol.* (2009) 602:250– 4. doi: 10.1016/j.ejphar.2008.11.050
- 119. Villalba M, Bockaert J, Journot L. Pituitary adenylate cyclase-activating polypeptide (PACAP-38) protects cerebellar granule neurons from apoptosis by activating the mitogen-activated protein kinase (MAP kinase) pathway. J Neurosci. (1997) 17:83–90. doi: 10.1523/JNEUROSCI.17-01-00083.1997

- 120. Tolner EA, Houben T, Terwindt GM, Vries B de, Ferrari MD, Maagdenberg AMJM van den. From migraine genes to mechanisms. *Pain*. (2015) 156:S64– 74. doi: 10.1097/01.j.pain.0000460346.00213.16
- 121. Hillman D, Chen S, Aung TT, Cherksey B, Sugimori M, Llinás RR. Localization of P-type calcium channels in the central nervous system. *Proc Natl Acad Sci USA*. (1991) 88:7076–80. doi: 10.1073/pnas.88.16.7076
- Satake S, Saitow F, Yamada J, Konishi S. Synaptic activation of AMPA receptors inhibits GABA release from cerebellar interneurons. *Nat Neurosci.* (2000) 3:551–8. doi: 10.1038/75718
- 123. Saitow F, Satake S, Yamada J, Konishi S. β-adrenergic receptor-mediated presynaptic facilitation of inhibitory GABAergic transmission at cerebellar interneuron-purkinje cell synapses. J Neurophysiol. (2000) 84:2016– 25. doi: 10.1152/jn.2000.84.4.2016
- 124. Zhang C, Zhou P, Yuan T. The cholinergic system in the cerebellum: from structure to function. *Rev Neurosci.* (2016) 27:769–76. doi: 10.1515/revneuro-2016-0008
- 125. Vitale F, Mattei C, Capozzo A, Pietrantoni I, Mazzone P, Scarnati E. Cholinergic excitation from the pedunculopontine tegmental nucleus to the dentate nucleus in the rat. *Neuroscience.* (2016) 317:12–22. doi: 10.1016/j.neuroscience.2015.12.055
- 126. Flace P, Livrea P, Basile GA, Galletta D, Bizzoca A, Gennarini G, et al. The cerebellar dopaminergic system. *Front Syst Neurosci.* (2021) 15:650614. doi: 10.3389/fnsys.2021.650614
- 127. Schweighofer N, Doya K, Kuroda S. Cerebellar aminergic neuromodulation: towards a functional understanding. *Brain Res Rev.* (2004) 44:103– 16. doi: 10.1016/j.brainresrev.2003.10.004
- Snyder RL, Faull RLM, Mehler WR. A comparative study of the neurons of origin of the spinocerebellar afferents in the rat, cat and squirrel monkey based on the retrograde transport of horseradish peroxidase. *J Comp Neurol.* (1978) 181:833–52. doi: 10.1002/cne.901810409
- 129. Jacquin MF, Semba K, Rhoades RW, Egger MD. Trigeminal primary afferents project bilaterally to dorsal horn and ipsilaterally to cerebellum, reticular formation, and cuneate, solitary, supratrigeminal and vagal nuclei. *Brain Res.* (2002) 246:285–91. doi: 10.1016/0006-8993(82)91177-5
- Marfurt CF, Rajchert DM. Trigeminal primary afferent projections to "nontrigeminal" areas of the rat central nervous system. *J Comp Neurol.* (1991) 303:489–511. doi: 10.1002/cne.903030313
- 131. Ikeda M. Projections from the spinal and the principal sensory nuclei of the trigeminal nerve to the cerebellar cortex in the cat, as studied by retrograde transport of horseradish peroxidase. *Neurosci Lett.* (1979) 11:31. doi: 10.1016/0304-3940(79)91594-5
- 132. Oscarsson O. Functional organization of the spino- and cuneocerebellar tracts. *Physiol Rev.* (1965) 45:495–522. doi: 10.1152/physrev.1965.45.3.495
- Lazarov NE. Comparative analysis of the chemical neuroanatomy of the mammalian trigeminal ganglion and mesencephalic trigeminal nucleus. *Prog Neurobiol.* (2002) 66:19–59. doi: 10.1016/S0301-0082(01)00021-1
- Pinganaud G, Bourcier F, Buisseret-Delmas C, Buisseret P. Primary trigeminal afferents to the vestibular nuclei in the rat: existence of a collateral projection to the vestibulo-cerebellum. *Neurosci Lett.* (1999) 264:133– 6. doi: 10.1016/S0304-3940(99)00179-2
- 135. Huerta MF, Frankfurter A, Harting JK. Studies of the principal sensory and spinal trigeminal nuclei of the rat: projections to the superior colliculus, inferior olive, and cerebellum. J Comp Neurol. (1983) 220:147– 67. doi: 10.1002/cne.902200204
- 136. Yatim N, Billig I, Compoint C, Buisseret P, Buisseret-Delmas C. Trigeminocerebellar and trigemino-olivary projections in rats. *Neurosci Res.* (1996) 25:267–83. doi: 10.1016/0168-0102(96)01061-9
- 137. Walberg F. The trigemino-olivary projection in the cat as studied with retrograde transport of horseradish peroxidase. *Exp Brain Res.* (1982) 45:101–7. doi: 10.1007/BF00235768
- Brown JT, Chan-Palay V, Palay S. A study of afferent input to the inferior olivary complex in the rat. J Compar Neurol. (1977) 176:1– 22. doi: 10.1002/cne.901760102
- Killackey HP, Erzurumlu RS. Trigeminal projections to the superior colliculus of the rat. J Comp Neurol. (1981) 201:221– 42. doi: 10.1002/cne.902010207
- 140. Guell X, Schmahmann JD, Gabrieli JD, Ghosh SS. Functional gradients of the cerebellum. *Elife*. (2018) 7:e36652. doi: 10.7554/eLife.36652

- D'Angelo E. Physiology of the cerebellum. In: Manto M, Huisman T, editors. Handbook of Clinical Neurology: The Cerebellum: From Embryology to Diagnostic Investigations, Vol. 154. (2018). p. 85–108.
- 142. Barmack NH. Central vestibular system: vestibular nuclei and posterior cerebellum. *Brain Res Bull.* (2003) 60:511– 41. doi: 10.1016/S0361-9230(03)00055-8
- 143. Maklad A, Fritzsch B. Partial segregation of posterior crista and saccular fibers to the nodulus and uvula of the cerebellum in mice, and its development. *Dev Brain Res.* (2003) 140:223– 36. doi: 10.1016/S0165-3806(02)00609-0
- 144. Patel VR, Zee DS. The cerebellum in eye movement control: nystagmus, coordinate frames and disconjugacy. *Eye.* (2014) 29:191–5. doi: 10.1038/eye.2014.271
- 145. Ango F, Reis RD. Sensing how to balance. *Elife.* (2019) 8:e46973. doi: 10.7554/eLife.46973
- 146. Middleton FA, Strick PL. Cerebellar output channels. Int Rev Neurobiol. (1997) 41:61–82. doi: 10.1016/S0074-7742(08)60347-5
- Middleton FA, Strick PL. Cerebellar output: motor and cognitive channels. Trends Cogn Sci. (1998) 2:348–54. doi: 10.1016/S1364-6613(98)01220-0
- 148. Buisseret-Delmas C, Angaut P. The cerebellar olivocorticonuclear connections in the rat. *Prog Neurobiol.* (1993) 40:63–87. doi: 10.1016/0301-0082(93)90048-W
- 149. Teune TM, Burg J van der, Moer J van der, Voogd J, Ruigrok TJH. Topography of cerebellar nuclear projections to the brain stem in the rat. *Prog Brain Res.* (2000) 124:141–72. doi: 10.1016/S0079-6123(00)24014-4
- 150. Homma Y, Nonaka S, Matsuyama K, Mori S. Fastigiofugal projection to the brainstem nuclei in the cat: an anterograde PHA-L tracing study. *Neurosci Res.* (1995) 23:89–102. doi: 10.1016/0168-0102(95)90019-5
- Bengtsson F, Hesslow G. Cerebellar control of the inferior olive. *Cerebellum*. (2005) 5:7. doi: 10.1080/14734220500462757
- Bagnall MW, Zingg B, Sakatos A, Moghadam SH, Zeilhofer HU, Lac S d. Glycinergic projection neurons of the cerebellum. *J Neurosci.* (2009) 29:10104–10. doi: 10.1523/JNEUROSCI.2087-09.2009
- 153. Husson Z, Rousseau CV, Broll I, Zeilhofer HU, Dieudonné S. Differential GABAergic and glycinergic inputs of inhibitory interneurons and Purkinje cells to principal cells of the cerebellar nuclei. *J Neurosci.* (2014) 34:9418– 31. doi: 10.1523/JNEUROSCI.0401-14.2014
- 154. Voogd J, Koehler PJ. Historic notes on anatomic, physiologic, and clinical research on the cerebellum. In: Manto M, Huisman T, editors. *Handbook* of Clinical Neurology: The Cerebellum: From Embryology to Diagnostic Investigations, Vol. 154. (2018). p. 3–26.
- 155. Judd EN, Lewis SM, Person AL. Diverse inhibitory projections from the cerebellar interposed nucleus. *Elife.* (2021) 10:e66231. doi: 10.7554/eLife.66231
- 156. Kang S, Jun S, Baek SJ, Park H, Yamamoto Y, Tanaka-Yamamoto K. Recent advances in the understanding of specific efferent pathways emerging from the cerebellum. *Front Neuroanat.* (2021) 15:759948. doi: 10.3389/fnana.2021.759948
- Kheradmand A, Zee DS. Cerebellum and ocular motor control. Front Neurol. (2011) 2:53. doi: 10.3389/fneur.2011.00053
- Beh SC, Frohman TC, Frohman EM. Cerebellar control of eye movements. J Neuro-ophthalmol. (2017) 37:87–98. doi: 10.1097/WNO.000000000000456
- 159. Basile GA, Quartu M, Bertino S, Serra MP, Boi M, Bramanti A, et al. Red nucleus structure and function: from anatomy to clinical neurosciences. *Brain Struct Funct*. (2021) 226:69–91. doi: 10.1007/s00429-020-02171-x
- 160. Fujita H, Kodama T, Lac S du. Modular output circuits of the fastigial nucleus for diverse motor and nonmotor functions of the cerebellar vermis. *Elife.* (2020) 9:e58613. doi: 10.7554/eLife.58613
- 161. Onodera S, Hicks TP. A comparative neuroanatomical study of the red nucleus of the cat, macaque and human. *PLoS ONE*. (2009) 4:e6623. doi: 10.1371/journal.pone.0006623
- Baek SJ, Park J, Kim J, Yamamoto Y, Tanaka-Yamamoto K. VTA-projecting cerebellar neurons mediate stress-dependent depression-like behaviors. *Biorxiv.* (2021) 11:e72981. doi: 10.7554/eLife.72981.sa2
- 163. Cerminara NL, Koutsikou S, Lumb BM, Apps R. The periaqueductal grey modulates sensory input to the cerebellum: a role in coping behaviour? *Eur J Neurosci.* (2009) 29:2197–206. doi: 10.1111/j.1460-9568.2009.06760.x

- 164. Stapley PJ, Drew T. The pontomedullary reticular formation contributes to the compensatory postural responses observed following removal of the support surface in the standing cat. J Neurophysiol. (2009) 101:1334– 50. doi: 10.1152/jn.91013.2008
- 165. Prentice SD, Drew T. Contributions of the reticulospinal system to the postural adjustments occurring during voluntary gait modifications. J Neurophysiol. (2001) 85:679–98. doi: 10.1152/jn.2001.85.2.679
- 166. Younis S, Hougaard A, Noseda R, Ashina M. Current understanding of thalamic structure and function in migraine. *Cephalalgia*. (2018) 39:1675– 82. doi: 10.1177/0333102418791595
- 167. Noseda R, Kainz V, Jakubowski M, Gooley JJ, Saper CB, Digre K, et al. A neural mechanism for exacerbation of headache by light. *Nat Neurosci.* (2010) 13:239–45. doi: 10.1038/nn.2475
- 168. Bay HH, Özkan M, Onat F, Çavdar S. Do the dento-thalamic connections of genetic absence epilepsy rats from strasbourg differ from those of control wistar rats? *Brain Connectivity.* (2019) 9:703-10. doi: 10.1089/brain.2019.0694
- Dum RP, Strick PL. Medial wall motor areas and skeletomotor control. CurrOpinNeurobiol. (1992) 2:836–9. doi: 10.1016/0959-4388(92)90142-8
- 170. Dum RP, Strick PL. Spinal cord terminations of the medial wall motor areas in macaque monkeys. J Neurosci. (1996) 16:6513–25. doi: 10.1523/JNEUROSCI.16-20-06513.1996
- Dum RP, Strick PL. An unfolded map of the cerebellar dentate nucleus and its projections to the cerebral cortex. J Neurophysiol. (2003) 89:634– 9. doi: 10.1152/jn.00626.2002
- 172. Middleton FA, Strick PL. Dentate output channels: motor and cognitive components. *Prog Brain Res.* (1997) 114:553– 66. doi: 10.1016/S0079-6123(08)63386-5
- 173. Palesi F, Tournier JD, Calamante F, Muhlert N, Castellazzi G, Chard D, et al. Contralateral cerebello-thalamo-cortical pathways with prominent involvement of associative areas in humans *in vivo. Brain Struct Funct.* (2015) 220:3369–84. doi: 10.1007/s00429-014-0861-2
- 174. Strick PL, Dum RP, Fiez JA. Cerebellum and nonmotor function. Annu Rev Neurosci. (2009) 32:413–34. doi: 10.1146/annurev.neuro.31.060407.125606
- Zhang XY, Wang JJ, Zhu JN. Cerebellar fastigial nucleus: from anatomic construction to physiological functions. *Cerebellum Ataxias*. (2016) 3:9. doi: 10.1186/s40673-016-0047-1
- 176. Bostan AC, Strick PL. The basal ganglia and the cerebellum: nodes in an integrated network. Nat Rev Neurosci. (2018) 19:338–50. doi: 10.1038/s41583-018-0002-7
- Hoshi E, Tremblay L, Féger J, Carras PL, Strick PL. The cerebellum communicates with the basal ganglia. *Nat Neurosci.* (2005) 8:1491– 3. doi: 10.1038/nn1544
- Pelzer EA, Melzer C, Timmermann L, Cramon DY von, Tittgemeyer M. Basal ganglia and cerebellar interconnectivity within the human thalamus. *Brain Struct Funct.* (2016) 222:381–92. doi: 10.1007/s00429-016-1223-z
- Arber S, Costa RM. Networking brainstem and basal ganglia circuits for movement. *Nat Rev Neurosci.* (2022) 23:342–60. doi: 10.1038/s41583-022-0 0581-w
- 180. Foster NN, Barry J, Korobkova L, Garcia L, Gao L, Becerra M, et al. The mouse cortico-basal ganglia-thalamic network. *Nature*. (2021) 598:188– 94. doi: 10.1038/s41586-021-03993-3
- 181. Bostan AC, Dum RP, Strick PL. Cerebellar networks with the cerebral cortex and basal ganglia. *Trends Cogn Sci.* (2013) 17:241-54. doi: 10.1016/j.tics.2013.03.003
- 182. Tanaka H, Ishikawa T, Lee J, Kakei S. The cerebro-cerebellum as a locus of forward model: a review. *Front Syst Neurosci.* (2020) 14:19. doi: 10.3389/fnsys.2020.00019
- Schmahmann JD, Pandya DN. Disconnection syndromes of basal ganglia, thalamus, and cerebrocerebellar systems. *Cortex.* (2008) 44:1037–66. doi: 10.1016/j.cortex.2008.04.004
- 184. Suter BA, Shepherd GMG. Reciprocal interareal connections to corticospinal neurons in mouse M1 and S2. J Neurosci. (2015) 35:2959–74. doi: 10.1523/JNEUROSCI.4287-14.2015
- Deschênes M, Veinante P, Zhang ZW. The organization of corticothalamic projections: reciprocity versus parity. *Brain Res Rev.* (1998) 28:286– 308. doi: 10.1016/S0165-0173(98)00017-4

- 186. Giolli RA, Gregory KM, Suzuki DA, Blanks RHI, Lui F, Betelak KF. Cortical and subcortical afferents to the nucleus reticularis tegmentipontis and basal pontine nuclei in the macaque monkey. *Visual Neurosci.* (2001) 18:725– 40. doi: 10.1017/S0952523801185068
- Jacquin MF, Wiegand MR, Renehan WE. Structure-function relationships in rat brain stem subnucleus interpolaris. VIII. Cortical inputs. J Neurophysiol. (1990) 64:3–27. doi: 10.1152/jn.1990.64.1.3
- Gao Z, Davis C, Thomas AM, Economo MN, Abrego AM, Svoboda K, et al. A cortico-cerebellar loop for motor planning. *Nature*. (2018) 563:1– 27. doi: 10.1038/s41586-018-0633-x
- Lambert GA, Hoskin KL, Zagami AS. Cortico-NRM influences on trigeminal neuronal sensation. *Cephalalgia*. (2008) 28:640– 52. doi: 10.1111/j.1468-2982.2008.01572.x
- Kuypers HGJM. An anatomical analysis of cortico-bulbar connexions to the pons and lower brain stem in the cat. J Anatomy. (1958) 92:198–218.
- 191. Glickstein M, Gerrits N, Kralj-Hans I, Mercier B, Stein J, Voogd J. Visual pontocerebellar projections in the macaque. J Comp Neurol. (1994) 349:51– 72. doi: 10.1002/cne.903490105
- 192. Cacciola A, Milardi D, Basile GA, Bertino S, Calamuneri A, Chillemi G, et al. The cortico-rubral and cerebello-rubral pathways are topographically organized within the human red nucleus. *Sci Rep UK*. (2019) 9:12117. doi: 10.1038/s41598-019-48164-7
- 193. Palesi F, Rinaldis AD, Castellazzi G, Calamante F, Muhlert N, Chard D, et al. Contralateral cortico-ponto-cerebellar pathways reconstruction in humans *in vivo*: implications for reciprocal cerebro-cerebellar structural connectivity in motor and non-motor areas. *Sci Rep UK*. (2017) 7:12841. doi: 10.1038/s41598-017-13079-8
- 194. Benagiano V, Rizzi A, Lorusso L, Flace P, Saccia M, Cagiano R, et al. The functional anatomy of the cerebrocerebellar circuit: a review and new concepts. J Comp Neurol. (2018) 526:769–89. doi: 10.1002/cne.24361
- 195. Stoodley CJ, Schmahmann JD. Functional topography of the human cerebellum. In: Manto M, Huisman T, editors. *Handbook of Clinical Neurology: The Cerebellum: From Embryology to Diagnostic Investigations*, Vol. 154. 2018, p. 59–70.
- 196. Burstein R, Yamamura H, Malick A, Strassman AM. Chemical stimulation of the intracranial dura induces enhanced responses to facial stimulation in brain stem trigeminal neurons. *J Neurophysiol.* (1998) 79:964– 82. doi: 10.1152/jn.1998.79.2.964
- 197. Borsook D, Burstein R, Becerra L. Functional imaging of the human trigeminal system: opportunities for new insights into pain processing in health and disease. *J Neurobiol.* (2004) 61:107–25. doi: 10.1002/neu.20085
- 198. May A. New insights into headache: an update on functional and structural imaging findings. Nat Rev Neurol. (2009) 5:199–209. doi: 10.1038/nrneurol.2009.28
- 199. Goadsby PJ, Charbit AR, Andreou AP, Akerman S, Holland PR. Neurobiology of migraine. *Neuroscience*. (2009) 161:327– 41. doi: 10.1016/j.neuroscience.2009.03.019
- 200. Noseda R, Monconduit L, Constandil L, Chalus M, Villanueva L. Central nervous system networks involved in the processing of meningeal and cutaneous inputs from the ophthalmic branch of the trigeminal nerve in the rat. *Cephalalgia*. (2008) 28:813–24. doi: 10.1111/j.1468-2982.2008.01588.x
- 201. Woda A. Pain in the trigeminal system: from orofacial nociception to neural network modeling. J Dent Res. (2003) 82:764–8. doi: 10.1177/154405910308201001
- 202. Jacquin MF, Chiaia NL, Haring JH, Rhoades RW. Intersubnuclear connections within the rat trigeminal brainstem complex. Somatosens Mot Res. (2009) 7:399–420. doi: 10.3109/08990229009144716
- 203. Villanueva L, Noseda R. Trigeminal mechanisms of nociception. In: McMahon S, Koltzenburg M, Tracey I, Turk DC, editors. *Wall and Melzack's Textbook of Pain*. 6th ed. Elsevier (2014). p. 793–802.
- 204. Henssen DJHA, Kurt E, Kozicz T, Dongen R van, Bartels RHMA, Walsum AM van C van. New insights in trigeminal anatomy: a double orofacial tract for nociceptive input. *Front Neuroanat.* (2016) 10:53. doi: 10.3389/fnana.2016.00053
- 205. Upadhyay J, Knudsen J, Anderson J, Becerra L, Borsook D. Non-invasive mapping of human trigeminal brainstem pathways. *Magnetic Resonance Med.* (2008) 60:1037. doi: 10.1002/mrm. 21682

- Villanueva L, Cliffer KD, Sorkin LS, Bars DL, Willis WD. Convergence of heterotopic nociceptive information onto neurons of caudal medullary reticular formation in monkey (*Macaca fascicularis*). J Neurophysiol. (1990) 63:1118–27. doi: 10.1152/jn.1990.63.5.1118
- 207. Villanueva L, Bouhassira D, Bing Z, Bars DL. Convergence of heterotopic nociceptive information onto subnucleus reticularis dorsalis neurons in the rat medulla. J Neurophysiol. (1988) 60:980–1009. doi: 10.1152/jn.1988.60.3.980
- Bouhassira D, Bars DL, Villanueva L. Heterotopic activation of A delta and C fibres triggers inhibition of trigeminal and spinal convergent neurones in the rat. J Physiol. (1987) 389:301–17. doi: 10.1113/jphysiol.1987.sp016658
- Ekerot CF, Oscarsson O, Schouenborg J. Stimulation of cat cutaneous nociceptive C fibres causing tonic and synchronous activity in climbing fibres. J Physiol. (1987) 386:539–46. doi: 10.1113/jphysiol.1987.sp016550
- Ekerot CF, Garwicz M, Schouenborg J. Topography and nociceptive receptive fields of climbing fibers projecting to the cerebellar anterior lobe in the cat. J Physiol. (1991) 441:257–74. doi: 10.1113/jphysiol.1991.sp018750
- 211. Garwicz M, Jörntell H, Ekerot C. Cutaneous receptive fields and topography of mossy fibres and climbing fibres projecting to cat cerebellar C3 zone. J Physiol. (1998) 512:277–93. doi: 10.1111/j.1469-7793.1998.277bf.x
- 212. Ness TJ, Follett KA, Piper J, Dirks BA. Characterization of neurons in the area of the medullary lateral reticular nucleus responsive to noxious visceral and cutaneous stimuli. *Brain Res.* (1998) 802:163– 74. doi: 10.1016/S0006-8993(98)00608-8
- Jie W, Pei-Xi C. Cerebellar evoked potential elicited by stimulation of C-fiber in saphenous nerve of cat. *Brain Res.* (1990) 522:144– 6. doi: 10.1016/0006-8993(90)91590-D
- 214. Lawrenson C, Paci E, Pickford J, Drake RA, Lumb BM, Apps R. Cerebellar modulation of memory encoding in the periaqueductal grey and fear behaviour. *Elife.* (2022) 11:e76278. doi: 10.7554/eLife.76278
- 215. Frontera JL, Aissa HB, Sala RW, Mailhes-Hamon C, Georgescu IA, Léna C, et al. Bidirectional control of fear memories by cerebellar neurons projecting to the ventrolateral periaqueductal grey. *Nat Commun.* (2020) 11:5207. doi: 10.1038/s41467-020-18953-0
- Vaaga CE, Brown ST, Raman IM. Cerebellar modulation of synaptic input to freezing-related neurons in the periaqueductal gray. *Elife*. (2020) 9:e54302. doi: 10.7554/eLife.54302
- 217. Koutsikou S, Crook JJ, Earl EV, Leith JL, Watson TC, Lumb BM, et al. Neural substrates underlying fear-evoked freezing: the periaqueductal grey-cerebellar link. *J Physiol.* (2014) 592(Pt 10):2197–213. doi: 10.1113/jphysiol.2013.268714
- 218. Bambico FR, Comai S, Diwan M, Hasan SMN, Conway JD, Darvish-Ghane S, et al. High frequency stimulation of the anterior vermis modulates behavioural response to chronic stress: involvement of the prefrontal cortex and dorsal raphe? *Neurobiol Dis.* (2018) 116:166–78. doi: 10.1016/j.nbd.2018.03.011
- Lamotte G, Shouman K, Benarroch EE. Stress and central autonomic network. *Autonomic Neurosci.* (2021) 235:102870. doi: 10.1016/j.autneu.2021.102870
- 220. Schwarz LA, Luo L. Organization of the locus coeruleus-norepinephrine system. *Curr Biol.* (2015) 25:R1051–6. doi: 10.1016/j.cub.2015.09.039
- 221. Ross JA, Bockstaele EJV. The locus coeruleus- norepinephrine system in stress and arousal: unraveling historical, current, and future perspectives. *Front Psychiatry*. (2021) 11:601519. doi: 10.3389/fpsyt.2020.601519
- Vila-Pueyo M, Strother LC, Kefel M, Goadsby PJ, Holland PR. Divergent influences of the locus coeruleus on migraine pathophysiology. *Pain.* (2019) 160:385–94. doi: 10.1097/j.pain.00000000001421
- 223. Welman FHSM, Smit AE, Jongen JLM, Tibboel D, Geest JN van der, Holstege JC. Pain experience is somatotopically organized and overlaps with pain anticipation in the human cerebellum. *Cerebellum.* (2018) 17:447– 60. doi: 10.1007/s12311-018-0930-9
- 224. Maleki N, Becerra L, Nutile L, Pendse G, Brawn J, Bigal M, et al. Migraine attacks the Basal Ganglia. *Molecular Pain.* (2011) 7:71. doi: 10.1186/1744-8069-7-71
- 225. Schwedt TJ, Chong CD, Chiang CC, Baxter L, Schlaggar BL, Dodick DW. Enhanced pain-induced activity of pain-processing regions in a case-control study of episodic migraine. *Cephalalgia.* (2014) 34:947–58. doi: 10.1177/0333102414526069

- Silva KE, Rosner J, Ullrich NJ, Chordas C, Manley PE, Moulton EA. Pain affect disrupted in children with posterior cerebellar tumor resection. *Ann Clin Transl Neur.* (2019) 6:344–54. doi: 10.1002/acn3.709
- Ruscheweyh R, Kühnel M, Filippopulos F, Blum B, Eggert T, Straube A. Altered experimental pain perception after cerebellar infarction. *Pain.* (2014) 155:1303–12. doi: 10.1016/j.pain.2014.04.006
- Guell X, Schmahmann J. Cerebellar functional anatomy: a didactic summary based on human fMRI evidence. *Cerebellum.* (2020) 19:1– 5. doi: 10.1007/s12311-019-01083-9
- 229. Sarnat HB. Cerebellar networks and neuropathology of cerebellar developmental disorders. In: Manto M, Huisman T, editors. *Handbook* of Clinical Neurology: The Cerebellum: From Embryology to Diagnostic Investigations, Vol. 154. (2018). p. 109–28.
- 230. Harno H, Hirvonen T, Kaunisto MA, Aalto H, Levo H, Isotalo E, et al. Subclinical vestibulocerebellar dysfunction in migraine with and without aura. *Neurology*. (2003) 61:1748– 52. doi: 10.1212/01.WNL.0000098882.82690.65
- Pelzer N, Haan J, Stam AH, Vijfhuizen LS, Koelewijn SC, Smagge A, et al. Clinical spectrum of hemiplegic migraine and chances of finding a pathogenic mutation. *Neurology.* (2018) 90:e575–82. doi: 10.1212/WNL.000000000004966
- 232. Searls DE, Pazdera L, Korbel E, Vysata O, Caplan LR. Symptoms and signs of posterior circulation ischemia in the new england medical center posterior circulation registry. *Arch Neurol Chicago*. (2012) 69:346–51. doi: 10.1001/archneurol.2011.2083
- Cocker LJLD, Geerlings MI, Hartkamp NS, Grool AM, Mali WP, Graaf YV der, et al. Cerebellar infarct patterns: the SMART-Medea study. *Neuroimage Clin.* (2015) 8:314–21. doi: 10.1016/j.nicl.2015.02.001
- Manto M. Cerebellar motor syndrome from children to the elderly. In: Manto M, Huisman T, editors. *Handbook of Clinical Neurology: The Cerebellum: From Embryology to Diagnostic Investigations*, Vol. 154. 2018. p. 151–66.
- 235. Kelman L. The triggers or precipitants of the acute migraine attack. *Cephalalgia*. (2007) 27:394–402. doi: 10.1111/j.1468-2982.2007.01303.x
- 236. Marmura MJ. Triggers, protectors, and predictors in episodic migraine. *Curr Pain Headache R.* (2018) 22:81. doi: 10.1007/s11916-018-0734-0
- 237. Seo JG, Park SP. Significance of fatigue in patients with migraine. J Clin Neurosci. (2018) 50:69–73. doi: 10.1016/j.jocn.2018.01.032
- 238. Karsan N, Goadsby PJ. Migraine is more than just headache: is the link to chronic fatigue and mood disorders simply due to shared biological systems? *Front Hum Neurosci.* (2021) 15:646692. doi: 10.3389/fnhum.2021.646692
- 239. Peres M, Zukerman E, Young W, Silberstein S. Fatigue in chronic migraine patients. *Cephalalgia*. (2002) 22:720– 4. doi: 10.1046/j.1468-2982.2002.00426.x
- Lucchesi C, Sassi AN, Siciliano G, Gori S. Fatigue is increased in episodic migraine without aura patients. *Headache J Head Face Pain*. (2013) 53:1163– 5. doi: 10.1111/head.12134
- 241. Kuppuswamy A. The fatigue conundrum. Brain. (2017) 140:2240– 5. doi: 10.1093/brain/awx153
- 242. Karsan N, Bose PR, Thompson C, Newman J, Goadsby PJ. Headache and non-headache symptoms provoked by nitroglycerin in migraineurs: a human pharmacological triggering study. *Cephalalgia*. (2020) 24:033310242091011– 4. doi: 10.1177/0333102420910114
- 243. Young WB, Gangal KS, Aponte RJ, Kaiser RS. Migraine with unilateral motor symptoms: a case-control study. *J Neurol Neurosurg Psychiatry*. (2007) 78:600. doi: 10.1136/jnnp.2006.100214
- 244. Blum ASS, Couperus CJ, Rosen BJ, Haskin-Leahy LF, Shapiro RE. Familial "Diplegic" migraine - description of a family with a novel CACNA1A mutation. *Headache J Head Face Pain*. (2020) 60:600– 6. doi: 10.1111/head.13741
- 245. Ambrosini A, Sándor PS, Pasqua V de, Pierelli F, Schoenen J. Performances in cerebellar and neuromuscular transmission tests are correlated in migraine with aura. *J Headache Pain*. (2008) 9:29–32. doi: 10.1007/s10194-008-0004-3
- 246. Florencio LL, Oliveira AS de, Pinheiro CF, Will-Lemos T, Dach F, Fernándezde-las-Peñas C, et al. Comparison of cervical muscle isometric force between migraine subgroups or migraine-associated neck pain: a controlled study. *Sci Rep UK*. (2021) 11:15434. doi: 10.1038/s41598-021-95078-4

- 247. Florencio LL, Oliveira AS de, Will-Lemos T, Pinheiro CF, Marçal JC da S, Dach F, et al. Muscle endurance and cervical electromyographic activity during submaximal efforts in women with and without migraine. *Clin Biomech.* (2021) 82:105276. doi: 10.1016/j.clinbiomech.2021.105276
- Florencio LL, Oliveira AS, Lemos TW, Carvalho GF, Dach F, Bigal ME, et al. Patients with chronic, but not episodic, migraine display altered activity of their neck extensor muscles. *J ElectromyogrKines*. (2016) 30:66– 72. doi: 10.1016/j.jelekin.2016.06.003
- 249. Bologna M, Berardelli A. The cerebellum and dystonia. In: Manto M, Huisman T, editors. *Handbook of Clinical Neurology: The Cerebellum: Disorders and treatment*, Vol. 155. Elsevier (2018). p. 259–72.
- White JJ, Sillitoe RV. Genetic silencing of olivocerebellar synapses causes dystonia-like behaviour in mice. *Nat Commun.* (2019) 8:1– 16. doi: 10.1038/ncomms14912
- 251. Münchau A, Valente EM, Shahidi GA, Eunson LH, Hanna MG, Quinn NP, et al. A new family with paroxysmal exercise induced dystonia and migraine: a clinical and genetic study. *J Neurol Neurosurg Psychiatry.* (2000) 68:609. doi: 10.1136/jnnp.68.5.609
- 252. Bøttger P, Doganli C, Lykke-Hartmann K. Migraine- and dystoniarelated disease-mutations of Na+/K+-ATPases: relevance of behavioral studies in mice to disease symptoms and neurological manifestations in humans. *NeurosciBiobehav Rev.* (2012) 36:855–71. doi: 10.1016/j.neubiorev.2011.10.005
- 253. Stampfl B, Fee D. Novel mutation in CACNA1A associated with activityinduced dystonia, cervical dystonia, and mild ataxia. *Case Rep Neurol Med.* (2021) 2021:7797770. doi: 10.1155/2021/7797770
- 254. Kamm C, Mayer P, Sharma M, Niemann G, Gasser T. New family with paroxysmal exercise-induced dystonia and epilepsy. *Movement Disord.* (2007) 22:873–7. doi: 10.1002/mds.21350
- 255. Magis D, Boon E, Coppola G, Daron A, Schoenen J. A novel CACNA1A mutation results in episodic ataxia with migrainous features without headache. *Cephalalgia*. (2012) 32:1147–9. doi: 10.1177/0333102412459572
- 256. Ferrari MD, Klever RR, Terwindt GM, Ayata C, Maagdenberg AMJM van den. Migraine pathophysiology: lessons from mouse models and human genetics. *Lancet Neurol.* (2015). 14:65-80. doi: 10.1016/S1474-4422(14)70220-0
- 257. Ophoff RA, Terwindt GM, Vergouwe MN, Eijk R van, Oefner PJ, Hoffman SMG, et al. Familial hemiplegic migraine and episodic ataxia type-2 are caused by mutations in the Ca2+ channel gene CACNL1A4. *Cell.* (1996) 87:543–52. doi: 10.1016/S0092-8674(00)81373-2
- 258. Brennan KC, Bates EA, Shapiro RE, Zyuzin J, Hallows WC, Huang Y, et al. Casein kinase i mutations in familial migraine and advanced sleep phase. Sci Transl Med. (2013) 5:183ra56. doi: 10.1126/scitranslmed.3005784
- 259. Eising E, Datson NA, Maagdenberg AM van den, Ferrari MD. Epigenetic mechanisms in migraine: a promising avenue? *BMC Med.* (2013) 11:26. doi: 10.1186/1741-7015-11-26
- 260. Ahuja AS, Rozen TD, Atwal PS. A sleep modulated channelopathy: a novel CACNA1A pathogenic variant identified in episodic ataxia type 2 and a potential link to sleep alleviated migraine. *BMC Neurol.* (2019) 19:246. doi: 10.1186/s12883-019-1491-3
- 261. Hautakangas H, Winsvold BS, Ruotsalainen SE, Bjornsdottir G, Harder AVE, Kogelman LJA, et al. Genome-wide analysis of 102,084 migraine cases identifies 123 risk loci and subtype-specific risk alleles. *Nat Genet.* (2022) 54:152–60. doi: 10.1038/s41588-021-0 0990-0
- 262. Richter F, Ebersberger A, Schaible HG. Blockade of voltage-gated calcium channels in rat inhibits repetitive cortical spreading depression. *Neurosci Lett.* (2002) 334:123–6. doi: 10.1016/S0304-3940(02)01120-5
- 263. Tozzi A, Iure A de, Filippo MD, Costa C, Caproni S, Pisani A, et al. Critical role of calcitonin gene-related peptide receptors in cortical spreading depression. *Proc Natl Acad Sci USA*. (2012) 109:18985– 90. doi: 10.1073/pnas.1215435109
- Moskowitz MA. Genes, proteases, cortical spreading depression and migraine: impact on pathophysiology and treatment. *Funct Neurol.* (2007) 22:133–6.
- 265. Khennouf L, Gesslein B, Lind BL, Maagdenberg AMJM van den, Lauritzen M. Activity-dependent calcium, oxygen, and vascular responses in a mouse

model of familial hemiplegic migraine type 1. Ann Neurol. (2016) 80:219–32. doi: 10.1002/ana.24707

- 266. Maagdenberg AMJM van den, Pietrobon D, Pizzorusso T, Kaja S, Broos LAM, Cesetti T, et al. A Cacnala knockin migraine mouse model with increased susceptibility to cortical spreading depression. *Neuron.* (2004) 41:701–10. doi: 10.1016/S0896-6273(04)00085-6
- 267. Goadsby PJ. Migraine aura: a knockin mouse with a knockout message. *Neuron*. (2004) 41:679–80. doi: 10.1016/S0896-6273(04)00112-6
- 268. Parker PD, Suryavanshi P, Melone M, Sawant-Pokam PA, Reinhart KM, Kaufmann D, et al. Non-canonical glutamate signaling in a genetic model of migraine with aura. *Neuron.* (2021) 109:611–28.e8. doi: 10.1016/j.neuron.2020.11.018
- 269. Capuani C, Melone M, Tottene A, Bragina L, Crivellaro G, Santello M, et al. Defective glutamate and K +clearance by cortical astrocytes in familial hemiplegic migraine type 2. *EMBO Mol Med.* (2016) 8:967–86. doi: 10.15252/emmm.201505944
- 270. Leo L, Gherardini L, Barone V, Fusco MD, Pietrobon D, Pizzorusso T, et al. Increased susceptibility to cortical spreading depression in the mouse model of familial hemiplegic migraine type 2. *PLoS Genet.* (2011) 7:e1002129– 12. doi: 10.1371/journal.pgen.1002129
- 271. Pietrobon D. Insights into migraine mechanisms and Ca V2.1 calcium channel function from mouse models of familial hemiplegic migraine. J Physiol. (2010) 588:1871–8. doi: 10.1113/jphysiol.2010.188003
- 272. Dichgans M, Freilinger T, Eckstein G, Babini E, Lorenz-Depiereux B, Biskup S, et al. Mutation in the neuronal voltage-gated sodium channel SCN1A in familial hemiplegic migraine. *Lancet.* (2005) 366:371– 7. doi: 10.1016/S0140-6736(05)66786-4
- 273. Hu Y, Tang W, Liu R, Dong Z, Chen X, Pan M, et al. Higher prevalence of migraine in essential tremor: a case-control study. *Cephalalgia*. (2014) 34:1142–9. doi: 10.1177/0333102414531153
- 274. Biary N, Koller W, Langenberg P. Correlation between essential tremor and migraine headache. J Neurol Neurosurg Psychiatry. (1990) 53:1060. doi: 10.1136/jnnp.53.12.1060
- Lau CI, Lin CC, Chen HJ, Wang HC, Chen WH, Liang JA. Increased risk of essential tremor in migraine: a population-based retrospective cohort study. *PLoS ONE*. (2017) 12:e0173586. doi: 10.1371/journal.pone.0173586
- 276. Louis ED. Essential tremor and the cerebellum. In: *The Cerebellum: Disorders and Treatment*, Vol. 155 (2018). p. 245–58.
- 277. Lorenz D, Deuschl G. Update on pathogenesis and treatment of essential tremor. *CurrOpin Neurol.* (2007) 20:447– 52. doi: 10.1097/WCO.0b013e3281e66942
- 278. Urban PP, Rolke R, Wicht S, Keilmann A, Stoeter P, Hopf HC, et al. Left-hemispheric dominance for articulation: a prospective study on acute ischaemic dysarthria at different localizations. *Brain.* (2006) 129:767– 77. doi: 10.1093/brain/awh708
- 279. Solomon NP, Makashay MJ, Helou LB, Clark HM. Neurogenic orofacial weakness and speech in adults with dysarthria. Am J Speech Lang Pat. (2017) 26:951–60. doi: 10.1044/2017_AJSLP-16-0144
- Thomsen LL, Eriksen MK, Roemer SF, Andersen I, Olesen J, Russell MB. A population-based study of familial hemiplegic migraine suggests revised diagnostic criteria. *Brain.* (2002) 125(Pt 6):1379–91. doi: 10.1093/brain/awf132
- Kirchmann M. Migraine with aura: new understanding from clinical epidemiologic studies. *Curr Opin Neurol.* (2006) 19:286–93. doi: 10.1097/01.wco.0000227040.16071.a9
- 282. Schwedt TJ, Peplinski J, Garcia-Filion P, Berisha V. Altered speech with migraine attacks: a prospective, longitudinal study of episodic migraine without aura. *Cephalalgia*. (2018) 39:722–31. doi: 10.1177/0333102418815505
- 283. Pollack IF, Polinko P, Albright LA, Towbin R, Fitz C. Mutism and pseudobulbar symptoms after resection of posterior fossa tumors in childrenincidence and pathophysiology. *Neurosurgery*. (1995) 37:885– 92. doi: 10.1227/00006123-199511000-00006
- Erşahin Y, Mutluer S, Çagli S, Duman Y. Cerebellar mutism: report of seven cases and review of the literature. *Neurosurgery.* (1996) 38:60–6. doi: 10.1097/00006123-199601000-00015

- 285. Catsman-Berrevoets C, Patay Z. Cerebellar mutism syndrome. In: Manto M, Huisman T, editors. *Handbook of Clinical Neurology: The Cerebellum: Disorders and treatment*, Vol. 155. Elsevier (2018). p. 273–88.
- Mariën P, Borgatti R. Chapter 11 language and the cerebellum. *Handb Clin* Neurol. (2018) 154:181–202. doi: 10.1016/B978-0-444-63956-1.00011-4
- 287. Sugihara I, Fujita H, Na J, Quy PN, Li B, Ikeda D. Projection of reconstructed single purkinje cell axons in relation to the cortical and nuclear aldolase C compartments of the rat cerebellum. J Comp Neurol. (2009) 512:282– 304. doi: 10.1002/cne.21889
- Edlow JA, Newman-Toker DE, Savitz SI. Diagnosis and initial management of cerebellar infarction. *Lancet Neurol.* (2008) 7:951–64. doi: 10.1016/S1474-4422(08)70216-3
- Savitz SI, Caplan LR, Edlow JA. Pitfalls in the diagnosis of cerebellar infarction. AcadEmerg Med. (2007) 14:63–8. doi: 10.1197/j.aem.2006.06.060
- Vukovic; V, Demarin V. Prevalence of vertigo dizziness and migrainous vertigo in patients with migraine. *Headache.* (2007) 47:1427–35. doi: 10.1111/j.1526-4610.2007.00939.x
- 291. Iljazi A, Ashina H, Lipton RB, Chaudhry B, Al-Khazali HM, Naples JG, et al. Dizziness and vertigo during the prodromal phase and headache phase of migraine: a systematic review and meta-analysis. *Cephalalgia.* (2020) 40:1095–103. doi: 10.1177/0333102420921855
- Dieterich M, Brandt T. Episodic vertigo related to migraine (90 cases): vestibular migraine? J Neurol. (1999) 246:883– 92. doi: 10.1007/s004150050478
- 293. Meng H, May PJ, Dickman JD, Angelaki DE. Vestibular signals in primate thalamus: properties and origins. J Neurosci. (2007) 27:13590– 602. doi: 10.1523/JNEUROSCI.3931-07.2007
- Russo A, Marcelli V, Esposito F, Corvino V, Marcuccio L, Giannone A, et al. Abnormal thalamic function in patients with vestibular migraine. *Neurology*. (2014) 82:2120–6. doi: 10.1212/WNL.00000000000496
- 295. Messina R, Rocca MA, Colombo B, Teggi R, Falini A, Comi G, et al. Structural brain abnormalities in patients with vestibular migraine. *J Neurol.* (2017) 264:295–303. doi: 10.1007/s00415-016-8349-z
- 296. Wang S, Wang H, Liu X, Yan W, Wang M, Zhao R. A resting-state functional MRI study in patients with vestibular migraine during interictal period. *Acta Neurol Belg.* (2021) 1–7. doi: 10.1007/s13760-021-01639-9
- 297. Kotchabhakdi N, Walberg F. Cerebellar afferent projections from the vestibular nuclei in the cat: an experimental study with the method of retrograde axonal transport of horseradish peroxidase. *Exp Brain Res.* (1978) 31:591–604. doi: 10.1007/BF00239814
- 298. Balaban CD, Beryozkin G. Vestibular nucleus projections to nucleus tractus solitarius and the dorsal motor nucleus of the vagus nerve: potential substrates for vestibulo-autonomic interactions. *Exp Brain Res.* (1994) 98:200–12. doi: 10.1007/BF00228409
- Gagliuso AH, Chapman EK, Martinelli GP, Holstein GR. Vestibular neurons with direct projections to the solitary nucleus in the rat. J Neurophysiol. (2019) 122:512–24. doi: 10.1152/jn.00082.2019
- 300. Maniyar FH, Sprenger T, Schankin C, Goadsby PJ. The origin of nausea in migraine-a PET study. J Headache Pain. (2014) 15:84. doi: 10.1186/1129-2377-15-84
- 301. Bower JM. Control of sensory data acquisition. Int Rev Neurobiol. (1997) 41:489–513. doi: 10.1016/S0074-7742(08)60367-0
- 302. Puledda F, Villar-Martínez MD, Goadsby PJ. Case report: transformation of visual snow syndrome from episodic to chronic associated with acute cerebellar infarct. *Front Neurol.* (2022) 13:811490. doi: 10.3389/fneur.2022.811490
- 303. Puledda F, Bruchhage M, O'Daly O, Ffytche D, Williams SCR, Goadsby PJ. Occipital cortex and cerebellum gray matter changes in visual snow syndrome. *Neurology.* (2020) 95:e1792– 9. doi: 10.1212/WNL.000000000010530
- 304. Denuelle M, Boulloche N, Payoux P, Fabre N, Trotter Y, Geraud G. A PET study of photophobia during spontaneous migraine attacks. *Neurology*. (2011) 76:213–8. doi: 10.1212/WNL.0b013e3182074a57
- 305. Boulloche N, Denuelle M, Payoux P, Fabre N, Trotter Y, Geraud G. Photophobia in migraine: an interictal PET study of cortical hyperexcitability and its modulation by pain. J Neurol Neurosurg Psychiatry. (2010) 81:978– 84. doi: 10.1136/jnnp.2009.190223

- 306. Eren OE, Ruscheweyh R, Straube A, Schankin CJ. Quantification of photophobia in visual snow syndrome: a case-control study. *Cephalalgia*. (2019) 40:393–8. doi: 10.1177/03331024198 96780
- 307. Schankin CJ, Maniyar FH, Chou DE, Eller M, Sprenger T, Goadsby PJ. Structural and functional footprint of visual snow syndrome. *Brain.* (2020) 47:996–8. doi: 10.1093/brain/awaa053
- Puledda F, Vandenbussche N, Moreno-Ajona D, Eren O, Schankin C, Goadsby PJ. Evaluation of treatment response and symptom progression in 400 patients with visual snow syndrome. *Brit J Ophthalmol.* (2021) 1–7. doi: 10.1136/bjophthalmol-2020-318653
- Noseda R, Copenhagen D, Burstein R. Current understanding of photophobia, visual networks and headaches. *Cephalalgia*. (2018) 39:1623– 34. doi: 10.1177/0333102418784750
- Palmer JE, Chronicle EP. Cognitive processing in migraine: a failure to find facilitation in patients with aura. *Cephalalgia*. (1998) 18:125– 32. doi: 10.1046/j.1468-2982.1998.1803125.x
- Gil-Gouveia R, Martins IP. Cognition and cognitive impairment in migraine. Curr Pain Headache R. (2019) 23:84. doi: 10.1007/s11916-019-0824-7
- 312. Morena J, Kamdar HA, Adeli A. Cerebellar ischemia presenting as transient global amnesia. *CognBehav Neurol.* (2021) 34:319– 22. doi: 10.1097/WNN.0000000000287
- Schmahmann JD, Sherman JC. Cerebellar cognitive affective syndrome. Int Rev Neurobiol. (1997) 41:433–40. doi: 10.1016/S0074-7742(08)60363-3
- 314. Demirtas-Tatlidede A, Freitas C, Cromer JR, Safar L, Ongur D, Stone WS, et al. Safety and proof of principle study of cerebellar vermal theta burst stimulation in refractory schizophrenia. *Schizophr Res.* (2010) 124:91–100. doi: 10.1016/j.schres.2010.08.015
- 315. Brady RO, Gonsalvez I, Lee I, Öngür D, Seidman LJ, Schmahmann JD, et al. Cerebellar-prefrontal network connectivity and negative symptoms in schizophrenia. Am J Psychiat. (2019) 176:512–20. doi: 10.1176/appi.ajp.2018.18040429
- Parker KL, Narayanan NS, Andreasen NC. The therapeutic potential of the cerebellum in schizophrenia. *Front Syst Neurosci.* (2014) 8:163. doi: 10.3389/fnsys.2014.00163
- Giffin NJ, Ruggiero L, Lipton RB, Silberstein SD, Tvedskov JF, Olesen J, et al. Premonitory symptoms in migraine: an electronic diary study. *Neurology*. (2003) 60:935–40. doi: 10.1212/01.WNL.0000052998.58526.A9
- Maniyar FH, Sprenger T, Monteith T, Schankin C, Goadsby PJ. Brain activations in the premonitory phase of nitroglycerin-triggered migraine attacks. *Brain*. (2013) 137:232–41. doi: 10.1093/brain/awt320
- Weiller C, May A, Limmroth V, Jüptner M, Kaube H, Schayck RV, et al. Brain stem activation in spontaneous human migraine attacks. *Nat Med.* (1995) 1:658–60. doi: 10.1038/nm0795-658
- Afridi SK, Giffin NJ, Kaube H, Friston KJ, Ward NS, Frackowiak RSJ, et al. A positron emission tomographic study in spontaneous migraine. *Arch Neurol.* (2005) 62:1270–5. doi: 10.1001/archneur.62.8.1270
- 321. Bahra A, Matharu MS, Büchel C, Frackowiak RSJ, Goadsby PJ. Brainstem activation specific to migraine headache. *Lancet.* (2001) 357:1016– 7. doi: 10.1016/S0140-6736(00)04250-1
- 322. Cao Y, Aurora SK, Nagesh V, Patel SC, Welch KMA. Functional MRI-BOLD of brainstem structures during visually triggered migraine. *Neurology*. (2002) 59:72–8. doi: 10.1212/WNL.59.1.72
- 323. Granziera C, Romascano D, Daducci A, Roche A, Vincent M, Krueger G, et al. Migraineurs without aura show microstructural abnormalities in the cerebellum and frontal lobe. *Cerebellum*. (2013) 12:812–8. doi: 10.1007/s12311-013-0491-x
- 324. Ke J, Yu Y, Zhang X, Su Y, Wang X, Hu S, et al. Functional alterations in the posterior insula and cerebellum in migraine without aura: a resting-state MRI study. *Front Behav Neurosci.* (2020) 14:567588. doi: 10.3389/fnbeh.2020.567588
- 325. Qin Z, He XW, Zhang J, Xu S, Li GF, Su J, et al. Structural changes of cerebellum and brainstem in migraine without aura. *J Headache Pain*. (2019) 20:93. doi: 10.1186/s10194-019-1045-5
- 326. Amin FM, Hougaard A, Magon S, Sprenger T, Wolfram F, Rostrup E, et al. Altered thalamic connectivity during spontaneous attacks of migraine without aura: a resting-state fMRI study. *Cephalalgia*. (2017) 38:1237– 44. doi: 10.1177/0333102417729113

- 327. Hougaard A, Amin FM, Larsson HBW, Rostrup E, Ashina M. Increased intrinsic brain connectivity between pons and somatosensory cortex during attacks of migraine with aura. *Hum Brain Mapp.* (2017) 38:2635– 42. doi: 10.1002/hbm.23548
- 328. Hougaard A, Amin FM, Christensen CE, Younis S, Wolfram F, Cramer SP, et al. Increased brainstem perfusion, but no blood-brain barrier disruption, during attacks of migraine with aura. *Brain*. (2017) 140:1633–42. doi: 10.1093/brain/awx089
- 329. Kruit MC, Buchem MA van, Hofman PAM, Bakkers JTN, Terwindt GM, Ferrari MD, et al. Migraine as a risk factor for subclinical brain lesions. JAMA . (2004) 291:427–34. doi: 10.1001/jama.291.4.427
- 330. Kruit MC. Infarcts in the posterior circulation territory in migraine. The population-based MRI CAMERA study. *Brain.* (2005) 128:2068– 77. doi: 10.1093/brain/awh542
- 331. Kruit MC, Buchem MA van, Launer LJ, Terwindt GM, Ferrari MD. Migraine is associated with an increased risk of deep white matter lesions, subclinical posterior circulation infarcts and brain iron accumulation: the population-based MRI CAMERA study. *Cephalalgia.* (2009) 291:427– 34. doi: 10.1111/j.1468-2982.2009.01904.x
- 332. Reinhard M, Schork J, Allignol A, Weiller C, Kaube H. Cerebellar and cerebral autoregulation in migraine. *Stroke.* (2012) 43:987–93. doi: 10.1161/STROKEAHA.111.644674
- Gross EC, Lisicki M, Fischer D, Sándor PS, Schoenen J. The metabolic face of migraine - from pathophysiology to treatment. *Nat Rev Neurol.* (2019) 15:1–17. doi: 10.1038/s41582-019-0255-4
- 334. Grech O, Mollan SP, Wakerley BR, Fulton D, Lavery GG, Sinclair AJ. The role of metabolism in migraine pathophysiology and susceptibility. *Life.* (2021) 11:415. doi: 10.3390/life11050415
- Vollono C, Primiano G, Marca GD, Losurdo A, Servidei S. Migraine in mitochondrial disorders: prevalence and characteristics. *Cephalalgia*. (2017) 38:1093–106. doi: 10.1177/0333102417723568
- Markley HG. CoEnzyme Q10 and riboflavin: the mitochondrial connection. *Headache J Head Face Pain.* (2012) 52:81– 7. doi: 10.1111/j.1526-4610.2012.02233.x
- 337. Younis S, Hougaard A, Christensen CE, Vestergaard MB, Paulson OB, Larsson HBW, et al. Interictal pontine metabolism in migraine without aura patients: a 3 Tesla proton magnetic resonance spectroscopy study. *Neuroimage Clin.* (2021) 32:102824. doi: 10.1016/j.nicl.2021.102824
- Proia P, Amato A, Contrò V, Monaco AL, Brusa J, Brighina F, et al. Relevance of lactate level detection in migrane and fibromyalgia. *Europ J Transl Myol.* (2019) 29:8202. doi: 10.4081/ejtm.2019.8202
- Dietrich MO, Horvath TL. Hypothalamic control of energy balance: insights into the role of synaptic plasticity. *Trends Neurosci.* (2013) 36:65– 73. doi: 10.1016/j.tins.2012.12.005
- 340. Nakamura K, Nakamura Y, Kataoka N. A hypothalamomedullary network for physiological responses to environmental stresses. *Nat Rev Neurosci.* (2022) 23:35–52. doi: 10.1038/s41583-021-00532-x
- 341. Herculano-Houzel S. The human brain in numbers: a linearly scaled-up primate brain. Front Hum Neurosci. (2009) 3:31. doi: 10.3389/neuro.09.031.2009
- Pietrobon D, Moskowitz MA. Chaos and commotion in the wake of cortical spreading depression and spreading depolarizations. *Nat Rev Neurosci.* (2014) 15:379–93. doi: 10.1038/nrn3770
- 343. Schain AJ, Melo-Carrillo A, Strassman AM, Burstein R. Cortical spreading depression closes paravascular space and impairs glymphatic flow: implications for migraine headache. *The J Neurosci.* (2017) 37:2904– 15. doi: 10.1523/JNEUROSCI.3390-16.2017
- 344. Zhang X, Levy D, Noseda R, Kainz V, Jakubowski M, Burstein R. Activation of meningeal nociceptors by cortical spreading depression: implications for migraine with aura. J Neurosci. (2010) 30:8807–14. doi: 10.1523/JNEUROSCI.0511-10.2010
- 345. Schain AJ, Melo-Carrillo A, Ashina S, Strassman AM, Burstein R. Celecoxib reduces cortical spreading depression-induced macrophage activation and dilatation of dural but not pial arteries in rodents. *Pain.* (2020) 161:1019–26. doi: 10.1097/j.pain.0000000000 01789
- 346. Hadjikhani N, Rio MS del, Wu O, Schwartz D, Bakker D, Fischl B, et al. Mechanisms of migraine aura revealed by functional MRI

in human visual cortex. *Proc Natl Acad Sci USA*. (2001) 98:4687–92. doi: 10.1073/pnas.071582498

- 347. Hadjikhani N, Albrecht D, Mainero C, Moskowitz MA. Extra-axial inflammatory signal in parameninges in migraine with visual aura. Ann Neurol. (2020) 87:1–27. doi: 10.1002/ana.25731
- 348. Chen G, Hanson CL, Dunbar RL, Ebner TJ. Novel form of spreading acidification and depression in the cerebellar cortex demonstrated by neutral red optical imaging. J Neurophysiol. (1999) 81:1992–8. doi: 10.1152/jn.1999.81.4.1992
- 349. Ebner TJ, Chen G, Gao W, Reinert K. Optical imaging of cerebellar functional architectures: parallel fiber beams, parasagittal bands and spreading acidification. *Prog Brain Res.* (2005) 148:125–38. doi: 10.1016/S0079-6123(04)48011-X
- 350. Keller JT, Saunders MC, Beduk A, Jollis JG. Innervation of the posterior fossa dura of the cat. *Brain Res Bull.* (1985) 14:97–102. doi: 10.1016/0361-9230(85)90181-9
- 351. Kimmel DL. Innervation of spinal dura mater and dura mater of the posterior cranial fossa. *Neurology*. (1961) 11:800– 9. doi: 10.1212/WNL.11.9.800
- 352. Wattiez A, O'Shea SA, Eyck PT, Sowers LP, Recober A, Russo AF, et al. Patients with vestibular migraine are more likely to have occipital headaches than those with migraine without vestibular symptoms. *Headache J Head Face Pain.* (2020) 60:1581–91. doi: 10.1111/head.13898

- 353. May A. Understanding migraine as a cycling brain syndrome: reviewing the evidence from functional imaging. *Neurol Sci.* (2017) 38(Suppl 1):125–30. doi: 10.1007/s10072-017-2 866-0
- 354. May A, Burstein R. Hypothalamic regulation of headache and migraine. *Cephalalgia*. (2019) 53:1710–9.

Conflict of Interest: The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Noseda. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.