Effect of melanoma stem cells on melanoma metastasis (Review)

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Abstract. Cancer stem cells (CSCs) are involved in the metastatic process, the resistance of many types of cancer to therapeutic treatments and consequently the onset of recurrences. The CSC concept therefore significantly extends our understanding of melanoma biology. More recently, melanoma stem cells (MSCs) have been described in melanoma as expressing specific biomarkers. These primitive melanoma cells are not only capable of self-renewal and differentiation plasticity, but may also confer virulence via immune evasion and multidrug resistance, and potentially, via vasculogenic mimicry and transition to migratory and metastasizing derivatives. This review will present the specific biomarkers of MSCs, including CD133, ATP binding cassette subfamily B member 5, CD271, CD20 and aldehyde dehydrogenase, which can regulate the transduction of tumor-related signals. These signal molecules can reversely act on tumor cells and regulate tumor angiogenesis, leading to the occurrence of melanoma metastasis. Targeting these specific biomarkers could inhibit the progression of melanoma and may help the development of novel therapeutic strategies for melanoma.

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1. Introduction

Melanoma is a malignant tumor of melanocytes that typically arises in the skin. It is highly malignant, prone to metastasis and recurrence, accounting for ~75% of skin cancer-associated mortality (1). The treatment of advanced or metastatic melanoma is particularly challenging, and there is a high tendency for patients to relapse and become resistant to current therapeutic agents (2). Although molecular targeted therapy and immunotherapy have been reported to prolong the survival time of patients, most patients will develop drug resistance within one year (3-5), resulting in melanoma metastasis. The existence of melanoma stem cells (MSCs) is one of the potential causes of melanoma invasion and metastasis.

Cancer stem cells (CSCs) have been shown to be an integral part of solid tumors (6). Furthermore, CSCs exhibit distinctive and remarkable capacities of self-renewal, differentiation and proliferation, which are believed to have a key role in all aspects of carcinogenesis, including tumor recurrence and metastasis (7,8). Previous studies have demonstrated that the essence of tumor metastasis is the transfer and homing of CSCs (6,9). In the last decade, with the rise of CSCs, several lines of evidence suggested that CSCs may be at the origin of tumor metastasis (10-12). Interestingly, the CSC subpopulation is responsible for many aspects of tumorigenesis and has been reported to serve a crucial role in melanoma development, progression, drug resistance and metastasis (13,14). The fact that the CSCs are resistant to chemotherapy also explains that traditional anticancer drugs can only inhibit or narrow the tumor, but not completely eradicate it, leading to tumor metastasis and recurrence (15,16). In addition, CSCs have been reported to express a variety of biomarkers, such as CD34, aldehyde dehydrogenase 1 (ALDH1), CD271, CD44 and lysine demethylase 5B (JARID1B); however, none of these markers have been shown to be CSCs-specific (17-19). Several potential biomarkers of CSCs have been demonstrated to be expressed by certain human solid tumors such as melanoma (16), including CD133, ATP binding cassette subfamily B member 5 (ABCB5), CD271, CD20 and ALDH (Table I). Although the mechanism of MSCs promoting tumor metastasis and recurrence has not been fully elucidated (13,20,21), the activation of the signaling pathways, including Notch, Hedgehog and Wnt, is modulated by

these biomarkers to maintain the characteristic of MSCs, thus promoting angiogenesis and epithelial-mesenchymal transition (EMT) of melanoma and accelerating tumor metastasis (7,20,22) (Fig. 1). It is therefore important to determine the role of MSCs in the invasion and metastasis of melanoma.

2. Melanoma metastasis is modulated by molecular markers of MSCs

CD133. As a surface protein with unknown function, CD133 (prominin-1) can be expressed on human melanoma, but hardly be detected in normal skin (23). It has been reported that CD133, a stem cell-related surface antigen, is closely related to tumor proliferation and progression in various types of tumor, including melanoma (15,24,25). Furthermore, CD133⁺ melanoma cells exhibit 411 upregulated genes, which are associated with angiogenesis, adhesion and migration (26). Melanoma CD133+ CSCs have the potential to initiate tumor progression. In addition, CD133 can activate MAPK signal pathway through notch receptor 1 (Notch1), regulate the expression of vascular endothelial growth factor (VEGF) and matrix metalloproteinases and promote the interaction of tumor endothelial cells, leading to the increase of tumor angiogenesis and lung metastasis (26). Furthermore, the expression of CD133, p-p38 and p-MEK3/6 in metastatic melanoma is significantly higher than that in paracancerous tissues (26,27), suggesting that Notch1 and its MAPK signaling pathway network might be considered as potential targets for MSCs-mediated melanoma targeted therapy (28,29). Furthermore, it is generally known that tumor angiogenesis is an essential factor for tumor growth and metastasis. In particular, CD133⁺ and ABCB5⁺ MSCs are involved in the formation of perivascular niches (30). When RNAi is used to block the expression of CD133, the tumorigenicity of stem cells in vivo is significantly decreased and the expression of CD144 and ABCB5 that are closely related to the vascular microenvironment is downregulated (30). However, ABCB5⁻ cells lose the ability to form CD144⁺ angiogenic mimicry. It has been reported that CD133+/ABCB5+ MSCs exist in CD144+ angiogenic mimicry, suggesting that CD133⁺ MSCs could promote tumor metastasis by increasing the formation of angiogenic mimicry and specific vascular microenvironment (30). Zimmerer et al (31) demonstrated by fluorescence microscopy that CD133+ melanoma D10 cells xenograft into nude mice can trigger an important angiogenesis process. A clonal dominance of a CD133⁺ population exists within the hierarchy of cells in cutaneous tissues from patients that have undergone successive progressive stages of melanoma, from primary to metastatic lesions (32). In addition, in the CD133⁻ melanoma cells subpopulation, exposure to taxol induces the activation of apoptosis signal-regulating kinasel/c-jun-N-terminal kinase, p38 and ERK pathways and Bax expression; however, in CD133⁺ cells, taxol only enhances the activity of the ERK pathway (33). Furthermore, it was demonstrated that the expression of CD133+ in patients with recurrent and metastatic melanoma is twice higher than in patients with primary melanoma (32). Mechanistically, CD133 downregulation in human metastatic melanoma cells can decrease the capacity of sphere-forming and the metastasis potential of melanocytes (28). In addition, CD133, is closely related to the expression of certain tumor associated antigens and could thus serve as a potential

target for immunotherapy (34,35). CD133 may therefore be considered as a predictive marker of melanoma and as a potential therapeutic target of high-risk melanoma.

ABCB5. ABCB5 is a member of the ATP binding cassettes (ABC) transporter family and a regulator of cell membrane potential. ABCB5 can regulate the fusion of normal skin progenitor cells and is considered as one marker of MSCs (36,37). It is now well accepted that ABC transporters mediate multidrug resistance through drug efflux in cancer cells, which is usually related to cancer stem cells (38-40). In vivo genetic lineage tracking demonstrated a specific capacity of ABCB5+ sub-populations for self-renewal and differentiation, as ABCB5+ cancer cells generate both ABCB5⁺ and ABCB5⁻ progeny whereas ABCB5⁻ tumor populations give rise, at lower rates, exclusively to ABCB5⁻ cells (37). Subsequently, ABCB5, a marker of MSCs, has high tumorigenicity potential and is co-expressed with other MSCs markers, such as CD133 (39,41,42). ABCB5+ tumor cells detected in human melanoma patients show a primitive molecular phenotype and correlate with clinical melanoma progression (43). Analysis of some clinical data demonstrated that overexpression of ABCB5 can promote tumor progression, and that ABCB5 expression is usually low in pigmented nevus subpopulation but high in primary and metastatic melanoma cell subpopulation (37,44). In serial human-to-mouse xenotransplantation experiments, ABCB5⁺ melanoma cells possess greater tumorigenic capacity than ABCB5⁻ bulk populations (44,45). Ma and Frank (45) reported that ABCB5⁺ melanoma cells exist in the peripheral blood of patients with melanoma. Subsequently, transplanting these cells to Nod/SCID/IL2 mice can induce distant metastasis, and the degree of metastasis is directly proportional to the number of ABCB5⁺ melanoma cells transplanted.

Frank et al (46) demonstrated that ABCB5+ human melanoma cells promote the formation of angiogenic mimicry by expressing endothelium-specific markers and other angiogenic proteins. Because angiogenic mimicry plays an essential role in angiogenesis of melanoma (21), the ABCB5⁺ subpopulation of human melanoma expresses preferentially the proteins tyrosine kinase with immunoglobulin like and EGF like domains 1 and CD144 (VE-cadherin) and other angiogenic differentiation markers. Furthermore, ABCB5+ and CD133+ MSCs express preferentially VEGFR1 and VEGF, which are important for the angiogenic mimicry of human melanoma cells and further promote melanoma metastasis (26,46). It was reported that in vivo targeting of VEGFR-1 blocks the development of ABCB5+ vascular mimicry and inhibits tumor growth in melanoma (46). Furthermore, one potential mechanism of metastasis promotion and melanoma recurrence by ABCB5 is that ABCB5 might inhibit the activation of T cells and contribute to immune escape of tumor cells (47). Furthermore, the levels of MHC class I receptor, melanoma associated antigen, costimulatory molecule B7.2 and PD-1 are lower in ABCB5⁺ cells than in ABCB5⁻ cells (48). These findings indicate that targeting ABCB5 MSCs markers in the treatment of melanoma might be beneficial, ongoing clinical trials have proved this view (48).

CD271. CD271, also known as low affinity nerve growth factor receptor or p75NTR, is a characteristic marker of MSCs (36,49). High Expression of CD271 has been reported

Table I. The role of specific biomarkers in melanoma.

CD133	CD133 ⁺ melanoma stem-like cells confer resistance to taxol (33).
	CD133 and ABCG2 positive melanoma cells have the potential ability to promote tumorigenesis (27)
	CD133 ⁺ population within cutaneous tissues promotes the continuous progress of melanoma (32)
	CD133 is a melanoma immunogenic target of which expression is often associated with expression of
	cancer/testis antigens (34,35)
ABCB5	Expression of ABCB5 in melanoma cells promotes resistance to chemical agent (40)
	ABCB5+ tumor cells detected in human melanoma patients show a primitive molecular phenotype and
	correlate with clinical melanoma progression (37)
	ABCB5 interacts with CD166, which promotes clinical malignant melanoma progression (39)
CD271	CD271 is associated with neurotrophins and their receptors, which plays an important role in promoting
	melanoma cell invasion <i>in vitro</i> and migration (52,55)
	A relatively high frequency of CD271/Sox10-positive cells correlates with higher metastatic potential and
	worse prognosis in human melanoma (65)
	CD271 ⁺ cells show higher tumorigenicity and metastatic ability in melanoma (49)
	The low affinity neurotrophin receptor CD271 plays a dual role as a mediator of phenotype switching,
	suppressing melanoma cell proliferation while concomitantly promoting metastasis formation in vivo (58)
CD20	Melanomas contain distinct cell subpopulations including one expressing CD20 with stem cell-like and
	tumor-initiating characteristics (36,72).
	The CD20-expressing melanoma subpopulation is characterized by self-renewal, differentiation into severa
	cell lineages and high tumorigenicity (73).
ALDH	ALDH1 is a promising new marker for cancer stem cells and catalyzes the oxidation of intracellular
	aldehydes, thus conferring multidrug resistance (83).
	ALDH1 is important for cell proliferation, survival and resistance to chemotherapeutic agents (85,86).
	ALDH ⁺ melanoma cells are more tumorigenic than ALDH- cells in both NOD/SCID mice and NOD/
	SCID/IL2ry ^{null} mice (88).
Sox10	Loss of Sox10 impairs neural crest stem cell maintenance and reduces the number of CD271-positive cell
	and counteracts tumorigenesis in melanoma (92,93).
	Following Sox10 knockdown, human melanoma cells are no longer able to initiate tumors in a
	xenotransplantation model (67).

ABCB5, ATP binding cassette subfamily B member 5; ALDH, aldehyde dehydrogenase; Sox10, SRY-Box transcription factor 10.

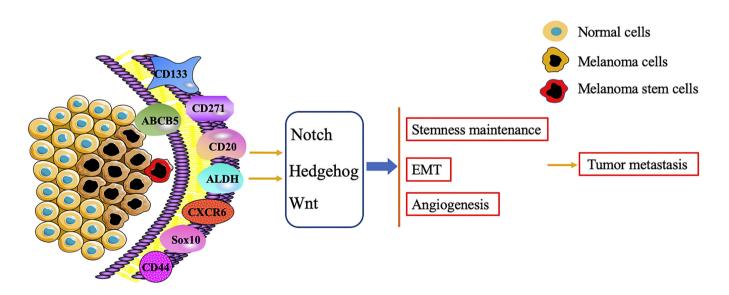


Figure 1. Melanoma stem cells account for only a small part of melanoma cells, which express a variety of cell markers, such as CD133, ABCB5, CD271, CD20, CXCR6, Sox10, CD44 and ALDH. Activation of the signaling pathways (including Notch, Hedgehog and Wnt) is modulated by these markers, in order to maintain the characteristic of melanoma stem cells, thus promoting EMT and angiogenesis of melanoma and accelerating tumor metastasis. ABCB5, ATP binding cassette subfamily B member 5; ALDH, aldehyde dehydrogenase; CXCR6, C-X-C motif chemokine receptor 6; EMT, epithelial-mesenchymal transition; Sox10, SRY-Box transcription factor 10.

in numerous human neural-crest-derived tissues and in some human cancers, including melanomas (50-52). Previous studies demonstrated that CD271⁺ melanoma cells have a higher tumorigenicity potential than CD271⁻ cells and are involved in the metastasis of melanoma in vivo, especially in peripheral nerves (49,53-55). CD271 is the most reliable cell surface marker for the identification of melanoma heterogeneous subsets (49). In addition, not only CD271 marks dedifferentiated melanoma cells emerging, for instance, through TGF\beta-mediated EMT, BRAF inhibitor-induced reprogramming (56) or in response to immunotherapies (57), but it is also functionally involved in promoting low rates of proliferation and high metastatic capacity (58). In fully immunocompromised mouse models, including NOD/SCID/IL2rynull mice, melanoma cells expressing the neurotrophin receptor CD271 have a higher tumor-initiation capacity than CD271⁻ cells, although the negative fraction is also able to generate tumors in this mouse model (49). Furthermore, CD271⁺ melanoma cells are prone to form liver and lung metastasis in mice, whereas CD271⁻ melanoma cells rarely form metastasis (59). In human samples, the proportion of CD271/SRY-Box transcription factor 10 (Sox10) positive cells is significantly increased in metastatic melanoma compared with primary melanoma cells (49,60). Interestingly, CD271 inactivation not only results in decreased melanoma cell survival, but also in increased sensitivity to BRAF inhibitor treatment, suggesting that CD271 might confer therapy resistance (61). Schnegg et al (62) demonstrated that CD133+ and CD271⁺ MSCs accumulate in the perivascular niche that melanoma cells with formation of angiogenic mimicry positively express CD271. Similarly, in the human uveal melanoma cell line c918 cultured in 2D and 3D cultures, evaluation of CD271 expression through immunofluorescence showed that CD271 is expressed on the tumor cells that form the vasculogenic mimicry (63). It was confirmed that melanoma cells that form the vasculogenic mimicry acquire the tumor stem cell-like phenotype and participate in angiogenesis (63).

A previous study reported that the number of MSCs positive for ABCB5, CD271 and receptor activator of nuclear factor κ B in circulating tumor cells (CTCs) of patients with advanced melanoma is significantly increased (64). In addition, CTCs are highly enriched in MSCs, which is a vital parameter inducing the formation of distant secondary tumors (64). Furthermore, melanoma-associated antigens, such as melanoma antigen recognized by T-cells 1, are less exposed to CD271 and ABCB5 positive melanoma cells, which supports the hypothesis that melanoma cells expressing the MSCs makers can escape the attack of host immune system (65). A previous study demonstrated that CD271 overexpression in melanoma cells inhibits the production of melanoma-specific cytotoxic T lymphocytes (CTLs), and that interferon- γ from CTLs subsequently triggers the expression of CD271 in melanoma cells, downregulating therefore the production of melanoma antigens (66). Similarly, it was reported that melanoma cells highly expressing CD271 are associated with high tumor metastasis potential and poor prognosis of patients (67).

CD20. In melanoma, numerous subpopulations with the capacity of self-renewal, differentiation, tumorigenicity and/or drug resistance have been described (49,68-70), including one subpopulation expressing the B cell marker CD20 (36,71-73).

Further characterization with respect to melanoma-associated antigen indicated that a more primitive melanoma phenotype revealed overexpression of CD20 (16). Importantly, CD20 was initially identified on a small percentage of human melanoma cells when cultured in embryonic stem cell medium and found on nonadherent spheres. These CD20+ melanoma cells with the ability of self-renewal and differentiation followed the definition of tumor stem cells. Consistent with the view that cancer stem cells occupy a small part of tumors, CD20⁺ cells only account for $\sim 2\%$ of the total number of melanoma cells (74). However, CD20⁺ melanoma cells were demonstrated to be highly tumorigenic in vivo following xenotransplantation, suggesting that these cells exhibit tumor-initiating capacity (16). A previous study reported that the melanoma cells WM115 in the non-adherent form (melanoma spheroid cells) express a higher level of CD20 compared with adherent WM115 cells and that it is more likely to develop tumor when melanoma spheroid cells are transplanted into mice (16). Similarly to all stem cells, melanoma spheroid cells are also capable of proliferation, differentiation and self-renewal (16). In the late stage of metastatic melanoma, the effect of targeted therapy with anti-CD20 monoclonal antibody is more significant than that of non-targeted therapy and can even achieve a clinical complete response (75-77). In melanoma patients resistant to chemotherapeutic drugs, intratumoral injections of rituximab, which is the specific antibody against CD20, induces a regression of tumor growth, accompanied by a significant decrease in serum levels of inflammatory markers (77). Although there is no clear evidence that CD20+ MSCs are directly involved in melanoma metastasis, the stem cell-like characteristics of CD20⁺ melanoma cells have been confirmed, and their high tumorigenicity and migration ability are the main reasons for tumor progression (16,76). CD20 may therefore be considered as a new target for melanoma treatment in the future.

ALDH. ALDH represents a group of isoenzymes that can oxidize acetaldehyde to acetic acid. The enzymatic activity of ALDH has been used to identify stem or progenitor cells from various malignancies including breast, colon and lung cancers (78-80). According to previous studies, ALDH, which is a marker in many CSCs (78), is associated with multidrug resistance and immune tolerance of different types of solid tumor (78,81-83), can inhibit oxidative stress and enhance resistance to chemotherapeutic drugs, such as oxazolidine, taxanes and platinum drugs (84-86). It has been reported that melanoma cells with high expression of ALDH exhibit MSCs characteristics (87,88). Furthermore, ALDH-positive melanoma cells are more resistant to chemotherapeutic agents, and silencing ALDH1A using small interfering RNA can sensitize melanoma cells to drug-induced cell death (88). ALDH^{high} cells (melanoma cells with high expression of ALDH) can produce more melanoma clone spheres than ALDH^{low} cells (melanoma cells with low expression of ALDH) (87); however, the ability of melanoma formation in vivo is significantly inhibited following ALDH silencing (88). For example, following downregulation of aryl hydrocarbon receptor (AHR) and/or ALDH1 in mouse melanoma B16F10 cells through retroviral transduction, Contador et al (89) demonstrated that ALDH1 downregulation could inhibit the metastasis of melanoma cells without AHR expression, reduce the number of CD133⁺/CD29⁺/CD44⁺ cells and the size of melanospheres, confirming that overactivation of ALDH1 could promote the progression of melanoma in the context of AHR deficiency. Similarly, ALDH1 silencing in melanoma cells using short hairpin (sh)RNA significantly delays the appearance and growth of xenograft melanoma and dramatically decreases the number and load of metastases in mice (90). These studies confirm that targeting ALDH1 may be considered as an effective strategy for the treatment of advanced melanoma.

Other molecular markers of MSCs. Sox10 is a key nuclear transcription factor involved in the malignant transformation of melanocytes that has the potential to be a marker of MSCs (91). The positive rate of Sox10 in sentinel lymph node micrometastasis of melanoma is close to 100%, which is significantly higher than other melanoma markers responsible for melanoma metastasis, such as S100, HMB45 and Melania (91). Furthermore, it has also been reported that the stem cell function of CD271⁺ melanoma cells may be correlated to the CD271/Sox10 interaction network, although the underlying mechanism remains unclear (59,92). High levels of organic cation transporter (OCT)3/4, Nanog homeobox (Nanog) and Sox10 are found in CD133⁺ transgenic mice and human melanoma cells, and were demonstrated to promote tumor neovascularization, indicating that Sox10 and other MSCs markers can regulate each other and accelerate tumor progression (26). However, Sox10 silencing in human melanoma cells suppresses neural crest stem cell properties, inhibits cell proliferation and survival and completely abolishes in vivo tumor formation (93). Sox10 may therefore represent a promising target for the treatment of congenital naevi and melanoma in human patients.

C-X-C motif chemokine receptor 6 (CXCR6) is also an important marker of MSCs. Compared with ATP binding cassette subfamily G member 2 (ABCG2)⁺ melanoma cells, CXCR6⁺ melanoma cells can produce larger tumors in a shorter time, and ABCG2⁺ and CXCR6⁺ double-positive cells have a higher tumorigenic potential than ABCG2⁺ or CXCR6⁺ single-positive melanoma cells in promoting tumor metastasis (94).

CD44 has been used as a specific marker of MSCs in preclinical previous studies (95,96). It has been reported that blocking insulin-like growth factor-1 can prevent the metastatic and EMT processes of melanoma cells by downregulating the stem cell markers Sox2, OCT3/4, CD44 CD133 and deleting stem cell functional characteristics (97). CD44 may therefore serve an important role in melanoma metastasis.

3. Melanoma stem cells and angiogenesis

The process of angiogenesis is an important hallmark of the growth and progression of tumors, including melanoma (98). Evidence has shown that tumor vessels are derived from capillaries and veins in the host tissue, following activation by pro-angiogenic factors of pre-existing endothelial cells migrating into the tumor and developing into new vessel networks (99). The endothelial progenitor cells found in the peripheral circulation also contribute to tumor angiogenesis by differentiating and proliferating in a local tumor (100). Similarly, mesenchymal stem cells promote the growth and angiogenesis of tumors thanks to

their self-renewal capacity, long-term viability and differentiation potential toward diverse cell types (101-104). For example, CD133⁺ glioma cells secrete higher levels of VEGF than CD133⁻ cells, which indicates that tumor stem cells promote tumor angiogenesis (105). Furthermore, MSCs, which are involved in tumor angiogenesis, can accelerate melanoma metastasis. The renal and melanoma derived-CSCs are able to differentiate into endothelial like cells when cultured in endothelial cell growth specific medium (26,106). In addition, melanoma cells with stem cell-like characteristics, particularly those locating at the margin of the tumor, such as melanoma initiating cells, express and deliver in the microenvironment several factors (including VEGF, bFGF and PDGF) associated with angiogenesis (20). Since MSCs have high degree of differentiation plasticity, they can contribute to the *de novo* formation of tumor angiogenesis via a process named vasculogenic mimicry (VM) (107). Interestingly, tumor cells with abundant VM have a high plasticity, and their ability to mimic vascular endothelial cells may be related to the stemness of tumor cells (107,108). It has been reported that increasing the expression of stem cell-like genes in melanoma can improve the plasticity of tumor cells (108,109). Furthermore, ABCB5⁺ MSCs express specific endothelial and proangiogenic factors as well as VE-cadherin, Tie2, VEGF and its receptors, which are specific markers of VM (46). Consistently with these observations, melanoma cells expressing the CD133 and ABCG2 stem cells markers overexpress proangiogenic proteins, such as VEGF and its receptor VEGFR-2, Tie2 and angiopoietin (27). Furthermore, CD271⁺ MSCs were demonstrated to be associated with VM, through activation of the VEGR receptor/PKC signaling pathway (64,110). Thus, MSCs in angiogenesis contribute to melanoma growth and metastasis.

4. Targeting molecular markers of melanoma stem cells

The molecular markers of MSCs are of relative specificity and can serve as targets of molecular targeted therapy. They not only contribute to the removal of MSCs but also prevent normal cells from being damaged, in order to achieve the highest benefit of tumor therapy. For example, tubacin, which is an inhibitor of histone deacetylase 6, can promote the release of CD133⁺ exosomes in the human metastatic melanoma cells FEMX-I, decrease the content of CD133 in cells and inhibit the proliferation and clonogenesis of FEMX-I cells (111). In addition, monoclonal antibodies against different epitopes of CD133 have exhibited a dose-dependent cytotoxicity in FEMX-I cells (28,112). Furthermore, CD133 monoclonal antibody can inhibit the proliferation of FEMX-I melanoma cells and prevent the growth of melanoma through its cytotoxic effect. It was also demonstrated that CD133 downregulation by shRNA can decrease the appearance of lung and spinal cord metastases from melanoma (28). In addition, andrographolide can block the expression of notch1-dependent CD133 in melanoma cells and decrease the activation of MAPK signaling pathway, leading thus to inhibition of tumor growth, angiogenesis and metastasis (26). Similarly, the elimination of chemoresistant ABCB5-positive cells may significantly inhibit the overall growth of xenotransplanted melanomas by a selective antibody (37). ABCB5 is involved in the in vitro and in vivo survival of melanoma cells following exposure to dacarbazine and the BRAF inhibitor vemurafenib (113), and

systemic administration of anti-ABCB5 antibody inhibits melanoma tumorigenesis in nude mice (37). Furthermore, cuprous oxide nanoparticles can significantly decrease the expression of Sox10 and CD271, which accelerates the apoptosis and inhibits the tumorigenicity of CD271 overexpressing A375 and WM266-4 cells (114). Lunasin can decrease the expression level of ALDH, a marker of MSCs, and the expression level of Nanog, a stem cell-related factor. Lunasin can also promote the upregulation of microphthalmia associated transcription factor that inhibits the colony-forming ability of tumor cells and the growth of xenograft tumors, and can increase the transformation from $ALDH^{\rm high}$ to $ALDH^{\rm low}$ melanoma cells (115). Furthermore, magnolol can decrease the expression of CD271, CD166, JARID1B and ABCB5 through reducing the expression level of notch 2, a downstream target protein of HES-1, and of the cell cycle-related protein cyclin D1, inhibiting therefore the proliferation of melanoma cells and inducing their autophagy (116). In a clinical trial, rituximab, an anti-CD20 monoclonal antibody, was used to treat CD20⁺ metastatic melanoma. The anti-CD20 monoclonal antibody can eliminate CD20⁺ melanoma cells and increase the level of peripheral B cells in patients with melanoma (77). In an in vivo and in vitro study of melanoma cells, MSCs-induced angiogenesis mimicry was shown be a potential biological target for some anticancer compounds, such as the natural phytochemicals lupeol, which can prevent tumor metastasis by inhibiting angiogenesis (117). However, current research mainly uses cell and animal models, and large-scale clinical trials are required to support these conclusions.

5. Conclusion

The present review on MSCs provided some insights into the metastasis, recurrence, drug resistance and treatment of melanoma. MSCs serve a crucial role in the occurrence and progression of melanoma, especially in tumor metastasis. MSCs can promote tumor progression via MSCs-specific markers and the subsequent regulation of related signaling pathways. It is therefore crucial to fully understand the underlying mechanisms of MSCs markers in the process of tumor metastasis, which would allow the discovery of effective targets for the targeted therapy of melanoma. By exploring MSCs markers and the related signal transduction pathways, we believe that effective treatment strategies to inhibit tumor metastasis or eradicate melanoma might be discovered in the near future.

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Authors' contributions

QY drafted the initial manuscript, and edited and critically revised the manuscript. HJ and DW gave guidance on the conception and design of the review. XS and SL were involved in conceiving and designing the review. All authors have read and approved the final manuscript. Data authentication is not applicable.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Conmpeting interests

The authors declare that they have no competing interests.

References

- Lo JA and Fisher DE: The melanoma revolution: From UV carcinogenesis to a new era in therapeutics. Science 46: 945-949, 2014.
- Holderfield M, Deuker MM, McCormick F and McMahon M: Targeting RAF kinases for cancer therapy: BRAF-mutated melanoma and beyond. Nat Rev Cancer 14: 455-467, 2014.
- Chapman PB, Hauschild A, Robert C, Haanen JB, Ascierto P, Larkin J, Dummer R, Garbe C, Testori A, Maio M, *et al*: Improved survival with vemurafenib in melanoma with BRAF V600E mutation. N Engl J Med 364: 2507-2516, 2011.
- Flaherty KT, Robert C, Hersey P, Nathan P, Garbe C, Milhem M, Demidov LV, Hassel JC, Rutkowski P, Mohr P, *et al*: Improved survival with MEK inhibition in BRAF-mutated melanoma. N Engl J Med 367: 107-114, 2012.
- Christiansen SA, Khan S and Gibney GT: Targeted therapies in combination with immune therapies for the treatment of metastatic melanoma. Cancer J 23: 59-62, 2017.
- 6. Nandy SB and Lakshmanaswamy R: Cancer stem cells and metastasis. Prog Mol Biol Transl Sci 151: 137-176, 2017.
- Parmiani G: Melanoma cancer stem cells: Markers and functions. Cancers (Basel) 8: 34, 2016.
- Fink J, Andersson-Rolf A and Koo BK: Adult stem cell lineage tracing and deep tissue imaging. BMB Rep 48: 655-667, 2015.
- Ricci E, Mattei E, Dumontet C, Eaton CL, Hamdy F, van der Pluije G, Cecchini M, Thalmann G, Clezardin P and Colombel M: Increased expression of putative cancer stem cell markers in the bone marrow of prostate cancer patients is associated with bone metastasis progression. Prostate 73: 1738-1746, 2013.
- Croker AK and Allan AL: Cancer stem cells: Implications for the progression and treatment of metastatic disease. J Cell Mol Med 12: 374-390, 2008.
- 11. Li F, Tiede B, Massague J and Kang Y: Beyond tumorigenesis: Cancer stem cells in metastasis. Cell Res 17: 3-14, 2007.
- Reya T, Morrison SJ, Clarke MF and Weissman IL: Stem cells, cancer, and cancer stem cells. Nature 414: 105-111, 2001.
- Wickremesekera AC, Brasch HD, Lee VM, Davis PF, Woon K, Johnson R, Tan ST and Itinteang T: Expression of cancer stem cell markers in metastatic melanoma to the brain. J Clin Neurosci 60: 112-116, 2019.
- Nguyen N, Couts KL, Luo Y and Fujita M: Understanding melanoma stem cells. Melanoma Manag 2: 179-188, 2015.
- Klein WM, Wu BP, Zhao S, Wu H, Klein-Szanto AJ and Tahan SR: Increased expression of stem cell markers in malignant melanoma. Mod Pathol 20: 102-107, 2007.

- Fang D, Nguyen TK, Leishear K, Finko R, Kulp AN, Hotz S, Van Belle PA, Xu X, Elder DE and Herlyn M: A tumorigenic subpopulation with stem cell properties in melanomas. Cancer Res 65: 9328-9337, 2005.
- Tirino V, Desiderio V, Paino F, De Rosa A, Papaccio F, Fazioli F, Pirozzi G and Papaccio G: Human primary bone sarcomas contain CD133⁺ cancer stem cells displaying high tumorigenicity in vivo. FASEB J 25: 2022-2030, 2011.
- Desiderio V, Papagerakis P, Tirino V, Zheng L, Matossian M, Prince ME, Paino F, Mele L, Papaccio F, Montella R, *et al*: Increased fucosylation has a pivotal role in invasive and metastatic properties of head and neck cancer stem cells. Oncotarget 6: 71-84, 2015.
- Collins AT, Berry PA, Hyde C, Stower MJ and Maitland NJ: Prospective identification of tumorigenic prostate cancer stem cells. Cancer Res 65: 10946-10951, 2005.
- 20. Marzagalli M, Raimondi M, Fontana F, Montagnani Marelli M, Moretti RM and Limonta P: Cellular and molecular biology of cancer stem cells in melanoma: Possible therapeutic implications. Semin Cancer Biol 59: 221-235, 2019.
- 21. Kumar D, Gorain M, Kundu G and Kundu GC: Therapeutic implications of cellular and molecular biology of cancer stem cells in melanoma. Mol Cancer 16: 7, 2017.
- 22. Lee N, Barthel SR and Schatton T: Melanoma stem cells and metastasis: Mimicking hematopoietic cell trafficking? Lab Invest 94: 13-30, 2014.
- 23. Shakhova O and Sommer L: Testing the cancer stem cell hypothesis in melanoma: The clinics will tell. Cancer Lett 338 : 74-81, 2013.
- Ricci-Vitiani L, Lombardi DG, Pilozzi E, Biffoni M, Todaro M, Peschle C and De Maria R: Identification and expansion of human colon-cancer-initiating cells. Nature 445: 111-115, 2007.
- 25. Zhang D, Tang DG and Rycaj K: Cancer stem cells: Regulation programs, immunological properties and immunotherapy. Semin Cancer Biol 52: 94-106, 2018.
- 26. Kumar D, Kumar S, Gorain M, Tomar D, Patil HS, Radharani NNV, Kumar TVS, Patil TV, Thulasiram HV and Kundu GC: Notch1-MAPK signaling axis regulates CD133(+) cancer stem cell-mediated melanoma growth and angiogenesis. J Invest Dermatol 136: 2462-2474, 2016.
- 27. Monzani E, Facchetti F, Galmozzi E, Corsini E, Benetti A, Cavazzin C, Gritti A, Piccinini A, Porro D, Santinami M, et al: Melanoma contains CD133 and ABCG2 positive cells with enhanced tumourigenic potential. Eur J Cancer 43: 935-946, 2007.
- Rappa G, Fodstad O and Lorico A: The stem cell-associated antigen CD133 (Prominin-1) is a molecular therapeutic target for metastatic melanoma. Stem Cells 26: 3008-3017, 2008.
- Houben R, Wischhusen J, Menaa F, Synwoldt P, Schrama D, Brocker EB and Becker JC: Melanoma stem cells: Targets for successful therapy? J Dtsch Dermatol Ges 6: 541-546, 2008.
- Lai CY, Schwartz BE and Hsu MY: CD133⁺ melanoma subpopulations contribute to perivascular niche morphogenesis and tumorigenicity through vasculogenic mimicry. Cancer Res 72: 5111-5118, 2012.
- Zimmerer RM, Matthiesen P, Kreher F, Kampmann A, Spalthoff S, Jehn P, Bittermann G, Gellrich NC and Tavassol F: Putative CD133⁺ melanoma cancer stem cells induce initial angiogenesis in vivo. Microvasc Res 104: 46-54, 2016.
- 32. Sharma BK, Manglik V, O'Connell M, Weeraratna A, McCarronEC, BroussardJN, DivitoKA, Simbulan-RosenthalCM, Rosenthal DS and Zapas JL: Clonal dominance of CD133⁺ subset population as risk factor in tumor progression and disease recurrence of human cutaneous melanoma. Int J Oncol 41: 1570-1576, 2012.
- 33. El-Khattouti A, Selimovic D, Haikel Y, Megahed M, Gomez CR and Hassan M: Identification and analysis of CD133(+) melanoma stem-like cells conferring resistance to taxol: An insight into the mechanisms of their resistance and response. Cancer Lett 343: 123-133, 2014.
- 34. Koshio J, Kagamu H, Nozaki K, Saida Y, Tanaka T, Shoji S, Igarashi N, Miura S, Okajima M, Watanabe S, *et al*: DEAD/H (Asp-Glu-Ala-Asp/His) box polypeptide 3, X-linked is an immunogenic target of cancer stem cells. Cancer Immunol Immunother 62: 1619-1628, 2013.
- 35. Gedye C, Quirk J, Browning J, Svobodova S, John T, Sluka P, Dunbar PR, Corbeil D, Cebon J and Davis ID: Cancer/testis antigens can be immunological targets in clonogenic CD133⁺ melanoma cells. Cancer Immunol Immunother 58: 1635-1646, 2009.
- Lang D, Mascarenhas JB and Shea CR: Melanocytes, melanocyte stem cells, and melanoma stem cells. Clin Dermatol 31: 166-178, 2013.

- 37. Schatton T, Murphy GF, Frank NY, Yamaura K, Waaga-Gasser AM, Gasser M, Zhan Q, Jordan S, Duncan LM, Weishaupt C, *et al*: Identification of cells initiating human melanomas. Nature 451: 345-349, 2008.
- Begicevic RR and Falasca M: ABC transporters in cancer stem cells: Beyond chemoresistance. Int J Mol Sci 18: 2362, 2017.
- 39. Frank NY, Margaryan A, Huang Y, Schatton T, Waaga-Gasser AM, Gasser M, Sayegh MH, Sadee W and Frank MH: ABCB5-mediated doxorubicin transport and chemoresistance in human malignant melanoma. Cancer Res 65: 4320-4333, 2005.
- 40. Roesch A, Fukunaga-Kalabis M, Schmidt EC, Zabierowski SE, Brafford PA, Vultur A, Basu D, Gimotty P, Vogt T and Herlyn M: A temporarily distinct subpopulation of slow-cycling melanoma cells is required for continuous tumor growth. Cell 141: 583-594, 2010.
- 41. Wang S, Tang L, Lin J, Shen Z, Yao Y, Wang W, Tao S, Gu C, Ma J, Xie Y and Liu Y: ABCB5 promotes melanoma metastasis through enhancing NF-κB p65 protein stability. Biochem Biophys Res Commun 492: 18-26, 2017.
- 42. Xiao J, Egger ME, McMasters KM and Hao H: Differential expression of ABCB5 in BRAF inhibitor-resistant melanoma cell lines. BMC Cancer 18: 675, 2018.
- 43. de Waard NE, Kolovou PE, McGuire SP, Cao J, Frank NY, Frank MH, Jager MJ and Ksander BR: Expression of multidrug resistance transporter ABCB5 in a murine model of human conjunctival melanoma. Ocul Oncol Pathol 1: 182-189, 2015.
- 44. Vasquez-Moctezuma I, Meraz-Rios MA, Villanueva-Lopez CG, Magana M, Martinez-Macias R, Sanchez-Gonzalez DJ, García-Sierra F and Herrera-González NE: ATP-binding cassette transporter ABCB5 gene is expressed with variability in malignant melanoma. Actas Dermosifiliogr 101: 341-348, 2010.
- 45. Ma J and Frank MH: Isolation of circulating melanoma cells. Methods Mol Biol: Sep 29, 2015 (Epub ahead of print). doi: https://doi.org/10.1007/7651_2015_300.
 46. Frank NY, Schatton T, Kim S, Zhan Q, Wilson BJ, Ma J,
- 46. Frank NY, Schatton T, Kim S, Zhan Q, Wilson BJ, Ma J, Saab KR, Osherov V, Widlund HR, Gasser M, *et al*: VEGFR-1 expressed by malignant melanoma-initiating cells is required for tumor growth. Cancer Res 71: 1474-1485, 2011.
- 47. Schatton T, Schutte U, Frank NY, Zhan Q, Hoerning A, Robles SC, Zhou J, Hodi FS, Spagnoli GC, Murphy GF and Frank MH: Modulation of T-cell activation by malignant melanoma initiating cells. Cancer Res 70: 697-708, 2010.
- Eggermont AM and Robert C: New drugs in melanoma: It's a whole new world. Eur J Cancer 47: 2150-2157, 2011.
- 49. Boiko AD, Razorenova OV, van de Rijn M, Swetter SM, Johnson DL, Ly DP, Butler PD, Yang GP, Joshua B, Kaplan MJ, *et al*: Human melanoma-initiating cells express neural crest nerve growth factor receptor CD271. Nature 466: 133-137, 2010.
- 50. Chesa PG, Rettig WJ, Thomson TM, Old LJ and Melamed MR: Immunohistochemical analysis of nerve growth factor receptor expression in normal and malignant human tissues. J Histochem Cytochem 36: 383-389, 1988.
- 51. Pietra G, Manzini C, Vitale M, Balsamo M, Ognio E, Boitano M, Queirolo P, Moretta L and Mingari MC: Natural killer cells kill human melanoma cells with characteristics of cancer stem cells. Int Immunol 21: 793-801, 2009.
- 52. Truzzi F, Marconi A, Lotti R, Dallaglio K, French LE, Hempstead BL and Pincelli C: Neurotrophins and their receptors stimulate melanoma cell proliferation and migration. J Invest Dermatol 128: 2031-2040, 2008.
- Nielsen PS, Riber-Hansen R and Steiniche T: Immunohistochemical CD271 expression correlates with melanoma progress in a case-control study. Pathology 50: 402-410, 2018.
- 54. Guo R, Fierro-Fine A, Goddard L, Russell M, Chen J, Liu CZ, Fung KM and Hassell LA: Increased expression of melanoma stem cell marker CD271 in metastatic melanoma to the brain. Int J Clin Exp Pathol 7: 8947-8951, 2014.
- Denkins Y, Reiland J, Roy M, Sinnappah-Kang ND, Galjour J, Murry BP, Blust J, Aucoin R and Marchetti D: Brain metastases in melanoma: Roles of neurotrophins. Neuro Oncol 6: 154-165, 2004.
- Shaffer SM, Dunagin MC, Torborg SR, Torre EA, Emert B, Krepler C, Beqiri M, Sproesser K, Brafford PA, Xiao M, *et al*: Rare cell variability and drug-induced reprogramming as a mode of cancer drug resistance. Nature 546: 431-435, 2017.
 Holzel M and Tuting T: Inflammation-induced plasticity in
- 57. Holzel M and Tuting T: Inflammation-induced plasticity in melanoma therapy and metastasis. Trends Immunol 37: 364-374, 2016.

- 58. Restivo G, Diener J, Cheng PF, Kiowski G, Bonalli M, Biedermann T, Reichmann E, Levesque MP, Dummer R and Sommer L: low neurotrophin receptor CD271 regulates phenotype switching in melanoma. Nat Commun 8: 1988, 2017.
- 59. Redmer T, Welte Y, Behrens D, Fichtner I, Przybilla D, Wruck W, Yaspo ML, Lehrach H, Schäfer R and Regenbrecht CR: The nerve growth factor receptor CD271 is crucial to maintain tumorigenicity and stem-like properties of melanoma cells. PLoS One 9: e92596, 2014.
- 60. Prasmickaite L, Skrbo N, Hoifodt HK, Suo Z, Engebraten O, Gullestad HP, Aamdal S, Fodstad Ø and Maelandsmo GM: Human malignant melanoma harbours a large fraction of highly clonogenic cells that do not express markers associated with cancer stem cells. Pigment Cell Melanoma Res 23: 449-451, 2010.
- Lehraiki A, Cerezo M, Rouaud F, Abbe P, Allegra M, Kluza J, Marchetti P, Imbert V, Cheli Y, Bertolotto C, *et al*: Increased CD271 expression by the NF-κB pathway promotes melanoma cell survival and drives acquired resistance to BRAF inhibitor vemurafenib. Cell Discov 1: 15030, 2015.
 Schnegg CI, Yang MH, Ghosh SK and Hsu MY: Induction of
- Schnegg CI, Yang MH, Ghosh SK and Hsu MY: Induction of vasculogenic mimicry overrides VEGF-A silencing and enriches stem-like cancer cells in melanoma. Cancer Res 75: 1682-1690, 2015.
- 63. Valyi-Nagy K, Kormos B, Ali M, Shukla D and Valyi-Nagy T: Stem cell marker CD271 is expressed by vasculogenic mimicry-forming uveal melanoma cells in three-dimensional cultures. Mol Vis 18: 588-592, 2012.
- 64. Gray ES, Reid AL, Bowyer S, Calapre L, Siew K, Pearce R, Cowell L, Frank MH, Millward M and Ziman M: Circulating melanoma cell subpopulations: Their heterogeneity and differential responses to treatment. J Invest Dermatol 135: 2040-2048, 2015.
- 65. Civenni G, Walter A, Kobert N, Mihic-Probst D, Zipser M, Belloni B, Seifert B, Moch H, Dummer R, van den Broek M and Sommer L: Human CD271-positive melanoma stem cells associated with metastasis establish tumor heterogeneity and long-term growth. Cancer Res 71: 3098-3109, 2011.
- 66. Furuta J, Inozume T, Harada K and Shimada S: CD271 on melanoma cell is an IFN-γ-inducible immunosuppressive factor that mediates downregulation of melanoma antigens. J Invest Dermatol 134: 1369-1377, 2014.
- 67. Roesch A: Melanoma stem cells. J Dtsch Dermatol Ges 13: 118-124, 2015.
- Clarke MF and Fuller M: Stem cells and cancer: Two faces of eve. Cell 124: 1111-1115, 2006.
- Held MA, Curley DP, Dankort D, McMahon M, Muthusamy V and Bosenberg MW: Characterization of melanoma cells capable of propagating tumors from a single cell. Cancer Res 70: 388-397, 2010.
- Frank NY, Schatton T and Frank MH: The therapeutic promise of the cancer stem cell concept. J Clin Invest 120: 41-50, 2010.
- Boonyaratanakornkit JB, Yue L, Strachan LR, Scalapino KJ, LeBoit PE, Lu Y, Leong SP, Smith JE and Ghadially R: Selection of tumorigenic melanoma cells using ALDH. J Invest Dermatol 130: 2799-2808, 2010.
- 72. Pinc A, Somasundaram R, Wagner C, Hormann M, Karanikas G, Jalili A, Bauer W, Brunner P, Grabmeier-Pfistershammer K, Gschaider M, *et al*: Targeting CD20 in melanoma patients at high risk of disease recurrence. Mol Ther 20: 1056-1062, 2012.
- Akbulut H, Babahan C, Abgarmi SA, Ocal M and Besler M: Recent advances in cancer stem cell targeted therapy. Crit Rev Oncog 24: 1-20, 2019.
- 74. Yaiza JM, Gloria RA, Maria Belen GO, Elena LR, Gema J, Juan Antonio M, María Ángel GC and Houria B: Melanoma cancer stem-like cells: Optimization method for culture, enrichment and maintenance. Tissue Cell 60: 48-59, 2019.
- 75. Schmidt P, Kopecky C, Hombach A, Zigrino P, Mauch C and Abken H: Eradication of melanomas by targeted elimination of a minor subset of tumor cells. Proc Natl Acad Sci USA 108: 2474-2479, 2011.
- Murphy GF, Wilson BJ, Girouard SD, Frank NY and Frank MH: Stem cells and targeted approaches to melanoma cure. Mol Aspects Med 39: 33-49, 2014.
- 77. Schlaak M, Schmidt P, Bangard C, Kurschat P, Mauch C and Abken H: Regression of metastatic melanoma in a patient by antibody targeting of cancer stem cells. Oncotarget 3: 22-30, 2012.

- 78. Ginestier C, Hur MH, Charafe-Jauffret E, Monville F, Dutcher J, Brown M, Jacquemier J, Viens P, Kleer CG, Liu S, *et al*: ALDH1 is a marker of normal and malignant human mammary stem cells and a predictor of poor clinical outcome. Cell Stem Cell 1: 555-567, 2007.
- 79. Huang EH, Hynes MJ, Zhang T, Ginestier C, Dontu G, Appelman H, Fields JZ, Wicha MS and Boman BM: Aldehyde dehydrogenase 1 is a marker for normal and malignant human colonic stem cells (SC) and tracks SC overpopulation during colon tumorigenesis. Cancer Res 69: 3382-3389, 2009.
- Jiang F, Qiu Q, Khanna A, Todd NW, Deepak J, Xing L, Wang H, Liu Z, Su Y, Stass SA and Katz RL: Aldehyde dehydrogenase 1 is a tumor stem cell-associated marker in lung cancer. Mol Cancer Res 7: 330-338, 2009.
- Carpentino JE, Hynes MJ, Appelman HD, Zheng T, Steindler DA, Scott EW and Huang EH: Aldehyde dehydrogenase-expressing colon stem cells contribute to tumorigenesis in the transition from colitis to cancer. Cancer Res 69: 8208-8215, 2009.
- Roudi R, Korourian A, Shariftabrizi A and Madjd Z: Differential expression of cancer stem cell markers ALDH1 and CD133 in various lung cancer subtypes. Cancer Invest 33: 294-302, 2015.
- Yoshida A, Hsu LC and Dave V: Retinal oxidation activity and biological role of human cytosolic aldehyde dehydrogenase. Enzyme 46: 239-244, 1992.
- 84. Singh S, Brocker C, Koppaka V, Chen Y, Jackson BC, Matsumoto A, Thompson DC and Vasiliou V: Aldehyde dehydrogenases in cellular responses to oxidative/electrophilic stress. Free Radic Biol Med 56: 89-101, 2013.
- 85. Le Moguen K, Lincet H, Deslandes E, Hubert-Roux M, Lange C, Poulain L, Gauduchon P and Baudin B: Comparative proteomic analysis of cisplatin sensitive IGROV1 ovarian carcinoma cell line and its resistant counterpart IGROV1-R10. Proteomics 6: 5183-5192, 2006.
- 86. Moreb JS, Gabr A, Vartikar GR, Gowda S, Zucali JR and Mohuczy D: Retinoic acid down-regulates aldehyde dehydrogenase and increases cytotoxicity of 4-hydroperoxycyclophosphamide and acetaldehyde. J Pharmacol Exp Ther 312: 339-345, 2005.
- 87. Santini R, Vinci MC, Pandolfi S, Penachioni JY, Montagnani V, Olivito B, Gattai R, Pimpinelli N, Gerlini G, Borgognoni L and Stecca B: Hedgehog-GLI signaling drives self-renewal and tumorigenicity of human melanoma-initiating cells. Stem Cells 30: 1808-1818, 2012.
- 88. Luo Y, Dallaglio K, Chen Y, Robinson WA, Robinson SE, McCarter MD, Wang J, Gonzalez R, Thompson DC, Norris DA, et al: ALDH1A isozymes are markers of human melanoma stem cells and potential therapeutic targets. Stem Cells 30: 2100-2113, 2012.
- 89. Contador-Troca M, Alvarez-Barrientos A, Merino JM, Morales-Hernandez A, Rodriguez MI, Rey-Barroso J, Barrasa E, Cerezo-Guisado MI, Catalina-Fernández I, Sáenz-Santamaría J, et al: Dioxin receptor regulates aldehyde dehydrogenase to block melanoma tumorigenesis and metastasis. Mol Cancer 14: 148, 2015.
- 90. Yue L, Huang ZM, Fong S, Leong S, Jakowatz JG, Charruyer-Reinwald A, Wei M and Ghadially R: Targeting ALDH1 to decrease tumorigenicity, growth and metastasis of human melanoma. Melanoma Res 25: 138-148, 2015.
- Willis BC, Johnson G, Wang J and Cohen C: SOX10: A useful marker for identifying metastatic melanoma in sentinel lymph nodes. Appl Immunohistochem Mol Morphol 23: 109-112, 2015.
- 92. Paratore C, Goerich DE, Suter U, Wegner M and Sommer L: Survival and glial fate acquisition of neural crest cells are regulated by an interplay between the transcription factor Sox10 and extrinsic combinatorial signaling. Development 128: 3949-3961, 2001.
- 93. Shakhova O, Zingg D, Schaefer SM, Hari L, Civenni G, Blunschi J, Claudinot S, Okoniewski M, Beermann F, Mihic-Probst D, *et al*: Sox10 promotes the formation and maintenance of giant congenital naevi and melanoma. Nat Cell Biol 14: 882-890, 2012.
- 94. Taghizadeh R, Noh M, Huh YH, Ciusani E, Sigalotti L, Maio M, Arosio B, Nicotra MR, Natali P, Sherley JL and La Porta CA: CXCR6, a newly defined biomarker of tissue-specific stem cell asymmetric self-renewal, identifies more aggressive human melanoma cancer stem cells. PLoS One 5: e15183, 2010.
- 95. Zhao F, Zhang R, Wang J, Wu D, Pan M, Li M, Guo M and Dou J: Effective tumor immunity to melanoma mediated by B16F10 cancer stem cell vaccine. Int Immunopharmacol 52: 238-244, 2017.

- 96. Dou J, He X, Liu Y, Wang Y, Zhao F, Wang X, Chen D, Shi F and Wang J: Effect of downregulation of ZEB1 on vimentin expression, tumour migration and tumourigenicity of melanoma B16F10 cells and CSCs. Cell Biol Int 38: 452-461, 2014.
- 97. Le Coz V, Zhu C, Devocelle A, Vazquez A, Boucheix C, Azzi S, Gallerne C, Eid P, Lecourt S and Giron-Michel J: IGF-1 contributes to the expansion of melanoma-initiating cells through an epithelial-mesenchymal transition process. Oncotarget 7: 82511-82527, 2016.
- 98. RS K: Tumor angiogenesis. N Engl J Med 358: 2039-2049, 2008.
- 99. Carmeliet P and Jain RK: Angiogenesis in cancer and other diseases. Nature 407: 249-257, 2000.
- 100. Rafii S, Lyden D, Benezra R, Hattori K and Heissig B: Vascular and haematopoietic stem cells: novel targets for anti-angiogenesis therapy? Nat Rev Cancer 2: 826-835, 2002.
- 101. Huang WH, Chang MC, Tsai KS, Hung MC, Chen HL and Hung SC: Mesenchymal stem cells promote growth and angiogenesis of tumors in mice. Oncogene 32: 4343-4354, 2013.
- 102. Jeon ES, Lee IH, Heo SC, Shin SH, Choi YJ, Park JH, Park DY and Kim JH: Mesenchymal stem cells stimulate angiogenesis in a murine xenograft model of A549 human adenocarcinoma through an LPA1 receptor-dependent mechanism. Biochim Biophys Acta 1801: 1205-1213, 2010.
- 103. Otsu K, Das S, Houser SD, Quadri SK, Bhattacharya S and Bhattacharya J: Concentration-dependent inhibition of angiogenesis by mesenchymal stem cells. Blood 113: 4197-4205, 2009.
- 104. Sun B, Zhang S, Ni C, Zhang D, Liu Y, Zhang W, Zhao X, Zhao C and Shi M: Correlation between melanoma angiogenesis and the mesenchymal stem cells and endothelial progenitor cells derived from bone marrow. Stem Cells Dev 14: 292-298, 2005.
- 105. Bao S, Wu O, Sathornsumetee S, Hao Y, Li Z, Hjelmeland AB, Shi Q, McLendon RE, Bigner DD and Rich JN: Stem cell-like glioma cells promote tumor angiogenesis through vascular endothelial growth factor. Cancer Res 66: 7843-7848, 2006.
- 106. Bussolati B, Bruno S, Grange C, Ferrando U and Camussi G: Identification of a tumor-initiating stem cell population in human renal carcinomas. FASEB J 22: 3696-3705, 2008.
- 107. Hendrix MJ, Seftor EA, Hess AR and Seftor RE: Vasculogenic mimicry and tumour-cell plasticity: Lessons from melanoma. Nat Rev Cancer 3: 411-421, 2003.
- 108. Seftor RE, Hess AR, Seftor EA, Kirschmann DA, Hardy KM, Margaryan NV and Hendrix MJ: Tumor cell vasculogenic mimicry: From controversy to therapeutic promise. Am J Pathol 181: 1115-1125, 2012.

- 109. Girouard SD and Murphy GF: Melanoma stem cells: Not rare, but well done. Lab Invest 91: 647-664, 2011.
- 110. Vartanian A, Stepanova E, Grigorieva I, Solomko E, Baryshnikov A and Lichinitser M: VEGFR1 and PKCa signaling control melanoma vasculogenic mimicry in a VEGFR2 kinase-independent manner. Melanoma Res 21: 91-98, 2011.
- 111. Chao OS, Chang TC, Di Bella MA, Alessandro R, Anzanello F, Rappa G, Goodman OB and Lorico A: The HDAC6 inhibitor tubacin induces release of CD133(+) extracellular vesicles from cancer cells. J Cell Biochem 118: 4414-4424, 2017.
- 112. Alamodi AA, Eshaq AM, Hassan SY, Al Hmada Y, El Jamal SM, Fothan ÂM, Arain OM, Hassan SL, Haikel Y, Megahed M and Hassan M: Cancer stem cell as therapeutic target for melanoma treatment. Histol Histopathol 31: 1291-1301, 2016.
- 113. Luo Y, Ellis LZ, Dallaglio K, Takeda M, Robinson WA, Robinson SE, Liu W, Lewis KD, McCarter MD, Gonzalez R, et al: Side population cells from human melanoma tumors reveal diverse mechanisms for chemoresistance. J Invest Dermatol 132: 2440-2450, 2012. 114. Yu B, Wang Y, Yu X, Zhang H, Zhu J, Wang C, Chen F, Liu C,
- Wang J and Zhu H: Cuprous oxide nanoparticle-inhibited melanoma progress by targeting melanoma stem cells. Int J Nanomedicine 12: 2553-2567, 2017.
- 115. Shidal C, Al-Rayyan N, Yaddanapudi K and Davis KR: Lunasin is a novel therapeutic agent for targeting melanoma cancer stem cells. Oncotarget 7: 84128-84141, 2016.
- 116. Kaushik G, Venugopal A, Ramamoorthy P, Standing D, Subramaniam D, Umar S, Jensen RA, Anant S and Mammen JM: Honokiol inhibits melanoma stem cells by targeting notch signaling. Mol Carcinog 54: 1710-1721, 2015.
- 117. Bhattacharyya S, Mitra D, Ray S, Biswas N, Banerjee S, Majumder B, Mustafi SM and Murmu N: Reversing effect of Lupeol on vasculogenic mimicry in murine melanoma progression. Microvasc Res 121: 52-62, 2019.



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