



Elevated DNA Polymerase Delta 1 Expression Correlates With Tumor Progression and Immunosuppressive Tumor Microenvironment in Hepatocellular Carcinoma

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Background and Objective: Hepatocellular carcinoma (HCC) is one of the most common cancers worldwide, and the DNA polymerase delta (POLD) family is significantly related to cancer prognosis. This study aimed to explore the significance of the POLD family in HCC *via* the DNA damage repair (DDR) pathway.

Methods: Data mining was conducted using bioinformatics methods. RNA sequencing and clinicopathological data were collected from The Cancer Genome Atlas, GTEx database and the Gumz Renal cohort. Statistical analyses were also performed in cancer samples (n>12,000) and the Affiliated Hospital of Youjiang Medical University for Nationalities (AHYMUN, n=107) cohort.

Results: The POLD family (POLD1–4) was identified as the most important functional component of the DDR pathway. Based on the analysis of independent cohorts, we found significantly elevated POLD expression in HCC compared with normal tissues. Second, we investigated the prognostic implication of elevated POLD1 expression in HCC and pan-cancers, revealing that increased POLD1 levels were correlated to worse prognoses for HCC patients. Additionally, we identified 11 hub proteins interacting closely with POLD proteins in base excision repair, protein-DNA complex and mismatch repair signaling pathways. Moreover, POLD1 mutation functioned as an independent biomarker to predict the benefit of targeted treatment. Importantly, POLD1 expression was associated with immune checkpoint molecules, including CD274, CD80, CD86, CTLA4, PDCD1 and

November 2021 | Volume 11 | Article 736363

1

TCGIT, and facilitated an immune-excluded tumor microenvironment. Additionally, we confirmed that elevated POLD1 expression was closely correlated with the aggressive progression and poor prognosis of HCC in the real-world AHYMUN cohort.

Conclusion: We identified a significant association between elevated POLD1 expression and poor patient survival and immune-excluded tumor microenvironment of HCC. Together, these findings indicate that POLD1 provides a valuable biomarker to guide the molecular diagnosis and development of novel targeted therapeutic strategies for HCC patients.

Keywords: POLD1, hepatocellular carcinoma, prognosis, immune microenvironment, biomarker

INTRODUCTION

Hepatocellular carcinoma (HCC) is the most common type of primary liver cancer, with over half a million new cases diagnosed worldwide each year. HCC is the most malignant form of liver cancer, which derives from the aggressive proliferation of liver epithelial cells (1). As the sixth leading cause of cancer-related mortality globally, HCC accounts for 4.7% of all cancer deaths. In 2020, over 910,000 people were diagnosed with HCC, and its incidence is continuously rising (2). In China, the incidence of HCC is particularly high, accounting for 55% of the total number of HCC patients worldwide.

Several molecular pathways are implicated in HCC carcinogenesis, including vascular endothelial growth factor receptor (VEGFR), TP53 and Akt/mTOR pathways (3). Among several pathogenic factors, cirrhosis underlies most cases of HCC, and hepatitis C virus (HCV) infection also causes cirrhosis and HCC. However, the eradication of HCV-related cirrhosis does not prevent the development of HCC (4, 5). Thus, more effective HCC treatments are needed.

Historically, serum alpha-fetoprotein (AFP) and diagnostic imaging were the primary diagnostic methods for HCC. However, the poor prognosis of patients owing to the late diagnosis of HCC is unacceptable, and AFP levels are not significantly elevated in most small and early-stage HCCs (6). Recently, several studies have focused on identifying promising biomarkers for early detection (7, 8). Currently, targeted drugs, such as sorafenib, are used as a first-line treatment strategy for the personalized treatment of advanced HCC (9). However, exploring unknown targets and pathways is important to achieve more significant survival benefits for advanced HCC patients who cannot be cured through surgery.

DNA damage repair (DDR) pathways consist of multiple interconnected cellular signaling networks that are activated in response to DNA damage. DDR is correlated with genomic instability, tumor mutational burden in HCC and immune cell function (10). Several studies have shown that the development of cancer is mainly driven by defects in DDR. Therefore, inhibitors targeting kinases involved in DDR, such as AZ20 and M3814, have been investigated for the treatment of cancers in clinical trials (11).

The DNA polymerase delta (POLD) family consisting of POLD1, POLD2, POLD3 and POLD4 is an important

mediator of DNA repair during chromosome replication, which is of great significance to the maintenance of DNA structure stability (12). *POLD1* encodes the catalytic subunit of DNA polymerase delta, which is involved in both polymerase and 3' to 5' exonuclease activities. POLD1 plays a crucial role in DNA replication and DNA double-strand break repair, together with the accessory proteins POLD2, POLD3 and POLD4 for full activity (13, 14). POLD2 encodes the 50-kDa catalytic subunit of DNA polymerase delta, assisting POLD1 in the process of DDR.

In addition, POLD3 and POLD4 may catalyze the repair of damaged replication forks *via* break-induced replication (15). Recent studies have shown that the loss or reduced expression of POLD3 predicts the occurrence and progression of colorectal cancer and other malignant tumors (16, 17). The decreased expression of POLD3 is associated with the pathogenesis of HCC by enhancing the proliferation and invasion abilities of tumor cells (18).

However, the relationship between the expression of POLD family members and the prognosis of HCC remains poorly understood. Based on the role of the POLD family in the maintenance and repair of DNA, we hypothesized that POLD expression significantly affects the prognosis of HCC *via* the DDR pathway. Our study aims to identify a biomarker with significant potential to improve the diagnosis and prognosis of HCC.

METHODS

Patients and Tissue Samples From Online Databases and Real-World Cohorts

The large-scale cohorts in this study included RNA sequencing data and accompanying clinicopathological data for 423 HCC patients collected from The Cancer Genome Atlas (TCGA, http://www.cancer.gov) and 10 patients with HCC from the Gumz Renal cohort in the Oncomine (http://www.oncomine. com) database. The threshold was set as follows: *P*-value<0.05, gene rank>10% and data type: mRNA, Cancer *vs.* Normal.

The Affiliated Hospital of Youjiang Medical University for Nationalities (AHYMUN, Guangxi, China) cohort consisted of 107 patients diagnosed with HCC in the Department of Hepatology, Affiliated Hospital of Youjiang Medical University for Nationalities, from June 2009 to August 2018. Clinicopathological data were collected from pathology reports or electronic medical records. Samples of HCC and normal liver tissues were collected during surgery and then processed and stored at the AHYMUN tissue bank before experiments.

Immunohistochemistry (IHC) Staining Analysis

IHC was performed with an anti-POLD1 antibody-N-terminal (ab226848, Abcam) at a 1:500 dilution. IHC staining was conducted in accordance with the manufacturer's instructions as previously described (19). Based on the IHC staining intensity and density, two experienced and independent pathologists/ clinicians evaluated the overall IHC score (from 0 to 12), with a score of 0 to 3 indicating negative staining and a score of 4 to 12 indicating positive staining for each tissue.

Establishment of a Protein-Protein Interaction (PPI) Network and Functional Annotations of POLD-Related PPI Networks

In this study, an online web tool for the retrieval of interacting genes (STRING, http://string-db.org; version 10.0) was used to establish a PPI network of differentially expressed genes (DEGs) related to POLD. Then, Spearman's correlation analysis was performed to describe the correlation between quantitative variables without a normal distribution. The gene ontology (GO) database was used for functional enrichment analyses of biological processes (BP), molecular functions (MF) and cellular components (CC). Kyoto Encyclopedia of Genes and Genomes (KEGG) and Reactome pathway enrichment databases were used to assess large-scale biologic molecular datasets. The publicly available WEB-based GEne SeT AnaLysis Toolkit (WebGestalt; http://www.webgestalt.org/; Version 6.8) was used to explore the potential function of the POLD-related gene panel (20).

Abundance and Frequency of POLD Mutations in HCC

To further investigate the role of POLD in HCC and pancancers, we analyzed the abundance and frequency of POLD mutations in HCC using cBioportal for cancer genomics (http:// www.cbioportal.org/). The frequency of typical gene mutations in HCC according to differential POLD1 expression was also analyzed. Significantly elevated genes between POLD1 altered and unaltered groups were screened and identified using the Limma R package.

Analysis of Immune Infiltration in the Tumor Microenvironment

We grouped the HCC patients (n=107, AHYMUN cohort) into POLD1^{high} and POLD1^{low} groups using the median POLD expression. Tumor Immune Estimation Resource 2.0 (TIMER 2.0, http://timer.cistrome.org/) was used to analyze the correlation between the abundance of tumor-infiltrating immune cells and POLD expression using Spearman's test. Then, R software was used to evaluate the interactions between immune checkpoint molecules and tumor-infiltrating lymphocytes (TILs) in human cancer samples from TCGA datasets. Spearman's test was also used to determine the relationship between POLD1 expression and tumor purity using the ESTIMATE algorithm (21).

Statistics Analysis

To determine the statistical significance of differential POLD expression between tumor and normal tissues, a Student's t-test was performed. Kaplan-Meier curves with their 95% confidence intervals (95%CIs) and log-rank tests were applied to evaluate the significance of disease-specific survival (DSS) and overall survival (OS) benefits in separate POLD expression groups and all subgroups classified by tumor microenvironment infiltration characteristics using the Kaplan-Meier Plotter (http://kmplot. com/analysis/index). Univariate and multivariate Cox regression analyses were performed to identify the proper terms to build the nomogram. A forest plot was generated with the "forestplot" R package and used to show the P-value, hazard ratio (HR) and 95% CI of each variable. A nomogram was developed based on the results of multivariate Cox proportional hazard analysis to predict the X-year overall recurrence and calculate the risk of recurrence for individual patients. Moreover, a Sankey diagram was used to explore the relationship between pathological factors and patient survival.

All statistical analyses were performed using SPSS software (version 23.0, Inc, Chicago, IL), GraphPad Prism 8.0, R software (version 3.4.3), or online web tools. All hypothetical analyses were two-sided, and P<0.05 was considered statistically significant.

RESULTS

Identification of Critical DDR Pathway Genes and Their Prognostic Implications in HCC

This study was conducted in three stages (Figure S1). To determine the most significant genes in the DDR pathway, we first screened genes involved in base excision repair, nucleotide excision repair, mismatch repair, homologous recombination, non-homologous end-joining and Fanconi anemia pathways. We arranged these genes according to the number of pathways they are involved in and found that the POLD family participated in four pathways, indicating the important value of POLDs in DDR (Figure 1A). Next, the differential expression in POLD family members between >12,000 tumor and normal tissues was assessed. The expression level of POLDs was significantly higher in HCC tumor tissues compared with normal tissues (Figure 1B). Furthermore, POLD1 had the highest Z-score in the heatmap compared with POLD2, POLD3 and POLD4, indicating its greater significance for HCC prognosis (Figure 1C). Based on these results, we mainly explored the important role of POLDs in DDR and the effect of POLD expression on the development and prognosis of HCC.

Prognostic Role of POLDs in HCC

To further confirm the research direction, we performed Kaplan-Meier analysis of 364 patients from the TCGA cohort.





High POLD1 expression remarkably predicted a worse OS of HCC patients, whereas patients with higher expression of POLD2, POLD3 and POLD4 did not exhibit significantly shorter OS compared with lower expression patients (Figures 2A-D). Then, we conducted subgroup survival analyses of POLD1 expression based on hepatitis virus infection, clinicopathological staging and sorafenib use in 364 patients with HCC (Figures 2E-H). The results suggested that elevated POLD1 expression significantly predicted poor OS in the hepatitis virus infected and uninfected subgroup. However, for HCC patients with an advanced stage, POLD1 expression was not significantly associated with prognosis (HR=1.71, P=0.13) but predicted significantly worse outcomes for those receiving sorafenib treatment (HR=4.13, P=0.026). Additionally, high POLD1 expression was remarkably related to poor PFS in 370 HCC patients (Figure 2I).

POLD1 Significantly Predicts Survival and Aggressive Clinicopathological Parameters for HCC Patients

Next, we explored the relationship between the expression of POLD1 and the survival of HCC patients. HCC patients with higher POLD1 expression experienced a significantly increased risk of death, and the z-score of POLD1 expression confirmed that elevated POLD1 levels were associated with higher mortality (**Figure 3A**). The Kaplan–Meier curve also demonstrated that high POLD1 expression led to worse OS in 371 patients, with a median survival of 2.8 years in the POLD^{high} group and 6.3 years in the POLD^{low} group (**Figure 3B**). The high sensitivity and specificity of the independent diagnostic and prognostic value of POLD1 expression were shown by the ROC curve (**Figure 3C**). However, the accuracy decreased over time [1-year area under the curve (AUC)=0.742, 3-year AUC=0.666 and 5-year





AUC=0.610], suggesting that POLD1 expression more accurately predicted the prognosis of early-stage HCC patients compared with those with advanced stages.

Next, we assessed the Gumz Liver cohort consisting of differential RNA-seq data from HCC and normal samples. The results revealed a significantly elevated POLD1 expression level in HCC compared with normal samples (n=22; P=0.043; **Figure 4A**). The Sankey diagram shows the relationships between tumor grades, stages, POLD1 expression levels and survival. Low-tumor grade and early-stage HCC patients tended to have reduced expression of POLD1, whereas high tumor grades and advanced stages were related to the increased POLD1 expression (**Figure 4B**).

Differential Expression of POLD1 in Pan-Cancers and Its Prognostic Value

Based on HCC tissues and adjacent normal tissues from TCGA (n=423) and GTEx (n=533) datasets, we first compared the

POLD1 expression level between pan-cancers and normal tissues. Significant differential expression of POLD1 was commonly observed between cancer tissues and adjacent normal tissues in pan-cancers, such as GMB and STAD, and the expression of POLD1 was significantly higher in the HCC tumor group compared with the normal group (**Figures S2A, B**). Second, to explore the prognostic implications of POLD1 expression in pan-cancers, we performed Cox regression analysis (**Figures S2C, D**). POLD1 expression was closely correlated with OS in several cancers, such as PRAD (HR=9.25; P=2.7e-04) and MESO (HR=2.71; P=1.7e-06), and DSS in KIRC (HR=1.93; P=7.4e-04) and ACC (HR=3.48; P=2.6e-06).

Construction of a Prediction Model Using Cox Regression and Nomogram Analysis

The univariate Cox regression analysis suggested that POLD1, POLD2, POLD3 and pTNM stages were closely related to the











pTNM stage were closely related to the survival of HCC patients (P < 0.05). (**B**) Multivariate Cox regression revealed a significant effect of POLD1 expression (HR = 1.63, P = 0.011) and pTNM stage (HR = 1.61, P < 0.01) on the prognosis of HCC. (**C**) We used a nomogram to evaluate the prognosis of HCC with a prediction model of POLD1 expression and pTNM stage. (**D**) a graphical representation of the factors was provided by nomogram to calculate the risk of recurrence for an individual patient. The prediction is more accurate in short-term survival prediction (C-index = 0.682, P < 0.001).

survival of HCC patients (P<0.05). Because of the interaction between these factors, multivariate Cox regression revealed a significant effect of POLD1 expression (HR=1.63, P=0.011) and pTNM stage (HR=1.61, P<0.01) on the prognosis of HCC (**Figures 5A, B**). Together, our findings have demonstrated a consistent relationship between high POLD1 expression and advanced clinicopathological staging. Thus, we included POLD1 expression and pTNM stages in a prediction model to evaluate the prognosis of HCC using a nomogram (**Figure 5C**). Finally, the nomogram provided a graphical representation of the factors, which was used to calculate the risk of recurrence for an individual patient based on the points associated with each risk factor, and the model was more accurate in short-term survival prediction (C-index=0.682, P<0.001; **Figure 5D**).

Univariate Cox regression analysis indicated that POLD1, pTNM staging and grade were closely related to the survival of HCC patients (P<0.05), and multivariate Cox regression revealed the significant effects of POLD1 expression (HR=1.54, P=0.0005) and pTNM staging (HR=1.27, P=0.016) on the prognosis of HCC (**Figures S3A, B**). These findings further confirm the relationship between high POLD1 expression and advanced clinicopathological staging in HCC prognosis. Therefore, we included POLD1 expression and pTNM stage in a nomogram to assess the prognosis of HCC

patients (Figure S3C). The model is more accurate in short-term survival prediction (Figure S3D).

Landscape of POLD1 Mutation and Related Genes in HCC

In addition, to investigate the underlying role of POLD1, we explored the potential value of POLD1 mutation and other related genes based on multi-omics data. First, we found that the mutation was mainly due to POLD1 missense mutation on POLBc_delta and zf-C4pol (Figure 6A). However, the mutation frequency was only 1.1% in the TCGA cohort. Second, we analyzed the frequency of mutations in common genes in HCC according to differential POLD1 expression in 272 cases, including TP53, TTN, CTNNB1, MUC16, ALB, PCLO, RYR2, MUC4, ABCA13, APOB and POLD1. The highest mutation frequency was found in TP53 (28%). Interestingly, the POLD1^{high} group exhibited significantly more TP53 mutations compared with the POLD1^{low} group, suggesting the importance of the TP53 expression level in clinical treatment and tumor prognosis (Figure 6B). Then, we explored significant DEGs between the POLD1 altered and unaltered groups and found that DNAH5, TTN, TP53, NTAN1, HDAC5, TMEM51, KIAA1211, LMBR1, MCF2L and OR5L1 were markedly upregulated, whereas TP53 expression was significantly



decreased in the altered group compared with the unaltered group (Figures 6C, D).

PPI Network Establishment and Functional Enrichment Analysis

To explore the mechanism of POLD1 in the DDR pathway, we constructed a PPI network and identified a gene panel of critical genes, including POLD1, POLD2, POLD3, POLD4, PCNA, MSH2, MSH6, RPA1, RPA3 and LIG1 (Figure 7A). Additionally, we studied the correlation between these genes. Except for POLD4, the other ten hub genes showed a close linear association with each other (Spearman's test; Figure 7B). To analyze the function of these genes, we conducted pathway enrichment analyses, including GO (BP, CC and MF), KEGG and Reactome (Figures 7C, D). We identified several critical factors, such as mismatch repair and DNA replication in BP, replication fork and a protein–DNA complex in CC, damaged DNA binding and nucleotidyltransferase activity in MF, nucleotide excision repair and base excision repair in KEGG and extension of telomeres in Reactome.

POLD1 Expression Predicts the Abundance of Immune Cells and Immune Checkpoint Molecules in the HCC Microenvironment

High expression levels of POLD1 were found to be closely associated with tumor purity, immune scores and stromal scores in pan-cancers (Figure 8A). In addition, several immune cells, including B cells, T cells, dendritic cells,

macrophages and neutrophils, were significantly associated with POLD1 expression to varying degrees in different cancers, especially HCC (r²>0.4, Figure 8B). Moreover, POLD1 expression was related to the abundance of neutrophils and CD56 natural killer cells in HCC (Figure 8C). Meanwhile, POLD1 levels exhibited a strong relationship with most immune checkpoint molecules, including CD274, CD80, CD86, CTLA4, PDCD1 and TCGIT (Figure 8D). The scatter plot shown in Figure 8 demonstrates the close relationship between POLD1 expression levels and tumor purity (HR=0.141), B cells (HR=0.468), CD8⁺ T cells (HR=0.277), CD4⁺ T cells (HR=0.358), macrophages (HR=0.397), neutrophils (HR=0.364) and dendritic cells (HR=0.438). Overall, these findings suggested that POLD1 expression was correlated with the infiltration of several immune cells and reshaped the immune-excluded microenvironment.

Validation of Differential POLD1 Expression and Its Prognostic Value in the AHYMUN Cohort

To validate the increased expression of POLD1 in HCC samples compared with normal liver tissues, we first collected samples from HPA and explored the prognostic implications of POLD1 expression in 107 HCC patients from the AHYMUN cohort. The nuclear expression of POLD1 was significantly higher in HCC than adjacent normal tissues based on the HPA database and AHYMUN cohorts (**Figures 9A, B**). Additionally, the results suggested that increased protein expression of POLD1 was closely associated with worse OS (P=0.018, HR=1.697) and PFS (P=0.042, HR=1.669; **Figures 9C, D**).



DISCUSSION

HCC is the most common primary liver cancer and has a poor prognosis. Over the past few decades, the incidences of liver cancer and liver cancer-related deaths have increased (4, 22). However, the treatment options for advanced liver cancer are very limited, and strategies for advanced personalized treatment of HCC are lacking (23). Therefore, better understanding of the mechanisms underlying HCC is critical because these findings may help identify novel treatments for HCC patients. In this study, we examined the potential prognostic value of POLD in HCC and evaluated its role in the tumor immune microenvironment.

POLD1 interacts with POLD2, POLD3, and POLD4 to form the POL δ holoenzyme together with replication factor C and proliferating cell nuclear antigen. POLD2–4 are smaller subunits, and POLD1 plays a major role in the biochemical activity of the polymerase (24). Consistent with these data, we found that POLD1 showed a more significant influence on tumor prognosis than the other three POLD proteins and therefore we focused on POLD1 for the subsequent analysis.

POL δ has important proofreading capabilities conferred by exonuclease activity and is also involved in repairing DNA damage, including nucleotide excision repair, double-strand break repair, base excision repair, and mismatch repair (25, 26). Multiple studies have linked germline and sporadic mutations in POLD1 and other subunits of POL δ with human pathologies (27, 28). For example, mutations in POL δ in mice and humans lead to genome instability, mutant phenotypes, and tumorigenesis. In addition, mutations in the proofreading domain of POLD1 have been identified as the root cause of some hereditary cancers and these mutations may affect treatment management. Recent studies have shown that lossof-function alterations in DDR genes are associated with human tumorigenesis (29). Germline mutations in POLE and POLD1 have been shown to predispose patients to multiple colorectal polyps and a wide range of neoplasms (30). A previous study showed that at least 1 in 92 primary liver cancer patients had DDR gene mutation (31). The landscape of DDR mutations and their association with genetic and clinicopathologic features suggest that PLC patients with altered DDR genes may be rational candidates for precision treatment (32). Therefore, in addition to the increased expression level of POLD1, POLD1 and POLE mutations may function as independent biomarkers to predict the benefit of targeted treatment.

The expression levels of POLD family members in tumor and normal tissues were explored. According to previous research, the increased expression of POLD3 indicated a poor prognosis of HCC patients, although not as significantly as POLD1 (33).



FIGURE 8 | Correlation between POLD1 expression and immune cells. (A) High expression of POLD1 was closely associated with tumor purity, immune score and stromal score in pan-cancers. (B) Immune cells, including B cells, T cells, dendritic cells, macrophages and neutrophils, were significantly associated with POLD1 expression in many cancers in varying degrees, especially in HCC. (C–D) POLD1 was related to the abundance of neutrophils and CD56 natural killer cells in HCC, and POLD1 expression showed strong relationships with most immune checkpoint molecules, including CD274, CD80, CD86, CTLA4, PDCD1 and TCGIT. *p < 0.05, **p < 0.01, ***p < 0.001. (E) A scatter plot shows the close relationship between POLD1 expression level and tumor purity (HR=0.141), B cells (HR=0.468), CD8⁺ T cells (HR=0.277), CD4⁺ T cells (HR=0.358), macrophages (HR=0.397), neutrophils (HR=0.364) and dendritic cells (HR=0.438).

POLD3 plays a specialized role in the repair of damaged replication forks, indicating that POLD3 activity may be particularly relevant for cancer cells enduring high levels of DNA replication stress (12). The cellular depletion of POLD1 or POLD3 resulted in differential genome instability manifested by DNA double-stranded breaks (17, 34).

Human cancers can be divided into three types according to the anti-tumor immune response status, or the immune phenotype: inflamed, immune-excluded, and immune desert status. Inflamed cancers generally refer to tumors with high PD-L1 expression in cancer cells and more immune cells and tumor infiltrating lymphocytes in the tumor; these tumors are sensitive to immune checkpoint inhibitors. Immune-excluded tumors are tumors in which the stroma shows a large number of T cells, but the T cells cannot penetrate the stroma and infiltrate the tumor because of the strong inhibitory microenvironment. These tumors often do not respond well to immune checkpoint inhibitors. Immune desert tumors lack the infiltration of T cells and immune cells even in the interstitium, which is a described as an "immune desert". In our analysis, we found that POLD1 is closely related to the expression of B cells, CD8+ T cells, CD4+ T cells and macrophages. These results suggest that POLD1 may play an important role in the immune-excluded tumor microenvironment. We speculate that this may be because POLD1 is involved in DNA cleavage repair, which affects the tumor microenvironment.

There are some advantages of this study. First, this research contained independent HCC cohorts, including the TCGA database (n=423), GTEx database and Gumz Renal cohort (n=10), and the real-world AHYMUN cohort (n=107). Second, we demonstrated the value of significantly elevated POLD expression for HCC prognosis and identified POLD1 as the most valuable gene for further analysis. Third, at both the mRNA and protein levels, we validated the association between POLD1 expression and HCC prognosis. Moreover, ~12,000 tumor samples from TCGA database were collected, and the effect of POLD1 expression on prognosis was verified in pan-cancers. Finally, functional enrichment analysis was performed. The role



of POLD1 in the infiltration of immune cells in the tumor microenvironment was demonstrated, which may guide cancer treatment and targeted drug development.

This study has several limitations. First, there is a high degree of heterogeneity between the patient groups in this study. Therefore, in the next study, we will select patients from multiple regions for multi-center research. Second, this was a retrospective study. Finally, we will further study the demographic, clinical and pathological details of the population.

Our study has demonstrated a link between elevated POLD1 expression and patient survival and the tumor microenvironment in HCC. In the next study, we will explore the role and mechanism of POLD in HCC progression through cytological tests and animal experiments. We will also analyze the potential mechanisms of POLD, POL&, and DNA cleavage repair in HCC through high-throughput sequencing and other methods. These findings will help provide new insights into the pathogenesis of HCC and new ideas for the personalized treatment of HCC patients.

CONCLUSION

This study first investigated the molecular and clinical role of the POLD family and revealed the significant relationship between elevated POLD1 expression and the poor survival and immune-excluded tumor microenvironment of HCC patients. Together, these findings support the use of POLD1 as a biomarker to guide

the molecular diagnosis and development of novel targeted therapeutic strategies for HCC patients.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**. Further inquiries can be directed to the corresponding authors.

ETHICS STATEMENT

The study design and experimental procedures were performed in accordance with the Declaration of Helsinki. The study was approved by the ethics committee of the Affiliated Hospital of Youjiang Medical College for Nationalities (Baise, Guangxi Province, China). Written informed consent was obtained from all participants.

AUTHOR CONTRIBUTIONS

WL and SZ carried out the molecular genetic studies, participated in the sequence alignment and drafted the

manuscript. JW and SC carried out the immunoassays. HD, CW and BH participated in the sequence alignment. XS and QG participated in the design of the study and performed the statistical analyses. HT, SZ, YZ, WL, and JW conceived the study, participated in the study design and coordination and helped to draft the manuscript. All authors contributed to the article and approved the submitted version.

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REFERENCES

- Cadoux M, Caruso S, Pham S, Gougelet A, Pophillat C, Riou R, et al. Expression of NKG2D Ligands is Downregulated by Beta-Catenin Signaling and Associates With HCC Aggressiveness. J Hepatol (2021). doi: 10.1016/ j.jhep.2021.01.017
- Luo P, Wu S, Yu Y, Ming X, Li S, Zuo X, et al. Current Status and Perspective Biomarkers in AFP Negative HCC: Towards Screening for and Diagnosing Hepatocellular Carcinoma at an Earlier Stage. *Pathol Oncol Res* (2020) 26 (2):599–603. doi: 10.1007/s12253-019-00585-5
- Campani C, Bensi C, Milani S, Galli A, Tarocchi M. Resection of NAFLD-Associated HCC: Patient Selection and Reported Outcomes. J Hepatocell Carcinoma (2020) 7:107–16. doi: 10.2147/JHC.S252506
- 4. Couri T, Pillai A. Goals and Targets for Personalized Therapy for HCC. *Hepatol Int* (2019) 13(2):125–37. doi: 10.1007/s12072-018-9919-1
- Gao X, Zhan M, Wang L, Ding Y, Niu J. Timing of DAA Initiation After Curative Treatment and Its Relationship With the Recurrence of HCV-Related HCC. J Hepatocell Carcinoma (2020) 7:347–60. doi: 10.2147/ JHC.S279657
- Pinto Marques H, Gomes da Silva S, De Martin E, Agopian VG, Martins PN. Emerging Biomarkers in HCC Patients: Current Status. *Int J Surg* (2020) 82S:70–6. doi: 10.1016/j.ijsu.2020.04.043
- Yang JD, Addissie BD, Mara KC, Harmsen WS, Dai J, Zhang N, et al. GALAD Score for Hepatocellular Carcinoma Detection in Comparison With Liver Ultrasound and Proposal of GALADUS Score. *Cancer Epidemiol Biomarkers Prev* (2019) 28(3):531–8. doi: 10.1158/1055-9965.EPI-18-0281
- Berhane S, Toyoda H, Tada T, Kumada T, Kagebayashi C, Satomura S, et al. Role of the GALAD and BALAD-2 Serologic Models in Diagnosis of Hepatocellular Carcinoma and Prediction of Survival in Patients. *Clin Gastroenterol Hepatol* (2016) 14(6):875–886 e6. doi: 10.1016/j.cgh.2015.12.042
- Ioannou GN. HCC Surveillance After SVR in Patients With F3/F4 Fibrosis. J Hepatol (2021) 74(2):458–65. doi: 10.1016/j.jhep.2020.10.016
- Ricciuti B, Recondo G, Spurr LF, Li YY, Lamberti G, Venkatraman D, et al. Impact of DNA Damage Response and Repair (DDR) Gene Mutations on Efficacy of PD-(L)1 Immune Checkpoint Inhibition in Non-Small Cell Lung Cancer. *Clin Cancer Res* (2020) 26(15):4135–42. doi: 10.1158/1078-0432.CCR-19-3529

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fonc.2021. 736363/full#supplementary-material

Supplementary Figure 1 | Flowchart of the study.

Supplementary Figure 2 | High expression of POLD1 in pan-cancers compared with normal liver tissues. (A, B) POLD1 expression levels between pan-cancers and normal tissues were compared in the TCGA (n = 423) and GTEx (n = 533) datasets. POLD1 expression was differentially expressed in cancer tissue and adjacent normal tissue in pan-cancer, and the expression of POLD1 was significantly higher in the HCC tumor group compared with the normal group. (C, D) The POLD expression level in pan-cancers and normal tissue based on data from the TCGA and GTEx databases. The results are unremarkable in some cancers. (C, D) Cox regression analysis showed that POLD1 expression is closely correlated with OS in many cancers.

Supplementary Figure 3 | Prediction model based on Cox regression and nomogram analysis of POLD1. (A) Univariate Cox regression analysis suggested that POLD1, age, grade, race and pTNM stage were closely related to the survival of HCC patients (*P* < 0.05). (B) Multivariate Cox regression revealed the significant effect of POLD1 expression and pTNM stage on the prognosis of HCC. (C) We used a nomogram to evaluate the prognosis of HCC with a prediction model of POLD1 expression and pTNM stage. (D) A graphical representation of the factors was provided by nomogram to calculate the risk of recurrence for an individual patient. The prediction is more accurate in short-term survival prediction.

- Brandsma I, Fleuren EDG, Williamson CT, Lord CJ. Directing the Use of DDR Kinase Inhibitors in Cancer Treatment. *Expert Opin Investig Drugs* (2017) 26(12):1341–55. doi: 10.1080/13543784.2017.1389895
- Murga M, Lecona E, Kamileri I, Diaz M, Lugli N, Sotiriou SK, et al. POLD3 Is Haploinsufficient for DNA Replication in Mice. *Mol Cell* (2016) 63(5):877–83. doi: 10.1016/j.molcel.2016.07.007
- Terai K, Shibata E, Abbas T, Dutta A. Degradation of P12 Subunit by CRL4Cdt2 E3 Ligase Inhibits Fork Progression After DNA Damage. J Biol Chem (2013) 288(42):30509–14. doi: 10.1074/jbc.C113.505586
- Meng X, Zhou Y, Lee EY, Lee MY, Frick DN. The P12 Subunit of Human Polymerase Delta Modulates the Rate and Fidelity of DNA Synthesis. *Biochemistry* (2010) 49(17):3545–54. doi: 10.1021/bi100042b
- Costantino L, Sotiriou SK, Rantala JK, Magin S, Mladenov E, Helleday T, et al. Break-Induced Replication Repair of Damaged Forks Induces Genomic Duplications in Human Cells. *Science* (2014) 343(6166):88–91. doi: 10.1126/ science.1243211
- Zhou Z, Wang L, Ge F, Gong P, Wang H, Wang F, et al. Pold3 is Required for Genomic Stability and Telomere Integrity in Embryonic Stem Cells and Meiosis. *Nucleic Acids Res* (2018) 46(7):3468–86. doi: 10.1093/nar/ gky098
- Tumini E, Barroso S, Calero CP, Aguilera A. Roles of Human POLD1 and POLD3 in Genome Stability. Sci Rep (2016) 6:38873. doi: 10.1038/srep38873
- Ito T, Nomizu T, Eguchi H, Kamae N, Dechamethakun S, Akama Y, et al. The First Case Report of Polymerase Proofreading-Associated Polyposis in POLD1 Variant, C.1433G>A P.S478N, in Japan. *Jpn J Clin Oncol* (2020) 50 (9):1080–3. doi: 10.1093/jjco/hyaa090
- Xu WH, Xu Y, Tian X, Anwaier A, Liu WR, Wang J, et al. Large-Scale Transcriptome Profiles Reveal Robust 20-Signatures Metabolic Prediction Models and Novel Role of G6PC in Clear Cell Renal Cell Carcinoma. J Cell Mol Med (2020) 24(16):9012–27. doi: 10.1111/jcmm.15536
- Liao Y, Wang J, Jaehnig EJ, Shi Z, Zhang B. WebGestalt 2019: Gene Set Analysis Toolkit With Revamped UIs and APIs. Nucleic Acids Res (2019) 47 (W1):W199–205. doi: 10.1093/nar/gkz401
- Xu WH, Xu Y, Wang J, Wan FN, Wang HK, Cao DL, et al. Prognostic Value and Immune Infiltration of Novel Signatures in Clear Cell Renal Cell Carcinoma Microenvironment. *Aging (Albany NY)* (2019) 11(17):6999– 7020. doi: 10.18632/aging.102233

- Kim DW, Talati C, Kim R. Hepatocellular Carcinoma (HCC): Beyond Sorafenib-Chemotherapy. J Gastrointest Oncol (2017) 8(2):256–65. doi: 10.21037/jgo.2016.09.07
- Margini C, Dufour JF. The Story of HCC in NAFLD: From Epidemiology, Across Pathogenesis, to Prevention and Treatment. *Liver Int* (2016) 36 (3):317–24. doi: 10.1111/liv.13031
- Nicolas E, Golemis EA, Arora S. POLD1: Central Mediator of DNA Replication and Repair, and Implication in Cancer and Other Pathologies. *Gene* (2016) 590(1):128–41. doi: 10.1016/j.gene.2016.06.031
- Darmawan H, Harrison M, Reha-Krantz LJ. DNA Polymerase 3'-5' Exonuclease Activity: Different Roles of the Beta Hairpin Structure in Family-B DNA Polymerases. DNA Repair (Amst) (2015) 29:36-46. doi: 10.1016/j.dnarep.2015.02.014
- Fortune JM, Pavlov YI, Welch CM, Johansson E, Burgers PM, Kunkel TA. Saccharomyces Cerevisiae DNA Polymerase Delta: High Fidelity for Base Substitutions But Lower Fidelity for Single- and Multi-Base Deletions. J Biol Chem (2005) 280(33):29980–7. doi: 10.1074/jbc.M505236200
- Albertson TM, Ogawa M, Bugni JM, Hays LE, Chen Y, Wang Y, et al. DNA Polymerase Epsilon and Delta Proofreading Suppress Discrete Mutator and Cancer Phenotypes in Mice. *Proc Natl Acad Sci USA* (2009) 106(40):17101–4. doi: 10.1073/pnas.0907147106
- Antoniali G, Marcuzzi F, Casarano E, Tell G. Cadmium Treatment Suppresses DNA Polymerase δ Catalytic Subunit Gene Expression by Acting on the P53 and Sp1 Regulatory Axis. DNA Repair (Amst) (2015) 35:90–105. doi: 10.1016/ j.dnarep.2015.08.007
- Ged Y, Chaim JL, DiNatale RG, Knezevic A, Kotecha RR, Carlo MI, et al. DNA Damage Repair Pathway Alterations in Metastatic Clear Cell Renal Cell Carcinoma and Implications on Systemic Therapy. J Immunother Cancer (2020) 8(1). doi: 10.1136/jitc-2019-000230
- Esteban-Jurado C, Gimenez-Zaragoza D, Munoz J, Franch-Exposito S, Alvarez-Barona M, Ocana T, et al. POLE and POLD1 Screening in 155 Patients With Multiple Polyps and Early-Onset Colorectal Cancer. Oncotarget (2017) 8(16):26732–43. doi: 10.18632/oncotarget.15810

- van der Doelen MJ, Isaacsson Velho P, Slootbeek PHJ, Pamidimarri Naga S, Bormann M, van Helvert S, et al. Impact of DNA Damage Repair Defects on Response to Radium-223 and Overall Survival in Metastatic Castration-Resistant Prostate Cancer. *Eur J Cancer* (2020) 136:16–24. doi: 10.1016/ j.ejca.2020.05.001
- Lin J, Shi J, Guo H, Yang X, Jiang Y, Long J, et al. Alterations in DNA Damage Repair Genes in Primary Liver Cancer. *Clin Cancer Res* (2019) 25(15):4701– 11. doi: 10.1158/1078-0432.CCR-19-0127
- Mayle R, Campbell IM, Beck CR, Yu Y, Wilson M, Shaw CA, et al. DNA REPAIR. Mus81 and Converging Forks Limit the Mutagenicity of Replication Fork Breakage. *Science* (2015) 349(6249):742–7. doi: 10.1126/science.aaa8391
- Dunlop MG, Dobbins SE, Farrington SM, Jones AM, Palles C, Whiffin N, et al. Common Variation Near CDKN1A, POLD3 and SHROOM2 Influences Colorectal Cancer Risk. Nat Genet (2012) 44(7):770–6. doi: 10.1038/ng.2293

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