





Complete Genome Sequence of *Stenotrophomonas maltophilia* Siphophage Silvanus

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ABSTRACT *Stenotrophomonas maltophilia* is an opportunistic Gram-negative bacterium capable of causing respiratory infections. *S. maltophilia* siphophage Silvanus was isolated, and its 45,678-bp genome is not closely related to known phages based on whole-genome comparative genomics analysis. It is predicted to use *cos*-type packaging due to the similarity of its large terminase subunit to that of phage HK97.

S tenotrophomonas maltophilia is an emerging Gram-negative, multidrug-resistant pathogen most associated with respiratory infections in humans (1). With a goal of using phage as potential control for this pathogen, we report here the isolation and genome annotation of Silvanus, a siphophage targeting *S. maltophilia*.

Phage Silvanus was isolated from a soil sample collected from a horse pasture in College Station, TX (GPS coordinates 30°33′04.4"N, 96°18′44.0"W), in January 2019. Silvanus was isolated and propagated with the soft-agar overlay methods described previously (2) using an S. maltophilia strain (ATCC 51331) grown aerobically at 30°C in nutrient broth or agar (BD). Samples were negatively stained with 2% (wt/vol) uranyl acetate and imaged by transmission electron microscopy (TEM) at the Texas A&M Microscopy and Imaging Center (3). DNA was purified using a Promega Wizard DNA cleanup system as described (4), and the libraries were prepared using a Swift 2S Turbo library preparation kit and sequenced on an Illumina MiSeq machine with paired-end 150-bp reads and V2 300-cycle chemistry. The sequence reads were quality controlled with FastQC (www.bioinformatics.babraham.ac.uk/projects/ fastqc) and trimmed with FASTX-Toolkit v0.11.6 (http://hannonlab.cshl.edu/fastx_toolkit/). Genomes were assembled from 85,453 trimmed reads with SPAdes v3.5.0 (5), and a contig with 138-fold sequencing coverage was obtained. The genome was closed by PCR and Sanger sequencing using forward primer 5'-CATCGTGTGGGCGAAATC-3' and reverse primer 5'-TGAACCCCTGAGTTTCGTGG-3'. PhageTerm was used to predict phage termini from raw sequencing reads (6). The genome was assembled and annotated with the CPT Galaxy-Apollo phage annotation platform (https://cpt.tamu.edu/galaxy-pub) (7-9). Gene calling was conducted with GLIMMER v3 and MetaGeneAnnotator v1.0 (10, 11). tRNAs were detected with ARAGORN v2.36 and tRNAscan-SE v2.0 (12, 13). Gene function predictions were determined using InterProScan v5.48 (14) and BLAST v2.9.0 (15) against the NCBI nonredundant (nr) and Swiss-Prot databases (16), TMHMM v2.0 (17), HHPred, LipoP v1.0, and SignalP v5.0 (18-20). The genome-wide DNA sequence similarity to the top BLAST nucleotide hits was calculated with progressiveMauve v2.4 (21). All analyses were conducted at default settings.

Phage Silvanus has a siphophage morphology (Fig. 1). The 45,678-bp genome has a coding density of 97.4% and a G+C content of 58.4%. No tRNA genes were identified, and 26 out of 68 total genes were assigned putative functions, including a complete lysis cassette with genes encoding an endolysin of the glycosyl hydrolase class, a holin with three transmembrane domains and N-out, C-in topology (class I), and two-component spanins. Silvanus is predicted to use $\it cos\mbox{-}type$ packaging because it encodes a

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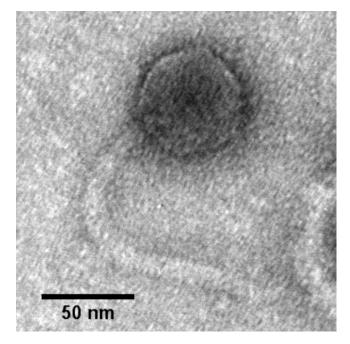


FIG 1 Transmission electron micrograph (TEM) of phage Silvanus. Phage particles were diluted with TEM buffer (20 mM NaCl, 10 mM Tris-HCl, pH 7.5, and 2 mM MgSO4) and captured on a freshly glow-discharged, Formvar carbon-coated grid. The grids were stained with 2% (wt/vol) uranyl acetate and observed on a JEOL 1200 EX TEM at 100 kV accelerating voltage at the Microscopy and Imaging Center at Texas A&M University.

large terminase subunit similar to that of the well-characterized cos phage HK97 (21% protein identity; E value, 10^{-8} ; 100% HHpred probability) and also encodes an HNH endonuclease similar to that of HK97 gp74 (40% protein identity; E value, 10^{-21}) at the opposite end of the genome, which is required for the 3′ cos cleavage (22). Moreover, according to HHPred, the predicted small terminase has a 99.5% probability match to the structure of the Pseudomonas phage PaP3 small terminase, which generates cohesive ends (23). The precise location of phage Silvanus cos sites, however, cannot be determined by PhageTerm analysis. Whole-genome comparative genomics analysis by progressiveMauve v2.4 (21) revealed that Silvanus has <7% overall nucleotide identity to known phages. Silvanus was found to carry a T1 p38-like tail tape measure protein.

Data availability. The Silvanus genome was deposited in GenBank with accession number MZ326867. The associated BioProject, SRA, and BioSample accession numbers are PRJNA222858, SRR14095258, and SAMN18509682, respectively.

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REFERENCES

- Brooke JS. 2012. Stenotrophomonas maltophilia: an emerging global opportunistic pathogen. Clin Microbiol Rev 25:2–41. https://doi.org/10 .1128/CMR.00019-11.
- 2. Adams MK. 1959. Bacteriophages. Interscience Publishers, Inc., New York, NY.
- Valentine RC, Shapiro BM, Stadtman ER. 1968. Regulation of glutamine synthetase. XII. Electron microscopy of the enzyme from *Escherichia coli*. Biochemistry 7:2143–2152. https://doi.org/10.1021/bi00846a017.
- 4. Summer EJ. 2009. Preparation of a phage DNA fragment library for whole

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- genome shotgun sequencing. Methods Mol Biol 502:27–46. https://doi.org/10.1007/978-1-60327-565-1_4.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Garneau JR, Depardieu F, Fortier LC, Bikard D, Monot M. 2017. PhageTerm: a tool for fast and accurate determination of phage termini and packaging mechanism using next-generation sequencing data. Sci Rep 7:8292. https://doi.org/10.1038/s41598-017-07910-5.
- 7. Ramsey J, Rasche H, Maughmer C, Criscione A, Mijalis E, Liu M, Hu JC, Young R, Gill JJ. 2020. Galaxy and Apollo as a biologist-friendly interface for high-quality cooperative phage genome annotation. PLoS Comput Biol 16:e1008214. https://doi.org/10.1371/journal.pcbi.1008214.
- Afgan E, Baker D, Batut B, van den Beek M, Bouvier D, Cech M, Chilton J, Clements D, Coraor N, Gruning BA, Guerler A, Hillman-Jackson J, Hiltemann S, Jalili V, Rasche H, Soranzo N, Goecks J, Taylor J, Nekrutenko A, Blankenberg D. 2018. The Galaxy platform for accessible, reproducible and collaborative biomedical analyses: 2018 update. Nucleic Acids Res 46: W537–W544. https://doi.org/10.1093/nar/gky379.
- 9. Dunn NA, Unni DR, Diesh C, Munoz-Torres M, Harris NL, Yao E, Rasche H, Holmes IH, Elsik CG, Lewis SE. 2019. Apollo: democratizing genome annotation. PLoS Comput Biol 15:e1006790. https://doi.org/10.1371/journal.pcbi.1006790.
- Delcher AL, Harmon D, Kasif S, White O, Salzberg SL. 1999. Improved microbial gene identification with GLIMMER. Nucleic Acids Res 27:4636–4641. https://doi.org/10.1093/nar/27.23.4636.
- Noguchi H, Taniguchi T, Itoh T. 2008. MetaGeneAnnotator: detecting species-specific patterns of ribosomal binding site for precise gene prediction in anonymous prokaryotic and phage genomes. DNA Res 15: 387–396. https://doi.org/10.1093/dnares/dsn027.
- Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. Nucleic Acids Res 32:11–16. https://doi.org/10.1093/nar/gkh152.
- Chan PP, Lowe TM. 2019. tRNAscan-SE: searching for tRNA genes in genomic sequences. Methods Mol Biol 1962:1–14. https://doi.org/10 .1007/978-1-4939-9173-0_1.

- Jones P, Binns D, Chang HY, Fraser M, Li W, McAnulla C, McWilliam H, Maslen J, Mitchell A, Nuka G, Pesseat S, Quinn AF, Sangrador-Vegas A, Scheremetjew M, Yong SY, Lopez R, Hunter S. 2014. InterProScan 5: genome-scale protein function classification. Bioinformatics 30:1236–1240. https://doi.org/10.1093/bioinformatics/btu031.
- Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. BMC Bioinformatics 10:421. https://doi.org/10.1186/1471-2105-10-421.
- The UniProt Consortium. 2018. UniProt: the universal protein knowledgebase. Nucleic Acids Res 46:2699. https://doi.org/10.1093/nar/gky092.
- Krogh A, Larsson B, von Heijne G, Sonnhammer EL. 2001. Predicting transmembrane protein topology with a hidden Markov model: application to complete genomes. J Mol Biol 305:567–580. https://doi.org/10.1006/jmbi .2000.4315.
- Zimmermann L, Stephens A, Nam SZ, Rau D, Kubler J, Lozajic M, Gabler F, Soding J, Lupas AN, Alva V. 2018. A completely reimplemented MPI Bioinformatics Toolkit with a new HHpred server at its core. J Mol Biol 430: 2237–2243. https://doi.org/10.1016/j.jmb.2017.12.007.
- Juncker AS, Willenbrock H, Von Heijne G, Brunak S, Nielsen H, Krogh A. 2003. Prediction of lipoprotein signal peptides in Gram-negative bacteria. Protein Sci 12:1652–1662. https://doi.org/10.1110/ps.0303703.
- Almagro Armenteros JJ, Tsirigos KD, Sonderby CK, Petersen TN, Winther O, Brunak S, von Heijne G, Nielsen H. 2019. SignalP 5.0 improves signal peptide predictions using deep neural networks. Nat Biotechnol 37: 420–423. https://doi.org/10.1038/s41587-019-0036-z.
- Darling AE, Mau B, Perna NT. 2010. progressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. PLoS One 5:e11147. https://doi.org/10.1371/journal.pone.0011147.
- 22. Weiditch SA, Bickers SC, Bona D, Maxwell KL, Kanelis V. 2020. HK97 gp74 possesses an α -helical insertion in the $\beta\beta\alpha$ fold that affects its metal binding, \cos site digestion, and in vivo activities. J Bacteriol 202:e00644-19. https://doi.org/10.1128/JB.00644-19.
- Niazi M, Florio TJ, Yang R, Lokareddy RK, Swanson NA, Gillilan RE, Cingolani G. 2020. Biophysical analysis of *Pseudomonas*-phage PaP3 small terminase suggests a mechanism for sequence-specific DNA-binding by lateral interdigitation. Nucleic Acids Res 48:11721–11736. https://doi.org/ 10.1093/nar/gkaa866.