### PLASTOME ANNOUNCEMENTS

OPEN ACCESS Check for updates

Taylor & Francis

Taylor & Francis Group

# Characterization of the complete chloroplast genome of *Clematoclethra scandens* subsp. *actinidioides* (Actinidiaceae)

Lei Zhang<sup>a,b</sup>, Ying Zhang<sup>a,b</sup>, Yun Jia<sup>a,b</sup>, Fangbing Ding<sup>a,b</sup>, Fengwei Wang<sup>a,b</sup>, Gang Yu<sup>a,b</sup> and Yongpeng Wu<sup>a,b</sup>

<sup>a</sup>Shaanxi Engineering Research Centre for Conservation and Utilization of Botanical Resources, Xi'an, China; <sup>b</sup>Xi'an Botanical Garden of Shaanxi Province (Institute of Botany of Shaanxi Province), Xi'an, China

#### ABSTRACT

*Clematoclethra scandens* subsp. *actinidioides* (Actinidiaceae) is an endemic medicinal species in China. Here, we first sequenced and characterized the complete chloroplast genome of *C. scandens* subsp. *actinidioides*. The chloroplast genome was 159,341 bp in length, containing a large single-copy of 88,351 bp and a small single-copy of 21,580 bp separated by a pair of identical inverted repeat regions of 24,705 bp each. A total of 131 genes were identified, including 84 protein-coding genes, 39 tRNA, and eight rRNA genes. The phylogenetic analysis of *C. scandens* subsp. *actinidioides* showed a relatively close relationship with *Clematoclethra scandens* subsp. *hemsleyi*.

ARTICLE HISTORY Received 16 May 2022

Accepted 2 August 2022

KEYWORDS Chloroplast genome; *Clematoclethra scandens* subsp. *actinidioides*; phylogenetics

Clematoclethra scandens subsp. actinidioides (Maximowicz) Y. C. Tang & Q. Y. Xiang 1890 is an endemic medicinal plant in the family Actinidiaceae, which is distributed in the temperate and subtropical regions in central and western China (Li et al. 2007; Yang et al. 2014). The roots of C. scandens subsp. actinidioides have long been used as an important traditional medicinal to treat chronic hepatitis, rheumatic arthritis, and hernia (Song et al. 2001). Previous studies have focused on the chemical composition, pollen morphology, and taxonomy for this species (Yang et al. 2014; Xiao et al. 2015). Due to its various flavonoids and triterpenoids, C. scandens subsp. actinidioides not only has a high medicinal value, but also has scientific research value as an endemic species (Xiao et al. 2015). Herein, we first sequenced and assembled the complete chloroplast genome of C. scandens subsp. actinidioides and analyzed its phylogenetic relationship.

The fresh leaves from a wild single tree of *C. scandens* subsp. *actinidioides* were collected from Feng River, Shaanxi Province (108°48'16.76"E, 33°50'22.77"N) and the voucher specimens were stored at Xi'an Botanical Herbarium under accession number XBH20200822 (http://www.xazwy.com/; Yongpeng Wu, Email: 43566351@qq.com). Total genomic DNA was extracted using CTAB method (Doyle and Doyle 1987) and sequenced with Illumina Hiseq 4000 platform. The chloroplast genome was de novo assembled using Novoplasty (Dierckxsens et al. 2019). The annotation was performed with the online annotation tool CPGAVAS2 (Shi et al. 2019). Phylogenetic analyses were carried out by maximum

likelihood (ML) using MEGA v7.0 (Kumar et al. 2016) with 1000 bootstrap replicates.

The chloroplast genome of *C. scandens* subsp. *actinidioides* was a typical quadripartite circular molecule with a length of 159,341 bp, including a large single-copy region (LSC) of 88,351 bp and a small single-copy region (SSC) of 21,580 bp, and two 24,705 bp inverted repeat regions (IRs). A total of 131 genes were annotated, containing 84 protein-coding genes, 39 tRNA genes, and eight rRNA genes. Unexpectedly, we observed the chloroplast genome lacks *clpP* gene, which is consistent with *C. scandens* subsp. *hemsleyi* chloroplast genome in the genus *Clematoclethra* (Wang et al. 2016). The overall GC content of *C. scandens* subsp. *actinidioides* plastid genome is 38.3%, while the corresponding values of LSC, SSC, and IR regions are 38.9%, 37.1%, and 37.5%, respectively.

To confirm the phylogenetic position of *C. scandens* subsp. *actinidioides*, 14 chloroplast genome sequences of Actinidiaceae, Lardizabalaceae, and Passifloraceae were aligned by MEGA v7.0 (Kumar et al. 2016). The result indicated that *C. scandens* subsp. *actinidioides* was found to be relatively closely related to *C. scandens* subsp. *hemsleyi* chloroplast compared to other species of *Actinidia* genera in Actinidiaceae (Figure 1). The chloroplast genome information reported in this study provided fundamental data for the bio-informatics and systematics of the Actinidiaceae.

CONTACT Yongpeng Wu 🐼 43566351@qq.com 🝙 Xi'an Botanical Garden of Shaanxi Province (Institute of Botany of Shaanxi Province), Xi'an, China © 2022 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

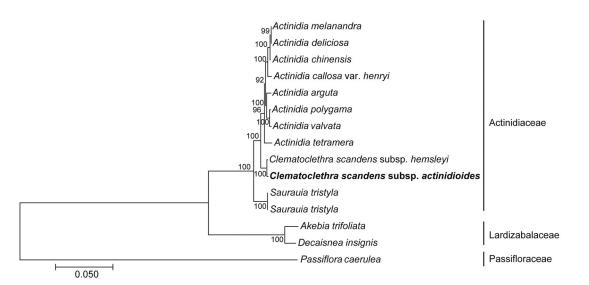


Figure 1. Phylogenetic tree produced by maximum-likelihood (ML) analysis based on 15 chloroplast genome sequences. The following sequences were used: Actinidia melanandra MK863365 (Zhao et al. 2019), Actinidia deliciosa NC026691, Actinidia chinensis NC026690, Actinidia callosa var. henryi NC043861 (Wu et al. 2019), Actinidia arguta NC034913, Actinidia polygama NC031186 (Wang et al. 2016), Actinidia valvata NC050357 (Wang et al. 2016), Actinidia tetramera NC031187 (Wang et al. 2016), Clematoclethra scandens subsp. hemsleyi KX345299 (Wang et al. 2016), Saurauia tristyla MG912839, Saurauia tristyla NC044098, Akebia trifoliata NC029427 (Sun et al. 2016), Decaisnea insignis NC035941 (Li et al. 2017), Passiflora caerulea MT884000, and Clematoclethra scandens subsp. actinidioides OL457297.

#### **Ethics statement**

Ethical approval for the study was obtained from the Ethical Committee of Xi'an Botanical Garden of Shaanxi Province.

## **Author contributions**

Y-PW and YZ conceived the study. LZ and YZ performed the experiments. Y-PW, YZ, LZ, YJ, F-BD, F-WW, and GY contributed materials and analysis tools. Y-PW, LZ and YJ wrote the manuscript. All authors approved the final version of the manuscript.

#### **Disclosure statement**

No potential conflict of interest was reported by the author(s).

#### Funding

This work was supported by the Xi'an Bureau of Science and Technology under Grant No. [20NYYF0003] and the Department of Science and Technology of Shaanxi Province [Nos. 2020NY-044, 2019TG-005].

## Data availability statement

The genome sequence data that support the findings of this study are openly available in GenBank of NCBI at https://www.ncbi.nlm.nih.gov/ under the accession no. OL457297. The associated BioProject, SRA, and Bio-Sample numbers are PRJNA778432, SRR16841622, and SAMN22959467, respectively.

## References

Dierckxsens N, Mardulyn P, Smits G. 2019. Unraveling heteroplasmy patterns with NOVOPlasty. NAR Genom Bioinform. 2:1–10. Doyle JJ, Doyle JL. 1987. A rapid DNA isolation procedure for small quantities of fresh leaf tissue. Phytochem Bull. 19:11–15.

- Kumar S, Stecher G, Tamura K. 2016. MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. Mol Biol Evol. 33(7): 1870–1874.
- Li JQ, Li XW, Soejarto DD. 2007. Actinidiaceae. In: Wu ZY, Raven PH, Hong DY, editors. Flora of China. Beijing, China; Missouri: Science Press; Missouri Botanical Garden; p. 334–360.
- Li B, Lin FR, Huang P, Guo WY, Zheng YQ. 2017. Complete chloroplast genome sequence of *Decaisnea insignis*: genome organization, genomic resources and comparative analysis. Sci Rep. 7(1):10073.
- Shi LC, Chen HM, Jiang M, Wang LQ, Wu X, Huang LF, Liu C. 2019. CPGAVAS2, an integrated plastome sequence annotator and analyzer. Nucleic Acids Res. 47(W1):W65–W73.
- Song LR, Hong X, Ding XL. 2001. Dictionary of modern Chinese herbal medicine. Beijing, China: Peoples Medical Publishing House; p. 2453.
- Sun Y, Moore MJ, Zhang S, Soltis PS, Soltis DE, Zhao T, Meng A, Li X, Li J, Wang H. 2016. Phylogenomic and structural analyses of 18 complete plastomes across nearly all families of early-diverging eudicots, including an angiosperm-wide analysis of IR gene content evolution. Mol Phylogenet Evol. 96:93–101.
- Wang WC, Chen SY, Zhang XZ. 2016. Chloroplast genome evolution in Actinidiaceae: clpP Loss, heterogenous divergence and phylogenomic practice. PLOS One. 11(9):e0162324.
- Wu HL, Li MM, Wang DY, Liu HH, Xu XT. 2019. The complete chloroplast genome sequence of *Actinidia callosa* var. *henryi*. Mitochondrial DNA B Resour. 4(1):652–653.
- Xiao SJ, Chen F, Ding LS, Zhou Y. 2015. A new ursane-type triterpenoid saponin from the aerial parts of *Clematoclethra scandens* subsp. actinidioides. Chin J Nat Med. 13(1):65–68.
- Yang CY, Chen L, Wang J, Yang YM, Li L. 2014. Pollen morphology of the Chinese endemic *Clematoclethra* (Actinidiaceae) and its taxonomic implications. Plant Divers Resour. 36:569–577.
- Zhao YM, Zhao ZX, Tang WL, Li XL, Zhao YP, Gao BY, Xie XD, Zhang XB. 2019. Characterization of the complete chloroplast genome of *Actinidia melanandra* (Actinidiaceae). Mitochondrial DNA B Resour. 4(2):2195–2196.