

Article

Increased Circulating Soluble Junctional Adhesion Molecules in Systemic Sclerosis: Association with Peripheral Microvascular Impairment

Eloisa Romano ^{1,†} , Irene Rosa ^{2,†} , Bianca Saveria Fioretto ^{1,2} , Marco Matucci-Cerinic ^{1,3} and Mirko Manetti ^{2,*} 

¹ Section of Internal Medicine, Department of Experimental and Clinical Medicine, University of Florence, 50134 Florence, Italy

² Section of Anatomy and Histology, Department of Experimental and Clinical Medicine, University of Florence, 50134 Florence, Italy

³ Unit of Immunology, Rheumatology, Allergy and Rare Diseases (UnIRAR), IRCCS San Raffaele Hospital, 20132 Milan, Italy

* Correspondence: mirko.manetti@unifi.it; Tel.: +39-055-2758073

† These authors contributed equally to this work.



Citation: Romano, E.; Rosa, I.; Fioretto, B.S.; Matucci-Cerinic, M.; Manetti, M. Increased Circulating Soluble Junctional Adhesion Molecules in Systemic Sclerosis: Association with Peripheral Microvascular Impairment. *Life* **2022**, *12*, 1790. <https://doi.org/10.3390/life12111790>

Academic Editor: Friedrich Jung

Received: 4 October 2022

Accepted: 3 November 2022

Published: 4 November 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Abstract: Systemic sclerosis (SSc, scleroderma) is a severe disease characterized by peripheral microcirculation abnormalities manifesting with Raynaud's phenomenon, nailfold videocapillaroscopic (NVC) changes, and even ischemic digital ulcers (DUs) that are often refractory to treatments. In the wake of previously described associations between the circulating levels of soluble junctional adhesion molecules (sJAMs) and SSc clinical features, here, we measured sJAM-A and sJAM-C levels by enzyme-linked immunosorbent assay in serum samples from a large case series of 110 SSc patients and 85 healthy controls, focusing on their possible association with peripheral vascular clinical features and their potential as biomarkers that are either diagnostic or mirror SSc-related microvasculopathy severity. Our data demonstrated that serum sJAM-A and sJAM-C are significantly increased in patients with SSc vs. healthy controls, especially in those featuring early/active NVC patterns and the presence of ischemic DUs. Moreover, circulating sJAM-C levels showed good diagnostic accuracy in discriminating between patients and controls, as assessed by receiver operator characteristics curve analysis. Finally, logistic regression revealed that, when comparing sJAM-A to sJAM-C, the latter might be better suited as a biomarker for SSc-related DUs. Our promising findings provide the necessary groundwork for longitudinal follow-up analyses of SSc patients aiming to assess whether circulating sJAM-C levels might be predictive for the development of new DUs, as well as DU recurrence and/or refractoriness to targeted therapies.

Keywords: systemic sclerosis; scleroderma; junctional adhesion molecules; sJAM-A; sJAM-C; peripheral microvascular damage; digital ulcers; enzyme-linked immunosorbent assay

1. Introduction

Systemic sclerosis (SSc, scleroderma) is a severe disease affecting the connective tissue that is characterized by the combination of immunological disturbances, widespread peripheral microvasculopathy, and progressive cutaneous and visceral fibrosis [1–3]. In the early phases of the disease course, SSc patients experience peripheral circulatory disturbances that precede fibrosis by months or years and include microvascular tone dysregulation, clinically manifesting with Raynaud's phenomenon and microangiopathy, as mirrored by nailfold capillaroscopic changes comprising capillary enlargement, bleeding, stenosis, and destruction [1–3]. Such microvascular dysfunction, characterized by endothelial cell (EC) activation/damage and perivascular infiltration/accumulation of lymphocytes and monocytes in the affected tissues [4,5], may frequently lead to significant peripheral ischemic

manifestations such as digital ulcers (DUs) [5,6], i.e., disabling and painful lesions often resulting in infections and gangrene, heavily compromising patients' quality of life [5,6]. Perivascular inflammation, which represents a typical hallmark of the early stages of the disease, is a consequence of an extravasation process mediated by the interaction between leukocytes and ECs, a mechanism highly reliant on the expression and function of cell surface adhesion molecules [7,8]. In addition to mediating leukocyte trafficking and inflammation, adhesion molecules play important roles in angiogenesis [9]. Indeed, endothelial adhesion molecules have been reported to regulate EC motility and to promote the influx of monocytes into tissues, where they can differentiate into macrophages and secrete proangiogenic factors, and to induce neovascularization with their soluble forms [9–12]. In this context, a number of studies suggested that, in the early stages of SSc, increased levels of EC-released soluble adhesion molecules may reflect the ongoing EC activation state and correlate with the presence and severity of specific organ complications [7,13,14].

Among the pool of cell-to-cell adhesion molecules, junctional adhesion molecule (JAM)-A (also known as JAM-1/F11 receptor) and JAM-C (JAM-3) are proteins belonging to the immunoglobulin superfamily and are broadly expressed on ECs, epithelial cells, fibroblasts, and circulating cells and are able to regulate leukocyte transmigration across the endothelium thanks to their ability to undergo heterophilic binding with the leukocyte integrins [15–17]. In addition, they can establish homophilic interactions at both endothelial and epithelial tight junctions, thus, participating in the regulation of paracellular permeability [15–17]. Since JAMs may also modulate cell adhesion, migration, and neovascularization, they have been implicated in several pathologic conditions, including cancer, hypertension, rheumatoid arthritis, and inflammatory bowel disease [18–21]. The soluble forms of these adhesion molecules have also been quantified in the circulation of patients affected by different pathologies, with their levels often correlating with disease severity [19,21–26]. As far as SSc is concerned, an abnormal expression of both JAM-A and JAM-C has been reported in the skin and blood of patients [7,27,28]. In particular, a previous study from our group demonstrated an aberrant expression of these JAMs not only in the skin of SSc patients, but also in SSc cultured dermal microvascular ECs, as well as in healthy microvascular ECs challenged with SSc sera [7]. Moreover, circulating levels of soluble JAM-A (sJAM-A) and sJAM-C were found to be significantly increased in patients with early-stage SSc and to correlate with different parameters of microvascular damage [7]. On the basis of such a preliminary scientific background and considering that our previous study was carried out on a relatively small cohort of patients, the aim of the present research was to evaluate sJAM-A and sJAM-C serum levels by enzyme-linked immune-sorbent assay (ELISA) in a larger case series of SSc patients, focusing on their association with peripheral vascular disease features and their possible potential as biomarkers that are either diagnostic or mirror SSc-related microvasculopathy severity.

2. Materials and Methods

2.1. Patients, Controls, and Serum Samples

Serum samples were obtained from 110 patients (101 women and 9 men; mean \pm SD age 57.1 ± 14.2 years) recruited from the Division of Rheumatology and Scleroderma Unit, Azienda Ospedaliero-Universitaria Careggi (AOUC), Florence, Italy, and fulfilling the American College of Rheumatology/European League Against Rheumatism 2013 classification criteria for SSc [29]. SSc patients with symptoms which overlap with those of other autoimmune, rheumatic, and/or connective tissue diseases were excluded from the study. Patients were under vasoactive/vasodilating drugs as follows: calcium channel blockers (72.6%), endothelin receptor antagonists (17.6%), phosphodiesterase 5 inhibitors (16.5%), and iloprost (22.0%). Before blood sampling, they were washed out for 10 days from oral vasoactive/vasodilating drugs and for 2 months from intravenous iloprost. At the time of blood sample collection, patients were not under immunosuppressants or other disease-modifying drugs. Eighty-five age- and sex-matched healthy individuals (78 women, 7 men; mean \pm SD age 58.5 ± 13.2 years) were used as controls. The presence

of primary Raynaud's phenomenon was considered as an exclusion criterion. Fresh venous blood samples from both patients and controls were drawn, allowed to clot for 30 min, and centrifugated at $1500 \times g$ for 15 min. Serum was then collected and stored in aliquots at -80°C until used. The study was conducted in agreement with the Declaration of Helsinki and approved by the local institutional review board at the AOUC, Florence, Italy (approval number: AOUC 69/13; approval date: 17 June 2013). All the involved individuals provided written informed consent.

2.2. Clinical Assessment

Patients were classified as having limited cutaneous SSc (lcSSc; $n = 72$) or diffuse cutaneous SSc (dcSSc, $n = 38$) according to the criteria of LeRoy et al. [30] and phenotypically assessed as recommended [31]. All patients reported the presence of Raynaud's phenomenon. At the time blood was drawn, the occurrence of ischemic DUs on the fingertips and other finger areas of SSc patients was recorded, and microvascular abnormalities on all 10 fingers were assessed by nailfold videocapillaroscopy (NVC). Briefly, patients were allowed to adapt to room temperature for at least 15 min and then their nailfolds were evaluated for the presence of pericapillary edema, microhemorrhages, enlarged and giant capillaries, ramified or bushy capillaries, disorganization of the vascular distribution, and loss of capillaries. The three different NVC patterns were identified as follows: (i) "early" NVC pattern, featuring few enlarged/giant capillaries and capillary microhemorrhages, no evident loss of capillaries and a relatively well-preserved capillary bed; (ii) "active" NVC pattern, characterized by giant capillaries and capillary microhemorrhages, absence/presence of few ramified capillaries, moderate capillary loss, and mild disorganization of the capillary structure; and (iii) "late" NVC pattern, with irregular capillary enlargement, absence/presence of few giant capillaries, no microhemorrhages, frequent ramified/bushy capillaries, severe loss of capillaries with large avascular areas, and disorganization of the normal capillary architecture [32]. A summary of the main features of the SSc patients is shown in Table 1.

Table 1. Demographic and clinical characteristics of SSc patients.

Characteristics	SSc Patients ($n = 110$)
Age, mean \pm SD (years)	57.1 \pm 14.2
Sex	
Male	9 (8.2)
Female	101 (91.8)
Disease subset	
lcSSc	72 (65.5)
dcSSc	38 (34.5)
Disease duration, mean \pm SD (years)	4.3 \pm 2.9
Digital ulcers	44 (40.0)
NVC pattern	
Early	36 (32.7)
Active	44 (40.0)
Late	30 (27.3)
Vasoactive/vasodilator drugs	
Calcium channel blockers	66 (60.0)
Endothelin receptor antagonists	16 (14.5)
Phosphodiesterase 5 inhibitors	15 (13.6)
Iloprost	20 (18.2)

Except where indicated otherwise, values are n (%) of subjects. dcSSc, diffuse cutaneous SSc; lcSSc, limited cutaneous SSc; NVC, nailfold videocapillaroscopy; SSc, systemic sclerosis.

2.3. Assay for Serum sJAM-A

Serum levels of sJAM-A were quantified by commercial quantitative colorimetric sandwich ELISA using a 96-well microtiter plate precoated with a capture antibody specific to sJAM-A and a biotin-conjugated detection antibody (catalog no. EKU05421; Biomatik, Wilmington, DE, USA). Each sample was measured in duplicate. According to the man-

manufacturer's protocol, once both standards and serum samples (100 μL /well) were added to the wells, the plate was left to incubate for 1 h at 37 °C. The liquid was then removed without washing, and the wells were incubated for 1 h at 37 °C with 100 μL of detection reagent A (biotin-conjugated detection antibody). The reaction, developed by adding 100 μL /well of detection reagent B (HRP conjugate) and 90 μL /well of tetramethylbenzidine (TMB) substrate solution for 10–20 min at 37 °C, was stopped by the addition of 50 μL of a sulfuric acid stop solution. The optical density (OD) of each well was measured using a microplate reader at 450 nm, and the concentration of serum sJAM-A was determined by comparing the OD of the samples to the standard curve. The detection range and the sensitivity of the assay were 0.313–20 ng/mL and 0.127 ng/mL, respectively.

2.4. Assay for Serum sJAM-C

Serum levels of sJAM-C were measured by commercial ELISA (catalog no. E2769Hu; BT Lab, Shanghai, China), following the manufacturer's instructions. Briefly, after adding standards (50 μL /well) and serum samples (40 μL /well) to the 96-well microplate precoated with an antibody specific to sJAM-C, 10 μL /well of a biotin-conjugated detection antibody and 50 μL /well of streptavidin–HRP were applied and left to incubate for 1 h at 37 °C. Next, following 5 washes, the reaction was developed by incubating the wells for 10 min at 37 °C in the dark with a 50 μL /well of substrate solution A and 50 μL /well of substrate solution B and, finally, terminated with 50 μL /well of stop solution. Color change was measured spectrophotometrically at a wavelength of 450 nm. The concentration of sJAM-C in the samples was determined by comparing the OD of each sample to those of the standard curve. The detection range and the sensitivity of the assay were 0.05–20 ng/mL and 0.021 ng/mL, respectively. Each sample was measured in duplicate.

2.5. Statistical Analysis

Statistical data analysis was performed by means of the SPSS software for Windows Version 28.0 (SPSS, Chicago, IL, USA). Descriptive statistics for continuous variables were expressed as the mean \pm SD or median and interquartile range (IQR), while those for categorical variables were reported as number and percentage. The non-parametric Mann–Whitney U test was used to assess serum sJAM-A or sJAM-C differences between two independent groups, while the Spearman ρ correlation coefficient was calculated to examine the relationship between two continuous variables. To verify the accuracy of circulating sJAM-A and sJAM-C levels for the diagnosis of SSc, the sensitivity (capability of the test to identify true positive subjects) and specificity (capability of the test to identify true negative subjects) of the test were evaluated for each molecule through receiver operator characteristics (ROC) curve analysis and the following estimation of the area under the curve (AUC) [33]. AUC values were interpreted as follows: 0.5–0.6 (failed), 0.6–0.7 (worthless), 0.7–0.8 (poor), 0.8–0.9 (good), >0.9 (excellent) [33]. Youden's index (i.e., Sensitivity – (1 – Specificity)) was also applied in order to evaluate the best cutoff value in our experimental data distributions. Since the Mann–Whitney U test unveiled that serum levels of both sJAMs were significantly increased in SSc patients with DUs and early/active NVC patterns, we performed multiple logistic regression analysis including sJAM-A and sJAM-C as independent variables and a single dependent variable each time (i.e., presence of DUs and NVC pattern). Odds ratios (ORs) with 95% confidence intervals (95% CIs) were determined. All *p*-values were two-tailed, and *p*-values < 0.05 were considered statistically significant.

3. Results

3.1. Serum Levels of Both sJAM-A and sJAM-C Are Increased in SSc Patients

Circulating levels of sJAM-A were significantly augmented in SSc patients (median 0.65 ng/mL, IQR 0.00–1.48 ng/mL) compared with healthy controls (median 0.20 ng/mL, IQR 0.00–0.42 ng/mL; *p* < 0.001; Figure 1A). sJAM-A levels were higher both in patients with lcSSc (median 0.66 ng/mL, IQR 0.00–1.36 ng/mL) and in those with dcSSc (me-

dian 0.64 ng/mL, IQR 0.00–1.63 ng/mL) than in controls (both $p < 0.001$; Figure 1A). Similarly, serum levels of sJAM-C were significantly increased in SSc patients (median 0.63 ng/mL, IQR 0.37–1.86 ng/mL) compared with healthy controls (median 0.00 ng/mL, IQR 0.00–0.46 ng/mL; $p < 0.001$; Figure 1B), as well as in both SSc cutaneous subsets vs. controls (lcSSc: median 0.60 ng/mL, IQR 0.37–1.68 ng/mL; dcSSc: median 0.66 ng/mL, IQR 0.37–1.91 ng/mL; both $p < 0.001$; Figure 1B). No significant difference in sJAM-A and sJAM-C levels was found between lcSSc and dcSSc patients.

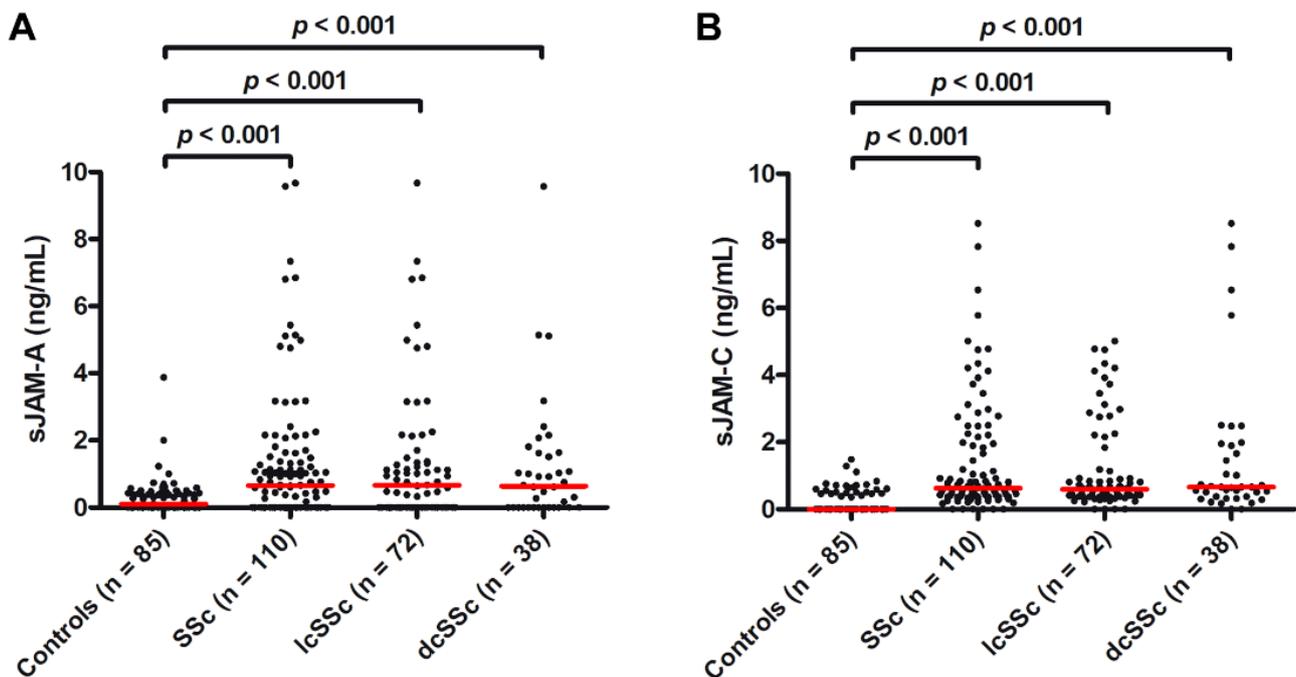


Figure 1. Serum levels of both (A) sJAM-A and (B) sJAM-C are increased in patients with SSc. (A) sJAM-A and (B) sJAM-C levels in healthy controls, patients with SSc, lcSSc, and dcSSc. Data are shown as dot plots. Each dot represents a subject. Horizontal red lines indicate the median value of each group. The non-parametric Mann–Whitney U test for independent samples was used to analyze serum sJAM-A and sJAM-C differences between groups. dcSSc, diffuse cutaneous SSc; lcSSc, limited cutaneous SSc; sJAM, soluble junctional adhesion molecule; SSc, systemic sclerosis.

3.2. Diagnostic Accuracy of sJAM-A and sJAM-C for SSc

The ROC curves and the corresponding AUC were plotted in order to evaluate the diagnostic accuracy of the two assessed circulating molecules. In particular, the diagnostic accuracy of sJAM-A was found to be poor (AUC = 0.670, 95% CI 0.604–0.754), while the diagnostic accuracy of sJAM-C was found to be good (AUC = 0.833, 95% CI 0.775–0.891) (Figure 2A,B). In addition, for sJAM-C, the ROC curve analysis revealed a cutoff value of 0.09 ng/mL, with 94.5% sensitivity and 69.4% specificity in discriminating between SSc patients and healthy controls.

3.3. Association of sJAM-A and sJAM-C Serum Levels with the Severity of Microvascular Impairment

As a measure of peripheral microvascular involvement, we further investigated the presence of a possible association of serum sJAM-A and sJAM-C with the NVC pattern and the occurrence of ischemic DUs. As far as NVC is concerned, when compared to controls, both circulating levels of sJAM-A and sJAM-C were found to be significantly higher in SSc patients with either early/active NVC patterns (median 0.89 ng/mL, IQR 0.00–2.12 ng/mL for sJAM-A and median 0.66 ng/mL, IQR 0.40–2.16 ng/mL for sJAM-C; $p < 0.001$ for both molecules; Figure 3A,B) or a late NVC pattern (median 2.23 ng/mL, IQR 0.60–3.44 ng/mL for sJAM-A and median 0.66 ng/mL, IQR 0.40–2.16 ng/mL for sJAM-C;

$p = 0.045$ for sJAM-A and $p < 0.001$ for sJAM-C; Figure 3A,B). When comparing sJAM levels between the two NVC groups, the values of both sJAM-A and sJAM-C were higher in the early/active pattern group compared to in the late one ($p = 0.049$ and $p = 0.043$, respectively; Figure 3A,B).

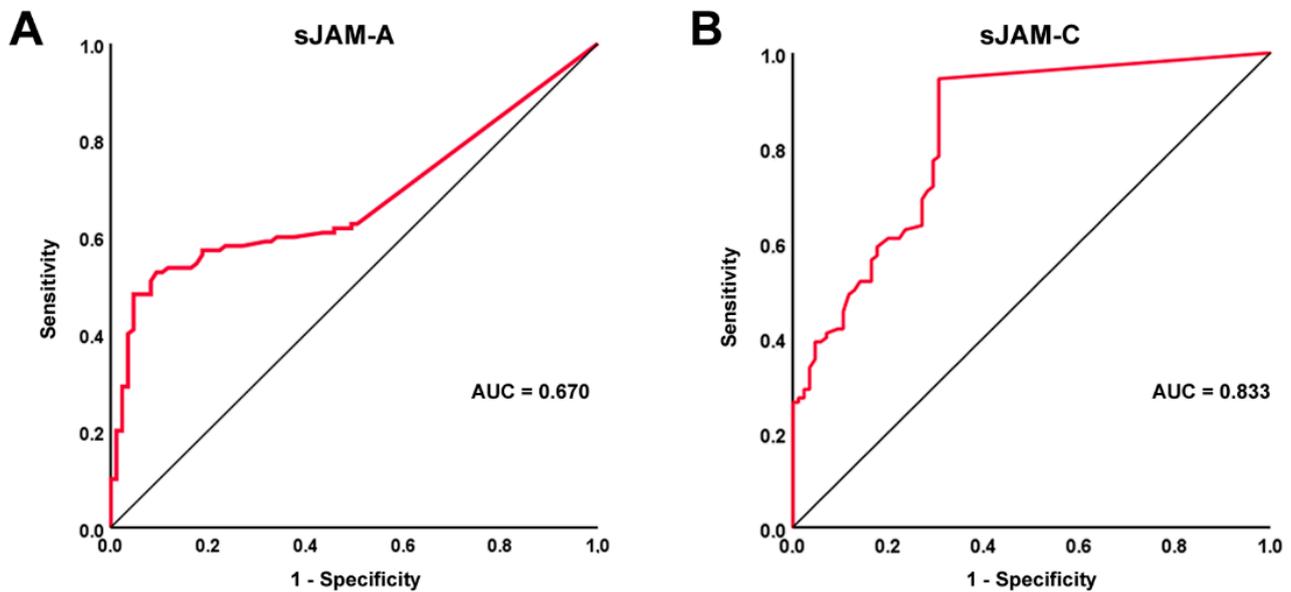


Figure 2. ROC curve (red line) plots for (A) sJAM-A and (B) sJAM-C in SSc patients vs. healthy controls. AUC values and reference lines for each curve are shown. AUC, area under the curve; ROC, receiver operator characteristic; sJAM, soluble junctional adhesion molecule; SSc, systemic sclerosis.

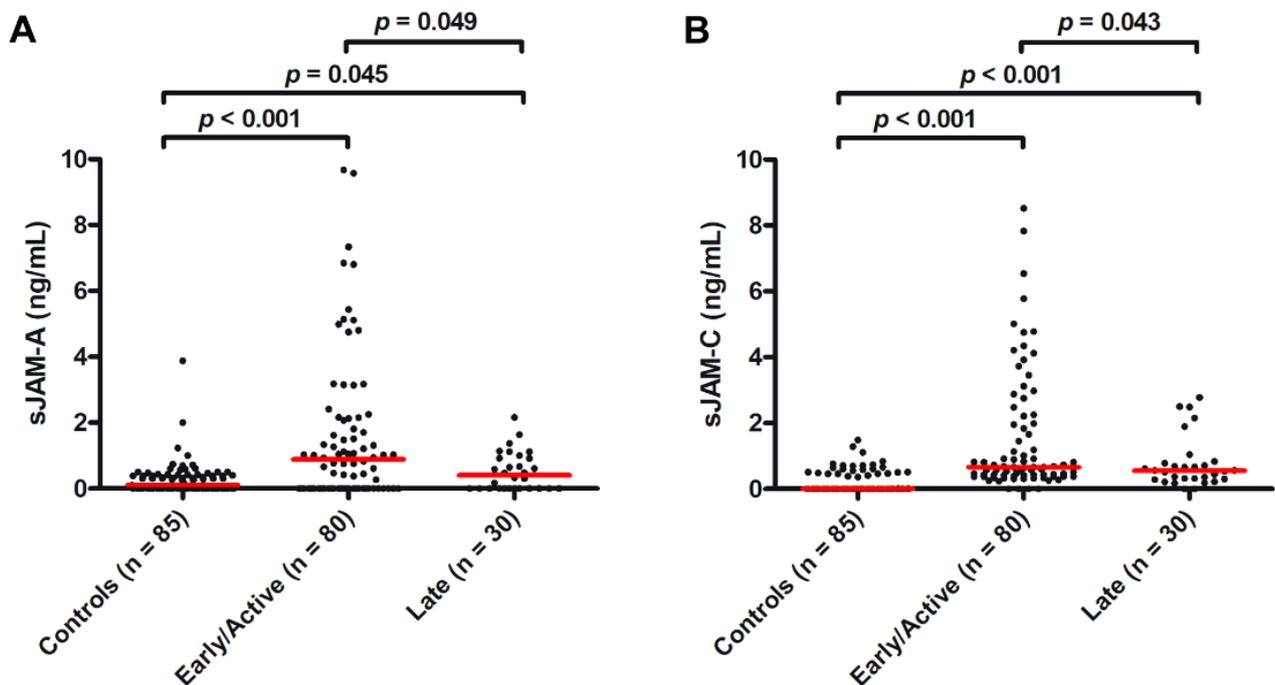


Figure 3. Serum levels of (A) sJAM-A and (B) sJAM-C in healthy controls and SSc patients stratified according to early/active and late nailfold videocapillaroscopic patterns. Data are shown as dot plots. Each dot represents a subject. Horizontal red lines indicate the median value of each group. The non-parametric Mann–Whitney U test for independent samples was used to analyze serum sJAM-A and sJAM-C differences between groups. sJAM, soluble junctional adhesion molecule; SSc, systemic sclerosis.

As far as ischemic DUs are concerned, the circulating levels of both sJAM-A and sJAM-C were found to be significantly higher when compared to controls both in SSc patients with DUs (median 1.01 ng/mL, IQR 0.00–2.37 ng/mL for sJAM-A and median 0.84 ng/mL, IQR 0.54–3.58 ng/mL for sJAM-C; $p < 0.001$ for both molecules; Figure 4A,B) and in those without DUs (median 0.53 ng/mL, IQR 0.00–1.12 ng/mL for sJAM-A and median 0.47 ng/mL, IQR 0.34–0.82 ng/mL for sJAM-C; $p = 0.003$ for sJAM-A and $p < 0.001$ for sJAM-C; Figure 4A,B). In addition, when comparing sJAM levels between the two SSc groups, the values of both sJAM-A and sJAM-C were significantly higher in patients with DUs compared to those without DUs ($p = 0.024$ and $p = 0.001$, respectively; Figure 4A,B).

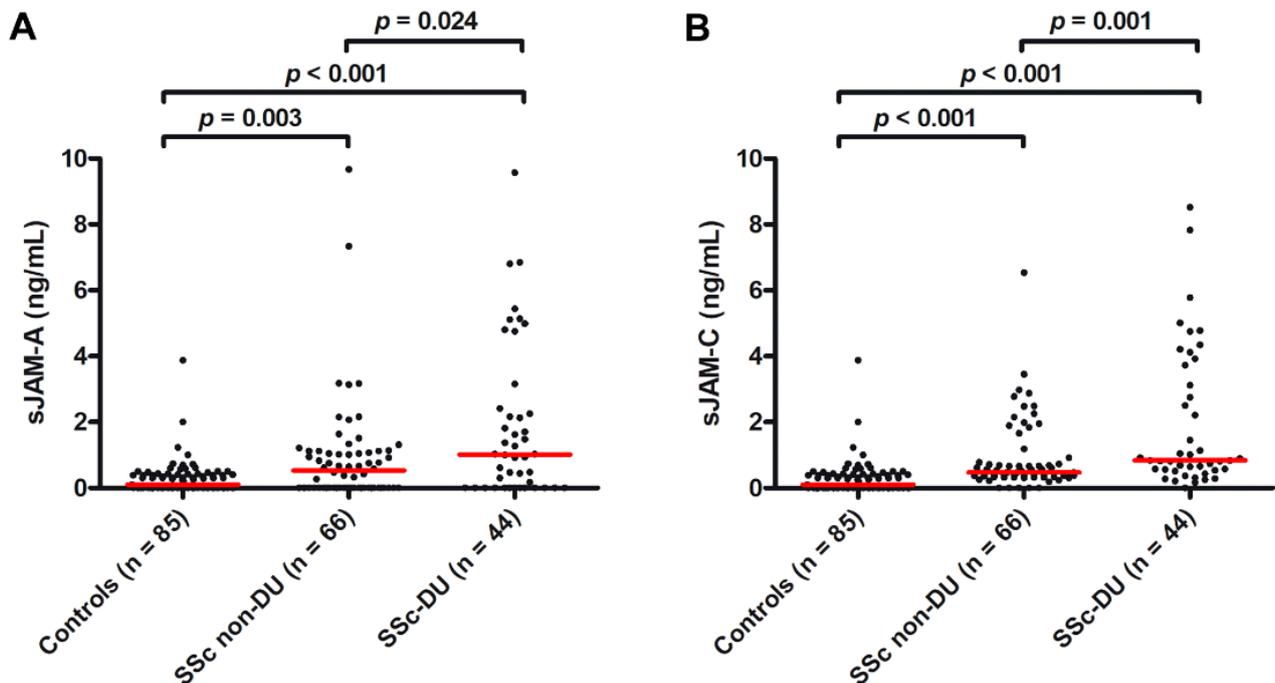


Figure 4. Serum levels of (A) sJAM-A and (B) sJAM-C in healthy controls and SSc patients with and without ischemic DUs. Data are represented as dot plots. Each dot represents a subject. Horizontal red lines indicate the median value of each group. The non-parametric Mann–Whitney U test for independent samples was used to analyze serum sJAM-A and sJAM-C differences between groups. DU, digital ulcers; sJAM, soluble junctional adhesion molecule; SSc, systemic sclerosis.

3.4. Correlation between Serum Levels of sJAM-A and sJAM-C and Logistic Regression Model

Circulating levels of sJAM-A and sJAM-C were directly correlated with one another in SSc patients ($\rho = 0.516$, $p < 0.001$; Figure 5) but not in healthy controls.

Since, when comparing SSc subgroup medians, we found that serum levels of both sJAMs were significantly increased in SSc patients with DUs and early/active NVC patterns, we finally performed multiple logistic regression analysis combining serum sJAM-A and sJAM-C as independent variables and one of the two abovementioned disease phenotypes (i.e., presence of DUs and NVC pattern) as a single dependent variable each time. The results of the logistic regression analysis are shown in Table 2.

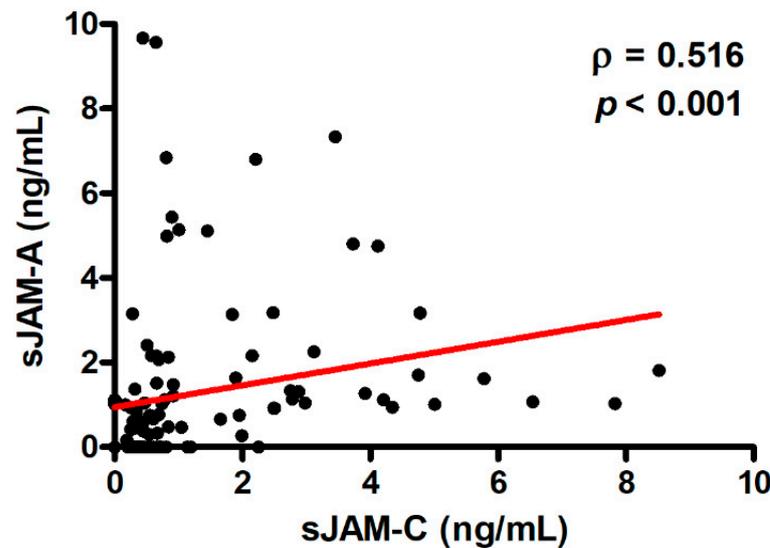


Figure 5. Correlation between sJAM-A and sJAM-C levels in patients with SSc. Data are displayed as a scatterplot where each dot represents a patient. Correlation coefficient (ρ) and p -values are indicated. sJAM, soluble junctional adhesion molecule; SSc, systemic sclerosis.

Table 2. Logistic regression analysis model combining serum sJAM-A and sJAM-C levels in SSc patients.

		Early/Active NVC	DUs
sJAM-A	OR (95% CI)	0.65 (0.41–1.04)	1.28 (1.01–1.61)
	p	0.073	0.04
sJAM-C	OR (95% CI)	0.75 (0.50–1.15)	1.48 (1.12–1.96)
	p	0.189	0.007

CI, confidence interval; DUs, digital ulcers; NVC, nailfold videocapillaroscopy; OR, odds ratio; sJAM, soluble junctional adhesion molecule; SSc, systemic sclerosis.

4. Discussion

In the wake of previously described associations between circulating levels of sJAMs and SSc [7,27], our data, obtained from a larger case–control cohort, allowed us to demonstrate that serum sJAM-A and sJAM-C were significantly increased in SSc vs. healthy controls, especially in patients characterized by early/active NVC patterns and the presence of ischemic DUs. Moreover, we showed for the first time that sJAM-C levels have a good diagnostic accuracy in discriminating between SSc patients and controls, as assessed by ROC curve analysis with cutoff value. Finally, logistic regression revealed that, when comparing sJAM-A to sJAM-C, the latter might be better suited as a biomarker for SSc-related DUs.

JAMs are cell adhesion molecules expressed by different cell types, including fibroblasts, in which they contribute to the establishment of their intercellular junctions and participate in intercellular crosstalk, retainment of myeloid cells, and wound healing [7,34,35], and ECs, in which they control tight junction maintenance and mediate leukocyte extravasation [21,36–39]. Due to their ability to trigger intracellular signal cascades at intercellular contact sites, JAMs are involved in different pathophysiological processes such as leukocyte recruitment to sites of inflammation and ischemia–reperfusion injury, atherogenesis, angiogenesis, and fibrosis [7,20,35,38–42]. As far as angiogenesis is concerned, JAM-A was found to be crucial for proper EC motility, directional movement, and focal contact formation and was shown to be involved in EC migration through integrin $\alpha v \beta 3$ [43] and to mediate basic fibroblast growth-factor-induced angiogenesis [40,44], an effect that was withdrawn in JAM-A deficient mice [45] or after EC-specific JAM-A depletion [40,46]. JAM-C has also been reported to act as a proangiogenic molecule, as its soluble form was

demonstrated to induce *in vitro* angiogenesis [21], and its blockade was able to reduce tumor growth and decrease angiogenesis both *in vivo* and *in vitro* [47]. In line with these findings, overexpression of both JAM-A and JAM-C was shown to enhance tumor angiogenesis and metastasization in different *in vitro* and *in vivo* studies [48–51]. As already mentioned above, JAM-A and JAM-C are also supposed to be implicated in fibrosis. Indeed, JAM-A overexpression in mouse fibroblasts was demonstrated to significantly augment the secretory capacity and proliferation of these cells [35], while increased JAM-C expression was detected in experimental hepatic fibrosis [20,42,52].

In SSc, a disease in which an early proinflammatory state and microvasculopathy progressively culminate in fibrosis and loss of angiogenesis [53–55], aberrant JAM-A and JAM-C protein expression was reported in skin biopsies as well as in dermal fibroblasts and ECs [7,27,28]. Moreover, in a previous preliminary study from our group, circulating sJAM-A and sJAM-C levels were shown to be particularly high in patients with early-stage SSc compared to healthy controls, although no difference was found between the totality of patients and controls in the relatively small cohort analyzed [7]. At variance with our previous report, here, we demonstrate, in a much larger case–control cohort, that circulating sJAM levels are significantly increased in the whole group of SSc patients respective to the healthy subjects. Such a discrepancy may be explained either because our earlier study was underpowered or by the fact that, since here we aimed to assess the diagnostic potential of these molecules, we did not include SSc patients with longstanding disease. As demonstrated by the ROC curve analysis, we additionally revealed that sJAM-C levels might be a useful diagnostic tool for discriminating between SSc and a healthy condition, evidence that is further supported by the fact that sJAM-C in patients with SSc was always significantly higher than in controls regardless of the clinical subgroup considered.

Interestingly, increased sJAM levels have been described in other different pathologic conditions. In particular, sJAM-A was found to be higher in patients with coronary artery disease, arterial hypertension, renal insufficiency, and multiple myeloma [19,23,24,26,56], while sJAM-C was shown to be augmented in rheumatoid arthritis and macular degeneration [21,25]. Consistent with our previously published results [7], we herein confirm in a larger sample that both sJAM-A and sJAM-C are predominantly higher in patients with early/active NVC patterns, which are characterized by an ongoing dysregulated angiogenic response resulting in microhemorrhages and immature and unstable giant microvessels, suggesting that an increase in sJAMs might not only reflect but possibly also actively contribute to the derangement of peripheral microcirculation in SSc patients. In addition, we not only validated the previously reported association between higher circulating levels of both sJAMs and SSc-related DUs, but, through logistic regression, also uncovered sJAM-C as a better biomarker for more severe peripheral vasculopathy characterized by the development of ischemic DUs. Strikingly, the potential of serum sJAM-C levels as a biomarker was also suggested for wet, age-related macular degeneration, which is characterized by vascular abnormalities into the macula [25].

The presence of soluble JAMs in the circulation depends on the cleavage of the extracellular domain of cell surface JAMs [21,57–59]. In particular, it has been reported that, under inflammatory conditions, the endothelial JAM-A and JAM-C extracellular domains are cleaved and secreted as soluble forms by a disintegrin and metalloproteinases (ADAMs) as well as neutrophil elastases [21,57,59]. Of note, patients with SSc show both an augmented protein expression of cell-membrane-bound JAM-A and JAM-C, and a significant upregulation of ADAM-17 and ADAM-12 proteins [7,60,61], which, together, might partly explain the increased concentrations of sJAM-A and sJAM-C in the circulation of SSc patients. To date, little is known about the mechanisms of action and/or the signaling cascades initiated by sJAMs. However, it has been hypothesized that sJAM-C may bind to membrane-bound JAM-B or JAM-C on the surface of ECs in order to initiate angiogenesis through the phosphorylation of Src, p38, and PI3K [21]. In addition, another study showed that the extracellular domain of JAM-C is cleaved and secreted in soluble form from adipose-derived stem cells, and released sJAM-C was demonstrated to couple with

JAM-B to promote cell adhesion, cell growth, and the expression of mesenchymal markers in such cells [62]. On the basis of the aforementioned background, our findings allow us to speculate that sJAM-C might be implicated in both the vascular and fibrotic aspects of SSc.

Based on the present data, we are aware that further studies are necessary to determine if serum levels of both sJAM-A and sJAM-C may correlate with additional clinical SSc manifestations. Furthermore, longitudinal studies monitoring changes in levels of sJAM-C in patients with a very early diagnosis of SSc [63] who progress into an established SSc disease will help us to gain further insights into the potential diagnostic value of this circulating molecule. Prospective follow-up analyses of SSc patients are also required to fully unravel whether sJAM-C might be suited as a risk biomarker for the development of new DUs, as well as DU recurrence and/or refractoriness to treatments. We also believe that additional *in vitro* and *in vivo* studies will help us to further explore the role of JAMs in SSc pathophysiology and to unveil whether the modulation of their expression/activity will allow us to find new targeted therapeutic strategies for the treatment of this devastating pathology. Finally, we are confident that our promising results will stimulate further research aimed at evaluating circulating levels of these adhesion molecules in other pathologic conditions characterized by microcirculatory alterations and tissue fibrosis.

Author Contributions: Conceptualization—E.R. and M.M.; methodology—E.R., I.R., B.S.F., M.M.-C. and M.M.; validation—E.R., I.R. and M.M.; formal analysis—E.R., I.R., B.S.F. and M.M.; investigation—E.R., I.R., B.S.F., M.M.-C. and M.M.; resources—M.M.-C. and M.M.; data curation—E.R., I.R. and M.M.; writing—original draft preparation—E.R., I.R. and M.M.; writing—review and editing—E.R., I.R., B.S.F., M.M.-C. and M.M.; visualization—E.R. and M.M.; supervision—E.R. and M.M.; project administration—M.M. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: The study was conducted in accordance with the Declaration of Helsinki and approved by the local institutional review board at the Azienda Ospedaliero-Universitaria Careggi, Florence, Italy (approval number: AOUC 69/13; approval date: 17 June 2013).

Informed Consent Statement: Informed consent was obtained from all subjects involved in the study.

Data Availability Statement: All relevant data are included within the manuscript.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Denton, C.P.; Khanna, D. Systemic sclerosis. *Lancet* **2017**, *390*, 1685–1699. [\[CrossRef\]](#)
2. Kobayashi, S.; Nagafuchi, Y.; Shoda, H.; Fujio, K. The pathophysiological roles of regulatory T cells in the early phase of systemic sclerosis. *Front. Immunol.* **2022**, *13*, 900638. [\[CrossRef\]](#) [\[PubMed\]](#)
3. Lescoat, A.; Varga, J.; Matucci-Cerinic, M.; Khanna, D. New promising drugs for the treatment of systemic sclerosis: Pathogenic considerations, enhanced classifications, and personalized medicine. *Expert. Opin. Investig. Drugs* **2021**, *30*, 635–652. [\[CrossRef\]](#) [\[PubMed\]](#)
4. Melissaropoulos, K.; Iliopoulos, G.; Sakkas, L.I.; Daoussis, D. Pathogenetic aspects of systemic sclerosis: A view through the prism of B cells. *Front. Immunol.* **2022**, *13*, 925741. [\[CrossRef\]](#)
5. Zanin-Silva, D.C.; Santana-Gonçalves, M.; Kawashima-Vasconcelos, M.Y.; Oliveira, M.C. Management of endothelial dysfunction in systemic sclerosis: Current and developing strategies. *Front. Med.* **2021**, *8*, 788250. [\[CrossRef\]](#) [\[PubMed\]](#)
6. Hughes, M.; Allanore, Y.; El Aoufy, K.; Denton, C.P.; Khanna, D.; Krieg, T.; Matucci-Cerinic, M. A practical approach to the management of digital ulcers in patients with systemic sclerosis: A narrative review. *JAMA Dermatol.* **2021**, *157*, 851–858. [\[CrossRef\]](#)
7. Manetti, M.; Guiducci, S.; Romano, E.; Rosa, I.; Ceccarelli, C.; Mello, T.; Milia, A.F.; Conforti, M.L.; Ibba-Manneschi, L.; Matucci-Cerinic, M. Differential expression of junctional adhesion molecules in different stages of systemic sclerosis. *Arthritis Rheum.* **2013**, *65*, 247–257. [\[CrossRef\]](#)
8. Bruni, C.; Frech, T.; Manetti, M.; Rossi, F.W.; Furst, D.E.; De Paulis, A.; Rivellese, F.; Guiducci, S.; Matucci-Cerinic, M.; Bellando-Randone, S. Vascular leaking, a pivotal and early pathogenetic event in systemic sclerosis: Should the door be closed? *Front. Immunol.* **2018**, *9*, 2045. [\[CrossRef\]](#)
9. Polverini, P.J. Cellular adhesion molecules: Newly identified mediators of angiogenesis. *Am. J. Pathol.* **1996**, *148*, 1023–1029.
10. Francavilla, C.; Maddaluno, L.; Cavallaro, U. The functional role of cell adhesion molecules in tumor angiogenesis. *Semin. Cancer Biol.* **2009**, *19*, 298–309. [\[CrossRef\]](#)

11. Harjunpää, H.; Lloret Asens, M.; Guenther, C.; Fagerholm, S.C. Cell adhesion molecules and their roles and regulation in the immune and tumor microenvironment. *Front. Immunol.* **2019**, *10*, 1078. [[CrossRef](#)] [[PubMed](#)]
12. Bulska-Będkowska, W.; Czajka-Francuz, P.; Jurek-Cisoń, S.; Owczarek, A.J.; Francuz, T.; Chudek, J. The predictive role of serum levels of soluble cell adhesion molecules (sCAMs) in the therapy of advanced breast cancer—a single-centre study. *Medicina* **2022**, *58*, 153. [[CrossRef](#)] [[PubMed](#)]
13. Kuryliszyn-Moskal, A.; Klimiuk, P.A.; Sierakowski, S. Soluble adhesion molecules (sVCAM-1, sE-selectin), vascular endothelial growth factor (VEGF) and endothelin-1 in patients with systemic sclerosis: Relationship to organ systemic involvement. *Clin. Rheumatol.* **2005**, *24*, 111–116. [[CrossRef](#)]
14. Yanaba, K.; Yoshizaki, A.; Muroi, E.; Ogawa, F.; Shimizu, K.; Sato, S. Increased circulating soluble vascular adhesion protein-1 levels in systemic sclerosis: Association with lower frequency and severity of interstitial lung disease. *Int. J. Rheum. Dis.* **2013**, *16*, 442–447. [[CrossRef](#)] [[PubMed](#)]
15. Wang, J.; Liu, H. The roles of junctional adhesion molecules (JAMs) in cell migration. *Front. Cell. Dev. Biol.* **2022**, *10*, 843671. [[CrossRef](#)]
16. Lauko, A.; Mu, Z.; Gutmann, D.H.; Naik, U.P.; Lathia, J.D. Junctional adhesion molecules in cancer: A paradigm for the diverse functions of cell-cell interactions in tumor progression. *Cancer Res.* **2020**, *80*, 4878–4885. [[CrossRef](#)] [[PubMed](#)]
17. Hartmann, C.; Schwietzer, Y.A.; Otani, T.; Furuse, M.; Ebnet, K. Physiological functions of junctional adhesion molecules (JAMs) in tight junctions. *Biochim. Biophys. Acta Biomembr.* **2020**, *1862*, 183299. [[CrossRef](#)] [[PubMed](#)]
18. Bonilha, C.S.; Benson, R.A.; Brewer, J.M.; Garside, P. Targeting Opposing immunological roles of the junctional adhesion molecule-A in autoimmunity and cancer. *Front. Immunol.* **2020**, *11*, 602094. [[CrossRef](#)]
19. Ong, K.L.; Leung, R.Y.; Babinska, A.; Salifu, M.O.; Ehrlich, Y.H.; Kornecki, E.; Wong, L.Y.; Tso, A.W.; Cherny, S.S.; Sham, P.C.; et al. Elevated plasma level of soluble F11 receptor/junctional adhesion molecule-A (F11R/JAM-A) in hypertension. *Am. J. Hypertens.* **2009**, *22*, 500–505. [[CrossRef](#)]
20. Hintermann, E.; Christen, U. The many roles of cell adhesion molecules in hepatic fibrosis. *Cells* **2019**, *8*, 1503. [[CrossRef](#)]
21. Rabquer, B.J.; Amin, M.A.; Teegala, N.; Shaheen, M.K.; Tsou, P.S.; Ruth, J.H.; Lesch, C.A.; Imhof, B.A.; Koch, A.E. Junctional adhesion molecule-C is a soluble mediator of angiogenesis. *J. Immunol.* **2010**, *185*, 1777–1785. [[CrossRef](#)] [[PubMed](#)]
22. Reglero-Real, N.; Colom, B.; Bodkin, J.V.; Nourshargh, S. Endothelial cell junctional adhesion molecules: Role and regulation of expression in inflammation. *Arterioscler Thromb. Vasc. Biol.* **2016**, *36*, 2048–2057. [[CrossRef](#)] [[PubMed](#)]
23. Cavusoglu, E.; Kornecki, E.; Sobocka, M.B.; Babinska, A.; Ehrlich, Y.H.; Chopra, V.; Yanamadala, S.; Ruwende, C.; Salifu, M.O.; Clark, L.T.; et al. Association of plasma levels of F11 receptor/junctional adhesion molecule-A (F11R/JAM-A) with human atherosclerosis. *J. Am. Coll. Cardiol.* **2007**, *50*, 1768–1776. [[CrossRef](#)] [[PubMed](#)]
24. Rath, D.; Rapp, V.; Schwartz, J.; Winter, S.; Emschermann, F.; Arnold, D.; Rheinlaender, J.; Büttcher, M.; Strebl, M.; Braun, M.B.; et al. Homophilic interaction between transmembrane-JAM-A and soluble JAM-A regulates thrombo-inflammation: Implications for coronary artery disease. *JACC Basic Transl. Sci.* **2022**, *7*, 445–461. [[CrossRef](#)] [[PubMed](#)]
25. Hou, X.; Du, H.J.; Zhou, J.; Hu, D.; Wang, Y.S.; Li, X. Role of junctional adhesion molecule-C in the regulation of inner endothelial blood-retinal barrier function. *Front. Cell. Dev. Biol.* **2021**, *9*, 695657. [[CrossRef](#)]
26. Salifu, M.O.; Kolff, Q.; Murty, P.; Haria, D.M.; Zimpa, M.; Shakeel, M.; Lee, H.; Kornecki, E.; Babinska, A. Relationship between the soluble F11 receptor and markers of inflammation in hemodialysis patients. *J. Investig. Med.* **2007**, *55*, 115–119. [[CrossRef](#)]
27. Hou, Y.; Rabquer, B.J.; Gerber, M.L.; Del Galdo, F.; Jimenez, S.A.; Haines, G.K., 3rd; Barr, W.G.; Massa, M.C.; Seibold, J.R.; Koch, A.E. Junctional adhesion molecule-A is abnormally expressed in diffuse cutaneous systemic sclerosis skin and mediates myeloid cell adhesion. *Ann. Rheum. Dis.* **2010**, *69*, 249–254. [[CrossRef](#)]
28. Rabquer, B.J.; Hou, Y.; Del Galdo, F.; Haines, G.K., 3rd; Gerber, M.L.; Jimenez, S.A.; Seibold, J.R.; Koch, A.E. The proadhesive phenotype of systemic sclerosis skin promotes myeloid cell adhesion via ICAM-1 and VCAM-1. *Rheumatology* **2009**, *48*, 734–740. [[CrossRef](#)]
29. van den Hoogen, F.; Khanna, D.; Fransen, J.; Johnson, S.R.; Baron, M.; Tyndall, A.; Matucci-Cerinic, M.; Naden, R.P.; Medsger, T.A., Jr.; Carreira, P.E.; et al. 2013 classification criteria for systemic sclerosis: An American College of Rheumatology/European League against Rheumatism collaborative initiative. *Ann. Rheum. Dis.* **2013**, *72*, 1747–1755. [[CrossRef](#)]
30. LeRoy, E.C.; Black, C.; Fleischmajer, R.; Jablonska, S.; Krieg, T.; Medsger, T.A., Jr.; Rowell, N.; Wollheim, F. Scleroderma (systemic sclerosis): Classification, subsets, and pathogenesis. *J. Rheumatol.* **1988**, *15*, 202–205.
31. Valentini, G.; Matucci Cerinic, M. Disease-specific quality indicators, guidelines and outcome measures in scleroderma. *Clin. Exp. Rheumatol.* **2007**, *25*, 159–162. [[PubMed](#)]
32. Sulli, A.; Secchi, M.E.; Pizzorni, C.; Cutolo, M. Scoring the nailfold microvascular changes during the capillaroscopic analysis in systemic sclerosis patients. *Ann. Rheum. Dis.* **2008**, *67*, 885–887. [[CrossRef](#)] [[PubMed](#)]
33. Polo, T.C.F.; Miot, H.A. Use of ROC curves in clinical and experimental studies. *J. Vasc. Bras.* **2020**, *19*, e20200186. [[CrossRef](#)] [[PubMed](#)]
34. Morris, A.P.; Tawil, A.; Berkova, Z.; Wible, L.; Smith, C.W.; Cunningham, S.A. Junctional Adhesion Molecules (JAMs) are differentially expressed in fibroblasts and co-localize with ZO-1 to adherens-like junctions. *Cell. Commun. Adhes.* **2006**, *13*, 233–247. [[CrossRef](#)] [[PubMed](#)]

35. Gong, T.; Wu, M.; Fan, X.; Chen, Z.; Xia, Z. Lentivirus-mediated subcutaneous JAM-A modification promotes skin wound healing in a mouse model by strengthening the secretory function and proliferation of fibroblasts. *Cell. Biol. Int.* **2022**, *46*, 1227–1235. [[CrossRef](#)] [[PubMed](#)]
36. Czubak-Prowizor, K.; Babinska, A.; Swiatkowska, M. The F11 receptor (F11R)/junctional adhesion molecule-A (JAM-A) (F11R/JAM-A) in cancer progression. *Mol. Cell. Biochem.* **2022**, *477*, 79–98. [[CrossRef](#)]
37. Shi, J.; Barakat, M.; Chen, D.; Chen, L. Bicellular tight junctions and wound healing. *Int. J. Mol. Sci.* **2018**, *19*, 3862. [[CrossRef](#)]
38. Kummer, D.; Ebnet, K. Junctional adhesion molecules (JAMs): The JAM-integrin connection. *Cells* **2018**, *7*, 25. [[CrossRef](#)]
39. Ebnet, K. Junctional adhesion molecules (JAMs): Cell adhesion receptors with pleiotropic functions in cell physiology and development. *Physiol. Rev.* **2017**, *97*, 1529–1554. [[CrossRef](#)]
40. Weber, C.; Fraemohs, L.; Dejana, E. The role of junctional adhesion molecules in vascular inflammation. *Nat. Rev. Immunol.* **2007**, *7*, 467–477. [[CrossRef](#)]
41. Wang, J.; Chen, X. Junctional adhesion molecules: Potential proteins in atherosclerosis. *Front. Cardiovasc. Med.* **2022**, *9*, 888818. [[CrossRef](#)] [[PubMed](#)]
42. Hintermann, E.; Bayer, M.; Ehser, J.; Aurrand-Lions, M.; Pfeilschifter, J.M.; Imhof, B.A.; Christen, U. Murine junctional adhesion molecules JAM-B and JAM-C mediate endothelial and stellate cell interactions during hepatic fibrosis. *Cell. Adh. Migr.* **2016**, *10*, 419–433. [[CrossRef](#)] [[PubMed](#)]
43. Naik, M.U.; Naik, U.P. Junctional adhesion molecule-A-induced endothelial cell migration on vitronectin is integrin alpha v beta 3 specific. *J. Cell. Sci.* **2006**, *119*, 490–499. [[CrossRef](#)] [[PubMed](#)]
44. Peddibhotla, S.S.; Brinkmann, B.F.; Kummer, D.; Tuncay, H.; Nakayama, M.; Adams, R.H.; Gerke, V.; Ebnet, K. Tetraspanin CD9 links junctional adhesion molecule-A to $\alpha\beta 3$ integrin to mediate basic fibroblast growth factor-specific angiogenic signaling. *Mol. Biol. Cell* **2013**, *24*, 933–944. [[CrossRef](#)] [[PubMed](#)]
45. Cooke, V.G.; Naik, M.U.; Naik, U.P. Fibroblast growth factor-2 failed to induce angiogenesis in junctional adhesion molecule-A-deficient mice. *Arterioscler. Thromb. Vasc. Biol.* **2006**, *26*, 2005–2011. [[CrossRef](#)]
46. Naik, M.U.; Vuppalanchi, D.; Naik, U.P. Essential role of junctional adhesion molecule-1 in basic fibroblast growth factor-induced endothelial cell migration. *Arterioscler. Thromb. Vasc. Biol.* **2003**, *23*, 2165–2171. [[CrossRef](#)]
47. Lamagna, C.; Hodivala-Dilke, K.M.; Imhof, B.A.; Aurrand-Lions, M. Antibody against junctional adhesion molecule-C inhibits angiogenesis and tumor growth. *Cancer Res.* **2005**, *65*, 5703–5710. [[CrossRef](#)]
48. Solimando, A.G.; Da Vià, M.C.; Leone, P.; Borrelli, P.; Croci, G.A.; Tabares, P.; Brandl, A.; Di Lernia, G.; Bianchi, F.P.; Tafuri, S.; et al. Halting the vicious cycle within the multiple myeloma ecosystem: Blocking JAM-A on bone marrow endothelial cells restores angiogenic homeostasis and suppresses tumor progression. *Haematologica* **2021**, *106*, 1943–1956. [[CrossRef](#)]
49. Leinster, D.A.; Colom, B.; Whiteford, J.R.; Ennis, D.P.; Lockley, M.; McNeish, I.A.; Aurrand-Lions, M.; Chavakis, T.; Imhof, B.A.; Balkwill, F.R.; et al. Endothelial cell junctional adhesion molecule C plays a key role in the development of tumors in a murine model of ovarian cancer. *FASEB J.* **2013**, *27*, 4244–4253.
50. Santoso, S.; Orlova, V.V.; Song, K.; Sachs, U.J.; Andrei-Selmer, C.L.; Chavakis, T. The homophilic binding of junctional adhesion molecule-C mediates tumor cell-endothelial cell interactions. *J. Biol. Chem.* **2005**, *280*, 36326–36333. [[CrossRef](#)]
51. Fuse, C.; Ishida, Y.; Hikita, T.; Asai, T.; Oku, N. Junctional adhesion molecule-C promotes metastatic potential of HT1080 human fibrosarcoma. *J. Biol. Chem.* **2007**, *282*, 8276–8283. [[CrossRef](#)] [[PubMed](#)]
52. Hintermann, E.; Bayer, M.; Conti, C.B.; Fuchs, S.; Fausther, M.; Leung, P.S.; Aurrand-Lions, M.; Taubert, R.; Pfeilschifter, J.M.; Friedrich-Rust, M.; et al. Junctional adhesion molecules JAM-B and JAM-C promote autoimmune-mediated liver fibrosis in mice. *J. Autoimmun.* **2018**, *91*, 83–96. [[CrossRef](#)] [[PubMed](#)]
53. Matucci-Cerinic, M.; Manetti, M.; Bruni, C.; Chora, I.; Bellando-Randone, S.; Lepri, G.; De Paulis, A.; Guiducci, S. The “myth” of loss of angiogenesis in systemic sclerosis: A pivotal early pathogenetic process or just a late unavoidable event? *Arthritis Res. Ther.* **2017**, *19*, 162. [[CrossRef](#)]
54. Lazzaroni, M.G.; Piantoni, S.; Angeli, F.; Bertocchi, S.; Franceschini, F.; Airò, P. A narrative review of pathogenetic and histopathologic aspects, epidemiology, classification systems, and disease outcome measures in systemic sclerosis. *Clin. Rev. Allergy Immunol.* **2022**, *online ahead of print*. [[CrossRef](#)]
55. Moschetti, L.; Piantoni, S.; Vizzardi, E.; Sciatti, E.; Riccardi, M.; Franceschini, F.; Cavazzana, I. Endothelial dysfunction in systemic lupus erythematosus and systemic sclerosis: A common trigger for different microvascular diseases. *Front. Med.* **2022**, *9*, 849086. [[CrossRef](#)] [[PubMed](#)]
56. Solimando, A.G.; Brandl, A.; Mattenheimer, K.; Graf, C.; Ritz, M.; Ruckdeschel, A.; Stühmer, T.; Mokhtari, Z.; Rudelius, M.; Dotterweich, J.; et al. JAM-A as a prognostic factor and new therapeutic target in multiple myeloma. *Leukemia* **2018**, *32*, 736–743. [[CrossRef](#)] [[PubMed](#)]
57. Koenen, R.R.; Pruessmeyer, J.; Soehnlein, O.; Fraemohs, L.; Zerneck, A.; Schwarz, N.; Reiss, K.; Sarabi, A.; Lindbom, L.; Hackeng, T.M.; et al. Regulated release and functional modulation of junctional adhesion molecule A by disintegrin metalloproteinases. *Blood* **2009**, *113*, 4799–4809. [[CrossRef](#)]
58. Houry, N.; Huang, K.C.; Nalbantoglu, J. The coxsackievirus and adenovirus receptor (CAR) undergoes ectodomain shedding and regulated intramembrane proteolysis (RIP). *PLoS ONE* **2013**, *8*, e73296. [[CrossRef](#)]

59. Colom, B.; Bodkin, J.V.; Beyrau, M.; Woodfin, A.; Ody, C.; Rourke, C.; Chavakis, T.; Brohi, K.; Imhof, B.A.; Nourshargh, S. Leukotriene B4-neutrophil elastase axis drives neutrophil reverse transendothelial cell migration in vivo. *Immunity* **2015**, *42*, 1075–1086. [[CrossRef](#)]
60. Bohgaki, T.; Amasaki, Y.; Nishimura, N.; Bohgaki, M.; Yamashita, Y.; Nishio, M.; Sawada, K.I.; Jodo, S.; Atsumi, T.; Koike, T. Up regulated expression of tumour necrosis factor {alpha} converting enzyme in peripheral monocytes of patients with early systemic sclerosis. *Ann. Rheum. Dis.* **2005**, *64*, 1165–1173. [[CrossRef](#)]
61. Cipriani, P.; Di Benedetto, P.; Ruscitti, P.; Liakouli, V.; Berardicurti, O.; Carubbi, F.; Ciccina, F.; Guggino, G.; Zazzeroni, F.; Alesse, E.; et al. Perivascular cells in diffuse cutaneous systemic sclerosis overexpress activated ADAM12 and are involved in myofibroblast transdifferentiation and development of fibrosis. *J. Rheumatol.* **2016**, *43*, 1340–1349. [[CrossRef](#)] [[PubMed](#)]
62. Yamazaki, M.; Sugimoto, K.; Mabuchi, Y.; Yamashita, R.; Ichikawa-Tomikawa, N.; Kaneko, T.; Akazawa, C.; Hasegawa, H.; Imura, T.; Chiba, H. Soluble JAM-C ectodomain serves as the niche for adipose-derived stromal/stem cells. *Biomedicines* **2021**, *9*, 278. [[CrossRef](#)] [[PubMed](#)]
63. Bellando-Randone, S.; Del Galdo, F.; Matucci-Cerinic, M. Insights into molecular and clinical characteristics of very early systemic sclerosis. *Curr. Opin. Rheumatol.* 2022, *online ahead of print*. [[CrossRef](#)] [[PubMed](#)]