

Revealing the genomic differences between two subgroups in *Lactobacillus gasseri*

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Being an autochthonous species in humans, *Lactobacillus gasseri* is widely used as a probiotic for fermented products. We thoroughly compared the gene contents of 75 *L. gasseri* genomes and identified two intraspecific groups by the average nucleotide identity (ANI) threshold of 94%. Group I, with 48 strains, possessed 53 group-specific genes including the gassericin T cluster (9 genes) and *N*-acyl homoserine lactone lactonase. Group II, with 27 strains, including the type strain ATCC 33323, possessed group-specific genes with plasmid- or phage-related annotations. The genomic differences provide evidences for demarcating a new probiotic group within *L. gasseri*.

Key words: comparative genomics, *Lactobacillus gasseri*, gassericin T, quorum sensing

Lactobacillus gasseri is an autochthonous species of lactic acid bacteria (LAB) that colonizes in the human gastrointestinal tract, vaginal tract, and oral cavity. Its health benefits, such as its antimicrobial activity and probiotic properties, have been well documented [1], making *L. gasseri* distinct as a probiotic yoghurt inoculum in Japan.

Previously, we reported the existence of two subtypes in *L. gasseri* by using the average nucleotide identity (ANI), the statistical similarity computed from whole genome sequences [2]. An ANI threshold of 95% corresponds to an experimental DNA-DNA hybridization (DDH) value of 70%, which is the general criterion for a species-level difference [3–5]. To reveal genomic characteristics within the two *L. gasseri* groups, we here report detailed analysis of them focusing on their gene contents.

In total, 75 draft genomes of *L. gasseri* were downloaded from our DFAST Archive of Genome Annotation (DAGA), the curated genome repository of *Lactobacillus* and *Pediococcus* from the DDBJ/ENA/GenBank, and Sequence Read Archive (SRA) [2]. They all satisfied a quality rating of ≥ 4 (out of 5) in our database, meaning that their genome completeness is $\geq 95\%$ and contamination level is $\leq 5\%$ as computed by the CheckM software [6]. When ANI values were calculated for all pairs of the obtained genomes by an open-source

Python script (<https://github.com/widowquinn/pyani>), the 75 genomes were cleanly classified into two groups at the 94% threshold: Group I, consisted of 48 strains, and Group II, consisting of the remaining 27 strains including the type strain (ATCC 33323^T, Fig. 1). The genome size ranged from 1.86–2.14 Mb in Group I (average 1.98 Mb) and ranged from 1.78–2.01 Mb in Group II (average 1.89 Mb). The average numbers of protein sequences were 1920 for Group I, 1844 for Group II, and 1893 for both groups (see Supplementary Table for details).

To elucidate the differences between Groups I and II from their gene contents, we computed orthologous gene groups from the overall 141,948 protein sequences using the GET_HOMOLOGUES software (version 1.3) with default settings [7]. Among the 6142 gene groups obtained, 3946 groups contained multiple genes, and 2196 were genome-specific genes, i.e., singletons (Fig. 2). Although the number of genes in each genome was not much different between Group I and Group II, the number of singletons in Group II (27 strains) is far more than in Group I (48 strains). After removing genes of hypothetical/unknown functions, we selected genes whose conservation rates between the two *gasseri* groups differed by more than 80%. The numbers of Group I- and Group II-specific genes became 53 and 46, respectively. For these genes, we manually verified with the Mauve software whether the selected genes form a gene cluster [8] and identified 5 conserved clusters in Group I and 4 conserved clusters in Group II, respectively (Table 1, 2 and Fig. 3). Notably, we observed a highly conserved gassericin T cluster (cluster ID: G1C1 in Table 1) and a putative *N*-acyl homoserine lactone (AHL) lactonase in Group I.

L. gasseri has been reported to produce several bacteriocins,

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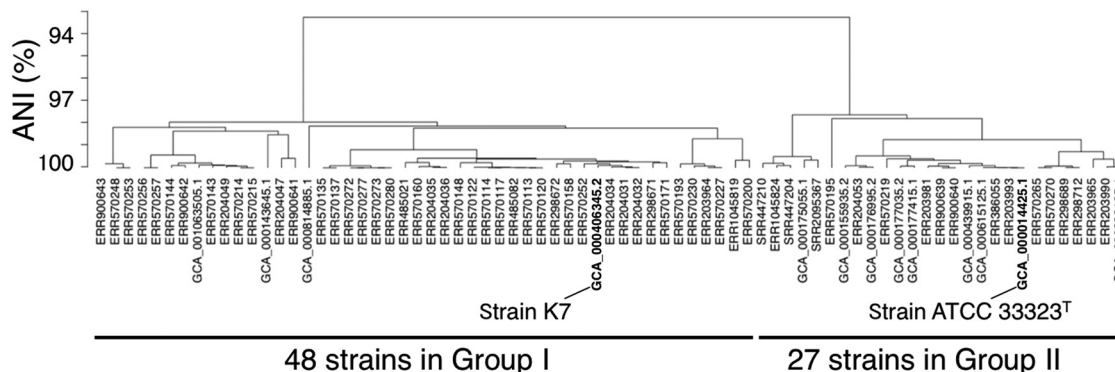


Fig. 1. The results of hierarchical clustering for 75 *L. gasseri* strains.

The genome distance is (1-ANI).

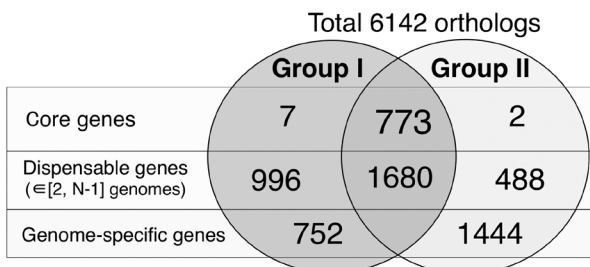


Fig. 2. Venn diagram of the homologous genes in Group I and Group II.

i.e., antimicrobial proteins against its phylogenetic relatives. Gassericin A was reported in *L. gasseri* LA39 [9], and analogs of gassericin T were reported in *L. gasseri* SBT2055 (gassericin T) [10], LA158 (gassericin T) [11], LF221 (gassericin K7 B) [12, 13], and EV1461 (gassericin E) [14]. The former, gassericin A, is a rare Class IIc circular bacteriocin. Only the G256_6_33 strain in Group I (SRA: ERR570193) and 4 strains in Group II (G278_2_12, G278_5_18, G287_2_14, and G287_5_2; SRA: ERR570265, ERR570270, ERR900639, and ERR900640, respectively) possessed its close homologs. On the other hand, gassericin T and its analogs are Class IIb two-peptide bacteriocins and have been widely found within *L. gasseri*. The multiple analogs presumably result from their promiscuous inhibitory spectra that depend on subtle amino acid substitutions or modifications.

The production of gassericin T requires 9 related genes in a 7 kb genomic region in *L. gasseri* LA158 (GenBank: AB710328.1) [11]. In Group I, this region was completely conserved in 39 out of 48 strains. Among the remaining 9 strains, seven conserved the partial region (5.2 kb) and lacked the two gassericin T peptide genes (*gatA* and *gatX*). Only two out of 48 strains, strain G277_2_5 (SRA: ERR570252) and strain 130918 (GenBank: GCA_000814885.1), lacked the whole region. The regional alignment, created by the

Easyfig visualizer (version 2.2.2), is depicted in Fig. 4 [15]. In contrast, all strains in Group II lacked the region except strain G257_1_23 (SRA: ERR570195), which retained a partial region (4.3 kb) without the two peptide genes. In most strains in Group II, however, the two bordering genes, *gatP* and *pedB*, were well conserved (Tables 1, 2). *GatP* is an autoinducer for gassericin T (see later), whereas the function of *pedB* has not been determined, and it is hypothetically annotated as “pediocin immunity protein” in the DAGA database.

Among the 48 Group I strains, an additionally conserved gene was the AHL lactonase related to quorum sensing. It is a communication system of bacteria to coordinate population. Gram-positive bacteria typically use secreted peptides as autoinducers, and *gatP* is known as the autoinducer gene for gassericin T production [14]. On the other hand, Gram-negative bacteria often use small molecules such as AHLs and Autoinducer-2 (AI-2 or furanosyl borate diester) [16]. The AHL lactonase is therefore a quorum-quenching enzyme, hydrolyzing the lactone ring of AHLs. As a member of the metallo-hydrolase superfamily [17], this lactonase has been found in various genera such as *Bacillus* [18], *Agrobacterium* [19], *Rhodococcus* [20], and *Streptomyces* [21].

In Group I, the AHL lactonase was fully conserved in 42 strains, and 4 strains conserved slightly shorter protein sequences. Two strains, G277_2_5 and UMB0099 (SRA: ERR1045819), lacked the gene, and the former strain did not possess the gassericin T cluster either. The amino acid sequence similarity among the 42 Group I strains was 91.1% (256/281 residues). The amino acid similarity between AHL lactonases in *L. gasseri* K7 and *Bacillus* sp. 240B1 (GenBank: AF196486.1) was 23.5% (66/281 residues). The gene was not found in Group II.

The extremely high correlation between the gassericin T gene cluster and the AHL lactonase gene in Group I is noteworthy. The gene cluster is chromosomal (at least in complete genomes), showing a good contrast to the gassericin A gene cluster (4 kb) on a plasmid of the producer strain *L. gasseri* LA39 (JCM11657; the strain was not included in our

Table 1. Group I-specific orthologous gene groups

Cluster ID	Number of genes	Gene names	Conservation (%)	
			Group I	Group II
G1C1	10	lactacin F two-component system inducer peptide precursor (<i>gatP</i>)	97.9	100.0
		histidine kinase (<i>gatK</i>)	95.8	3.7
		two-component system response regulator (<i>gatR</i>)	95.8	0.0
		peptide ABC transporter ATP-binding protein (<i>gatT</i>)	95.8	3.7
		lactacin F transporter auxillary protein (<i>gatC</i>)	95.8	3.7
		bacteriocin (<i>gatA</i>)	81.3	0.0
		bacteriocin (<i>gatX</i>)	81.3	0.0
		lactacin F immunity protein (<i>gatI</i>)	87.5	3.7
		enterocin A immunity protein	95.8	3.7
		pediocin immunity protein PedB	97.9	96.3
G1C2	6	peptidase C45	100.0	0.0
		adenine deaminase	93.8	0.0
		spermidine/putrescine ABC transporter substrate-binding protein	93.8	0.0
		spermidine/putrescine ABC transporter ATP-binding protein	93.8	0.0
		spermidine/putrescine ABC transporter permease protein	93.8	0.0
		amino acid permease	100.0	0.0
G1C3	4	poly(glycerol-phosphate) alpha-glucosyltransferase	89.6	0.0
		accessory Sec system protein Asp2	87.5	0.0
		preprotein translocase subunit SecA	87.5	7.4
		preprotein translocase subunit SecY	83.3	0.0
G1C4	3	arginine/ornithine antiporter	87.5	0.0
		phosphatidylserine decarboxylase	100.0	0.0
		phosphatidylserine decarboxylase	87.5	0.0
G1C5	2	acetolactate synthase large subunit	100.0	3.7
		alpha-acetolactate decarboxylase	100.0	3.7

Gene names follow the output of DAGA annotation, and the cluster ID is our tentative assignment in this Table.

Table 2. Group II-specific orthologous gene groups

Cluster ID	Number of genes	Gene names	Conservation (%)	
			Group I	Group II
G2C1	3	death-on-curing family protein	4.2	100.0
		NADPH-quinone reductase	12.5	92.6
		TetR family transcriptional regulator	2.1	100.0
G2C2	3	integrase	0.0	96.3
		dephospho-CoA kinase	2.1	92.6
		type III restriction protein, res subunit	0.0	81.5
G2C3	3	RelE/StbE family addiction module toxin	0.0	85.2
		DNA-damage-inducible protein J	0.0	88.9
		protein-tyrosine phosphatase	0.0	81.5
G2C4	2	flavoprotein	4.2	100.0
		LysR family transcriptional regulator	4.2	92.6

Gene names follow the output of DAGA annotation.

study due to the absence of the whole genome sequence) [22]. In our study, the gassericin A cluster was found in only one strain in Group I (3 kb partial match in ERR570193) and 4 strains in Group II (a 4 kb complete match in ERR570265 and ERR570270 and a 3.7 kb partial match in ERR900639 and ERR900640). From the draft sequence information, it is hard

to tell whether they are plasmidal or chromosomal. However, Group II strains possess more genome-specific genes, and their conserved clusters also contain plasmid- or phage-related annotations such as “TetR transcriptional regulation,” “integrase,” or “RelE/StbE toxin-antitoxin” (Table 2).

These facts together with the ANI analysis demarcate

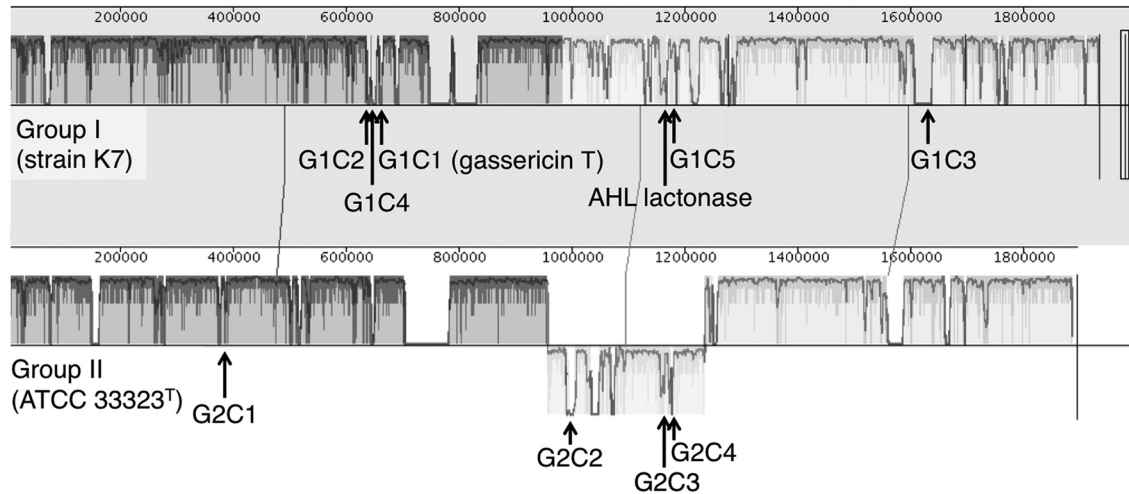


Fig. 3. Genome alignment by the Mauve software between strain K7 (Group I) and strain ATCC 33323^T (Group II). Positions of group-specific clusters are indicated by Cluster IDs (also see Table 1).

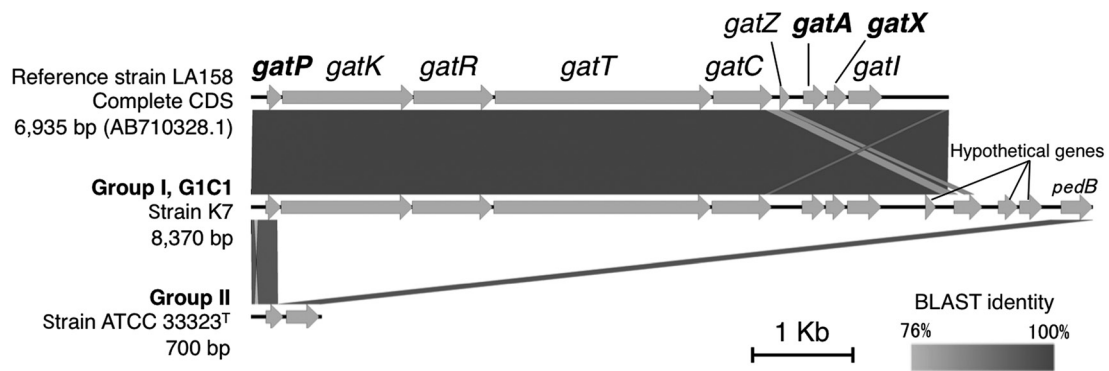


Fig. 4. The structure of the gassericin T cluster. Strain LA158 is used as a reference. See Table 1 for details of the gene functions.

a new probiotic group within *L. gasseri*. To investigate the functionality of the gassericin T cluster and to biochemically characterize Group I strains, we are conducting an investigation with a polyphasic taxonomic approach (in preparation).

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