

Short Report

Open Access

## Circulating microRNAs in breast cancer and healthy subjects

Weizhu Zhu<sup>1</sup>, Wenyi Qin<sup>1</sup>, Ulus Atasoy<sup>2</sup> and Edward R Sauter\*<sup>1</sup>

Address: <sup>1</sup>Department of Surgery, University of North Dakota, Grand Forks, ND, USA and <sup>2</sup>Department of Surgery, University of Missouri, Columbia, MO, USA

Email: Weizhu Zhu - wzhu@medicine.nodak.edu; Wenyi Qin - wqin@medicine.nodak.edu; Ulus Atasoy - atasoy@health.missouri.edu; Edward R Sauter\* - esauter@medicine.nodak.edu

\* Corresponding author

Published: 19 May 2009

Received: 15 April 2009

BMC Research Notes 2009, 2:89 doi:10.1186/1756-0500-2-89

Accepted: 19 May 2009

This article is available from: <http://www.biomedcentral.com/1756-0500/2/89>

© 2009 Sauter et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

### Abstract

**Background:** It has been demonstrated that extracellular mRNA can be detected in the circulation. Our hypothesis was that circulating miRNAs are also present and differentially expressed in the serum of breast cancer patients compared to controls.

**Findings:** We measured miRNA in the serum of samples with and without the addition of miRNA prior to analysis. To test our RNA extraction efficiency, we spiked-in serial dilutions of single-strand C elegans miR-39 (cel-miR-39) and human miR-145 (has-miR-145) into goat serum and a 10 year old human serum specimen. We next analyzed miR-16, -145, and -155 in archived serum specimens from 21 participants, 13 of whom did and 8 of whom did not have breast cancer. We were able to detect the miRNAs from all the serum samples to which the miRNAs had been added. We were also able to detect endogenous miR-16, -145, and -155 in all serum samples. While the expression of all three miRNAs was similar in samples from healthy women compared to those with breast cancer, women with progesterone receptor (PR,  $p = 0.016$ ) positive tumors had higher miR-155 expression than tumors that were negative for these receptors.

**Conclusion:** 1) RNA species can be detected in archived serum; 2) miR-155 may be differentially expressed in the serum of women with hormone sensitive compared to women with hormone insensitive breast cancer. Screening serum for miRNAs that predict the presence of breast cancer is feasible, and may be useful for breast cancer detection.

### Background and research hypothesis

There is increasing evidence supporting microRNA (miRNA) analysis for breast cancer diagnosis, prognosis and therapy [1-3]. miRNAs are ~22 nucleotide long RNA molecules encoded in the genome that can control gene expression. Their expression levels vary greatly among species and tissues [4]. Over 700 miRNAs have been identified from human tissue. Some of these short, endogenous non-protein coding miRNAs are linked to human malignancy and are differentially expressed in human

cancer vs. matched normal tissue [5]. For example, a recent publication identified 29 miRNAs that were aberrantly expressed in primary breast cancer tissue. Of the 29, miR-10b, miR-125b, and miR-145 were down-regulated, while miR-21 and miR-155 were up-regulated [6]. Depending on the genes that the miRNAs act on, they may serve to stimulate or suppress tumor formation and growth. Another team reported 15 miRNAs that are expressed in a variety of normal human tissues. Among them, miR-16 is expressed at a higher level than the

remaining 14 miRNAs [7]. These differences provide the molecular basis and rationale for the identification of miRNAs in breast cancer detection.

Cancer-related RNAs are present and detectable in serum [8-11], and it was recently reported that the levels of cyclin D1 and thymidylate synthase levels of free mRNA in plasma correlated with nonresponse to breast cancer therapy [12]. As RNase is present in the blood of healthy and cancer containing individuals [13], one would expect all circulating RNA (including miRNAs) in the blood to be rapidly destroyed. However, accumulated data suggest that circulating RNA may not be as fragile as was previously assumed. Tsui *et al.* reported that RNA in serum is more RNase resistant and stable than previously thought [14]. For example, tyrosinase mRNA has been found in the serum of melanoma patients but not in healthy individuals [9], and epithelial mRNA in the plasma of breast cancer patients was associated with poor prognosis [15]. Both observations demonstrate that cancer-related mRNA exists in the circulation and is detectable. Finally, a recent pilot study demonstrated that miRNAs can be detected in the serum of ovarian cancer patients [16]. Our hypothesis was that circulating miRNAs are also present and detectable in the serum of breast cancer patients.

**Methods**

**Blood Collection and serum preparation**

After informed consent was obtained from 21 participants (13 with breast cancer and 8 normal controls, Table 1), 6

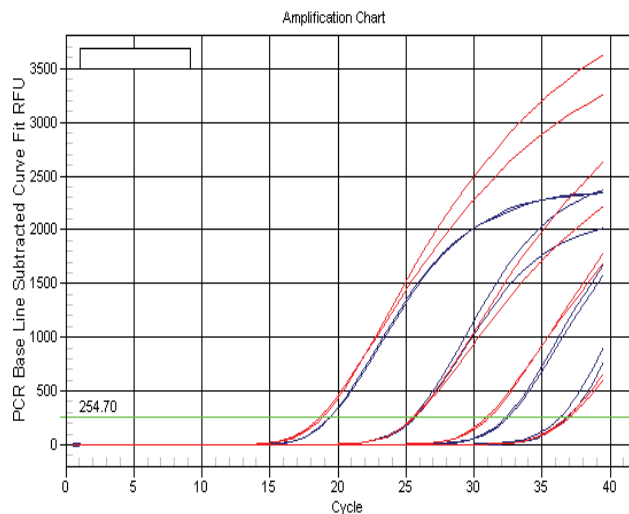
ml of venous blood was collected from the antecubital fossa and placed in a serum separator tube. The blood was centrifuged at 1600 rpm for 5 min and the serum aliquoted into 1.7 ml Eppendorf tubes, followed by a 15 min high speed centrifugation at 12,000 rpm to completely remove cell debris, leaving only circulating RNA. To test our RNA extraction efficiency while minimizing possible interference from endogenous RNA, we used a 10 year old human serum sample and goat serum, to which cel-miR-39 and has-miR-145 were concurrently spiked in.

**RNA Extraction**

We first evaluated our RNA extraction efficiency. Either 10<sup>5</sup>, 10<sup>3</sup>, 10, 10<sup>-1</sup>, or 0 pg of single stranded cel-miR-39 and has-miR-145 synthesized by Invitrogen (Carlsbad, CA 92008) were spiked into 400 µL of both goat serum (Abcam, Cambridge, MA) and 10 year old archived human serum collected from a single subject. We isolated total RNA which included miRNA in 400 µL of serum from 21 participants using the MagMAX™Viral RNA Isolation kit following the manufacturer's instructions (Ambion, Austin, TX). The RNA was stored at -80°C until use. Both MagMAX™Viral RNA Isolation kit and mirVana PARIS kit™ have been recommended to isolate small RNAs in serum by the supplier. We chose the MagMAX kit because it provided better miRNA recovery from the serum samples. We were able to increase our miRNA recovery by 1) extending the incubation and binding time from four to 10 minutes, and 2) increasing the ratio of isopropanol to wash buffer # 1 in the kit from 1:2 to 1:1.

**Table 1: Demographics of Participant Samples**

	Yes	No	Unknown
<b>Breast Cancer</b>	13	8	
<b>Disease Stage</b>			
1	4		
2	5		
3	4		
<b>Estrogen receptor positive</b>	10	2	1
<b>Progesterone receptor positive</b>	9	3	1
<b>HER2/neu positive</b>	1	11	1
<b>Race</b>			
Caucasian	19		
African American	2		



**Figure 1**  
Different quantities (10<sup>5</sup>, 10<sup>3</sup>, 10, 10<sup>-1</sup>, and 0 pg) of cel-miR-39 (blue curves) and has-miR-145 (red curves) inputs led to similar corresponding Ct values of 19.2, 25.8, 32.4, 37.10 and undetectable, respectively.

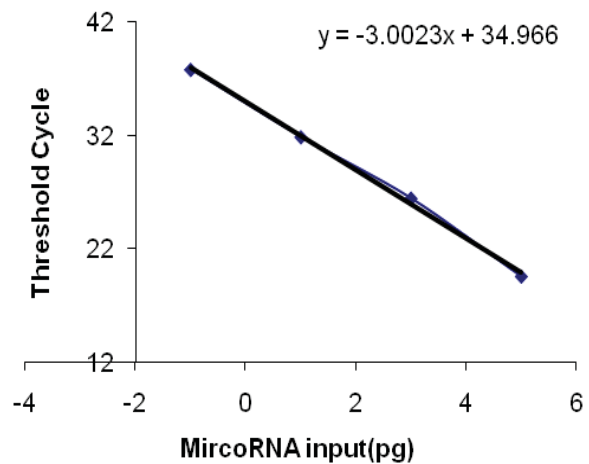
**Reverse Transcription (RT) and Quantitative PCR (qPCR)**

RT and qPCR kits made specifically for accurate miRNA analysis (Applied Biosystems) were used to evaluate expression of the following miRNAs from serum samples: cel-miR-39, has-miR-16, miR-145 and miR-155. 18s rRNA was used to normalize the RNA input. The RT reaction for 18s rRNA was performed with RETROscript® Kit (Ambion, catalog # AM1710). An equal volume of the eluted RNA (5 µL/reaction = 1/10 of the eluted volume) was reverse-transcribed with specific looped RT primers for each miRNA evaluated. The 15 µL RT reactions were performed using a TaqMan® microRNA Reverse Transcription Kit (Applied Biosystems catalog # 4366596) and incubated for 30 min at 16°C, 30 min at 42°C, 5 min at 85°C, and then maintained at 4°C. For real-time PCR, 5 µL RT products were used as templates in 20 µL reactions containing primers and probes for each miRNA and 18s rRNA according to manufacturer instructions. All reactions were run on the MyiQ™ Real-Time PCR Detection System (Bio-Rad, Hercules, CA) using the following conditions: 95°C for 10 min, followed by 40 cycles at 95°C for 15 s, and 60°C for 1 min. The relative miRNA quantity in serum from participants with vs. without breast cancer was determined using the comparative Ct method (Bulletin #2: ABI Prism 7700 Sequence Detection System, Applied Biosystems, 1997).

**Results and discussion**

Early detection is a major factor contributing to the 2.3% annual decline in breast cancer death rates over the past 10 years [17]. Nonetheless 40,480 women in the U.S. were projected to die from breast cancer in 2008 [18], in part because currently available breast cancer screening tools such as mammography and breast examination miss 10–40% of early breast cancers and are least effective in detecting cancer in young women, whose tumors are often more aggressive. An invasive needle or surgical biopsy must be performed when an area of suspicion is identified in order to confirm, by cytologic or histologic evaluation, the presence of malignancy, even though 66–85% of abnormalities are benign [19].

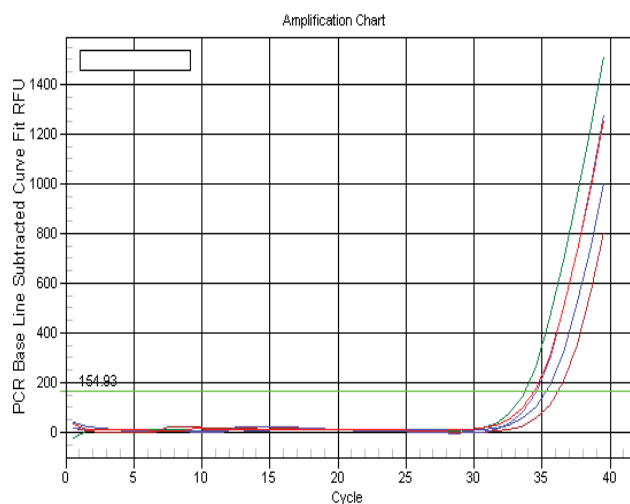
In principle, there are three different ways in which biomarkers can be used. First, they can be used to differentiate normal from diseased states. Second, they can be used to determine into which prognostic groups subjects should be placed. Finally, biomarkers can be used to monitor response to therapy. The identification of biologic markers such as miRNA species collected noninvasively in the circulation that would distinguish between women with or without breast cancer is therefore of crucial importance in early cancer detection, to improve disease free and overall survival from the disease. There is a recent report which outlines the usefulness of miRNA signatures



**Figure 2**  
There was linear recovery (slope = -3.0) of the miR-39 and miR-145 at concentrations equal to and greater than  $10^{-1}$ pg/400 µL serum.

to distinguish normal from diseased tissues and tissue origin, relying upon miRNA signatures from formalin fixed, paraffin embedded and frozen tissue specimens [20], demonstrating the stability of miRNAs. Our miRNA findings in archival serum support and extend these findings, with the potential that miRNA analysis may prove useful for noninvasive breast cancer detection.

We first tested if spiked-in cel-miR-39 and has-miR-145 were detectable in a 10 year old human serum and a goat serum specimen. The specimen was divided into five sam-



**Figure 3**  
The Ct value for endogenous GAPDH mRNA in each of the serial human serum samples to which miR-145 was spiked in was approximately 35.

**Table 2: The Ct Value of miR-16, miR-145 and miR-155 in the Serum Samples with and without Breast Cancer**

Ct value	miR-16		miR-145		miR-155	
	NL <sup>1</sup>	TU <sup>1</sup>	NL	TU	NL	TU
Range	21.57–25.55	21.76–26.14	32.96–36.50	32.62–37.60	33.95–35.84	31.88–38.94
Average	23.75	24.23	34.90	35.51	34.87	34.98
Median	23.79	24.46	34.97	35.35	34.93	34.90
T-test	0.49		0.35		0.87	

1: NL = Normal serum, TU = Tumor serum. 21 serum samples (13 from subjects with breast cancer and eight controls). 18s rRNA was used to normalize the relative expression of miRNA species.

ples of equal volume. Decreasing inputs of single-strand cel-miR-39 and has-miR-145 ( $10^5$ ,  $10^3$ ,  $10$ ,  $10^{-1}$  and  $0$  pg) added to each sample (Figures 1 and 2) led to the corresponding mean Ct values (19.2, 25.8, 32.4, 37.1 and 38.7), respectively, with similar values for cel-39 and has-145. The Ct value of 37.1 obtained using the most dilute sample ( $10^{-1}$  pg) was slightly lower than background of 38.7, indicating that the level of endogenous miR-145 in the human serum to which the synthetic single stranded miR-145 mimic was spiked in was very low, and therefore unlikely to have influenced the amplification of the synthesized miR-145. The equidistant curves down to a concentration of  $10^{-1}$  pg/400  $\mu$ L serum suggest linear recovery (Figure 1) and efficient amplification (Figure 2, slope = -3.0) of the added miRNAs. The Ct values (Figure 3) for endogenous GAPDH mRNA in the serial serum samples to which has-miR-145 was spiked in were approximately 35. The results demonstrate that circulating RNA, including endogenous GAPDH mRNA and spiked-in cel-miR-39 and has-miR-145, can be efficiently extracted and amplified from serum.

We then performed a pilot study in serum specimens from 21 women with and without breast cancer (Table 2) to assess the expression of miR-16, -145 and -155 using 18s rRNA to normalize the relative expression of the miRNAs in archived serum specimens from women with and without breast cancer. Each miRNA and 18s rRNA was isolated and amplified efficiently in all subjects. The expression of all three miRNAs in serum was similar in samples from healthy women compared to those with breast cancer, unlike a prior report which found differences in the expression of mi-R-145 and -155 miRNAs in breast cancer tissue vs. benign tissue women [6]. On the other hand, progesterone receptor (PR,  $p = 0.016$ ) positive tumors had higher miR-155 expression than tumors that were negative for this receptor. miR-155 has been classified as a multifunctional miRNA, having an important role in both normal and pathologic processes in immunity, inflammation, cancer and cardiovascular disease [21]. The signifi-

cance of our finding regarding miR-155 and the PR will require a larger study.

In determining the proper endogenous control for qPCR, we initially tried three small non-coding nucleolar RNAs (Rnu44, Rnu48 and Rnu66). Unfortunately, none were useful because we could not detect them. GAPDH was also tested but relatively high Ct values were observed, so 18s rRNA was tried levels adequate to serve as a control in all human serum samples. We confirmed that our techniques yielded adequate RNA extraction efficiency by measuring levels of spiked-in cel-miR-39 and has-miR-145. The spiked-in miRNAs had the advantage of lacking homology to and thereby interference from endogenous human miRNAs.

### Conclusion

The efficient extraction and accurate identification of miRNAs from serum represents a key first step toward the development of a noninvasive, blood-based detection test for breast cancer, and we believe represents an important advancement in achieving this goal.

### Abbreviations

ER: estrogen receptor; mi-, m- and rRNA: micro-, messenger and ribosomal ribonucleic acid; Rnu: nucleolar RNA; qPCR: quantitative polymerase chain reaction; PR: progesterone receptor; RT: reverse transcription.

### Competing interests

The authors declare that they have no competing interests.

### Authors' contributions

All authors read and approved the final manuscript. WZ and WQ helped conceive the project, designed and performed the experiments, UA helped design the project, provided scientific and technical insight and manuscript review, and ERS provided the samples, helped conceive the project, prepared the manuscript, provided scientific and clinical input, and submitted the manuscript.

## References

1. Calin GA, Croce CM: **MicroRNA signatures in human cancers.** *Nat Rev Cancer* 2006, **6(11)**:857-866.
2. Calin GA, Croce CM: **MicroRNA-cancer connection: the beginning of a new tale.** *Cancer research* 2006, **66(15)**:7390-7394.
3. Jeyaseelan K, Herath VB, Armugam A: **MicroRNAs as therapeutic targets in human diseases.** *Expert Opin Ther Targets* 2007, **11(8)**:1119-1129.
4. Kim J, Krichevsky A, Grad Y, Hayes GD, Kosik KS, Church GM, Ruvkun G: **Identification of many microRNAs that copurify with polyribosomes in mammalian neurons.** *Proceedings of the National Academy of Sciences of the United States of America* 2004, **101(1)**:360-365.
5. Esquela-Kerscher A, Slack FJ: **Oncomirs – microRNAs with a role in cancer.** *Nat Rev Cancer* 2006, **6(4)**:259-269.
6. Iorio MV, Ferracin M, Liu CG, Veronese A, Spizzo R, Sabbioni S, Magri E, Pedriali M, Fabbri M, Campiglio M, et al.: **MicroRNA gene expression deregulation in human breast cancer.** *Cancer Res* 2005, **65(16)**:7065-7070.
7. Liang Y, Ridzon D, Wong L, Chen C: **Characterization of microRNA expression profiles in normal human tissues.** *BMC Genomics* 2007, **8**:166.
8. Lo KW, Lo YM, Leung SF, Tsang YS, Chan LY, Johnson PJ, Hjelm NM, Lee JC, Huang DP: **Analysis of cell-free Epstein-Barr virus associated RNA in the plasma of patients with nasopharyngeal carcinoma.** *Clin Chem* 1999, **45(8 Pt 1)**:1292-1294.
9. Kopreski MS, Benko FA, Kwak LW, Gocke CD: **Detection of tumor messenger RNA in the serum of patients with malignant melanoma.** *Clin Cancer Res* 1999, **5(8)**:1961-1965.
10. Chen XQ, Bonnefoi H, Pelte MF, Lyautey J, Lederrey C, Movarekhi S, Schaeffer P, Mulcahy HE, Meyer P, Stroun M, et al.: **Telomerase RNA as a detection marker in the serum of breast cancer patients.** *Clin Cancer Res* 2000, **6(10)**:3823-3826.
11. Fabian TJ, Dew MA, Pollock BG, Reynolds CF 3rd, Mulsant BH, Butters MA, Zmuda MD, Linares AM, Trottni M, Kroboth PD: **Endogenous concentrations of DHEA and DHEA-S decrease with remission of depression in older adults.** *Biol Psychiatry* 2001, **50(10)**:767-774.
12. Garcia V, Garcia JM, Pena C, Silva J, Dominguez G, Lorenzo Y, Diaz R, Espinosa P, de Sola JG, Cantos B, et al.: **Free circulating mRNA in plasma from breast cancer patients and clinical outcome.** *Cancer letters* 2008, **263(2)**:312-20.
13. Reddi KK, Holland JF: **Elevated serum ribonuclease in patients with pancreatic cancer.** *Proceedings of the National Academy of Sciences of the United States of America* 1976, **73(7)**:2308-2310.
14. Tsui NB, Ng EK, Lo YM: **Stability of endogenous and added RNA in blood specimens, serum, and plasma.** *Clin Chem* 2002, **48(10)**:1647-1653.
15. Silva JM, Dominguez G, Silva J, Garcia JM, Sanchez A, Rodriguez O, Provencio M, Espana P, Bonilla F: **Detection of epithelial messenger RNA in the plasma of breast cancer patients is associated with poor prognosis tumor characteristics.** *Clin Cancer Res* 2001, **7(9)**:2821-2825.
16. Resnick KE, Alder H, Hagan JP, Richardson DL, Croce CM, Cohn DE: **The detection of differentially expressed microRNAs from the serum of ovarian cancer patients using a novel real-time PCR platform.** *Gynecol Oncol* 2009, **112(1)**:55-59.
17. Weir HK, Thun MJ, Hankey BF, Ries LA, Howe HL, Wingo PA, Jemal A, Ward E, Anderson RN, Edwards BK: **Annual report to the nation on the status of cancer, 1975–2000, featuring the uses of surveillance data for cancer prevention and control.** *J Natl Cancer Inst* 2003, **95(17)**:1276-1299.
18. Society AC: **Cancer Facts & Figures 2008.** American Cancer Society Atlanta 2008.
19. Fahy BN, Bold RJ, Schneider PD, Khatri V, Goodnight JE Jr: **Cost-benefit analysis of biopsy methods for suspicious mammographic lesions; discussion 994–5.** *Arch Surg* 2001, **136(9)**:990-994.
20. Rosenfeld N, Aharonov R, Meiri E, Rosenwald S, Spector Y, Zepeniuk M, et al.: **MicroRNAs accurately identify cancer tissue origin.** *Nature Biotechnology* 2008, **26(4)**:462-9.
21. Faraoni I, Antonetti FR, Cardone J, Bonmassar E: **miR-155 gene: A typical multifunctional microRNA.** *Biochim Biophys Acta* 2009 in press.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:  
[http://www.biomedcentral.com/info/publishing\\_adv.asp](http://www.biomedcentral.com/info/publishing_adv.asp)

