

## **Specific pharmacological and $G_{i/o}$ protein responses of some native GPCRs in neurons**

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## Supplementary Tables

**Supplementary Table 1. Potency of the indicated ligands of the GABA<sub>B</sub> receptor in the different neurons and in HEK293 cells using the indicated G<sub>i/o</sub> protein sensors.** Values are mean ± SEM from n independent experiments as indicated.

		pEC <sub>50</sub> / pIC <sub>50</sub> ± SEM (n)						
		CGP64213	GABA	baclofen	SKF 97541	APPA	baclofen + Rac BHFF 1 μM	baclofen + Rac BHFF 10 μM
CGNs	G <sub>i1</sub>	7.55 ± 0.04 (5)	5.03 ± 0.07 (5)	5.32 ± 0.06 (11)	5.63 ± 0.04 (5)	6.13 ± 0.14 (3)	5.56 ± 0.03 (3)	6.19 ± 0.10 (3)
	G <sub>oA</sub>	7.59 ± 0.11 (6)	5.07 ± 0.02 (3)	5.57 ± 0.06 (4)	5.69 ± 0.12 (3)	6.16 ± 0.14 (4)	5.15 ± 0.10 (3)	5.85 ± 0.09 (3)
Cortical neurons	G <sub>i1</sub>		5.58 ± 0.03 (4)	5.95 ± 0.03 (5)	6.52 ± 0.10 (3)			
	G <sub>oA</sub>		5.38 ± 0.09 (3)	5.75 ± 0.08 (4)	6.22 ± 0.13 (3)			
Hippocampal neurons	G <sub>i1</sub>		5.48 ± 0.05 (3)	5.99 ± 0.04 (4)	6.64 ± 0.04 (3)			
	G <sub>oA</sub>		5.15 ± 0.14 (3)	5.58 ± 0.12 (3)	6.16 ± 0.06 (3)			
HEK293 cells transfected GABA <sub>B</sub> R	G <sub>i1</sub>		6.47 ± 0.01 (3)	6.08 ± 0.10 (3)	7.14 ± 0.09 (3)	7.67 ± 0.09 (3)		
	G <sub>oA</sub>		6.45 ± 0.04 (3)	6.07 ± 0.07 (3)	7.07 ± 0.08 (3)	7.77 ± 0.10 (3)		

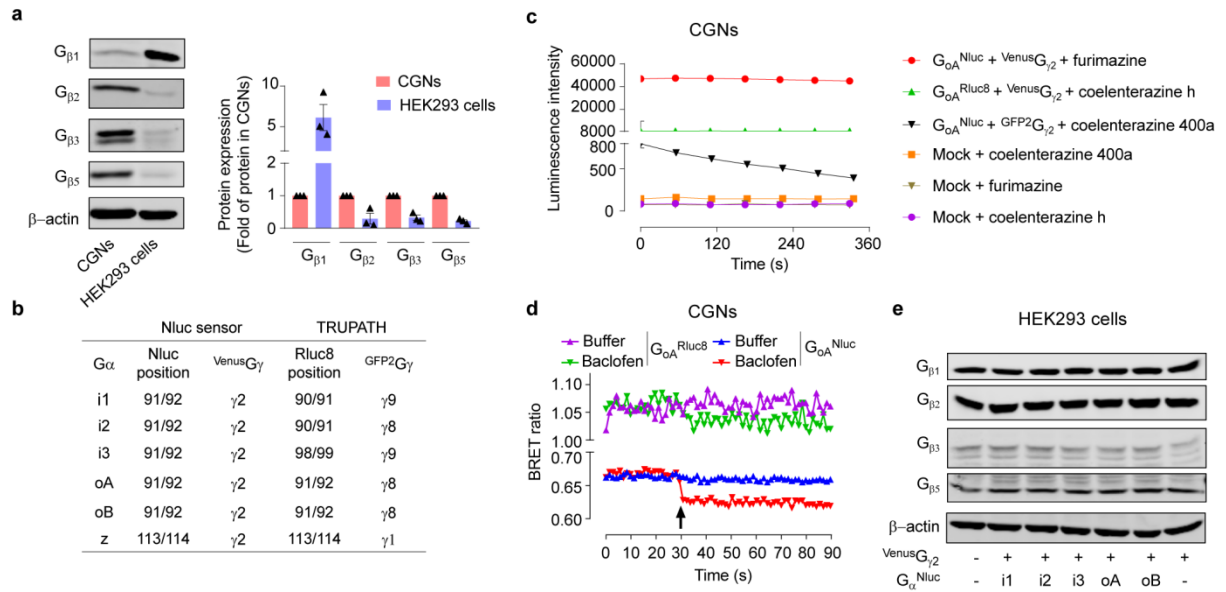
**Supplementary Table 2. Potency and efficacy of the indicated orthosteric ligands on the cannabinoid receptor CB1 in CGNs and HEK293 cells using the indicated  $G_{i/o}$  protein sensors.** Values are mean  $\pm$  SEM from n independent experiments as indicated. Statistical significance of  $pEC_{50}$  and  $E_{max}$  are analysed using one-way analysis of variance (ANOVA) with a Dunnett's *post-hoc* multiple comparison test (compared with CP 55940 group). \*\*\*\* $p < 0.0001$ ; \*\*\* $p < 0.001$ , \*\* $p < 0.01$  and not significant (ns).

		$pEC_{50} \pm SEM$ (n)			$E_{max} \pm SEM$ (n)		
		CP 55940	Win 55,212-2	Bay 59-3074	CP 55940	Win 55,212-2	Bay 59-3074
CGNs	$G_{i1}$	8.18 $\pm$ 0.08 (4)	7.20 $\pm$ 0.17 (4) ***	6.47 $\pm$ 0.11 (4) ****	100	104.8 $\pm$ 3.3 (4) ns	70.7 $\pm$ 1.1 (4) ****
	$G_{oA}$	7.26 $\pm$ 0.13 (4)	6.40 $\pm$ 0.06 (4) ***	6.20 $\pm$ 0.08 (3) ***	100	130.7 $\pm$ 3.3 (4) ****	42.1 $\pm$ 3.9 (3) ****
HEK293 cells transfected CB1	$G_{i1}$	6.90 $\pm$ 0.13 (5)	6.45 $\pm$ 0.21 (5) ns	6.15 $\pm$ 0.18 (4) *	100	95.5 $\pm$ 3.1 (5) ns	50.3 $\pm$ 2.3 (4) ****
	$G_{oA}$	6.78 $\pm$ 0.08 (5)	6.12 $\pm$ 0.11 (4) ****	5.93 $\pm$ 0.06 (4) ****	100	107.7 $\pm$ 9.6 (5) ns	56.3 $\pm$ 4.9 (4) ****

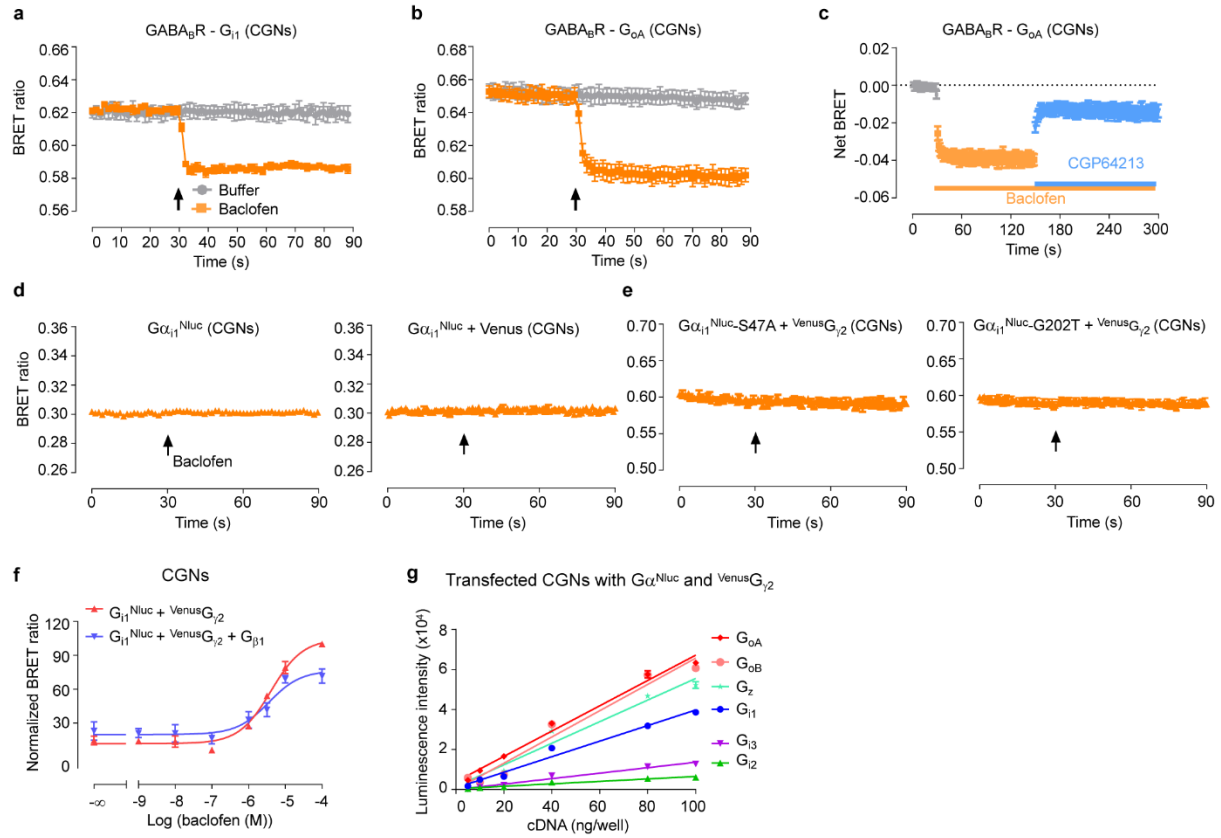
**Supplementary Table 3. G protein response bias analysis of CP 55940 and Bay 59-3074 using Win 55,212-2 as reference (ref.) in CGNs and transfected HEK293 cells.**

	G <sub>i1</sub>					G <sub>oA</sub>					Ligand bias		
	E <sub>max</sub>	EC <sub>50</sub> (M)	E <sub>max</sub> /EC <sub>50</sub>	Log(E <sub>max</sub> /EC <sub>50</sub> )	ΔLog(E <sub>max</sub> /EC <sub>50</sub> )	E <sub>max</sub>	EC <sub>50</sub> (M)	E <sub>max</sub> /EC <sub>50</sub>	Log(E <sub>max</sub> /EC <sub>50</sub> )	ΔLog(E <sub>max</sub> /EC <sub>50</sub> )	ΔΔlog(E <sub>max</sub> /EC <sub>50</sub> )	Bias factor	For
<b>CGNs</b>													
Win 55,212-2	1.00	6.31E-08	1.58E+07	7.20	0.00	1.00	3.98E-07	2.51E+06	6.40	0.00	0.00	1	ref.
CP 55940	0.95	6.61E-09	1.44E+08	8.16	0.96	0.77	5.50E-08	1.39E+07	7.14	0.74	0.22	1.6	G <sub>i1</sub>
Bay 59-3074	0.67	3.39E-07	1.99E+06	6.30	-0.90	0.32	6.31E-07	5.11E+05	5.71	-0.69	0.21	1.6	G <sub>oA</sub>
<b>HEK293 cells</b>													
Win 55,212-2	1.00	3.55E-07	2.82E+06	6.45	0.00	1.00	7.59E-07	1.32E+06	6.12	0.00	0.00	1	ref.
CP 55940	1.05	1.26E-07	8.32E+06	6.92	0.47	0.93	1.66E-07	5.59E+06	6.75	0.63	0.16	1.4	G <sub>oA</sub>
Bay 59-3074	0.53	7.08E-07	7.44E+05	5.87	-0.58	0.52	1.17E-06	4.45E+05	5.65	-0.47	0.11	1.3	G <sub>oA</sub>

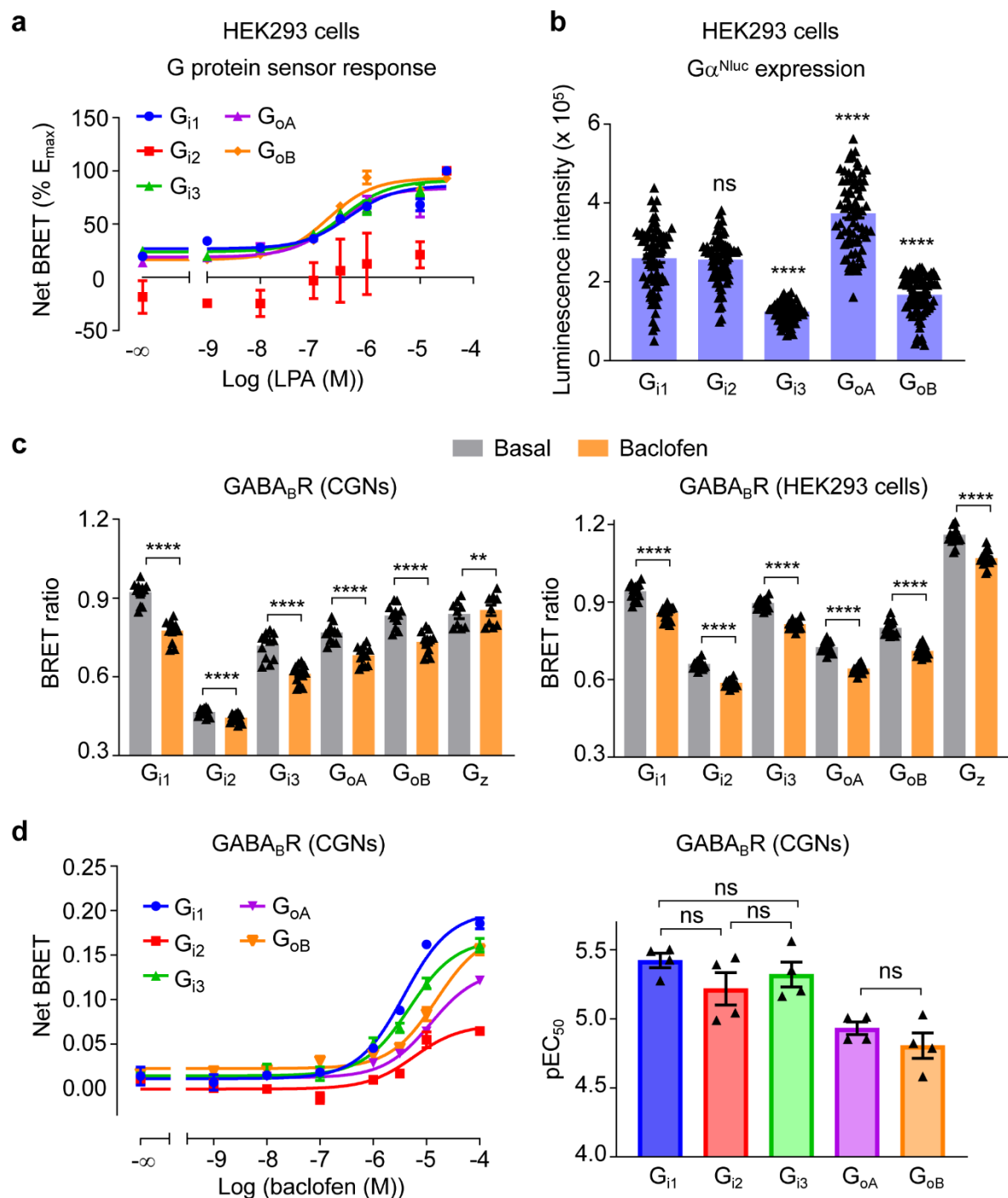
## Supplementary Figures



**Supplementary Fig. 1. Validation of the G $\alpha$  sensors in CGNs.** (a) Detection of endogenous G $\beta$ <sub>1</sub>, G $\beta$ <sub>2</sub>, G $\beta$ <sub>3</sub> and G $\beta$ <sub>5</sub> in CGNs and HEK293 cells by western blotting. Blots are representative from three independent experiments. Values are mean  $\pm$  SEM from three biologically independent experiments. (b) Comparison of the constructs for the Nluc sensors developed in this study with the TRUPATH sensors for the indicated G $\alpha$  sensors. (c) Luminescence intensity measured in CGNs transfected with the indicated G $\alpha$  luciferase constructs and G $\gamma$ <sub>2</sub> fused with indicated fluorescence protein (50 ng each per well in 96-well plate). Coelenterazine 400a (10  $\mu$ M) was added for G $\alpha$ <sup>Rluc8</sup> and GFP2G $\gamma$ <sub>2</sub> (TRUPATH sensor); coelenterazine H (5  $\mu$ M) was added for G $\alpha$ <sup>Rluc8</sup> and VenusG $\gamma$ <sub>2</sub>; furimazine (10  $\mu$ M) was added for G $\alpha$ <sup>Nluc</sup> and VenusG $\gamma$ <sub>2</sub> in CGNs. For these three G $\alpha$  sensors, the signal was measured immediately after addition of these substrates. The effect of these three substrates was also measured in nontransfected CGNs (Mock). Data are representative from two independent experiments. (d) Kinetics of the BRET signal in CGNs co-expressing VenusG $\gamma$ <sub>2</sub> and G $\alpha$ <sup>Nluc</sup> or G $\alpha$ <sup>Rluc8</sup>. Buffer or baclofen (100  $\mu$ M) were injected at the indicated time (arrow). Data are representative from independent experiments (Nluc: n = 4; Rluc8: n = 2). (e) Comparison of the expression of endogenous G $\beta$ <sub>1</sub>, G $\beta$ <sub>2</sub>, G $\beta$ <sub>3</sub> and G $\beta$ <sub>5</sub> in HEK293 cells transfected with or without the indicated G $\alpha$ <sup>Nluc</sup> and VenusG $\gamma$ <sub>2</sub>. Data are representative from three independent experiments. The raw data and p values are available in source data provided as a Source Data file.



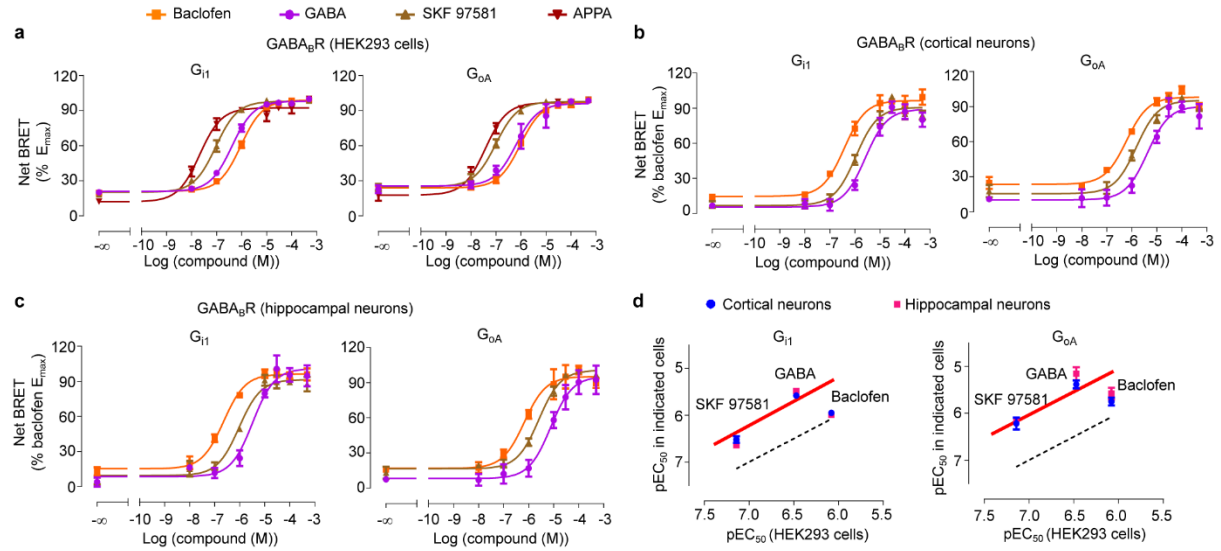
**Supplementary Fig. 2. GABA<sub>B</sub> receptor-induced G<sub>i/o</sub> responses in CGNs.** (a-c) Kinetics of the BRET signal between Gα<sup>Nluc</sup> and VenusG<sub>γ2</sub> in CGNs co-expressing VenusG<sub>γ2</sub> and Gα<sub>i1</sub><sup>Nluc</sup> or Gα<sub>oA</sub><sup>Nluc</sup>. Buffer or baclofen (100 μM) or competitive antagonist CGP64213 (10 μM) were injected at the indicated time (arrow). (d-e) Kinetics of the BRET signal in CGNs expressing G<sub>i1</sub><sup>Nluc</sup> alone, or G<sub>i1</sub><sup>Nluc</sup> and Venus, or G<sub>i1</sub><sup>Nluc</sup>-S47A and VenusG<sub>γ2</sub>, or G<sub>i1</sub><sup>Nluc</sup>-G202T and VenusG<sub>γ2</sub>. Baclofen (100 μM) was injected at the indicated time (arrow). (f) Change of BRET signal between Gα<sub>i1</sub><sup>Nluc</sup> and VenusG<sub>γ2</sub> induced by baclofen, in CGNs co-expressing VenusG<sub>γ2</sub> and Gα<sub>i1</sub><sup>Nluc</sup> (50 ng each/well), or VenusG<sub>γ2</sub>, Gβ<sub>1</sub> and Gα<sub>i1</sub><sup>Nluc</sup> (50 ng each/well). The data presented are as mean ± SEM of the BRET ratios from three independent experiments. (g) Effect of the quantity of transfected cDNA (5, 10, 20, 40, 80 and 100 ng/well in 96-well plate) for the constructs Gα<sub>i1</sub><sup>Nluc</sup>, Gα<sub>i2</sub><sup>Nluc</sup>, Gα<sub>i3</sub><sup>Nluc</sup>, Gα<sub>oA</sub><sup>Nluc</sup>, Gα<sub>oB</sub><sup>Nluc</sup> or Gα<sub>z</sub><sup>Nluc</sup>, together with VenusG<sub>γ2</sub> (50 ng/well) on the luminescence signal of the luciferase at 480 nm in CGNs. Data are representative from at least three independent experiments in (a-e, g). (a), n = 5; (b), n = 4; (c), n = 4; (d), n = 3; (e), n = 3; (g), n = 3. The raw data and p values are available in source data provided as a Source Data file.



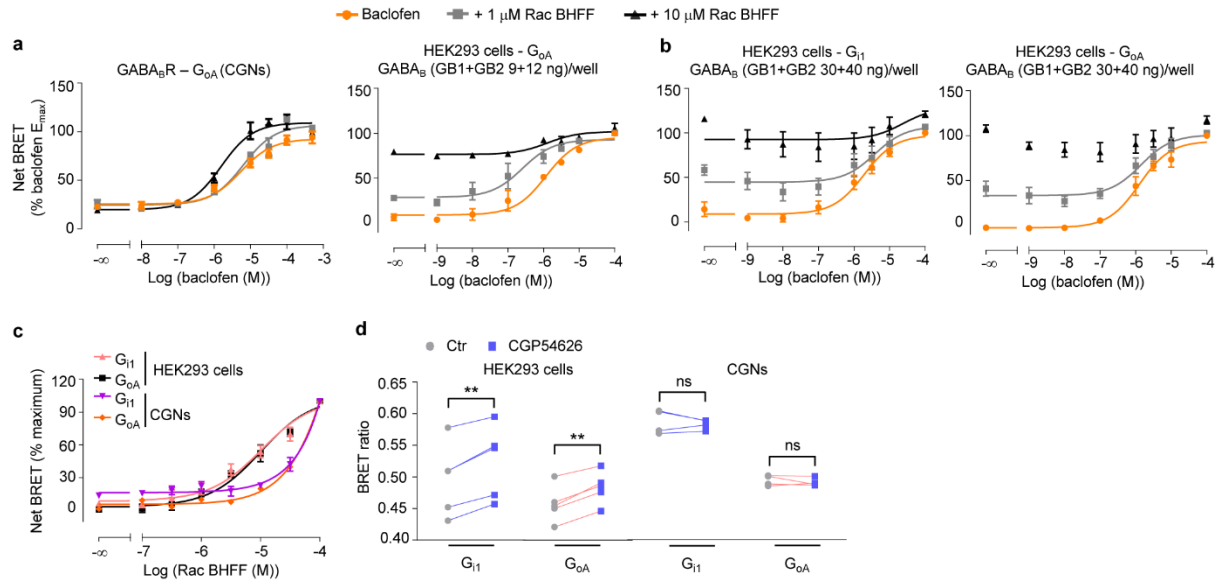
**Supplementary Fig. 3. BRET signal and expression of the different  $G_{i/o}$  protein sensors in HEK293 cells and CGNs.** (a) Change of BRET signal upon different concentration of LPA treatment in HEK293 cells co-transfected with  $Venus^{G\gamma_2}$  and the Nluc-tagged  $G\alpha_{i1}$ ,  $G\alpha_{i2}$ ,  $G\alpha_{i3}$ ,  $G\alpha_{oA}$  or  $G\alpha_{oB}$ . Values are mean  $\pm$  SEM from three biologically independent experiments each performed in triplicate and normalized to maximum response. (b) Expression of the Nluc-tagged  $G\alpha_{i1}$ ,  $G\alpha_{i2}$ ,  $G\alpha_{i3}$ ,  $G\alpha_{oA}$  or  $G\alpha_{oB}$  in (a), measured by the luminescent signal at 480 nm. Values are mean  $\pm$  SEM from three biologically independent experiments each performed in triplicate. The values of individual wells are shown. Data are analysed by using one-way analysis of variance (ANOVA) with a Dunnett's *post-hoc* multiple comparison test to determine significance (compared with  $G_{i1}$  group). \*\*\*\* $p < 0.0001$  and not significant (ns). (c) Change of BRET signal upon baclofen treatment in the cells in Fig. 2a: HEK293 cells co-transfected with  $GB1$ ,  $GB2$ ,  $G\beta_1$ ,  $Venus^{G\gamma_2}$  and

the Nluc-tagged  $G\alpha_{i1}$ ,  $G\alpha_{i2}$ ,  $G\alpha_{i3}$ ,  $G\alpha_{oA}$ ,  $G\alpha_{oB}$  or  $G\alpha_z$ ; CGNs co-transfected with  $^{Venus}G\gamma_2$  and the Nluc-tagged  $G\alpha_{i1}$ ,  $G\alpha_{i2}$ ,  $G\alpha_{i3}$ ,  $G\alpha_{oA}$ ,  $G\alpha_{oB}$  or  $G\alpha_z$ . Values are mean  $\pm$  SEM from four biologically independent experiments each performed triplicate or quadruplicate. The values of individual wells are shown. Data are analysed by using a paired  $t$ -test between basal and baclofen treatment. \*\*\*\* $p < 0.0001$  and \*\* $p < 0.01$ . **(d)** Dose-response and  $pEC_{50}$  of baclofen-induced BRET change between the indicated  $G\alpha_{i/o}^{Nluc}$  and  $^{Venus}G\gamma_2$  in CGNs ( $n = 4$ ). Data are analysed by using one-way analysis of variance (ANOVA) with a Dunnett's *post-hoc* multiple comparison test to determine significance. not significant (ns). The raw data and p values are available in source data provided as a Source Data file.

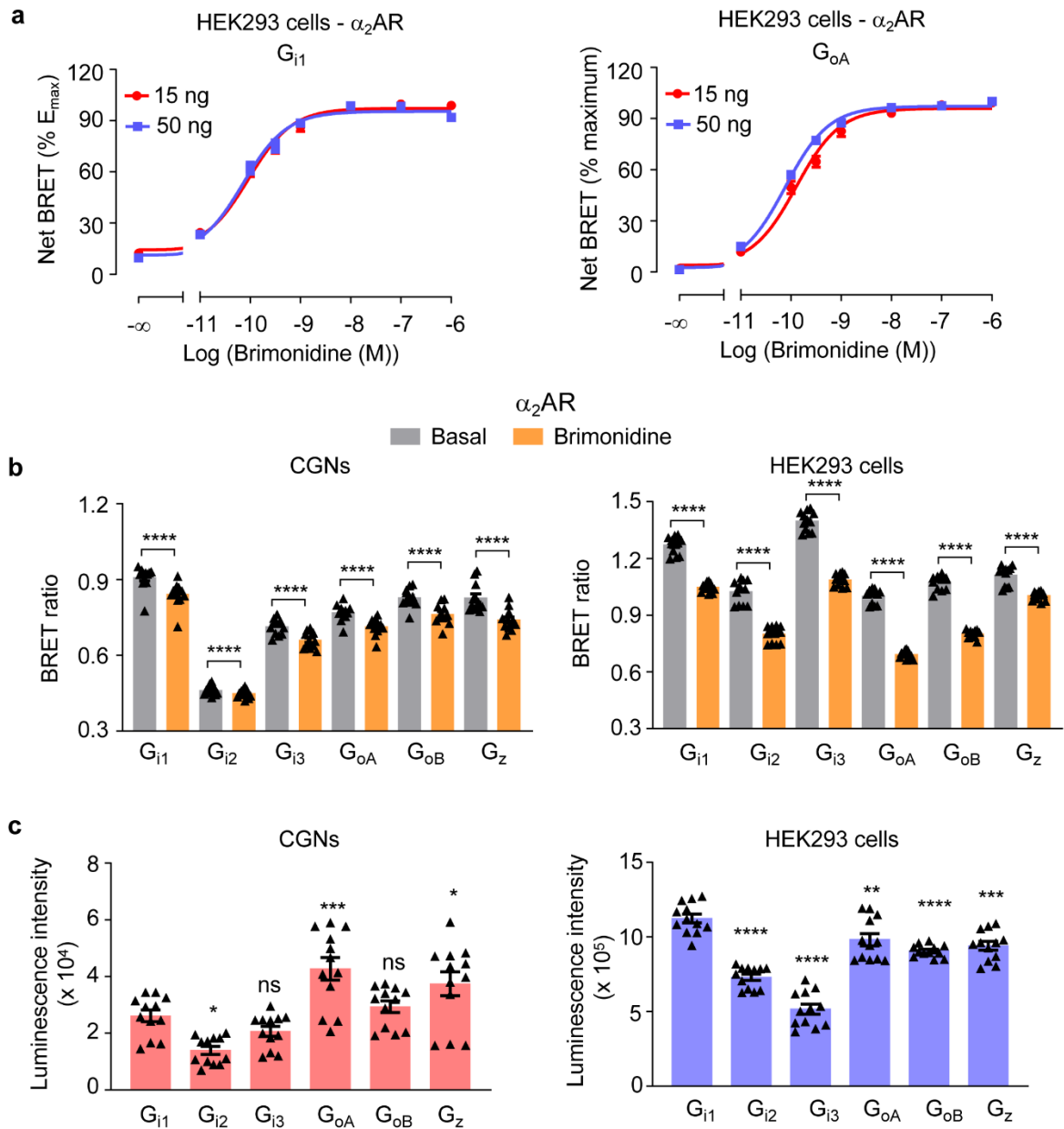




**Supplementary Fig. 4. Ligand potencies of the endogenous GABA<sub>B</sub> receptor in cortical and hippocampal neurons compared to HEK293 cells.** (a) Change of BRET signal between  $\alpha\text{Glu}^{\text{NLuc}}$  and  $\text{Venus}^{\text{G}\gamma_2}$  induced by the indicated agonists of the GABA<sub>B</sub> receptor in HEK293 cells co-transfected with GB1, GB2, G $\beta_1$ ,  $\text{Venus}^{\text{G}\gamma_2}$  and G $\alpha_{i1}^{\text{NLuc}}$  or G $\alpha_{oA}^{\text{NLuc}}$ . (b-c) Change of BRET signal between  $\alpha\text{Glu}^{\text{NLuc}}$  and  $\text{Venus}^{\text{G}\gamma_2}$  induced by the indicated agonists of the GABA<sub>B</sub> receptor in cortical neurons or hippocampal neurons co-transfected with  $\text{Venus}^{\text{G}\gamma_2}$  and G $\alpha_{i1}^{\text{NLuc}}$  or G $\alpha_{oA}^{\text{NLuc}}$ . (d) Correlation of the agonist potencies (pEC<sub>50</sub>) between HEK293 cells and cortical neurons or hippocampal neurons determined by the G<sub>i1</sub> and G<sub>oA</sub> BRET sensors in (a-c). Dotted lines are the correlation of pEC<sub>50</sub> determined with HEK293 cells. Red lines are the fit of pEC<sub>50</sub> in cortical neurons or hippocampal neurons with the same slope as dotted lines. Data are mean  $\pm$  SEM from at least three biologically independent experiments each performed in triplicate, in a-c. (a), baclofen, GABA and SKF 97581, n = 4; APPA, n = 3. (b), G<sub>i1</sub>, n = 4; G<sub>oA</sub>, n = 3. (c), G<sub>i1</sub>, n = 3; G<sub>oA</sub>, n = 3. Data are normalized to maximum baclofen response in (a, b, c). The raw data and p values are available in source data provided as a Source Data file.

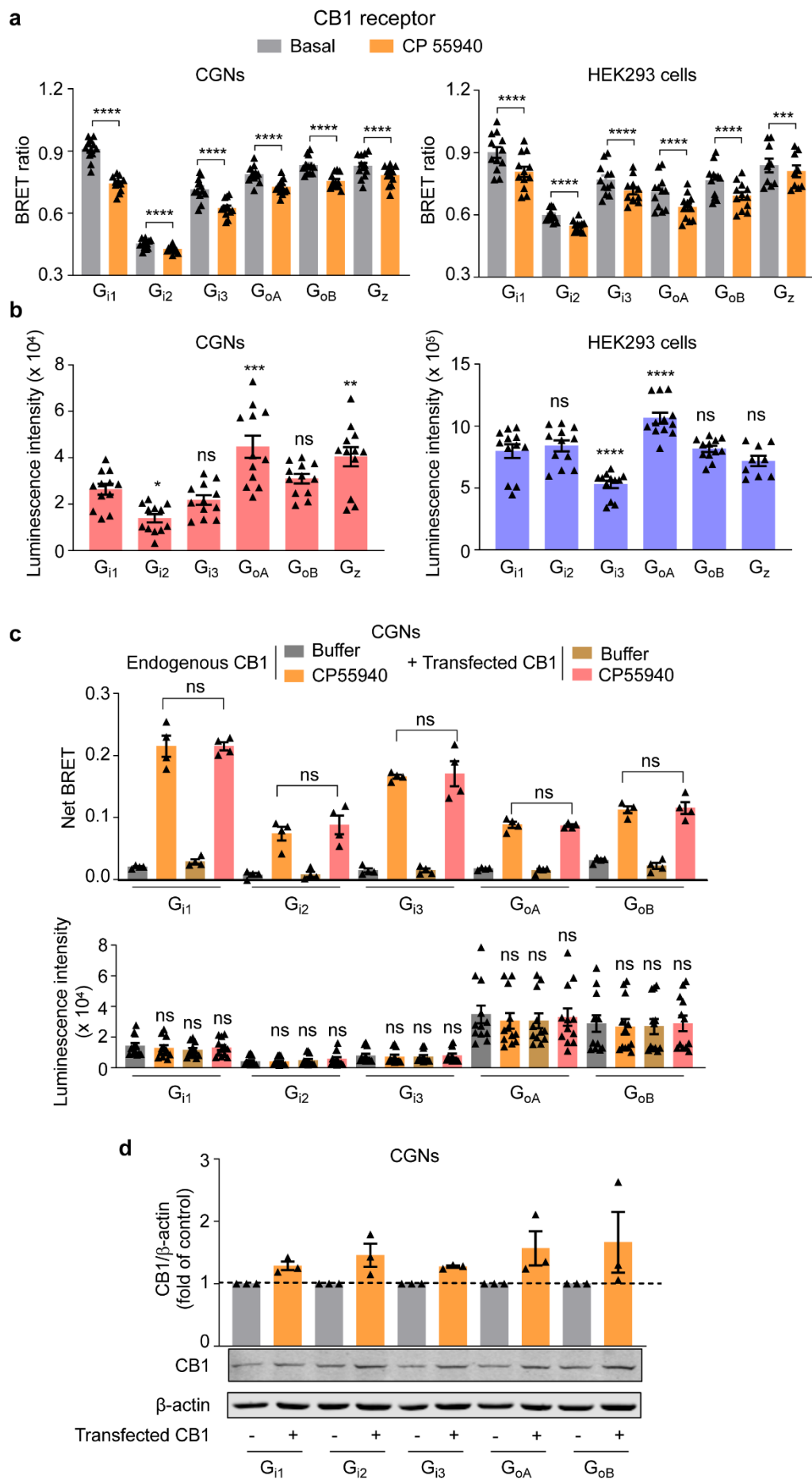


**Supplementary Fig. 5. Effect of PAM Rac BHFF and antagonist CGP54626 on the change in GABA<sub>B</sub> receptor activity in CGNs and HEK293 cells.** (a) The effect of Rac BHFF in baclofen-induced G<sub>OA</sub> response in transfected CGNs and HEK293 cells (GB1 + GB2: 9 ng + 12 ng per well for 96-well plate). (b) The effect of Rac BHFF in baclofen-induced G<sub>I1</sub> and G<sub>OA</sub> response in transfected HEK293 cells (GB1 + GB2: 30 ng + 40 ng per well for 96-well plate). (c) The effect of Rac BHFF alone in G<sub>I1</sub> and G<sub>OA</sub> response in transfected HEK293 cells and CGNs. Data are mean ± SEM from three biologically independent experiments each performed in triplicate, in *a-c*. Data are normalized to maximum baclofen response in (*a*, *b*) and normalized to Rac BHFF maximum response in (*c*). (d) BRET ratio of the G<sub>I1</sub> and G<sub>OA</sub> sensors in absence (control, Ctr) or presence of CGP54626 (50 μM for 30 min) in CGNs co-expressing Venus<sup>Gγ2</sup> and Gα<sub>i1</sub><sup>Nluc</sup> or Gα<sub>OA</sub><sup>Nluc</sup> and in HEK293 cells co-transfected with GB1, GB2, Gβ1, Venus<sup>Gγ2</sup> and Gα<sub>i1</sub><sup>Nluc</sup> or Gα<sub>OA</sub><sup>Nluc</sup>. Data are from biologically independent experiments (CGNs, n = 4; HEK293 cells, n = 5), and analysed using a paired-t test. \*\*p < 0.01 and not significant (ns). The raw data and p values are available in source data provided as a Source Data file.

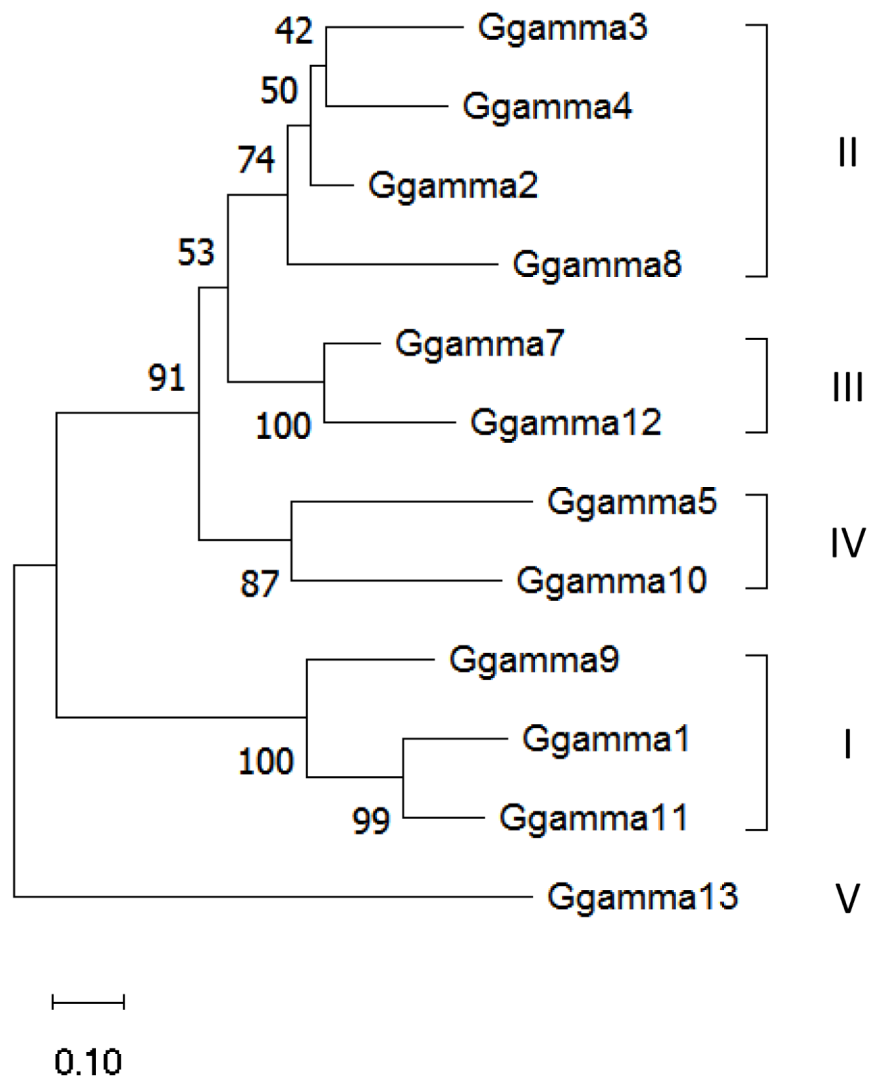


**Supplementary Fig. 6. Change of BRET signal induced by  $\alpha_2$ AR and expression of the different G<sub>i/o</sub> protein sensors in CGNs and HEK293 cells. (a)** Dose-response of brimonidine-induced BRET change for the indicated G<sub>i1</sub> or G<sub>0A</sub> sensors in HEK293 cells co-transfected with mouse  $\alpha_2$ AR (15 ng or 50 ng), G $\beta_1$ , VenusG $\gamma_2$  and G $\alpha_{i1}$ <sup>Nluc</sup> or G $\alpha_{0A}$ <sup>Nluc</sup>, for the indicated amount of  $\alpha_2$ AR cDNA. Values are mean  $\pm$  SEM from four biologically independent experiments each performed in triplicate. **(b)** Change of BRET signal upon brimonidine treatment in the cells in Fig. 5e (CGNs co-transfected with VenusG $\gamma_2$  and the Nluc-tagged G $\alpha_{i1}$ , G $\alpha_{i2}$ , G $\alpha_{i3}$ , G $\alpha_{0A}$ , G $\alpha_{0B}$  or G $\alpha_z$ ; HEK293 cells co-transfected with mouse  $\alpha_2$ AR, G $\beta_1$ , VenusG $\gamma_2$  and the Nluc-tagged G $\alpha_{i1}$ , G $\alpha_{i2}$ , G $\alpha_{i3}$ , G $\alpha_{0A}$ , G $\alpha_{0B}$  or G $\alpha_z$ ). Values are mean  $\pm$  SEM from four biologically independent experiments each performed in triplicate or quadruplicate. Data are analysed by using a paired *t*-test between basal and brimonidine treatment. \*\*\*\**p* < 0.0001. **(c)** Expression of the Nluc-tagged G $\alpha_{i1}$ , G $\alpha_{i2}$ , G $\alpha_{i3}$ , G $\alpha_{0A}$ , G $\alpha_{0B}$  or G $\alpha_z$  in transfected CGNs or HEK293 cells in Fig. 5e, measured by the luminescent signal at 480 nm. Values are mean  $\pm$  SEM from four biologically independent experiments each performed in triplicate or quadruplicate. The values of individual wells are shown. Data are analysed by using one-way analysis of variance (ANOVA) with a Dunnett's *post-hoc* multiple comparison test to

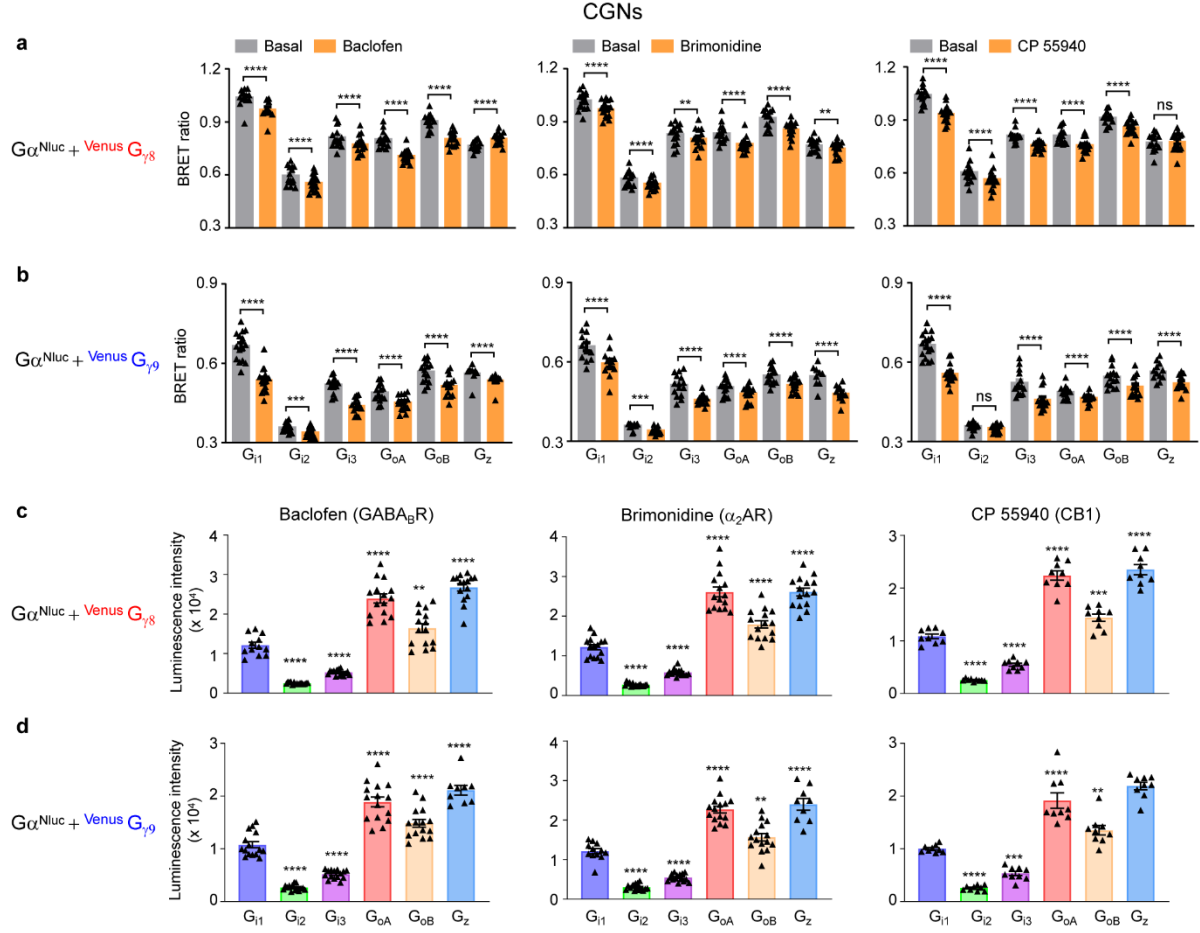
determine significance (compared with G<sub>il</sub> group). \*\*\* $p < 0.001$ , \* $p < 0.05$  and not significant (ns). The raw data and p values are available in source data provided as a Source Data file.



**Supplementary Fig. 7. Change of BRET signal induced by CB1 receptor and expression of the different  $G_{i/o}$  protein sensors in CGNs and HEK293 cells.** (a) Change of BRET signal upon CP 55940 treatment in the cells in Fig. 6b (CGNs co-transfected with  $^{Venus}G\gamma_2$  and the Nluc-tagged  $G\alpha_{i1}$ ,  $G\alpha_{i2}$ ,  $G\alpha_{i3}$ ,  $G\alpha_{oA}$ ,  $G\alpha_{oB}$  or  $G\alpha_z$ ; HEK293 cells co-transfected with mouse CB1,  $G\beta_1$ ,  $^{Venus}G\gamma_2$  and the Nluc-tagged  $G\alpha_{i1}$ ,  $G\alpha_{i2}$ ,  $G\alpha_{i3}$ ,  $G\alpha_{oA}$ ,  $G\alpha_{oB}$  or  $G\alpha_z$ ). Values are mean  $\pm$  SEM from four biologically independent experiments each performed in triplicate. Data are analysed by using a paired *t*-test between basal and CP 55940 treatment. \*\*\*\**p* < 0.0001 and \*\*\**p* < 0.001. (b) Expression of the Nluc-tagged  $G\alpha_{i1}$ ,  $G\alpha_{i2}$ ,  $G\alpha_{i3}$ ,  $G\alpha_{oA}$ ,  $G\alpha_{oB}$  or  $G\alpha_z$  in transfected CGNs and HEK293 cells in Fig. 6b, measured by the luminescent signal at 480 nm. Values are mean  $\pm$  SEM from four biologically independent experiments each performed in triplicate. The values of individual well are shown. Data are analysed by using one-way analysis of variance (ANOVA) with a Dunnett's *post-hoc* multiple comparison test to determine significance (compared with  $G_{i1}$  group). \*\*\*\**p* < 0.0001, \*\*\**p* < 0.001, \*\**p* < 0.01, \**p* < 0.05 and not significant (ns). (c) CP 55940-induced net BRET between  $G\alpha^{Nluc}$  and  $^{Venus}G\gamma$  in CGNs with or without CB1 transfection (50 ng/well), for the indicated  $G_{i/o}$  sensors. CGNs were co-transfected only with indicated  $^{Venus}G\gamma_2$  and the Nluc-tagged  $G\alpha_{i1}$ ,  $G\alpha_{i2}$ ,  $G\alpha_{i3}$ ,  $G\alpha_{oA}$  or  $G\alpha_{oB}$ . Values are mean  $\pm$  SEM from four biologically independent experiments each performed in triplicate. The expression of the indicated Nluc-tagged  $G\alpha_{i/o}$  in these CGNs was measured by the luminescent signal at 480 nm. Values are mean  $\pm$  SEM from three biologically independent experiments each performed in triplicate. The values of individual well are shown. (d) Western blotting detection of CB1 and  $\beta$ -actin in the CGNs in (c). The raw data and *p* values are available in source data provided as a Source Data file.

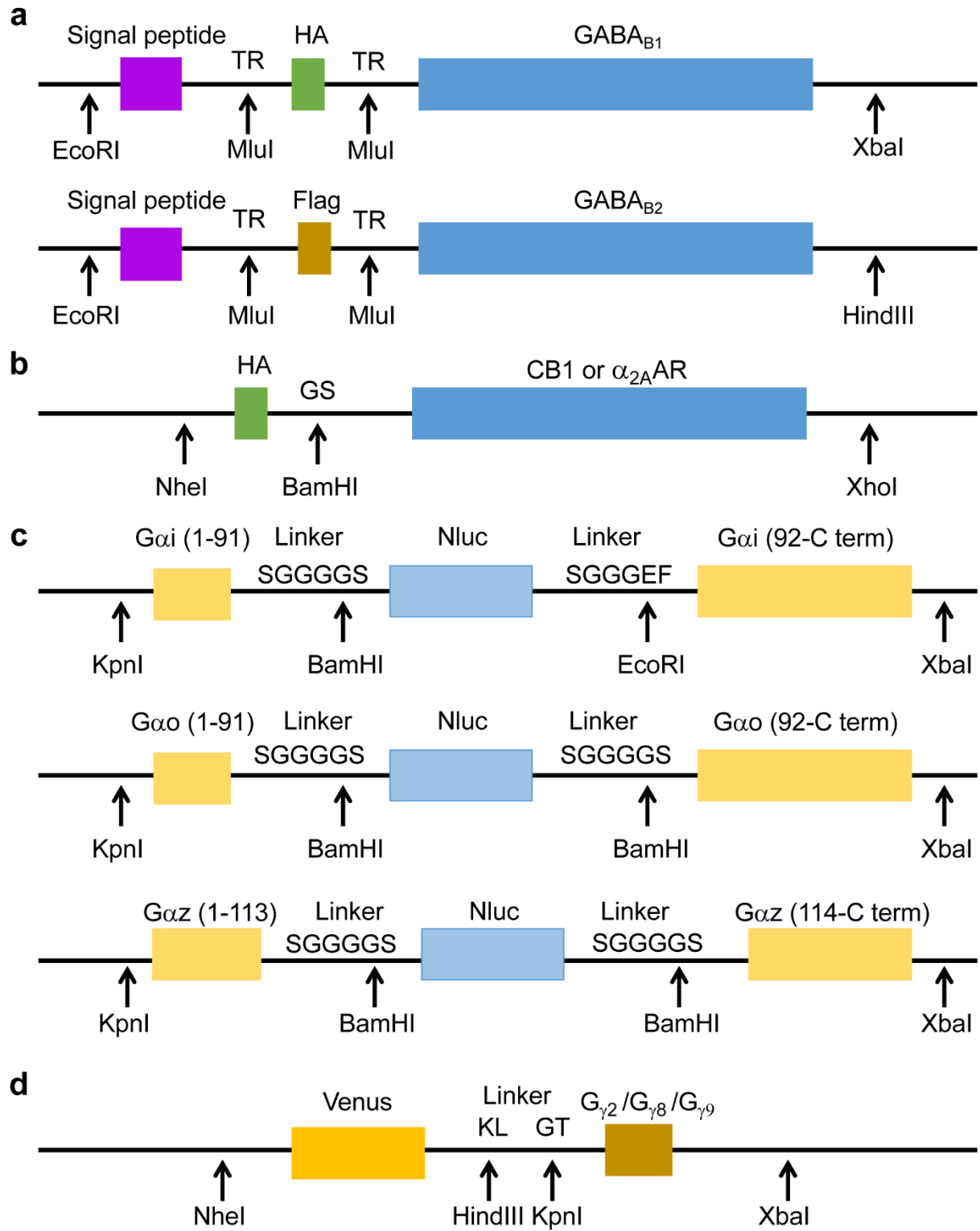


**Supplementary Fig. 8. Phylogenetic trees of human Gγ subunits.** Gγ subunits are classified in I-V groups using MegaX software. The numbers at the nodes of the branches represent the percentage of the bootstrap test (1000 replicates). Scale bar, 0.1 = 10% genetic distance.



**Supplementary Fig. 9. Change of BRET signal induced by the indicated receptors and expression of the different  $G_{i/o}$  protein sensors in CGNs. (a-b)** Change of BRET signal upon baclofen, brimonidine or CP 55940 treatment in CGNs co-transfected with  $Venus G_{\gamma 8}$  or  $Venus G_{\gamma 9}$  together with the Nluc-tagged  $G_{\alpha_{i1}}$ ,  $G_{\alpha_{i2}}$ ,  $G_{\alpha_{i3}}$ ,  $G_{\alpha_{oA}}$ ,  $G_{\alpha_{oB}}$  or  $G_{\alpha_z}$  (referred to Fig. 7a-b). Values are mean  $\pm$  SEM from biologically independent experiments (baclofen and brimonidine,  $n = 5$ ; CP 55940,  $n = 3$ ) each performed in triplicate. Data are analysed by using a paired  $t$ -test between basal and indicated drug treatment. \*\*\*\* $p < 0.0001$ , \*\*\* $p < 0.001$ , \*\* $p < 0.01$  and not significant (ns). **(c-d)** Expression of the Nluc-tagged  $G_{\alpha_{i1}}$ ,  $G_{\alpha_{i2}}$ ,  $G_{\alpha_{i3}}$ ,  $G_{\alpha_{oA}}$ ,  $G_{\alpha_{oB}}$  or  $G_{\alpha_z}$  in transfected CGNs in Fig. 7a-b, measured by the luminescent signal at 480 nm. Values are mean  $\pm$  SEM from biologically independent experiments (baclofen and brimonidine,  $n = 5$ ; CP 55940,  $n = 3$ ) each performed in triplicate. The values of individual well are shown. Data are analysed by using one-way analysis of variance (ANOVA) with a Dunnett's *post-hoc* multiple comparison test to determine significance (compared with  $G_{i1}$  group). \*\*\*\* $p < 0.0001$ , \*\*\* $p < 0.001$  and \*\* $p < 0.01$ . The raw data and  $p$  values are available in source data provided as a Source Data file.





**Supplementary Fig. 10.** Schematic representation of the constructs used **(a)** for GABA<sub>B</sub> receptor (GABA<sub>B1</sub> and GABA<sub>B2</sub> subunits), **(b)** for CB1 or α<sub>2A</sub> AR receptors, **(c)** for Gα<sub>i1</sub><sup>Nluc</sup>, Gα<sub>i2</sub><sup>Nluc</sup>, Gα<sub>i3</sub><sup>Nluc</sup>, Gα<sub>oA</sub><sup>Nluc</sup>, Gα<sub>oB</sub><sup>Nluc</sup> and Gα<sub>z</sub><sup>Nluc</sup>, **(d)** for <sup>Venus</sup>Gγ<sub>2</sub>, <sup>Venus</sup>Gγ<sub>8</sub> and <sup>Venus</sup>Gγ<sub>9</sub>. TR, GS, SGGGS, SGGGEF, KLGT are amino acid linkers.

Rat HA-GB1

MVLLLLILSVLLLKEDVRGSAQSTRYPYDVPDYATRGGAQTPNATSEGCQIIHPPWEGGIRYRGLTRDQVKAI  
NFLPVDYIEIYVCRGEREVVGPVKVRKCLANGSWTDMTPSRCVIRCSKSYLTLENGKVFLTGGDLPALDGA  
RVEFRCDPDFHLVGSSRSVCSQGQWSTPKPHCQVNRTPHSERRAVYIGALFPMSSGGWPGGQACQPAVEMAL  
EDVNSRRDILPDYELKLIHHDSCDPGQATKYLYELLYNDPIKIILMPGCSSVSTLVAEAARMWNILVLSYGS  
SSPALSNRQRFPTFFRTHPSATLHNPTRVKLFKKGWGWKKIATIQQTTEVFTSTLDDLEERVKEAGIEITFRQSFF  
SDPAVPVKNLKRQDARIIVGLFYETEARKVFEVYKERLFGKKYVWFLIGWYADNWFKTYDPSINCTVEEM  
TEAVEGHITTEIVMLNPANTRISISNMTSQEFVEKLTARKLRHPEETGGFQEAPLAYDAIWALALALNKTSGG  
GGRSGVRLEDNFYNNQTITDQIYRAMNSSSFEGVSGHVVDASGSRMAWTLIEQLQGGSYKKIGYYDSTKD  
DLSWSKTDK WIGGSPPADQTLVIKTRFSLSQKLFISVSVLSSLGIVLAVVCLSFNIYNHSHVRYIQNSQPNLNL  
TAVGCSLALAAVFPGLDGYHIGRSQFPFVCQARLWLLGLGFSLGYGSMFTKIWWVHTVFTKKEEKKEWR  
KTLEPWKLYATVGLLVGMDVLTALAIWQIVDPLHRTIETFAKEEPKEDIDVSILPQLEHCSSKKMNTWLGIFYG  
YKGLLLLLGIFLAYETKSVSTEKINDHRAVGMAIYNVAVLCLITAPVTMILSSQQDAAFAFASLAIVFSSYITLV  
VLFVPKMRRLITRGEWQSETQDTMKTGSSTNNNEEEKSRLEKENRELEKIIAEKEERVSELRHQLQSRQQL  
RSRRHPPTPPDPSSGGLPRGPSEPPDRLSCDGSRVHLLYK\*

Rat Flag-GB2

MVLLLLILSVLLLKEDVRGSAQSTRYPYDVPDYATRWTRGAPRPPSSPPLSIMGLMPLTKEVAKGSIGRGV  
LPAVELAIEQIRNESLLRPYFLDLRLYDTECDNAKGLKAFYDAIKYGNHLMVFGGVCPSVTSIIAESLQGWN  
LVQLSFAATTPVLADKKKYPYFFRTVPSDNAVNPAIKLLKHFRWRRVGTLTQDVQRSEVRNDLTGVLYGE  
DIEISDTESFSDNPCTSVKKLKGNVDRIILGQFDQNMMAKVCFCAFEESMFGSKYQWIIPGWYEPAWWEQV  
HVEANSSRCLRRSLLAAMEGYIGVDFEPLSSKQIKTISGKTPQQYEREYNTKRSGVGPSKFHGYAYDGIWVI  
AKTLQRAMETLHASSRHQRIQDFNYTDHTLGKIILNAMNETNFFGVTGQVFRNGERMGTIKFTQFQDSRE  
VKVGEYNAVADTLEIINDTIRFQGSEPPKDKTILEQLRKISLPLYSILSALTILGMIMASAFLEFFNIKNRNQKLI  
KMSSPYMNNLILGGMLSYASIFLFGLDGFSVSEKTFETLCTVRTWILTGYTTAFGAMFAKTWRVHAIFKN  
VKMKKKIHKDQKLLVIVGGMLLIDLCLICWQAVDPLRRTVERYSMEPDAGRDISIRPLEHCENTHMTIWL  
GIVYAYKGLMLFGCFLAWETRNVSIPALNDSKYIGMSVYVNGIMCIIGAAVSFLTRDQPNVQFCIVALVIIFC  
STITLCLVFPVKLITLRTNPDAATQNRRFQFTQNNKKEDSKTSTSVTSVNQASTSRLEGLQSENHRLRMKITE  
LDKDLEEVTMQLQDTPEKTTYIKQNHQYQELNDILSLGNFTSTDDGGKAILKNHLDQNPQLQWNTTEPSRTC  
KDPIEDINSPEHIQRRLSLQLPILHHAYLPSIGGVDASCVSPCVSPTASPRHRHVPPSFRVMVSGL\*

Mouse HA- $\alpha_2A$ AR

MYPYDVPDYAGSFRQEQLAEGSFAPMGSLQPDAGNSSWNGTEAPGGGTRATPYSLQVTLTLVCLAGLLM  
LFTVFGNVLVIIAVFTSRALKAPQNLFLVSLASADILVATLVIPFSLANEVMGYWYFGKVWCEIYLALDVLFC  
TSSIVHLCAISLDYWSITQAEYNLKRTPRRIKAIIVTVWISAVISFPPLISIEKKGGGGQQPAEPSCKINDQ  
KWYVISSISGFFAPCLMILVYVRIYQIAKRRTVPPSRGPVADACSAPPGGADRRPNGLGPERGAGPTGAEE  
PLPTQLNGAPGEPAPAGPRDGDALDLEESSSSSEHAERPPGPRRPRDGPRAKKGKTRASQVKPGDSLPRRGPGA  
AGPGASGSGHGEERGGGAKASRWGRQRNREKRFTFVLAVVIGVFVVCWFPFFFTYTLIAVGCPVPSQLFNFF  
FWFGYCNSSLNPVIYITIFNHDFFRAFKILCRGDRKRIV\*

Mouse HA-CB1

MYPYDVPDYAGSKSILDGLADTTTFTITDILLYVGSNDIQYEDIKGDMSKLGYPQKFPLTSFRGSPFQEK  
MTAGDNSPLVPAGDTTNITEFYNKSLSSFKENEDNIQCGENFMDMECFMILNPSQQLAIAVLSLTGTFTVLE  
NLLVLCVILHSRSLRCRPSYHFIGSLAVADLLGSVIFVYSFVDFHVFHRKDSPNVFLFKLGGVTASFTASVGS  
FLTAIDRYISIRPLAYKRIVTRPKAVVAFCLMWIAIVIAVLPLLGWNCCKLQSVCSDFPLIDETYLFWIGV  
TSVLLLFIVYAYMYILWKAHSHAVRMIQRGTQKSIHHTSEDGKVQVTRPDQARMDIRLAKTLVLVLVLIICW  
GPLLAIMVYDVFGKMNKLIKTVFAFCMLCLLNSTVNPPIIYALRSKDLRHAFRSMFPCSCEGTAQPLDNMMDG  
SDCLHKHANNTASMHRAAESCIKSTVKIAKVTMSVSTDTSAEAL\*

**Supplementary Fig. 11.** Sequences of the constructs used for the rat GABA<sub>B</sub> receptor (GB1 and GB2 subunits), mouse  $\alpha_2A$ AR and mouse CB1 receptor. The amino acid sequence of the signal peptide of GB1 and GB2 (dark red), HA tag of GB1,  $\alpha_2A$ AR and CB1 (red), Flag tag of GB2 (blue), amino acid linkers (green) and receptor (black) are indicated.

G<sub>α1</sub><sup>Nluc</sup>  
 MGCTLSAEDKAAVERSKMIDRNLRDEGEKAAAEVKLLLLGAGESGKSTIVKQMKIIHEAGYSEEECKQYKAVVYSNTIQSIHAIIRAM  
 GRLSGGGGSVFTLEDFVGDWRQTAGYNLDQVLEQGGVSSLFQNLGVSVTPIQRIVLSGENGLKIDIHVIIPYEGLSGDQMGQIEKIFK  
 VVYPVDDHHFKVILHYGTLVIDGVTNPMIDYFGRPYEGIAVFDGKKITVTGTLWNGNKIIDERLINPDGSLFRVTINGVTGWRLCERI  
 LASGGGEFKIDFGDSARADDARQLFVLAGAEEGFMATAELAGVIKRLWKDSGVQACFNRSREYQLNDSAAYYLNDLDRIAQPNYIP  
 TQQDVLRLTRVKTGTGIVETHFTFKDLHFKMFDVGGQRSEKRWIHCFEVGTAIIFCVALSDYDLVLAEDEEMNRMHESMKLFDSCNN  
 KWFTDTSIILFLNKKDLFEEKIKKSPLTICYPEYAGSNTYEEAAAYIQCFEDLNKRKDTKEIYTHFTCATDTKNVQFVFDVAVTDVIIKN  
 NLKDCGLF\*

G<sub>α2</sub><sup>Nluc</sup>  
 MGCTVSAEDKAAAERSKMIDKNLRDEGEKAAAEVKLLLLGAGESGKSTIVKQMKIIHEDGYSEEECRQYRAVVYSNTIQSIMAIVK  
 AMGNLSSGGGSVFTLEDFVGDWRQTAGYNLDQVLEQGGVSSLFQNLGVSVTPIQRIVLSGENGLKIDIHVIIPYEGLSGDQMGQIEKI  
 FKVVYPVDDHHFKVILHYGTLVIDGVTNPMIDYFGRPYEGIAVFDGKKITVTGTLWNGNKIIDERLINPDGSLFRVTINGVTGWRLC  
 ERILASGGGEFKIDFADPSRADDARQLFALSCTAEQGVLPDDLGVIRRLWADHGVQACFGRSREYQLNDSAAYYLNDLERIAQSD  
 YIPTQQDVLRLTRVKTGTGIVETHFTFKDLHFKMFDVGGQRSEKRWIHCFEVGTAIIFCVALSDYDLVLAEDEEMNRMHESMKLFDSC  
 NNKWFTDTSIILFLNKKDLFEEKITHSPLTICFPEYTGANKYDEAASYIQSKFEDLNKRKDTKEIYTHFTCATDTKNVQFVFDVAVTDVII  
 KNNLKDCGLF\*

G<sub>α3</sub><sup>Nluc</sup>  
 MGCTLSAEDKAAVERSKMIDRNLRDEGEKAAAEVKLLLLGAGESGKSTIVKQMKIIHEDGYSEDECKQYKVVVYSNTIQSIHAIIRA  
 MGRLSSGGGSVFTLEDFVGDWRQTAGYNLDQVLEQGGVSSLFQNLGVSVTPIQRIVLSGENGLKIDIHVIIPYEGLSGDQMGQIEKIF  
 KVVYPVDDHHFKVILHYGTLVIDGVTNPMIDYFGRPYEGIAVFDGKKITVTGTLWNGNKIIDERLINPDGSLFRVTINGVTGWRLCE  
 RILASGGGEFKIDFGAARADDARQLFVLAGSAEEGVMPELAGVIKRLWRDGGVQACFSRSREYQLNDSASYYLNDLDRISQSNYI  
 PTQQDVLRLTRVKTGTGIVETHFTFKDLFYKMFVGGQRSEKRWIHCFEVGTAIIFCVALSDYDLVLAEDEEMNRMHESMKLFDSC  
 NNKWFTDTSIILFLNKKDLFEEKIKRSPLTICPEYTGNTYEEAAAYIQCFEDLNRRKDTKEIYTHFTCATDTKNVQFVFDVAVTDVIIK  
 NNLKECGLY\*

G<sub>α0A</sub><sup>Nluc</sup>  
 MGCTLSAEERAALERSKAIEKNLKEDGISAADVKLLLLGAGESGKSTIVKQMKIIHEDGFSGEDVKQYKPVVYSNTIQSLAAIVRA  
 MDTLSSGGGSVFTLEDFVGDWRQTAGYNLDQVLEQGGVSSLFQNLGVSVTPIQRIVLSGENGLKIDIHVIIPYEGLSGDQMGQIEKIF  
 KVVYPVDDHHFKVILHYGTLVIDGVTNPMIDYFGRPYEGIAVFDGKKITVTGTLWNGNKIIDERLINPDGSLFRVTINGVTGWRLCE  
 RILASGGGSIEYGDKERKADAKMVCVSRMEDTEPFSAELL SAMMRLWGDGSGIQECFNRSREYQLNDSAKYYLDSLDRIGAA  
 DYQPTQDILRLTRVKTGTGIVETHFTFKNLHFRLFDVGGQRSEKRWIHCFEVGTAIIFCVALSGYDQVLHEDETTNRMHESMLFDSI  
 CNKFFIDTSIILFLNKKDLFEEKIKKSPLTICFPEYTGPNYEDAAAYIQAQFESKNRSPNKEIYCHMTCATDTNNIQVVFDAVTDIIIA  
 NNLRGCGLY\*

G<sub>α0B</sub><sup>Nluc</sup>  
 MGCTLSAEERAALERSKAIEKNLKEDGISAADVKLLLLGAGESGKSTIVKQMKIIHEDGFSGEDVKQYKPVVYSNTIQSLAAIVRA  
 MDTLSSGGGSVFTLEDFVGDWRQTAGYNLDQVLEQGGVSSLFQNLGVSVTPIQRIVLSGENGLKIDIHVIIPYEGLSGDQMGQIEKIF  
 KVVYPVDDHHFKVILHYGTLVIDGVTNPMIDYFGRPYEGIAVFDGKKITVTGTLWNGNKIIDERLINPDGSLFRVTINGVTGWRLCE  
 RILASGGGSIEYGDKERKADAKMVCVSRMEDTEPFSAELL SAMMRLWGDGSGIQECFNRSREYQLNDSAKYYLDSLDRIGAA  
 DYQPTQDILRLTRVKTGTGIVETHFTFKNLHFRLFDVGGQRSEKRWIHCFEVGTAIIFCVALSGYDQVLHEDETTNRMHESMLFDSI  
 NNKWFTDTSIILFLNKKDLFEEKIKKSPLTICFPEYTGSAFTEAVAYIQAQYESKNKSAHKEIYTHVTCATDTNNIQVVFDAVTDVIIAK  
 NLRGCGLY\*

G<sub>αz</sub><sup>Nluc</sup>  
 MGCQRSSSEEKAAARRSRIDRLRSESQRQRREIKLLLLGTSNSGKSTIVKQMKIIHSGGFNLEACKEYKPLIYNDAISLTRIIRALAA  
 LRIDFHNPDRAVDVQLFALTGPSGGGSVFTLEDFVGDWRQTAGYNLDQVLEQGGVSSLFQNLGVSVTPIQRIVLSGENGLKIDIHV  
 IIPYEGLSGDQMGQIEKIFKVVYPVDDHHFKVILHYGTLVIDGVTNPMIDYFGRPYEGIAVFDGKKITVTGTLWNGNKIIDERLINPDG  
 SLLFRVTINGVTGWRLCERILASGGGS AASKGEITPELLGVMRRLWADPGAQACFSRSSEYHLEDNAAYLNDLERIAAADYIPTVE  
 DILRSRDMTTGIVENKFTFKELTFKMVDVGGQRSEKRWIHCFEVGTAIIFCVELSGYDLKLYEDNQTSRMAESRLFDSCNNNWF  
 NTSILFLNKKDLLAEKIRRIPLTICFPEYKGQNTYEEAAVYIQRQFEDLNRRKETKEIYSHFTCATDTSNIQVVFDAVTDVIIQNNLKYI  
 GLC\*

**Supplementary Fig. 12.** Sequences of the constructs used for human G<sub>α1</sub><sup>Nluc</sup>, G<sub>α2</sub><sup>Nluc</sup>, G<sub>α3</sub><sup>Nluc</sup>, G<sub>α0A</sub><sup>Nluc</sup>, G<sub>α0B</sub><sup>Nluc</sup> and G<sub>αz</sub><sup>Nluc</sup>. The G<sub>α</sub> proteins (black), amino acid linkers (red) and Nluc (blue) are indicated.

VenusG<sub>γ2</sub>

MVSKGEELFTGVVPILVELDGDVNGHKFSVSGEGEGDATYGKLTCLKICTTGKLPVPWPTLVTTGLGYGLQCF  
ARYPDHMKQHDFFKSAMPEGYVQERTIFFKDDGNYKTRAEVKFEGDTLVNRIELKGIDFKEDGNILGHKLE  
YNYNSHNVYITADKQKNGIKANFKIRHNIEDGGVQLADHYQQNTPIGDGPVLLPDNHYLSYQSALS KDPNE  
KRDHMLVLEFVTAAGITLGMDELYK~~KLGT~~MASNNTASIAQARKLVEQLKMEANIDRIKVS KAAADLMAYC  
EAHAKEDPLLTPVPAENPFREKKFFCAIL\*

VenusG<sub>γ8</sub>

MVSKGEELFTGVVPILVELDGDVNGHKFSVSGEGEGDATYGKLTCLKICTTGKLPVPWPTLVTTGLGYGLQCF  
ARYPDHMKQHDFFKSAMPEGYVQERTIFFKDDGNYKTRAEVKFEGDTLVNRIELKGIDFKEDGNILGHKLE  
YNYNSHNVYITADKQKNGIKANFKIRHNIEDGGVQLADHYQQNTPIGDGPVLLPDNHYLSYQSALS KDPNE  
KRDHMLVLEFVTAAGITLGMDELYK~~KLGT~~TMSNNMAKIAEARKTVEQLKLEVNIDRMKVS QAAAELLAFC  
ETHAKDDPLVTPVPAENPF RDKRLFCVLL\*

VenusG<sub>γ9</sub>

MVSKGEELFTGVVPILVELDGDVNGHKFSVSGEGEGDATYGKLTCLKICTTGKLPVPWPTLVTTGLGYGLQCF  
ARYPDHMKQHDFFKSAMPEGYVQERTIFFKDDGNYKTRAEVKFEGDTLVNRIELKGIDFKEDGNILGHKLE  
YNYNSHNVYITADKQKNGIKANFKIRHNIEDGGVQLADHYQQNTPIGDGPVLLPDNHYLSYQSALS KDPNE  
KRDHMLVLEFVTAAGITLGMDELYK~~KLGT~~MAQDLSEKDLLKMEVEQLKKEVKNT RIPISKAGKEIKEYVE  
AQAGNDPFLKGIPEDKNPFKEKGGCLIS\*

**Supplementary Fig. 13.** Sequences of the constructs used for human VenusG<sub>γ2</sub>, VenusG<sub>γ8</sub> and VenusG<sub>γ9</sub>. The Venus (orange), amino acid linker (red) and Gγ subunit (purple) are indicated.