

Intracerebral Hemorrhage: The Effects of Aging on Brain Injury

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Intracerebral hemorrhage (ICH) is a devastating subtype of stroke with high rates of mortality and morbidity. ICH patients often suffer devastating and debilitating neurological impairments, from which the majority of victims are unable to fully recover to functional independence. Unfortunately, there is no established medical therapy for ICH, which is partly attributed to the lack of understanding of the complex pathology of the disorder. Despite advanced age being a major risk factor of ICH, most preclinical studies on ICH employed young animal subjects. Due to this discrepancy, the molecular level changes in the aging brain after ICH are largely unknown, limiting the translation of preclinical studies into potential human treatments. The purpose of this review is to highlight the effects of advanced age on ICH- induced brain injury and recovery and to draw attention to current knowledge gaps, which warrant further investigation.

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INTRODUCTION

Intracerebral hemorrhage (ICH) is the second most common form of stroke, caused by blood vessel rupture and subsequent bleeding into the surrounding brain tissue (Qureshi et al., 2009). ICH accounts for 10–20% of stroke cases worldwide (Feigin et al., 2009; Sacco et al., 2009), with incidence varying across different countries and ethnicities. For instance, its prevalence is much higher in low-middle income countries, which have a higher proportion of fatal cases (Feigin et al., 2009). The incidence of ICH in African Americans is twice as high compared to white Americans (Flaherty et al., 2005). Notably, the worldwide incidence of ICH has risen by \sim 47% over the last 20 years (An et al., 2017), and hospital admissions have increased by 18% in the past 10 years (Qureshi et al., 2007). Moreover, the United States population is aging at an unprecedented pace and the fastest-growing age group in the United States is those over the age of 65 (Wasil and Lichtman, 2005). It is projected that by 2030, 20% of the United States population will be over the age of 65, compared to 2010, when this demographic only accounted for 13% of the population (Albright et al., 2016). As the elderly population continues to grow, the prevalence of ICH could rise alongside it since advanced age is a major risk factor of ICH. By 2030, nearly 4% of the United States population is estimated to have had a stroke (Ovbiagele et al., 2013).

Intracerebral hemorrhage imposes a significant economic burden on society, contributing to an estimated \$17.2 billion in annual direct costs to the U.S. healthcare system associated with stroke

Abbreviations: BMDM, brain-infiltrating monocyte-derived macrophage; CAA, cerebral amyloid angiopathy; CMB, cerebral microbleeds; CMH, cerebral microbemorrhages; ICH, intracerebral hemorrhage; IL-10, interleukin-10; IL-13, interleukin-13; IL-16, interleukin-16; IL-4, interleukin-4; IL-6, interleukin-6; IL-8, interleukin-8; IL-33, interleukin-33; LDL-C, LDL Cholestero; LPS, lipopolysaccharide; MAC, membrane attack complexes; MMP-9, matrix mettaloproteinase-9; MMP, matrix metalloproteinase; TGF- β , transforming growth factor beta; TLR4, toll like receptor 4; TNF- α , tumor necrosis factor alpha.

(Taylor et al., 1996; Caro and Huybrechts, 1999; Reed et al., 2001; Mozaffarian et al., 2015). The 30-day mortality rate of patients suffering from ICH is high, about 40%, with only around 20% of survivors achieving functional independence 6 months after the onset of brain hemorrhage (Bonsack and Alleyne, 2016). A critical barrier to improve patient outcomes after ICH is the lack of effective treatment, partly attributed to the complexity and the poorly defined pathophysiology of the disorder. Though surgical approaches are offered for selected patients, elderly patients, which comprise approximately one-third of the ICH population, are less likely to undergo surgical treatment, perhaps due to established end-of-life preferences. Taken together, the current treatment option for ICH, even in dedicated stroke centers, is limited to supportive care.

The risk factors of ICH include hypertension, cerebral amyloid angiopathy, advanced age, cigarette smoking, diabetes, alcohol abuse, drug abuse, Asian ethnicity, genetic factors, menopause, and oral anticoagulant treatment (Feldmann et al., 2005; Morotti and Goldstein, 2016). Also, there was a significant association between lower LDL-C (Low-Density Lipoprotein-Cholesterol; <70 mg/dL) and higher risk of ICH (Ma et al., 2019). However, a recent meta-analysis reported that lipid-lowering therapy was not associated with a significantly increased risk of ICH in primary prevention trials (Judge et al., 2019), while a more recent meta-analysis concluded that lipid-lowering therapy increased the risk of ICH, but only at high doses that achieved \geq 35% reduction in LDL-cholesterol levels (Cheng et al., 2020). Due to these discrepancies, further studies are highly required to determine the precise role of LDL-C in the pathogenesis of ICH. The most common risk factor of ICH is hypertension (Morotti and Goldstein, 2016), which accounts for approximately 83% of ICH cases (Cheung and Zou, 2003). Hypertension nearly doubles the risk of ICH (Jackson and Sudlow, 2006) and usually leads to ruptures of vessels at the bifurcation of small arteries within the brain (Qureshi et al., 2009). It is believed that long-standing elevated blood pressure promotes shear stress and degenerative changes to the walls of small-tomedium penetrating vessels causing vascular ruptures and ICH (Qureshi et al., 2009). Hypertension is also a key contributor to aneurysmal rupture leading to ICH (van Asch et al., 2010b; Jabbarli et al., 2016) and hypertensive ICH occurs mostly in deep brain structures (Matsukawa et al., 2012). Another risk factor that accounts for \sim 20% of ICH cases is cerebral amyloid angiopathy (CAA), which is prevalent in elderly patients (Morotti and Goldstein, 2016; An et al., 2017). CAA is manifested by the deposition of amyloid-ß peptides in small-to-mediumsized arteries and arterioles in the cortex and leptomeninges, resulting in lobar hemorrhage (Matsukawa et al., 2012). CAA accounts for approximately 12-15% of lobar ICH in the elderly (Itoh et al., 1993; Mehndiratta et al., 2012), and the risk of developing sporadic CAA-related ICH increases in carriers of the $\varepsilon 4$ and $\varepsilon 2$ alleles of apolipoprotein E, the monogenic risk factor of ICH (Nicoll et al., 1997; Sudlow et al., 2006; Talha et al., 2020).

As a person gets older, the risk of developing ICH increases. Elderly individuals have a fivefold higher risk of ICH as opposed to their younger counterparts (van Asch et al., 2010a). Age enhances the risk of chronic health conditions and systemic conditions such as hypertension, diabetes and atrial fibrillation (Ariesen et al., 2003), which can contribute to the pathophysiology of ICH. Elderly patients make up a significant portion of the ICH population, and as per a recent study $\sim 34\%$ of ICH patients were 80 years or older (Stein et al., 2012). Studies have shown a steady increase of ICH cases per 100,000 individuals from 5.9 in 35-54 year-olds to 176.3 in 75-94 yearolds (Wasil and Lichtman, 2005). Age increases not only the prevalence of ICH exponentially (Broderick et al., 1993), but also the 30-day mortality rate (Gonzalez-Perez et al., 2013). The mortality rate in men raised from 23% in ICH patients under 75 years of age to 41% in those over 75 (Zia et al., 2009). Furthermore, age is an independent predictor of poor functional outcomes when measured via total Functional Independence Measure (FIM) score and Motor FIM score (Bagg et al., 2002) and age (>65 years) is an independent predictor for recurrent ICH (Zia et al., 2009). The sex differences in outcomes have not been fully characterized in the pathophysiology of ICH. For younger patients, female sex was protective, but at age >60 years, female sex was a risk factor for death or discharge to hospice (Umeano et al., 2013; Craen et al., 2019). The mortality rate of ICH increased from 20% in female patients under 75 to 26% in those over 75 (Zia et al., 2009). However, the role of aging in the pathophysiology of ICH remains largely understudied. The lack of preclinical studies limits our understanding of the intricate molecular mechanisms of ICH-induced brain injury and the translation of preclinical studies into potential human treatments. Although preclinical animal models of ICH are potent tools for characterizing the disease pathology, most ICH research has employed young animal subjects. This discrepancy may be in part due to the increased amount of time and cost that need to be invested for aging-related studies and the limited commercial availability of aged animals. Given that the elderly population accounts for approximately one-third of ICH patients coupled with the possible increase in prevalence of ICH, herein we provide an overview of the multifactorial effects of aging in the pathophysiology of ICH and identify the knowledge gap, which could help develop new research avenues to improve the prognosis of ICH patients.

Cerebrovascular Circulation and Intracerebral Hemorrhage

Aging is an intricate phenomenon and there are numerous effects of aging on the body. Age-related changes in the cerebral vasculature include vascular stiffness, decreased vascular density, thickening of the vessel wall, endothelial dysfunction, and increased blood-brain barrier permeability (Xu et al., 2017). Overall, these age-induced changes to the vasculature can make the brain parenchyma more susceptible to ICH-induced brain damage apart from enhancing the risk of ICH. Consistently, aged rats (18-months) exhibited significantly higher neurobehavioral deficits after ICH than young rats (3-months) and this was coupled with augmented brain edema in the aged group at 3 days post-ICH (Gong et al., 2005). Additionally, residual brain lesion volume was significantly enhanced in the aged rats

at 28-days post-ICH compared to their younger counterparts, suggesting aging-associated impairment in lesion resolution (Wasserman et al., 2008). However, it is largely unknown how aging precisely modulates neurological deficits, cerebral edema and lesion resolution after ICH, warranting investigation.

Age increases the prevalence of hypertension (Buford, 2016) and CAA (Love et al., 2003), the common causatives of ICH. Hypertension in the context of aging contributes to arterial stiffening and remodeling (Sun, 2015), factors that could predispose to ICH. The vasculopathic changes that are often associated with CAA include loss of smooth muscle cells, vessel wall thickening, lumenal narrowing, concentric splitting of the vessel wall and microaneurysm formation (Viswanathan and Greenberg, 2011). In addition, lobar cerebral microbleeds (CMB or cerebral microhemorrhages: CMH), small focal intracerebral hemorrhages and key contributors of cognitive decline and dementia in older adults (Akoudad et al., 2015), are often found in conjunction with CAA (Viswanathan and Greenberg, 2011). However, the mechanism of blood vessel rupture in CAA is yet to be defined. It is assumed that replacement of the smooth muscle cells of the media by amyloid deposits result in vessel wall weakening and subsequently, vascular rupture (Winkler et al., 2001). It remains largely unclear why some CAA-associated vessel ruptures result in ICH, while others culminate in microhemorrhages. Though the presence of cerebral microbleeds is not significantly associated with the risk of ICH or clinical outcome (Derraz et al., 2021), microbleeds may serve as predictors of ICH recurrence (Tsushima et al., 2003; Greenberg et al., 2004).

Hypertension is another risk factor for CMHs (Vernooij et al., 2008) and aging promoted hypertension inducedcerebral microhemorrhages in a mouse model that recapitulated cerebromicrovascular alterations in elderly humans (Toth et al., 2015). Mechanistically, age-induced reduction in IGF-1 (insulinlike growth factor -1) signaling and reactive oxygen speciesmediated activation of MMPs (Matrix Metalloproteinases) in the cerebrovasculature could make the cerebral blood vessels more vulnerable to hypertension-induced rupture (Tarantini et al., 2017). However, despite the association between increased MMP-9 activation and the genesis of ICH in various experimental murine models (Lee et al., 2003, 2007), genetic deletion of MMP-9 did not attenuate neurological manifestations associated with hypertension-induced ICH in aged mice (Tarantini et al., 2021). Though this is an important observation, there is a possibility that in MMP-9 null mice, other MMP isoforms could express in dysregulated manner and overcompensate the effects of MMP-9 deletion (Tang et al., 2004) and hence, further studies are required with selective pharmacological agents to determine the role of MMP-9 in the pathogenesis of ICH.

Despite an emerging interest in elucidating the association between cerebral microbleeds and ICH, the mechanisms of the development of cerebral microbleeds are largely obscure and complex. A recent study documented that induction of severe systolic hypertension in mice could alter the neurovascular unit resulting in microhemorrhages in the brain (de Montgolfier et al., 2019). Moreover, cerebral venous congestion can contribute to brain microhemorrhages in mice (Nyul-Toth et al., 2022), implicating a novel role of venous circulation in the genesis of cerebral microbleeds, requiring further studies.

Immune Response and Intracerebral Hemorrhage

Aging is a complex process and the immune system experiences significant changes with advanced age. Consistently, in the healthy aged brain, the augmented activation of microglia, the cells that play critical roles in innate immune response, has been reported in diverse mammalian species, including humans (Peters et al., 1991; Dickson et al., 1992; Ogura et al., 1994; Sheffield and Berman, 1998). Furthermore, advanced age is associated with neuronal death, a decline in cognitive function (Ginaldi et al., 1999; Rawji et al., 2016), and a chronic low-grade inflammatory state known as inflamm-aging that is characterized by elevated levels of proinflammatory cytokines (Gabuzda and Yankner, 2013). It is believed that senescence of immune cells and age-dependent changes in macromolecules contribute to inflamm-aging, which, in turn, could partly be responsible for the impaired innate and adaptive immune responses seen in the elderly (Deleidi et al., 2015; Frasca and Blomberg, 2016).

Similar to other bodily tissues, a direct injury to the brain will result in the rapid release of local inflammatory factors and the recruitment of immune cells (Carson et al., 2006). Consequently, microglia undergo alterations in phenotypic, phagocytic, and antigen presentation properties (Sheffield and Berman, 1998). The activated microglia are regarded as the key cellular regulators for neuroinflammation following ICH, owing to their ability to secrete cytokines, chemokines, reactive oxygen species, and prostaglandins (Aronowski and Hall, 2005; Wang and Dore, 2007). The release of these factors further exacerbates microglial activation and recruit blood-derived monocytes/macrophages into the brain, together modulating the inflammatory response (Tessier et al., 1997; Melton et al., 2003; Nakanishi, 2003; Shiratori et al., 2010; Starossom et al., 2012; Chang et al., 2017) and contributing to ICH-induced brain injury (Platt et al., 1998; Hickenbottom et al., 1999; Leira et al., 2004; Zhao et al., 2007). ICH results in the release of a cascade of stimuli that activate microglia/macrophages, which include blood components such as thrombin, hemoglobin, plasma proteins, and hemoglobin degradation products such as hemin and iron (Bonsack and Alleyne, 2016). Several of these factors interact with a class of pattern recognition receptors, Tolllike receptors (TLRs), located on microglia/macrophages and activate proinflammatory signaling such as NFkB or NLRP3 (Dasari et al., 2021). Along these lines, TLR-4 is a key regulator of inflammatory brain damage after ICH (Lin et al., 2012; Wang et al., 2013). Notably, the proinflammatory activation of microglia/macrophage after ICH correlates with blood-brain barrier damage, brain swelling/edema, hematoma expansion, neurological deterioration, and poor functional recovery (Platt et al., 1998; Hickenbottom et al., 1999; Leira et al., 2004; Zhao et al., 2007), implicating microglia as a key contributor of ICHinduced secondary brain injury and loss of neurological function.

Though microglia/macrophage characterization after ICH has primarily been carried out in young animal subjects, it is reported that the number of activated microglia/macrophages

is significantly increased in elderly rats after ICH compared to younger rats (Gong et al., 2004) in line with severe brain injury observed in aged rats. Furthermore, microglia exhibited widespread activation in the ipsilateral brain parenchyma in aged rats after ICH (Wasserman et al., 2008). However, it is largely understudied whether and how aging orchestrates the microglial release of various inflammatory mediators and brain injury after ICH. Studies carried out in young mice have shown that microglia/macrophages undergo polarization after ICH and exhibit pro-inflammatory M1 phenotype or anti-inflammatory M2 phenotype (Bonsack and Alleyne, 2016). The classical activation of microglia/macrophage that gears toward M1 phenotype releases proinflammatory cytokines IL-1 β , IL-6, IL-8, and TNF- α and reactive oxygen species, thereby contributing to brain damage (Wan et al., 2016; Zhang et al., 2016; Lan et al., 2017). In contrast, an alternate activation of microglia yields an anti-inflammatory M2 phenotype, releasing anti-inflammatory cytokines such as IL-10, IL-4, IL-13, and transforming growth factor β (TGF β), culminating in brain recovery (Ni et al., 2016). In line with the detrimental and beneficial role of M1 and M2 microglia/macrophage, respectively, a reduction of M1 or an increase of M2 microglia/macrophage was associated with neuroprotection in the acute phase of ICH. However, studies are yet to be conducted to elucidate how aging alters classical or alternate activation of microglia after ICH and whether microglial phenotypes are viable targets to improve outcomes after ICH in the elderly (Spittau, 2017).

Microglia themselves show age-related changes in phenotype and functionality. Aged microglia are described as dystrophic or senescent (Candlish and Hefendehl, 2021), which exhibit many phenotypic changes compared to young microglia, such as increased soma volume and less arborization, meaning fewer and shorter processes (Koellhoffer et al., 2017; Spittau, 2017). Dystrophic microglia, to some extent, are comparable to activated microglia. Functionally, these dystrophic microglia show reduced chemotaxis and process motility, suggesting that they could respond differentially to neuropathology (Spittau, 2017). Moreover, the number of *dystrophic* microglia significantly increases as individuals age, especially in people with neurodegenerative diseases (Shahidehpour et al., 2021). A potential reason for the increased activation of microglia in the aged brain could be aging-induced myelin breakdown and subsequent activation of the microglia in response to the changes in the brain microenvironment and as an attempt to engulf myelin debris (Conde and Streit, 2006). In addition, studies have shown that aging could shift microglia to a constant low-grade inflammatory state (Pan et al., 2020; Candlish and Hefendehl, 2021), suggesting that microglia could play a critical role in "inflamm-aging," which is partly responsible for age-associated impairments such as decreased remyelination, memory deficits, and gray matter loss (Koellhoffer et al., 2017). Furthermore, aging could prime microglia to a proinflammatory M1 phenotype. Consistently, microglia in aged rats exhibited increased expression of MHC II (Henry et al., 2009), a marker of M1 microglial phenotype. Moreover, aged microglia were associated with enhanced mRNA expression of proinflammatory cytokines such as TNFa, IL-1β, and IL-6

as well as anti-inflammatory cytokines, IL-10 and TGFB1 (Sierra et al., 2007). Moreover, mixed glial cultures from aged mice produced elevated levels of proinflammatory cytokines upon lipopolysaccharide (LPS) treatment compared to those established from young adult mice. Also, microglia from aged mice retained a classically activated or M1 phenotype in the presence of IL-4 (Fenn et al., 2012). In contrast, microglia from young adult mice were responsive to anti-inflammatory cytokine, IL-4 and its treatment shifted microglial phenotype toward an alternatively activated M2 (Fenn et al., 2012). Overall, an enhanced response to proinflammatory signals coupled with a reduced microglial sensitivity to IL-4 could result in exaggerated and prolonged neuroinflammation, amplifying neurodegeneration in the aging brain upon a brain injury. Consistently, it was demonstrated that the expression of IL-1ß protein after ICH was greater in aged rats than in young rats (Lee et al., 2009). Altogether, the age-induced alterations in inflammatory microglial responses could contribute to ICHinduced brain injury and the disproportionate deficits and recovery rates in older patients.

Intracerebral hemorrhage results in both primary and secondary brain injury. The primary brain injury results from the development and mass effect of the hematoma. In contrast, the secondary brain injury, which persists for an extended period and often results in long-term neurological deficits, involves a multitude of mechanisms mostly induced by hematoma components, such as neuroinflammation, oxidative brain damage, and blood-brain damage. Importantly, the volume of the initial hematoma correlates with morbidity and mortality following ICH, and hematoma expansion was associated with poor patient prognosis (Fujii et al., 1994). Altogether, the timely removal of hematoma, the ongoing source of brain damage, is critical for brain recovery after ICH. To this end, apart from the role of microglia in inflammatory brain responses after ICH, studies document that microglia and brain infiltrating macrophages could regulate hematoma resolution and brain recovery owing to their ability to phagocytose cellular debris that accumulates in the brain after a brain injury. Moreover, phagocytosis or removal of dying cells is necessary to prevent the release of intracellular inflammatory agents such as damageassociated molecular patterns (DAMPs) (Sims et al., 2010). Therefore, identification and characterization of endogenous molecular regulators of microglial or macrophage-mediated phagocytosis could improve outcomes after ICH. Of note, elderly subjects with ICH had a larger hematoma volume with poorer outcomes than younger patients (Inoue et al., 2018) partly due to age-mediated parenchymal degeneration and subsequent reduction in the structural integrity of the brain tissue, which could otherwise restrict hematoma growth. Moreover, aged microglia exhibited reduced expression of genes associated with phagocytosis (Orre et al., 2014) and TGFβ-induced phagocytosis was abolished in aged microglia compared to their younger counterparts (Tichauer et al., 2014). Also, aging can enhance the infiltration of brain-infiltrating monocyte-derived macrophage (macrophage/BMDM) after a brain injury (Chou et al., 2018) and modulate its responses, such as the release of inflammatory mediators and phagocytosis (Albright et al., 2016; Rawji et al., 2016). However, how aging functionally alters microglial or BMDM-mediated inflammatory responses, phagocytosis, and hematoma resolution after ICH remains enigmatic, warranting studies.

Another hallmark of brain aging is increased oxidative stress and lipid peroxidation. A prevailing hypothesis is that the age-induced accumulation of free radical damage promotes neuroinflammation. Consistently, there was an overall increase in pro-oxidant and inflammatory genes, while there is a reduction in anti-oxidant genes in the brain of older rodents compared to adults (Lee et al., 1999; Godbout et al., 2005). Furthermore, reactive oxygen species could drive persistent microglial activation (Qin et al., 2013) and promoted M1 microglial activation (Taetzsch et al., 2015). Also, cells damaged by oxidative stress could produce inflammatory factors (Shao et al., 2020), further implicating a role of oxidative stress in neuroimmune responses, warranting investigation. Moreover, age-mediated alterations in the levels of circulating factors such as cytokines could regulate brain injury (Huang et al., 2020). Along these lines, plasma from young rodents could alleviate acute brain injury post-ICH in aged rodents (Yuan et al., 2019), lending support to the conclusion that circulating factors contribute to neural deficits and increased injury after ICH in the elderly.

Iron and Intracerebral Hemorrhage

Iron is a key contributor to both acute as well as delayed brain damage after ICH (Nakamura et al., 2003). The brain concentration of iron, a hemoglobin degradation product, reaches very high levels post-ICH due to erythrocyte lysis and subsequent release of hemoglobin into the extracellular space. A threefold increase of brain non-heme iron after intracerebral hemorrhage was observed in rats (Wu et al., 2003). Iron accumulation in the brain triggers a cascade of deleterious reactions such as free radical production, mitochondria damage, and macrophage/microglial activation, disrupting cellular homeostasis and culminating in neuronal death, oxidative and inflammatory brain injury, and neurological deficits after ICH (Dai et al., 2019). In the acute phase of ICH, hemolysis-generated iron can potentiate thrombin-induced neurotoxicity (Nakamura et al., 2005) and contribute to cerebral edema (Xi et al., 2002b). Although the molecular mechanisms of iron-induced neurotoxicity are not fully understood, iron levels in the brain remain high for at least several weeks post-ICH (Wu et al., 2003), which could contribute to long-term neurological deficits. Importantly, aging is often associated with excess iron accumulation in the substantia nigra, putamen, globus pallidus, caudate nucleus, and cortices (Zecca et al., 2004; Ramos et al., 2014; Ward et al., 2014), which could further modulate brain damage after ICH. In addition, age-mediated enhancement in erythrocyte fragility (Orbach et al., 2017), may alter the rate of erythrocyte lysis subsequent to ICH, resulting in increased heme or iron-induced brain damage. Consistently, the level of the ironregulatory protein, heme-oxygenase 1, was elevated in the aged rat after ICH compared to young rats (Gong et al., 2004). Of note, genetic overexpression of ferroportin 1, an iron exporter, led to less iron accumulation, less neuronal apoptosis, and improved

neurological outcomes in aged mice (Bao et al., 2020), further implicating a role of iron in ICH pathophysiology.

Evidence has been shown that advanced age is associated with enhanced complement activation (Gong et al., 2008), which plays a role in the formation of membrane attack complexes (MAC) (Hua et al., 2000; Ducruet et al., 2009), resulting in erythrocyte lysis and hence, hemoglobin or iron-mediated neurotoxicity (Yuan et al., 2019) and cerebral edema development after ICH (Xi et al., 2001, 2002a; Yang et al., 2006a,b). Moreover, complement components such as C3a anaphylatoxin could also contribute to ICH pathology by enhancing vascular permeability (Foreman et al., 1996) and leukocyte infiltration. Consistent with the role of complement activation in brain injury, intracerebral administration of a complement inhibitor reduced erythrolysis, iron accumulation, microglial activation, cerebral edema, and neuronal death in aged rats after ICH (Yuan et al., 2019). Furthermore, complement components may play a role in the clearance of apoptotic cell bodies and contribute to ischemic stroke-induced neurogenesis (Rahpeymai et al., 2006), implicating its unexplored role in brain recovery after ICH. Therefore, further studies are required to determine the precise molecular mechanisms by which complement activation modulate brain damage or recovery and whether systemic administration of a complement inhibitor is a feasible strategy to improve neurological outcomes in aged mice after ICH.

White matter injury is a frequent complication of ICH (Tao et al., 2017) and as per a report, more than 77% of ICH patients suffered white matter injury (Smith et al., 2004). White matter injury is observed in both acute and chronic phases of ICH and is characterized by demyelination, axonal damage and oligodendrocyte death (Ni et al., 2015). Though the precise mechanism of white matter injury after ICH is enigmatic, ironinduced oxidative stress could culminate in white matter damage (Li et al., 2021). In a rat model of ICH, white matter injury correlated with brain edema and poor neurological outcomes (Tao et al., 2016). Moreover, white matter injury is a major cause of sensory-motor deficits commonly seen in ICH patients (Li et al., 2021) and was associated with cognitive impairment (Smith et al., 2004). Of note, aging is often associated with cerebral white matter lesions characterized by demyelination, gliosis, and capillary degeneration (Hoffman et al., 1985; Baltan et al., 2008; Asdaghi et al., 2012). Also, aging can augment white matter vulnerability to ICH-induced brain damage. Altogether, additional studies are required to delineate the age-induced changes in iron metabolism and molecular mechanisms of ironinduced neurological deficits in the aging population after ICH.

Cognition and Intracerebral Hemorrhage

There is a high prevalence of dementia after ICH (ranging from 9 to 29% for pre-ICH and 14–88% for post-ICH) (Donnellan and Werring, 2020) and dementia could be a predictor of mortality in ICH survivors (Judge et al., 2019). After ICH, cognitive deficits could arise from the acute hemorrhagic lesion or in a progressive manner owing to slowly accumulating vascular and non-vascular pathology (Xiong et al., 2016). Notably, cognitive impairment after ICH remains largely understudied. In a preclinical rodent model of ICH, no significant learning or



memory deficits were observed 1–7 months post-ICH (MacLellan et al., 2009). However, in another study using the same model, there were significant learning deficits at 2 weeks post-ICH, but the learning deficits reduced remarkably at 8 weeks post-ICH (Hartman et al., 2009). These conflicting results warrant additional investigation. Moreover, these studies were conducted in young animal subjects, which lack underlying neuropathology, which could otherwise be needed for the development of cognitive impairment after ICH, apart from hemorrhage-induced brain damage. To this end, employing aged animal subjects could better establish the association between ICH and cognition, which may improve the prognosis of ICH survivors.

Prognostic Factors and Intracerebral Hemorrhage

Given the devastatingly high morbidity and mortality associated with ICH, the predictors of patient prognosis carry high clinical significance. The predictors of adverse patient outcome include advanced age, enhanced ICH volume, presence of intraventricular hemorrhage, low Glasgow Coma Scale score (GCS score) and deep/infratentorial ICH location (Poon et al., 2014). Evaluation of these prognostic factors helps establish an ICH score or a risk stratification scale predicting 30-day mortality (Hemphill et al., 2001). Of note, Yang et al. (2020) found that the predictors of patient mortality differ between young and aged ICH patients. To this end, brain herniation in the young group, and low GCS scores, renal or heart disease, and leukocytosis in the elderly were associated with higher 1month mortality (Yang et al., 2020). Also, studies on elderly patients with age \geq 75 years demonstrated that a hematoma volume \geq 30 ml, or a prior history of ICH was associated with a higher likelihood of short-term death (Batista et al., 2021). Apart from these, blood-derived inflammation markers could serve as prognostic indicators. Along these lines, increased plasma level of TNF-a was associated with mortality in ICH patients (Fang et al., 2007). As per another study, elevated plasma level of IL-6 is an independent predictor for early hematoma growth, which, in turn, is associated with poor outcomes following ICH (Silva et al., 2005). Though inflammatory biomarkers that could

predict better recovery after ICH are least characterized, as per a recent report, increased serum levels of IL-33, a newly identified member of the IL-1 family, were found in patients with improved functional outcomes compared to those with poor outcomes (Miao et al., 2021). It is important to highlight that these studies investigating potential ICH biomarkers are not limited to the elderly population, but often include any patient of adult age. Therefore, further studies are needed to determine whether these predictive markers are as effective when solely looking at elderly patients.

CONCLUSION

Intracerebral hemorrhage is a complex disorder with no effective treatment. Aging has a multifaceted effect on the development and the progression of the disease (**Figure 1**). Therefore, aging could impose a myriad of unique challenges to ICH treatment. However, the molecular level changes that occur in the brain after ICH remain largely unknown. Given that the aged population is the most rapidly growing population in America and possible increase in the incidence of ICH in the aging population, there is a need to conduct additional preclinical studies with old animal subjects for a better understanding of the role of aging in ICH pathology, which in turn would aid in the development of novel treatment strategies.

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NW, FB, and SS-R: original draft preparation and writing. SS-R: conceptualization, editing, and funding acquisition. All authors contributed to the article and approved the submitted version.

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REFERENCES

- Akoudad, S., Portegies, M. L., Koudstaal, P. J., Hofman, A., van der Lugt, A., Ikram, M. A., et al. (2015). Cerebral Microbleeds Are Associated With an Increased Risk of Stroke: the Rotterdam Study. *Circulation* 132, 509–516. doi: 10.1161/ CIRCULATIONAHA.115.016261
- Albright, J. M., Dunn, R. C., Shults, J. A., Boe, D. M., Afshar, M., and Kovacs, E. J. (2016). Advanced Age Alters Monocyte and Macrophage Responses. *Antioxid. Redox. Signal.* 25, 805–815. doi: 10.1089/ars.2016.6691
- An, S. J., Kim, T. J., and Yoon, B. W. (2017). Epidemiology, Risk Factors, and Clinical Features of Intracerebral Hemorrhage: an Update. J. Stroke 19, 3–10. doi: 10.5853/jos.2016.00864
- Ariesen, M. J., Claus, S. P., Rinkel, G. J., and Algra, A. (2003). Risk factors for intracerebral hemorrhage in the general population: a systematic review. *Stroke* 34, 2060–2065. doi: 10.1161/01.STR.0000080678.09344.8D
- Aronowski, J., and Hall, C. E. (2005). New horizons for primary intracerebral hemorrhage treatment: experience from preclinical studies. *Neurol. Res.* 27, 268–279. doi: 10.1179/016164105X25225
- Asdaghi, N., Butcher, K. S., and Hill, M. D. (2012). Risks and benefits of thrombolysis in the elderly. *Int. J. Stroke* 7, 142–149. doi: 10.1111/j.1747-4949. 2011.00744.x
- Bagg, S., Pombo, A. P., and Hopman, W. (2002). Effect of age on functional outcomes after stroke rehabilitation. *Stroke* 33, 179–185. doi: 10.1161/hs0102. 101224
- Baltan, S., Besancon, E. F., Mbow, B., Ye, Z., Hamner, M. A., and Ransom, B. R. (2008). White matter vulnerability to ischemic injury increases with age because of enhanced excitotoxicity. *J. Neurosci.* 28, 1479–1489. doi: 10.1523/ JNEUROSCI.5137-07.2008
- Bao, W. D., Zhou, X. T., Zhou, L. T., Wang, F., Yin, X., Lu, Y., et al. (2020). Targeting miR-124/Ferroportin signaling ameliorated neuronal cell death through inhibiting apoptosis and ferroptosis in aged intracerebral hemorrhage murine model. *Aging Cell* 19:e13235. doi: 10.1111/acel.13235
- Batista, A., Osorio, R., Varela, A., Guilherme, P., Marreiros, A., Pais, S., et al. (2021). Prediction of short-term prognosis in elderly patients with spontaneous intracerebral hemorrhage. *Eur. Geriatr. Med.* 12, 1267–1273. doi: 10.1007/ s41999-021-00529-w
- Bonsack, Ft, and Alleyne, C. H. Jr. (2016). Sukumari-Ramesh S: augmented expression of TSPO after intracerebral hemorrhage: a role in inflammation? *J. Neuroinflamm.* 13:151. doi: 10.1186/s12974-016-0619-2
- Broderick, J. P., Brott, T., Tomsick, T., Miller, R., and Huster, G. (1993). Intracerebral hemorrhage more than twice as common as subarachnoid hemorrhage. J. Neurosurg. 78, 188–191. doi: 10.3171/jns.1993.78.2.0188
- Buford, T. W. (2016). Hypertension and aging. Ageing Res. Rev. 26, 96-111.
- Candlish, M., and Hefendehl, J. K. (2021). Microglia Phenotypes Converge in Aging and Neurodegenerative Disease. *Front. Neurol.* 12:660720. doi: 10.3389/ fneur.2021.660720
- Caro, J. J., and Huybrechts, K. F. (1999). Stroke treatment economic model (STEM): predicting long-term costs from functional status. *Stroke* 30, 2574– 2579. doi: 10.1161/01.str.30.12.2574
- Carson, M. J., Thrash, J. C., and Walter, B. (2006). The cellular response in neuroinflammation: the role of leukocytes, microglia and astrocytes in neuronal death and survival. *Clin. Neurosci. Res.* 6, 237–245. doi: 10.1016/j.cnr.2006.09. 004
- Chang, C. F., Wan, J., Li, Q., Renfroe, S. C., Heller, N. M., and Wang, J. (2017). Alternative activation-skewed microglia/macrophages promote hematoma resolution in experimental intracerebral hemorrhage. *Neurobiol. Dis.* 103, 54– 69. doi: 10.1016/j.nbd.2017.03.016
- Cheng, Y., Qiao, L., Jiang, Z., Dong, X., Feng, H., Gui, Q., et al. (2020). Significant reduction in the LDL cholesterol increases the risk of intracerebral hemorrhage: a systematic review and meta-analysis of 33 randomized controlled trials. *Am. J. Transl. Res.* 12, 463–477.
- Cheung, R. T., and Zou, L. Y. (2003). Use of the original, modified, or new intracerebral hemorrhage score to predict mortality and morbidity after intracerebral hemorrhage. *Stroke* 34, 1717–1722. doi: 10.1161/01.STR. 0000078657.22835.B9
- Chou, A., Krukowski, K., Morganti, J. M., Riparip, L. K., and Rosi, S. (2018). Persistent Infiltration and Impaired Response of Peripherally-Derived

Monocytes after Traumatic Brain Injury in the Aged Brain. *Int. J. Mol. Sci.* 19:1616. doi: 10.3390/ijms19061616

- Conde, J. R., and Streit, W. J. (2006). Microglia in the aging brain. J. Neuropathol. Exp. Neurol. 65, 199–203.
- Craen, A., Mangal, R., Stead, T. G., and Ganti, L. (2019). Gender Differences in Outcomes after Non-traumatic Intracerebral Hemorrhage. *Cureus* 11:e5818. doi: 10.7759/cureus.5818
- Dai, S., Hua, Y., Keep, R. F., Novakovic, N., Fei, Z., and Xi, G. (2019). Minocycline attenuates brain injury and iron overload after intracerebral hemorrhage in aged female rats. *Neurobiol. Dis.* 126, 76–84. doi: 10.1016/j.nbd.2018.06.001
- Dasari, R., Bonsack, F., and Sukumari-Ramesh, S. (2021). Brain injury and repair after intracerebral hemorrhage: the role of microglia and brain-infiltrating macrophages. *Neurochem. Int.* 142:104923. doi: 10.1016/j.neuint.2020.104923
- de Montgolfier, O., Pouliot, P., Gillis, M. A., Ferland, G., Lesage, F., Thorin-Trescases, N., et al. (2019). Systolic hypertension-induced neurovascular unit disruption magnifies vascular cognitive impairment in middle-age atherosclerotic LDLr(-/-):hApoB(+/+) mice. Geroscience 41, 511–532. doi: 10. 1007/s11357-019-00070-6
- Deleidi, M., Jaggle, M., and Rubino, G. (2015). Immune aging, dysmetabolism, and inflammation in neurological diseases. *Front. Neurosci.* 9:172. doi: 10.3389/ fnins.2015.00172
- Derraz, I., Cagnazzo, F., Gaillard, N., Morganti, R., Dargazanli, C., Ahmed, R., et al. (2021). Microbleeds, Cerebral Hemorrhage, and Functional Outcome After Endovascular Thrombectomy. *Neurology* 96, e1724–e1731. doi: 10.1212/WNL. 000000000011566
- Dickson, D. W., Crystal, H. A., Mattiace, L. A., Masur, D. M., Blau, A. D., Davies, P., et al. (1992). Identification of normal and pathological aging in prospectively studied nondemented elderly humans. *Neurobiol. Aging* 13, 179–189. doi: 10. 1016/0197-4580(92)90027-u
- Donnellan, C., and Werring, D. (2020). Cognitive impairment before and after intracerebral haemorrhage: a systematic review. *Neurol. Sci.* 41, 509–527. doi: 10.1007/s10072-019-04150-5
- Ducruet, A. F., Zacharia, B. E., Hickman, Z. L., Grobelny, B. T., Yeh, M. L., Sosunov, S. A., et al. (2009). The complement cascade as a therapeutic target in intracerebral hemorrhage. *Exp. Neurol.* 219, 398–403. doi: 10.1016/j.expneurol. 2009.07.018
- Fang, H. Y., Ko, W. J., and Lin, C. Y. (2007). Inducible heat shock protein 70, interleukin-18, and tumor necrosis factor alpha correlate with outcomes in spontaneous intracerebral hemorrhage. J. Clin. Neurosci. 14, 435–441. doi: 10. 1016/j.jocn.2005.12.022
- Feigin, V. L., Lawes, C. M., Bennett, D. A., Barker-Collo, S. L., and Parag, V. (2009). Worldwide stroke incidence and early case fatality reported in 56 populationbased studies: a systematic review. *Lancet Neurol.* 8, 355–369. doi: 10.1016/ S1474-4422(09)70025-0
- Feldmann, E., Broderick, J. P., Kernan, W. N., Viscoli, C. M., Brass, L. M., Brott, T., et al. (2005). Major risk factors for intracerebral hemorrhage in the young are modifiable. *Stroke* 36, 1881–1885. doi: 10.1161/01.STR.0000177480.62341.6b
- Fenn, A. M., Henry, C. J., Huang, Y., Dugan, A., and Godbout, J. P. (2012). Lipopolysaccharide-induced interleukin (IL)-4 receptor-alpha expression and corresponding sensitivity to the M2 promoting effects of IL-4 are impaired in microglia of aged mice. *Brain Behav. Immun.* 26, 766–777. doi: 10.1016/j.bbi. 2011.10.003
- Flaherty, M. L., Woo, D., Haverbusch, M., Sekar, P., Khoury, J., Sauerbeck, L., et al. (2005). Racial variations in location and risk of intracerebral hemorrhage. *Stroke* 36, 934–937. doi: 10.1161/01.STR.0000160756.72109.95
- Foreman, K. E., Glovsky, M. M., Warner, R. L., Horvath, S. J., and Ward, P. A. (1996). Comparative effect of C3a and C5a on adhesion molecule expression on neutrophils and endothelial cells. *Inflammation* 20, 1–9. doi: 10.1007/ BF01487740
- Frasca, D., and Blomberg, B. B. (2016). Inflammaging decreases adaptive and innate immune responses in mice and humans. *Biogerontology* 17, 7–19. doi: 10.1007/s10522-015-9578-8
- Fujii, Y., Tanaka, R., Takeuchi, S., Koike, T., Minakawa, T., and Sasaki, O. (1994). Hematoma enlargement in spontaneous intracerebral hemorrhage. *J. Neurosurg.* 80, 51–57. doi: 10.3171/jns.1994.80.1.0051
- Gabuzda, D., and Yankner, B. A. (2013). Physiology: inflammation links ageing to the brain. *Nature* 497, 197–198. doi: 10.1038/nature12100

- Ginaldi, L., De Martinis, M., D'Ostilio, A., Marini, L., Loreto, M. F., and Quaglino, D. (1999). Immunological changes in the elderly. *Aging* 11, 281–286. doi: 10. 1007/bf03339801
- Godbout, J. P., Chen, J., Abraham, J., Richwine, A. F., Berg, B. M., Kelley, K. W., et al. (2005). Exaggerated neuroinflammation and sickness behavior in aged mice following activation of the peripheral innate immune system. *FASEB J.* 19, 1329–1331. doi: 10.1096/fj.05-3776fje
- Gong, Y., Hua, Y., Keep, R. F., Hoff, J. T., and Xi, G. (2004). Intracerebral hemorrhage: effects of aging on brain edema and neurological deficits. *Stroke* 35, 2571–2575. doi: 10.1161/01.STR.0000145485.67827.d0
- Gong, Y., Xi, G. H., Keep, R. F., Hoff, J. T., and Hua, Y. (2005). Aging enhances intracerebral hemorrhage-induced brain injury in rats. *Acta Neurochir. Suppl.* 95, 425–427. doi: 10.1007/3-211-32318-x_87
- Gong, Y., Xi, G., Wan, S., Gu, Y., Keep, R. F., and Hua, Y. (2008). Effects of aging on complement activation and neutrophil infiltration after intracerebral hemorrhage. *Acta Neurochir. Suppl.* 105, 67–70. doi: 10.1007/978-3-211-09469-3_14
- Gonzalez-Perez, A., Gaist, D., Wallander, M. A., McFeat, G., and Garcia-Rodriguez, L. A. (2013). Mortality after hemorrhagic stroke: data from general practice (The Health Improvement Network). *Neurology* 81, 559–565. doi: 10.1212/ WNL.0b013e31829e6eff
- Greenberg, S. M., Eng, J. A., Ning, M., Smith, E. E., and Rosand, J. (2004). Hemorrhage burden predicts recurrent intracerebral hemorrhage after lobar hemorrhage. *Stroke* 35, 1415–1420. doi: 10.1161/01.STR.0000126807.697 58.0e
- Hartman, R., Lekic, T., Rojas, H., Tang, J., and Zhang, J. H. (2009). Assessing functional outcomes following intracerebral hemorrhage in rats. *Brain Res.* 1280, 148–157. doi: 10.1016/j.brainres.2009.05.038
- Hemphill, J. C. III, Bonovich, D. C., Besmertis, L., Manley, G. T., and Johnston, S. C. (2001). The ICH score: a simple, reliable grading scale for intracerebral hemorrhage. *Stroke* 32, 891–897. doi: 10.1161/01.str.32.4.891
- Henry, C. J., Huang, Y., Wynne, A. M., and Godbout, J. P. (2009). Peripheral lipopolysaccharide (LPS) challenge promotes microglial hyperactivity in aged mice that is associated with exaggerated induction of both pro-inflammatory IL-1beta and anti-inflammatory IL-10 cytokines. *Brain Behav. Immun.* 23, 309–317. doi: 10.1016/j.bbi.2008.09.002
- Hickenbottom, S. L., Grotta, J. C., Strong, R., Denner, L. A., and Aronowski, J. (1999). Nuclear factor-kappaB and cell death after experimental intracerebral hemorrhage in rats. *Stroke* 30, 2472–2477. doi: 10.1161/01.str.30.11.2472
- Hoffman, W. E., Pelligrino, D., Miletich, D. J., and Albrecht, R. F. (1985). Brain metabolic changes in young vs aged rats during hypoxia. *Stroke* 16, 860–863. doi: 10.1161/01.str.16.5.860
- Hua, Y., Xi, G., Keep, R. F., and Hoff, J. T. (2000). Complement activation in the brain after experimental intracerebral hemorrhage. J. Neurosurg. 92, 1016–1022. doi: 10.3171/jns.2000.92.6.1016
- Huang, J., Yang, G., Xiong, X., Wang, M., Yuan, J., Zhang, Q., et al. (2020). Agerelated CCL12 Aggravates Intracerebral Hemorrhage-induced Brain Injury via Recruitment of Macrophages and T Lymphocytes. *Aging Dis.* 11, 1103–1115. doi: 10.14336/AD.2019.1229
- Inoue, Y., Miyashita, F., Minematsu, K., and Toyoda, K. (2018). Clinical Characteristics and Outcomes of Intracerebral Hemorrhage in Very Elderly. *J. Stroke Cerebrovasc. Dis.* 27, 97–102. doi: 10.1016/j.jstrokecerebrovasdis.2017. 08.006
- Itoh, Y., Yamada, M., Hayakawa, M., Otomo, E., and Miyatake, T. (1993). Cerebral amyloid angiopathy: a significant cause of cerebellar as well as lobar cerebral hemorrhage in the elderly. *J. Neurol. Sci.* 116, 135–141. doi: 10.1016/0022-510x(93)90317-r
- Jabbarli, R., Reinhard, M., Roelz, R., Shah, M., Niesen, W. D., Kaier, K., et al. (2016). Intracerebral Hematoma Due to Aneurysm Rupture: are There Risk Factors Beyond Aneurysm Location? *Neurosurgery* 78, 813–820. doi: 10.1227/ NEU.000000000001136
- Jackson, C. A., and Sudlow, C. L. (2006). Is hypertension a more frequent risk factor for deep than for lobar supratentorial intracerebral haemorrhage? J. Neurol. Neurosurg. Psychiatry 77, 1244–1252. doi: 10.1136/jnnp.2006.089292
- Judge, C., Ruttledge, S., Costello, M., Murphy, R., Loughlin, E., Alvarez-Iglesias, A., et al. (2019). Lipid Lowering Therapy, Low-Density Lipoprotein Level and Risk of Intracerebral Hemorrhage - A Meta-Analysis. J. Stroke Cerebrovasc. Dis. 28, 1703–1709. doi: 10.1016/j.jstrokecerebrovasdis.2019.02.018

- Koellhoffer, E. C., McCullough, L. D., and Ritzel, R. M. (2017). Old Maids: Aging and Its Impact on Microglia Function. *Int. J. Mol. Sci.* 18:769. doi: 10.3390/ ijms18040769
- Lan, X., Han, X., Li, Q., Li, Q., Gao, Y., Cheng, T., et al. (2017). Pinocembrin protects hemorrhagic brain primarily by inhibiting toll-like receptor 4 and reducing M1 phenotype microglia. *Brain Behav. Immun.* 61, 326–339. doi: 10.1016/j.bbi.2016.12.012
- Lee, C. K., Klopp, R. G., Weindruch, R., and Prolla, T. A. (1999). Gene expression profile of aging and its retardation by caloric restriction. *Science* 285, 1390– 1393. doi: 10.1126/science.285.5432.1390
- Lee, C. Z., Xue, Z., Zhu, Y., Yang, G. Y., and Young, W. L. (2007). Matrix metalloproteinase-9 inhibition attenuates vascular endothelial growth factorinduced intracerebral hemorrhage. *Stroke* 38, 2563–2568. doi: 10.1161/ STROKEAHA.106.481515
- Lee, J. C., Cho, G. S., Choi, B. O., Kim, H. C., and Kim, W. K. (2009). Aging exacerbates intracerebral hemorrhage-induced brain injury. J. Neurotrauma 26, 1567–1576. doi: 10.1089/neu.2008.0630
- Lee, J. M., Yin, K. J., Hsin, I., Chen, S., Fryer, J. D., Holtzman, D. M., et al. (2003). Matrix metalloproteinase-9 and spontaneous hemorrhage in an animal model of cerebral amyloid angiopathy. *Ann. Neurol.* 54, 379–382. doi: 10.1002/ana. 10671
- Leira, R., Davalos, A., Silva, Y., Gil-Peralta, A., Tejada, J., Garcia, M., et al. (2004). Stroke Project CDGotSNS: early neurologic deterioration in intracerebral hemorrhage: predictors and associated factors. *Neurology* 63, 461–467. doi: 10.1212/01.wnl.0000133204.81153.ac
- Li, J., Xiao, L., He, D., Luo, Y., and Sun, H. (2021). Mechanism of White Matter Injury and Promising Therapeutic Strategies of MSCs After Intracerebral Hemorrhage. *Front. Aging Neurosci.* 13:632054. doi: 10.3389/fnagi.2021.632054
- Lin, S., Yin, Q., Zhong, Q., Lv, F. L., Zhou, Y., Li, J. Q., et al. (2012). Heme activates TLR4-mediated inflammatory injury via MyD88/TRIF signaling pathway in intracerebral hemorrhage. *J. Neuroinflammation* 9:46. doi: 10.1186/1742-2094-9-46
- Love, S., Nicoll, J. A., Hughes, A., and Wilcock, G. K. (2003). APOE and cerebral amyloid angiopathy in the elderly. *Neuroreport* 14, 1535–1536. doi: 10.1097/ 00001756-200308060-00027
- Ma, C., Gurol, M. E., Huang, Z., Lichtenstein, A. H., Wang, X., Wang, Y., et al. (2019). Low-density lipoprotein cholesterol and risk of intracerebral hemorrhage: a prospective study. *Neurology* 93, e445–e457.
- MacLellan, C. L., Langdon, K. D., Churchill, K. P., Granter-Button, S., and Corbett, D. (2009). Assessing cognitive function after intracerebral hemorrhage in rats. *Behav. Brain Res.* 198, 321–328. doi: 10.1016/j.bbr.2008.11.004
- Matsukawa, H., Shinoda, M., Fujii, M., Takahashi, O., Yamamoto, D., Murakata, A., et al. (2012). Factors associated with lobar vs. non-lobar intracerebral hemorrhage. *Acta Neurol. Scand.* 126, 116–121. doi: 10.1111/j.1600-0404.2011. 01615.x
- Mehndiratta, P., Manjila, S., Ostergard, T., Eisele, S., Cohen, M. L., Sila, C., et al. (2012). Cerebral amyloid angiopathy-associated intracerebral hemorrhage: pathology and management. *Neurosurg. Focus* 32:E7. doi: 10.3171/2012.1. FOCUS11370
- Melton, L. M., Keith, A. B., Davis, S., Oakley, A. E., Edwardson, J. A., and Morris, C. M. (2003). Chronic glial activation, neurodegeneration, and APP immunoreactive deposits following acute administration of double-stranded RNA. *Glia* 44, 1–12. doi: 10.1002/glia.10276
- Miao, Y., Zhang, Z.-X., Feng, X., and Sun, W.-M. (2021). IL-33 as a Novel Serum Prognostic Marker of Intracerebral Hemorrhage. Oxid. Med. Cell. Long. 2021:5597790. doi: 10.1155/2021/5597790
- Morotti, A., and Goldstein, J. N. (2016). Diagnosis and Management of Acute Intracerebral Hemorrhage. *Emerg. Med. Clin. North Am.* 34, 883–899. doi: 10.1016/j.emc.2016.06.010
- Mozaffarian, D., Benjamin, E. J., Go, A. S., Arnett, D. K., Blaha, M. J., Cushman, M., et al. (2015). Heart Disease and Stroke Statistics-2016 Update: a Report From the American Heart Association. *Circulation* 133, e38–360.
- Nakamura, T., Keep, R. F., Hua, Y., Schallert, T., Hoff, J. T., and Xi, G. (2003). Deferoxamine-induced attenuation of brain edema and neurological deficits in a rat model of intracerebral hemorrhage. *Neurosurg. Focus* 15:EC4.
- Nakamura, T., Xi, G., Park, J. W., Hua, Y., Hoff, J. T., and Keep, R. F. (2005). Holo-transferrin and thrombin can interact to cause brain damage. *Stroke* 36, 348–352. doi: 10.1161/01.STR.0000153044.60858.1b

- Nakanishi, H. (2003). Microglial functions and proteases. Mol. Neurobiol. 27, 163–176. doi: 10.1385/MN:27:2:163
- Ni, W., Mao, S., Xi, G., Keep, R. F., and Hua, Y. (2016). Role of Erythrocyte CD47 in Intracerebral Hematoma Clearance. *Stroke* 47, 505–511. doi: 10.1161/ STROKEAHA.115.010920
- Ni, W., Okauchi, M., Hatakeyama, T., Gu, Y., Keep, R. F., Xi, G., et al. (2015). Deferoxamine reduces intracerebral hemorrhage-induced white matter damage in aged rats. *Exp. Neurol.* 272, 128–134. doi: 10.1016/j.expneurol.2015.02.035
- Nicoll, J. A., Burnett, C., Love, S., Graham, D. I., Dewar, D., Ironside, J. W., et al. (1997). High frequency of apolipoprotein E epsilon 2 allele in hemorrhage due to cerebral amyloid angiopathy. *Ann. Neurol.* 41, 716–721. doi: 10.1002/ana. 410410607
- Nyul-Toth, A., Fulop, G. A., Tarantini, S., Kiss, T., Ahire, C., Faakye, J. A., et al. (2022). Cerebral venous congestion exacerbates cerebral microhemorrhages in mice. *Geroscience* Epub online ahead of print. doi: 10.1007/s11357-021-00504-0
- Ogura, K., Ogawa, M., and Yoshida, M. (1994). Effects of ageing on microglia in the normal rat brain: immunohistochemical observations. *Neuroreport* 5, 1224–1226. doi: 10.1097/00001756-199406020-00016
- Orbach, A., Zelig, O., Yedgar, S., and Barshtein, G. (2017). Biophysical and Biochemical Markers of Red Blood Cell Fragility. *Transfus Med. Hemother.* 44, 183–187. doi: 10.1159/000452106
- Orre, M., Kamphuis, W., Osborn, L. M., Jansen, A. H. P., Kooijman, L., Bossers, K., et al. (2014). Isolation of glia from Alzheimer's mice reveals inflammation and dysfunction. *Neurobiol. Aging* 35, 2746–2760. doi: 10.1016/j.neurobiolaging. 2014.06.004
- Ovbiagele, B., Goldstein, L. B., Higashida, R. T., Howard, V. J., Johnston, S. C., Khavjou, O. A., et al. (2013). Forecasting the future of stroke in the United States: a policy statement from the American Heart Association and American Stroke Association. *Stroke* 44, 2361–2375. doi: 10.1161/STR. 0b013e31829734f2
- Pan, J., Ma, N., Yu, B., Zhang, W., and Wan, J. (2020). Transcriptomic profiling of microglia and astrocytes throughout aging. J. Neuroinflammation 17:97. doi: 10.1186/s12974-020-01774-9
- Peters, A., Josephson, K., and Vincent, S. L. (1991). Effects of aging on the neuroglial cells and pericytes within area 17 of the rhesus monkey cerebral cortex. Anat. Rec. 229, 384–398. doi: 10.1002/ar.1092290311
- Platt, N., da Silva, R. P., and Gordon, S. (1998). Recognizing death: the phagocytosis of apoptotic cells. *Trends Cell Biol.* 8, 365–372. doi: 10.1016/s0962-8924(98) 01329-4
- Poon, M. T., Fonville, A. F., and Al-Shahi Salman, R. (2014). Long-term prognosis after intracerebral haemorrhage: systematic review and meta-analysis. J. Neurol. Neurosurg. Psychiatry 85, 660–667. doi: 10.1136/jnnp-2013-306476
- Qin, L., Liu, Y., Hong, J. S., and Crews, F. T. (2013). NADPH oxidase and aging drive microglial activation, oxidative stress, and dopaminergic neurodegeneration following systemic LPS administration. *Glia* 61, 855–868. doi: 10.1002/glia.22479
- Qureshi, A. I., Mendelow, A. D., and Hanley, D. F. (2009). Intracerebral haemorrhage. *Lancet* 373, 1632–1644.
- Qureshi, A. I., Suri, M. F., Nasar, A., Kirmani, J. F., Ezzeddine, M. A., Divani, A. A., et al. (2007). Changes in cost and outcome among US patients with stroke hospitalized in 1990 to 1991 and those hospitalized in 2000 to 2001. *Stroke* 38, 2180–2184. doi: 10.1161/STROKEAHA.106.467506
- Rahpeymai, Y., Hietala, M. A., Wilhelmsson, U., Fotheringham, A., Davies, I., Nilsson, A. K., et al. (2006). Complement: a novel factor in basal and ischemiainduced neurogenesis. *EMBO J.* 25, 1364–1374. doi: 10.1038/sj.emboj.7601004
- Ramos, P., Santos, A., Pinto, N. R., Mendes, R., Magalhaes, T., and Almeida, A. (2014). Iron levels in the human brain: a post-mortem study of anatomical region differences and age-related changes. *J. Trace. Elem. Med. Biol.* 28, 13–17. doi: 10.1016/j.jtemb.2013.08.001
- Rawji, K. S., Mishra, M. K., Michaels, N. J., Rivest, S., Stys, P. K., and Yong, V. W. (2016). Immunosenescence of microglia and macrophages: impact on the ageing central nervous system. *Brain* 139, 653–661. doi: 10.1093/brain/awv395
- Reed, S. D., Blough, D. K., Meyer, K., and Jarvik, J. G. (2001). Inpatient costs, length of stay, and mortality for cerebrovascular events in community hospitals. *Neurology* 57, 305–314. doi: 10.1212/wnl.57.2.305
- Sacco, S., Marini, C., Toni, D., Olivieri, L., and Carolei, A. (2009). Incidence and 10year survival of intracerebral hemorrhage in a population-based registry. *Stroke* 40, 394–399. doi: 10.1161/STROKEAHA.108.523209

- Shahidehpour, R. K., Higdon, R. E., Crawford, N. G., Neltner, J. H., Ighodaro, E. T., Patel, E., et al. (2021). Dystrophic microglia are associated with neurodegenerative disease and not healthy aging in the human brain. *Neurobiol. Aging* 99, 19–27. doi: 10.1016/j.neurobiolaging.2020.12.003
- Shao, A., Lin, D., Wang, L., Tu, S., Lenahan, C., and Zhang, J. (2020). Oxidative Stress at the Crossroads of Aging. Stroke and Depression. Aging Dis. 11, 1537–1566. doi: 10.14336/AD.2020.0225
- Sheffield, L. G., and Berman, N. E. (1998). Microglial expression of MHC class II increases in normal aging of nonhuman primates. *Neurobiol. Aging* 19, 47–55. doi: 10.1016/s0197-4580(97)00168-1
- Shiratori, M., Tozaki-Saitoh, H., Yoshitake, M., Tsuda, M., and Inoue, K. (2010). P2X7 receptor activation induces CXCL2 production in microglia through NFAT and PKC/MAPK pathways. J. Neurochem. 114, 810–819. doi: 10.1111/ j.1471-4159.2010.06809.x
- Sierra, A., Gottfried-Blackmore, A. C., McEwen, B. S., and Bulloch, K. (2007). Microglia derived from aging mice exhibit an altered inflammatory profile. *Glia* 55, 412–424. doi: 10.1002/glia.20468
- Silva, Y., Leira, R., Tejada, J., Lainez, J. M., Castillo, J., and Davalos, A. (2005). Stroke Project CDGotSNS: molecular signatures of vascular injury are associated with early growth of intracerebral hemorrhage. *Stroke* 36, 86–91. doi: 10.1161/01.STR.0000149615.51204.0b
- Sims, G. P., Rowe, D. C., Rietdijk, S. T., Herbst, R., and Coyle, A. J. (2010). HMGB1 and RAGE in inflammation and cancer. *Annu. Rev. Immunol.* 28, 367–388. doi: 10.1146/annurev.immunol.021908.132603
- Smith, E. E., Gurol, M. E., Eng, J. A., Engel, C. R., Nguyen, T. N., Rosand, J., et al. (2004). White matter lesions, cognition, and recurrent hemorrhage in lobar intracerebral hemorrhage. *Neurology* 63, 1606–1612. doi: 10.1212/01.wnl. 0000142966.22886.20
- Spittau, B. (2017). Aging Microglia-Phenotypes, Functions and Implications for Age-Related Neurodegenerative Diseases. *Front. Aging Neurosci.* 9:194. doi: 10.3389/fnagi.2017.00194
- Starossom, S. C., Mascanfroni, I. D., Imitola, J., Cao, L., Raddassi, K., Hernandez, S. F., et al. (2012). Galectin-1 deactivates classically activated microglia and protects from inflammation-induced neurodegeneration. *Immunity* 37, 249– 263. doi: 10.1016/j.immuni.2012.05.023
- Stein, M., Misselwitz, B., Hamann, G. F., Scharbrodt, W., Schummer, D. I., and Oertel, M. F. (2012). Intracerebral hemorrhage in the very old: future demographic trends of an aging population. *Stroke* 43, 1126–1128. doi: 10.1161/ STROKEAHA.111.644716
- Sudlow, C., Martinez Gonzalez, N. A., Kim, J., and Clark, C. (2006). Does apolipoprotein E genotype influence the risk of ischemic stroke, intracerebral hemorrhage, or subarachnoid hemorrhage? Systematic review and metaanalyses of 31 studies among 5961 cases and 17,965 controls. *Stroke* 37, 364–370. doi: 10.1161/01.STR.0000199065.12908.62
- Sun, Z. (2015). Aging, arterial stiffness, and hypertension. *Hypertension* 65, 252– 256. doi: 10.1161/hypertensionaha.114.03617
- Taetzsch, T., Levesque, S., McGraw, C., Brookins, S., Luqa, R., Bonini, M. G., et al. (2015). Redox regulation of NF-kappaB p50 and M1 polarization in microglia. *Glia* 63, 423–440. doi: 10.1002/glia.22762
- Talha, K. A., Selina, F., Nasir, M., Kausar, A., Islam, T., and Perveen, R. A. (2020). Systematic Review on Apolipoprotein E: a Strong Genetic Cause of Hemorrhagic Stroke. *Mymensingh Med. J.* 29, 1026–1032.
- Tang, J., Liu, J., Zhou, C., Alexander, J. S., Nanda, A., Granger, D. N., et al. (2004). Mmp-9 deficiency enhances collagenase-induced intracerebral hemorrhage and brain injury in mutant mice. *J. Cereb. Blood Flow Metab.* 24, 1133–1145. doi: 10.1097/01.WCB.0000135593.05952.DE
- Tao, C., Hu, X., Li, H., and You, C. (2017). White Matter Injury after Intracerebral Hemorrhage: Pathophysiology and Therapeutic Strategies. Front. Hum. Neurosci. 11:422. doi: 10.3389/fnhum.2017.00422
- Tao, C., Zhang, R., Hu, X., Song, L., Wang, C., Gao, F., et al. (2016). A Novel Brainstem Hemorrhage Model by Autologous Blood Infusion in Rat: white Matter Injury, Magnetic Resonance Imaging, and Neurobehavioral Features. *J. Stroke Cerebrovasc. Dis.* 25, 1102–1109. doi: 10.1016/j.jstrokecerebrovasdis. 2016.01.025
- Tarantini, S., Valcarcel-Ares, N. M., Yabluchanskiy, A., Springo, Z., Fulop, G. A., Ashpole, N., et al. (2017). Insulin-like growth factor 1 deficiency exacerbates hypertension-induced cerebral microhemorrhages in mice, mimicking the aging phenotype. *Aging Cell* 16, 469–479. doi: 10.1111/acel.12583

- Tarantini, S., Yabluchanskiy, A., Lindsey, M. L., Csiszar, A., and Ungvari, Z. (2021). Effect of genetic depletion of MMP-9 on neurological manifestations of hypertension-induced intracerebral hemorrhages in aged mice. *Geroscience* 43, 2611–2619. doi: 10.1007/s11357-021-00402-5
- Taylor, T. N., Davis, P. H., Torner, J. C., Holmes, J., Meyer, J. W., and Jacobson, M. F. (1996). Lifetime cost of stroke in the United States. *Stroke* 27, 1459–1466. doi: 10.1161/01.str.27.9.1459
- Tessier, P. A., Naccache, P. H., Clark-Lewis, I., Gladue, R. P., Neote, K. S., and McColl, S. R. (1997). Chemokine networks in vivo: involvement of C-X-C and C-C chemokines in neutrophil extravasation in vivo in response to TNF-alpha. *J. Immunol.* 159, 3595–3602.
- Tichauer, J. E., Flores, B., Soler, B., Eugenin-von Bernhardi, L., Ramirez, G., and von Bernhardi, R. (2014). Age-dependent changes on TGFbeta1 Smad3 pathway modify the pattern of microglial cell activation. *Brain Behav. Immun.* 37, 187–196. doi: 10.1016/j.bbi.2013.12.018
- Toth, P., Tarantini, S., Springo, Z., Tucsek, Z., Gautam, T., Giles, C. B., et al. (2015). Aging exacerbates hypertension-induced cerebral microhemorrhages in mice: role of resveratrol treatment in vasoprotection. *Aging Cell.* 14, 400–408. doi: 10.1111/acel.12315
- Tsushima, Y., Aoki, J., and Endo, K. (2003). Brain microhemorrhages detected on T2*-weighted gradient-echo MR images. AJNR Am. J. Neuroradiol. 24, 88–96.
- Umeano, O., Phillips-Bute, B., Hailey, C. E., Sun, W., Gray, M. C., Roulhac-Wilson, B., et al. (2013). Gender and age interact to affect early outcome after intracerebral hemorrhage. *PLoS One* 8:e81664. doi: 10.1371/journal.pone. 0081664
- van Asch, C. J., Luitse, M. J., Rinkel, G. J., van der Tweel, I., Algra, A., and Klijn, C. J. (2010a). Incidence, case fatality, and functional outcome of intracerebral haemorrhage over time, according to age, sex, and ethnic origin: a systematic review and meta-analysis. *Lancet Neurol.* 9, 167–176. doi: 10.1016/S1474-4422(09)70340-0
- van Asch, C. J., Oudendijk, J. F., Rinkel, G. J., and Klijn, C. J. (2010b). Early intracerebral hematoma expansion after aneurysmal rupture. *Stroke* 41, 2592– 2595. doi: 10.1161/STROKEAHA.110.589291
- Vernooij, M. W., van der Lugt, A., Ikram, M. A., Wielopolski, P. A., Niessen, W. J., Hofman, A., et al. (2008). Prevalence and risk factors of cerebral microbleeds: the Rotterdam Scan Study. *Neurology* 70, 1208–1214. doi: 10.1212/01.wnl. 0000307750.41970.d9
- Viswanathan, A., and Greenberg, S. M. (2011). Cerebral amyloid angiopathy in the elderly. *Ann. Neurol.* 70, 871–880.
- Wan, S., Cheng, Y., Jin, H., Guo, D., Hua, Y., Keep, R. F., et al. (2016). Microglia Activation and Polarization After Intracerebral Hemorrhage in Mice: the Role of Protease-Activated Receptor-1. *Transl. Stroke Res.* 7, 478–487. doi: 10.1007/ s12975-016-0472-8
- Wang, J., and Dore, S. (2007). Inflammation after intracerebral hemorrhage. J. Cereb. Blood Flow Metab 27, 894–908. doi: 10.1038/sj.jcbfm.9600403
- Wang, Y. C., Wang, P. F., Fang, H., Chen, J., Xiong, X. Y., and Yang, Q. W. (2013). Toll-like receptor 4 antagonist attenuates intracerebral hemorrhage-induced brain injury. *Stroke* 44, 2545–2552. doi: 10.1161/STROKEAHA.113.001038
- Ward, R. J., Zucca, F. A., Duyn, J. H., Crichton, R. R., and Zecca, L. (2014). The role of iron in brain ageing and neurodegenerative disorders. *Lancet Neurol.* 13, 1045–1060. doi: 10.1016/S1474-4422(14)70117-6
- Wasil, T., and Lichtman, S. M. (2005). Treatment of elderly cancer patients with chemotherapy. *Cancer Invest.* 23, 537–547. doi: 10.1080/07357900500202770
- Wasserman, J. K., Yang, H., and Schlichter, L. C. (2008). Glial responses, neuron death and lesion resolution after intracerebral hemorrhage in young vs. aged rats. *Eur. J. Neurosci.* 28, 1316–1328. doi: 10.1111/j.1460-9568.2008.06442.x
- Winkler, D. T., Bondolfi, L., Herzig, M. C., Jann, L., Calhoun, M. E., Wiederhold, K. H., et al. (2001). Spontaneous hemorrhagic stroke in a mouse model of cerebral amyloid angiopathy. *J. Neurosci.* 21, 1619–1627. doi: 10.1523/ JNEUROSCI.21-05-01619.2001
- Wu, J., Hua, Y., Keep, R. F., Nakamura, T., Hoff, J. T., and Xi, G. (2003). Iron and iron-handling proteins in the brain after intracerebral hemorrhage. *Stroke* 34, 2964–2969. doi: 10.1161/01.STR.0000103140.52838.45

- Xi, G., Hua, Y., Keep, R. F., Younger, J. G., and Hoff, J. T. (2001). Systemic complement depletion diminishes perihematomal brain edema in rats. *Stroke* 32, 162–167. doi: 10.1161/01.str.32.1.162
- Xi, G., Hua, Y., Keep, R. F., Younger, J. G., and Hoff, J. T. (2002a). Brain edema after intracerebral hemorrhage: the effects of systemic complement depletion. *Acta Neurochir. Suppl.* 81, 253–256. doi: 10.1007/978-3-7091-6738-0_66
- Xi, G., Keep, R. F., and Hoff, J. T. (2002b). Pathophysiology of brain edema formation. *Neurosurg. Clin. North Am.* 13, 371–383. doi: 10.1016/s1042-3680(02)00007-4
- Xiong, L., Reijmer, Y. D., Charidimou, A., Cordonnier, C., and Viswanathan, A. (2016). Intracerebral hemorrhage and cognitive impairment. *Biochim. Biophys. Acta* 1862, 939–944.
- Xu, X., Wang, B., Ren, C., Hu, J., Greenberg, D. A., Chen, T., et al. (2017). Agerelated Impairment of Vascular Structure and Functions. *Aging Dis.* 8, 590–610. doi: 10.14336/AD.2017.0430
- Yang, N. R., Kim, J. H., Ahn, J. H., Oh, J. K., Chang, I. B., and Song, J. H. (2020). Is nontraumatic intracerebral hemorrhage different between young and elderly patients? *Neurosurg. Rev.* 43, 781–791. doi: 10.1007/s10143-019-01 120-5
- Yang, S., Nakamura, T., Hua, Y., Keep, R. F., Younger, J. G., He, Y., et al. (2006a). The role of complement C3 in intracerebral hemorrhage-induced brain injury. J. Cereb. Blood Flow Metab. 26, 1490–1495. doi: 10.1038/sj.jcbfm.960 0305
- Yang, S., Nakamura, T., Hua, Y., Keep, R. F., Younger, J. G., Hoff, J. T., et al. (2006b). Intracerebral hemorrhage in complement C3-deficient mice. Acta Neurochir. Suppl. 96, 227–231. doi: 10.1007/3-211-30714-1_49
- Yuan, J. J., Zhang, Q., Gong, C. X., Wang, F. X., Huang, J. C., Yang, G. Q., et al. (2019). Young plasma ameliorates aging-related acute brain injury after intracerebral hemorrhage. *Biosci. Rep.* 39:BSR20190537. doi: 10.1042/ BSR20190537
- Zecca, L., Stroppolo, A., Gatti, A., Tampellini, D., Toscani, M., Gallorini, M., et al. (2004). The role of iron and copper molecules in the neuronal vulnerability of locus coeruleus and substantia nigra during aging. *Proc. Natl. Acad. Sci. U. S. A.* 101, 9843–9848. doi: 10.1073/pnas.0403495101
- Zhang, Y., Yang, Y., Zhang, G. Z., Gao, M., Ge, G. Z., Wang, Q. Q., et al. (2016). Stereotactic Administration of Edaravone Ameliorates Collagenase-Induced Intracerebral Hemorrhage in Rat. CNS Neurosci. Ther. 22, 824–835. doi: 10. 1111/cns.12584
- Zhao, X., Zhang, Y., Strong, R., Zhang, J., Grotta, J. C., and Aronowski, J. (2007). Distinct patterns of intracerebral hemorrhage-induced alterations in NF-kappaB subunit, iNOS, and COX-2 expression. J. Neurochem. 101, 652–663. doi: 10.1111/j.1471-4159.2006.04414.x
- Zia, E., Engstrom, G., Svensson, P. J., Norrving, B., and Pessah-Rasmussen, H. (2009). Three-year survival and stroke recurrence rates in patients with primary intracerebral hemorrhage. *Stroke* 40, 3567–3573. doi: 10.1161/STROKEAHA. 109.556324

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