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## Parallel paleogenomic transects reveal complex genetic history of early European farmers

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### Abstract

Ancient DNA studies have established that Neolithic European populations were descended from Anatolian migrants<sup>1–8</sup> who received a limited amount of admixture from resident hunter-gatherers<sup>3–5,9</sup>. Many open questions remain, however, about the spatial and temporal dynamics of population interactions and admixture during the Neolithic period. Using the highest-resolution genome-wide ancient DNA data set assembled to date—a total of 180 samples, 130 newly reported here, from the Neolithic and Chalcolithic of Hungary (6000–2900 BCE,  $n = 100$ ), Germany (5500–3000 BCE,  $n = 42$ ), and Spain (5500–2200 BCE,  $n = 38$ )—we investigate the population dynamics of Neolithization across Europe. We find that genetic diversity was shaped predominantly by local processes, with varied sources and proportions of hunter-gatherer ancestry among the three regions and through time. Admixture between groups with different ancestry profiles was pervasive and resulted in observable population transformation across almost all cultural transitions. Our results shed new light on the ways that gene flow reshaped European populations throughout the Neolithic period and demonstrate the potential of time-series-based sampling and modeling approaches to elucidate multiple dimensions of historical population interactions.

The population dynamics of the Neolithization process are of great importance for understanding European prehistory<sup>10–13</sup>. The first quantitative model of the Neolithic transition to integrate archaeological and genetic data was the demic diffusion hypothesis<sup>10</sup>, which posited that growing population densities among Near Eastern farmers led to a range expansion that spread agriculture to Europe. Ancient DNA analysis has validated major migrations from populations related to Neolithic Anatolians as driving the introduction of farming in Europe<sup>1–8</sup>, but the demic diffusion model does not account for the complexities

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### Author contributions

A.S.-N., J.B., E.B., K.W.A., C.L.-F., W.H., and D.R. designed and supervised the study. B.G.M., K.K., K.O., M.B., T.M., A.O., J.J., T.P., F.H., P.C., J.K., K.Se., A.A., P.R., J.R., J.P.B., S.F., G.S., Z.T., E.G.N., J.D., E.M., G.P., L.M., B.M., Z.B., L.D., J.F.-E., J.A.M.-A., C.A.F., J.J.E., R.B., J.O., K.Sc., H.M., A.C., J.B., E.B., K.W.A., C.L.-F., and W.H. provided samples and assembled archaeological and anthropological information. A.S.-N., A.P., B.S., V.K., N.R., K.St., M.F., M.M., J.O., N.B., E.H., S.N., and B.L. performed laboratory work. M.L., A.S.-N., S.M., and D.R. analyzed genetic data. M.L., A.S.-N., and D.R. wrote the manuscript with input from all coauthors.

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of the interactions between farmers and hunter-gatherers in Europe throughout the Neolithic<sup>11–16</sup>. For example, ancient DNA has shown that farmers traversed large portions of Europe with limited initial admixture from hunter-gatherers<sup>3,5,7,8</sup>, and furthermore that farmers and hunter-gatherers lived in close proximity in some locations long after the arrival of agriculture<sup>15,16</sup>. However, genetic data have yet to be used systematically to model the population interactions and transformations during the course of the Neolithic period. Key open questions include whether migrating farmers mixed with hunter-gatherers at each stage of the expansion (and if so how soon after arriving) and whether the previously observed increase in hunter-gatherer ancestry among farmers in several parts of Europe by the Middle Neolithic<sup>5–9</sup> represented a continuous versus discrete process and a continent-wide phenomenon versus a collection of parallel, local events.

We compiled a high-resolution data set of 180 Neolithic and Chalcolithic European genomes (pre-dating the arrival of steppe ancestry in the third millennium BCE [ref 5]) from what are now Hungary, Germany, and Spain, of which 130 individuals are newly reported here, 45 with new direct radiocarbon dates (Table 1; Fig. 1A, B; Extended Data Tables 1, 2; Supplementary Tables 1, 2; Supplementary Information sections 1–3). We enriched for DNA fragments covering a set of ~1.23 million single nucleotide polymorphism (SNP) targets<sup>7</sup> and called one allele at random per site, obtaining largely high-quality data, with at least 100,000 SNPs hit at least once (average coverage ~0.1 or higher) for 90 of the 130 samples (Methods). The majority (90) of our new samples comprise an approximately 3000-year transect of the prehistory of the Carpathian Basin (Supplementary Information section 1), from both the eastern (Great Hungarian Plain, or Alföld) and western (Transdanubia) portions of present-day Hungary. For our primary analyses, we retained 104 samples from 15 population groupings (Methods; Table 1), which we merged with 50 Neolithic individuals from the literature<sup>4,5,7,17,18</sup>. We co-analyzed these samples with 25 Neolithic individuals (~6500–6000 BCE) from northwestern Anatolia<sup>7</sup> to represent the ancestors of the first European farmers (FEF; Supplementary Information section 4) and four primary European hunter-gatherer individuals<sup>4,7,17,19,20</sup> (“WHG,” western hunter-gatherers; Table 1).

A principal component analysis (PCA) of our samples shows that, as expected, all of the Neolithic individuals fall along a cline of admixture between FEF and WHG (Extended Data Fig. 1). Y-chromosome diversity also indicates contributions from ancestral Anatolian farmer and local hunter-gatherer populations, dominated by haplogroups G and I (the latter especially common in Iberia; Supplementary Information section 3). The European populations are consistent with a common origin in Anatolia (Supplementary Information section 4), reflected in the low differentiation among EN groups in the PCA. Over the course of the Neolithic, we observe a trend of increasing hunter-gatherer ancestry in each region, although at a slower rate in Hungary than in Germany and Spain, and with limited intra-population heterogeneity (Fig. 2A; Supplementary Information section 6). We also find that this hunter-gatherer ancestry is more similar to the eastern WHG individuals (KO1 and VIL) farther east and more similar to the western WHG individuals (LB1 and LOS) farther west (Fig. 2B). While this pattern does not demonstrate directly where mixture between hunter-gatherers and farmers took place, it suggests, given that European hunter-gatherers display a strong correlation between genetic and geographic structure (Fig. 1D), that hunter-gatherer

ancestry in farmers was to a substantial extent derived from populations in relatively close proximity.

To analyze admixed hunter-gatherer ancestry more formally, we modeled Neolithic farmers in an admixture graph framework. We started with a “scaffold” model (Extended Data Fig. 2) consisting of Neolithic Anatolians, the four reference WHG individuals, and two outgroups (Mbuti and Kostenki 14 [refs 20, 22]), with significant signals of admixture in LB1 and KO1 (Supplementary Information sections 5–6). We then added each Neolithic population to this model in turn, fitting them as a mixture of FEF and either one or two hunter-gatherer ancestry components. To check for robustness, we repeated our analyses using transversions or outgroup-ascertained SNPs only, with in-solution capture data for LOS, and with additional or alternative hunter-gatherers in the model (Extended Data Table 3; Supplementary Information section 6), and in all cases the results were qualitatively consistent. We find that almost all ancient groups from Hungary have ancestry significantly closest to one of the more eastern WHG individuals (KO1 or VIL); the samples from present-day Germany have the greatest affinity to LOS; and all three Iberian groups contain LB1-related ancestry (Fig. 2C; Extended Data Table 3). This pattern implies that admixture into European farmers occurred multiple times from local hunter-gatherer populations. Moreover, combining the proportions and sources of hunter-gatherer ancestry, populations from the three regions are distinguishable at all stages of the Neolithic. Thus, any further long-range migrations that may have occurred after the initial spread of agriculture in the studied regions (and before large incursions from the steppe) were not substantial enough to homogenize the ancestry of farming populations.

Additional insights about population interactions can be gained by studying the dates of admixture events. We used *ALDER* (ref. 23) to estimate dates of admixture for Neolithic individuals based on the recombination-induced breakdown of contiguous blocks of FEF and WHG ancestry over time (Extended Data Tables 1, 2, 4; Extended Data Fig. 3). The *ALDER* algorithm is not able to accommodate large amounts of missing data, so we developed a strategy for running it with the relatively low coverage of ancient DNA (Supplementary Information section 7). The dates we obtain (Fig. 2D) are based on a model of a single wave of admixture, which means that if the true history for a population includes multiples waves or continuous admixture, we will obtain an intermediate value. In particular, for later populations, this history could include mixture with previously admixed groups (either farmers with substantially different hunter-gatherer ancestry proportions or hunter-gatherers with farmer ancestry).

For our most complete time series, from Hungary, we infer admixture dates throughout the Neolithic that are on average mostly 18–30 generations old (500–840 years), indicating ongoing population transformation and admixture (Fig. 2D; Extended Data Table 4). This pattern is accompanied by a gradual increase in hunter-gatherer ancestry over time, although never reaching the levels observed in MN Germany or Iberia (Fig. 2A). While the majority of the EN samples from Hungary do not have significantly more hunter-gatherer ancestry than Neolithic Anatolians (Fig. 2A; Extended Data Tables 1, 2), one Star evo individual, BAM17b, is inferred to have  $7.8 \pm 1.7\%$  hunter-gatherer ancestry and a very recent *ALDER* date of  $4.5 \pm 1.9$  generations ( $5865 \pm 65$  BCE;  $1.9 \pm 0.9$  generations using a group-level

estimate; Extended Data Table 4), consistent with having one or two hunter-gatherer ancestors in the past few generations. Additionally, one newly sampled Körös individual, TIDO2a, is similar to KO1 in having ~80% WHG and ~20% FEF ancestry and an *ALDER* date of  $16.1 \pm 3.8$  generations, reinforcing the distinctive heterogeneity of the Tiszasz 1 site, the source for both TIDO2a and KO1. We also infer an average admixture date of  $5675 \pm 55$  BCE for the ALPc MN, again suggesting that in Hungary, interaction between Anatolian migrants and local hunter-gatherers began in the Early Neolithic (cf. refs 14, 24–26). The greatest differences between Alföld and Transdanubia are observed in the MN, with substantially more hunter-gatherer ancestry in ALPc than LBKT (Fig. 2; Extended Data Table 3), and overall, we observe slight trends toward more hunter-gatherer ancestry to the north and east (Extended Data Fig. 4), as expected based on the greater archaeological evidence of hunter-gatherer settlement and interactions<sup>24</sup>. By the LN and CA, however, and especially in the Baden period (when the region became culturally unified<sup>27</sup>), our results are broadly similar over the two halves of present-day Hungary.

From Germany, we analyzed a large sample of the EN LBK culture and 11 individuals from the MN period, four of them from the Blätterhöhle site, which has been shown to have featured a combination of farmer and hunter-gatherer occupation to a relatively late date<sup>15</sup>. The average date of admixture for LBK ( $5545 \pm 65$  BCE) is more recent than the dates for EN/MN populations from Hungary, and the total hunter-gatherer ancestry proportion in LBK (~4–5%) is intermediate between LBKT and ALPc. This ancestry is most closely related to a combination of KO1 and LOS, although the assignment of the hunter-gatherer source(s) is not statistically significant (Fig. 2B; Extended Data Table 3). These results are consistent with genetic and archaeological evidence for LBK origins from the early LBKT (ref. 26), followed by additional, Central European WHG admixture after about 5500 BCE. Our “Germany MN” grouping shows increased hunter-gatherer ancestry (~17%, most closely related to LOS) and a more recent average date of admixture, reflecting gene flow from hunter-gatherers after the LBK period. We successfully sequenced a total of 17 Blätterhöhle MN samples, many of them with distinct individual labels from ref. 15, although surprisingly, the genome-wide data indicated that these corresponded to only four unique individuals (Supplementary Information section 8), for which we merged libraries to increase coverage. In accordance with previous results<sup>15</sup>, we find that the three farmer individuals (classified based on stable isotopes) harbored 40–50% hunter-gatherer ancestry, while Bla8, who showed signatures associated with a hunter-gatherer-fisher lifestyle, was closer genetically to hunter-gatherers but was also admixed, with ~27% ancestry from farmers. Our results thus provide evidence of asymmetric gene flow between farmers and hunter-gatherers at Blätterhöhle centered around the relatively late date of ~4000 BCE (*ALDER* dates of 10–25 generations).

In Iberia, we again see widespread evidence of local hunter-gatherer admixture, with confidently inferred LB1-related ancestry in all three population groups (EN, MN, and CA). For Iberia EN, we infer an average admixture date of  $5650 \pm 65$  BCE, which rises to  $5860 \pm 110$  BCE when considering only the five oldest samples (of which the earliest, CB13 [ref. 18] has an individual estimate of  $5890 \pm 105$  BCE). Given that farming is thought to have begun in Spain around 5500 BCE (ref. 28), these dates suggest the presence of at least a small proportion of hunter-gatherer ancestry in earlier Cardial Neolithic populations

acquired along their migration route (although our admixture graph analysis only confidently detected an LB1-related component). The later Iberians have large proportions of hunter-gatherer ancestry, approximately 23% for MN (from the site of La Mina, in north-central Iberia) and 27% for CA, and also relatively old *ALDER* dates (approximately 50 generations, or 1400 years), indicating that most of the admixture occurred well before their respective sample dates. Both populations have evidence of ancestry related to a different WHG individual in addition to LB1 (Fig. 2C; Extended Data Table 3), suggesting a non-local source for at least some of the hunter-gatherer ancestry gained between the EN and MN.

Synthesizing our time series data, we compared the observed *ALDER* dates and hunter-gatherer ancestry proportions of Neolithic populations to those estimated for simulated data under different temporal admixture scenarios (Fig. 3; Extended Data Fig. 5; Supplementary Information section 9). We assumed dates of 5900 BCE (Hungary) or 5500 BCE (Germany and Spain) for the onset of mixture. While none of the scenarios match the data perfectly, a good fit for Hungary is provided by a model (bottom solid green curve in both panels of Fig. 3) of an initial admixture pulse (approximately 1/4 of the total hunter-gatherer ancestry observed by the end of the time series) followed by continuous gene flow. By contrast, scenarios such as a single admixture pulse or continuous mixture decreasing by 5% or more per generation provide too much hunter-gatherer ancestry at early dates. Alföld and Transdanubia should be considered as separate series, but their parameters follow mostly similar trajectories, with the exception of the MN, where LBKT has a relatively old admixture date (albeit with large uncertainty) and ALPc a relatively high hunter-gatherer ancestry proportion (possibly influenced by the bias of sampling in favor of the middle and northern parts of the Alföld). Considering the other regions, even after normalizing for the different total hunter-gatherer ancestry proportions, we observe a high degree of local distinctiveness, for example in the older *ALDER* dates for Iberia MN/CA and the markedly higher hunter-gatherer ancestry in Blätterhöhle (Extended Data Fig. 5). We note that while the simulated data are generated under a model of gene flow from an unadmixed hunter-gatherer source population into a series of farmer populations in a single line of descent, observed admixture could also be influenced by flow in the other direction (from farmers to hunter-gatherers) or could reflect immigration of new farmer populations (either via their own previous hunter-gatherer admixture or new admixture between farming populations with different proportions of hunter-gatherer ancestry). Based on archaeological evidence, such a scenario is possible, for example, for the introduction of hunter-gatherer ancestry into TDLN from Southeastern European farmers via the dispersal of the northern Balkan Vin a or Sopot cultures to Transdanubia<sup>14,29,30</sup>.

Our results provide greatly increased detail in understanding population interactions and admixture during the European Neolithic. In each of our three study regions, the arrival of farmers prompted admixture with local hunter-gatherers, which unfolded over many centuries: almost all sampled populations have more hunter-gatherer ancestry and more recent dates of admixture than their local predecessors, suggesting recurrent changes in genetic composition and significant hunter-gatherer gene flow beyond initial contact. These transformations left distinct signatures in each region, implying that they resulted from a complex web of local interactions rather than a uniform demographic phenomenon. Our

transect of Hungary, in particular, with representative samples from many archaeological cultures across the region and throughout the Neolithic and Chalcolithic, illustrates the power of dense ancient DNA time series. Future work with continually improving data sets and statistical models promises to yield many more insights about historical population transformations in space and time.

## Methods

### Experimental procedures

Prehistoric teeth and petrous bone samples from Hungary were taken under sterile conditions in the Hungarian Museums and anthropological collections. Samples other than Gorzsa were documented, cleaned, and ground into powder either in the Anthropological Department of the Johannes Gutenberg University of Mainz, during the course of the German Research Foundation project AL 287-10-1, or in Budapest, in the Laboratory of Archaeogenetics of the Institute of Archaeology, Research Centre for the Humanities, Hungarian Academy of Sciences, following published protocols<sup>26</sup>. DNA was extracted in Budapest using 0.08–0.11 g powder via published methods<sup>31</sup>, using High Pure Viral NA Large Volume Kit columns (Roche)<sup>32,33</sup>. DNA extractions were tested by PCR, amplifying the 16117–16233 bp fragment of the mitochondrial genome, and visualized on a 2% agarose gel. DNA libraries were prepared from clean and successful extraction batches using UDG-half and no-UDG treated methods<sup>5,34</sup>. We included milling (hydroxylapatite blanks to control for cleanness) and extraction negative controls in every batch. Bar-code adapter ligated libraries were amplified with TwistAmp Basic (Twist DX Ltd), purified with Agencourt AMPure XP (Beckman Coulter), and checked on 3% agarose gel<sup>5</sup>. Library concentration was measured on a Qubit 2.0 fluorometer. Promising libraries after initial quality control analysis were shipped to Harvard Medical School, where further processing took place. All other samples were prepared similarly in dedicated clean rooms at Harvard Medical School and the University of Adelaide in accordance with published methods<sup>5,7,33</sup>. For samples LHUE2010.11 (one library) and MIR202-037-n105, we used magnetic bead cleanups instead of MinElute column cleanups between enzymatic reactions with magnetic bead cleanups and SPRI bead cleanup instead of the final PCR cleanup<sup>35,36</sup>.

We initially screened the libraries via in-solution hybridization to a set of probes targeting mitochondrial DNA (mtDNA)<sup>37</sup> plus roughly 3000 nuclear SNP targets, using a protocol described previously<sup>5,33</sup> with amplified baits synthesized by CustomArray, Inc. Libraries with good screening results—limited evidence of contamination, reasonable damage profiles, and substantial coverage on targeted segments—were enriched for a genome-wide set of ~1.2 million SNPs<sup>7,33</sup> and sequenced to greater depth. Raw sequencing data were processed by trimming bar-codes and adapters, merging read pairs with at least 15 base pairs of overlapping sequence, and mapping to the human reference genome (version hg19). Reads were filtered for mapping and base quality, duplicate molecules were removed, and two terminal bases were clipped to eliminate damage (five for UDG-minus libraries)<sup>5</sup>. All libraries had a rate of at least 4.8% C-to-T substitutions in the final base of screening sequencing reads (Supplementary Table 1), consistent with damage patterns expected for authentic ancient DNA (refs 34, 38). Pseudo-haploid genotypes at each SNP were called by

choosing one allele at random from among mapped reads. Sex determinations for each individual were made by manually examining the fractions of reads mapping to the X and Y chromosomes and imposing thresholds for males and females (with any indeterminate samples labeled as unknown).

Mitochondrial DNA sequences were reassembled in Geneious R10 to rCRS (ref. 39) and RSRG (ref. 40), and SNPs with at least 3× coverage and a minimum variant frequency of 0.7 were called. The assembly and the resulting list of SNPs were double-checked against [phylotree.org](http://phylotree.org) (mtDNA tree Build 17; 18 Feb 2016). Haplotype calls are given in Extended Data Tables 1 and 2 and Supplementary Table 2. On the Y chromosome, 15,100 SNPs were targeted and sequenced, and the detected derived and ancestral alleles were compared to the ISOGG Y-tree ([www.isogg.org](http://www.isogg.org)) version 12.34, updated on 5th February 2017. Haplogroup definitions are detailed in Supplementary Information section 3.

We merged libraries from the same individual (for those with more than one) and then combined our new samples with genome-wide data from the literature (ancient individuals as described and as listed in Extended Data Tables 1 and 2 and present-day individuals from the SGDP [ref. 41]) using all autosomal SNPs (~1.15 million) from our target set. For two replications of our admixture graph analyses, we restricted either to the subset of transversions (~280K SNPs) or to the subset from panels 4 and 5 of the Affymetrix Human Origins array (ascertained as heterozygous in a San or Yoruba individual; ~260K SNPs). For PCA (Extended Data Fig. 1), we merged with a large set of present-day samples<sup>33</sup> and used all autosomal Human Origins SNPs (~593K).

To test for possible contamination, we used contamMix (ref. 42) and ANGSD (ref. 43) to estimate rates of apparent heterozygosity in haploid genome regions (mtDNA and the X chromosome in males, respectively). Any samples with > 5% mtDNA mismatching or > 2% X contamination were excluded from further analyses, with the exception of Bla5 (Supplementary Information section 8). We also removed samples identified as clear outliers in PCA or with significant population genetic differences between all sequencing data and genotypes called only from sequences displaying ancient DNA damage signatures. A total of 19 samples were excluded based on one of these criteria. For individual-level  $f$ -statistic analyses (Fig. 2A–B), we restricted to samples with a maximum level of uncertainty, defined as a standard error of at most  $7 \times 10^{-4}$  for the statistic  $f_4(\text{Mbuti, WHG; Anatolia, X})$ . This threshold (corresponding to an average coverage of approximately 0.05, or ~60K SNPs hit at least once) was met by 89 of the 112 samples passing QC (and 49 of the 50 samples from the literature). We did not impose such a threshold for *ALDER* analyses, but because low coverage results in a weaker signal, only one of the 23 high-uncertainty individuals in our primary data set provided an *ALDER* date (as compared to 89 of the 130 low-uncertainty individuals).

### Population assignments

In most cases, population groupings were used that correspond to archaeological culture assignments based on chronology, geography, and material culture traits. Occasionally, we merged populations that appeared similar genetically in order to increase power: we pooled samples from all phases and groups of the eastern Hungarian MN into a single ALPc

population; merged six Sopot with eight Lengyel individuals for the western Hungarian TDLN; combined one Hunyadhalom (Middle CA from the Danube-Tisza interfluvium in central Hungary) with Lasinja; pooled four LBK samples from Stuttgart with the majority from farther to the northeast (primarily Halberstadt); and merged several cultures of the German MN into a single group. Other populations vary in their degrees of date and site heterogeneity, with Iberia MN the most homogeneous and Iberia EN and CA among the least (Extended Data Tables 1, 2; Supplementary Table 1). For our main analyses, we excluded the Vinča and Tiszapolgár population groups because they lacked sufficient high-quality data.

We note that the designations EN, MN, LN, and CA have different meanings in different areas. For our study regions, each term generally refers to an earlier period in Hungary than in Germany and Spain (for example, ALPc and LBKT MN in Hungary are roughly contemporaneous with LBK and Iberia EN). In order to maintain agreement with the archaeological literature, we use the established definitions, with the appropriate word of caution that they should be treated separately in each region.

### Sample dates

We report 52 newly obtained accelerator mass spectrometry (AMS) radiocarbon dates for Neolithic individuals (45 direct, 7 indirect), focusing on representative high-quality samples from each site and any samples with chronological uncertainty. These are combined with 58 radiocarbon dates from the literature<sup>4,5,7,17,18,26,29,30,44,45</sup>. We report the 95.4% calibrated confidence intervals (CI) from OxCal (ref. 46) version 4.2 with the IntCal13 calibration curve<sup>47</sup> in Extended Data Tables 1 and 2. For use in *ALDER* analyses (Supplementary Information section 7), we use the mean and standard deviation of the calibrated date distributions; while the distributions are non-normal, we find that on average the mean plus or minus two standard deviations contains more than 95.4% of the probability density. For samples without direct radiocarbon dates but with dates from other samples or materials at the same site, we form a conservative 95.4% CI by taking the minimum and maximum bounds of any of the calibrated CIs from the site. Finally, for the remaining samples, we use plausible date ranges based on archaeological context; we assume independence across individuals but as a result take a conservative approach and treat the assigned range as  $\pm$  one standard error (e.g., an estimated range of 4800–4500 BCE becomes  $4650 \pm 150$  BCE).

### Population genetic analyses

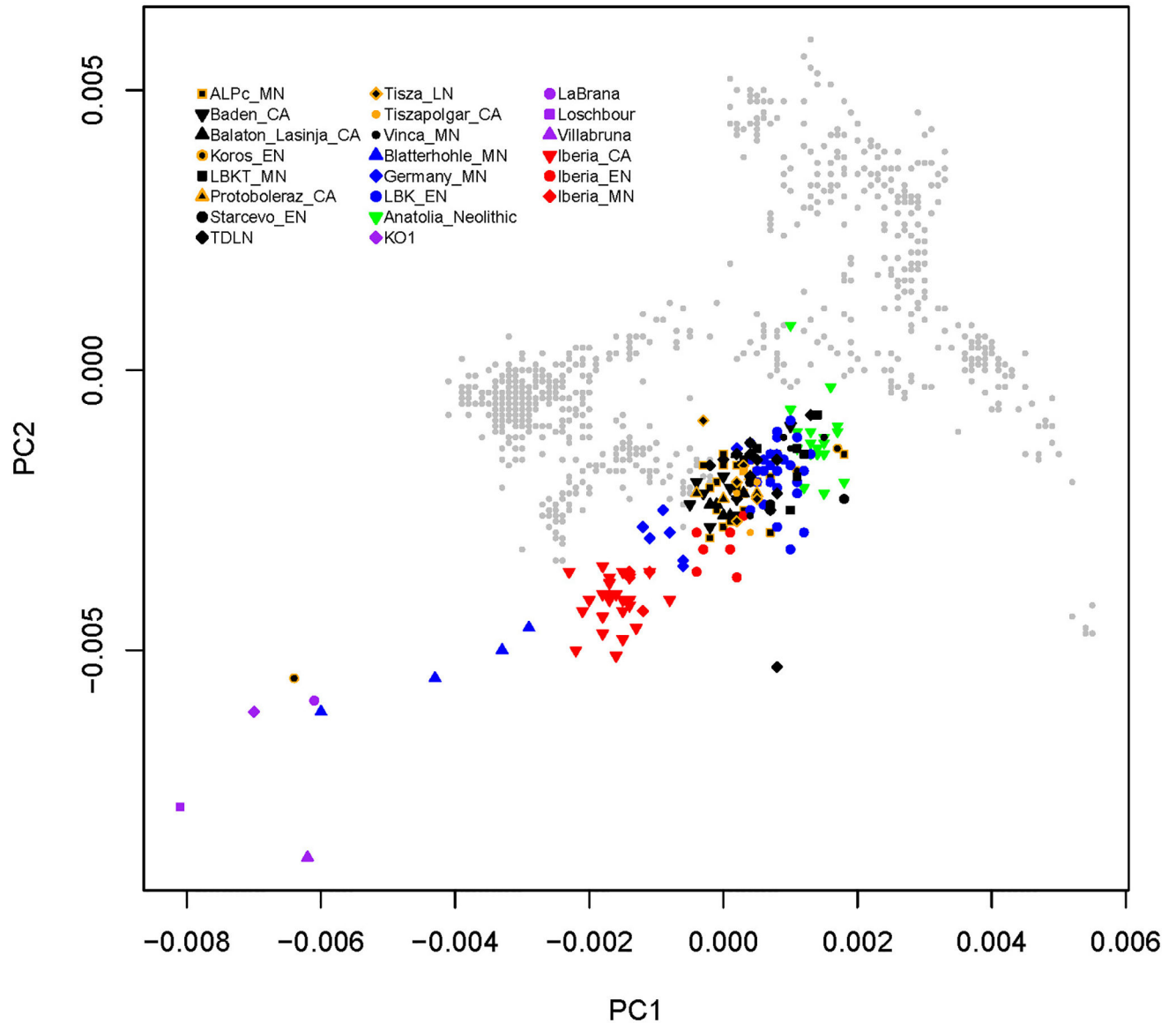
We performed PCA by computing components for present-day populations and then projecting ancient individuals using the “lsqproject” and “shrinkmode” options in smartpca (ref. 48). Admixture graphs and f-statistics were implemented through ADMIXTOOLS (ref. 49). To obtain calendar dates of admixture, we combine the *ALDER* results (in generations in the past) with the ages of the Neolithic individuals, assuming an average generation time of 28 years<sup>50,51</sup>. All analytical procedures are described in full detail in Supplementary Information sections 4–9.



### Data availability

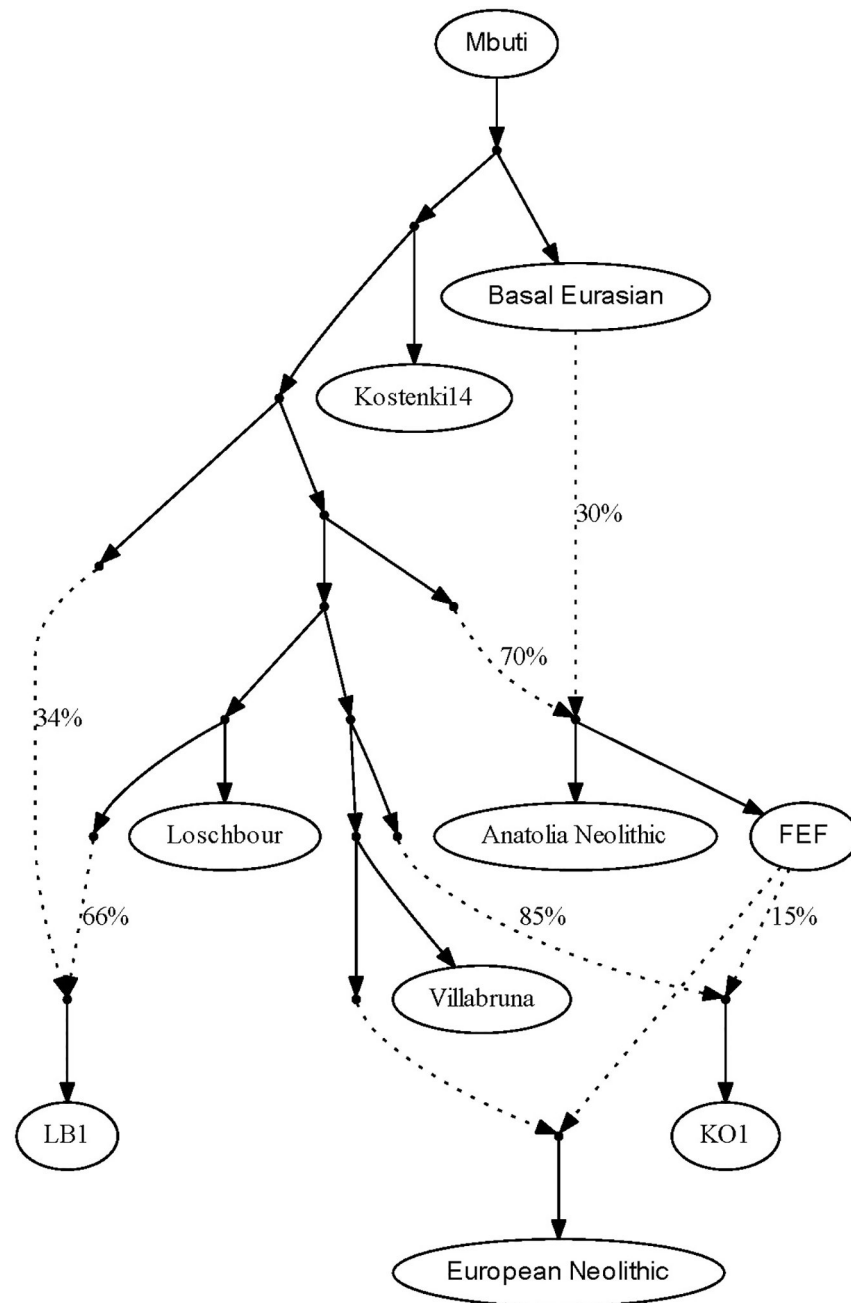
The aligned sequences are available through the European Nucleotide Archive under accession number PRJEB22629. Genotype datasets used in analysis are available at <https://reich.hms.harvard.edu/datasets>.

### Extended Data



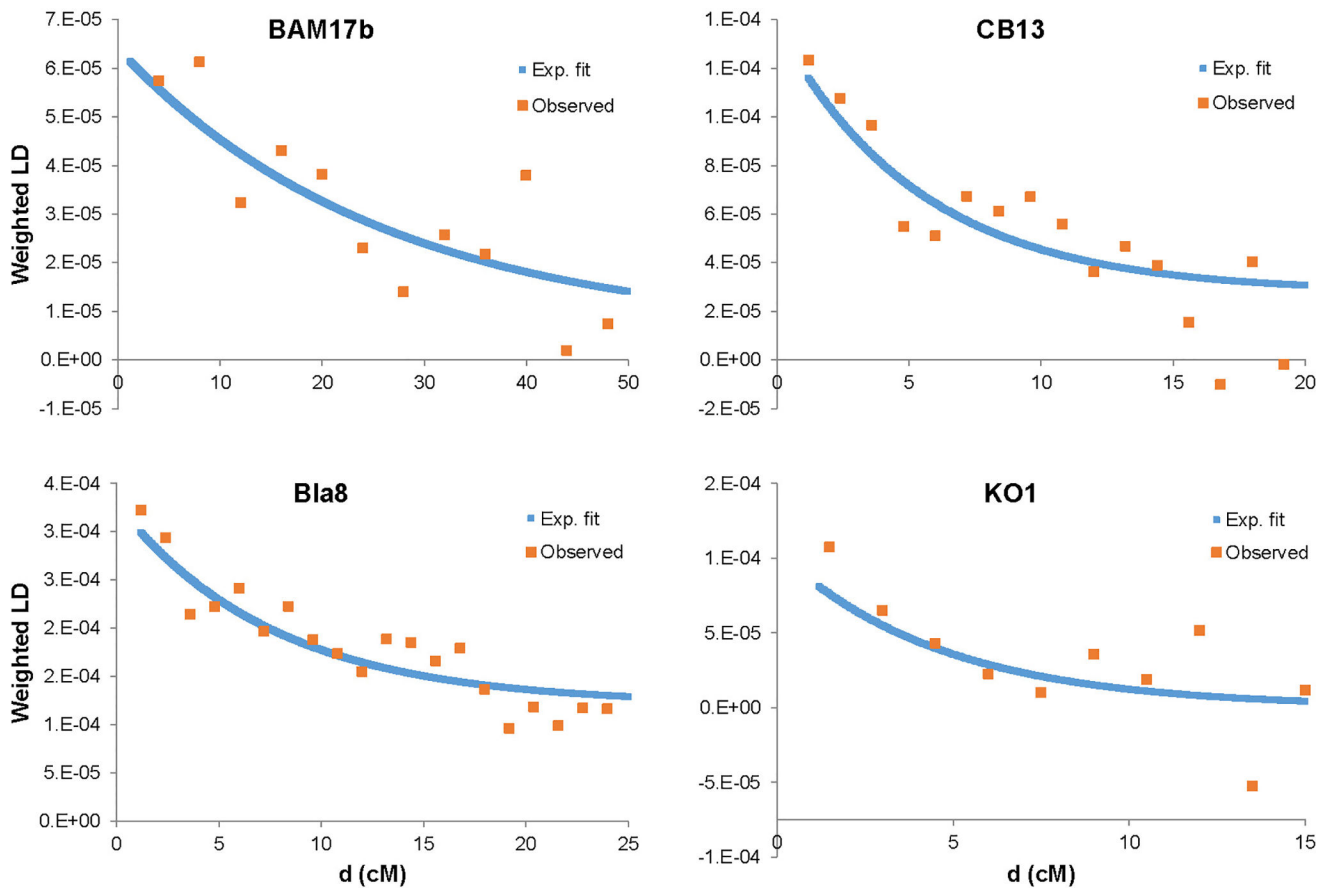
#### Extended Data Figure 1. First two principal components from PCA

We computed PCs for a set of 782 present-day western Eurasian individuals genotyped on the Affymetrix Human Origins array (background gray points) and then projected ancient individuals onto these axes. Shown is a closeup omitting the present-day Bedouin population.



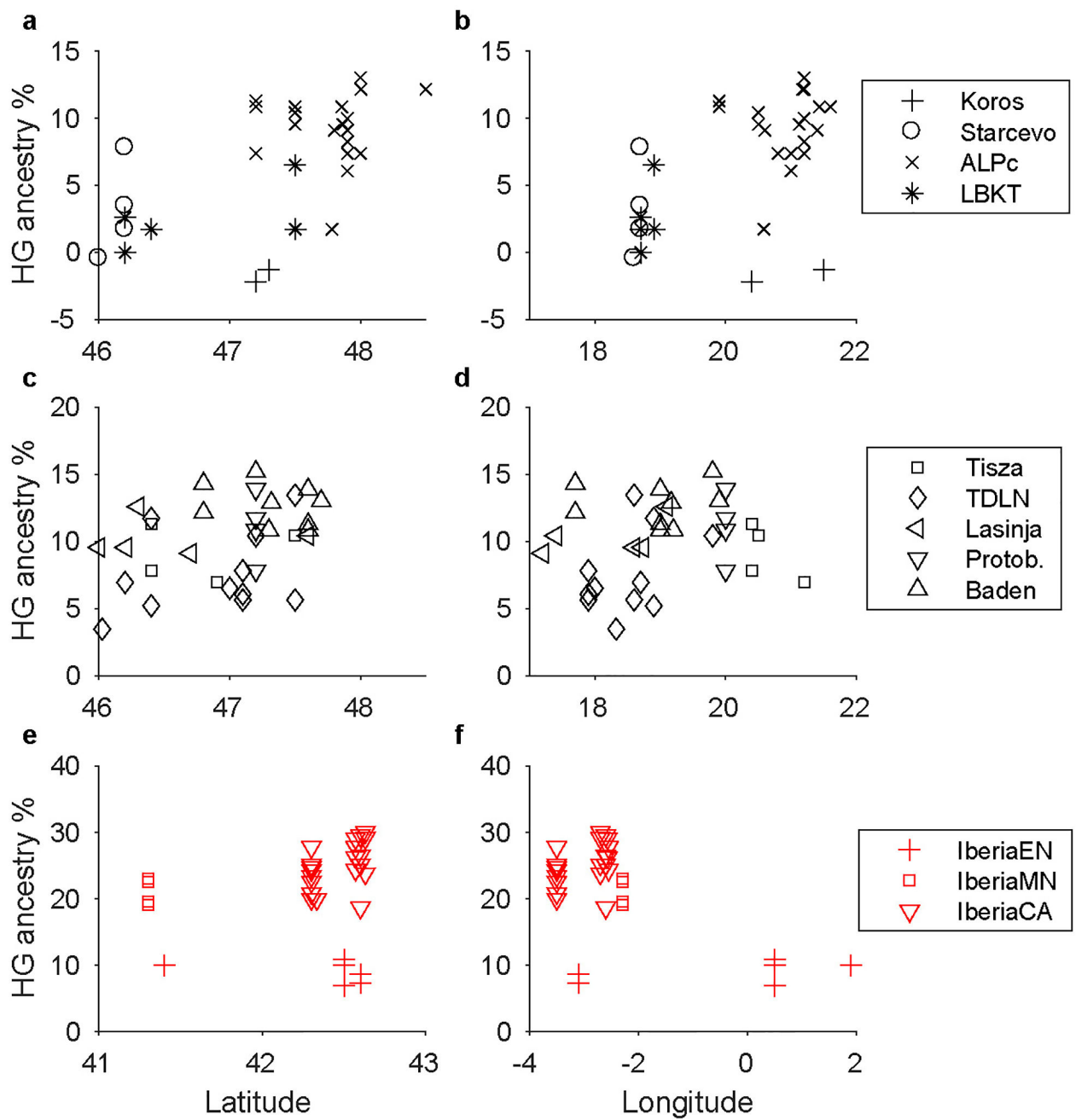
**Extended Data Figure 2. Scaffold admixture graph used for modeling European Neolithic populations**

Dotted lines denote admixture events. Neolithic Anatolians, LB1, and KO1 are modeled as admixed, with Basal Eurasian ancestry, deeper European hunter-gatherer ancestry, and FEF ancestry, respectively. European test populations are fit as a mixture of FEF and ancestry related to one or two of the four WHG individuals (here VIL-related as an example). See Supplementary Information section 6 for full details.



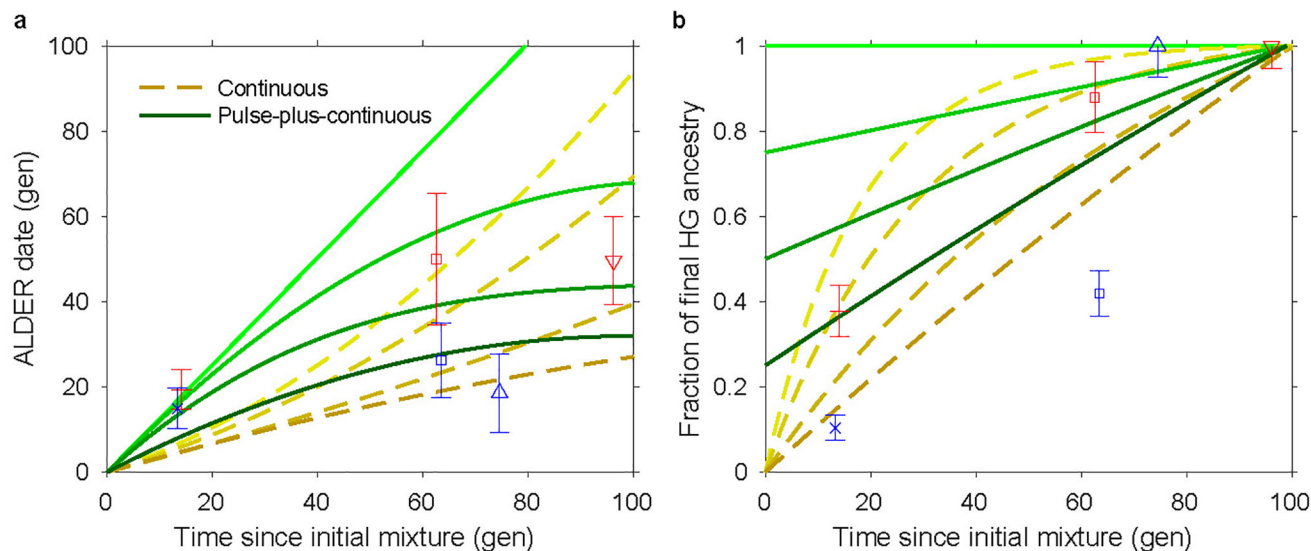
**Extended Data Figure 3. Examples of *ALDER* weighted LD decay curves**

Weighted LD is shown as a function of genetic distance  $d$ , using Neolithic Anatolians and WHG as references, for four individuals: BAM17b (Star evo EN), CB13 (Iberia EN), Bla8 (Blätterhöhle hunter-gatherer), and KO1. The results shown here use helper individuals M11–363 (Neolithic Anatolian), L11–322 (Neolithic Anatolian), BIC, and LB1, respectively, and have fitted dates (blue curves) of  $3.8 \pm 1.2$ ,  $18.3 \pm 6.0$ ,  $13.1 \pm 2.7$ , and  $21.6 \pm 8.8$  generations (compared to final individual-level dates of  $4.5 \pm 1.9$ ,  $17.5 \pm 3.5$ ,  $12.1 \pm 2.9$ , and  $21.0 \pm 7.0$  generations; see Supplementary Information section 7). Note different x-axis scales for the four individuals.



**Extended Data Figure 4. Hunter-gatherer ancestry as a function of latitude and longitude for Neolithic individuals**

**a, b, EN/MN Hungary. c, d, LN/CA Hungary. e, f, Iberia. Protob., Protoboleráz.**



### Extended Data Figure 5. Germany and Iberia time series and simulated data

**a**, Dates of admixture. **b**, Hunter-gatherer ancestry proportions, normalized by the total in the most recent (rightmost) population. Symbols are as in Figs 1 and 2, here showing population-level averages plus or minus two standard errors. Yellow dashed lines represent continuous admixture simulations: from top to bottom, diminishing 5% per generation, diminishing 3%, diminishing 1%, and uniform. Green solid lines represent pulse-plus-continuous admixture simulations: from top to bottom, all hunter-gatherer ancestry in a pulse at time zero; 3/4 of final hunter-gatherer ancestry in an initial pulse, followed by uniform continuous gene flow; half in initial pulse and half continuous; and 1/4 in initial pulse.

### Extended Data Table 1

Information for Neolithic individuals from Hungary.

ID	Population	Site	Lat.	Long.	Date	Sex	Mt Hap	$\gamma$ Hap	Cov.	HG%	ALD	Ref.
GEN68	Körös	Törökszentmiklós road 4 site 3	47.2	20.4	5706–5541	F	k1a	..	6.16	-2.16±1.5	0±0.0	
HUNG276, KO2	Körös	Berettyóújfalú-Morotva-liget	47.3	21.5	5713–5566	F	K1a	..	0.91	-1.49±1.6	0±0.0	[7, 17]
TIDO2a	Körös	Tiszasz 1 s-Domaháza	47.6	20.7	5736–5547	M	K1	I2a2	0.45	79.3±2.1	16±3.8	
BAM17b	Star evo	Alsónyék-Bátaszék, Mémőki telep	46.2	18.7	5832–5667	M	T1a2	H2	1.47	7.76±1.7	4.5±1.9	
BAM25	Star evo	Alsónyék-Bátaszék, Mémőki telep	46.2	18.7	5702–5536	M	N1a1a1	H2	0.22	1.62±1.9	0±0.0	[5, 7]
BAM4a	Star evo	Alsónyék-Bátaszék, Mémőki telep	46.2	18.7	5641–5547	M	K1a4	G2a2a1	0.20	3.39±2.0	0±0.0	
LGCS1a	Star evo	Lánycsók	46.0	18.6	5800–5500	M	W5	G2a2b2b1a	0.77	-0.63±1.6	0±0.0	
BAL25b	LBKT	Bátaszék-Lajvér	46.2	18.7	5208–4948	M	K1b1a	G2a2a1	2.77	0.06±1.5	0±0.0	
BOVO1b	LBKT	Böleske-Gy r svölgy	46.7	19.0	5300–4900	F	H	..	0.01	10.9±6.3	0±0.0	
BUD4a	LBKT	Budakeszi-Sz 1 skert	47.5	18.9	5300–4900	M	T1a	G2a2b2a	0.17	6.72±2.3	36±6.1	
BUD9a	LBKT	Budakeszi-Sz 1 skert	47.5	18.9	5300–4900	F	U2	..	1.10	1.87±1.6	13±5.3	
GEN18	LBKT	Alsónyék, site 11	46.2	18.7	5309–5074	M	T2c1	G2a2b2b1	1.48	2.66±1.5	35±12	
KON3	LBKT	Enese elkerül , Kóny, Proletár-dűlő, M85, site 2	47.6	17.4	5300–4900	F	T2b	..	0.03	2.79±4.0	0±0.0	
SZEH4	LBKT	Szemely-Hegyes	46.4	18.7	5207–4944	F	N1a1a1a3	..	0.07	1.88±3.0	0±0.0	[5, 7]
CEG07B	ALPc	Cegléd, site 4/1	47.2	19.9	5300–4900	M	J2b1	G2a2b2a	0.30	11.4±1.9	0±0.0	
CEG08b	ALPc	Cegléd, site 4/1	47.2	19.9	5300–4900	F	J1c1	..	0.19	11.0±2.2	23±3.0	
EBSA2a	ALPc	Ebes-Sajtgyár	47.5	21.5	5300–4900	F	K1a	..	0.05	16.2±3.1	0±0.0	
EBVO5a	ALPc	Ebes-Zsongvölgy	47.5	21.5	5300–4900	M	V1a	CT	0.04	9.25±3.3	0±0.0	

ID	Population	Site	Lat.	Long.	Date	Sex	Mt Hap	$\gamma$ Hap	Cov.	HG%	ALD	Ref.
HAJE10a	ALPc	Hajdúnánás-Eszlári út	47.9	21.4	5221–5000	M	J2b1	I	0.29	10.8±1.8	0±0.0	
HAJE7a	ALPc	Hajdúnánás-Eszlári út	47.9	21.4	5302–5057	M	K1a	I2	1.57	9.15±1.7	6.2±5.7	
HELI11a	ALPc	Hej kürt-Lidl	47.9	21.0	5209–4912	M	N1a1a1	I2a2a1b	0.99	6.01±1.8	14±2.0	
HELI2a	ALPc	Hej kürt-Lidl	47.9	21.0	5300–4900	M	U8b1b	I	0.09	7.39±2.6	4.4±1.7	
HUNG302, NE2	ALPc	Debrecen Tocopart Erdoalja	47.5	21.6	5291–5056	F	H	.	4.88	11.0±1.7	0±0.0	[7, 17]
HUNG372, NE5	ALPc	Kompolt-Kigyóser	47.2	20.8	5295–4950	M	J1c1	C1a2	4.25	7.48±1.6	0±0.0	[7, 17]
HUNG86, NE3	ALPc	Garadna-Elkerül út site 2	48.5	21.2	5281–5026	F	X2b-T226C	.	3.32	12.1±1.7	18±3.2	[7, 17]
MEMO24b	ALPc	Mez kövesd-Mocsolyás	47.8	20.6	5500–5300	M	U8b1b	CT	0.04	11.7±3.3	26±12	
MEMO2b	ALPc	Mez kövesd-Mocsolyás	47.8	20.6	5500–5300	F	K1a1	.	2.28	8.99±1.7	24±5.2	
MEMO7a	ALPc	Mez kövesd-Mocsolyás	47.8	20.6	5481–5361	F	HV	.	0.26	1.64±1.9	13±6.1	
PF325, NE1	ALPc	Polgár-Ferenci-hát	47.9	21.2	5306–5071	F	U5b2c	.	1.52	8.12±1.8	11±3.9	[7, 17]
PF839/1198, NE4	ALPc	Polgár-Ferenci-hát	47.9	21.2	5211–5011	F	J1c5	.	3.49	9.95±1.7	25±10	[7, 17]
POP15a	ALPc	Polgár-Piócás	47.9	21.1	5300–4900	M	K1a1	I2a2a	0.31	9.75±2.0	11±3.7	
PULE1.18a	ALPc	Pusztataskony-Ledence	47.5	20.5	5300–4900	F	T2c1d1	.	0.29	10.6±1.8	0±0.0	
PULE1.23a	ALPc	Pusztataskony-Ledence	47.5	20.5	5300–4900	F	H1e	.	0.17	9.52±2.2	11±3.4	
TISO13a	ALPc	Tiszadob-Ókenéz	48.0	21.2	5208–4942	M	J1c2	I2a2a	1.21	12.9±1.7	22±7.6	
TISO1b	ALPc	Tiszadob-Ókenéz	48.0	21.2	5300–4900	M	H7	I2a2a1b1	0.11	7.24±2.4	0±0.0	
TISO3a	ALPc	Tiszadob-Ókenéz	48.0	21.2	5300–4900	F	U5b2b1a	.	0.27	12.1±2.1	8.4±5.2	
SEKU10a	Vin a	Szederkény-Kukorica-dűlő	45.6	18.3	5320–5080	M	K2a	G2a2b2a1a	0.24	2.28±1.9	0±0.0	
SEKU6a	Vin a	Szederkény-Kukorica-dűlő	45.6	18.3	5321–5081	F	H26	.	1.15	9.16±1.7	9.0±9.4	
VEGI17a	Vin a	Versend-Gilencsa	45.6	18.3	5400–5000	F	U2	.	0.01	-6.14±5.6	0±0.0	
VEGI3a	Vin a	Versend-Gilencsa	45.6	18.3	5400–5000	M	T2b	H2	0.41	0.53±1.8	0±0.0	
Gorzsa18	Tisza	Hódmez vásárhely-Gorzsa	46.4	20.4	5000–4500	M	U5b2c	I2a1	6.87	7.77±1.6	13±4.3	
Gorzsa4	Tisza	Hódmez vásárhely-Gorzsa	46.4	20.4	5000–4500	F	T1a	.	0.06	11.3±3.0	22±11	
KOKE3a	Tisza	Hódmez vásárhely-Kökénydomb Vörös tanya	46.4	20.2	5000–4500	M	K1b1	I	0.06	13.7±3.2	0±0.0	
PULE1.24	Tisza	Pusztataskony-Ledence	47.5	20.5	5000–4500	F	K1a4	.	0.40	10.4±1.9	18±7.2	
VSM3a	Tisza	Vézt -Mágor	46.9	21.2	5000–4500	M	H26	G2a	0.09	6.92±2.6	0±0.0	
ALE14a	TDLN	Alsónyék-Elkerül site 2	46.2	18.7	5030–4848	M	U8b1b	G2a	0.05	-1.11±3.2	0±0.0	
ALE4a	TDLN	Alsónyék-Elkerül site 2	46.2	18.7	5016–4838	M	T2c1	F	0.03	10.6±3.6	0±0.0	
BAL3a	TDLN	Bátaszék-Lajvér	46.2	18.7	4800–4500	M	T2f	H1b1	0.91	6.89±1.7	22±9.0	
CSAT19a	TDLN	Csabdi-Téizöldes	47.5	18.6	4800–4500	M	H	H	0.52	5.82±1.8	34±9.6	
CSAT25a	TDLN	Csabdi-Téizöldes	47.5	18.6	4826–4602	M	T2b	I2	0.43	13.5±1.9	26±8.1	
FAGA1a	TDLN	Fajsz-Garadomb	46.4	18.9	5100–4750	M	HVOa	I	0.09	5.08±2.4	0±0.0	
FAGA2a	TDLN	Fajsz-Garadomb	46.4	18.9	5195–4842	F	H	.	0.49	11.9±1.8	14±4.1	
FEB3a	TDLN	Fels ős-Báróker	47.0	18.0	4800–4500	M	H44	J2a	0.16	6.31±2.1	0±0.0	
HUNG347, NE7	TDLN	Apc-Berekalja	47.2	19.8	4491–4357	M	N1a1a1a	I	4.85	10.6±1.6	19±3.1	[7, 17]
SZEH5a	TDLN	Szemely-Hegyes	46.0	18.3	4904–4709	M	K1b1a	G	0.01	10.8±6.5	0±0.0	
SZEH7b	TDLN	Szemely-Hegyes	46.0	18.3	4930–4715	F	K1a	.	0.52	3.44±1.7	0±0.0	
VEJ12a	TDLN	Veszprém Jutasi út	47.1	17.9	4800–4500	M	U8b1a2b	H	0.10	6.17±2.3	0±0.0	
VEJ2a	TDLN	Veszprém Jutasi út	47.1	17.9	4800–4500	M	T2b	C	0.34	5.63±1.8	0±0.0	
VEJ5a	TDLN	Veszprém Jutasi út	47.1	17.9	4936–4742	M	J1c2	G2a2a1	0.62	7.78±1.8	15±2.9	
GEN67	Tiszapolgár	Törökszentmiklós road 4 site 3	47.2	20.4	4444–4257	M	H1	I2a2a1b	2.28	13.0±1.7	50±15	
PULE1.10a	Tiszapolgár	Pusztataskony-Ledence	47.5	20.5	4500–4000	M	T2c1	I2a	0.28	9.03±2.0	0±0.0	
PULE1.13a	Tiszapolgár	Pusztataskony-Ledence	47.5	20.5	4500–4000	M	T2c1	G2a2b2a1a1c1a	0.38	10.3±1.9	0±0.0	
PULE1.9a	Tiszapolgár	Pusztataskony-Ledence	47.5	20.5	4500–4000	M	H26	G2a2b	0.11	11.6±2.4	0±0.0	
GEN100	Lasinja	Alsónyék, site 11	46.2	18.7	4300–3900	F	T2b	.	1.81	9.51±1.6	45±11	
GEN49	Lasinja	Nemesnádudvar-Pápföld	46.3	19.1	4228–3963	M	T2b23	CT	0.97	12.8±1.8	27±6.8	
KEFP2a	Lasinja	Keszthely-Fenekpuszta	46.7	17.2	4300–3900	F	J2b1a	.	0.74	9.12±1.7	21±5.4	
KON2a	Lasinja	Enese elkerül , Kóny, Proletár-dűlő, M85, site 2	47.6	17.4	4333–4072	F	K2a	.	2.13	10.3±1.7	21±6.4	
M6-116.12a	Lasinja	Lánycsók, Csata-alja	46.0	18.6	4232–4046	F	T2f8a	.	0.64	9.68±1.7	29±11	
VEJ9a	Lasinja	Veszprém Jutasi út	47.1	17.9	4339–4237	M	H40	CT	0.05	8.83±3.2	0±0.0	
GEN60	Protoboleráz	Abony, Turjányos-d 1	47.2	20.0	3909–3651	M	H	G2a2b2a	1.88	14.0±1.6	37±8.8	
GEN61	Protoboleráz	Abony, Turjányos-d 1	47.2	20.0	3800–3600	M	J1c	I2c	0.76	10.8±1.7	65±13	
GEN62	Protoboleráz	Abony, Turjányos-d 1	47.2	20.0	3762–3636	F	N1a1a1a3	.	4.81	8.00±1.6	37±9.6	

ID	Population	Site	Lat.	Long.	Date	Sex	Mt Hap	Y Hap	Cov.	HG%	ALD	Ref.
GEN63	Protoboleráz	Abony, Turjános-d 1	47.2	20.0	3658–3384	M	U5a1c1	I2c	1.92	11.9±1.7	34±8.1	
GEN12a	Baden	Budakalász-Luppa csárda	47.6	19.0	3340–2945	M	H26a	G2a2b2a1a1b1	1.98	13.8±1.6	34±7.2	
GEN13a	Baden	Budakalász-Luppa csárda	47.6	19.0	3332–2929	M	HV	G2a2b2a1a	2.65	11.3±1.6	27±6.6	
GEN15a	Baden	Budakalász-Luppa csárda	47.6	19.0	3367–3103	M	J2a1a1	G2a2b2a1a1c1a	1.66	10.8±1.7	22±9.3	
GEN16a	Baden	Alsónémedi	47.3	19.2	3346–2945	F	T2b	.	4.30	12.9±1.6	38±16	
GEN17a	Baden	Alsónémedi	47.3	19.2	3359–3098	M	U5b3f	G2a2a	0.82	10.7±1.7	21±6.4	
GEN21	Baden	Balatonlelle-Fels -Gamász	46.8	17.7	3600–2850	M	K1a	I2a1	0.67	12.3±1.7	0±0.0	
GEN22	Baden	Balatonlelle-Fels -Gamász	46.8	17.7	3332–2929	M	U5a1	I2a1a1	2.31	14.5±1.7	25±6.6	
GEN55	Baden	Vámosgyörk	47.7	19.9	3600–2850	F	T2c1d1	.	0.81	13.1±1.7	22±6.6	
HUNG353, CO1	Baden	Apc-Berekalja	47.2	19.8	3315–2923	F	H	.	4.56	15.1±1.7	0±0.0	[7, 17]
Vors1	Baden	Vörs	46.7	17.3	3300–2850	F	T2f	.	0.03	4.47±4.2	0±0.0	

Cov: average coverage per SNP. HG%: inferred percentage of hunter-gatherer ancestry (mean ± standard error). ALD inferred date of admixture (generations in the past; mean ± standard error; zero implies no date obtained). Ref: reference for published data; if blank, newly published sample in this study (asterisk denotes a published individual with new sequencing data added). Radiocarbon dates are in normal text, while dates estimated from archaeological context are in italics. Further information can be found in Supplementary Table 1.

### Extended Data Table 2

Information for Neolithic individuals from Germany and Spain.

ID	Population	Site	Lat.	Long.	Date	Sex	Mt Hap	Y Hap	Cov.	HG%	ALD	Ref.
HAL03a	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5295–5057	F	T2b	.	0.01	-5.13±6.8	0±0.0	
HAL07a	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5212–4992	F	N1a1a1	.	0.05	1.72±3.2	0±0.0	
HAL15a	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5199–4857	M	N1a1a1a3	G2	0.02	5.26±5.0	0±0.0	
HAL17b	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5500–4850	F	V1	.	0.02	9.21±4.2	0±0.0	
HAL18a	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5500–4850	F	K2a	.	0.02	0.27±4.6	0±0.0	
HAL19	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5500–4850	F	K1a2	.	0.86	7.10±1.7	16±7.6	[7]*
HAL2	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5211–4963	M	N1a1a1a2	G2a2a1	0.76	1.91±1.7	11±2.4	[5, 7]*
HAL20b	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5500–4850	M	K1a2	G2a2a	0.06	2.53±3.1	0±0.0	
HAL21a	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5500–4850	M	T2b	G2a2a	0.01	-4.41±5.8	0±0.0	
HAL22b	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5500–4850	F	T2b	.	0.02	-7.71±4.7	0±0.0	
HAL24	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5201–4850	M	X2d1	G2a2a1	0.42	6.39±1.8	0±0.0	[5, 7]*
HAL25	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5210–5002	M	K1a	G2a2a1	0.49	2.58±1.7	18±6.6	[5, 7]*
HAL27a	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5500–4850	M	N1a1a3	G2a2a	0.05	3.84±3.0	0±0.0	
HAL31a	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5295–5057	F	K1	.	0.12	4.54±2.3	11±3.1	
HAL32b	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5500–4850	F	H26	.	0.23	3.34±2.0	23±4.4	
HAL34	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5219–5021	F	N1a1a1	.	0.25	5.63±2.0	9.2±5.0	[5, 7]*
HAL35b	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5500–4850	F	J1c	.	0.10	3.93±2.4	0±0.0	
HAL38a	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5500–4850	F	V1	.	0.29	1.10±1.9	0±0.0	
HAL39b	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5210–5002	M	H1e	G2a2a1	0.08	3.96±2.6	0±0.0	
HAL4	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5202–4852	F	N1a1a1a	.	6.92	6.55±1.6	18±5.9	[5, 7]*
HAL40a	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5500–4850	F	T2b	.	0.17	2.50±2.1	0±0.0	
HAL5	LBK	Halberstadt-Sonntagsfeld	51.9	11.0	5211–4991	F	T2c1	.	2.23	2.98±1.6	15±5.4	[5, 7]*
KAR16A	LBK	Karsdorf	51.3	11.7	5500–4850	M	H46b	T1a	0.09	0.28±2.6	13±5.1	[7]
KAR6	LBK	Karsdorf	51.3	11.7	5217–5041	M	H1/H1au1b	CT	0.10	5.78±2.5	0±0.0	[5, 7]
LBK1976	LBK	Viesenhäuser Hof	48.8	9.2	5500–4850	F	T2e	.	0.44	3.46±1.7	18±4.4	[5, 7]
LBK1992	LBK	Viesenhäuser Hof	48.8	9.2	5500–4850	F	T2b	.	2.66	5.68±1.6	12±4.3	[5, 7]
LBK2155	LBK	Viesenhäuser Hof	48.8	9.2	5500–4850	F	T2b	.	3.63	4.84±1.5	13±4.4	[5, 7]
Stuttgart	LBK	Viesenhäuser Hof	48.8	9.2	5310–5076	F	T2c1d1	.	9.65	3.00±1.6	22±8.1	[4]*
UWS4	LBK	Unterwiederstedt	51.7	11.5	5223–5021	F	J1c17	.	18.6	5.70±1.6	13±14	[5, 7]
ESP30	GermanyMN	Esperstedt	51.4	11.7	3970–3710	M	H1e1a	I	0.09	22.0±2.7	0±0.0	[5, 7]
HAL13a	GermanyMN	Halberstadt-Sonntagsfeld	51.9	11.0	4600–4300	F	V1a	.	0.11	9.04±2.4	13±4.3	

ID	Population	Site	Lat.	Long.	Date	Sex	Mt Hap	Y Hap	Cov.	HG%	ALD	Ref.
QLB15D	GermanyMN	Quedlinburg	51.8	11.1	3654–3527	M	HV	R	0.16	20.9±2.2	36±8.7	[5, 7]
QLB18A	GermanyMN	Quedlinburg	51.8	11.1	3640–3376	F	T2e1	.	0.41	19.6±1.8	23±4.9	[5, 7]
SALZ3B	GermanyMN	Salzmünde-Schiebzig	51.5	11.8	3400–3025	M	U3a1	G2a2a1	0.09	14.9±2.7	0±0.0	[7]
SALZ57A	GermanyMN	Salzmünde-Schiebzig	51.5	11.8	3345–3097	F	H3	.	0.02	25.0±4.4	0±0.0	
SALZ77A	GermanyMN	Salzmünde-Schiebzig	51.5	11.8	3400–3025	M	H3	IJK (x J)	0.02	21.3±5.0	0±0.0	
Bla16	Blätterhöhle	Blätterhöhle Cave	51.4	7.6	3958–3344	M	U5b2a2	R1b1	0.80	39.5±1.9	15±5.8	
Bla28	Blätterhöhle	Blätterhöhle Cave	51.4	7.6	3337–3024	M	J1c1b1	R1	0.10	51.9±2.7	11±4.5	
Bla5	Blätterhöhle	Blätterhöhle Cave	51.4	7.6	3704–3117	F	H5	.	5.07	41.2±1.9	24±4.7	
Bla8	Blätterhöhle	Blätterhöhle Cave	51.4	7.6	4038–3532	M	U5b2b2	I2a1	4.58	72.6±2.0	12±2.9	
CB13	Iberia EN	Cova Bonica	41.4	1.9	5469–5327	F	K1a2a	.	0.98	9.97±1.7	17±3.5	[18]
E-06-Ind1	Iberia EN	EI Prado de Pancorbo	42.6	-3.1	4827–4692	F	K1a4a1	.	0.47	8.72±1.8	17±2.3	
E-14-Ind2	Iberia EN	EI Prado de Pancorbo	42.6	-3.1	5216–5031	F	H1	.	0.38	7.52±1.8	19±2.8	
Troc1	Iberia EN	Els Trocs	42.5	0.5	5311–5218	F	J1c3	.	0.69	7.15±1.7	12±9.1	[5, 7]
Troc3	Iberia EN	Els Trocs	42.5	0.5	5294–5066	M	T2c1d/T2c1d2	R1b1a	1.31	9.91±1.8	49±22	[5, 7]
Troc5	Iberia EN	Els Trocs	42.5	0.5	5310–5078	M	N1a1a1	I2a1b1	13.8	6.83±1.6	6.8±2.8	[5, 7]
Troc7	Iberia EN	Els Trocs	42.5	0.5	5303–5075	F	V	.	1.57	11.0±1.7	18±4.8	[5, 7]
Mina18	Iberia MN	La Mina	41.3	-2.3	3893–3661	F	U5b1	.	13.6	22.8±1.7	42±18	[5, 7]
Mina3	Iberia MN	La Mina	41.3	-2.3	3900–3600	M	K1a1b1	H2	0.38	19.5±1.9	80±20	[5, 7]
Mina4	Iberia MN	La Mina	41.3	-2.3	3900–3600	M	H1	I2a2a1b2	3.95	22.6±1.9	25±6.2	[5, 7]
Mina6	Iberia MN	La Mina	41.3	-2.3	3900–3600	F	K1b1a1	.	1.36	18.9±1.7	46±8.2	[5, 7]
1.-K11	Iberia CA	La Chabola de la Hechicera	42.6	-2.6	3263–2903	M	X2b	I2a2	0.18	27.8±2.1	68±28	
3.-K11	Iberia CA	La Chabola de la Hechicera	42.6	-2.6	3627–3363	F	J2a1a1	.	0.12	24.4±2.4	27±11	
5.-K18	Iberia CA	La Chabola de la Hechicera	42.6	-2.6	3090–2894	M	J1c1	I2a2	0.10	18.5±2.5	43±11	
ES.1/4	Iberia CA	EI Sotillo	42.6	-2.6	2571–2347	M	H3	I	0.07	25.4±2.8	0±0.0	
ES-6G-110	Iberia CA	EI Sotillo	42.6	-2.6	2916–2714	M	H3	I2a2a	0.05	25.4±3.2	0±0.0	
Inventario0/4	Iberia CA	EI Sotillo	42.6	-2.6	2481–2212	M	X2b	I2a2a	0.12	29.6±2.5	56±23	
LHUE11J.5	Iberia CA	Alto de la Huesera	42.6	-2.6	3092–2877	F	U5b1	.	1.19	26.7±1.9	40±9.7	
LHUE2010.10	Iberia CA	Alto de la Huesera	42.6	-2.6	3014–2891	F	J1c1	.	0.11	25.2±2.5	64±13	
LHUE2010.11	Iberia CA	Alto de la Huesera	42.6	-2.6	3092–2918	M	V	G2a2a	5.36	28.9±1.8	38±12	
LHUE2014.11J	Iberia CA	Alto de la Huesera	42.6	-2.6	3100–2850	F	U5b2b	.	0.06	26.3±3.0	0±0.0	
LY.II.A.10.15066	Iberia CA	Las Yurdinas II	42.6	-2.7	3350–2750	M	U5b2b3a	I2a2a	1.93	30.0±1.8	0±0.0	
LY.II.A.10.15067	Iberia CA	Las Yurdinas II	42.6	-2.7	3350–2750	F	J2a1a1	.	0.30	23.8±2.0	0±0.0	
LY.II.A.10.15068	Iberia CA	Las Yurdinas II	42.6	-2.7	3350–2750	F	K1a4a1	.	0.39	29.2±1.9	26±10	
LY.II.A.10.15069	Iberia CA	Las Yurdinas II	42.6	-2.7	3354–2943	F	J1c3	.	4.24	25.1±1.7	28±15	
MIR1	Iberia CA	EI Mirador Cave	42.3	-3.5	2900–2346	F	K1a	.	0.24	24.2±2.1	0±0.0	[7]
MIR13	Iberia CA	EI Mirador Cave	42.3	-3.5	2900–2346	F	H3c3	.	0.10	27.8±2.4	0±0.0	[7]
MIR14	Iberia CA	EI Mirador Cave	42.3	-3.5	2568–2346	M	H3	I2a2a	0.94	23.3±1.8	57±15	[7]
MIR17	Iberia CA	EI Mirador Cave	42.3	-3.5	2900–2346	F	J1c1	.	0.22	23.6±2.2	0±0.0	[7]
MIR18	Iberia CA	EI Mirador Cave	42.3	-3.5	2865–2575	F	H1t	.	1.58	20.0±1.6	0±0.0	[7]
MIR19	Iberia CA	EI Mirador Cave	42.3	-3.5	2900–2346	M	H3	I	0.06	21.8±3.1	0±0.0	[7]
MIR2	Iberia CA	EI Mirador Cave	42.3	-3.5	2857–2496	F	K1b1a1	.	0.98	22.6±1.7	56±8.9	[7]
MIR202-037-n105	Iberia CA	EI Mirador Cave	42.3	-3.5	2900–2346	M	K1a	I2a2a	5.73	19.9±1.7	0±0.0	
MIR21	Iberia CA	EI Mirador Cave	42.3	-3.5	2900–2346	M	H3	I	0.11	24.7±2.4	55±17	[7]
MIR22	Iberia CA	EI Mirador Cave	42.3	-3.5	2900–2346	F	K1a2a	.	2.79	22.6±1.7	62±10	[7]
MIR24	Iberia CA	EI Mirador Cave	42.3	-3.5	2900–2346	M	J2b1a3	G2a2b2b	0.06	20.0±3.0	0±0.0	[7]
MIR25	Iberia CA	EI Mirador Cave	42.3	-3.5	2900–2346	M	U3a1	I2a1a1	0.73	25.3±1.7	34±15	[7]
MIR5, MIR6	Iberia CA	EI Mirador Cave	42.3	-3.5	2900–2679	M	X2b	I2a2a2a	10.4	20.7±1.7	0±0.0	[7]

Cov: average coverage per SNP. HG%: inferred percentage of hunter-gatherer ancestry (mean ± standard error). ALD: inferred date of admixture (generations in the past; mean ± standard error; zero implies no date obtained). Ref: reference for published data; if blank, newly published sample in this study (asterisk denotes a published individual with new



sequencing data added). Radiocarbon dates are in normal text, while dates estimated from archaeological context are in italics. Further information can be found in Supplementary Table 1.

### Extended Data Table 3

Admixture graph results for Neolithic populations

Population	Main scaffold		Alternative scaffold	
	HG ancestry	WHG affinity	HG ancestry	WHG affinity
Körös EN	0.0 ± 1.2%		0.0 ± 1.2%	
Star evo EN	2.3 ± 1.0%	KO1/VIL *	2.3 ± 1.0%	VIL
ALPc MN	8.8 ± 0.6%	KO1 * + VIL	9.5 ± 0.6%	KO1 * + VIL
LBKT MN	0.8 ± 0.9%	VIL *	0.5 ± 0.9%	VIL
Tisza LN	8.4 ± 1.3%	KO1/VIL	9.8 ± 1.3%	KO1/VIL + EHG
TDLN	8.2 ± 0.7%	KO1/VIL *	8.4 ± 0.7%	KO1 *
Lasinja CA	10.7 ± 0.9%	KO1/VIL *	10.6 ± 0.9%	KO1/VIL *
Protoboleráz CA	12.7 ± 0.9%	KO1/VIL *	12.5 ± 0.9%	KO1/VIL
Baden CA	13.0 ± 0.7%	KO1/VIL *	13.4 ± 0.7%	KO1 *
LBK EN	4.2 ± 0.6%	KO1 + LOS	5.0 ± 0.6%	KO1 *
Germany MN	17.0 ± 1.1%	LOS *	18.3 ± 1.1%	LOS + KO1
Blätterhöhle MN	40.6 ± 1.5%	KO1/VIL * + LOS	42.6 ± 1.5%	KO1 * + LOS
Iberia EN	10.0 ± 0.8%	LB1 *	10.4 ± 0.8%	LB1 *
Iberia MN	23.3 ± 1.1%	LB1 * + LOS	24.8 ± 1.1%	LB1 * + LOS
Iberia CA	26.5 ± 0.7%	LB1 * + LOS/KO1/VIL *	27.5 ± 0.7%	LB1 * + VIL *

Hunter-gatherer ancestry in Neolithic populations as inferred from admixture graph analyses. Shown are the inferred ancestry proportions for the best-fitting FEF+WHG model, along with the WHG individual(s) inferred to be related to the hunter-gatherer sources, with \* denoting statistical significance (Methods). The two sets of results are for the primary scaffold model (Extended Data Fig. 2) and an alternative admixture graph scaffold including EHG (Supplementary Information section 6). Plus signs indicate two components, while slashes indicate single components with one of two or three possibilities.

### Extended Data Table 4

Average dates of admixture for Neolithic populations

Population	Individual-based	Group-based	Average sample date (BCE)
Körös EN			5631 ± 31
Star evo EN	4.5 ± 1.9	1.9 ± 0.9	5738 ± 35
ALPc MN	17.8 ± 2.0	16.4 ± 2.6	5180 ± 31
LBKT MN	30.3 ± 5.8	31.5 ± 10.9	5142 ± 93
Tisza LN	18.2 ± 6.6	12.6 ± 3.1	4750 ± 145
TDLN	20.9 ± 2.7	19.1 ± 3.8	4681 ± 32
Lasinja CA	29.3 ± 5.2	23.0 ± 4.1	4123 ± 59
Protoboleráz CA	44.3 ± 6.4	19.8 ± 5.4	3674 ± 35
Baden CA	27.6 ± 3.8	26.2 ± 6.9	3176 ± 49
LBK EN	14.9 ± 2.4	15.4 ± 3.6	5128 ± 38
Germany MN	26.2 ± 4.4	55.0 ± 41.2	3724 ± 46
Blätterhöhle MN	18.5 ± 4.6	23.1 ± 6.2	3414 ± 84
Iberia EN	19.4 ± 2.3	17.5 ± 5.9	5107 ± 20

Population	Individual-based	Group-based	Average sample date (BCE)
Iberia MN	49.9 ± 7.7	40.0 ± 6.9	3749 ± 74
Iberia CA	49.6 ± 5.2	56.5 ± 7.9	2808 ± 27

Dates of admixture (in generations in the past) as inferred from *ALDER* through two different methods. On the left are the average individual-level dates used in our main analyses, and on the right are direct estimates for population groups. By default, for group-level estimates, we used all individuals that yielded a date in our standard *ALDER* procedure, but because of missing data, for some populations we used a subset of individuals (typically those with highest coverage): Star evo (BAM17b, BAM4a, and LGCS1a; we note that in this case only BAM17b had an *ALDER* signal individually), ALPc (HAJE7a, HELI11a, MEMO2b, NE1, NE3, NE4, and TISO13a), Tisza (Gorzsai8 and PULE1.24), Baden (GEN12a, GEN13a, GEN15a, GEN17a, GEN22, and GEN55), LBK (HAL19, HAL2, HAL4, HAL5, LBK1992, and Stuttgart), and Iberia CA (LHUE11J.5, LHUE2010.11, LY.II.A.10.15066, LY.II.A.10.15069, MIR14, MIR2, and MIR22). For the group-level estimate for Iberia MN, we use a fitting start point of 0.8 cM instead of the program-inferred minimum of 0.6 because of a noticeably lower standard error. For our main analyses, we omit the outlier Protoboleráz individual GEN61, yielding an average date of 36.0 ± 5.2 generations, to help capture uncertainty due to the disagreement between the individual-level and group-level estimates shown here. Average sample dates (except for Körös) are based on the same weighting as the individual-level average dates of admixture for compatibility (Supplementary Information section 7).

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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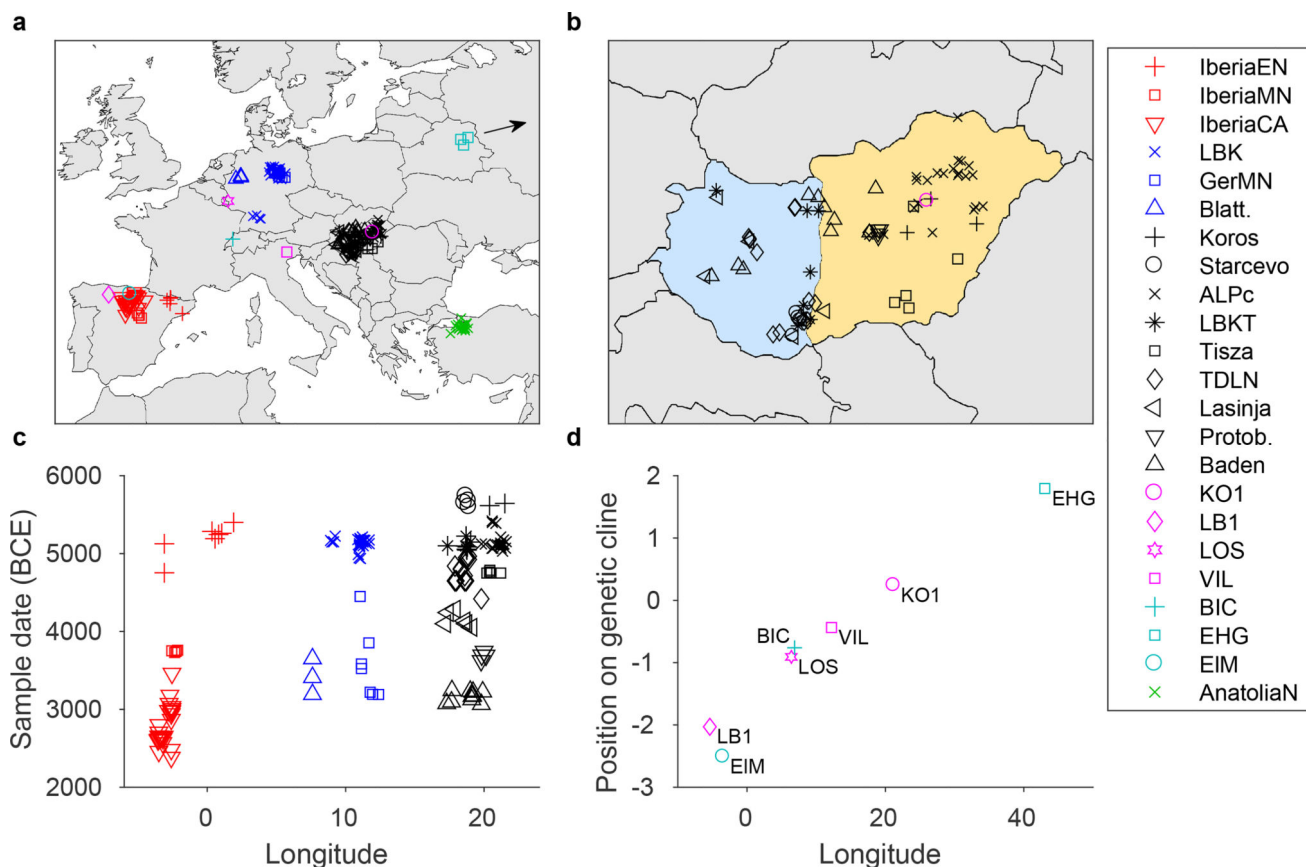
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## References

1. Bramanti B, et al. Genetic discontinuity between local hunter-gatherers and Central Europe's first farmers. *Science*. 2009; 326:137–140. [PubMed: 19729620]
2. Haak W, et al. Ancient DNA from European early Neolithic farmers reveals their Near Eastern affinities. *PLoS Biol*. 2010; 8:e1000536. [PubMed: 21085689]
3. Skoglund P, et al. Origins and genetic legacy of Neolithic farmers and hunter-gatherers in Europe. *Science*. 2012; 336:466–469. [PubMed: 22539720]
4. Lazaridis I, et al. Ancient human genomes suggest three ancestral populations for present-day Europeans. *Nature*. 2014; 513:409–413. [PubMed: 25230663]
5. Haak W, et al. Massive migration from the steppe was a source for Indo-European languages in Europe. *Nature*. 2015; 522:207–211. [PubMed: 25731166]
6. Günther T, et al. Ancient genomes link early farmers from Atapuerca in Spain to modern-day Basques. *Proc. Natl. Acad. Sci. U. S. A.* 2015; 112:11917–11922. [PubMed: 26351665]
7. Mathieson I, et al. Genome-wide patterns of selection in 230 ancient Eurasians. *Nature*. 2015; 528:499–503. [PubMed: 26595274]
8. Hofmanová Z, et al. Early farmers from across Europe directly descended from Neolithic Aegeans. *Proc. Natl. Acad. Sci. U. S. A.* 2016 201523951.
9. Brandt G, et al. Ancient DNA reveals key stages in the formation of Central European mitochondrial genetic diversity. *Science*. 2013; 342:257–261. [PubMed: 24115443]
10. Ammerman, AJ., Cavalli-Sforza, LL. *The Neolithic transition and the genetics of populations in Europe*. Princeton: 1984.
11. Price, TD. Lessons in the transition to agriculture. In: Price, TD., editor. *Europe's First Farmers*. Cambridge: 2000. p. 301-18.
12. Zvebil M. The agricultural transition and the origins of Neolithic society in Europe. *Documenta Praehistorica*. 2001; 28:1–26.
13. Richards M. The Neolithic invasion of Europe. *Ann. Rev. Anthropol.* 2003:135–162.

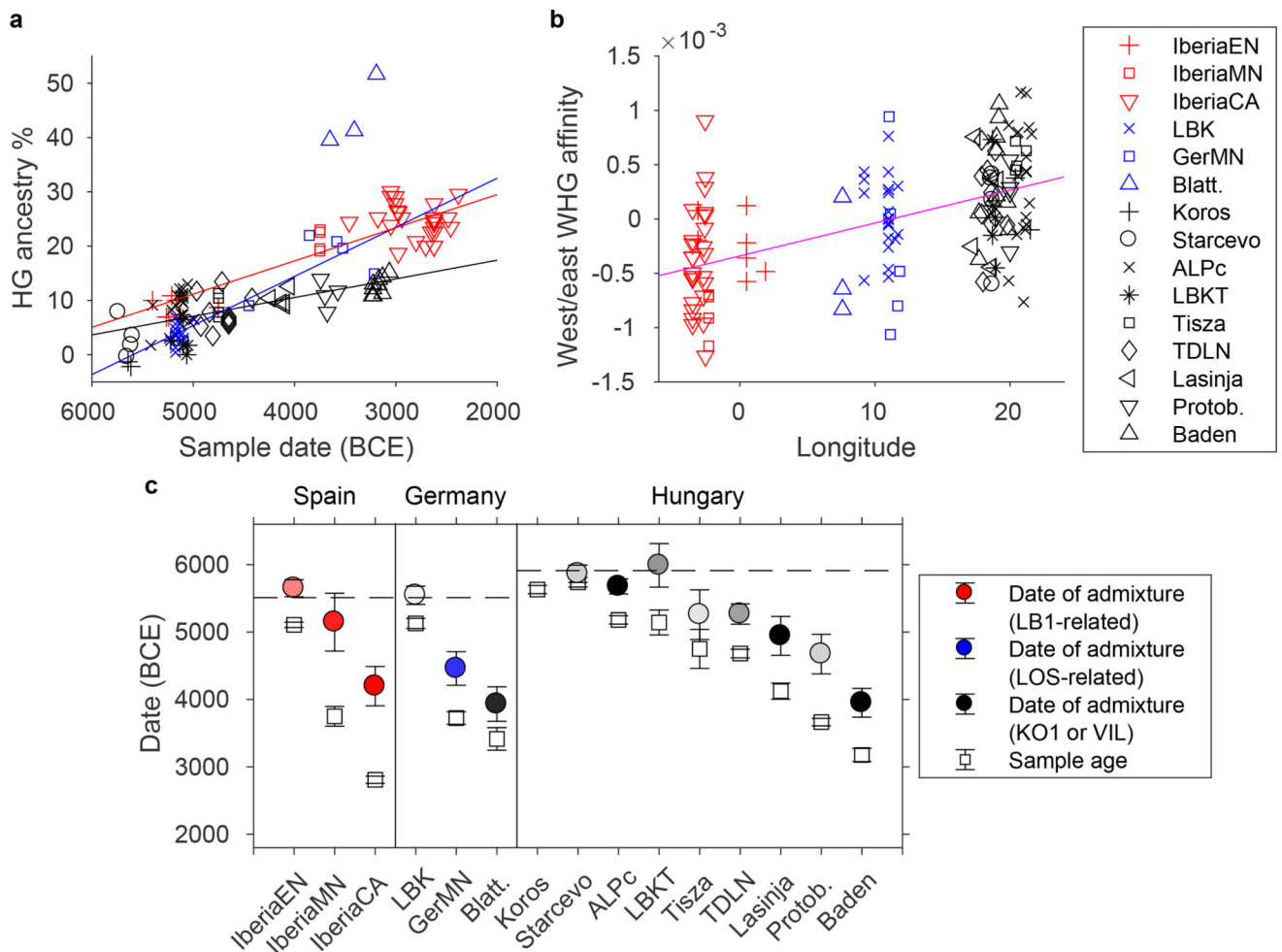
14. Tringham, R. Southeastern Europe in the transition to agriculture in Europe: bridge, buffer or mosaic. In: Price, TD., editor. *Europe's First Farmers*. Cambridge: 2000. p. 19-56.
15. Bollongino R, et al. 2000 years of parallel societies in Stone Age Central Europe. *Science*. 2013; 342:479–481. [PubMed: 24114781]
16. Skoglund P, et al. Genomic diversity and admixture differs for Stone-Age Scandinavian foragers and farmers. *Science*. 2014; 344:747–750. [PubMed: 24762536]
17. Gamba C, et al. Genome flux and stasis in a five millennium transect of European prehistory. *Nat. Comm*. 2014; 5:5257.
18. Olalde I, et al. A common genetic origin for early farmers from Mediterranean Cardial and Central European LBK cultures. *Mol. Biol. Evol*. 2015; 32:3132–3142. [PubMed: 26337550]
19. Olalde I, et al. Derived immune and ancestral pigmentation alleles in a 7,000-year-old Mesolithic European. *Nature*. 2014; 507:225–228. [PubMed: 24463515]
20. Fu Q, et al. The genetic history of Ice Age Europe. *Nature*. 2016; 534:200–205. [PubMed: 27135931]
21. Jones ER, et al. Upper Palaeolithic genomes reveal deep roots of modern Eurasians. *Nat. Comm*. 2015; 6:8912.
22. Seguin-Orlando A, et al. Genomic structure in Europeans dating back at least 36,200 years. *Science*. 2014; 346:1113–1118. [PubMed: 25378462]
23. Loh P-R, et al. Inferring admixture histories of human populations using linkage disequilibrium. *Genetics*. 2013; 193:1233–1254. [PubMed: 23410830]
24. Bánffy E. Eastern, Central and Western Hungary – variations of Neolithisation models. *Documenta Praehistorica*. 2006; 33:125–142.
25. Domboróczki L, Kaczanowska M, Kozowski J. The Neolithic settlement at Tiszasz 1 s-Domaháza-puszta and the question of the northern spread of the Körös Culture. *Atti Soc. Preist. Protost. Friuli-VG*. 2010; 17:101–155.
26. Szécsényi-Nagy A, et al. Tracing the genetic origin of Europe's first farmers reveals insights into their social organization. *Proc. Royal Soc. B*. 2015; 282:20150339.
27. Raczky, P. Historical context of the Late Copper Age Cemetery at Budakalász. In: Bondár, M., Raczky, P., editors. *The Copper Age Cemetery of Budakalász*. Pytheas, Budapest: 2009. p. 475-485.
28. Martins H, et al. Radiocarbon dating the beginning of the Neolithic in Iberia: New results, new problems. *J. Medit. Arch*. 2015; 28:105–131.
29. Jakucs J, et al. Between the Vinča and Linearbandkeramik worlds: The diversity of practices and identities in the 54th–53rd centuries cal BC in Southwest Hungary and beyond. *J. World Prehist*. 2016; 29:267–336. [PubMed: 27746586]
30. Oross K, et al. Midlife changes: The Sopot burial ground at Alsónyék. *Bericht der Römisch-Germanischen Kommission*. 2016; 94:151–178.
31. Dabney J, et al. Complete mitochondrial genome sequence of a Middle Pleistocene cave bear reconstructed from ultrashort DNA fragments. *Proc. Natl. Acad. Sci. U. S. A*. 2013; 110:15758–15763. [PubMed: 24019490]
32. Korlevi P, et al. Reducing microbial and human contamination in DNA extractions from ancient bones and teeth. *BioTechniques*. 2015; 59:87–93. [PubMed: 26260087]
33. Lazaridis I, et al. Genomic insights into the origin of farming in the ancient Near East. *Nature*. 2016; 536:419–424. [PubMed: 27459054]
34. Rohland N, Harney E, Mallick S, Nordenfelt S, Reich D. Partial uracil–DNA–glycosylase treatment for screening of ancient DNA. *Phil. Trans. R. Soc. B*. 2015; 370 20130624.
35. DeAngelis MM, Wang DG, Hawkins TL. Solid-phase reversible immobilization for the isolation of PCR products. *Nucl. Acids Res*. 1995; 23:4742–4743. [PubMed: 8524672]
36. Rohland N, Reich D. Cost-effective, high-throughput DNA sequencing libraries for multiplexed target capture. *Genome Res*. 2012; 22:939–946. [PubMed: 22267522]
37. Meyer M, et al. A mitochondrial genome sequence of a hominin from Sima de los Huesos. *Nature*. 2014; 505:403–406. [PubMed: 24305051]

38. Sawyer S, Krause J, Guschanski K, Savolainen V, Pääbo S. Temporal patterns of nucleotide misincorporations and DNA fragmentation in ancient DNA. *PloS one*. 2012; 7:e34131. [PubMed: 22479540]
39. Andrews RM, et al. Reanalysis and revision of the Cambridge reference sequence for human mitochondrial DNA. *Nat. Genet.* 1999; 23:147. [PubMed: 10508508]
40. Behar DM, et al. A “Copernican” reassessment of the human mitochondrial DNA tree from its root. *Am. J. Hum. Genet.* 2012; 90:675–684. [PubMed: 22482806]
41. Mallick S, et al. The Simons Genome Diversity Project: 300 genomes from 142 diverse populations. *Nature*. 2016; 538:201–206. [PubMed: 27654912]
42. Fu Q, et al. DNA analysis of an early modern human from Tianyuan Cave, China. *Proc. Natl. Acad. Sci. U. S. A.* 2013; 110:2223–2227. [PubMed: 23341637]
43. Korneliussen TS, Albrechtsen A, Nielsen R. ANGSD: analysis of next generation sequencing data. *BMC Bioinformatics*. 2014; 15:356. [PubMed: 25420514]
44. Domboróczki, L. Research at Tiszasz 1 s-Domaháza-pusztá in 2003. In: Anders, A., Siklósi, Z., editors. *The First Neolithic Sites in Central/South-East European Transect. Volume III: The Körös Culture in Eastern Hungary*. Oxford: 2012. p. 107-111.
45. Oross K, et al. The early days of Neolithic Alsónyék: the Star evo occupation. *Bericht der Römisch-Germanischen Kommission*. 2016; 94:93–121.
46. Ramsey CB, Lee S. Recent and planned developments of the program OxCal. *Radiocarbon*. 2013; 55:720–730.
47. Reimer PJ, et al. Intcal13 and marine13 radiocarbon age calibration curves 0–50,000 years cal bp. *Radiocarbon*. 2013; 55:1869–1887.
48. Patterson N, Price A, Reich D. Population structure and eigenanalysis. *PLoS Genet*. 2006; 2:e190. [PubMed: 17194218]
49. Patterson N, et al. Ancient admixture in human history. *Genetics*. 2012; 192:1065–1093. [PubMed: 22960212]
50. Fenner J. Cross-cultural estimation of the human generation interval for use in genetics-based population divergence studies. *Am. J. Phys. Anthropol.* 2005; 128:415–423. [PubMed: 15795887]
51. Moorjani P, et al. A genetic method for dating ancient genomes provides a direct estimate of human generation interval in the last 45,000 years. *Proc. Natl. Acad. Sci. U. S. A.* 2016 201514696.



**Figure 1. Spatial and temporal contexts of European Neolithic samples**

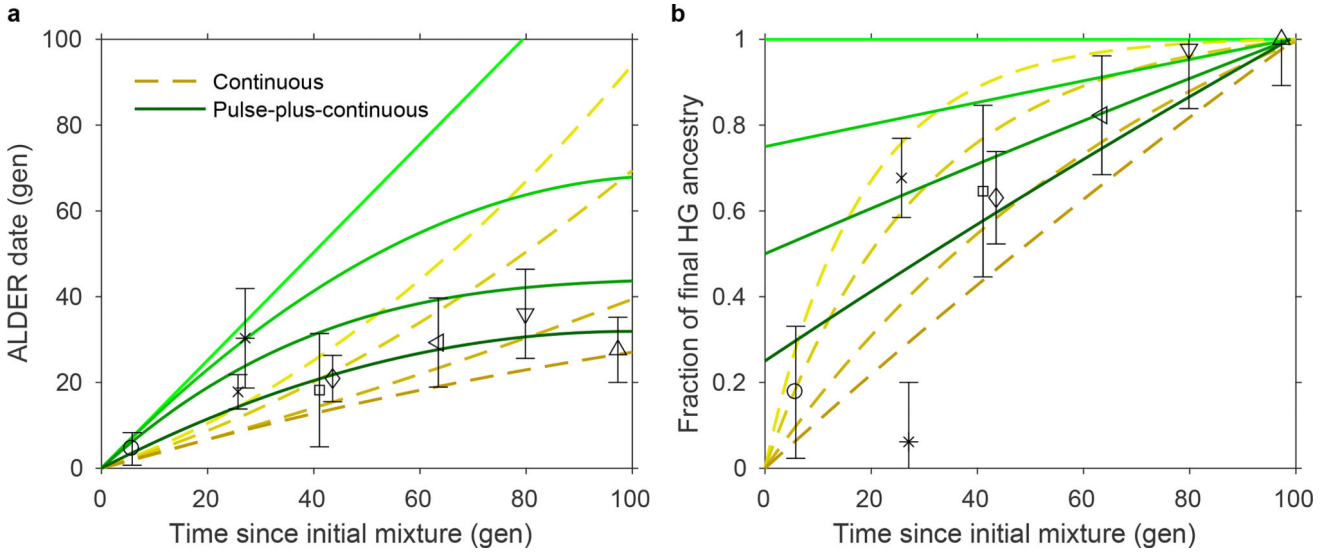
**a, b**, Locations of samples used for analyses, with close-up of Hungary (orange shading for Alföld and light blue for Transdanubia). **c**, Sample dates arranged by longitude. **d**, Hunter-gatherer genetic cline (derived from MDS analysis; Supplementary Information section 5) as a function of longitude. The four primary WHG individuals are shown together with “BIC” (Bichon, ~11,700 BCE from Switzerland<sup>21</sup>), “EHG” (eastern hunter-gatherers, ~7000–5000 BCE from Russia<sup>5,7</sup>), and “EIM” (El Mirón, ~17,000 BCE from Spain<sup>20</sup>). Random jitter is added to separate overlapping positions in **a–c**. GerMN, Germany MN; Blatt., Blätterhöhle; Protob., Protoboleráz.



**Figure 2. Admixture parameters for test individuals and populations**

**a**, Estimated individual hunter-gatherer ancestry versus sample date, with best-fitting regression lines for each region (excluding Blätterhöhle). Standard errors are around 2% for hunter-gatherer ancestry and 100 years for dates (Methods; Extended Data Tables 1, 2). **b**, Relative affinity of hunter-gatherer ancestry in Neolithic individuals, measured as  $f_4(\text{LB1+LOS}, \text{KO1+VIL}; \text{Anatolia}, X)$  (positive, more similar to eastern WHG; negative, more similar to western WHG; standard errors  $\sim 5 \times 10^{-4}$ ), with best-fitting regression line ( $|Z| > 3$  for aggregate differences among the three regions). **c**, Population-level average sample ages and dates of admixture, plus or minus two standard errors. Colored fill indicates the inferred primary hunter-gatherer ancestry component, with darker shades corresponding to higher confidence (all admixed populations except LBK and Tisza significant at  $p < 0.05$ ; see Extended Data Table 3 and Supplementary Information section 6). Dashed lines denote the approximate date of arrival of farming in each region.





**Figure 3. Hungary time series and simulated data**

**a**, Dates of admixture. **b**, Hunter-gatherer ancestry proportions, normalized by the total in the most recent (rightmost) population. Symbols are as in Figs 1 and 2, here showing population-level averages plus or minus two standard errors. Yellow dashed lines represent continuous admixture simulations: from top to bottom, diminishing 5% per generation, diminishing 3%, diminishing 1%, and uniform. Green solid lines represent pulse-plus-continuous admixture simulations: from top to bottom, all hunter-gatherer ancestry in a pulse at time zero; 3/4 of final hunter-gatherer ancestry in an initial pulse, followed by uniform continuous gene flow; half in initial pulse and half continuous; and 1/4 in initial pulse.

**Table 1**

Neolithic population groups and western hunter-gatherer individuals in the study

Population	Country	Samples <sup>*</sup>	Appx. dates (BCE)
Körös EN	Hungary <sup>E</sup>	6/5/3 <sup>†</sup>	6000–5500
Star evo EN	Hungary <sup>W</sup>	5/4/4	6000–5500
ALPc MN	Hungary <sup>E</sup>	25/20/22	5500–5000
LBKT MN	Hungary <sup>W</sup>	8/7/7	5500–5000
Vin a MN	Hungary <sup>W</sup>	6/6/0	5500–5000
Tisza LN	Hungary <sup>E</sup>	6/6/5	5000–4500
TDLN	Hungary <sup>W</sup>	15/14/14	5000–4500
Tiszapolgár CA	Hungary <sup>E</sup>	5/5/0	4500–4000
Lasinja CA	Hungary <sup>W</sup>	7/7/6	4300–3900
Protoboleráz CA	Hungary <sup>E</sup>	4/4/4	3800–3600
Baden CA	Hungary	13/12/10	3600–2850
LBK EN	Germany	30/15/29	5500–4850
Germany MN	Germany	8/4/7	4600–3000
Blätterhöhle MN	Germany	4/4/4 <sup>†</sup>	4100–3000
Iberia EN	Spain	7/2/7	5500–4500
Iberia MN	Spain	4/0/4	3900–3600
Iberia CA	Spain	27/15/27	3000–2200
KO1 HG	Hungary <sup>E</sup>	1/0/1	5700
LB1 HG	Spain	1/0/1	5900
LOS HG	Luxembourg	1/0/1	6100
VIL HG	Italy <sup>E</sup>	1/0/1	12,000

\* Total number/new in this study/used in final analyses

<sup>†</sup> Includes one hunter-gatherer individual treated separately

<sup>E</sup> Eastern

<sup>W</sup> Western

EN/MN/LN, Early/Middle/Late Neolithic; CA, Chalcolithic; HG, hunter-gatherer (LB1, La Braña 1; LOS, Loschbour; VIL, Villabruna)