



Phyto-Therapeutic and Nanomedicinal Approaches to Cure Alzheimer's Disease: Present Status and Future Opportunities

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Ovais M, Zia N, Ahmad I, Khalil AT, Raza A, Ayaz M, Sadiq A, Ullah F and Shinwari ZK (2018) Phyto-Therapeutic and Nanomedicinal Approaches to Cure Alzheimer's Disease: Present Status and Future Opportunities. Front. Aging Neurosci. 10:284. doi: 10.3389/fnagi.2018.00284 Alzheimer's disease (AD) is characterized by cognitive inability manifested due to the accumulation of β -amyloid, formation of hyper phosphorylated neurofibrillary tangles, and a malfunctioned cholinergic system. The degeneration integrity of the neuronal network can appear long after the onset of the disease. Nanotechnology-based interventions have opened an exciting area via theranostics of AD in terms of tailored nanomedicine, which are able to target and deliver drugs across the blood-brain barrier (BBB). The exciting interface existing between medicinal plants and nanotechnology is an emerging marvel in medicine, which has delivered promising results in the treatment of AD. In order to assess the potential applications of the medicinal plants, their derived components, and various nanomedicinal approaches, a review of literature was deemed as necessary. In the present review, numerous phytochemicals and various feats in nanomedicine for the treatment of AD have been discussed mechanistically for the first time. Furthermore, recent trends in nanotechnology such as green synthesis of metal nanoparticles with reference to the treatment of AD have been elaborated. Foreseeing the recent progress, we hope that the interface of medicinal plants and nanotechnology will lead to highly effective theranostic strategies for the treatment of AD in the near future.

Keywords: Alzheimer's disease, nanotechnology, medicinal plants, green synthesis, theranostic NPs

INTRODUCTION

Alzheimer's disease (AD) is a chronic neurodegenerative disorder characterized by memory loss, impaired cognitive function, and behavioral instability, predominantly among the elderly population (Minati et al., 2009). In the brain of AD patients, the excessive production of proteins [β -amyloid (A β) and hyperphosphorylated tau] leads to the loss of synaptic connections and neuronal impairment in the hippocampus and cerebral cortex. The aforementioned events

thus, lead to the degenerative diseases like AD and dementia (Selkoe, 2001). The complex pathophysiological aspects of AD brain have been extensively studied; however, an effective medicine is still unavailable for the complete treatment of AD (Ayaz et al., 2016). In clinical trials, several claimed potential anti-Alzheimer drugs failed to prove their effectiveness (Cummings et al., 2016). In order to combat the situation, alternative and cutting edge approaches like nanomedicine have been proposed as a potential treatment modality. In this review paper, emerging phyto-therapeutic and nanomedicinal approaches for treating AD are discussed. In addition, recent trends in emerging theranostics for AD using NPs delivery across the blood–brain barrier (BBB) and their mechanism of action are elaborated.

Prevalence and Etiology of AD

The AD is one of the major causes of dementia in an estimated 60–80% of cases. According to the Alzheimer's Association Report 2017, about 5.5 million individuals have AD (Alzheimer's and Association, 2017). Early clinical symptoms of AD patients include impairment in memory, e.g., current conversations, names or events, while later symptoms comprises poor communication, misperception, confusion, misjudgment, changed attitude, and finally, difficulty in speaking, swallowing, and walking (Khachaturian, 1985). In the United States, AD was reported in 4.7 million individuals aged 65 years or older during 2010, which is estimated to reach 13.8 million cases by 2050. Similarly, a considerable number of deaths were observed among older AD patients. In 2010, \sim 600,000 deaths occurred among people with AD at the age of 65 years or older, which is expected to reach 1.6 million by 2050 (Hebert et al., 2013).

The A β and tau proteins are the key factors to be considered in AD treatment. Currently, researchers are trying to develop drugs that can minimize amyloid synthesis/accumulation and tau aggregation/phosphorylation. The AD is associated with multiple factors of unknown etiology, and various cases can be observed with mutations in different genes, such as in presenilins (PS1 and PS2) and A\beta precursor protein (APP) (Bird, 2008). The apolipoprotein E (APOE) genes are also considered to be a risk factor for AD. Similarly, different proteins like APOE, APP, β amyloid cleaving enzyme (BACE), presenilin 1 and 2 (PS1 and PS2), and secretases (BACE1) are the key factors in the pathogenesis of AD (Kim et al., 2009). Therefore, researchers have focused on the synthesis of novel inhibitors for PS1, BACE, and BACE1 for the possible treatment of AD. Investigators are also trying to determine the efficacy of cholinesterase (ChE) inhibitors in the brain (Ayaz et al., 2017b; Ovais et al., 2018b). The appliance of new-generation acetyl and butyrylcholinesterase (AChE/BChE) inhibitors has been tested in the clinical trials for AD (Giacobini, 2004). Additional approaches such as hormone therapy and use of antioxidants, cholesterol-lowering mediators, anti-inflammatory substances, and vaccinations have also been studied for the treatment of AD (Yiannopoulou and Papageorgiou, 2013).

Role of the BBB in AD

The BBB is a delicate membrane synthesized by the endothelial cells, which makes up the inside layer of cerebral microvessels.

The BBB controls the entrance of plasma components, red blood cells, and leukocytes into the central nervous system (CNS), while exporting the neurotoxic molecules from the brain to the blood. Two additional sites were found in the CNS, which make up a barrier between the blood and cerebrospinal fluid (CSF), e.g., the arachnoid epithelium constituting the central layer of the meninges and choroid plexus epithelium. These distinctive BBB membranes encompass a group of physical, transport, and metabolic barriers that separate the neural environment from the blood (Abbott et al., 2010). The integrity of the BBB is controlled by the brain microvessel endothelial cells (BMECs) having distinctive luminal (apical) and abluminal (basolateral) membrane compartments (Zlokovic, 2011). They are an essential part of the neurovascular unit (NVU) in conjunction with pericytes, vascular smooth muscle cells from the vessel wall, neurons, and glial cells. Besides maintaining the chemical composition of the brain interstitial fluid, the NVU regulates BBB permeability and cerebral blood flow, which is necessary to maintain useful neuronal circuits (Daneman and Prat, 2015). Impairment in the BBB and NVU has severe pathological characteristics. These include outflow of circulating materials from the plasma into the CNS, changes in transporters that lead to poor nutrient supply and addition of neurotoxins in the CNS, and changes in the expression or secretion of proteins by NVU cells responsible for inflammation, oxidative stress, and neuronal damage. Recently, it has also been reported that the abnormal NVU cells result in different CNS pathologies such as dementia, neurodegeneration, and cognitive failure in AD (Erickson and Banks, 2013). Previously, the CNS was thought of as an immune-privileged organ, missing a lymphatic system and protected from the peripheral blood cells by the BBB. Conversely, the phenomenon is clear now that the BBB has the ability to respond to soluble factors and plasma proteins and interconnect with the peripheral immune system cells, thus forming neuroimmune system interactions, validating the concept that neuroinflammation leads to AD pathology (Heneka et al., 2015). Hence, the brain cannot be considered as an immune-privileged organ, and abnormal NVUs have to be withheld for peripheral immune cells and circulating soluble molecules that mediate immune responses (Zenaro et al., 2015).

Alternative Therapy Using Medicinal Plants

Due to severe side effects of current therapies, alternative therapies such as use of medicinal plants for the treatment of AD and other diseases are a vital focus of researchers (Ali M. et al., 2017; Ayaz et al., 2017a; Sadiq et al., 2018). The medicinal plants have been reported to enhance the memory and learning process that normally decline with AD (Ahmad et al., 2016; Ayaz et al., 2017b; Zohra et al., 2018). From the current literature, we have summarized potential anti-Alzheimer compounds isolated from various medicinal plants (**Table 1**). Currently, several studies report on the phytochemicals that have been clinically proven with significant anti-AD potentials. Some of these bioactive compounds and their mechanisms of action against AD have been discussed in the following sections.

TABLE 1 | The major medicinal plants with potential bioactive compounds and their mechanisms of action against AD.

Plant Part used Mechanism		Mechanism of action	Reference	
Withania somnifera	nania Withanolides - Effectively inhibits β-site amyloid precursor protein cleaving enzyme (BACE1) nnifera and and acetylcholinesterase (AChE). withanamides withanamides		Mahrous et al., 2017	
		- Bind to the active motif of β -amyloid (25–35), protect the rat neuronal cells (PC-12) from β -amyloid induced cell damage and prevent the fibril formation.	Jayaprakasam et al., 2010	
Curcuma longa	Curcumin	- Effectively inhibits $A\beta$ oligomer and fibril formation in Tg2576 mice brain.	Yang et al., 2005	
		- Reducing the astrocytic marker GFAP, Aβ, and plaque burden in Alzheimer transgenic APPSw mouse model (Tg2576).	Lim et al., 2001	
Convolvulus pluricaulis	Convolvine and convolamine	- Reduce AD biomarkers (A β PP, tau, and A β) in Naive male Wistar rats.	Bihaqi et al., 2012	
		- CNS depressant, neurodegenerative, and increases learning abilities.	Agarwa et al., 2014	
		 Increased AChE activity in the hippocampal CA1 and CA3 regions associated with learning and memory. 	Dubey et al., 1994	
Centella	Asiatic acid	- Used for rejuvenating the neuronal cells, increasing intelligence, longevity, and	Shinomol and Bharath, 2011	
asiatica	asiaticoside	memory.	Dhanasekaran et al., 2009	
		- Inhibits β -amyloid level in the brains of PSAPP mice.		
Celastrus paniculatus	Celapanin and celapanigin	 Increases cholinergic activity and improves memory performance by increasing ACh level in rat brain. 	Bhanumathy et al., 2010	
		 Prevents glutamine-induced neurotoxicity in embryonic rat forebrain neuronal cells. 	Godkar et al., 2006	
Nardostachys jatamansi	Valeranone nardosinone	- Improves learning and memory in mice.	Karkada et al., 2012	
Coriandrum sativum	Coumarin	- Suppresses A β 42, glial cell proliferation, and extracellular signal-regulated kinase activation.	Liu et al., 2016	
		- Improves $A\beta(1-42)$ -induced spatial memory impairment by attenuation of the oxidative stress in the rat hippocampus.	Liu et al., 2016	
		- Improves memory deficits in rats induced by scopolamine and diazepam.	Mani and Parle, 2009	
Ficus carica	Quercetin	- Induces neuronal bioactivity and is against oxidative stress-related AD.	Khojah and Edrada-Ebel, 2017	
		- Improvement of cognitive and behavioral deficits.	Subash et al., 2016	
Ginkgo biloba	Bilobalide ginkgolide	- Increases cell proliferation and neuroblast differentiation in mice.	Yoo et al., 2011	
		- Effective in the treatment and prevention of AD.	Shi et al., 2010	
Coffea arabica	Caffeine and chlorogenic acids	- Possesses neuroprotective effect in alleviating AD.	Shi et al., 2010	
		 Modulates nicotinamide mononucleotide adenylyl transferase 2 levels in cortical neurons. 	Ali Y.O. et al., 2017	
Glycyrrhiza glabra	Glycyrrhizin	 Relives neuroinflammation and memory deficit prompted by systemic lipopolysaccharide treatment in mice. 	Song et al., 2013	
Lepidium meyenii	Choline	 Enhances learning and memory, while reduces lipid peroxidation and acetylcholinesterase in ovariectomized (OVX) mice. 	Rubio et al., 2011	
Huperzia serrata	Huperzine	 Promotes hippocampal neurogenesis in vitro and in vivo and activates MAPK/ERK signaling pathway in mice. 	Ma et al., 2013	
Panax ginseng	Ginsenoside	- Cholinergic and activates Rb1, Rg1-3, Re, and Rh2 effective in the treatment of AD.	Nie et al., 2017	
		 Ginsenoside Rg1 improves behavioral abnormalities and modifies the hippocampal proteomic profile in transgenic AD mice. 		
Magnolia officinalis	Magnolol and honokiol	 Recovering memory impairment induced by scopolamine, and its effect may be related to the ability of AChE inhibition. 	Lee et al., 2009	
		 Significantly decreases Aβ-induced cell death, neuroprotective, reduces ROS production, suppresses intracellular calcium, and inhibits caspase-3 activity. 	Hoi et al., 2010	
Zingiber officinale	Shogaol and gingerol	- Significantly improves learning and memory in rates.	Gharibi et al., 2013	
		- Inhibits GSK-3 β and helps to recover the SHSY-5Y cells from A β -42 toxicity.	Yerer et al., 2017	
Crocus sativus	Crocin	- Reduces cognitive decline in AD patients.	Farokhnia et al., 2014	
		- Inhibits $A\beta$ and has a positive effect on cognitive function in AD.	Akhondzadeh et al., 2010	

(Continued)

TABLE 1 | Continued

Plant	Part used	Mechanism of action	Reference
Cissampelos pareira	Benzylisoquinoline	- Inhibits acetylcholinesterase activity and enhances memory in mice.	Kulkarni et al., 2011
Myristica fragrans	Monoterpenoid	- Memory enhancement and inhibition of acetylcholinesterase activity.	Asgarpanah and Kazemivash, 2012
Bacopa monnieri	Bacoside and reserpine	- Inhibits acetylcholinesterase and β -amyloid level while activates choline acetyltransferase.	Aguiar and Borowski, 2013
		- Improved cognition, protects memory loss, and neurodegeneration	Shinomol and Bharath, 2011
Camellia Sinensis	Catechins	 Prevents cognitive impairment and protects brain by modulating the risk factors for AD. 	Ho et al., 2013
		- Regulates BACE1 mRNA expression in SY5Y neuroblastoma cells.	Geiser et al., 2017
Allium sativum	S-allylcysteine	- Improves short-term recognition memory and relieves the neuroinflammation in A β -induced rats.	Nillert et al., 2017

Withanolides and Withanamides

Withania somnifera (WS), an essential medicinal plant, has been in use for the treatment of AD for the past 3000 years. The crude extract of this plant possesses withanamides A and C that can effectively bind to AB and inhibit fibril synthesis in the rat neuronal cells (PC-12) (Jayaprakasam et al., 2010; Ali et al., 2018). In a current study, withanolide S, isolated from different parts of WS, showed dual inhibitory activities against BACE1 and AChE (Mahrous et al., 2017; Puerta et al., 2017). In another study, computational tools were used to determine the bioactive compounds of WS against AD. The ligands (anaferine, anahygrine, cuscohygrine, and isaopelletierine) of WS were found to be neurologically active as agonists at the neuronal nicotinic acetylcholine receptors (nAChR) (Remya et al., 2016). Vareed et al. (2014) investigated the BBB permeability of withanamides that are present in W.somnifera fruit extract using animal models via high-performance liquid chromatography and quadrupole time of flight mass spectrometer system. Results of the study concluded that four withanamides significantly crossed BBB and their major peaks were identical both in brain tissue homogenates and plant crude extracts (Vareed et al., 2014).

Curcumin

Curcumin is the active ingredient isolated from *Curcuma longa*, carrying significant effects against AD. It has an antiamyloidogenic potential, which inhibits A β aggregation and downregulates BACE1 expression. It also possesses antitau function by inhibiting tau hyperphosphorylation and plays a key role in tau tangle clearance. In a randomized, placebocontrolled double-blinded study, Larry Baum et al. reported that curcumin exhibits neuroprotective effects in AD patients via several mechanisms, including the inhibition of A β aggregation, the inflammatory pathways, and free radicalinduced neurodegeneration (Baum et al., 2008).

In another study, curcumin significantly reduced proinflammatory cytokines levels like interleukin (IL) 1 β and oxidized proteins. Moreover, curcumin therapy considerably declined soluble and insoluble A β load in brain tissues and reduced astrocytic marker glial fibrillary acidic protein (GFAP) (Lim et al., 2001). The free radicals-scavenging, anti-amyloid, and anti-inflammatory effects of curcumin were also analyzed in transgenic and lipopolysaccharide (LPS) induced AD model. In both animal models, curcumin therapy reduced A β load and the chronic inflammation via inhibition of inflammatory cytokines (Begum et al., 2008). Furthermore, it can be used as a neuroprotective agent that promotes neuroplasticity and inhibits AChE activity (Serafini et al., 2017). Several other studies signify the potential use of curcumin in the management of neurodegenerative disorders like AD and its possible mechanism of action (Baum and Ng, 2004; Aggarwal and Sung, 2009).

In a study, Tsai et al. (2011) investigated the BBB penetration potentials of curcumin-loaded poly (lactic-co-glycolic acid) (PLGA) nanoparticles (NPs). Curcumin-brain concentrations and retention times were considerably increased in the cerebral cortex and hippocampus tissues of the animal brain (Tsai et al., 2011). Likewise, curcumin therapy is also reported to inhibit cerebral ischemia and reperfusion injury via prevention of BBB damage (Jiang et al., 2007). Yet another study on nano-curcumin preparations revealed significant improvements in cognitive function and reduction in amyloid load in transgenic animal model of AD (Cheng et al., 2013).

Asiatic Acid and Asiaticoside

Centella asiatica is an important medicinal plant containing asiatic acid (AA) and asiaticoside. These neuroprotective phytochemicals have the potential to cross the BBB. Asiatic acid increases the viability of differentiated human neuroblastoma SH-SY5Y cells (Ternchoocheep et al., 2017). The neuroprotective effects of asiaticoside have been reported in primary cultured mouse cortical neurons exposed to glutamate-induced excitotoxicity invoked by *N*-methyl-D-aspartate (NMDA). It downregulates NMDA receptor subtype 2B (NMDAR2B or NR2B), which shows that asiaticoside can protect neurons from excitotoxicity induced by NMDA exposure via obstructing cell apoptosis and calcium overload (Qi et al., 2014). Asiatic acid isolated from *C. asiatica* is reported to mediate its neuroprotective effects via reduction of the BBB permeability and mitochondrial injury (Krishnamurthy et al., 2009).

Celapanin and Celapanigin

Phytochemical studies of *Celastrus paniculatus* (CP) show the presence of alkaloids like celapanin and celapanigin, which are extensively used as neuroprotective agents, memory enhancers, and in different CNS disorders. A recent study has proposed the protective effects of CP against 3-nitropropionic acid (3-NP)-induced neurotoxicity. It also prevents glutamineinduced neurotoxicity in embryonic rat forebrain neuronal cells. Glutamate receptor and/or NMDA receptor antagonists have shown neuroprotective effects against such damage (Godkar et al., 2006; Malik et al., 2017).

Valeranone and Nardosinone

Nardostachys jatamansi, a flowering herb that contains valeranone and nardosinone enhances the levels of key neurotransmitters such as gamma-aminobutyric acid (GABA), norepinephrine, dopamine, serotonin, and 5hydroxyindoleacetic acid in the brain. Valeranone and nardosinone also inhibit inflammatory cytokines IL-1 β , IL-6, and tumor necrosis factor alpha (TNF- α). Furthermore, they deactivate p38 mitogen-activated protein kinases (MAPKs) (Akram and Nawaz, 2017) https://www.ncbi.nlm.nih.gov/pubmed/26371857. It is also reported that nardosinone acts as an enhancer of nerve growth factor (NGF) (Zhang C. et al., 2014), dibutyryl cyclic AMP, and staurosporine-induced neurite outgrowth from PC12D cells by amplifying both the MAP kinase-dependent and independent signaling pathways of dibutyryl-cAMP (dbcAMP) and staurosporine (Li et al., 2003). Nordosinone has been recently reported to significantly cross BBB and exhibit strong antidepressant activity (Jalali et al., 2018).

Coumarin

Coriandrum sativum, known as coriander, is an important medicinal plant, which contains coumarin as an active ingredient that has memory-enhancing and neuroprotective effects (Mani and Parle, 2009). *C. sativum* extract intake by AD model flies increases the reactive oxygen species (ROS) levels and number of glial cells. It also inhibits the epidermal growth factor receptor and induced phosphorylation of extracellular signalregulated kinase (ERK) (Laribi et al., 2015). Currently, a series of 7-substituted coumarin derivatives have been designed and synthesized to display ChE and monoamine oxidase (MAO-B) inhibitory activities. The molecular modeling determined that the tested compounds are able to inhibit AChE-induced $A\beta$ aggregation and, therefore, can be considered as promising multifunctional lead compounds for the treatment of AD patients (Joubert et al., 2017).

Among the natural coumarins, osthole is reported to exhibit neuroprotective effects via inhibition of oxidative stress-induced cerebral ischemia and inhibition of BBB disruption (Mao et al., 2011; Chen Z. et al., 2015). Several coumarin derivatives are reported to selectively inhibit MAO-B enzyme implicated in AD and prevent $A\beta_{1-42}$ aggregation. Some of the test compounds cross BBB, chelate metal ions, and exhibit low toxicity in cell lines, which signify their use as potential anti-AD agents (Huang et al., 2015; Jameel et al., 2016).

Quercetin

Quercetin, an important polyphenol found in *Ficus carica*, has the ability to improve learning and memory when used in the treatment of AD (Biswas et al., 2015). It is considered as a multifunctional neuroprotective agent (Ossola et al., 2009). Currently, the effect of oral administration of nanoencapsulated quercetin was determined in a mouse model of AD. It was observed that the oral administration of nanoencapsulated quercetin in zein NPQ reduced the cognition- and memoryimpairment characteristics of senescence-accelerated prone mouse (SAMP8). These observations appeared to be related with a decreased expression of the hippocampal astrocyte marker GFAP (Puerta et al., 2017).

Quercetin is reported to significantly cross BBB (Faria et al., 2014), accumulate in the target brain tissues of animal models, and reduce the oxidative stress-mediated neurodegeneration (Ishisaka et al., 2011). Moreover, quercetin is also reported to exhibit neuroprotective effects via inhibition of NO excessive production and overexpression of iNOS and downregulation of pro-inflammatory gene expressions like TNF- α , COX-2, and IL-1 β in zebrafish (Zhang et al., 2011). Nanoencapsulated quercetin is reported to inhibit ischemic reperfusion-mediated neuronal damage in animal models (Ghosh et al., 2013).

Bilobalide and Ginkgolide

Ginkgo biloba (GB) contains very important constituents such as bilobalide and ginkgolide. They have been extensively studied for their CNS effects (Maclennan et al., 2002) and can be used for the possible treatment of AD patients (Ahlemeyer and Krieglstein, 2003; Shi et al., 2010). The dietary treatment of mice with GB extract upregulates the expression of genes encoding neuronal tyrosine/threonine phosphatase 1 and microtubule-associated tau in the cerebral cortex. The GB extract can also upregulate two mitochondrial DNA-encoded genes, subunit III of cytochrome-c oxidase and NADH dehydrogenase, subunit 1 (ND1), indicating a fundamental mechanism that may underlie bilobalide-induced neuroprotection and its role in cognitive impairment (DeFeudis, 2002). Recently, it has also been reported that the ectopic expression of APE1 enables neuronal cells to overcome the oxidative damage caused by $A\beta_{25-35}$. Moreover, ginkgolide has modified the mitochondrial OXPHOS against AB25-35-induced oxidative stress and regulated the reactive oxygen and/or nitrogen species (ROS/RNS) level in the existence of ectopic APE1. This study portrays an alternative approach in the therapeutic potential for AD by harnessing the synergistic neuroprotective roles of apurinic/apyrimidinic endonuclease 1 (APE1) and ginkgolide (Kaur et al., 2015).

Ginkgolides A, B, C, J, and bilobalide isolated from *G. biloba* significantly cross BBB using *in vitro* and *in vivo* permeability models and inhibit oxidative stress-induced cognitive dysfunctions (Sharma et al., 2000; Madgula et al., 2010). In another brain permeability study, ginkgolide B passed BBB and inhibited reperfusion-induced ischemic brain damage (Fang et al., 2010; Lang et al., 2010).

Caffeine and Chlorogenic Acids

Coffea arabica is well known for the coffee, which is a common drink linked to a number of health benefits, including its key role to intervene AD. Different coffee varieties contain caffeine and chlorogenic acids in different concentrations, which possess neuroprotective effects observed in various *in vitro* and *in vivo* studies (Rebai et al., 2017). Recently, nicotinamide mononucleotide adenylyl transferase 2 (NMNAT2) was reported as a key neuronal factor that delivers strong neuroprotection in numerous preclinical models of neurological disorders. Systemically administered caffeine was identified as a positive modulator that restores NMNAT2 expression in tauopathy mouse model (rTg4510) to normal levels (Ali Y.O. et al., 2017). Caffeine and 5-caffeoylquinic acid (chlorogenic acid)-enriched coffee extensively crosses BBB and exhibits numerous CNS effects including neurotropic, anti-AD, and neuroprotective properties (Ito et al., 2008; Chu et al., 2009; Kwon et al., 2010; Cropley et al., 2012; Esquivel and Jiménez, 2012; Oboh et al., 2013).

Glycyrrhizin

Glycyrrhizin (GRZ) is a triterpenoid saponin, majorly found in *Glycyrrhiza glabra*. The GRZ exhibited spatial memoryenhancing and ameliorating effects on cognitive impairment induced by A β injection into the hippocampus *in vitro* and *in vivo* (Grzybowska et al., 1998; Chen et al., 2017). Currently, it is being reported that GRZ effectively reduced neuroinflammation and ameliorated the memory deficits induced by systemic LPS treatment. The effects of GRZ were found to be mediated through inhibition of pro-inflammatory cytokines TNF- α , IL-1 β , and microglial activation in the brain tissues. This study supports that GRZ may be a putative therapeutic drug for neurodegenerative diseases that are associated with cognitive deficits such as in AD (Song et al., 2013).

The BBB permeability of 18β -glycyrrhetinic acid, an active metabolite of GRZ, was reported by Tabuchi et al. (2012) using *in vitro* and *in vivo* models. Study results revealed that the test compound, after oral administration, significantly crossed the BBB in both the *in vitro* BBB permeability model as well as animal models (Tabuchi et al., 2012). Oral pretreatment with GRZ is reported to thwart surgery-mediated cognitive dysfunctions via inhibition of neuroinflammation and AD symptoms (Chen et al., 2017).

Choline

Choline is required for the production of acetylcholine, an important neurotransmitter for memory, brain, and nervous system functions. It also plays important roles in modulating gene expression, cell membrane signaling, lipid transport and metabolism, and early brain development. Nutritional intake of choline in AD patients can influence cognitive functions via an effect on phosphatidylcholine (PC)-containing eicosapentaenoic and docosahexaenoic acids and polyunsaturated species of PC, levels of which are reduced in brain that is linked with higher memory performance (Blusztajn et al., 2017). In a current study, perinatal choline supplementation was reported to reduce amyloidosis and increase choline acetyltransferase expression in the hippocampus of APPswePS1dE9 (APP/PS1) AD mice. The data propose that dietary intake of choline during fetal development and early postnatal life can establish a preemptive approach for AD (Mellott et al., 2017).

Huperzine

Huperzine A (HupA) is a lycopodium alkaloid found in *Huperzia serrata*. The HupA was reported as an effective cholinergic agent. It is also effective in acute seizures as tested in a rat model. Therefore, HupA is a safe drug for AD patients who show both mnemonic and epileptic symptoms (Rafii et al., 2011). Moreover, it has also been reported that HupA can penetrate the BBB due to its interaction with efflux transmembrane transporters (ABCB1 and ABCG2). In transgenic animal models, the brain-to-plasma concentration ratio of Huperzine A was significantly increased as compared with the wild type mice due to ABCB1 that played a predominant role in the efflux of Huperzine A across BBB (Li et al., 2017).

Ginsenoside

Ginsenosides Rg1 (GRg1) and Rb1 (GRb1) are found to be the active ingredients of ginseng, which exhibits potential efficacy in prevention and treatment of CNS disorders and neurodegenerative diseases like AD. Currently, Rb1 and baicalin have been reported to promote proliferation and differentiation of endogenous neural stem cells (NSCs) in AD rat models. It also shows a decreased nuclear pyknosis and pyramidal cell defects in hippocampus of rats by increasing the expression of nestin, GFAP, and neuron specific-enolase (NSE) proteins and thereby improves cognitive function in AD rats (Zhao et al., 2018). Recently, the neuroprotective effects and its transport across BBB was reported by Rui et al. (2009) using in vitro and in vivo rat models. Ginsenoside Rg1 exhibited significant neuroprotective potentials, whereas, its passage across BBB was extremely poor. Ginsenoside Rg1 ameliorates cerebral ischemia via downregulation of protease-activated receptor 1 (PAR-1) expression (Xie et al., 2015) and offers protection against BBB disruption in brain injury model (Zhou et al., 2014; Chen W. et al., 2015).

Magnolol and Honokiol

Magnolol and honokiol are the bioactive compounds found in Magnolia officinalis. They are neuroprotective and decrease ROS production, intracellular calcium and caspase-3 activities. Therefore, these phytochemicals or their analogs may be explored as therapeutic drugs for AD (Hoi et al., 2010). Honokiol was reported as to reduce ABO-induced hippocampal neuronal apoptosis, ROS production, and loss of mitochondrial membrane potential in a dose-dependent manner. Moreover, honokiol inhibited ABO-mediated NF-KB activation as well as inhibited the up regulation of amyloid precursor protein (APP) and beta secretase enzyme (BACE1). Therefore, honokiol may be a potential candidate in AD therapy (Wang et al., 2017). Honokiol is reported to cross BBB and effectively target CNS disorders like gliosarcoma (Wang et al., 2011) and cerebral ischemia via inhibition of NF-kB (Zhang et al., 2013) and offer protection against brain ischemic-reperfusion injury via disruption of PSD95-nNOS interactions (Hu et al., 2013).

Shogaol and Gingerol

Zingiber officinale contains two major therapeutic agents, shogaol and gingerol, that could recover the SHSY-5Y cells from $A\beta_{1-42}$

oligomer and aggregate toxicity. Currently, 6-gingerol and 6shogaol have been found as neuroprotective agents, which can inhibit the GSK-3 β activity (Yerer et al., 2017). It was found that 6-shogaol influenced neuritogenic activity in PC-12 cells by activating the MAPK, extracellular signal-regulated kinase 1 and 2 (ERK1/2), phosphatidylinositol 3-kinase (PI3K), and protein kinase B (AKT) signaling pathways. Hence, 6-shogaol can act as a NGF mimicking agent, which can be used as a preventive therapeutic agent in neurodegenerative diseases (Seow et al., 2017).

Crocin

Crocin is a water-soluble carotenoid found in *Crocus sativus* and possesses the capability to improve learning and memory as well as protect brain cells. Recently, crocin has been reported for its considerable effects on $A\beta$ and tau proteins. Crocin revealed multifunctional protective activities in the brain and could be a promising agent when applied as a supplement or drug for the treatment of AD (Finley and Gao, 2017). Crocin has been reported to improve locomotor activities in animal models (Karami et al., 2013), provide protection against oxidative stress and cerebral ischemia (Zheng et al., 2007), and maintain BBB integrity (Zhang et al., 2017).

Benzylisoquinoline

Cissampelos pareira contains benzylisoquinoline that exerts nootropic activity in AD. It is a memory-enhancing agent, inhibits AChE activity, and enhances memory in mice (Kulkarni et al., 2011). A novel series of benzylisoquinoline derivatives have also been evaluated. The results have shown that most of the compounds have significant inhibitory potentials against ChEs (James et al., 2016), human cholinesterases (h-ChEs), and self-induced A β aggregation (Xu et al., 2014).

Monoterpenoid

Monoterpenoids in *Lavandula luisieri* have been identified as potent inhibitors of β -secretase (BACE-1), an aspartic protease involved in the conversion of APP to A β in animal models. Currently, an active monoterpene, necrodane ketone, 2,3,4,4-tetramethyl-5-methylenecyclopent-2-enone was tested for its inhibitory mechanism against BACE-1. This monoterpene revealed a dose-dependent inhibition of BACE-1 in cellular and mouse models of AD (Videira et al., 2014). Recently, 80 types of aroma compounds including monoterpenes were screened for *in vitro* inhibitory activity at a concentration of 200 μ M against recombinant human BACE1 (Marumoto et al., 2017).

Bacoside and Reserpine

Bacopa monnieri, known as a memory-enhancing herb, contains key therapeutic agents bacoside and reserpine. It promotes free radicals scavenger mechanisms and protects cells in the prefrontal cortex, hippocampus, and striatum against cytotoxicity and DNA damage implicated in AD. It protects the cholinergic neurons and reduces anticholinesterase activity comparable with donepezil, rivastigmine, and galantamine (Chaudhari et al., 2017). The mechanism of action of reserpine was determined in *Caenorhabditis elegans* where it alleviated the toxicity of $A\beta$. The afore-mentioned study was conducted to evaluate the ~2.8 kb promoter region of FLP-11, driving the expression of green florescent protein, which can be a critical mediator of alleviation of A β toxicity through acetylcholine and reserpine (Saharia et al., 2016). The PLGA-loaded bacoside-A NPs are reported to effectively deliver the drug across BBB and mediate its neuropharmacological actions (Jose et al., 2014; Chaudhary and Bist, 2017).

Catechins

Catechins found in *Camellia sinensis* prevent cognitive impairment among the elderly. It provides protection to the brain and modulates the risk factors of AD (Preedy, 2012). Currently, the human-derived cell line (SH-SY5Y) was treated with native oligomers (epicatechin and theaflavin) that reduce A β -induced BACE1 expression. The data show that the antioxidant activity of catechins and theaflavins may be more essential in downregulating BACE1 mRNA expression than their capability to inhibit A β oligomerization (Geiser et al., 2017). Catechins can cross the BBB and mediate various neuroprotective effects (Faria et al., 2011; Wu et al., 2012).

S-allylcysteine

Garlic extract (GE) contains *S*-allylcysteine, which has been reported for multiple biological activities, including antiinflammatory effect and its influence on $A\beta_{1-42}$ -induced cognitive dysfunction and neuroinflammation (Kodera et al., 2002; Pérez-Severiano et al., 2004). The GE was found to be useful by improving the short-term recognition memory and relieve the neuroinflammation in Aβ-induced adult male Wister rats (Nillert et al., 2017). Recently, a research group has reviewed different antioxidant mechanisms (scavenging of free radicals and prooxidant species, induction of antioxidant enzymes, activation of Nrf2 factor, inhibition of prooxidant enzymes, and chelating effects) involved in the protective actions of aged GE (AGE) and S-allyl-L-cysteine (SAC), thereby, emphasizing their potential use as therapeutic agents (Colín-González et al., 2012).

Treating Alzheimer's: A Triple Challenge

Treating brain diseases, in general, and AD, in particular, is a special challenge. First of all, the intricate and specific system that the brain possesses in the form of BBB analyzes and monitors the transport of every molecule that crosses the brain. It consists of endothelial cells, astrocytes, pericytes, and microglial cells (Johansson, 1990). The highly tight junctions between the endothelial cells act as a border patrol system, which analyze the molecules on the basis of their lipophilicity, size, surface charge, and hydrogen-bonding potential. Priority is given to small lipophilic molecules with size less than 500 Da and the entry of these molecules is restricted, which do not fall under the specific criteria of structural and physicochemical properties (Abbott et al., 2010). In addition, even if the molecule of choice does get entry across the BBB, there is a high chance of it being thrown out due to the number of efflux transporters such as the ATP-binding cassette family (P-glycoprotein ABCB1), the multidrug resistance-related proteins (MRPs, ABCC1, 2, and 5) and the breast cancer resistance protein (BCRP, ABCG2) present in the endothelium of the brain _ENREF_3 (Begley, 2004; Ayaz et al., 2017d).

In addition to the transport-related issues, limited knowledge of the basic pathology of AD is another challenge in more effective drug therapy (Sperling et al., 2013). Many mechanisms leading to the pathology have been proposed, and the disease is attributed to be the result of a number of overlapping phenomena such as inflammation, iron dysregulation, oxidative damage, cholesterol metabolism (Galimberti and Scarpini, 2010), and "amyloid plaque" hypothesis. Despite this, there is no FDAapproved drug or carrier system with the disease-modifying potential for AD (Casey et al., 2010). Moreover, the third and major limitation in the successful therapy of AD is the absence of tools for early detection of the disease. Biochemical alterations at the cellular level start around 10-20 years before the onset of cognitive decline (Beason-Held et al., 2013). It has been proposed that the failure of various therapeutic approaches may be attributed to the fact that until the time of appearance of clinical symptoms, irreversible damage has already been done.

Currently Approved Drugs for Alzheimer's and Their Shortcomings

Until now, the FDA has approved drugs only for the symptomatic treatment of AD. The five that have been approved target neurotransmitter imbalance at different stages of disease; they include AChE inhibitors; tacrine (1st generation), donepezil, rivastigmine, and galantamine (2nd generation) (Ayaz et al., 2015). One of the five approved drugs by the FDA for the symptomatic treatment of AD is an NMDA receptor blocker, memantine. The pharmacokinetic profile of drugs affecting acetylcholine and NMDA is given in **Table 2**.

Despite the beneficial effects of improvement in everyday life of patients, reduction in emotional impact on caregivers, and overall care cost, the large variations between the effectiveness and efficacy of these drugs make their cost-effectiveness controversial (Loveman et al., 2006). This large gap in the efficacy profile is majorly due to peripheral stimulation of cholinergic system that results in gastrointestinal tract (GIT)related side effects. For instance, GIT-based adverse drug reactions (ADRs) were the major causes for trial discontinuation, with mean frequency of nausea being 11% for donepezil, 44% for rivastigmine, and 24% for galantamine across various trials

TABLE 2 The pharmacokinetic profile of EDA-approved drugs for AD

of AChE inhibitors (AChEI) (Casey et al., 2010). Moreover, these drugs have shown poor long-term maintenance of the improvement of symptoms with studies showing that benefits of AChEI can be endured for up to 4 years with decline in the cognition below baseline levels after 1 year of therapy (Winblad and Jelic, 2004).

All these drawbacks can be attributed to the poor pharmacokinetic profile of these therapeutic moieties, including low bioavailability, volatility, oxidation, hydrolysis, limited transport across BBB, and tendency for drug-drug interactions (Mangialasche et al., 2010). As AD majorly affects the elderly, the comorbidities, polypharmacy, and non-compliance have a huge impact in decreasing the efficacy of the treatment regimen (Noetzli and Eap, 2013). Therefore, there is a major need for the development of multipurpose moieties with potent therapeutic potential against AD.

Mechanistic Basis of the Nanoparticle-Based Treatment

The unknown pathology and increase in the elderly population point toward a growing need for the development of therapeutic molecules specific to AD etiology. Many researchers around the world have developed NPs aiming at a different molecular mechanism of AD, in the hope of pinpointing the basic cause or limiting factor (Nazem and Mansoori, 2011; **Table 3**). There are four major and basic points in the pathophysiology of AD that can be targeted by use of NPs. (1) Neurotransmitter modulators; (2) anti-amyloid beta immunotherapy; (3) metal stress reducers, and (4) inflammatory inhibitors.

Neurotransmitter Modulators

It is known that presynaptic deficits of Ach contribute significantly toward cognitive dysfunctions and induction of memory impairment (Ullah et al., 2016). Interestingly, it is noted in animal models that degeneration of neurons in the basal forebrain and medial septum leads to substantial decline in Ach levels in the cerebral cortex and hippocampus, respectively. Moreover, the biochemical evaluation of brain from patients with AD also showed considerable deficits in the neocortical levels of AChE, an enzyme involved in the synthesis of Ach (Ayaz et al., 2014). The NPs capable of transporting the acetylcholine-modulating drugs across the BBB

Name	Vdl	Log p	Molecular weight g/mol	Formulation (oral)	References			
				Excretion [renal and hepatic]	T _{1/2}			
Memantine	9 to 11 L/Kg	3.3	179.307 g/mol	148% of administered drug is excreted unchanged in urine	60–100 h	(1)		
Rivastigmine	1.8 to 2.7 L/kg	2.3	250.342 g/mol	Renal clearance = 2.1–2.8 L/h	1.5 h	(2)		
Donepezil	12 L/kg	3.6	379.5 g/mol	0.13 L/h/kg in urine	70 h	(3)		
Galantamine	175 L	1.8	287.359 g/mol	300 mL/min [After IV. or oral administration] in urine	7h	(4)		

(1) Drug Bank. Memantine. 2018, September 29 Available from: https://www.drugbank.ca/drugs/DB01043.

(2) Drug Bank. Rivastigmine. 2018, September 29 Available from: https://www.drugbank.ca/drugs/DB00989.

(3) Drug Bank. Donepezil. 2018, September 29 Available from: https://www.drugbank.ca/drugs/DB00843.

(4) Drug Bnak. Galantamine. 2018, September 29 Available from: https://www.drugbank.ca/drugs/DB00674.

TABLE 3 | Phytochemicals used as drug delivery agents for synthesis of NPs in treatment of AD.

Phytochemical NPs	Mechanism of action	Reference
Curcumin-NPs	-Oral administration of nanocurcumin to Tg2576 AD model mice even at a low dose (23 mg/kg per week) resulted in significant improvements over placebo control in working and cue memory. The novel nanocurcumin has a great potential for AD therapy.	Cheng et al., 2013
	-Curcumin-encapsulated PLGA nanoparticles (Cur-PLGA-NPs) potently induce neural stem cells (NSCs) proliferation and neuronal differentiation <i>in vitro</i> and in the hippocampus and subventricular zone of adult rats, as compared to uncoated bulk curcumin.	Tiwari et al., 2013
Quercetin-NPs	-PLGA-functionalized quercetin (PLGA@QT) NPs showed negligible cells toxicity and inhibited Aβ ₄₂ fibrillation and reduced Aβ ₄₂ -induced toxicity. Morris water maze and Novel object recognition tests showed an enhanced learning and memory of AD mice and proposed it as therapeutic drug for AD.	Sun et al., 2016
	-Quercetin nanoparticles (NQC) enhancement has shown increased efficacy and prolonged residence time in systemic circulation and increased bioavailability in rates. The NQC offers the potential clinical application in human neurodegenerative disease in future.	Palle and Neerati, 2017
Ginsenoside-NPs	-Poly (lactic-co-glycolic acid) (PLGA)-NPs encapsulating ginsenoside Rg3 and Thioflavin T(Aβ diagnostic) were examined for their neuroprotective effects. Key mechanisms were investigated for their neuroprotective effects and evaluated their ability to cross the BBB using an <i>in vitro</i> BBB model. The PLGA-Rg3 NPs offer an exciting new theranostic material capable of encapsulating natural nutraceuticals for the detection and treatment of AD.	Aalinkeel et al., 2018
	-Ginsenoside Rg1 nanoparticle has potential to penetrate the blood-brain barrier to improve the cerebral function of diabetic rats complicated with cerebral infarction.	Shen et al., 2017
Bacoside-NPs	-Bacoside-A-loaded PLGA nanoparticles surfaces were modified by coating with polysorbate 80 to facilitate the crossing of the blood–brain barrier (BBB). When compared to pure drug solutions ($2.56 \pm 1.23 \mu$ g/g tissue), an <i>in vivo</i> study using Wistar albino rats demonstrated higher brain concentrations of Bacoside-A ($23.94 \pm 1.74 \mu$ g/g tissue) suggesting a significant role of surface-coated nanoparticles on brain targeting.	Jose et al., 2014
	-Bacoside as a model drug was incorporated into solid lipid nanoparticles (SLNs) prepared from stearic acid using Tween 80 emulsifiers, and <i>in vitro</i> drug release study showed up to 84.68% drug release from SLNs.	Vitthal et al., 2013
Huperzine-NPs	-Huperzine A using lactoferrin-conjugated <i>N</i> -trimethylated chitosan surface-modified PLGA NPs (Lf-TMC NPs) were developed for intranasal delivery to the brain for the treatment of AD. After intranasal administration, Lf-TMC NPs facilitated the distribution of HupA in the brain, and the values of the drug targeting index in the mouse olfactory bulb, cerebrum (with hippocampus removal), cerebellum, and hippocampus were about 2.0, 1.6, 1.9, and 1.9, respectively.	Meng et al., 2018
Choline -NPs	-Choline transporter (ChT) was used to transport choline-derivate-modified NPs across the blood-brain barrier. These modified NPs exhibited higher permeability across the brain capillary endothelial cells (BCECs) monolayer <i>in vitro</i> and higher gene distribution and expression <i>in vivo</i> .	Li et al., 2011
Lectin -NPs	-Basic fibroblast growth factor (bFGF) was entrapped in nanoparticles conjugated with <i>Solanum tuberosum</i> lectin (STL), which selectively binds to <i>N</i> -acetylglucosamine on the nasal epithelial membrane for its brain delivery. The spatial learning and memory of AD rats in the STL-bFGF-NP group were significantly improved compared with the AD model group proposing it a good brain drug delivery system for peptide- and protein-based drugs.	Zhang C. et al., 2014

should restore the imbalance of the Ach in AD patients and should ameliorate the deterioration of memory and improve cognitive function. It was reported that rivastigmine-loaded lipid NPs (RL) have successfully normalized the cortical Ach level and subsequently improved the deterioration of spatial memory in animal models treated with AlCl₃ (Ismail et al., 2013). In their study, rats that were coadministered with RL and AlCl₃ showed normalization of BACE1 gene expression as compared with its 316% overexpression in rats treated with AlCl₃ only. Furthermore, the histological evaluation showed that coadministration of RL prevented the formation of A β plaque and, hence, strengthens the argument that normalization of Ach is associated with an anti-Alzheimer effect.

Amyloid Beta-Targeted Immunotherapy

The A β s were considered as therapeutic agents for active immunity after it was noted that transgenic AD mouse developed anti-A β antibodies when immunized with A β_{42} . The antibody development resulted in improved cognition and decreased

amyloid plaque number. The mechanism involves binding of anti-amyloid antibody to the amyloid plaque followed by the disaggregation of plaques and, hence, removal from the brain (Modi et al., 2010). Previously, external brain molecules like heparin and gelsolin were thought to trap or "sink-in" the $A\beta$ and, hence, decrease its concentration in the brain (Fonseca-Santos et al., 2015). Both active and passive immunotherapy targeting $A\beta$ forms the leading approach in the treatment of AD.

For active immunotherapy, $A\beta$ subfragments were used in combination with NPs as their delivery across BBB is low. This approach was used by Songjiang and Lixiang (2009) to develop intramembranous $A\beta$ fragment-loaded chitosan NPs for enhanced brain delivery. The ELISA data from the study showed that the formulation has significant potential to induce immunogenicity (Songjiang and Lixiang, 2009). Similarly, Agyare et al. (2008) developed chitosan polymeric NPs with tripolyphosphate (TPP) and loaded them with the polyaminemodified F(ab') portion of anti-amyloid antibody (IgG4.1). The biodistribution studies in mice and transport study across bovine BMECs (BBMECs) showed that NPs were able to trancytose across BBMEC, and, hence, are proposed to target brain amyloid (Agyare et al., 2008).

In addition to active immunity with $A\beta$ fragments, passive immunization is widely exploited for potential therapeutic regimen in AD, as indicated by several $A\beta$ -specific antibodies being present in various phases of clinical trials (**Table 4**). However, very few studies have exploited antibodies in combination with NPs for the treatment of AD. Currently, the first synthetic study of $A\beta_{1-42}$ monoclonal antibody-tagged polymeric NPs (125 nm) has been reported for the treatment of AD in transgenic (Tg) 2576 mice. The study showed that animals treated with mAb-tagged NPs exhibit significant reduction in the soluble $A\beta$ peptide and its oligomer level in the brain and considerable increase of the $A\beta$ levels in plasma. Moreover, the results demonstrated complete recovery of memory deficits in Tg AD mice, which further strengthens the fact that antibody-mediated treatment may provide the ultimate long-term and personalized therapeutic strategy for AD treatment (Carradori et al., 2018).

Metal Stress Modulators

It is evident that metabolism of zinc, cupric, and iron metal ions regulates A β toxicity, senile plaques (SPs), and neurofibrillary tangles (NFTs) formation. It has also been reported that SP and AB plaques are enriched with copper, iron, and zinc (James et al., 2016). In addition to promoting the conversion of A β to oligomers and fibrils and hyperphosphorylation of tau, these metal ions gets reduced when they undergo binding with A β , leading to generation of H₂O₂ – the main source of oxidative stress in A β toxicity (França et al., 2017).

Metal chelators loaded onto NPs can easily gain access to brain, bind to the metal particles, and leave the brain along with their conjugated metal ions and, hence, provide an easy means to reduce metal stress. Cui et al. (2005) reported the synthesis of Cu (I) chelator D-penicillamine conjugated ethanolamine-based

Name	Company	Phase trial	Observations	Target site	Nature	Reference
Solanezumab	Lilly, Roche, Alzheimer's Association	11/111	Slowing of cognitive and functional decline versus placebo Baseline to endpoint changes in levels of CSF total tau and p-tau did not differ between treatment groups	monomeric Ab	lgG1 Anti amyloid monoclonal antibody	Siemers et al., 2016
Aducanumab	Biogen Inc.	lb	Slowing in clinical decline Dose-dependent decrease in amyloid beta	amyloid-β (Αβ) plaques, tau	Anti amyloid monoclonal antibody	Sevigny et al., 2016
CAD106	Novartis, Amgen, NIA, Alzheimer's association	II	Sustained Aβ-IgG titers and prolonged time to decline were observed in extensions versus core studies	Ab1-6, active vaccine	Anti amyloid vaccine	Farlow et al., 2015
Verubecestat	Merck	11/111	No clinically beneficial effect Reduction in plasma, brain, and CSF concentration of Ab40, Ab42, and sAPPb	BACE1	BACE1 inhibitor	Kennedy et al., 2016
Bapineuzumab	Janssen Alzheimer Immunotherapy and Pfizer	III	No improvement in clinical outcomes of patient Reduction in CSF level of phospho-tau May modify Ab, but no clinical benefit (limitation of study design)	Amyloid beta	humanized anti-amyloid- beta monoclonal antibody	Salloway et al., 2014
Gantenerumab	Roche, Lilly, Alzheimer's Association	I	mAb targeting aggregated Ab	Aggregated Ab	Anti-amyloid mAb	Bohrmann et al., 2012
Crenezumab	Genentech	11/111	-	Soluble oligomer and fibrillar Ab	Anti-amyloid mAb	Cummings et al., 2016
AADvac1	Axon Neuroscience	1,11	-	Tau epitope	Anti-tau mAb	Cummings et al., 2016
ABBV-8E12	AbbVie	II	-	tau	Anti tau monoclonal anti body	Cummings et al., 2016
BAN2401	Eisai	II	-	N terminal protofibrils	Monoclonal anti body	Cummings et al., 2016
NewGam 10% IVIG	Sutter Health	II	-	Multiple forms of $A\beta$	Polyclonal antibody	Cummings et al., 2016
LY3314814	Lilly	11/111	Being studied for MCI to mild AD	BACE	BACE inhibitor	Cummings et al., 2016
CNP520	Novartis	11/111	Being studied for asymptomatic (homozygote APOE ε4/ε4)	BACE	BACE inhibitor	Cummings et al., 2016

TABLE 4 | Properties of some important antibodies and BACE inhibitors at different phases of trials.

NPs. The study showed successful uptake of NPs in the brain, and the NPs along with released penicillamine were able to resolubalize more than 40 % of Cu–Ab conjugates when treated with 0.1 N NaOH (Cui et al., 2005). Similarly, Liu et al. (2009) reported the syntheses of polystyrene-based – iron chelator conjugate (Nano-N2PY) and its ability to inhibit A β aggregation-mediated cytotoxicity in human cortical neuronal cells. These preliminary *in vitro* studies can provide a strong background for further *in vivo* evaluation of metal-conjugated NPs as AD therapeutics.

Anti-inflammatory Agents

It is established that oxidative stress related to AB toxicity results in mitochondrial damage, which leads to cell death and neuronal damage. Anti-inflammatory and antioxidant agents, such as CoQ₁₀, have established a role in mitochondrial bioenergetics and reduction of oxidative stress secondary to AB toxicity (Garrido-Maraver et al., 2014). The CoQ₁₀-loaded trimethylated chitosan (TMC)-PLGA-based NPs (TMC-coated D,L-lactide-coglycolide) were synthesized, and it was investigated that the drug-loaded TMC-PLGA NP formulation was biocompatible, showed higher accumulation in the brain, and with great reduction in memory impairment, restoring it to almost normal level in comparison with CoQ10-loaded PLGA NPs without TMC. The formulation also helped decrease AB concentration and $A\beta$ fibril formation; this can be attributed to the effect of Co Q10 on dissolution of SP (Garrido-Maraver et al., 2014).

The NPs as Anti-Alzheimer Moieties; Overcoming the Existing Barriers

Traditional drugs used for AD have low bioavailability, limited transport, and controversial long-term effects; therefore, a tremendous number of studies have reported on the use of nano-scale particles to overcome these limitations. There are different strategies that are employed to use NPs of various origins to improve pharmacokinetic profiles; (i) increase targeted delivery of drugs across BBB (Rocha, 2013), (ii) increase efficacy of already available approved agents, and (iii) provide a multifunctional platform that can help in theranostic applications.

The NPs by virtue of their small size, modifiable surface properties, and tunable morphology act as ideal candidates to overcome all the hurdles that a brain drug delivery (DDR) system needs to evade (Lockman et al., 2002). With the large loading capacity, NPs are good candidates for carrying increased amounts of drug across the BBB and increasing the brain-to-blood ratio of the already available drugs (Fazil et al., 2012).

Various strategies have been adopted to enhance the effect of these tiny cargos for better therapeutics.

- (1) Lipophilic NPs like liposomes for enhanced permeation of hydrophilic drugs across BBB (Smith et al., 2010).
- (2) Hydrophilic polymer-based NPs are employed to impart stealth character and act as a parenteral sustained drugrelease system by prolonging the blood retention time (Wilson et al., 2010). Thus, by providing a higher concentration gradient between blood and brain, they

facilitate the passive diffusion of drug across BBB (Masserini, 2013).

- (3) Mucoadhesive polymeric NPs are used for enhanced nasal delivery of drugs through direct endocytosis-mediated uptake across BBB (Mistry et al., 2009).
- (4) Targeted NPs with surface ligands for receptor-mediated transcytosis (Ulbrich et al., 2009).

Conventional NPs

Nanoparticles that consist of polymers, metal, or lipids without any delivery enhancing or targeting moiety to guide their transport are sometimes referred to as conventional NPs. These were the first ones to be used in AD nano-therapeutics.

Polymer-Based Delivery System

Polymeric NPs have been used extensively for AD drug delivery to enhance the efficacy of already available drugs.

Chitosan NPs

Chitosan is a biocompatible, biodegradable, and non-toxic (De Campos et al., 2001) linear polyamine polymer with reactive amine and hydroxyl surface groups (Dutta et al., 2004). Owing to the presence of reactive sites, this harmless polymer has chameleon-like characteristics. By virtue of these surface groups, it can have various modifiable properties like (i) adjustable drugloading capacity_ENREF_3, (ii) hydrophilic–lipophilic balance (HLB) value, (iii) pH sensitivity, (iv) controlled drug release, (v) targeting ability, and (vi) surface adsorption (Shu and Zhu, 2000; Liang et al., 2008; Wang et al., 2008).

The chitosan backbone can be chemically altered to adjust the HLB value of the resultant polymer (Agnihotri et al., 2004). It is suggested that the hydrophilic polymer-based DDR system that prolong the blood retention time may act as a parenteral sustained drug-release system. This characteristic of chitosan has been utilized in increasing the brain delivery of anti-Alzheimer drugs that can freely cross the BBB (Wilson et al., 2010). Chitosan has also been used to increase the brain-to-blood ratio of drugs with poor ability to cross BBB. This was achieved by using the muco-adhesive properties of the polymer by Fazil et al. (2012) in developing rivastigmine-loaded chitosan NP for intranasal delivery of drug to brain.

PLGA NPs

Poly (lactic-co-glycolic acid) forms one of the most attractive nanomaterials to synthesize NPs for Alzheimer's treatment. Its properties of biocompatibility, biodegradability, modifiable surface properties, stealth character, and ability to develop modified and sustained drug releasing nano-depots make it a perfect nanodevice (Danhier et al., 2012).

Md et al. (2014) prepared nano-particulate DDR system for donepezil using PLGA. The formulation showed sustained release and efficient brain targeting in SD rats (Md et al., 2014). Similarly, in another study, Wang et al. (2010) showed that coenzyme Q(10)-loaded TMC surface-modified NPs (TMC/PLGA-NP) greatly improved the memory impairment in mice. Senile plaques and biochemical parameters also showed



FIGURE 1 | Injected targeted liposomes cross the blood-brain barrier of APP/PSEN1 mice and labeled parenchymal Ab deposits. Composite images (A–E) of olfactory bulb (A) showing plaque pathology within the granule cell layer. Nanoparticles bound to plaques at the level of the septo-striatum (B). Cortical pathology at the septo-diencephalic (C). Hippocampal and cortical pathology (D) within the caudal diencephalon. Binding to cerebellar plaques (found within APP mice) within the rostral mesencephalon (E). An example of cortical plaque pathology (F) visualized at 10× magnification. Hippocampal plaque pathology (G) is similar to previously reported studies. Scale bar = 100 mm (adopted with permission from Tanifum et al., 2012).

brain targeting potential of the TMC/PLGA-NPs (Wang et al., 2010).

Cyanoacrylate NPs

Cyanoacrylate polymer provides a biocompatible and bioerosive medium for the synthesis of targeted NPs for treatment of AD. Its modifiable surface properties provide a very good medium for development of the sustained release drug delivery vehicle with long blood circulating times (Yordanov, 2012). Wilson et al. (2008) used poly (*n*-butyl cyanoacrylate) NPs for enhanced delivery of tacrine and rivastigmine. The group developed rivastigmine- and tacrine-loaded polysorbate 80-coated poly (*n*butyl cyanoacrylate) NPs in two different studies. It was noted that there was a 3.82- and 4-fold increase in the concentration of rivastigmine and tacrine in brain, respectively, with the nanoformulation as compared with the free drug (Wilson et al., 2008).

Liposomal NPs for AD Drug Delivery

Liposomes are colloidal particles prepared by exposing phospholipids to the aqueous phase (Anwar et al., 2017). They facilitate loading of hydrophilic drugs into the aqueous interior of liposomes, thus preventing them from acid degradation, first pass effect, and delayed renal clearance (Smith et al., 2010).

Mutlu et al. (2011) have developed multilamillar rivastigmineloaded liposomal NPs associated with permeability enhancer-like sodium-taurocholate or dimethyl-beta-cyclodextrin. Rivastigmine-sodium-taurocholate liposomes showed the highest apparent permeability coefficient in Madin-Darby Canine Kidney cells monolayer and the highest AChE inhibition in Balb-C type mice when administered through oral/intraperitoneal route (Mutlu et al., 2011). Similarly, Smith et al. (2010) have reported increased oral bioavailability by twofold and improved neuronal (SweAPP N2a cells) α -secretase inducing ability by 91% *in vitro* of epigallocatechin-3-gallate (EGCG) with use of nano-lipidic particles.

Stealth NPs

As BBB favors delivery of lipophilic, neutral, and small-sized particles, most of the early NPs employed for brain delivery of therapeutic agents are hydrophobic in nature (Sahni et al., 2011). Conventional NPs overcome a number of hindrance factors associated with conventional oral drug delivery such as acid degradation, first pass effect, and low brain-to-blood ratio (Di Stefano et al., 2011). The phenomenon of surface opsonization and subsequent removal from the bloodstream through reticuloendothelial system and Kupffer cells in liver and spleen leads to substantial decrease in the systemic and local therapeutic drug concentration. In the blood, hydrophobic and charged NPs undergo rapid surface opsonization within seconds to minutes as compared to hydrophilic or neutral NPs. This surface property is used to prepare the so-called "stealth" NPs with long blood-circulating times by coating them with specific polymer. Typical characteristics of polymer used to induce stealth character to NPs include flexibility and hydrophilicity. Several polymers that are used for such purposes in the brain DDR system include natural; chitosan, semisynthetic; polyethylene glycol (PEG), poly vinyl alcohol (PVA), polyvinylpyrrolidone (PVP), PEG-based copolymer; like poloxamer, poloxamines, and polysorbates (Salmaso and Caliceti, 2013). Tanifum et al. (2012) synthesized stealth liposomal NPs incorporated with AB-targeting fluorescent lipid conjugate, 1,2-distearoyl-sn-glycero-3-phosphoethanolamine-*N*-[methoxy-XO4-(polyethylene glycol-3400)] sodium salt (DSPE-PEG₃₄₀₀-XO4), where methoxy-XO4 serves

as targeting and fluorescent moiety. When administered intravenously, nano-conjugate systems showed long circulating time with high efficiency for A β plaques in brain sections of APP/PSEN1 transgenic mice (Tanifum et al., 2012). **Figure 1** shows the injected-targeted liposomes crossing the BBB of APP/PSEN1 mice and labeled parenchymal Ab deposits.

Nasal Route for Enhanced Delivery Across BBB

Many studies have used the nasal route for enhanced delivery of drugs across BBB because increased permeability of the nasal epithelia to drugs due to tight junction opening between apical cells provides a better route for increased drug delivery (Pardeshi and Belgamwar, 2013). It was noted that the brain delivery of drug through the nasal route is affected by the size, surface charge, and hydrophilic-lipophilic balance value of the NPs. The NPs with a size of less than 20 nm are proposed to follow the paracellular transport as compared with NPs of size 100-200 nm, which have been proposed to follow the transcellular route of drug transport. Similarly, particles with a dominant hydrophilic character follow the aqueous paracellular route of transport. Moreover, increase in cationic charge over the chitosan NPs showed increase in muco-adhesion to the nasal mucous and, hence, increased delivery across BBB (Mistry et al., 2009).

This attribute was used by Zhang C. et al. (2014) where they reported the synthesis of Solanum tuberosum lectinmodified polyethylene glycol-polylactide-polyglycolide NPs for enhanced intranasal delivery of basic fibroblast growth factor. The study showed 1.7-fold higher concentration of drug when administered intranasally as compared with intravenously as determined by the area under concentration-time graph of the brain based on the radioiodine isotopic method. Moreover, the neuroprotective effect of the intranasal preparation was higher as compared with the intravenous preparation as determined by the mirror water maze task in AD rats, indicating that the system is a good brain DDR system for peptide- and protein-based drugs. Similarly, in another study, Arumugam et al. (2008) showed that intranasal delivery of rivastigmine-loaded multilamellar liposomes results in higher concentration and longer half-life in the brain of Wister rats in comparison to intranasal (non- liposomal) and oral preparation.

Targeted NPs for Delivery Across BBB

The advent of new strategies such as targeting agents, specific cell receptor ligands, and monoclonal antibodies have shifted the paradigm from non-specific neurotransmitter-based symptomatic treatment toward more personalized and targeted therapeutics that aim to disrupt unique biochemical processes necessary for plaque formation and its development (Bhaskar et al., 2010). Many researchers have explored the possibility of limiting the off-site binding and enhancing drug delivery of NPs across BBB by employing a targeting agent for specific BBB and A β receptors (Ulbrich et al., 2009). These agents include,

but are not limited to apolipoprotein-E receptor-binding peptide and mAbs directed to transferrin and insulin receptors for BBB targeting and anti-amyloid beta antibody against $A\beta_{42}$ for $A\beta$ plaques targeting (Rocha, 2013). Similarly, targeting agents directed toward tau protein and GSH are also under various clinical trials.

In an in vitro study, Canovi et al. (2011) showed that nano-liposomes decorated with an anti-AB monoclonal antibody (Aβ-MAb) have a strong potential as vector for targeting $A\beta_{1-42}$ peptides *in vitro* and plaque deposits in postmortem AD tissue. In addition to antibody-based targeting agents, several types of amyloid-specific cell receptor ligands are employed for targeted delivery of drug carriers across BBB. Gobbi et al. (2010) demonstrated in an in vitro immunestaining study that phosphatidic acid (PA)- and cardiolipin (CL)-decorated liposomes successfully interacted with $A\beta_{1-42}$ aggregates, indicating that these liposomes are a potential vector for targeted delivery of anti-amyloid agents. In another similar study, Markoutsa et al. (2012) reported synthesis of dualtargeting liposomal NPs of two types: anti-Aβ-MAb (Aβ-MAb)decorated immune-liposomes (LIP) and dually decorated LIP (dd-LIP) with OX-26 (directed toward transferrin receptor) and Aβ–MAb using biotin–STREP–biotin method. Both types of NPs showed an increase in binding (and transport) across human brain endothelial hCMEC/D3 cells model of BBB, indicating the potential of the targeting agent in increasing the binding and transport of NPs across BBB. The dd-LIP showed lysosomal uptake and it was proposed that the transport was receptor mediated, but validation of permeability across BBB needs further in vivo-based studies (Markoutsa et al., 2011, 2012). The group also reported, in another study, the synthesis of mono and dually decorated LIP tagged with anti-transferrin mAb alone and in combination with peptide derivative of apolipoprotein E3 (APOE) to target the low-density lipoprotein receptor-related protein (LPR), respectively. The PEG-LIP were taken as control (Markoutsa et al., 2014). In vitro studies in hCMEC/D3 cell monolayer showed additive effect of dual targeting in increasing the delivery across the monolayer, but in vivo studies in FVB mice indicated increased delivery of dual-LIP across BBB in comparison to control, but not to mono-LIP. This variable response in in vitro and in vivo studies was attributed to the presence of serum proteins. It was suggested that use of serum proteins in *in vitro* tests will be a predictive tool for targeting ability.

Moreover, benefits of the double-targeting approach were also developed for bifunctional liposomes tagged with apolipoprotein-E receptor-binding peptide for BBB targeting and with PA for A β binding (Balducci et al., 2014). The study in APP/presenilin 1 transgenic mice showed that the formulation successfully reduced brain A β burden and ameliorated memory impairment.

Diagnostic NPs for AD

As stated earlier, the inability to pinpoint the basic pathological event or maker of AD usually results in delayed diagnosis and treatment (Ayaz et al., 2017c). Many NPs have been checked for the delivery of diagnostic machinery across the BBB. This



from Zhang J. et al., 2014).



should ideally consist of unique fluorescent active or magnetic nanomaterial that can be detected non-invasively, packaged into biocompatible nano-vesicles decorated with a detecting ligand specific to early stage biomarkers of AD (Giljohann and Mirkin, 2009; Azria et al., 2017).

The fact is that radiolabeled $A\beta_{40}$ peptide binds reversibly and specifically to diffuse and neuritic plaques and cerebrovascular amyloid in brain tissue obtained from AD patient. Radiolabeled $A\beta_{40}$ peptide can be imaged *in vivo* non-invasively from the basis for its use in diagnostic and theranostic modalities related to AD. In order to improve its delivery across the BBB, targeting agents such as mAb, fusion proteins, and natural diamines are linked to it (Rocha, 2013). Some of these targeting strategies have been used in combination of NPs.

Yin et al. (2015) synthesized dimercaptosuccinic acid-coated magnetic NPs (MNPs) tagged with anti-amyloid antibodies. They showed that the synthesized NPs were biocompatible and biologically active with the potential of being used for early detection of AD (Yin et al., 2015). Similarly, in another study, Tanifum et al. (2016) reported the synthesis of amyloid fibril-binding ligand ET6-21. The ligand was used to prepare amyloid-targeted liposomes, tagged with Gd chelates and indocyanine green for visualization by magnetic resonance imaging (MRI) and infrared microscopy, respectively. The study showed successful binding of the particles to the plaques after intravenous administration in transgenic 2576 and APP (mutant APP) mouse models of AD. The study indicated the potential for development of MRI imaging agents for detection of amyloid plaques. Similarly, another group synthesized curcuminloaded super paramagnetic iron oxide NPs of <100 nm for targeting of amyloid plaques (Cheng et al., 2015). The synthesized NPs were successful in differentiating the transgenic 2576 mouse brains having Aβ plaques from non-transgenic mouse models, indicating their potential as a diagnostic agent in AD.

Moreover, recent studies showed that A β oligomer instigate the early stage of memory loss and thus, is a better candidate as a biomarker of early-stage identification of AD. Viola et al. (2015) synthesized A β oligomer-targeted molecular MRI probes by attaching A β Os-specific monoclonal antibody (NU4) to the nitro-dopamine and polyethylene glycol stabilized 12–16 nm magnetite NPs (Viola et al., 2015). The study showed significant clinical potential of the A β O-MRI probe for identification of synapto-toxic oligomers in 5xFAD mouse model following intranasal delivery.

Theranostic NPs for Delivery Across BBB

Many preclinical studies have established the role of NPs as carriers and use of specific surface ligands for targeting the NP for AD separately. This leads to the development of multimodal preparations having characteristics of drug carriers tagged with targeting moieties and diagnostic agents. These magical NPs with all-in-one characteristics are the next-generation particles termed as "theranostic agent."

Papadia et al. (2017) synthesized multifunctional liposomal NPs decorated with curcumin-lipid ligand (TREG) with affinity toward amyloid species along with ligands to target the transferrin and the LDL receptors of the BBB. The study showed successful targeting of brain and inhibition of amyloid peptide aggregation (Papadia et al., 2017). Similarly, Rotman et al. (2015) have also developed two liposomal formulations based on 1,2-dimyristoyl-sn-glycero-3-phosphocholine and egg-yolk PC encapsulating indium-111-labeled A β -binding llama single-domain antibody fragments (VHH-pa2H) and were coated with glutathione–PEG. The animal study in APP_{swe}/PS1dE9 double transgenic mice showed the highest standard uptake in brain with GSH–PEG EYPC, indicating the theranostic potential of the prepared formulation (Rotman et al., 2015).

Green-Synthesized NPs as Future AD Theranostic Agents

Recent developments at the interface of green chemistry and nanotechnology indicate significant potential in the biomedical sciences from theranostic perspectives (Ovais et al., 2016, 2017b, 2018a,d; Barabadi et al., 2017b; Khalil et al., 2017a). However, their potential is largely untapped in case of the neurodegenerative diseases like Alzheimer's. The use of other synthesis methods (physical and chemical) is often discouraged as they possess certain inherent disadvantages such as toxicity and cost (Khalil et al., 2017b; Ovais et al., 2017a). Moreover, in a recent work, it was reported that some chemicals used during the synthesis of NPs through a chemical route can remain attached on the surface of the NPs and, therefore, could not be used for biomedical application. Therefore, tremendous interest has been growing over the last decade to fabricate materials of interest using a green process (Emmanuel et al., 2017). Usually, the green chemistry-based approach includes the use of medicinal plants or pure phytochemicals that have potential medicinal value. These phytochemicals are involved in the chelating as well as stabilizing of NPs (Barabadi et al., 2017a; Ovais et al., 2017b, 2018c; Khalil et al., 2018).

Recently, Suganthy et al. (2017) reported the neuroprotective nature of biogenic gold NPs synthesized through Terminalia arjuna. Their results indicated the significant neuroprotective nature and biocompatibility of biogenic gold NPs. The biogenic gold NPs successfully inhibited the ChE enzymes, reduced the Aß fibrillation process, and also destabilized the mature fibrils at very low concentrations. The functionalization of chemically synthesized gold NPs with trehalose has significantly improved the inhibition of protein aggregation as well as the mature fibril disintegration and possesses the potential to be used in photothermal therapies (Mandal et al., 2017). Functionalization of bioinspired gold NPs with such antiamyloidogenic molecules can be a fruitful strategy for further enhancing their neuroprotective nature. Another research contribution directed toward the neuroprotective behavior included the biogenic platinum NPs biosynthesized through Bacopa monnieri. The authors concluded that the neuroprotective potential of biogenic platinum nanoparticles is mediated via inhibition of free radicals generation and antioxidant effects (Nellore et al., 2013). Plant phytochemical-based functionalization of NPs has yielded amazing results. Recently, polyphenol-based functionalization of selenium NPs was achieved. In the research, nanoscaled

selenium was coated with EGCG; a polyphenol found in the tea. The EGCG is known for its neuroprotective potential and possesses the ability to inhibit various amyloid-forming proteins such as amyloid beta, transthyretin, α -synuclein, and huntingtin, which are involved in the progression of AD. The EGCG-stabilized Se NPs were further coated with Tet-1 proteins, having strong neuronal affinity. The Tet1-Se@EGCH nanosystem induced effective inhibition of the fibrillation process of AB and also disintegrated the already-matured fibrils as shown in Figure 2. In addition, the indicated nanosystem was found to inhibit the DNA fragmentation and ROS generation while found to be effective at very low concentrations (Zhang J. et al., 2014). There is potential to replace the EGCG with other plant-based chemical entities. Curcumin or curcuminbased derivatives have yielded impressive properties to halt the disease progression of Alzheimer's. Curcumin in conjunction with the benzothiazolinone possesses strong affinity for binding to amyloid and, hence, can be used to deliver curcumin or other natural products with anti-Alzheimer potential as summarized in Figure 3 (Megill et al., 2013; Fang et al., 2014).

CONCLUSION AND FUTURE PROSPECTS

Recent research on phyto and nanomedicines represents a potential area of interest in the treatment of AD.

REFERENCES

- Aalinkeel, R., Kutscher, H. L., Singh, A., Cwiklinski, K., Khechen, N., Schwartz, S. A., et al. (2018). Neuroprotective effects of a biodegradable poly (lactic-coglycolic acid)-ginsenoside Rg3 nanoformulation: a potential nanotherapy for Alzheimer's disease? *J. Drug Target.* 26, 182–193. doi: 10.1080/1061186X.2017. 1354002
- Abbott, N. J., Patabendige, A. A., Dolman, D. E., Yusof, S. R., and Begley, D. J. (2010). Structure and function of the blood-brain barrier. *Neurobiol. Dis.* 37, 13–25. doi: 10.1016/j.nbd.2009.07.030
- Agarwa, P., Sharma, B., Fatima, A., and Jain, S. K. (2014). An update on Ayurvedic herb Convolvulus pluricaulis Choisy. Asian Pac. J. Trop. Biomed. 4, 245–252. doi: 10.1016/S2221-1691(14)60240-9
- Aggarwal, B. B., and Sung, B. (2009). Pharmacological basis for the role of curcumin in chronic diseases: an age-old spice with modern targets. *Trends Pharmacol. Sci.* 30, 85–94. doi: 10.1016/j.tips.2008.11.002
- Agnihotri, S. A., Mallikarjuna, N. N., and Aminabhavi, T. M. (2004). Recent advances on chitosan-based micro-and nanoparticles in drug delivery. *J. Control. Release* 100, 5–28. doi: 10.1016/j.jconrel.2004.08.010
- Aguiar, S., and Borowski, T. (2013). Neuropharmacological review of the nootropic herb *Bacopa monnieri*. *Rejuvenation Res.* 16, 313–326. doi: 10.1089/rej.2013. 1431
- Agyare, E. K., Curran, G. L., Ramakrishnan, M., Caroline, C. Y., Poduslo, J. F., and Kandimalla, K. K. (2008). Development of a smart nano-vehicle to target cerebrovascular amyloid deposits and brain parenchymal plaques observed in Alzheimer's disease and cerebral amyloid angiopathy. *Pharm. Res.* 25, 2674– 2684. doi: 10.1007/s11095-008-9688-y
- Ahlemeyer, B., and Krieglstein, J. (2003). Pharmacological studies supporting the therapeutic use of *Ginkgo biloba* extract for Alzheimer's disease. *Pharmacopsychiatry* 36, 8–14. doi: 10.1055/s-2003-40454
- Ahmad, S., Ullah, F., Sadiq, A., Ayaz, M., Imran, M., Ali, I., et al. (2016). Chemical composition, antioxidant and anticholinesterase potentials of

Numerous phytochemicals have proven promising results in AD therapy, while with the emerging feat of tailored nanomedicines, the effectiveness can be increased several folds by efficient delivery of the active ingredients to the target sites. Moreover, a relatively untapped area of research at the interface of medicinal plants and nanotechnology is the biogenic synthesis of metal NPs for AD treatment.

The occurrence of metal and metal-based NPs in the brain can be toxic as well as beneficial. The NPs like TiO₂, ZnO, Ag, and Au NPs etc. can induce oxidative damage in the brain. However, selenium NPs are reported to have an antioxidant role in the brain. Largely, these mentioned NPs were synthesized through chemistry-based methods. The green chemistry-based methods can alter the properties of these NPs. The science of biogenic NPs is still in the nascent phase, and theranostic NPs of variable and interesting features can be produced to alleviate the menace of AD.

AUTHOR CONTRIBUTIONS

MO, NZ, and IA conceived the idea and drafted the manuscript. AK, AR, MA, AS, FU, and ZS contributed in manuscript drafting, language correction, and technical corrections. All the authors read and approved the manuscript.

essential oil of *Rumex hastatus* D. Don collected from the North West of Pakistan. *BMC Complement. Alternat. Med.* 16:29. doi: 10.1186/s12906-016-0998-z

- Akhondzadeh, S., Sabet, M. S., Harirchian, M. H., Togha, M., Cheraghmakani, H., Razeghi, S., et al. (2010). A 22-week, multicenter, randomized, doubleblind controlled trial of *Crocus sativus* in the treatment of mild-to-moderate Alzheimer's disease. *Psychopharmacology* 207, 637–643. doi: 10.1007/s00213-009-1706-1
- Akram, M., and Nawaz, A. (2017). Effects of medicinal plants on Alzheimer's disease and memory deficits. *Neural Regen. Res.* 12, 660–670. doi: 10.4103/1673-5374.205108
- Ali, M., Khan, T., Fatima, K., Ali, Q. U. A., Ovais, M., Khalil, A. T., et al. (2018). Selected hepatoprotective herbal medicines: evidence from ethnomedicinal applications, animal models, and possible mechanism of actions. *Phytother. Res.* 32, 199–215. doi: 10.1002/ptr.5957
- Ali, M., Muhammad, S., Shah, M. R., Khan, A., Rashid, U., Farooq, U., et al. (2017). Neurologically potent molecules from *Crataegus oxyacantha*; isolation, anticholinesterase inhibition, and molecular docking. *Front. Pharmacol.* 8:327. doi: 10.3389/fphar.2017.00327
- Ali, Y. O., Bradley, G., and Lu, H.-C. (2017). Screening with an NMNAT2-MSD platform identifies small molecules that modulate NMNAT2 levels in cortical neurons. *Sci. Rep.* 7:43846. doi: 10.1038/srep4 3846
- Alzheimer's and Association (2017). 2017 Alzheimer's disease facts and figures. Alzheimers Dement. 13, 325–373. doi: 10.1016/j.jalz.2017. 02.001
- Anwar, A., Ovais, M., Khan, A., and Raza, A. (2017). Docetaxel-loaded solid lipid nanoparticles: a novel drug delivery system. *IET Nanobiotechnol.* 11, 621–629. doi: 10.1049/iet-nbt.2017.0001
- Arumugam, K., Subramanian, G., Mallayasamy, S., Averineni, R., Reddy, M., and Udupa, N. (2008). A study of rivastigmine liposomes for delivery into the brain through intranasal route. *Acta Pharm.* 58, 287–297. doi: 10.2478/v10007-008-0014-3

- Asgarpanah, J., and Kazemivash, N. (2012). Phytochemistry and pharmacologic properties of *Myristica fragrans* hoyutt: a review. *Afr. J. Biotechnol.* 11, 12787– 12793. doi: 10.5897/AJB12.1043
- Ayaz, M., Junaid, M., Ahmed, J., Ullah, F., Sadiq, A., Ahmad, S., et al. (2014). Phenolic contents, antioxidant and anticholinesterase potentials of crude extract, subsequent fractions and crude saponins from *Polygonum hydropiper L. BMC Complement. Altern. Med.* 14:145. doi: 10.1186/1472-6882-14-145
- Ayaz, M., Junaid, M., Ullah, F., Sadiq, A., Khan, M. A., Ahmad, W., et al. (2015). Comparative chemical profiling, cholinesterase inhibitions and antiradicals properties of essential oils from *Polygonum hydropiper* L: a preliminary anti-Alzheimer's study. *Lipids Health Dis.* 14:141. doi: 10.1186/s12944-015-0145-8
- Ayaz, M., Junaid, M., Ullah, F., Sadiq, A., Ovais, M., Ahmad, W., et al. (2016). Chemical profiling, antimicrobial and insecticidal evaluations of *Polygonum hydropiper L. BMC Complement. Altern. Med.* 16:502. doi: 10.1186/s12906-016-1491-4
- Ayaz, M., Junaid, M., Ullah, F., Sadiq, A., Shahid, M., Ahmad, W., et al. (2017a). GC-MS analysis and gastroprotective evaluations of crude extracts, isolated saponins, and essential oil from *Polygonum hydropiper L. Front. Chem.* 5:58. doi: 10.3389/fchem.2017.00058
- Ayaz, M., Junaid, M., Ullah, F., Subhan, F., Sadiq, A., Ali, G., et al. (2017b). Anti-Alzheimer's studies on β-sitosterol isolated from *Polygonum hydropiper* L. *Front. Pharmacol.* 8:697. doi: 10.3389/fphar.2017.00697
- Ayaz, M., Sadiq, A., Junaid, M., Ullah, F., Subhan, F., and Ahmed, J. (2017c). Neuroprotective and anti-aging potentials of essential oils from aromatic and medicinal plants. *Front. Aging Neurosci.* 9:168. doi: 10.3389/fnagi.2017. 00168
- Ayaz, M., Subhan, F., Sadiq, A., Ullah, F., Ahmed, J., and Sewell, R. (2017d). Cellular efflux transporters and the potential role of natural products in combating efflux mediated drug resistance. *Front. Biosci.* 22, 732–756. doi: 10.2741/4513
- Azria, D., Blanquer, S., Verdier, J.-M., and Belamie, E. (2017). Nanoparticles as contrast agents for brain nuclear magnetic resonance imaging in Alzheimer's disease diagnosis. *J. Mater. Chem. B* 5, 7216–7237. doi: 10.1039/C7TB0 1599B
- Balducci, C., Mancini, S., Minniti, S., La Vitola, P., Zotti, M., Sancini, G., et al. (2014). Multifunctional liposomes reduce brain β-amyloid burden and ameliorate memory impairment in Alzheimer's disease mouse models. J. Neurosci. 34, 14022–14031. doi: 10.1523/JNEUROSCI.0284-14. 2014
- Barabadi, H., Alizadeh, A., Ovais, M., Ahmadi, A., Shinwari, Z. K., and Saravanan, M. (2017a). Efficacy of green nanoparticles against cancerous and normal cell lines: a systematic review and meta-analysis. *IET Nanobiotechnol.* 12, 377–391. doi: 10.1049/iet-nbt.2017.0120
- Barabadi, H., Ovais, M., Shinwari, Z. K., and Saravanan, M. (2017b). Anti-cancer green bionanomaterials: present status and future prospects. *Green Chem. Lett. Rev.* 10, 285–314. doi: 10.1080/17518253.2017.1385856
- Baum, L., Lam, C. W. K., Cheung, S. K.-K., Kwok, T., Lui, V., Tsoh, J., et al. (2008). Six-month randomized, placebo-controlled, double-blind, pilot clinical trial of curcumin in patients with Alzheimer disease. J. Clin. Psychopharmacol. 28, 110–113. doi: 10.1097/jcp.0b013e318160862c
- Baum, L., and Ng, A. (2004). Curcumin interaction with copper and iron suggests one possible mechanism of action in Alzheimer's disease animal models. *J. Alzheimers Dis.* 6, 367–377. doi: 10.3233/JAD-2004-6403
- Beason-Held, L. L., Goh, J. O., An, Y., Kraut, M. A., O'Brien, R. J., Ferrucci, L., et al. (2013). Changes in brain function occur years before the onset of cognitive impairment. *J. Neurosci.* 33, 18008–18014. doi: 10.1523/JNEUROSCI.1402-13. 2013
- Begley, D. J. (2004). ABC transporters and the blood-brain barrier. Curr. Pharm. Des. 10, 1295–1312. doi: 10.2174/1381612043384844
- Begum, A. N., Jones, M. R., Lim, G. P., Morihara, T., Kim, P., Heath, D. D., et al. (2008). Curcumin structure-function, bioavailability, and efficacy in models of neuroinflammation and Alzheimer's disease. *J. Pharmacol. Exp. Ther.* 326, 196–208. doi: 10.1124/jpet.108.137455
- Bhanumathy, M., Harish, M., Shivaprasad, H., and Sushma, G. (2010). Nootropic activity of *Celastrus paniculatus* seed. *Pharm. Biol.* 48, 324–327. doi: 10.3109/ 13880200903127391

- Bhaskar, S., Tian, F., Stoeger, T., Kreyling, W., de la Fuente, J. M., Grazú, V., et al. (2010). Multifunctional nanocarriers for diagnostics, drug delivery and targeted treatment across blood-brain barrier: perspectives on tracking and neuroimaging. *Part. Fibre Toxicol.* 7:3. doi: 10.1186/1743-8977-7-3
- Bihaqi, S. W., Singh, A. P., and Tiwari, M. (2012). Supplementation of *Convolvulus pluricaulis* attenuates scopolamine-induced increased tau and Amyloid precursor protein (AβPP) expression in rat brain. *Indian J. Pharmacol.* 44, 593–598. doi: 10.4103/0253-7613.100383
- Bird, T. D. (2008). Genetic aspects of Alzheimer disease. *Genet. Med.* 10, 231–239. doi: 10.1097/GIM.0b013e31816b64dc
- Biswas, K., Islam, A., Sharmin, T., and Biswas, P. K. (2015). In-vitro cholinesterase inhibitory activity of dry fruit extract of *Phyllanthus emblica* relevant to the treatment of Alzheimer's disease. J. Phytopharmacol. 4, 5–8.
- Blusztajn, J. K., Slack, B. E., and Mellott, T. J. (2017). Neuroprotective actions of dietary choline. *Nutrients* 9:E815. doi: 10.3390/nu9080815
- Bohrmann, B., Baumann, K., Benz, J., Gerber, F., Huber, W., Knoflach, F., et al. (2012). Gantenerumab: a novel human anti-Aβ antibody demonstrates sustained cerebral amyloid-β binding and elicits cell-mediated removal of human amyloid-β. J. Alzheimers Dis. 28, 49–69. doi: 10.3233/JAD-2011-11 0977
- Canovi, M., Markoutsa, E., Lazar, A. N., Pampalakis, G., Clemente, C., Re, F., et al. (2011). The binding affinity of anti-Aβ1-42 MAb-decorated nanoliposomes to Aβ1-42 peptides in vitro and to amyloid deposits in postmortem tissue. *Biomaterials* 32, 5489–5497. doi: 10.1016/j.biomaterials.2011. 04.020
- Carradori, D., Balducci, C., Re, F., Brambilla, D., Le Droumaguet, B., Flores, O., et al. (2018). Antibody-functionalized polymer nanoparticle leading to memory recovery in Alzheimer's disease-like transgenic mouse model. *Nanomedicine* 14, 609–618. doi: 10.1016/j.nano.2017.12.006
- Casey, D. A., Antimisiaris, D., and O'Brien, J. (2010). Drugs for Alzheimer's disease: are they effective? *Pharm. Ther.* 35, 208–211.
- Chaudhari, K. S., Tiwari, N. R., Tiwari, R. R., and Sharma, R. S. (2017). Neurocognitive effect of nootropic drug brahmi (*Bacopa monnieri*) in Alzheimer's disease. *Ann. Neurosci.* 24, 111–122. doi: 10.1159/00047 5900
- Chaudhary, B., and Bist, R. (2017). Protective manifestation of bacoside A and bromelain in terms of cholinesterases, gamma-amino butyric acid, serotonin level and stress proteins in the brain of dichlorvos-intoxicated mice. *Cell Stress Chaperones* 22, 371–376. doi: 10.1007/s12192-017-0773-1
- Chen, W., Guo, Y., Yang, W., Zheng, P., Zeng, J., and Tong, W. (2015). Protective effect of ginsenoside Rb1 on integrity of blood-brain barrier following cerebral ischemia. *Exp. Brain Res.* 233, 2823–2831. doi: 10.1007/s00221-015-4352-3
- Chen, Z., Mao, X., Liu, A., Gao, X., Chen, X., Ye, M., et al. (2015). Osthole, a natural coumarin improves cognitive impairments and BBB dysfunction after transient global brain ischemia in C57 BL/6J mice: involvement of Nrf2 pathway. *Neurochem. Res.* 40, 186–194. doi: 10.1007/s11064-014-1483-z
- Chen, X., Hua, H.-P., Liang, L., and Liu, L.-J. (2017). The oral pretreatment of glycyrrhizin prevents surgery-induced cognitive impairment in aged mice by reducing neuroinflammation and Alzheimer's-related pathology via HMGB1 inhibition. J. Mol. Neurosci. 63, 385–395. doi: 10.1007/s12031-017-0989-7
- Cheng, K. K., Chan, P. S., Fan, S., Kwan, S. M., Yeung, K. L., Wang, Y.-X. J., et al. (2015). Curcumin-conjugated magnetic nanoparticles for detecting amyloid plaques in Alzheimer's disease mice using magnetic resonance imaging (MRI). *Biomaterials* 44, 155–172. doi: 10.1016/j.biomaterials.2014. 12.005
- Cheng, K. K., Yeung, C. F., Ho, S. W., Chow, S. F., Chow, A. H., and Baum, L. (2013). Highly stabilized curcumin nanoparticles tested in an in vitro blood-brain barrier model and in Alzheimer's disease Tg2576 mice. AAPS J. 15, 324–336. doi: 10.1208/s12248-012-9444-4
- Chu, Y.-F., Brown, P. H., Lyle, B. J., Chen, Y., Black, R. M., Williams, C. E., et al. (2009). Roasted coffees high in lipophilic antioxidants and chlorogenic acid lactones are more neuroprotective than green coffees. J. Agric. Food Chem. 57, 9801–9808. doi: 10.1021/jf902095z

- Colín-González, A. L., Santana, R. A., Silva-Islas, C. A., Chánez-Cárdenas, M. E., Santamaría, A., and Maldonado, P. D. (2012). The antioxidant mechanisms underlying the aged garlic extract-and S-allylcysteine-induced protection. Oxid. Med. Cell. Longev. 2012:907162. doi: 10.1155/2012/90 7162
- Cropley, V., Croft, R., Silber, B., Neale, C., Scholey, A., Stough, C., et al. (2012). Does coffee enriched with chlorogenic acids improve mood and cognition after acute administration in healthy elderly? A pilot study. *Psychopharmacology* 219, 737–749. doi: 10.1007/s00213-011-2395-0
- Cui, Z., Lockman, P. R., Atwood, C. S., Hsu, C.-H., Gupte, A., Allen, D. D., et al. (2005). Novel D-penicillamine carrying nanoparticles for metal chelation therapy in Alzheimer's and other CNS diseases. *Eur. J. Pharm. Biopharm.* 59, 263–272. doi: 10.1016/j.ejpb.2004.07.009
- Cummings, J., Morstorf, T., and Lee, G. (2016). Alzheimer's drug-development pipeline: 2016. *Alzheimers Dement. (N.Y.)* 2, 222-232. doi: 10.1016/j.trci.2016. 07.001
- Daneman, R., and Prat, A. (2015). The blood-brain barrier. Cold Spring Harb. Perspect. Biol. 7:a020412. doi: 10.1101/cshperspect.a020412
- Danhier, F., Ansorena, E., Silva, J. M., Coco, R., Le Breton, A., and Préat, V. (2012). PLGA-based nanoparticles: an overview of biomedical applications. J. Control. Release 161, 505–522. doi: 10.1016/j.jconrel.2012. 01.043
- De Campos, A. M., Sánchez, A., and Alonso, M. J. (2001). Chitosan nanoparticles: a new vehicle for the improvement of the delivery of drugs to the ocular surface. Application to cyclosporin A. *Int. J. Pharm.* 224, 159–168. doi: 10.1016/S0378-5173(01)00760-8
- DeFeudis, F. V. (2002). Effects of *Ginkgo biloba* extract (EGb 761) on gene expression: possible relevance to neurological disorders and age-associated cognitive impairment. *Drug Dev. Res.* 57, 214–235. doi: 10.1002/ddr. 10151
- Dhanasekaran, M., Holcomb, L. A., Hitt, A. R., Tharakan, B., Porter, J. W., Young, K. A., et al. (2009). *Centella asiatica* extract selectively decreases amyloid β levels in hippocampus of Alzheimer's disease animal model. *Phytother. Res.* 23, 14–19. doi: 10.1002/ptr.2405
- Di Stefano, A., Iannitelli, A., Laserra, S., and Sozio, P. (2011). Drug delivery strategies for Alzheimer's disease treatment. *Expert Opin. Drug Deliv.* 8, 581–603. doi: 10.1517/17425247.2011.561311
- Dubey, G., Pathak, S., and Gupta, B. (1994). Combined effect of Brahmi (*Bacopa monniera*) and Shankhpushpi (*Convolvulus pluricaulis*) on cognitive functions. *Pharmacopsychoecol* 7, 249–251.
- Dutta, P. K., Dutta, J., and Tripathi, V. (2004). Chitin and chitosan: chemistry, properties and applications. J. Sci. Ind. Res. 63, 20-31.
- Emmanuel, R., Saravanan, M., Ovais, M., Padmavathy, S., Shinwari, Z. K., and Prakash, P. (2017). Antimicrobial efficacy of drug blended biosynthesized colloidal gold nanoparticles from *Justicia glauca* against oral pathogens: a nanoantibiotic approach. *Microb. Pathog.* 113, 295–302. doi: 10.1016/j.micpath. 2017.10.055
- Erickson, M. A., and Banks, W. A. (2013). Blood-brain barrier dysfunction as a cause and consequence of Alzheimer's disease. J. Cereb. Blood Flow Metab. 33, 1500–1513. doi: 10.1038/jcbfm.2013.135
- Esquivel, P., and Jiménez, V. M. (2012). Functional properties of coffee and coffee by-products. *Food Res. Int.* 46, 488–495. doi: 10.1016/j.foodres.2011.05.028
- Fang, L., Gou, S., Liu, X., Cao, F., and Cheng, L. (2014). Design, synthesis and anti-Alzheimer properties of dimethylaminomethyl-substituted curcumin derivatives. *Bioorg. Med. Chem. Lett.* 24, 40–43. doi: 10.1016/j.bmcl.2013. 12.011
- Fang, W., Deng, Y., Li, Y., Shang, E., Fang, F., Lv, P., et al. (2010). Blood brain barrier permeability and therapeutic time window of Ginkgolide B in ischemiareperfusion injury. *Eur. J. Pharm. Sci.* 39, 8–14. doi: 10.1016/j.ejps.2009. 10.002
- Faria, A., Meireles, M., Fernandes, I., Santos-Buelga, C., Gonzalez-Manzano, S., Dueñas, M., et al. (2014). Flavonoid metabolites transport across a human BBB model. *Food Chem.* 149, 190–196. doi: 10.1016/j.foodchem.2013. 10.095
- Faria, A., Pestana, D., Teixeira, D., Couraud, P.-O., Romero, I., Weksler, B., et al. (2011). Insights into the putative catechin and epicatechin transport across blood-brain barrier. *Food Funct.* 2, 39–44. doi: 10.1039/C0FO00100G

- Farlow, M. R., Andreasen, N., Riviere, M.-E., Vostiar, I., Vitaliti, A., Sovago, J., et al. (2015). Long-term treatment with active Aβ immunotherapy with CAD106 in mild Alzheimer's disease. *Alzheimers Res. Ther.* 7:23. doi: 10.1186/s13195-015-0108-3
- Farokhnia, M., Shafiee Sabet, M., Iranpour, N., Gougol, A., Yekehtaz, H., Alimardani, R., et al. (2014). Comparing the efficacy and safety of *Crocus sativus*L. with memantine in patients with moderate to severe Alzheimer's disease: a double-blind randomized clinical trial. *Hum. Psychopharmacol.* 29, 351–359. doi: 10.1002/hup.2412
- Fazil, M., Md, S., Haque, S., Kumar, M., Baboota, S., Sahni, J. K., et al. (2012). Development and evaluation of rivastigmine loaded chitosan nanoparticles for brain targeting. *Eur. J. Pharm. Sci.* 47, 6–15. doi: 10.1016/j.ejps.2012. 04.013
- Finley, J. W., and Gao, S. (2017). A perspective on *Crocus sativus* L.(Saffron) constituent crocin: a potent water-soluble antioxidant and potential therapy for Alzheimer's disease. *J. Agric. Food Chem.* 65, 1005–1020. doi: 10.1021/acs.jafc. 6b04398
- Fonseca-Santos, B., Gremião, M. P. D., and Chorilli, M. (2015). Nanotechnologybased drug delivery systems for the treatment of Alzheimer's disease. *Int. J. Nanomedicine* 10, 4981–5003. doi: 10.2147/IJN.S87148
- França, M. B., Lima, K. C., and Eleutherio, E. C. (2017). Oxidative stress and amyloid toxicity: insights from yeast. J. Cell. Biochem. 118, 1442–1452. doi: 10.1002/jcb.25803
- Galimberti, D., and Scarpini, E. (2010). Treatment of Alzheimer's disease: symptomatic and disease-modifying approaches. *Curr. Aging Sci.* 3, 46–56. doi: 10.2174/1874609811003010046
- Garrido-Maraver, J., Cordero, M. D., Oropesa-Ávila, M., Vega, A. F., De La Mata, M., Pavón, A. D., et al. (2014). Coenzyme q10 therapy. *Mol. Syndromol.* 5, 187–197. doi: 10.1159/000360101
- Geiser, R. J., Chastain, S. E., and Moss, M. A. (2017). Regulation of Bace1 Mrna expression in Alzheimer'S disease by green tea catechins and black tea theaflavins. *Biophys. J.* 112:362a. doi: 10.1016/j.bpj.2016.11. 1965
- Gharibi, A., Khalili, M., Kiasalari, Z., and Hoseinirad, M. (2013). The effect of *Zingiber officinalis* L. on learning and memory in rats. *J. Bas. Clin. Pathophysiol.* 2, 2013–2014.
- Ghosh, A., Sarkar, S., Mandal, A. K., and Das, N. (2013). Neuroprotective role of nanoencapsulated quercetin in combating ischemia-reperfusion induced neuronal damage in young and aged rats. *PLoS One* 8:e57735. doi: 10.1371/ journal.pone.0057735
- Giacobini, E. (2004). Cholinesterase inhibitors: new roles and therapeutic alternatives. *Pharmacol. Res.* 50, 433–440. doi: 10.1016/j.phrs.2003.11.017
- Giljohann, D. A., and Mirkin, C. A. (2009). Drivers of biodiagnostic development. *Nature* 462, 461–464. doi: 10.1038/nature08605
- Gobbi, M., Re, F., Canovi, M., Beeg, M., Gregori, M., Sesana, S., et al. (2010). Lipid-based nanoparticles with high binding affinity for amyloid- β 1–42 peptide. *Biomaterials* 31, 6519–6529. doi: 10.1016/j.biomaterials.2010. 04.044
- Godkar, P., Gordon, R., Ravindran, A., and Doctor, B. (2006). Celastrus paniculatus seed oil and organic extracts attenuate hydrogen peroxide-and glutamateinduced injury in embryonic rat forebrain neuronal cells. *Phytomedicine* 13, 29–36. doi: 10.1016/j.phymed.2003.11.011
- Grzybowska, B., Słoczyński, J., Grabowski, R., Wcisło, K., Kozłowska, A., Stoch, J., et al. (1998). Chromium oxide/alumina catalysts in oxidative dehydrogenation of isobutane. J. Catal. 178, 687–700. doi: 10.1006/jcat.1998. 2203
- Hebert, L. E., Weuve, J., Scherr, P. A., and Evans, D. A. (2013). Alzheimer disease in the United States (2010–2050) estimated using the 2010 census. *Neurology* 80, 1778–1783. doi: 10.1212/WNL.0b013e31828726f5
- Heneka, M. T., Carson, M. J., El Khoury, J., Landreth, G. E., Brosseron, F., Feinstein, D. L., et al. (2015). Neuroinflammation in Alzheimer's disease. *Lancet Neurol.* 14, 388–405. doi: 10.1016/S1474-4422(15)70016-5
- Ho, Y.-S., Hung, C. H.-L., and Chang, R. C.-C. (2013). Green tea and neurodegeneration in Alzheimer's disease, in *Tea in Health and Disease Prevention*, ed. V. R. Preedy (New York, NY: Elsevier), 691–704.
- Hoi, C. P., Ho, Y. P., Baum, L., and Chow, A. H. (2010). Neuroprotective effect of honokiol and magnolol, compounds from *Magnolia officinalis*, on

beta-amyloid-induced toxicity in PC12 cells. *Phytother. Res.* 24, 1538–1542. doi: 10.1002/ptr.3178

- Hu, Z., Bian, X., Liu, X., Zhu, Y., Zhang, X., Chen, S., et al. (2013). Honokiol protects brain against ischemia-reperfusion injury in rats through disrupting PSD95–nNOS interaction. *Brain Res.* 1491, 204–212. doi: 10.1016/j.brainres. 2012.11.004
- Huang, M., Xie, S.-S., Jiang, N., Lan, J.-S., Kong, L.-Y., and Wang, X.-B. (2015). Multifunctional coumarin derivatives: monoamine oxidase B (MAO-B) inhibition, anti-β-amyloid (Aβ) aggregation and metal chelation properties against Alzheimer's disease. *Bioorg. Med. Chem. Lett.* 25, 508–513. doi: 10.1016/ j.bmcl.2014.12.034
- Ishisaka, A., Ichikawa, S., Sakakibara, H., Piskula, M. K., Nakamura, T., Kato, Y., et al. (2011). Accumulation of orally administered quercetin in brain tissue and its antioxidative effects in rats. *Free Radic. Biol. Med.* 51, 1329–1336. doi: 10.1016/j.freeradbiomed.2011.06.017
- Ismail, M. F., ElMeshad, A. N., and Salem, N. A.-H. (2013). Potential therapeutic effect of nanobased formulation of rivastigmine on rat model of Alzheimer's disease. *Int. J. Nanomedicine* 8, 393–406. doi: 10.2147/IJN.S3 9232
- Ito, H., Sun, X.-L., Watanabe, M., Okamoto, M., and Hatano, T. (2008). Chlorogenic acid and its metabolite m-coumaric acid evoke neurite outgrowth in hippocampal neuronal cells. *Biosci. Biotechnol. Biochem.* 72, 885–888. doi: 10.1271/bbb.70670
- Jalali, S., Zarrinhaghighi, A., Sadraei, S., Ghasemi, Y., Sakhteman, A., and Faridi, P. (2018). A system pharmacology study for deciphering anti depression activity of *Nardostachys jatamansi. Curr. Drug Metab.* 19, 469–476. doi: 10.2174/ 1389200219666180305151011
- Jameel, E., Umar, T., Kumar, J., and Hoda, N. (2016). Coumarin: a privileged scaffold for the design and development of antineurodegenerative agents. *Chem. Biol. Drug Des.* 87, 21–38. doi: 10.1111/cbdd. 12629
- James, S. A., Churches, Q. I., de Jonge, M. D., Birchall, I. E., Streltsov, V., McColl, G., et al. (2016). Iron, copper, and zinc concentration in Aβ plaques in the APP/PS1 mouse model of Alzheimer's disease correlates with metal levels in the surrounding neuropil. ACS Chem. Neurosci. 8, 629–637. doi: 10.1021/ acschemneuro.6b00362
- Jayaprakasam, B., Padmanabhan, K., and Nair, M. G. (2010). Withanamides in Withania somnifera fruit protect PC-12 cells from β -amyloid responsible for Alzheimer's disease. *Phytother. Res.* 24, 859–863.
- Jiang, J., Wang, W., Sun, Y. J., Hu, M., Li, F., and Zhu, D. Y. (2007). Neuroprotective effect of curcumin on focal cerebral ischemic rats by preventing blood-brain barrier damage. *Eur. J. Pharmacol.* 561, 54–62. doi: 10.1016/j.ejphar.2006. 12.028
- Johansson, B. B. (1990). The physiology of the blood-brain barrier, in *Circulating Regulatory Factors and Neuroendocrine Function*, eds J. C. Porter and D. Ježová (Boston, MA: Springer), 25–39. doi: 10.1007/978-1-4684-5799-5_2
- Jose, S., Sowmya, S., Cinu, T., Aleykutty, N., Thomas, S., and Souto, E. (2014). Surface modified PLGA nanoparticles for brain targeting of Bacoside-A. *Eur. J. Pharm. Sci.* 63, 29–35. doi: 10.1016/j.ejps.2014. 06.024
- Joubert, J., Foka, G. B., Repsold, B. P., Oliver, D. W., Kapp, E., and Malan, S. F. (2017). Synthesis and evaluation of 7-substituted coumarin derivatives as multimodal monoamine oxidase-B and cholinesterase inhibitors for the treatment of Alzheimer's disease. *Eur. J. Med. Chem.* 125, 853–864. doi: 10.1016/ j.ejmech.2016.09.041
- Karami, M., Bathaie, S. Z., Tiraihi, T., Habibi-Rezaei, M., Arabkheradmand, J., and Faghihzadeh, S. (2013). Crocin improved locomotor function and mechanical behavior in the rat model of contused spinal cord injury through decreasing calcitonin gene related peptide (CGRP). *Phytomedicine* 21, 62–67. doi: 10.1016/ j.phymed.2013.07.013
- Karkada, G., Shenoy, K., Halahalli, H., and Karanth, K. (2012). Nardostachys jatamansi extract prevents chronic restraint stress-induced learning and memory deficits in a radial arm maze task. J. Nat. Sci. Biol. Med. 3, 125–132. doi: 10.4103/0976-9668.101879
- Kaur, N., Dhiman, M., Perez-Polo, J. R., and Mantha, A. K. (2015). Ginkgolide B revamps neuroprotective role of apurinic/apyrimidinic endonuclease 1 and mitochondrial oxidative phosphorylation against Aβ25–35-induced

neurotoxicity in human neuroblastoma cells. J. Neurosci. Res. 93, 938-947. doi: 10.1002/jnr.23565

- Kennedy, M. E., Stamford, A. W., Chen, X., Cox, K., Cumming, J. N., Dockendorf, M. F., et al. (2016). The BACE1 inhibitor verubecestat (MK-8931) reduces CNS β-amyloid in animal models and in Alzheimer's disease patients. *Sci. Transl. Med.* 8:363ra150. doi: 10.1126/scitranslmed.aad9704
- Khachaturian, Z. S. (1985). Diagnosis of Alzheimer's disease. Arch. Neurol. 42, 1097–1105. doi: 10.1001/archneur.1985.04060100083029
- Khalil, A. T., Ovais, M., Ullah, I., Ali, M., Jan, S. A., Shinwari, Z. K., et al. (2017a). Bioinspired synthesis of pure massicot phase lead oxide nanoparticles and assessment of their biocompatibility, cytotoxicity and in-vitro biological properties. *Arab. J. Chem.* (in press). doi: 10.1016/j.arabjc.2017.08.009
- Khalil, A. T., Ovais, M., Ullah, I., Ali, M., Shinwari, Z. K., Khamlich, S., et al. (2017b). Sageretia thea (Osbeck.) mediated synthesis of zinc oxide nanoparticles and its biological applications. *Nanomedicine* 12, 1767–1789. doi: 10.2217/ nnm-2017-0124
- Khalil, A. T., Ovais, M., Ullah, I., Ali, M., Shinwari, Z. K., Hassan, D., et al. (2018). Sageretia thea (Osbeck.) modulated biosynthesis of NiO nanoparticles and their in vitro pharmacognostic, antioxidant and cytotoxic potential. Artif. Cells Nanomed. Biotechnol. 46, 838–852. doi: 10.1080/21691401.2017.134 5928
- Khojah, H., and Edrada-Ebel, R. (2017). Identification of bioactive metabolites from *Ficus carica* and their neuroprotective effects of Alzheimer's disease. *Int. J. Med. Health Sci.* 11, 2277–4505.
- Kim, J., Basak, J. M., and Holtzman, D. M. (2009). The role of apolipoprotein E in Alzheimer's disease. *Neuron* 63, 287–303. doi: 10.1016/j.neuron.2009. 06.026
- Kodera, Y., Suzuki, A., Imada, O., Kasuga, S., Sumioka, I., Kanezawa, A., et al. (2002). Physical, chemical, and biological properties of S-allylcysteine, an amino acid derived from garlic. *J. Agric. Food Chem.* 50, 622–632. doi: 10.1021/ jf0106648
- Krishnamurthy, R. G., Senut, M. C., Zemke, D., Min, J., Frenkel, M. B., Greenberg, E. J., et al. (2009). Asiatic acid, a pentacyclic triterpene from *Centella asiatica*, is neuroprotective in a mouse model of focal cerebral ischemia. *J. Neurosci. Res.* 87, 2541–2550. doi: 10.1002/jnr.22071
- Kulkarni, P. D., Ghaisas, M. M., Chivate, N. D., and Sankpal, P. S. (2011). Memory enhancing activity of *Cissampelos pareira* in mice. *Int. J. Pharm. Pharma. Sci.* 3, 206–211.
- Kwon, S.-H., Lee, H.-K., Kim, J.-A., Hong, S.-I., Kim, H.-C., Jo, T.-H., et al. (2010). Neuroprotective effects of chlorogenic acid on scopolamineinduced amnesia via anti-acetylcholinesterase and anti-oxidative activities in mice. *Eur. J. Pharmacol.* 649, 210–217. doi: 10.1016/j.ejphar.2010. 09.001
- Lang, D., Ude, C., Wurglics, M., Schubert-Zsilavecz, M., and Klein, J. (2010). Brain permeability of bilobalide as probed by microdialysis before and after middle cerebral artery occlusion in mice. J. Pharm. Pharm. Sci. 13, 607–614. doi: 10.18433/J31C7Q
- Laribi, B., Kouki, K., M'Hamdi, M., and Bettaieb, T. (2015). Coriander (*Coriandrum sativum L.*) and its bioactive constituents. *Fitoterapia* 103, 9–26. doi: 10.1016/j.fitote.2015.03.012
- Lee, Y. K., Yuk, D. Y., Kim, T. I., Kim, Y. H., Kim, K. T., Kim, K. H., et al. (2009). Protective effect of the ethanol extract of *Magnolia officinalis* and 4-O-methylhonokiol on scopolamine-induced memory impairment and the inhibition of acetylcholinesterase activity. *J. Nat. Med.* 63, 274–282. doi: 10. 1007/s11418-009-0330-z
- Li, J., Yue, M., Zhou, D., Wang, M., and Zhang, H. (2017). Abcb1a but not Abcg2 played a predominant role in limiting the brain distribution of Huperzine A in mice. *Food Chem. Toxicol.* 107, 68–73. doi: 10.1016/j.fct.2017. 06.005
- Li, J., Zhou, L., Ye, D., Huang, S., Shao, K., Huang, R., et al. (2011). Cholinederivate-modified nanoparticles for brain-targeting gene delivery. *Adv. Mater.* 23, 4516–4520. doi: 10.1002/adma.201101899
- Li, P., Matsunaga, K., Yamakuni, T., and Ohizumi, Y. (2003). Nardosinone, the first enhancer of neurite outgrowth-promoting activity of staurosporine and dibutyryl cyclic AMP in PC12D cells. *Dev. Brain Res.* 145, 177–183. doi: 10. 1016/S0165-3806(03)00239-6
- Liang, X.-F., Wang, H.-J., Tian, H., Luo, H., and Chang, J. (2008). Synthesis, structure and properties of novel quaternized carboxymethyl chitosan with

drug loading capacity. *Acta Phys. Chim. Sin.* 24, 223–229. doi: 10.1016/S1872-1508(08)60011-X

- Lim, G. P., Chu, T., Yang, F., Beech, W., Frautschy, S. A., and Cole, G. M. (2001). The curry spice curcumin reduces oxidative damage and amyloid pathology in an Alzheimer transgenic mouse. J. Neurosci. 21, 8370–8377. doi: 10.1523/ JNEUROSCI.21-21-08370.2001
- Liu, G., Men, P., Kudo, W., Perry, G., and Smith, M. A. (2009). Nanoparticlechelator conjugates as inhibitors of amyloid-β aggregation and neurotoxicity: a novel therapeutic approach for Alzheimer disease. *Neurosci. Lett.* 455, 187–190. doi: 10.1016/j.neulet.2009.03.064
- Liu, Q. F., Jeong, H., Lee, J. H., Hong, Y. K., Oh, Y., Kim, Y.-M., et al. (2016). Coriandrum sativum suppresses Aβ42-induced ROS increases, glial cell proliferation, and ERK activation. Am. J. Chin. Med. 44, 1325–1347. doi: 10. 1142/S0192415X16500749
- Lockman, P., Mumper, R., Khan, M., and Allen, D. (2002). Nanoparticle technology for drug delivery across the blood-brain barrier. *Drug Dev. Ind. Pharm.* 28, 1–13. doi: 10.1081/DDC-120001481
- Loveman, E., Green, C., Kirby, J., Takeda, A., Picot, J., Payne, E., et al. (2006). The clinical and cost-effectiveness of donepezil, rivastigmine, galantamine and memantine for Alzheimer's disease. *Health Technol. Assess.* 10, iii–iv, ix–xi, 1–160.
- Ma, T., Gong, K., Yan, Y., Zhang, L., Tang, P., Zhang, X., et al. (2013). Huperzine A promotes hippocampal neurogenesis in vitro and in vivo. *Brain Res.* 1506, 35–43. doi: 10.1016/j.brainres.2013.02.026
- Maclennan, K. M., Darlington, C. L., and Smith, P. F. (2002). The CNS effects of *Ginkgo biloba* extracts and ginkgolide B. *Prog. Neurobiol.* 67, 235–257. doi: 10.1016/S0301-0082(02)00015-1
- Madgula, V. L., Avula, B., Yu, Y.-B., Wang, Y.-H., Tchantchou, F., Fisher, S., et al. (2010). Intestinal and blood-brain barrier permeability of ginkgolides and bilobalide: in vitro and in vivo approaches. *Planta Med.* 76, 599–606. doi: 10.1055/s-0029-1240601
- Mahrous, R., Ghareeb, D. A., Fathy, H. M., EL-Khair, R. M. A., and Omar, A. A. (2017). The protective effect of Egyptian Withania somnifera against Alzheimer's. Med. Aromat Plants 6:285. doi: 10.4172/2167-0412.1000285
- Malik, J., Karan, M., and Dogra, R. (2017). Ameliorating effect of *Celastrus paniculatus* standardized extract and its fractions on 3-nitropropionic acid induced neuronal damage in rats: possible antioxidant mechanism. *Pharm. Biol.* 55, 980–990. doi: 10.1080/13880209.2017.1285945
- Mandal, S., Debnath, K., Jana, N. R., and Jana, N. R. (2017). Trehalosefunctionalized gold nanoparticle for inhibiting intracellular protein aggregation. *Langmuir* 33, 13996–14003. doi: 10.1021/acs.langmuir.7b0 2202
- Mangialasche, F., Solomon, A., Winblad, B., Mecocci, P., and Kivipelto, M. (2010). Alzheimer's disease: clinical trials and drug development. *Lancet Neurol.* 9, 702–716. doi: 10.1016/S1474-4422(10)70119-8
- Mani, V., and Parle, M. (2009). Memory-enhancing activity of *Coriandrum sativum* in rats. *Pharmacologyonline* 2, 827–839.
- Mao, X., Yin, W., Liu, M., Ye, M., Liu, P., Liu, J., et al. (2011). Osthole, a natural coumarin, improves neurobehavioral functions and reduces infarct volume and matrix metalloproteinase-9 activity after transient focal cerebral ischemia in rats. *Brain Res.* 1385, 275–280. doi: 10.1016/j.brainres.2011. 02.015
- Markoutsa, E., Pampalakis, G., Niarakis, A., Romero, I. A., Weksler, B., Couraud, P. -O., et al. (2011). Uptake and permeability studies of BBB-targeting immunoliposomes using the hCMEC/D3 cell line. *Eur. J. Pharm. Biopharm.* 77, 265–274. doi: 10.1016/j.ejpb.2010.11.015
- Markoutsa, E., Papadia, K., Clemente, C., Flores, O., and Antimisiaris, S. G. (2012). Anti-Aβ-MAb and dually decorated nanoliposomes: effect of Aβ1-42 peptides on interaction with hCMEC/D3 cells. *Eur. J. Pharm. Biopharm.* 81, 49–56. doi: 10.1016/j.ejpb.2012.02.006
- Markoutsa, E., Papadia, K., Giannou, A., Spella, M., Cagnotto, A., Salmona, M., et al. (2014). Mono and dually decorated nanoliposomes for brain targeting, in vitro and in vivo studies. *Pharm. Res.* 31, 1275–1289. doi: 10.1007/s11095-013-1249-3
- Marumoto, S., Okuno, Y., and Miyazawa, M. (2017). Inhibition of β -secretase activity by monoterpenes, sesquiterpenes, and C13 norisoprenoids. *J. Oleo Sci.* 66, 851–855. doi: 10.5650/jos.ess16188

- Masserini, M. (2013). Nanoparticles for brain drug delivery. ISRN Biochem. 2013:238428. doi: 10.1155/2013/238428
- Md, S., Ali, M., Baboota, S., Sahni, J. K., Bhatnagar, A., et al. (2014). Preparation, characterization, in vivo biodistribution and pharmacokinetic studies of donepezil-loaded PLGA nanoparticles for brain targeting. *Drug Dev. Ind. Pharm.* 40, 278–287. doi: 10.3109/03639045.2012.758130
- Megill, A., Lee, T., DiBattista, A. M., Song, J. M., Spitzer, M. H., Rubinshtein, M., et al. (2013). A tetra (ethylene glycol) derivative of benzothiazole aniline enhances Ras-mediated spinogenesis. J. Neurosci. 33, 9306–9318. doi: 10.1523/ JNEUROSCI.1615-12.2013
- Mellott, T. J., Huleatt, O. M., Shade, B. N., Pender, S. M., Liu, Y. B., Slack, B. E., et al. (2017). Perinatal choline supplementation reduces amyloidosis and increases choline acetyltransferase expression in the hippocampus of the APPswePS1dE9 Alzheimer's disease model mice. *PLoS One* 12:e0170450. doi: 10.1371/journal. pone.0170450
- Meng, Q., Wang, A., Hua, H., Jiang, Y., Wang, Y., Mu, H., et al. (2018). Intranasal delivery of Huperzine A to the brain using lactoferrin-conjugated N-trimethylated chitosan surface-modified PLGA nanoparticles for treatment of Alzheimer's disease. *Int. J. Nanomedicine* 13:705–718. doi: 10.2147/IJN. S151474
- Minati, L., Edginton, T., Grazia Bruzzone, M., and Giaccone, G. (2009). Reviews: current concepts in Alzheimer's disease: a multidisciplinary review. Am. J. Alzheimers Dis. Other Demen. 24, 95–121. doi: 10.1177/1533317508328602
- Mistry, A., Stolnik, S., and Illum, L. (2009). Nanoparticles for direct nose-to-brain delivery of drugs. *Int. J. Pharm.* 379, 146–157. doi: 10.1016/j.ijpharm.2009. 06.019
- Modi, G., Pillay, V., and Choonara, Y. E. (2010). Advances in the treatment of neurodegenerative disorders employing nanotechnology. Ann. N. Y. Acad. Sci. 1184, 154–172. doi: 10.1111/j.1749-6632.2009.05108.x
- Mutlu, N. B., Deðim, Z., Yılmaz, Ş., Eşsiz, D., and Nacar, A. (2011). New perspective for the treatment of Alzheimer diseases: liposomal rivastigmine formulations. *Drug Dev. Ind. Pharm.* 37, 775–789. doi: 10.3109/03639045.2010. 541262
- Nazem, A., and Mansoori, G. A. (2011). Nanotechnology for Alzheimer's disease detection and treatment. *Insciences J.* 1, 169–193. doi: 10.5640/insc.0104169
- Nellore, J., Pauline, C., and Amarnath, K. (2013). Bacopa monnieri phytochemicals mediated synthesis of platinum nanoparticles and its neurorescue effect on 1-methyl 4-phenyl 1,2,3,6 tetrahydropyridine-induced experimental parkinsonism in zebrafish. J. Neurodegener. Dis. 2013:972391. doi: 10.1155/2013/972391
- Nie, L., Xia, J., Li, H., Zhang, Z., Yang, Y., Huang, X., et al. (2017). Ginsenoside Rg1 ameliorates behavioral abnormalities and modulates the hippocampal proteomic change in triple transgenic mice of Alzheimer's disease. Oxid. Med. Cell. Longev. 2017:6473506. doi: 10.1155/2017/6473506
- Nillert, N., Pannangrong, W., Welbat, J. U., Chaijaroonkhanarak, W., Sripanidkulchai, K., and Sripanidkulchai, B. (2017). Neuroprotective effects of aged garlic extract on cognitive dysfunction and neuroinflammation induced by β -amyloid in rats. *Nutrients* 9:24. doi: 10.3390/nu90 10024
- Noetzli, M., and Eap, C. B. (2013). Pharmacodynamic, pharmacokinetic and pharmacogenetic aspects of drugs used in the treatment of Alzheimer's disease. *Clin. Pharmacokinet.* 52, 225–241. doi: 10.1007/s40262-013-0038-9
- Oboh, G., Agunloye, O. M., Akinyemi, A. J., Ademiluyi, A. O., and Adefegha, S. A. (2013). Comparative study on the inhibitory effect of caffeic and chlorogenic acids on key enzymes linked to Alzheimer's disease and some pro-oxidant induced oxidative stress in rats' brain-in vitro. *Neurochem. Res.* 38, 413–419. doi: 10.1007/s11064-012-0935-6
- Ossola, B., Kääriäinen, T. M., and Männistö, P. T. (2009). The multiple faces of quercetin in neuroprotection. *Expert Opin. Drug Saf.* 8, 397–409. doi: 10.1517/14740330903026944
- Ovais, M., Ahmad, I., Khalil, A. T., Mukherjee, S., Javed, R., Ayaz, M., et al. (2018a). Wound healing applications of biogenic colloidal silver and gold nanoparticles: recent trends and future prospects. *Appl. Microbiol. Biotechnol.* 102, 4305–4318. doi: 10.1007/s00253-018-8939-z
- Ovais, M., Ayaz, M., Khalil, A. T., Shah, S. A., Jan, M. S., Raza, A., et al. (2018b). HPLC-DAD finger printing, antioxidant, cholinesterase, and α -glucosidase

inhibitory potentials of a novel plant Olax nana. BMC Complement. Altern. Med. 18:1. doi: 10.1186/s12906-017-2057-9

- Ovais, M., Khalil, A. T., Islam, N. U., Ahmad, I., Ayaz, M., Saravanan, M., et al. (2018c). Role of plant phytochemicals and microbial enzymes in biosynthesis of metallic nanoparticles. *Appl. Microbiol. Biotechnol.* doi: 10.1007/s00253-018-9146-7 [Epub ahead of print].
- Ovais, M., Khalil, A. T., Raza, A., Islam, N. U., Ayaz, M., Saravanan, M., et al. (2018d). Multifunctional theranostic applications of biocompatible greensynthesized colloidal nanoparticles. *Appl. Microbiol. Biotechnol.* 102, 4393– 4408. doi: 10.1007/s00253-018-8928-2
- Ovais, M., Khalil, A. T., Raza, A., Khan, M. A., Ahmad, I., Islam, N. U., et al. (2016). Green synthesis of silver nanoparticles via plant extracts: beginning a new era in cancer theranostics. *Nanomedicine* 12, 3157–3177. doi: 10.2217/nnm-2016-0279
- Ovais, M., Nadhman, A., Khalil, A. T., Raza, A., Khuda, F., Sohail, M. F., et al. (2017a). Biosynthesized colloidal silver and gold nanoparticles as emerging leishmanicidal agents: an insight. *Nanomedicine* 12, 2807–2819. doi: 10.2217/ nnm-2017-0233
- Ovais, M., Raza, A., Naz, S., Islam, N. U., Khalil, A. T., Ali, S., et al. (2017b). Current state and prospects of the phytosynthesized colloidal gold nanoparticles and their applications in cancer theranostics. *Appl. Microbiol. Biotechnol.* 101, 3551–3565. doi: 10.1007/s00253-017-8250-4
- Palle, S., and Neerati, P. (2017). Quercetin nanoparticles attenuates scopolamine induced spatial memory deficits and pathological damages in rats. *Bull. Fac. Pharm. Cairo Univ.* 55, 101–106. doi: 10.1016/j.bfopcu.2016.10.004
- Papadia, K., Giannou, A. D., Markoutsa, E., Bigot, C., Vanhoute, G., Mourtas, S., et al. (2017). Multifunctional LUV liposomes decorated for BBB and amyloid targeting-B. In vivo brain targeting potential in wild-type and APP/PS1 mice. *Eur. J. Pharm. Sci.* 102, 180–187. doi: 10.1016/j.ejps.2017. 03.010
- Pardeshi, C. V., and Belgamwar, V. S. (2013). Direct nose to brain drug delivery via integrated nerve pathways bypassing the blood–brain barrier: an excellent platform for brain targeting. *Expert Opin. Drug Deliv.* 10, 957–972. doi: 10.1517/ 17425247.2013.790887
- Pérez-Severiano, F., Salvatierra-Sánchez, R., Rodr*ι*guez-Pérez, M., Cuevas-Mart*ι* nez, E. Y., Guevara, J., Limón, D., et al. (2004). S-Allylcysteine prevents amyloid-β peptide-induced oxidative stress in rat hippocampus and ameliorates learning deficits. *Eur. J. Pharmacol.* 489, 197–202. doi: 10.1016/j.ejphar.2004. 03.001
- Preedy, V. R. (2012). *Tea in Health and Disease Prevention*. Cambridge, MA: Academic Press.
- Puerta, E., Suárez-Santiago, J. E., Santos-Magalhães, N. S., Ramirez, M. J., and Irache, J. M. (2017). Effect of the oral administration of nanoencapsulated quercetin on a mouse model of Alzheimer's disease. *Int. J. Pharm.* 517, 50–57. doi: 10.1016/j.ijpharm.2016.11.061
- Qi, F.-Y., Yang, L., Tian, Z., Zhao, M.-G., Liu, S.-B., and An, J.-Z. (2014). Neuroprotective effects of Asiaticoside. *Neural Regen. Res.* 9, 1275–1282. doi: 10.4103/1673-5374.137574
- Rafii, M., Walsh, S., Little, J., Behan, K., Reynolds, B., Ward, C., et al. (2011). A phase II trial of huperzine A in mild to moderate Alzheimer disease. *Neurology* 76, 1389–1394. doi: 10.1212/WNL.0b013e318216eb7b
- Rebai, O., Belkhir, M., Sanchez-Gomez, M. V., Matute, C., Fattouch, S., and Amri, M. (2017). Differential molecular targets for neuroprotective effect of chlorogenic acid and its related compounds against glutamate induced excitotoxicity and oxidative stress in rat cortical neurons. *Neurochem. Res.* 42, 3559–3572. doi: 10.1007/s11064-017-2403-9
- Remya, C., Dileep, K., Variayr, E., and Sadasivan, C. (2016). An in silico guided identification of nAChR agonists from Withania somnifera. Front. Life Sci. 9, 201–213. 10.1080/21553769.2016.1207569 doi: 10.1080/21553769.2016. 1207569
- Rocha, S. (2013). Targeted drug delivery across the blood brain barrier in Alzheimer's disease. *Curr. Pharm. Des.* 19, 6635–6646. doi: 10.2174/ 13816128113199990613
- Rotman, M., Welling, M. M., Bunschoten, A., de Backer, M. E., Rip, J., Nabuurs, R. J., et al. (2015). Enhanced glutathione PEGylated liposomal brain delivery of an anti-amyloid single domain antibody fragment in a mouse model for Alzheimer's disease. J. Control. Release 203, 40–50. doi: 10.1016/j.jconrel.2015. 02.012

- Rubio, J., Qiong, W., Liu, X., Jiang, Z., Dang, H., Chen, S.-L., et al. (2011). Aqueous extract of black maca (*Lepidium meyenii*) on memory impairment induced by ovariectomy in mice. *Evid. Based Complement. Alternat. Med.* 2011:253958. doi: 10.1093/ecam/nen063
- Rui, W., Yan-Nan, L., Guang-Ji, W., Hai-Ping, H., Xiao-Lan, W., and Fang, Z. (2009). Neuroprotective effects and brain transport of ginsenoside Rg1. *Chin. J. Nat. Med.* 7, 315–320. doi: 10.3724/SPJ.1009.2008.00315
- Sadiq, A., Zeb, A., Ullah, F., Ahmad, S., Ayaz, M., Rashid, U., et al. (2018). Chemical characterization, analgesic, antioxidant, and anticholinesterase potentials of essential oils from *Isodon rugosus* Wall. ex. Benth. *Front. Pharmacol.* 9:623. doi: 10.3389/fphar.2018.00623
- Saharia, K., Kumar, R., Gupta, K., Mishra, S., and Subramaniam, J. R. (2016). A novel way of amelioration of amyloid beta induced toxicity in *Caenorhabditis* elegans. Ann. Neurosci. 23, 149–154. doi: 10.1159/000449180
- Sahni, J. K., Doggui, S., Ali, J., Baboota, S., Dao, L., and Ramassamy, C. (2011). Neurotherapeutic applications of nanoparticles in Alzheimer's disease. *J. Control. Release* 152, 208–231. doi: 10.1016/j.jconrel.2010.11.033
- Salloway, S., Sperling, R., Fox, N. C., Blennow, K., Klunk, W., Raskind, M., et al. (2014). Two phase 3 trials of bapineuzumab in mild-to-moderate Alzheimer's disease. N. Engl. J. Med. 370, 322–333. doi: 10.1056/NEJMoa1304839
- Salmaso, S., and Caliceti, P. (2013). Stealth properties to improve therapeutic efficacy of drug nanocarriers. *J. Drug Deliv.* 2013:374252. doi: 10.1155/2013/374252
- Serafini, M. M., Catanzaro, M., Rosini, M., Racchi, M., and Lanni, C. (2017). Curcumin in Alzheimer's disease: Can we think to new strategies and perspectives for this molecule? *Pharmacol. Res.* 124, 146–155. doi: 10.1016/j. phrs.2017.08.004
- Selkoe, D. J. (2001). Alzheimer's disease: genes, proteins, and therapy. *Physiol. Rev.* 81, 741–766. doi: 10.1152/physrev.2001.81.2.741
- Seow, S. L. S., Hong, S. L., Lee, G. S., Malek, S. N. A., and Sabaratnam, V. (2017). 6-shogaol, a neuroactive compound of ginger (jahe gajah) induced neuritogenic activity via NGF responsive pathways in PC-12 cells. *BMC Complement. Altern. Med.* 17:334. doi: 10.1186/s12906-017-1837-6
- Sevigny, J., Chiao, P., Bussière, T., Weinreb, P. H., Williams, L., Maier, M., et al. (2016). The antibody aducanumab reduces Aβ plaques in Alzheimer's disease. *Nature* 537, 50–56. doi: 10.1038/nature19323
- Sharma, H. S., Drieu, K., Alm, P., and Westman, J. (2000). Role of nitric oxide in blood-brain barrier permeability, brain edema and cell damage following hyperthermic brain injury. An experimental study using EGB-761 and Gingkolide B pretreatment in the rat, in *Brain Edema XI. Acta Neurochirurgica Supplements*, ed. A. D. Mendelow (Vienna: Springer), 81–86.
- Shen, J., Zhao, Z., Shang, W., Liu, C., Zhang, B., Zhao, L., et al. (2017). Ginsenoside Rg1 nanoparticle penetrating the blood-brain barrier to improve the cerebral function of diabetic rats complicated with cerebral infarction. *Int. J. Nanomedicine* 12, 6477–6486. doi: 10.2147/IJN.S139602
- Shi, C., Liu, J., Wu, F., and Yew, D. T. (2010). *Ginkgo biloba* extract in Alzheimer's disease: from action mechanisms to medical practice. *Int. J. Mol. Sci.* 11, 107–123. doi: 10.3390/ijms11010107
- Shinomol, G. K., and Bharath, M. M. (2011). Exploring the role of "Brahmi"(Bacopa monnieri and Centella asiatica) in brain function and therapy. Recent Pat. Endocr. Metab. Immune Drug Discov. 5, 33–49. doi: 10.2174/187221411794351833
- Shu, X., and Zhu, K. (2000). A novel approach to prepare tripolyphosphate/chitosan complex beads for controlled release drug delivery. *Int. J. Pharm.* 201, 51–58. doi: 10.1016/S0378-5173(00)00403-8
- Siemers, E. R., Sundell, K. L., Carlson, C., Case, M., Sethuraman, G., Liu-Seifert, H., et al. (2016). Phase 3 solanezumab trials: secondary outcomes in mild Alzheimer's disease patients. *Alzheimers Dement*. 12, 110–120. doi: 10.1016/j. jalz.2015.06.1893
- Smith, A., Giunta, B., Bickford, P. C., Fountain, M., Tan, J., and Shytle, R. D. (2010). Nanolipidic particles improve the bioavailability and α -secretase inducing ability of epigallocatechin-3-gallate (EGCG) for the treatment of Alzheimer's disease. *Int. J. Pharm.* 389, 207–212. doi: 10.1016/j.ijpharm.2010.01.012
- Song, J.-H., Lee, J.-W., Shim, B., Lee, C.-Y., Choi, S., Kang, C., et al. (2013). Glycyrrhizin alleviates neuroinflammation and memory deficit induced by systemic lipopolysaccharide treatment in mice. *Molecules* 18, 15788–15803. doi: 10.3390/molecules181215788

- Songjiang, Z., and Lixiang, W. (2009). Amyloid-beta associated with chitosan nano-carrier has favorable immunogenicity and permeates the BBB. AAPS PharmSciTech 10, 900–905. doi: 10.1208/s12249-009-9279-1
- Sperling, R. A., Karlawish, J., and Johnson, K. A. (2013). Preclinical Alzheimer disease—the challenges ahead. *Nat. Rev. Neurol.* 9, 54–58. doi: 10.1038/ nrneurol.2012.241
- Subash, S., Essa, M. M., Braidy, N., Al-Jabri, A., Vaishnav, R., Al-Adawi, S., et al. (2016). Consumption of fig fruits grown in Oman can improve memory, anxiety, and learning skills in a transgenic mice model of Alzheimer's disease. *Nutr. Neurosci.* 19, 475–483. doi: 10.1179/1476830514Y.0000000131
- Suganthy, N., Ramkumar, V. S., Pugazhendhi, A., Benelli, G., and Archunan, G. (2017). Biogenic synthesis of gold nanoparticles from *Terminalia arjuna* bark extract: assessment of safety aspects and neuroprotective potential via antioxidant, anticholinesterase, and antiamyloidogenic effects. *Environ. Sci. Pollut. Res.* 25, 10418–10433. doi: 10.1007/s11356-017-9789-4
- Sun, D., Li, N., Zhang, W., Zhao, Z., Mou, Z., Huang, D., et al. (2016). Design of PLGA-functionalized quercetin nanoparticles for potential use in Alzheimer's disease. *Colloids Surf. B Biointerfaces* 148, 116–129. doi: 10.1016/j.colsurfb.2016. 08.052
- Tabuchi, M., Imamura, S., Kawakami, Z., Ikarashi, Y., and Kase, Y. (2012). The blood–brain barrier permeability of 18β-glycyrrhetinic acid, a major metabolite of glycyrrhizin in Glycyrrhiza root, a constituent of the traditional Japanese medicine yokukansan. *Cell. Mol. Neurobiol.* 32, 1139–1146. doi: 10.1007/s10571-012-9839-x
- Tanifum, E. A., Dasgupta, I., Srivastava, M., Bhavane, R. C., Sun, L., Berridge, J., et al. (2012). Intravenous delivery of targeted liposomes to amyloid-β pathology in APP/PSEN1 transgenic mice. *PLoS One* 7:e48515. doi: 10.1371/journal.pone. 0048515
- Tanifum, E. A., Ghaghada, K., Vollert, C., Head, E., Eriksen, J. L., and Annapragada, A. (2016). A novel liposomal nanoparticle for the imaging of amyloid plaque by magnetic resonance imaging. J. Alzheimers Dis. 52, 731–745. doi: 10.3233/JAD-151124
- Ternchoocheep, K., Surangkul, D., and Ysothonsreekul, S. (2017). The recovery and protective effects of asiatic acid on differentiated human neuroblastoma SH-SY5Y cells cytotoxic-induced by cholesterol. Asian Pac. J. Trop. Biomed. 7, 416–420. doi: 10.1016/j.apjtb.2017.01.012
- Tiwari, S. K., Agarwal, S., Seth, B., Yadav, A., Nair, S., Bhatnagar, P., et al. (2013). Curcumin-loaded nanoparticles potently induce adult neurogenesis and reverse cognitive deficits in Alzheimer's disease model via canonical Wnt/β-catenin pathway. ACS Nano 8, 76–103. doi: 10.1021/nn40 5077y
- Tsai, Y.-M., Chien, C.-F., Lin, L.-C., and Tsai, T.-H. (2011). Curcumin and its nano-formulation: the kinetics of tissue distribution and blood-brain barrier penetration. *Int. J. Pharm.* 416, 331–338. doi: 10.1016/j.ijpharm.2011. 06.030
- Ulbrich, K., Hekmatara, T., Herbert, E., and Kreuter, J. (2009). Transferrinand transferrin-receptor-antibody-modified nanoparticles enable drug delivery across the blood-brain barrier (BBB). *Eur. J. Pharm. Biopharm.* 71, 251–256. doi: 10.1016/j.ejpb.2008.08.021
- Ullah, F., Ayaz, M., Sadiq, A., Hussain, A., Ahmad, S., Imran, M., et al. (2016). Phenolic, flavonoid contents, anticholinesterase and antioxidant evaluation of Iris germanica var; florentina. *Nat. Prod. Res.* 30, 1440–1444. doi: 10.1080/ 14786419.2015.1057585
- Vareed, S. K., Bauer, A. K., Nair, K. M., Liu, Y., Jayaprakasam, B., and Nair, M. G. (2014). Blood-brain barrier permeability of bioactive withanamides present in *Withania somnifera* fruit extract. *Phytother. Res.* 28, 1260–1264. doi: 10.1002/ ptr.5118
- Videira, R., Castanheira, P., Graþos, M. R., Resende, R., Salgueiro, L., Faro, C., et al. (2014). Dose-dependent inhibition of BACE-1 by the monoterpenoid 2, 3, 4, 4-tetramethyl-5-methylenecyclopent-2-enone in cellular and mouse models of Alzheimer's disease. J. Nat. Prod. 77, 1275–1279. doi: 10.1021/np40 0903w
- Viola, K. L., Sbarboro, J., Sureka, R., De, M., Bicca, M. A., Wang, J., et al. (2015). Towards non-invasive diagnostic imaging of early-stage Alzheimer's disease. *Nat. Nanotechnol.* 10, 91–98. doi: 10.1038/nnano.2014.254
- Vitthal, K. U., Pillai, M., and Kininge, P. (2013). Study of solid lipid nanoparticles as a carrier for bacoside. *Int. J. Pharma Biosci.* 3, 414–426.

- Wang, M., Li, Y., Ni, C., and Song, G. (2017). Honokiol attenuates oligomeric amyloid β1-42-induced Alzheimer's disease in mice through attenuating mitochondrial apoptosis and inhibiting the nuclear factor kappa-B signaling pathway. *Cell. Physiol. Biochem.* 43, 69–81. doi: 10.1159/00048 0320
- Wang, X., Chi, N., and Tang, X. (2008). Preparation of estradiol chitosan nanoparticles for improving nasal absorption and brain targeting. *Eur. J. Pharm. Biopharm.* 70, 735–740. doi: 10.1016/j.ejpb.2008.07.005
- Wang, X., Duan, X., Yang, G., Zhang, X., Deng, L., Zheng, H., et al. (2011). Honokiol crosses BBB and BCSFB, and inhibits brain tumor growth in rat 9L intracerebral gliosarcoma model and human U251 xenograft glioma model. *PLoS One* 6:e18490. doi: 10.1371/journal.pone.0018490
- Wang, Z. H., Wang, Z. Y., Sun, C. S., Wang, C. Y., Jiang, T. Y., and Wang, S. L. (2010). Trimethylated chitosan-conjugated PLGA nanoparticles for the delivery of drugs to the brain. *Biomaterials* 31, 908–915. doi: 10.1016/j.biomaterials. 2009.09.104
- Wilson, B., Samanta, M. K., Santhi, K., Kumar, K. P. S., Paramakrishnan, N., and Suresh, B. (2008). Poly (n-butylcyanoacrylate) nanoparticles coated with polysorbate 80 for the targeted delivery of rivastigmine into the brain to treat Alzheimer's disease. *Brain Res.* 1200, 159–168. doi: 10.1016/j.brainres.2008.break01.039
- Wilson, B., Samanta, M. K., Santhi, K., Kumar, K. S., Ramasamy, M., and Suresh, B. (2010). Chitosan nanoparticles as a new delivery system for the anti-Alzheimer drug tacrine. *Nanomedicine* 6, 144–152. doi: 10.1016/j.nano.2009. 04.001
- Winblad, B., and Jelic, V. (2004). Long-term treatment of Alzheimer disease: efficacy and safety of acetylcholinesterase inhibitors. *Alzheimer Dis. Assoc. Disord.* 18, S2–S8. doi: 10.1097/01.wad.0000127495. 10774.a4
- Wu, L., Zhang, Q.-L., Zhang, X.-Y., Lv, C., Li, J., Yuan, Y., et al. (2012). Pharmacokinetics and blood-brain barrier penetration of (+)-catechin and (-)-epicatechin in rats by microdialysis sampling coupled to high-performance liquid chromatography with chemiluminescence detection. *J. Agric. Food Chem.* 60, 9377–9383. doi: 10.1021/jf301787f
- Xie, C.-L., Li, J.-H., Wang, W.-W., Zheng, G.-Q., and Wang, L.-X. (2015). Neuroprotective effect of ginsenoside-Rg1 on cerebral ischemia/reperfusion injury in rats by downregulating protease-activated receptor-1 expression. *Life Sci.* 121, 145–151. doi: 10.1016/j.lfs.2014.12.002
- Xu, Z.-C., Wang, X.-B., Yu, W.-Y., Xie, S.-S., Li, S.-Y., and Kong, L.-Y. (2014). Design, synthesis and biological evaluation of benzylisoquinoline derivatives as multifunctional agents against Alzheimer's disease. *Bioorg. Med. Chem. Lett.* 24, 2368–2373. doi: 10.1016/j.bmcl.2014. 03.058
- Yang, F., Lim, G. P., Begum, A. N., Ubeda, O. J., Simmons, M. R., Ambegaokar, S. S., et al. (2005). Curcumin inhibits formation of amyloid β oligomers and fibrils, binds plaques, and reduces amyloid in vivo. *J. Biol. Chem.* 280, 5892–5901. doi: 10.1074/jbc.M404751200
- Yerer, M. B., Tiryaki, M. K., and Demirpolat, E. (2017). GSK-3beta inhibitory effects of 6-gingerol and 6-shogaol help to the recovery of SHSY-5Y cells after amyloid beta1–42 oligomer or aggregate toxicity. J. Cell. Biotechnol. 2, 145–157. doi: 10.3233/JCB-15035
- Yiannopoulou, K. G., and Papageorgiou, S. G. (2013). Current and future treatments for Alzheimer's disease. *Ther. Adv. Neurol. Disord.* 6, 19–33. doi: 10.1177/1756285612461679
- Yin, Z., Yu, T., and Xu, Y. (2015). Preparation of amyloid immuno-nanoparticles as potential MRI contrast agents for Alzheimer's disease diagnosis. J. Nanosci. Nanotechnol. 15, 6429–6434. doi: 10.1166/jnn.2015.11296
- Yoo, D. Y., Nam, Y., Kim, W., Yoo, K.-Y., Park, J., Lee, C. H., et al. (2011). Effects of *Ginkgo biloba* extract on promotion of neurogenesis in the hippocampal dentate gyrus in C57BL/6 mice. *J. Vet. Med. Sci.* 73, 71–76. doi: 10.1292/jvms.10-0294
- Yordanov, G. (2012). Poly (alkyl cyanoacrylate) nanoparticles as drug carriers: 33 years later. Bulg. J. Chem. 1, 61–72.
- Zenaro, E., Pietronigro, E., Della Bianca, V., Piacentino, G., Marongiu, L., Budui, S., et al. (2015). Neutrophils promote Alzheimer's disease–like pathology and cognitive decline via LFA-1 integrin. *Nat. Med.* 21, 880–886. doi: 10.1038/nm. 3913

- Zhang, C., Chen, J., Feng, C., Shao, X., Liu, Q., Zhang, Q., et al. (2014). Intranasal nanoparticles of basic fibroblast growth factor for brain delivery to treat Alzheimer's disease. *Int. J. Pharm.* 461, 192–202. doi: 10.1016/j.ijpharm.2013. 11.049
- Zhang, J., Zhou, X., Yu, Q., Yang, L., Sun, D., Zhou, Y., et al. (2014). Epigallocatechin-3-gallate (EGCG)-stabilized selenium nanoparticles coated with Tet-1 peptide to reduce amyloid-β aggregation and cytotoxicity. *ACS Appl. Mater. Interfaces* 6, 8475–8487. doi: 10.1021/am501341u
- Zhang, P., Liu, X., Zhu, Y., Chen, S., Zhou, D., and Wang, Y. (2013). Honokiol inhibits the inflammatory reaction during cerebral ischemia reperfusion by suppressing NF-κB activation and cytokine production of glial cells. *Neurosci. Lett.* 534, 123–127. doi: 10.1016/j.neulet.2012.11.052
- Zhang, X., Fan, Z., and Jin, T. (2017). Crocin protects against cerebral-ischemiainduced damage in aged rats through maintaining the integrity of bloodbrain barrier. *Restor. Neurol. Neurosci.* 35, 65–75. doi: 10.3233/RNN-16 0696
- Zhang, Z. J., Cheang, L. C. V., Wang, M. W., and Lee, S. M.-Y. (2011). Quercetin exerts a neuroprotective effect through inhibition of the iNOS/NO system and pro-inflammation gene expression in PC12 cells and in zebrafish. *Int. J. Mol. Med.* 27, 195–203.
- Zhao, J., Lu, S., Yu, H., Duan, S., and Zhao, J. (2018). Baicalin and ginsenoside Rb1 promote the proliferation and differentiation of neural stem cells in Alzheimer's disease model rats. *Brain Res.* 1678, 187–194. doi: 10.1016/j.brainres.2017. 10.003
- Zheng, Y.-Q., Liu, J.-X., Wang, J.-N., and Xu, L. (2007). Effects of crocin on reperfusion-induced oxidative/nitrative injury to cerebral microvessels after global cerebral ischemia. *Brain Res.* 1138, 86–94. doi: 10.1016/j.brainres.2006. 12.064

- Zhou, Y., Li, H.-Q., Lu, L., Fu, D.-L., Liu, A.-J., Li, J.-H., et al. (2014). Ginsenoside Rg1 provides neuroprotection against blood brain barrier disruption and neurological injury in a rat model of cerebral ischemia/reperfusion through downregulation of aquaporin 4 expression. *Phytomedicine* 21, 998–1003. doi: 10.1016/j.phymed.2013.12.005
- Zlokovic, B. V. (2011). Neurovascular pathways to neurodegeneration in Alzheimer's disease and other disorders. *Nat. Rev. Neurosci.* 12, 723–738. doi: 10.1038/nrn3114
- Zohra, T., Ovais, M., Khalil, A. T., Qasim, M., Ayaz, M., and Shinwari, Z. K. (2018). Extraction optimization, total phenolic, flavonoid contents, HPLC-DAD analysis and diverse pharmacological evaluations of *Dysphania ambrosioides* (L.) Mosyakin & Clemants. *Nat. Prod. Res.* doi: 10.1080/14786419. 2018.1437428 [Epub ahead of print].

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