Enterobacter cloacae harbouring bla_{KPC-2} and qnrB-1 isolated from a cystic fibrosis patient: a case report

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Abstract

We describe the first detection of a KPC-2- and QnrB-producing Enterobacter cloacae from a patient with cystic fibrosis. The blaKPC-2 and qnrB-1 genes were located in a 79.8-kb plasmid. The presence of blaKPC-2 and qnrB-1 genes was determined by PCR and sequencing. Mobilization of plasmid containing *bla*_{KPC2} gene was assayed by conjugation. © 2018 The Authors. Published by Elsevier Ltd.

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Introduction

Enterobacteriaceae are uncommonly isolated from respiratory specimens and appear to play a minor role in lung infection in individuals with cystic fibrosis (CF). However, increasing reports of Klebsiella pneumoniae carbapenemase (KPC)-producing Enterobacteriaceae in individuals with CF is a matter of concern [1,2]. This study describes the first report of a KPC-2 and QnrB-I-producing Enterobacter cloacae from an individual with CF.

Case report

The patient was a 21-year-old woman who presented weight loss and cough since birth. A diagnosis of CF was confirmed at 2 years of age, with sweat test (>90 mEq/L) and the genotype F508del/F508del. Pancreatic insufficiency was soon diagnosed and treatment was started with nutritional support, pancreatic enzymes, vitamins, physiotherapy, mucolytics and bronchodilators. Pseudomonas aeruginosa was first detected at the time of diagnosis and chronic colonization was assumed 5 years later (August 2003) when she started on inhaled tobramycin every other month. Although she had good treatment compliance and achieved good nutritional status (body mass index (BMI) $19-21 \text{ kg/m}^2$), she still had frequent respiratory exacerbations that required either oral ciprofloxacin or intravenous ceftazidime and amikacin. By 15-years of age, she presented a BMI of 20.8 kg/m²; forced expiratory volume in I second (FEVI) 67.5%; forced vital capacity (FVC): 85.8% and sputum with imipenem-resistant P. aeruginosa. At this time (May 2011), E. cloacae was isolated and the patient was using only inhaled tobramycin. Two months after the isolation of E. cloacae, the patient had another exacerbation (lost weight and BMI 20.8 kg/ m²; dyspnoea, more secretions) and received intravenous antibiotics for 2 weeks. During subsequent evaluations FEVI decreased from 67.5% to 48% (rate: 4.5% each year) and FVC decreased from: 85.8% to 78% (rate: 1.95% each year) while BMI was stable (20.8 kg/m²) during the period. From this

moment and until the latest cultures, only mucoid imipenemresistant *P. aeruginosa* strains and methicillin-susceptible *Staphylococcus aureus* were isolated.

Enterobacter cloacae identification and antimicrobial susceptibility profile were investigated using the Vitek2 system (bio-Mérieux, Marcy l'Etoile, France). The strain was susceptible to amikacin, gentamicin, ciprofloxacin and colistin, with MICs of 8, <1, 1 and <0.5 mg/L, respectively, using criteria defined by the CLSI [3], with the exception of colistin, where European Committee on Antimicrobial Susceptibility Testing criteria were used. The strain was resistant to a wide variety of antimicrobials, including carbapenems (Table 1). According to phenotypic methods using EDTA and phenyl boronic acid as enzymatic blockers [4] the isolate was defined as a possible KPC producer. This was confirmed by PCR amplifying and sequencing the bla_{KPC-} $_{2}$ gene [5]. The presence of the quinolone-resistance gene qnrBwas also verified by PCR [6] (Table 1). Analyses with SI nuclease pulsed-field gel electrophoresis and Southern blot confirmed the size of the plasmid and showed the presence of both the bla_{KPC-2} and qnrB-1 genes in a 78.0-kb plasmid. Plasmid mobilization was successfully performed by conjugation with an Escherichia coli J53 strain. Selection with imipenem (I mg/L) allowed the isolation of transconjugants harbouring the *bla*KPC-2 and *qnrB-1* genes, with an increased MIC for meropenem and imipenem (Table 1).

Discussion

The presence of Enterobacteriaceae has seldom been described in patients with CF. However, a recent study showed the persistent isolation of Escherichia coli (i.e. for longer than 6 months) among German CF patients [2]. Besides that, reports of KPC-producing K. pneumoniae in individuals with CF supports a role for members of the Enterobacteriaceae as possible CF pathogens [1,2]. Our group reported the first isolation of KPC-2-producing K. pneumoniae from two Brazilian patients with CF patients pulmonary exacerbations. However, no KPC-producing strains grew from subsequent sputum cultures [2]. More recently, the establishment of a chronic lung infection in a patient with CF with a colistin-resistant KPC-3-producing K. pneumoniae was reported [1]. The isolates were persistently isolated from sputum cultures collected from the patient for at least 6 consecutive months. In contrast with previous reports, ours is the first case in which blaKPC-2 and qnrB genes were detected in E. cloacae from a patient with CF. This strain was recovered only once in the sputum cultures during an exacerbation episode.

Although the isolation of *Enterobacteriaceae* species may not be associated with disease severity in patients with CF, the detection of *E. cloacae* carrying a conjugative plasmid harbouring the bla_{KPC} gene highlights the possibility of the spread TABLE 1. Phenotypic and molecular characterization of a QnrB-I and KPC-2 Enterobacter cloacae strain 12615 isolated from a brazilian CF patient and its transconjugant

	E. cloacae 12615		TC-12615- bla _{KPC} , qnrB		J53 Escherichia coli (recipient)	
Specimen Resistance genesª	Sputum bla _{KPC-2}		– bla _{KPC} , g	ınrB	– NA	
Ū.	qnrB-		KFC, 1			
MIC (mg/L) ^b [category]						
Amikacin	8	[S]	≤2	[S]	≤2	[S]
Cefepime	2	[1]	≤2 ≤I	[S]	≤2 ≤1 ≤4 ≤1 ≤1 4	[S]
Cefoxitin	≥64	[R]	≤4 ≤ ≤	[S]	≤4	[S]
Ceftazidime	16	[R]	≤ 1	[S]	≤ 1	[S]
Ceftriaxone	≥64	[R]	≤ 1	[S]	≤ 1	[S]
Cefuroxime	≥64	[R]	≥64	[R]		[S]
Ciprofloxacin	1	[S]	\leq 0.25	[S]	\leq 0.25	[S]
Colistin	\leq 0.5	[S]	\leq 0.5	[S]	≤0.125	[S]
Gentamycin	≤ 1	[S]	≤ 1	[S]	≤ 1	[S]
Imipenem	\geq I6	[R]	8	[R]	0.25	[S]
Meropenem	≥16 2	[R]	4	[R]	≤0.25	[S]
Tigecycline	2	[1]	\leq 0,25	[S]	≤0.25	[S]
Piperacillin/tazobactam	\geq I 28	[R]	\geq I28	[R]	≤4	[S]

¹Resistances genes for clinical sample were detected by PCR and sequencing. For transconjugant strain, standard PCR was used.
²Antimicrobial susceptibility tests were interpreted according to CLSI criteria, except for tigecycline and polymyxins, which were interpreted according to European Committee on Antimicrobial Susceptibility Testing guidelines.

of carbapenem resistance among different microorganisms, such as P. aeruginosa, in infected lungs. The presence of qnr genes can also be a determining factor for the selection of additional chromosome-borne mechanisms of resistance to quinolones after treatment by these antibiotics. The qnr-positive isolates may be a favourable background for an in vivo selection of additional chromosome-borne mechanisms of resistance to quinolones after treatment by fluoroquinolones. The qnr genes appear to be highly promiscuous, having the capacity to become rapidly disseminated among related and unrelated hosts. The transmissibility of the qnr genes makes the genomic mechanisms facilitating their movement of considerable interest and of relevance in the community and in healthcare settings. The detection of qnrB-1 and bla_{KPC-2} plasmid genes in E. cloacae isolated from a patient with CF highlights the importance of screening resistance genes in CF microbiology to aid in the infection control measures adopted for these patients.

Conflict of interest

The authors declare that they have no conflict of interest.

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