

James N. Fullerton, Alastair J. O'Brien, and Derek W. Gilroy

Centre for Clinical Pharmacology, Division of Medicine, Rayne Institute, 5 University Street, University College London, London, WC1E 6JF, UK

Open access under CC BY license.

Sepsis, trauma, burns, and major surgical procedures activate common systemic inflammatory pathways. Nosocomial infection, organ failure, and mortality in this patient population are associated with a quantitatively different reprioritization of the circulating leukocyte transcriptome to the initial inflammatory insult, greater in both magnitude and duration, and secondary to multiple observed defects in innate and adaptive immune function. Dysregulation of inflammatory resolution processes and associated bioactive lipid mediators (LMs) mechanistically contribute to this phenotype. Recent data indicate the potential efficacy of therapeutic interventions that either reduce immunosuppressive prostaglandins (PGs) or increase specialized proresolving LMs. Here, we reassess the potential for pharmacological manipulation of these LMs as therapeutic approaches for the treatment of critical illness (CI).

Inflammation unites CI

Systemic inflammation is nearly ubiquitous in CI, uniting the leading causes of intensive care (ICU) admission [1]. Induced by infectious and noninfectious stimuli with approximately equal frequency [2], the host inflammatory reaction is driven by common mediators and shared signaling pathways [3,4]. Critically ill patients are now understood to experience a highly coordinated, reproducible response at the transcriptomic, metabolomic, and proteomic level, regardless of the inflammatory source [5–7].

Adverse clinical outcomes are associated with a quantitative dysregulation of the inflammatory profile in both magnitude and duration [7]. Predominance and prolongation of anti-inflammatory processes mechanistically contribute to multiple defects in the innate and adaptive immune system, and consequent vulnerability to nosocomial [hospital-acquired infection (HAI)] [8,9] and CI-induced immune dysfunction (CIIID) [10]. In total, ~30% of CI patients will contract HAI; a rate six times greater than on standard wards [11,12]. HAI in this setting is associated with a

 $K\!eywords:$ eicosanoids; prostaglandins; cyclooxygenase; resolution of inflammation; sepsis; immunosuppression.

1471-4906

^{*}This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited. twofold greater inpatient mortality risk and a case mortality in excess of 50% [11,13]. Causative pathogens are most commonly bacterial; however, fungal infections (particularly *Candida* spp.) are increasing in incidence [12]. Viral infection, especially reactivation, poses an additional risk, and polymicrobial infection is common.

LMs, including eicosanoids and the more recently discovered 'specialized pro-resolution lipid mediators' (SPMs), are key signaling molecules in the resolution of inflammation, playing a pivotal role in regulating the inflammatory profile and promoting return to homeostasis [14]. Their dysregulation in any of several dimensions may have pathogenic consequences (Box 1), with failure of resolution leading to chronic inflammation and excess tissue damage being best recognized [14,15]. The potential active contribution of LMs to an anti-inflammatory, immunosuppressive phenotype has, until recently, received little attention [10].

Observational studies have related LM-modifying aspirin and statin administration, as well as nonsteroidal antiinflammatory drugs (NSAIDs), to clinical benefit in the CI population (Box 2) [16], and compelling clinical data support the use of targeted immunomodulatory therapy [17,18]. This information, coupled with a more advanced appreciation of how inflammatory resolution pathways, and LMs in particular, may affect immune competence (and be used to modify it) in CI, necessitates a reappraisal of the clinical validity of these drugs.

Here, we summarize contemporary preclinical and clinical data describing the effects of modulating LMs in CI syndromes, predominantly sepsis. We describe the pathogenic contribution of two distinct yet interlinked patterns of dysregulation to CIIID, namely injurious and failed inflammatory resolution, and discuss therapeutic opportunities presented by LM manipulation.

CI: a failure of resolution?

 ω -3-Derived SPMs from different series appear to have individually separate yet collectively beneficial effects on multiple modalities of immune function. Evidence indicates that a paucity of these LMs contributes to derangement of the inflammatory profile and CIIID, with therapeutic replacement restoring or augmenting immune function. In the next sections we discuss data relating to specific LMs of the resolvin (Rv) and protectin series, and later the lipoxin (LX) and leukotriene (LT) families in interventional animal models of infection/inflammation.

Defining features of SPM bioaction include the ability to: (i) counter-regulate mediators that summon leukocytes,



Corresponding author: Fullerton, J.N. (james.fullerton@doctors.org.uk).

^{© 2013} The Authors. Published by Elsevier Ltd. All rights reserved. http://dx.doi.org/ 10.1016/j.it.2013.10.008

Box 1. LMs: background and pathogenic potential

AA (20:6, ω -6), docosahexaenoic acid (22:6, ω -3) and eicosapentaenoic acid (20:5, ω -3) are polyunsaturated fatty acids that form the substrates for the enzymatic generation of several groups of bioactive LMs. Eicosanoids – the generic term for a group of bioactive lipids containing 20 carbons derived from AA – are divided into several separate mediator families, the major groups being the PGs, LTs and LXs. More recently discovered ω -3-derived SPM families include Rvs, protectins, and maresins. LMs represent vital endogenous biochemical determinants of inflammatory kinetics and the principle mediators of resolution.

The ability of NSAIDs to reduce the primary symptoms and signs of inflammation via COX inhibition and hence PG suppression has led to a common assumption that they, and in turn eicosanoids in general, are universally proinflammatory. This represents a grossly simplified view. These lipid mediators are variably constitutive and inducible, expressed widely yet in a cell-type- and tissue-specific manner, and their actions are diverse, multifaceted, and vary down to the receptor level. Individual molecules have been shown to variably exert pro- and anti-inflammatory effects along with proresolution properties in a context-dependent manner.

Given their immunomodulatory potency and diversity of action, for an effective and self-limited inflammatory reaction to be facilitated, eicosanoid generation must be localized, balanced, proportionate, and timely. Disturbance in any of these dimensions in isolation, or more likely combination, may contribute adversely to disease states. Several pathogenic aberrations may be hypothesized:

- 1) Location of action.
- Compartment leakage or altered distribution of generation. Endocrine as opposed to typical autocrine or paracrine activity.
 Increased or decreased concentration.
 - Altered synthesis, via host, pathogen or iatrogenic intervention, through modulation of substrate or enzymatic process.
 - Promotion or loss of catabolism (local or systemic) or the failure of feedback loops. Altered bioavailability or protein binding (e.g., albumin).
- 3) Deranged temporal profile of production.
 - Failure or dysregulation of eicosanoid class switching [64,65].
- 4) Up- or downregulation of receptors, alteration in receptor profile, including distribution.
- Modification of action (e.g., co-stimulation additive, synergistic, or anergic) by other stimuli/mediators in the surrounding inflammatory milieu.

In broad terms, two key patterns may result from the above. First, deficient or failed resolution where either an insufficient concentration of LM are available to facilitate inflammatory termination, or their action is inadequate. This has been recently discussed in varying inflammatory conditions [14]. Second, a state we describe as injurious resolution may exist. Here, an excessive immunoregulatory effect is exerted by eicosanoids involved in the initiation or control of resolution, rendering host defenses locally or systemically compromised [10].

in particular polymorphonuclear cells (PMNs, neutrophils), to an inflamed site; (ii) dampen pain; (iii) stimulate nonphlogistic monocyte recruitment; and (iv) activate macrophages to efferocytose apoptotic granulocytes and clear both pathogens and tissue debris [14]. Despite being part of the endogenous anti-inflammatory process via action (i), with associated prevention of inflammatory amplification, it is attributes (iii) and (iv) in tandem with promotion of phagocyte trafficking to lymph nodes [19] that distinguishes them from classical anti-inflammatory mediators such as interleukin (IL)-10 or IL-1 receptor antagonist [14].

SPMs have repeatedly been demonstrated to lack an immunosuppressive action, and indeed to augment host-directed antimicrobial defenses [20]. These molecules

Box 2. Observational and mechanistic data supporting therapeutic LM manipulation

Regular use of aspirin in patients who develop community-acquired pneumonia has been associated with lower ICU requirement (odds ratio 0.19, 95% confidence interval 0.04–0.87) and shorter in-patient stay (13.9±6.2 vs. 18.2±10.2 days) [86]. In a general CI patient set prior prescription and consumption of aspirin and statins has been linked with reduced severity of illness (development of severe sepsis, acute lung injury, or adult respiratory distress syndrome) and mortality in a multivariate analysis [83]. Aspirin administration within 24 h of SIRS recognition has been separately linked with a significant decrease in mortality in all such ICU patients of -6.2% (absolute risk difference after propensity matching), and an even greater mortality reduction in those with proven sepsis of -14.8% (27.4% aspirin vs. 42.2% no aspirin) [84]. Independently, a near 50% reduction in the risk of inpatient mortality in septic patients given aspirin during their ICU admission has been described [88], with similar levels of benefits potentially resulting from utilization of alternative NSAIDs in addition (ibuprofen, diclofenac, or indomethacin) [85]. NSAID benefit is however lost if coadministered with aspirin [85], and concerns regarding delayed presentation [89] and side effects persist [84]. The cause of clinical improvement is likely multifactorial and suggested to be secondary to antithrombotic (antiplatelet), anti-inflammatory effects and augmentation of inflammatory resolution pathways [16,87]. This is re-enforced via positive interactions with statins [83] and other antiplatelet agents [86,87].

Mechanistically, aspirin and statins exert their immunomodulatory actions through increased synthesis of bioactive SPMs from all three PUFAs (Figure 3). Both may modify COX-2 – aspirin via serine acetylation, and statins via cysteine *S*-nitrosylation – to generate 15R-hydroxyeicosatetraenoic acid, which is subsequently converted by 5-LOX into 15-epi-LXA₄ (aspirin-triggered LX, ATL). Aspirin may in addition upregulate expression of the ATL receptor ALX (FPRL1) [42], and promote the generation of aspirin-triggered Rvs (17*R*epimers) and protectins (e.g., aspirin-triggered protectin D1). Glucocorticoids have also been demonstrated to increase SPM generation [79]. The effects of these molecules in models of CI are discussed in the main text.

stimulate mucosal production of bactericidal peptides [21] and enhance bacterial phagocytosis by PMNs and macrophages, working synergistically with antibiotics, to increase their therapeutic action and hence bacterial clearance [22]. They have further been shown to suppress nuclear viral mRNA transcript export, and hence replication, reducing mortality from influenza infection [23]: a potentially novel therapeutic addition to standard antivirals, focused on modifying host immune capability, avoiding the problems posed by these infectious agents diversity, variability and capacity to evolve.

Rvs and protectins

RvE1 administered prior to a murine model of aspiration pneumonia (hydrochloric acid with subsequent *Escherichia coli* challenge) was associated with a reduction in proinflammatory cytokines, decreased pulmonary PMN accumulation, enhanced bacterial clearance, and improved survival [24]. El Kebir and colleagues have further described the ability of RvE1 to promote resolution of established infective and sterile models of murine lung injury [25]. Mechanistically, RvE1 was noted to enhance NADPH-oxidase reactive oxygen species generation and promote phagocytosis-induced neutrophil apoptosis (with subsequent efferocytosis by macrophages) via the LTB₄ receptor BLT1. Increased activation of caspase-8 and caspase-3 in tandem with attenuation of both extracellular signal-regulated kinase (ERK)

Trends in Immunology January 2014, Vol. 35, No. 1

and Akt-mediated apoptosis-suppressing signals shifting the balance of pro-/anti-survival information toward apoptosis via induction of mitochondrial dysfunction. In addition, RvE1, at concentrations as low as 1 nM, enhances macrophage phagocytosis, with the products of its metabolism continuing to exert pro-resolution properties but with reduced bioactivity *in vivo* [26].

RvD1 pretreatment prior to lipopolysaccharide (LPS)induced acute lung injury is protective, improving pathological changes and survival [27]. The central mechanism appears to be suppression of nuclear factor (NF)-kB activation in a partly peroxisome proliferator-activated receptor (PPAR)_y-dependent manner, with associated reduction in downstream signaling/transcriptomic alteration [28]. RvD2, but not its isomer trans-RvD2, has been shown specifically to improve survival in murine polymicrobial sepsis (cecal-ligation and puncture; CLP). Its actions appear multifaceted – modulating leukocyte-endothelium interactions in a direct (adhesion receptor expression) and indirect manner (endothelial NO production), altering the cytokine profile [reduced IL-17, IL-10, PGE₂ and LTB₄], and enhancing bacterial phagocytosis and intraphysosomal vacuolar production of reactive oxygen species [20]. More recently, the ability of RvD2 to restore neutrophil directionality, prevent CIIID, and thus increase survival from a secondary septic challenge post-burn injury has been demonstrated [29].

Discrete specialized pro-resolution mediators are unlikely to be produced in isolation and have overlapping proresolving actions. RvE1, aspirin-triggered (ATL, 15-epi-lipoxin A₄) and protectin D1 may independently rescue cyclo-oxygenase (COX)- and lipoxygenase (LOX)-derived 'resolution deficits' in vitro and in vivo, with actions extending to promotion of phagocyte trafficking away from the primary inflammatory site [19]. The ability to bind and act as agonists on alternate SPM receptors (e.g., RvD1 on the LXA₄ receptor [27]) may provide one pharmacological explanation for this phenomenon. However, despite their common actions the source of different classes of SPMs in inflammation appears diverse. Recent evidence suggests that RvE1 and 2 are synthesized by PMNs via the 5-LOX pathway [30], whereas eosinophils are responsible for generation of 12/15-LOX-derived mediators protectin D1 and the newly discovered RvE3 [31,32]. Deficiency of these cell types in the resolution phase may lead to impaired biosynthesis with deleterious consequences [31]. The same may be true of polyunsaturated fatty acids at the inflammatory site.

Experimentally, the ω -3 Rv precursors eicosapentaenoic and docosahexaenoic acids have been demonstrated to increase in exudates during the resolution phase; being both plasma (partially bound to leaked albumin) and locally derived [14,33]. Indirect evidence to support the therapeutic benefit of increasing this SPM series concentration in humans comes from the addition of fish oils to parenteral nutrition in septic patients. A raised plasma eicosapentaenoic concentration was observed along with modification of the cytokine profile, and small physiological and clinical benefits in a recent randomized clinical trial [34].

LTs and LXs

Therapeutic use of the arachidonic acid (AA)-derived LX series may also be beneficial. Post-insult treatment with

LXA₄ has been demonstrated to limit inhaled LPS-induced lung injury [35], and to reduce pro- and anti-inflammatory cytokine production, enhance macrophage recruitment, reduce blood bacterial load, and improve mortality in a rat CLP model [36]. In this later study, macrophage recruitment was increased without impairing phagocytic function, and systemic inflammation reduced without increasing bacterial spread, mirroring the previously described observations with other SPMs [20,24].

A similar paradoxical relation between an attenuated innate immune response (PMN trafficking to the infected site), yet efficacy of the overall inflammatory process, as determined by survival, has been demonstrated in both wild type mice treated with MK 886 (a 5-LOX inhibitor) and in 5-LOX-deficient mice [37]. This effect could be partially replicated if antagonists of cysteinyl-LTs (a family including LTC- F_4) were given, but crucially not with antagonism of the classically proinflammatory LTB₄. This elegantly demonstrates the hierarchical, multifaceted and often opposing effects of eicosanoids in sepsis. In this setting it appears that the prevention of the cysteinyl-LTs deleterious effects on the vasculature, hence host hemodynamics, assumes primacy as the main cause of benefit in 5-LOX antagonism. By contrast, selective LTB₄ inhibition prior to and post-CLP appears to have little effect on vascular tone and permeability, but may blunt the innate immune response - specifically neutrophil trafficking – exacerbating the infective insult [38,39].

The complex interplay between AA-derived LMs in sepsis has also been highlighted in recent data showing that flavocoxid, a dual COX-2 and 5-LOX inhibitor, reduces the expression of NF- κ B, COX-2, and 5-LOX with improved survival in a murine CLP model [40]. Plasma IL-10 and LXA₄ concentrations were increased while tumor necrosis factor (TNF)- α , IL-6, LTB₄, and PGE₂ were decreased. Whether the observed improvement in outcome was due to enhanced pro-resolution effects driven by increased LXA₄, decreased cytokine storm (TNF- α and IL-6), augmentation of the immune response via reducing PGE₂ and 5-LOX derived LTs (discussed below), selective shunting of AA down the COX or LOX pathways, or a combination of the above is unclear.

Separately, biologically active ATL and aspirin-triggered Rvs have been noted to have pro-resolution effects, inhibiting leukocyte trafficking in a NO-dependent manner in both murine [41] and human [42] inflammation, and downregulating superoxide production in neutrophils along with macrophage inflammatory peptide 2 and IL- 1β production [43]. The importance of ATL and LXA₄ inhibitory stimulation has been previously demonstrated in CI; its absence leads to unbridled inflammation and elevated mortality in animal models of infection due to dendritic cell (DC) hypersensitivity [44].

Thus, accumulating evidence supports the notion that SPMs are both necessary to the host immune response and beneficial in severe inflammatory states. Increasing LX, Rv, and protectin concentrations augments host defense and improves survival in preclinical models of CI via multimodally enhancing innate immune function and ameliorating CIIID. The immunomodulatory actions of SPMs on inflammatory resolution processes are summarized in Figure 1.



Figure 1. Immunomodulatory effects of two broad classes of lipid mediators throughout the inflammatory response. Persistence or excessive levels of cyclooxygenasederived prostanoids and leukotrienes due to a failure to class-switch may contribute to injurious resolution. Here, mediators designed to promote inflammatory resolution multimodally drive an immunosuppressive, anti-inflammatory phenotype leading to susceptibility to secondary nosocomial infections (highlighted in red). Predominantly ω -3 polyunsaturated fatty acid lipoxygenase-derived specialized proresolving lipid mediators nonphlogistically augment key processes in inflammatory resolution. An absence or deficit of these eicosanoids may lead to chronic inflammation, failure of pathogen clearance, and both local or distal tissue damage.

Contribution of excessive, injurious resolution pathways

The majority of the interventional studies described above have used a paradigm of substrate supplementation, direct molecule addition, or aspirin-augmented biosynthesis to highlight the benefits of SPMs in CI. Their pathogenic contribution to severe inflammatory states thus appears to be one of absence or insufficiency. Conversely, excess (absolute or relative) or persistence of early-phase resolution-regulating LMs, specifically prostanoids, may be equally deleterious to host immune function: a state of 'injurious resolution' [10]. The negative immunomodulatory effects of PGE_2 [45], PGD_2 [46], and the cyclopentenone PGs of the J_2 series [47] have recently been reviewed and are not described in detail here. Indications of their role in CI is consequently derived from the reverse methodology their reduction or antagonism. Specifically 'two-hit' inflammatory/infective insult models may be of particular relevance to studying CIIID [48].

Severe inflammatory stress in mice is associated with a predominantly anti-inflammatory state and relative immunosuppression following the initial proinflammatory response. This is characterized by increased susceptibility to and worse outcomes from infection [48–50], mimicking clinical observations [51]. Vulnerability to secondary infective challenge (replicating HAI) is dependent on time postinitial insult, decreasing with increased temporal separation [49,50], and may be reduced by the rapeutic strategies designed to restore immune function [49]. PGE_2 appears to be a key mediator of this phenomenon. Bronchoalveolar lavage fluid in mice subjected to pulmonary Aspergillus fumigatus conidia challenge post-CLP contains higher amounts of PGE₂, the production of which seems dependent upon both alveolar macrophages and epithelial cells [52]. Treatment with ketoprofen, a nonselective COX-inhibitor, after CLP, but prior to fungal challenge, reduced the PGE₂ concentration by >95%, enhanced neutrophil recruitment, macrophage phagocytosis and proinflammatory cytokine secretion, leading to a fourfold increase in survival. The Eprostanoid (EP)4 receptor in particular was described as playing a pivotal role [52].

These data support previous work also demonstrating the efficacy of COX-2 inhibition (NS-398) in attenuating prolonged, excessive PGE₂ production secondary to an alternate severe inflammatory stressor (trauma/hemorrhage), with resultant reduction in mortality from subsequent infective challenge [53,54]. In addition to EP4, a PGE₂–EP2–cAMP axis also appears to contribute to reduced effector cell function, with both specific EP2 receptor antagonists or EP2-deficient mice preventing PGE₂-induced impairment of both Fc γ -receptor-dependent phagocytosis and NADPH-oxidase-mediated bacterial killing [55,56].

The temporally defined window of immunosuppression is matched by sequential alterations in COX-2 expression and PGE₂ synthesis. Higher circulating plasma concentrations of PGE₂ are observed on both Days 1 and 7 postsevere inflammatory stress compared to controls, indicating exaggerated and prolonged prostanoid production [54]. Correspondingly, splenic macrophages have increased COX-2 mRNA induction in response to LPS stimulation even at Day 7 [53,54]. Increased PGE₂ concentrations may alternatively stem from selective induction of microsomal PGE₂ synthase (mPGES)-1 by CI states independent of COX-2 expression [52]. In comparison, peritoneal neutrophils from burn-injured animals have been reported to exhibit a late-phase decrease in COX-2 expression and PGE₂ synthesis, coupled with a lack of induction to secondary infective challenge [57], likely indicating compartmentalization of the inflammatory response [58,59].

AA release and its COX-mediated metabolism are also modified by CI in humans. In patients with fracture or burn injury, peripheral blood mononuclear cells exhibit increased COX-2 mRNA and PGE₂ synthesis in response to LPS stimulation [60]. By contrast, COX-2 gene expression in septic patients is reduced such that both basal circulating concentrations of its metabolites (including PGE_2), and those induced by LPS stimulation of blood leukocytes ex vivo, were lower than in healthy controls [61]. Furthermore, the degree of AA-metabolism derangement was associated with disease severity (greater in septic shock vs. severe sepsis), and failure of its recovery between admission to ICU and Day 3 post-admission was predictive of adverse clinical outcome (prolonged admission or death). The authors speculate that reduced prostanoid generation forms part of the anti-inflammatory CIIID phenotype. In attempting to reconcile these data, we propose two alternative but interlinked explanations: (i) that altered AA metabolism and PGE_2 generation in the blood compartment represents an adaptive change to prevent systemic inflammation [58,62]; or (ii) a response to excess production extravascularly (i.e., the primary infective/inflammatory site). Up- and downregulation of prostanoid receptors has been observed in CI humans [60], along with alterations in COX-2 and mPGES-1 expression [61], indicating a dynamic, responsive system. Late-phase immunosuppressive mediators are thought to arise from tissue-resident macrophages in vivo [63]. Chronic release into the circulation and endocrine action initiated by the primary event may be expected to induce compensatory adaptions in blood leukocytes.

The above data indicate that, contrary to popular conceptions, COX-derived eicosanoids, and in particular PGE₂, have significant anti-inflammatory and immunosuppressive effects. In both animal and human studies, CI induces alteration of prostanoid production. Sustained or excessive production of these resolution-regulating eicosanoids may be pathogenic through increasing susceptibility to secondary infection. The efficacy displayed by COX inhibition in clinical observational studies and either COX inhibition or PGE₂ antagonism in preclinical work warrants further investigation. The immunomodulatory actions of COX-derived eicosanoids are summarized in Figure 1.

A single or dual defect?

It is unlikely that either an excess of immunosuppressive eicosanoids or a deficiency of specialized pro-resolution mediators exists independently in CI; their production being inextricably interlinked. Over the course of the inflammatory response, LM profiles undergo a 'class switch' from initial-phase COX-derived PGs and LTs to specialized pro-resolution mediators of the LX, Rv and protectin series, with PGE₂ playing a controlling role [64,65]. In particular, there is now evidence to suggest that different phagocytic cell types and their subpopulations display specific eicosanoid profile signatures that are dynamically altered at defined intervals throughout inflammation, influenced by the surrounding milieu and ingested material [66].

Recent data demonstrate that severe inflammatory stress undermines these normally tightly regulated resolution programs. In a murine peritonitis model contrasting low-dose, self-resolving inflammation with high-dose, nonresolving inflammation (10 mg vs. 1 mg zymosan), sustained high amounts of PGE_2 and LTB_4 (>5× normal) were reported in the high-dose exudates, along with persistently compromised specialized pro-resolution mediator production (LXA₄, protectin D1 and Rvs \sim 3-fold less than normal concentrations) [67]. Alterations in miRNA expression, specifically miR-219-2, and subsequent target gene expression were implicated. This situation appears analogous to the dysregulated transcriptomic and inflammatory response associated with adverse outcomes in CI [7]. Humans display different inflammatory response profiles to set stimuli that are largely determined by resolution processes and mediators, broad categories of 'resolution phenotype' being established preclinically [68], and tentatively in various clinical settings [69,70]. We speculate that inflammatory stress of sufficient magnitude in individuals rendered susceptible via their resolution phenotype may trigger both higher and persistent amounts of initial-phase eicosanoids with concomitant failure of specialized proresolution mediator generation, leading to both injurious and failed resolution (Figure 2). Such dysregulation of LM synthesis may both exacerbate the acute systemic inflammatory response syndrome (SIRS) and subsequently contribute mechanistically to late-phase CIIID [10].

Harnessing therapeutic opportunities

The recognition of aberrant inflammatory resolution pathways as a contributory factor to the CI clinical phenotype opens several translational therapeutic avenues (Figures 2 and 3).



Figure 2. Proposed dysregulation of resolution-phase lipid mediators in critical illness. Solid lines indicate the normal inflammatory profile, with an early rise in cyclooxygenase (COX)-derived prostaglandins (PGs) and 5-lipoxygenase-derived leukotriene B_4 , which trigger a subsequent rise in specialized proresolving lipid mediators (SPMs) including lipoxins, resolvins, and protectins. Dashed lines display the altered profile of eicosanoids in critical illness-induced immune dysfunction, with persistence of early-phase lipid mediators that exert negative immunomodulatory effects and a paucity or relative insufficiency of SPMs, which nonphlogistically augment several key resolution pathways including bacterial clearance. The combination of this dual defect contributes to vulnerability to hospital-acquired infection. Identification of aberrant eicosanoid profiles in critically ill patients may allow their therapeutic correction or antagonism to ameliorate effector cell functional impairment as indicated by the dotted lines.

The first derives from the principle of non-maleficence: cessation of, or alteration in, the use of existing therapies and/or practices that impair the resolution phase, or cause its dysregulation. Candidate drugs may be identified via resolution indices (for example [19]) in standardized models. Widely used drugs in anesthetics and critical care have already been implicated [71], and the use of alternate agents or the same drugs at appropriate phases of inflammation may be advocated. As an example, NSAIDs may adversely blunt the initial inflammatory response through impairment of proinflammatory mediator generation, but may hypothetically be used to ameliorate the late immunosuppressive contribution of COX-derived prostanoids, especially PGE₂, to CIIID [52–54,72].

The potential for both immunostimulatory and immunosuppressive agents in the management of CI in general, and sepsis in particular, is well recognized and multiple potential targets have been identified (see [73] for recent review). Real-time immunological assays to determine host immune competence and guide immunomodulatory therapeutics have already shown promise in both experimental [74] and clinical [17,18] studies. As already discussed, interindividual variance in inflammatory response to a given stimulus may partly be attributed to expression of pro-resolution processes [68]. Eicosanoid serum concentrations, *in-vitro/ex-vivo* functional assays and flow cytometry may be used to stratify patients into those who will benefit from targeted interventions to correct resolution aberrancies: a form of individualized medicine [75]. Evidence that such a strategy may be effective is already available: a subgroup of patients in the major randomized controlled trial of ibuprofen in sepsis who exhibited exaggerated prostanoid production displayed reduced mortality secondary to COX inhibition (Box 3) [76].

The administration of SPMs or their precursors, as either replacement for a lack of endogenous production for instance, from a failure to 'class switch' [64,67], or as a supplement to host-generated mediators [20] in the face of an acute increase in catabolism [26], offers one strategy to combat failure of inflammatory resolution. Oral administration of eicosapentaenoic and docosahexaenoic acid have already been shown to increase plasma levels of SPMs to clinically relevant levels [77]. Alternatively, the use of drugs that act as resolution agonists may prove advantageous (Figure 3). In particular, there is compelling evidence that aspirin [42], statins [78], and potentially glucocorticoids [79] increase the generation of SPMs or their equally bioactive epimeric forms to augment aspects of innate immune function and promote the resolution of inflammation (Figure 1).



Figure 3. Potential therapeutic interventions to modify resolution defects and improve innate effector cell functionality in critical illness. Targets on the left of the diagram (red) describe means of reducing excessive or prolonged production of immunosuppressive COX-generated PGs or antagonizing their action. Targets on the right (blue) indicate means of supplementing and/or augmenting levels of SPMs that nonphlogistically enhance multiple effector modalities. AA, arachidonic acid; ATL, aspirin-triggered lipoxin (15-epi-LXA₄); COX, cyclooxygenase; EP2/4, E prostanoid 2/4 receptor; LOX, lipoxygenase; LT, leukotriene; LX, lipoxin; PD, protectin; PG, prostaglandin; PPAR, peroxisome proliferator-activated receptor; RV, resolvin; SPM, specialized proresolving lipid mediator.

The concentrations of mediators achieved at the inflammatory site will be key to the efficacy of these therapeutic strategies. As Ricciotti and FitzGerald observe 'it is one thing to show that putative pro-resolution products can be formed *in vitro* and that the synthetic compounds do exert pro-resolution actions when administered in vivo and another to document that the concentrations formed *in vivo* in the setting of inflammation are sufficient and necessary to mediate resolution' [80]. Recent reports describing the isolation of high levels of SPMs in discrete human biological samples that follow predictable, modifiable, temporal profiles however partially allay these concerns [77,81]. Data suggesting SPMs may be exploited by pathogens in supraphysiological levels to induce a survival advantage (Box 4) indicates that attempts to exploit their therapeutic potential must not inadvertently induce a state of injurious resolution. Further caveats are discussed below.

The inhibition or antagonism of early-phase eicosanoids contributing to CIIID, the corollary and complementary strategy of ameliorating injurious resolution (Figure 2), may yet prove more tractable. As the above discussion has indicated, a growing body of evidence suggests that PGE₂, PGD₂, 15d-PGJ₂, and LTB₄ may be viable targets, whether through inhibition of enzymatic synthesis, enhancement of catabolism (e.g., via albumin), receptor antagonism, or monoclonal antibody therapy. Translational potential will be predicated upon description of both local and systemic eicosanoid profiles induced by severe inflammatory stress, establishment of robust and generalizable resolution indices in health and disease to gauge pathogenic dysfunction, and the trialing of pharmacological agents capable of selectively modifying PG and LT concentrations in relevant compartments.

Immunomodulatory drugs offer two clear advantages over current management of CIIID and subsequent HAI. The first is the ability to prevent infections prophylactically. Corrective therapy may be administered throughout ICU admissions to achieve a minimum baseline immune functional status, preventing new invasive pathogens from gaining traction. The second is their ability to support bacterial killing in an antibiotic-independent manner via augmenting innate immune function. Antibiotic resistance is one of the greatest challenges facing humans and poses a particular problem in the ICU patient cohort. Strategies to both therapeutically enhance pro-resolution activity [22] and reduce prostanoids-driven injurious resolution [82]

Box 3. History of COX inhibition in Cl

The concept of using COX inhibition in the setting of CI predominantly sepsis - is not new; the first experiments being performed in the 1960s utilizing aspirin. Since then a vast amount of data has accumulated. Although animal work has been overwhelmingly positive, COX inhibition pre- or post-septic challenge improving survival in >70% (31/43) of publications and being detrimental in only 6.9% (3/43) [72], this success has not been replicated in humans. The largest randomized controlled trial showed no significant improvement in sepsis-related mortality (37% ibuprofen vs. 40% placebo) despite demonstrable prostanoid suppression and improvement in physiological variables [90]. Two further smaller studies also showed no effect on mortality (ibuprofen [91] and lornoxicam [92]). All trials have however supported the safety of NSAID use in CI and fears that COX inhibition via aspirin or NSAIDs may predispose to severe sepsis have not been substantiated [89].

Why then has this preclinical promise not been fulfilled? Many of the standard difficulties of ICU and sepsis trials apply to these studies. Investigated populations were heterogeneous in age, infection site, causative pathogen, and comorbidity burden, producing a high noise-to-signal ratio. Enrollment over a range of timepoints post acquirement of infection was inevitable, preventing standardized intervention in the same phase of disease and potentially missing a narrow therapeutic window where adverse outcomes may be averted. Trial design was imperfect, the major randomized controlled trial being underpowered, aiming to detect an unrealistic 35% difference in mortality [16]. Most importantly, we now understand that the rationale behind the therapeutic strategy may have been flawed. NSAID therapy was directed at the proinflammatory phase, aiming to ameliorate SIRS not ClIID, and given to a diverse patient set in a nonstratified manner [73].

Re-evaluation of the previously described publications supports this view. A greater beneficial therapeutic effect (survival) of NSAIDs was observed in animal models involving live pathogens than in those where pathogen-associated molecular patterns (PAMPs; e.g., LPS) were administered alone [72]. This differential and enhanced efficacy may be explained by suppression of immunosuppressive eicosanoids and consequent augmentation of innate immune function, as opposed to abrogation of their negative physiological effects, which would be equivalent in both live pathogen and PAMP models. Furthermore, subgroup analysis of the primary randomized controlled trial showed a significant improvement in mortality from hypothermic sepsis (54% vs. 90%) in those treated with ibuprofen [90]. This presentation was associated with an exaggerated prostanoid response on admission to the study [76]; a suggestion that had laboratory-based testing been used to target therapy at a more immunologically homogeneous population, a beneficial signal could have been detected.

Pharmaceutical trial evidence targeting the inflammatory cascade in Cl has been instructive largely through its failure. Novel immunomodulatory approaches aiming to reduce late-phase mortality in stratified patient populations have already demonstrated promise [17,18,75]. Discarding NSAIDs – a relatively safe, cheap, and potentially efficacious class of drugs – based on limited data seems naïve. Further investigation may yet establish therapeutic eicosanoid manipulation as part of an immune-restorative arsenal in Cl.

have recently demonstrated preclinical success in lowering antibiotic requirements and/or increasing their efficacy. It is currently unknown if these will translate into the clinical setting.

Concluding remarks

An excessive dysregulated inflammatory response is associated with adverse clinical outcomes in CI patients. Resolution-regulating lipid mediators contribute to this phenomenon being mechanistically involved in CIIID via both sustained production of early-phase prostanoids, (in

Box 4. Pathogen modulation of LM profiles

The exploitation of LM properties by multiple pathogens underscores their biological potency and the potential pathogenicity of their dysregulation.

Both Toxoplasma gondii and Pseudomonas aeruginosa have been shown to utilize their independently generated 15-LOX variants on host substrate (AA) to create a microenvironment with supraphysiological concentrations of LXs, blocking DC activation and preventing their phagocytosis [93,94]. Mycobacterium tuberculosis (MTB) has been demonstrated to support replication and immune evasion via a similar yet distinct mechanism. Protective circulating levels of (anti-inflammatory) LXs appear to be generated via MTB stimulation of host biosynthesis (as opposed to via pathogen-generated enzymes). 5-LOX-deficient mice consequently displayed enhanced T helper (Th)1 responses and lower bacterial burden than wild type controls; a protective phenotype abolished by administration of a stable LXA₄ analog [95]. Other authors have demonstrated the opposite of this effect - diminished Th1 responses (lower IL-12 and γ interferon) and consequent survival in tuberculosis-infected mice treated with MK 886 [96]. It seems likely that both early inhibition of pro-inflammatory LTB4 and subsequent excessive LXA₄ may be pathogenic in this setting. This demonstration of the importance of both pro- and anti-inflammatory LOX products in mounting a successful defense against invading pathogens has recently been elegantly supported by work in zebrafish [97].

A further eicosanoid contributor to the pathogen-host interaction is PGE₂. PGE₂ production triggered by virulent but not avirulent MTB has been implicated as a pathogen-defense mechanism, with PGE synthase^{-/-} mice having significantly higher bacterial burden post-challenge as a result of a shift in macrophage fate profile [98]. In addition, the fungi *Candida albicans* and *Cryptococcus neoformans* secrete PGE₂ [99] as a virulence mechanism [100]. In this context, PGE₂-mediated inhibition of the transcription factor interferon regulatory factor (IRF)4, with resultant decrease in Th17 cell IL-17 production during cell differentiation was implicated. Blockade of host PGE₂ in mice infected with *C. neoformans* led to increased survival [100].

particular PGE_2) and reduced or insufficient late-phase SPMs.

Resolution therapies, rectifying LM profiles, are predicted to restore innate immune function, reducing susceptibility to HAI and its sequelae. To be proven clinically effective they will have to avoid the pitfalls that have befallen multiple pharmacological agents in CI, including previous trials of NSAIDs in sepsis (Box 3). Patients will have to be stratified and appropriate therapy delivered according to real-time assessments of inflammatory status (pro vs. anti), immune functional defects, and causative mechanisms. Therapy will ideally be targeted at specific biochemical pathways (e.g., PGE₂-EP2/4 blockade as opposed to nonspecific COX-inhibition), and guided by quantifiable outcomes [17] in order to reduces off-target effects and minimize potential toxicity. Equally, a 'magic bullet' should not be expected. LM manipulation may form one part of multimodal immunomodulatory therapy, working synergistically with other agents. Finally, diverse demographic, clinical, and microbiological considerations may also modify treatment decisions. Although these stipulations ensure drug development will undoubtedly represent a substantial challenge, the chances of developing an effective therapy are significantly higher as a result. The potential clinical benefit suggested by observational data provides sufficient justification to pursue this strategy [83-88].

Acknowledgments

J.N.F. and D.W.G. are in receipt of funding from the Wellcome Trust.

References

- 1 Wunsch, H. et al. (2011) Comparison of medical admissions to intensive care units in the United States and United Kingdom. Am. J. Respir. Crit. Care Med. 183, 1666–1673
- 2 Dulhunty, J.M. *et al.* (2008) Does severe non-infectious SIRS differ from severe sepsis? Results from a multi-centre Australian and New Zealand intensive care unit study. *Intensive Care Med.* 34, 1654–1661
- 3 Xu, J. et al. (2009) Extracellular histones are major mediators of death in sepsis. Nat. Med. 15, 1318–1321
- 4 Zhang, Q. et al. (2010) Circulating mitochondrial DAMPs cause inflammatory responses to injury. *Nature* 464, 104–107
- 5 Langley, R.J. et al. (2013) An integrated clinico-metabolomic model improves prediction of death in sepsis. Sci. Transl. Med. 5, 195ra195
- 6 Tang, B.M. et al. (2008) Gene-expression profiling of gram-positive and gram-negative sepsis in critically ill patients. Crit. Care Med. 36, 1125–1128
- 7 Xiao, W. et al. (2011) A genomic storm in critically injured humans. J. Exp. Med. 208, 2581–2590
- 8 Perl, M. et al. (2006) Contribution of anti-inflammatory/immune suppressive processes to the pathology of sepsis. Front. Biosci. 11, 272–299
- 9 Hotchkiss, R.S. *et al.* (2013) Immunosuppression in sepsis: a novel understanding of the disorder and a new therapeutic approach. *Lancet Infect. Dis.* 13, 260–268
- 10 Fullerton, J.N. et al. (2013) Pathways mediating resolution of inflammation: when enough is too much. J. Pathol. 231, 8–20
- 11 Burgmann, H. et al. (2010) Impact of nosocomial infections on clinical outcome and resource consumption in critically ill patients. Intensive Care Med. 36, 1597–1601
- 12 Vincent, J.L. (2003) Nosocomial infections in adult intensive-care units. Lancet 361, 2068–2077
- 13 Alberti, C. et al. (2002) Epidemiology of sepsis and infection in ICU patients from an international multicentre cohort study. Intensive Care Med. 28, 108–121
- 14 Serhan, C.N. (2010) Novel lipid mediators and resolution mechanisms in acute inflammation: to resolve or not? Am. J. Pathol. 177, 1576– 1591
- 15 Nathan, C. and Ding, A. (2010) Nonresolving inflammation. *Cell* 140, 871–882
- 16 Eisen, D.P. (2012) Manifold beneficial effects of acetyl salicylic acid and nonsteroidal anti-inflammatory drugs on sepsis. *Intensive Care* Med. 38, 1249–1257
- 17 Meisel, C. et al. (2009) Granulocyte-macrophage colony-stimulating factor to reverse sepsis-associated immunosuppression: a doubleblind, randomized, placebo-controlled multicenter trial. Am. J. Respir. Crit. Care Med. 180, 640–648
- 18 Hall, M.W. et al. (2011) Immunoparalysis and nosocomial infection in children with multiple organ dysfunction syndrome. *Intensive Care* Med. 37, 525–532
- 19 Schwab, J.M. et al. (2007) Resolvin E1 and protectin D1 activate inflammation-resolution programmes. Nature 447, 869–874
- 20 Spite, M. et al. (2009) Resolvin D2 is a potent regulator of leukocytes and controls microbial sepsis. Nature 461, 1287–1291
- 21 Canny, G. et al. (2002) Lipid mediator-induced expression of bactericidal/permeability-increasing protein (BPI) in human mucosal epithelia. Proc. Natl. Acad. Sci. U.S.A. 99, 3902–3907
- 22 Chiang, N. et al. (2012) Infection regulates pro-resolving mediators that lower antibiotic requirements. Nature 484, 524–528
- 23 Morita, M. et al. (2013) The lipid mediator protectin D1 inhibits influenza virus replication and improves severe influenza. Cell 153, 112–125
- 24 Seki, H. et al. (2010) The anti-inflammatory and proresolving mediator resolvin E1 protects mice from bacterial pneumonia and acute lung injury. J. Immunol. 184, 836–843
- 25 El Kebir, D. et al. (2012) Resolvin E1 promotes phagocytosis-induced neutrophil apoptosis and accelerates resolution of pulmonary inflammation. Proc. Natl. Acad. Sci. U.S.A. 109, 14983–14988
- 26 Hong, S. et al. (2008) Resolvin E1 metabolome in local inactivation during inflammation-resolution. J. Immunol. 180, 3512–3519

- 27 Wang, B. et al. (2011) Resolvin D1 protects mice from LPS-induced acute lung injury. Pulm. Pharmacol. Ther. 24, 434–441
- 28 Liao, Z. *et al.* (2012) Resolvin D1 attenuates inflammation in lipopolysaccharide-induced acute lung injury through a process involving the PPARgamma/NF-kappaB pathway. *Respir. Res.* 13, 110
- 29 Kurihara, T. et al. (2013) Resolvin D2 restores neutrophil directionality and improves survival after burns. FASEB J. 27, 2270–2281
- **30** Tjonahen, E. *et al.* (2006) Resolvin E2: identification and antiinflammatory actions: pivotal role of human 5-lipoxygenase in resolvin E series biosynthesis. *Chem. Biol.* 13, 1193–1202
- 31 Yamada, T. et al. (2011) Eosinophils promote resolution of acute peritonitis by producing proresolving mediators in mice. FASEB J. 25, 561–568
- 32 Isobe, Y. et al. (2012) Identification and structure determination of novel anti-inflammatory mediator resolvin E3, 17,18dihydroxyeicosapentaenoic acid. J. Biol. Chem. 287, 10525–10534
- 33 Kasuga, K. et al. (2008) Rapid appearance of resolvin precursors in inflammatory exudates: novel mechanisms in resolution. J. Immunol. 181, 8677–8687
- 34 Barbosa, V.M. et al. (2010) Effects of a fish oil containing lipid emulsion on plasma phospholipid fatty acids, inflammatory markers, and clinical outcomes in septic patients: a randomized, controlled clinical trial. Crit. Care 14, R5
- 35 Jin, S.W. et al. (2007) Posttreatment with aspirin-triggered lipoxin A4 analog attenuates lipopolysaccharide-induced acute lung injury in mice: the role of heme oxygenase-1. Anesth. Analg. 104, 369-377
- 36 Walker, J. et al. (2011) Lipoxin a4 increases survival by decreasing systemic inflammation and bacterial load in sepsis. Shock 36, 410–416
- 37 Benjamim, C.F. et al. (2005) Opposing and hierarchical roles of leukotrienes in local innate immune versus vascular responses in a model of sepsis. J. Immunol. 174, 1616–1620
- 38 Rios-Santos, F. et al. (2003) A critical role of leukotriene B4 in neutrophil migration to infectious focus in cecal ligaton and puncture sepsis. Shock 19, 61-65
- 39 Lammermann, T. et al. (2013) Neutrophil swarms require LTB4 and integrins at sites of cell death in vivo. Nature 498, 371–375
- **40** Bitto, A. *et al.* (2012) Flavocoxid, a dual inhibitor of COX-2 and 5-LOX of natural origin, attenuates the inflammatory response and protects mice from sepsis. *Crit. Care* 16, R32
- 41 Paul-Clark, M.J. et al. (2004) 15-epi-lipoxin A4-mediated induction of nitric oxide explains how aspirin inhibits acute inflammation. J. Exp. Med. 200, 69–78
- 42 Morris, T. et al. (2009) Effects of low-dose aspirin on acute inflammatory responses in humans. J. Immunol. 183, 2089–2096
- 43 Hachicha, M. et al. (1999) Lipoxin (LX)A4 and aspirin-triggered 15epi-LXA4 inhibit tumor necrosis factor 1alpha-initiated neutrophil responses and trafficking: regulators of a cytokine-chemokine axis. J. Exp. Med. 189, 1923–1930
- 44 Machado, F.S. et al. (2006) Anti-inflammatory actions of lipoxin A4 and aspirin-triggered lipoxin are SOCS-2 dependent. Nat. Med. 12, 330–334
- 45 Kalinski, P. (2012) Regulation of immune responses by prostaglandin E2. J. Immunol. 188, 21–28
- 46 Joo, M. and Sadikot, R.T. (2012) PGD synthase and PGD2 in immune resposne. *Mediators Inflamm.* 2012, 503128
- 47 Scher, J.U. and Pillinger, M.H. (2009) The anti-inflammatory effects of prostaglandins. J. Investig. Med. 57, 703–708
- 48 Muenzer, J.T. *et al.* (2006) Pneumonia after cecal ligation and puncture: a clinically relevant "two-hit" model of sepsis. *Shock* 26, 565–570
- 49 Muenzer, J.T. et al. (2010) Characterization and modulation of the immunosuppressive phase of sepsis. Infect. Immun. 78, 1582–1592
- 50 Davis, C.G. et al. (2011) Increased susceptibility to Candida infection following cecal ligation and puncture. Biochem. Biophys. Res. Commun. 414, 37–43
- 51 Boomer, J.S. et al. (2011) Immunosuppression in patients who die of sepsis and multiple organ failure. JAMA 306, 2594–2605
- 52 Brogliato, A.R. et al. (2012) Ketoprofen impairs immunosuppression induced by severe sepsis and reveals an important role for prostaglandin E2. Shock 38, 620–629
- 53 Strong, V.E. et al. (2000) Blocking prostaglandin E2 after trauma attenuates pro-inflammatory cytokines and improves survival. Shock 14, 374–379

- 54 Mack Strong, V.E. et al. (2001) NS-398 treatment after trauma modifies NF-kappaB activation and improves survival. J. Surg. Res. 98, 40–46
- 55 Medeiros, A.I. et al. (2009) Efferocytosis impairs pulmonary macrophage and lung antibacterial function via PGE2/EP2 signaling. J. Exp. Med. 206, 61–68
- 56 Serezani, C.H. et al. (2007) Prostaglandin E2 suppresses bacterial killing in alveolar macrophages by inhibiting NADPH oxidase. Am. J. Respir. Cell Mol. Biol. 37, 562–570
- 57 He, L.K. et al. (2001) The expression of cyclooxygenase and the production of prostaglandin E2 in neutrophils after burn injury and infection. J. Burn Care Rehabil. 22, 58-64
- 58 Cavaillon, J.M. and Annane, D. (2006) Compartmentalization of the inflammatory response in sepsis and SIRS. J. Endotoxin Res. 12, 151–170
- 59 Medeiros, A. et al. (2012) Prostaglandin E2 and the suppression of phagocyte innate immune responses in different organs. Mediat. Inflamm. 2012, 327568
- 60 Strong, V.E. et al. (2001) Prostaglandin E2 receptors EP2 and EP4 are down-regulated in human mononuclear cells after injury. Surgery 130, 249–255
- 61 Bruegel, M. et al. (2012) Sepsis-associated changes of the arachidonic acid metabolism and their diagnostic potential in septic patients. Crit. Care Med. 40, 1478–1486
- 62 Munford, R.S. and Pugin, J. (2001) Normal responses to injury prevent systemic inflammation and can be immunosuppressive. *Am. J. Respir. Crit. Care Med.* 163, 316–321
- 63 Kox, M. et al. (2011) Differential ex vivo and in vivo endotoxin tolerance kinetics following human endotoxemia. Crit. Care Med. 39, 1866–1870
- 64 Levy, B.D. et al. (2001) Lipid mediator class switching during acute inflammation: signals in resolution. Nat. Immunol. 2, 612–619
- 65 Serhan, C.N. and Savill, J. (2005) Resolution of inflammation: the beginning programs the end. Nat. Immunol. 6, 1191–1197
- 66 Dalli, J. and Serhan, C.N. (2012) Specific lipid mediator signatures of human phagocytes: microparticles stimulate macrophage efferocytosis and pro-resolving mediators. *Blood* 120, e60–e72
- 67 Fredman, G. et al. (2012) Self-limited versus delayed resolution of acute inflammation: temporal regulation of pro-resolving mediators and microRNA. Sci. Rep. 2, 639
- 68 Morris, T. et al. (2010) Dichotomy in duration and severity of acute inflammatory responses in humans arising from differentially expressed proresolution pathways. Proc. Natl. Acad. Sci. U.S.A. 107, 8842–8847
- 69 Wu, S.H. et al. (2009) Inverse temporal changes of lipoxin A4 and leukotrienes in children with Henoch–Schonlein purpura. Prostaglandins Leukot. Essent. Fatty Acids 80, 177–183
- 70 Pillai, P.S. et al. (2012) Chemical mediators of inflammation and resolution in post-operative abdominal aortic aneurysm patients. *Inflammation* 35, 98–113
- 71 Chiang, N. et al. (2008) Anesthetics impact the resolution of inflammation. PLoS ONE 3, e1879
- 72 Aronoff, D.M. (2012) Cyclooxygenase inhibition in sepsis: is there life after death? *Mediat. Inflamm.* 2012, 696897
- 73 Larosa, S.P. and Opal, S.M. (2012) Immune aspects of sepsis and hope for new therapeutics. *Curr. Infect. Dis. Rep.* 14, 474–483
- 74 Osuchowski, M.F. et al. (2009) Stratification is the key: inflammatory biomarkers accurately direct immunomodulatory therapy in experimental sepsis. Crit. Care Med. 37, 1567–1573
- 75 Venet, F. et al. (2013) Monitoring the immune response in sepsis: a rational approach to administration of immunoadjuvant therapies. *Curr. Opin. Immunol.* 25, 477–483
- 76 Arons, M.M. et al. (1999) Effects of ibuprofen on the physiology and survival of hypothermic sepsis. Ibuprofen in Sepsis Study Group. Crit. Care Med. 27, 699–707
- 77 Mas, E. et al. (2012) Resolvins D1, D2, and other mediators of selflimited resolution of inflammation in human blood following n-3 fatty acid supplementation. Clin. Chem. 58, 1476–1484

- 78 Ye, Y. et al. (2008) Phosphorylation of 5-lipoxygenase at ser523 by protein kinase A determines whether pioglitazone and atorvastatin induce proinflammatory leukotriene B4 or anti-inflammatory 15-epilipoxin a4 production. J. Immunol. 181, 3515–3523
- 79 Perretti, M. et al. (2002) Endogenous lipid- and peptide-derived anti-inflammatory pathways generated with glucocorticoid and aspirin treatment activate the lipoxin A4 receptor. Nat. Med. 8, 1296-1302
- 80 Ricciotti, E. and FitzGerald, G.A. (2011) Prostaglandins and inflammation. Arterioscler. Thromb. Vasc. Biol. 31, 986–1000
- 81 Weiss, G.A. et al. (2013) High levels of anti-inflammatory and proresolving lipid mediators lipoxins and resolvins and declining docosahexaenoic acid levels in human milk during the first month of lactation. *Lipids Health Dis.* 12, 89
- 82 Stables, M.J. et al. (2010) Priming innate immune responses to infection by cyclooxygenase inhibition kills antibiotic-susceptible and -resistant bacteria. Blood 116, 2950–2959
- 83 O'Neal, H.R., Jr et al. (2011) Prehospital statin and aspirin use and the prevalence of severe sepsis and acute lung injury/acute respiratory distress syndrome. Crit. Care Med. 39, 1343–1350
- 84 Eisen, D.P. et al. (2012) Acetyl salicylic acid usage and mortality in critically ill patients with the systemic inflammatory response syndrome and sepsis. Crit. Care Med. 40, 1761–1767
- 85 Sossdorf, M. et al. (2013) Benefit of low-dose aspirin and non-steroidal anti-inflammatory drugs in septic patients. Crit. Care 17, 402
- 86 Winning, J. et al. (2009) Anti-platelet drugs and outcome in severe infection: clinical impact and underlying mechanisms. Platelets 20, 50–57
- 87 Winning, J. et al. (2010) Antiplatelet drugs and outcome in mixed admissions to an intensive care unit. Crit. Care Med. 38, 32–37
- 88 Otto, G.P. et al. (2012) Effects of low-dose acetylsalicylic acid and atherosclerotic vascular diseases on the outcome in patients with severe sepsis or septic shock. *Platelets* 24, 480–485
- 89 Legras, A. et al. (2009) A multicentre case-control study of nonsteroidal anti-inflammatory drugs as a risk factor for severe sepsis and septic shock. Crit. Care 13, R43
- 90 Bernard, G.R. et al. (1997) The effects of ibuprofen on the physiology and survival of patients with sepsis. The Ibuprofen in Sepsis Study Group. N. Engl. J. Med. 336, 912–918
- 91 Haupt, M.T. et al. (1991) Effect of ibuprofen in patients with severe sepsis: a randomized, double-blind, multicenter study. The Ibuprofen Study Group. Crit. Care Med. 19, 1339–1347
- 92 Memis, D. et al. (2004) Effects of lornoxicam on the physiology of severe sepsis. Crit. Care 8, R474–R482
- 93 Bannenberg, G.L. et al. (2004) Exogenous pathogen and plant 15lipoxygenase initiate endogenous lipoxin A4 biosynthesis. J. Exp. Med. 199, 515-523
- 94 Vance, R.E. et al. (2004) The opportunistic pathogen Pseudomonas aeruginosa carries a secretable arachidonate 15-lipoxygenase. Proc. Natl. Acad. Sci. U.S.A. 101, 2135-2139
- 95 Bafica, A. et al. (2005) Host control of Mycobacterium tuberculosis is regulated by 5-lipoxygenase-dependent lipoxin production. J. Clin. Invest. 115, 1601–1606
- 96 Peres, C.M. et al. (2007) Inhibition of leukotriene biosynthesis abrogates the host control of Mycobacterium tuberculosis. Microbes Infect. 9, 483–489
- 97 Tobin, D.M. et al. (2010) The lta4h locus modulates susceptibility to mycobacterial infection in zebrafish and humans. Cell 140, 717–730
- 98 Chen, M. et al. (2008) Lipid mediators in innate immunity against tuberculosis: opposing roles of PGE2 and LXA4 in the induction of macrophage death. J. Exp. Med. 205, 2791–2801
- 99 Noverr, M.C. et al. (2001) Pathogenic yeasts Cryptococcus neoformans and Candida albicans produce immunomodulatory prostaglandins. Infect. Immun. 69, 2957–2963
- 100 Valdez, P.A. et al. (2012) Prostaglandin E2 suppresses antifungal immunity by inhibiting interferon regulatory factor 4 function and interleukin-17 expression in T cells. *Immunity* 36, 668–679