

RESEARCH ARTICLE

Open Access

Brucella ceti infection in dolphins from the Western Mediterranean sea

Marcos Isidoro-Ayza^{1†}, Nazareth Ruiz-Villalobos^{2†}, Lola Pérez¹, Caterina Guzmán-Verri², Pilar M Muñoz³, Fernando Alegre⁴, Montserrat Barberán³, Carlos Chacón-Díaz^{2,5}, Esteban Chaves-Olarte^{2,5}, Rocio González-Barrientos⁶, Edgardo Moreno^{2,7}, José María Blasco³ and Mariano Domingo^{1,8*}

Abstract

Background: *Brucella ceti* infections have been increasingly reported in cetaceans. Brucellosis in these animals is associated with meningoencephalitis, abortion, discospondylitis, subcutaneous abscesses, endometritis and other pathological conditions. *B. ceti* infections have been frequently described in dolphins from both, the Atlantic and Pacific Oceans. In the Mediterranean Sea, only two reports have been made: one from the Italian Tyrrhenian Sea and the other from the Adriatic Sea.

Results: We describe the clinical and pathological features of three cases of *B. ceti* infections in three dolphins stranded in the Mediterranean Catalanian coast. One striped dolphin had neurobrucellosis, showing lethargy, incoordination and lateral swimming due to meningoencephalitis, A *B. ceti* infected bottlenose dolphin had discospondylitis, and another striped dolphin did not show clinical signs or lesions related to *Brucella* infection. A detailed characterization of the three *B. ceti* isolates was performed by bacteriological, molecular, protein and fatty acid analyses.

Conclusions: All the *B. ceti* strains originating from Mediterranean dolphins cluster together in a distinct phylogenetic clade, close to that formed by *B. ceti* isolates from dolphins inhabiting the Atlantic Ocean. Our study confirms the severity of pathological signs in stranded dolphins and the relevance of *B. ceti* as a pathogen in the Mediterranean Sea.

Keywords: *Brucella ceti*, Cetacean, Neurobrucellosis, Discospondylitis, Mediterranean sea

Background

After the first descriptions of *Brucella* infections in dolphins and seals and the definition of *Brucella ceti* and *Brucella pinnipedialis* as two new species within the genus [1-3], there has been an increasing recognition of brucellosis in marine mammals (see [4] and [5] for recent reviews). *Brucella* strains from marine mammal origin have been isolated from humans [6]. Antibodies against *Brucella* have been detected in 28 out of 42 cetacean species investigated, and *B. ceti* has been isolated

from 10 of these species [5]. *B. ceti* infection in cetaceans is associated to meningoencephalomyelitis [1,3,7-9], abortion [1,2], discospondylitis, subcutaneous abscesses, endometritis, and a wide range of other pathological conditions [4,5,10]. However, with the exception of the striped dolphin (*Stenella coeruleoalba*) [1,3-5,9-13] the proportion of other cetacean species showing clinicopathological signs associated with brucellosis is low, suggesting that most of these infected animals overcome clinical disease, eventually remaining as *Brucella* carriers and shedders.

Presumptive *Brucella* infections in Western Mediterranean Sea dolphins was first established by serology in two striped dolphins and one bottlenose dolphin (*Tursiops truncatus*) stranded on the Mediterranean Catalanian coast [14]. Recently, *Brucella* strains were isolated from striped dolphins in the Tyrrhenian and Adriatic Seas [11,15]. Here, we describe the clinical and pathological features of three

* Correspondence: mariano.domingo@cresa.uab.cat

†Equal contributors

¹Departament de Sanitat i Anatomia Animal, Facultat de Veterinària, Universitat Autònoma de Barcelona (UAB), and Servei de Diagnòstic de Patologia Veterinària, Facultat de Veterinària, Universitat Autònoma de Barcelona (UAB), 08193, Bellaterra, Barcelona, Spain

⁸Centre de Recerca en Sanitat Animal (CRESA), UAB-IRTA, 08193, Bellaterra, Cerdanyola del Vallès, Barcelona, Spain

Full list of author information is available at the end of the article

cases of brucellosis in dolphins stranded on the Mediterranean Catalanian coast, and provide detailed information on the phenotypic and molecular characterization of these three *B. ceti* isolates.

Methods

Dolphin stranding and serological, pathological and bacteriological examinations

Two striped dolphins (N-372/09, N-301/12) and one bottlenose dolphin (N-275/12) reacting positive in standard Rose Bengal Test (RBT), performed according to standard procedures [16] were included in this study. The relevant animal and stranding data are shown in Table 1. At the time of necropsy all three dolphin carcasses were in a good preservation state (2 in a scale of 1 -very good- to 5 -poor-). Gross pathological studies were performed in the three dolphins and complete sets of organs and tissues for each animal were preserved in 10% buffered formalin and processed for histopathological examination and immunohistochemical detection of Cetacean Morbillivirus (CeMV) as described previously [17]. Selected samples of brain tissue and spleen from the three dolphins were also examined immunohistochemically using an anti-*B. melitensis* 16 M polyclonal rabbit antiserum and avidin-biotin peroxidase system as described elsewhere [7,12].

Tissue samples of two dolphins (N-275/12 and N-301/12) were collected at the time of necropsy and submitted for bacteriological examination (Table 1). Tissues of the third dolphin (*S. coeruleoalba*, N-372/09), frozen at -80°C since 2009, were defrosted and submitted also to bacteriological studies, but cerebrospinal fluid (CSF) was not available in this case. Swabs taken at necropsy were each smeared in at least two plates of both Farrell's [18]

and CITA [19] culture media. The remaining necropsy samples were homogenized under sterile conditions in the minimum amount possible of sterile buffered saline (PBS pH 6.8) in a Stomacher unit (Seward Medical, Worthing, UK), and 0.5 mL of each tissue homogenate seeded also on at least two plates of each selective culture medium. The plates were checked for growth after 5–8 days of incubation at 37°C both in air and 10% CO₂ atmospheres. *Brucella* colonies were identified by colonial morphology and standard typing procedures [20,16]. One culture was considered as positive when at least one *Brucella* colony forming unit (CFU) was isolated. The suspected *Brucella* colonies isolated were further identified and characterized by molecular and chemical methods (see below).

Control strains

The following strains obtained from the CITA and from PIET/CIET strain collections were used as controls for molecular studies: *B. ceti* Atlantic dolphin type (B14/94), *B. ceti* Atlantic porpoise type (B1/94), *B. ceti* Cantabric Sea isolate from *S. coeruleoalba* stranded in Northern Spain (C1), *B. pinnipedialis* seal type (B2/94), *Brucella abortus* 2308 (biovar 1 virulent reference strain), *B. abortus* S19 (biovar 1 reference vaccine strain), *Brucella melitensis* Rev1 (biovar 1 reference vaccine strain), *Brucella suis* (S2 biovar 1), *B. canis* (CR206-10; Costa Rica isolate), *B. neotomae* 5 K33 (reference strain), *Brucella ovis* PA (virulent reference strain) and *Brucella microti* (CCM4915, reference strain).

Molecular studies

Brucella DNA samples from each isolate and control strains were extracted with DNeasy Blood & Tissue kit

Table 1 Biological data of *Brucella ceti* infected Mediterranean dolphins

Identification	Place, coordinates and date of stranding	Pathological diagnoses (Macro/Micro)	Tissues sampled for bacteriological examination	<i>B. ceti</i> strain isolated from
N-372/09 <i>Stenella coeruleoalba</i>	Salou, Spain (41.0733, 1.1343)	Non-suppurative encephalitis by CeMV. RT-PCR and IHC for CeMV both positive only in CNS	Encephalon, spleen, diaphragmatic and preescapular lymph nodes, lung	bmarMR26 Spleen
Male, 1.94 m, 74.5 Kg	Found alive Sept 11th-2009			
N-275/12 <i>Tursiops truncatus</i>	Badalona, Spain (41.4458, 2.2507)	Mycotic pyogranulomatous-necrotizing meningoencephalomyelitis	Encephalon, CSF (swab from lateral ventricle), vertebral abscess (swab), spleen.	bmarMR25 Vertebral abscess
Male 3 m	Found dead 7May 23rd-2012	Chronic, severe, focally extensive, suppurative discospondylitis		
		RT-PCR and IHC for CeMV negative		
N-301/12 <i>Stenella coeruleoalba</i>	Cunit, Spain (41.1931, 1.6360)	Non-suppurative meningoencephalitis. RT-PCR and IHC for CeMV negative	Encephalon, spleen, diaphragmatic lymph node, lung	bmarMR24 Encephalon
Female, 1.84 m, 54.5 Kg	Found alive June 3rd-2012			

from QIAGEN®, and stored at -70°C until used. The three Mediterranean dolphin isolates were identified as *B. ceti* using the multiplex PCR as described elsewhere [21]. DNA samples from these isolates and the marine control strains were also tested by PCR-RFLP of *omp2b* locus [22] and by multiplex PCR using the following two pairs of primers: TCA ACT GCG TGA ACA ATG CT (f) / GCG GGC TCT ATC TCA AGG TC (r), and CGT CAA CTC GCT GGC CAA GAG (f) / GCA GGA GAA CCG CAA CCT AA (r). Multiple loci variable number of tandem repeats (MLVA-16) analysis of *Brucella* species and strains was performed as described previously [23-26]. The basic protocol for MLVA-16 was slightly modified to use DreamTaq™ PCR Master Mix (Fermentas®). Amplicon analysis was performed on the ChemiDoc Gel Documentation System XRS, BioRad® using the Quantity One® software, which allowed molecular size determination of amplicons. *Brucella* control strains were used for validation [12,23,26]. The profiles were entered in the database MLVA-NET for the corresponding analysis [27].

Mass spectrometry analysis of *Brucella* protein extracts

For MALDI-TOF studies, the three *B. ceti* isolates and all control strains were grown in trypticase soy agar plates for four days in the presence or in the absence of CO₂ (as required) following modifications of previous protocols [16,18,28]. For each bacterial strain, three clearly separated colonies were suspended in a 1.5 ml Eppendorf tube containing 1 ml of ultrapure distilled water, and centrifuged at 14,000 rpm for 5 min. Then, the bacterial pellet was thoroughly resuspended in 300 µL of water. After this, 700 µL of absolute ethanol were added, the suspension mixed in a vortex, let rest for five minutes at room temperature and centrifuged at 14,000 rpm for 5 minutes. The bacterial pellet was resuspended in 250 µL of water and directly sonicated in the Eppendorf tube with the aid of a titanium micro tip, at room temperature for 2 minutes. Nine hundred µL of absolute ethanol were added and the extract dried to completeness in a speed-vac centrifuge at 45°C for ~2.5 hours. Fifty µL of 70% formic acid were added to suspend the dried pellet, mixed thoroughly by pipetting and then 50 µL of acetonitrile were added and mixed. The extract was centrifuged at 14,000 rpm for 5 minutes and the supernatant transferred into a clean tube. In order to select optimal conditions for mass spectrometer analysis, several dilutions of the extract were tested. A volume of 0.5 µL of each dilution was spotted onto a steel Opti-TOF 384 plate target (ABSciex) and air-dried at room temperature. The spot sample was overlaid with 0.5 µL of matrix solution (saturated solution of alpha-cyano-4-hydroxy-cinnamic acid) in organic solvent (50% acetonitrile and 2.5% trifluoroacetic acid) and air-dried.

The samples were analyzed in a MALDI-TOF on an Applied Biosystems 4800 Plus mass spectrometer. Spectra were acquired in linear positive mode, using a laser intensity of 3,800 and 500 shots/spectrum, in the m/z range 2,000 to 11,000, after external MS calibration with CalMix-5 standards (ABSciex) spotted on the same plate. Spectra were visualized using Data Explorer v.4.9 (Applied Biosystems).

Gas chromatographic analysis of fatty acid methyl esters

The three *B. ceti* isolates and all control strains were grown as described above. For each bacterial strain, 75 clearly separated colonies were chosen from three plates and placed in a sealed glass tube. Saponification of the samples and processing for total fatty acid methyl ester determination were carried out according to the MIDI instruction manual of Technical Note #101 (MIS, MIDI Inc., Newark, DE). Analysis was performed by gas chromatography (Agilent Technologies 6850) using a 25 m x 0,2 mm cross linked phenyl-methyl silicone fused silica capillary column HP 19091B-102 (Agilent Technologies Inc., Santa Clara, CA). A binary matrix was generated using the fatty acid profile of the tested strains.

Phylogenetic analysis and cladograms

Dendrograms based on the retention time of the fatty acid methyl esters and on the protein masses detected were constructed using an Agglomerative hierarchical clustering (AHC) algorithm, using Microsoft® Excel 2000/XLSTAT®-Pro (Version 4.07, 2013, Addinsoft, Inc., Brooklyn, NY, USA). Proximities were calculated using Squared Euclidean Distance, and aggregation was calculated using the unweighted pair-group average method. MLVA 16 phylogenetic trees based on differences in MLVA-16 was built according to the procedures described in the *Brucella* MLVA database [27].

Results

Clinical and pathological findings

The main biopathological features of the three Mediterranean dolphins from which *B. ceti* strains were isolated are summarized in Table 1.

Striped dolphin N-301/12 was found stranded alive, showing uncoordinated swimming, with circling and severe lateralization, needing continuous support (Figure 1A). This dolphin died four days after stranding, in spite of the supportive medical care. Episodes of tonic-clonic seizures were observed shortly before death. At necropsy, this animal showed a good body condition. No significant gross lesions were observed, apart from a moderate amount of subcutaneous oedema in the cranial region of the trunk, and multiple erosions of the tongue epithelium. A moderate infestation of the caudal blubber by larval forms of the tapeworm *Phyllobothrium delphini* and bile and pancreatic

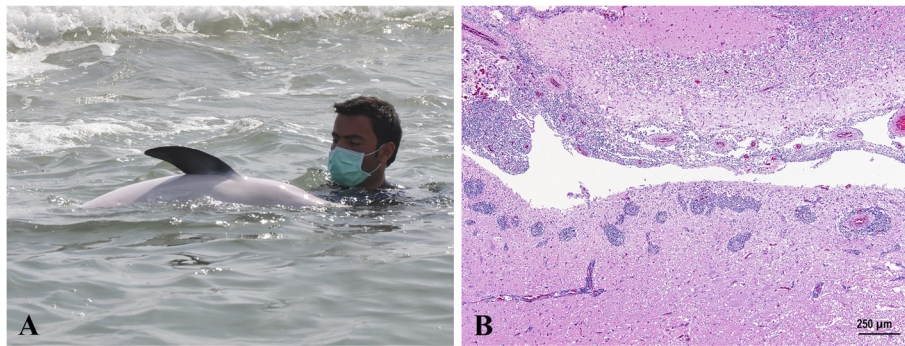


Figure 1 Striped dolphin, case N-301/12. **A.** Dolphin with clinical signs of neurobrucellosis, showing lateralization when swimming and inability to maintain equilibrium and flotation. The dolphin needed continuous holding and support. **B.** Histopathologic section of the brain from the same case. Thick perivascular infiltrates mainly composed of lymphocytes, plasma cells and macrophages are seen affecting leptomeninges, the neuropil surrounding the fourth ventricle and the choroid plexus (H&E).

ducts by trematodes of the genus *Campyla* were observed. These are common parasites of striped dolphins. Histologically, the most relevant finding was a severe, diffuse, chronic, non-suppurative, meningoencephalomyelitis. The inflammatory process affected the leptomeninges, and the subependymal neuropil (periventriculitis), forming thick perivascular cuffs that shallowly penetrated into the underlying grey and white matter (Figure 1B). The process was more severe in cerebellum, brainstem, spinal cord and medulla oblongata and with less involvement of the cerebral cortex. The inflammatory infiltrate was composed of a large number of lymphocytes and plasma cells and a lesser number of macrophages. A similar inflammatory infiltrate was observed in the choroid plexus (choroiditis) as well as in the dorsal and ventral cervical nerve roots (radiculitis). All studied sections of the nervous system showed variable degrees of gliosis, satellitosis with formation of glial nodules, spongiosis and perivascular oedema.

Small, multifocal, randomly distributed, non-suppurative inflammatory infiltrates were found in the liver and kidney. In addition, a small focus of granulomatous and necrotizing lymphadenitis was seen in the mesenteric lymph node. These lesions were probably associated to parasitic migrations. No other relevant microscopical findings were found in examined tissues. *Brucella* antigens were not detected by IHC in CNS. RT-PCR for CeMV resulted negative in all the investigated tissues.

The bottlenose dolphin (N-275/12) was found dead. Gross external examination revealed that this dolphin was in a good body condition. Several "Tattoo'-like lesions measuring 2–4 cm in diameter were observed on the skin of the left maxilla. A hard spherical mass of approximately 18 cm in diameter was found in the caudal peduncle. The skin was unaffected at this site (Figure 2A). On radiographs the mass was characterized as a marked bone proliferation rising from the periosteum of two consecutive coccygeal vertebrae generating ankylosis of

the implicated joint (Figure 2B). A longitudinal section (Figure 2C) showed a chronic, severe, focally extensive and suppurative disco-spondylitis with marked disruption of the adjacent soft tissues (skeletal muscle fascicles, tendons and fascia). Histopathology from this lesion was not performed. In the CNS, a slightly increased amount of CSF was noticed at the lateral ventricles. After pre-fixation, the brain was sliced. At that time, poorly demarcated, bilateral areas of malacia were identified in the area of the *nucleus caudatus*. A third focus of malacia was disclosed in the area of the *nucleus accumbens* of the left hemisphere. Microscopically, there was a pyogranulomatous and necrotizing meningoencephalomyelitis and radiculitis caused by the fungus *Cunninghamella bertholletiae* that has been described in detail elsewhere [29]. No other significant lesions were found at necropsy in this dolphin. Tissues tested by RT-PCR for CeMV yielded negative results. IHC for *Brucella* yielded negative results at CNS. The coccygeal diskospondylitic lesion was not investigated by IHC.

Striped dolphin N-372/09 was found alive showing lethargy, incoordination and lateral swimming. The animal died before the rescue protocol could be initiated. At necropsy, no significant macroscopic lesions were observed in this dolphin, except for the absence of content in the stomach cavities. Moderate infestation by the common larval forms of the tapeworms *Phyllobothrium delphini* and *Monorygma grimaldi* were also observed in the caudal blubber and the peritoneal cavity respectively. Histopathological examination revealed chronic, non-suppurative encephalitis as the main pathological sign. Immunohistochemical (IHC) staining for CeMV detected viral antigen at the Central Nervous System (CNS) of this dolphin. This result was confirmed by a CeMV positive RT-PCR in CNS tissue. Consequently, this case was diagnosed as a CNS-localized chronic CeMV infection [30].

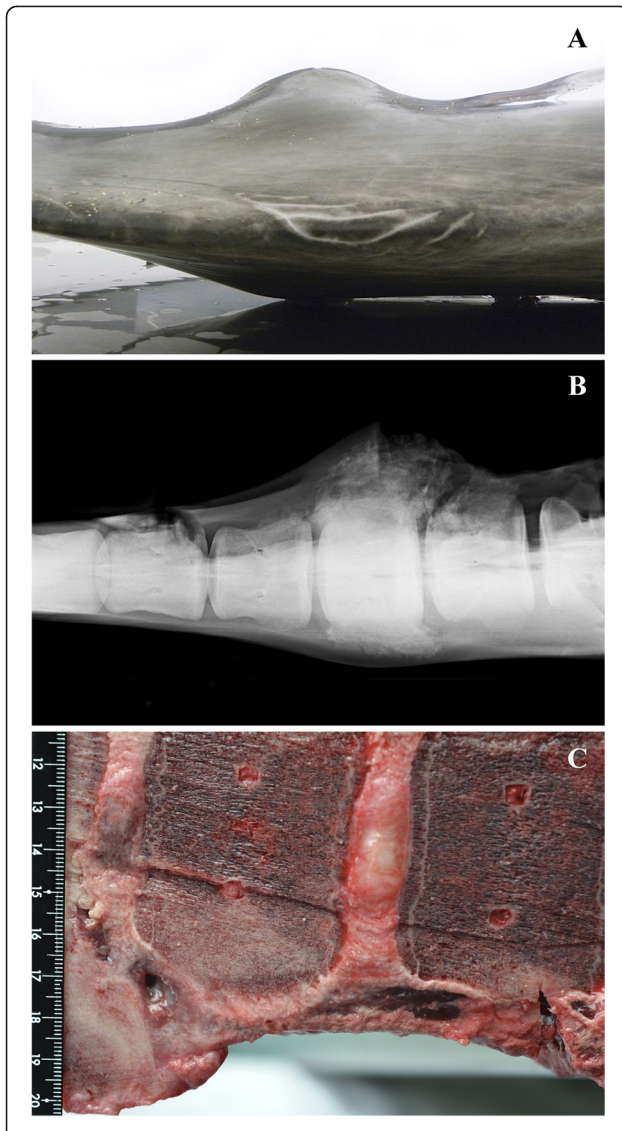


Figure 2 Bottlenose dolphin, case N-275/12. **A.** Mass in the caudal peduncle, dorso-ventral view of the lesion at necropsy. **B.** Radiograph from the lesion in dorso-ventral view, showing periosteal proliferation with ankylosis of coccygeal vertebrae. **C.** Longitudinal section of the affected vertebra, showing bony proliferation at the ventrolateral side of the vertebral body, leading to ankylosis. There is formation of small abscesses. *Brucella ceti* was cultured from one of the abscesses.

IHC for *Brucella* antigens showed positive staining in cytoplasm of cells inside splenic lymphoid follicles (Figure 3).

Bacteriological findings

Bacterial isolates identified as *Brucella* strains were obtained in high numbers from the spleen (N-372/09), vertebral abscess (N-275/12) and brain (N-301/12), respectively. The strains were isolated in the presence and absence of CO₂ and selective culture media with the exception of

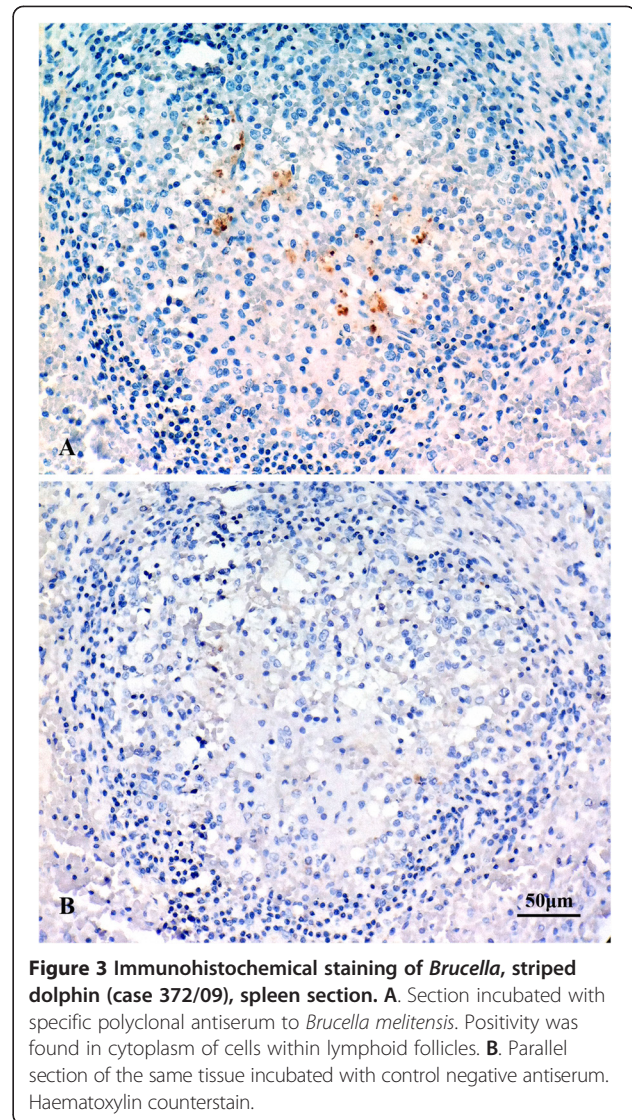


Figure 3 Immunohistochemical staining of *Brucella*, striped dolphin (case 372/09), spleen section. **A.** Section incubated with specific polyclonal antiserum to *Brucella melitensis*. Positivity was found in cytoplasm of cells within lymphoid follicles. **B.** Parallel section of the same tissue incubated with control negative antiserum. Haematoxylin counterstain.

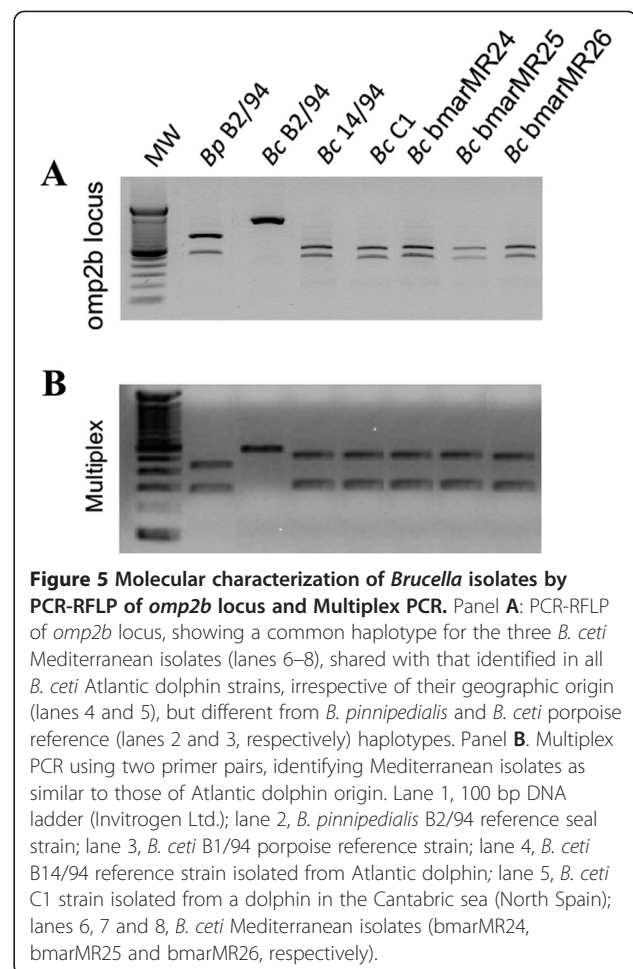
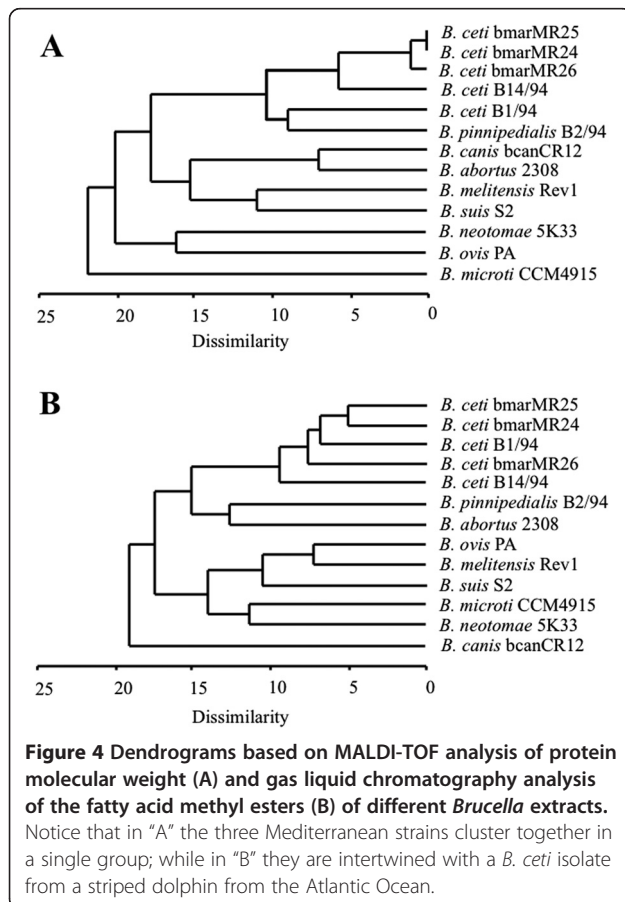
strain from case N-301/12, whose growth was inhibited on Farrell's medium. The isolated strains were not capnophilic, were positive for the oxidase and urease tests, and displayed a smooth type, agglutinating with both anti-A and anti-M mono-specific sera. The strains were as well lysed by the Iz but not the Tb, Wb and R/C phages, and grew on standard concentrations of both thionin and basic fuchsin. The three Mediterranean isolates were identified as *B. ceti* using the multiplex PCR. They were named as bmarMR24 (isolated from case N-301/12), bmarMR25 (from case N-275/12) and bmarMR26 (from case 372/09).

MLVA16, protein and fatty acid polymorphisms and phylogeny

Fatty acid and protein molecular phenotyping are well recognized methods for typing bacteria and have been used extensively to differentiate bacteria, including *Brucella* strains [24,31,32]. The MLVA16 respective sizes for

the analysed strains, the protein molecular weight peaks determined by MALDI-TOF and the retention times of the fatty acid methyl ester peaks determined by GLC of the strains studied are presented respectively in Additional file 1: Tables S1, Additional file 2: Table S2, and Additional file 3: Table S3. Dendrograms built on the basis of MALDI-TOF analysis demonstrated that the three *B. ceti* Mediterranean isolates clustered together in a distinct clade, close to the Atlantic dolphin type B14/94 strain (Figure 4A). However, on the basis of fatty acids (Figure 4B), they clustered with both *B. ceti* porpoise type B1/94 and *B. ceti* Atlantic dolphin type B14/94 strains. In spite of this, all *B. ceti* strains remained as a separate cluster from *B. pinnipedialis* seal B2/94 strain and other *brucellae* from terrestrial animals.

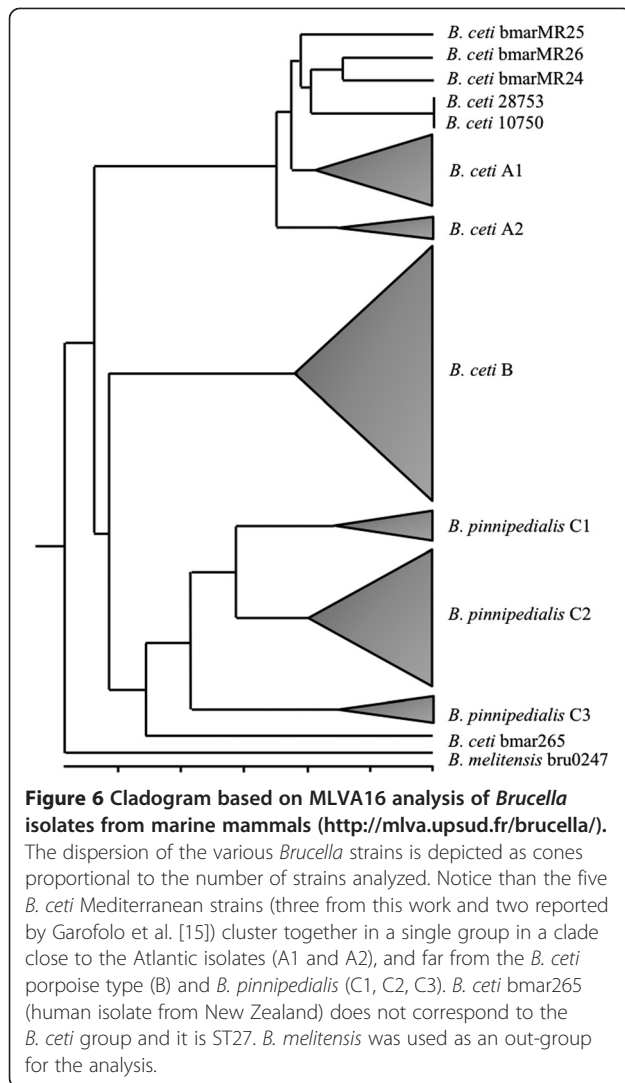
The PCR-RFLP of the *omp2b* locus confirmed also the close relationship of the three Mediterranean isolates of *B. ceti*, resulting in a common haplotype identical in all strains, irrespective of their geographic origin (Figure 5A). Similarly, the multiplex PCR using two primer pairs identified the Mediterranean isolates as similar to the Atlantic dolphin type *B. ceti* B14/94 strain (Figure 5B). The MLVA16 phylogenetic analysis showed that the three *B. ceti* Spanish Mediterranean strains clustered together in a



distinct clade together with the two reported *B. ceti* isolated in the Mediterranean Italian littoral [15] and close to the Atlantic A1 dolphin cluster (Figure 6). The distinct topology of the Mediterranean *B. ceti* isolates is maintained even when a comprehensive phylogenetic analysis against the complete *Brucella* MLVA16 base data is performed (Figure 7). As expected, the *B. ceti* Mediterranean isolates MLVA 16 patterns were identical to strains previously typed by MLSA as ST 26.

Discussion

During the last decade *B. ceti* strains have been isolated from stranded dolphins of both the Atlantic and Pacific Oceans [5]. Information about *B. ceti* strains from the Mediterranean Sea is scarce. Recently, a *B. ceti* strain showing phenotypic characteristics of Atlantic *B. ceti* strains was isolated from a striped dolphin in the Tyrrhenian littoral of Italy [11]. Also, two other *B. ceti* strains have been characterized from the Italian Southern Apulian Coast [15]. MLVA analysis of the two isolates assigned them to a novel genotype within cluster A. Here we confirm and extend these observations, endorsing the



presence of *B. ceti* infecting and causing pathology in at least two dolphin species (*S. coeruleoalba* and *T. truncatus*) inhabiting the Mediterranean Sea. The multilocus sequence analysis of these three *B. ceti* Mediterranean strains indicates that they also belong to the dolphin type of the ST26 cluster. Up to now, all *B. ceti* Mediterranean strains stem in a separate branch from the main MLVA16 A1 and A2 clusters of *B. ceti* isolates from dolphins inhabiting the Atlantic Ocean. Although the number of analysed Mediterranean *B. ceti* strains by MLVA16 is still low to draw a definitive branching order, the taxonomical position of these newly defined strains [11] is supported by proteome analysis and, to a less extent, by fatty acids analysis, two robust techniques used in bacterial taxonomy [33] and useful for characterization of *Brucella* [9-11,24,31,32]. Therefore, it seems that the *B. ceti* isolates described here belong to a particular genotype prevalent in the Mediterranean Sea. The close relationship

between Mediterranean and Atlantic *B. ceti* strains keeps important parallelism with the phylogenetic and taxonomical studies on the striped and bottlenose dolphin populations in both seas. Indeed, there is strong evidence of dispersal of bottlenose dolphin populations between both seas, keeping a population structure and genetic diversity in concordance with the boundaries that coincide with transitions between different habitat regions [34]. In the case of striped dolphins, there is also evidence of some genetic flow between the Mediterranean and Atlantic populations; albeit, this is significantly more restricted than in bottlenose dolphins. This is mainly due to ecological and behavioral factors that limit the exchange between these two *S. coeruleoalba* populations across Gibraltar Strait [35]. Considering this, the phenotypic and genetic structure from both North Atlantic and Mediterranean *B. ceti* isolates is not unexpected, as populations of *S. coeruleoalba* and *T. truncatus* share similar habitats and feed resources, and may then share various microorganisms, including *B. ceti*.

Infection by *B. ceti* is common in cetaceans but only a small proportion of infected cetaceans display clinicopathological signs associated to brucellosis, suggesting that many infected cetaceans overcome infection, perhaps remaining as carriers and potential *Brucella* shedders [5]. This is in sharp contrast with the absence of obvious disease in seals or walrus infected with *B. pinnipedialis* [36]. Several clinico-pathological entities have been associated to *B. ceti* infection in a relevant fraction of the stranded dolphins and porpoises [5], with neurobrucellosis being one of the most significant signs present [8,12,11]. One of our Mediterranean striped dolphins (N-301/12) presented unequivocally *B. ceti* induced non-suppurative meningoencephalitis with *B. ceti* being isolated from the CNS, and regarded as the primary cause of death. This is in agreement with the suggested higher susceptibility of this dolphin species for developing neurobrucellosis in comparison to other cetaceans [12].

B. ceti has been found to invade joints and cause chronic inflammatory lesions, and has been frequently isolated from these lesions in cetaceans [10,12,37]. In line with these observations, a causal relationship was hypothesized between the discospondylitis of the peduncle observed in the bottlenose dolphin case (N-275/12) and the *B. ceti* strain isolated from that lesion. This discospondylitis could be causing a disabling condition, but this lesion was probably not life-threatening. Primary cause of death in this dolphin was attributed to a mycotic encephalitis caused by *Cunninghamella bertholletiae*.

Similarly, death of the other striped dolphin (N-372/09) was linked to CeMV-related encephalitis and not to *B. ceti* infection (isolated retrospectively only from spleen, without evidence of lesions related to *Brucella*). CeMV

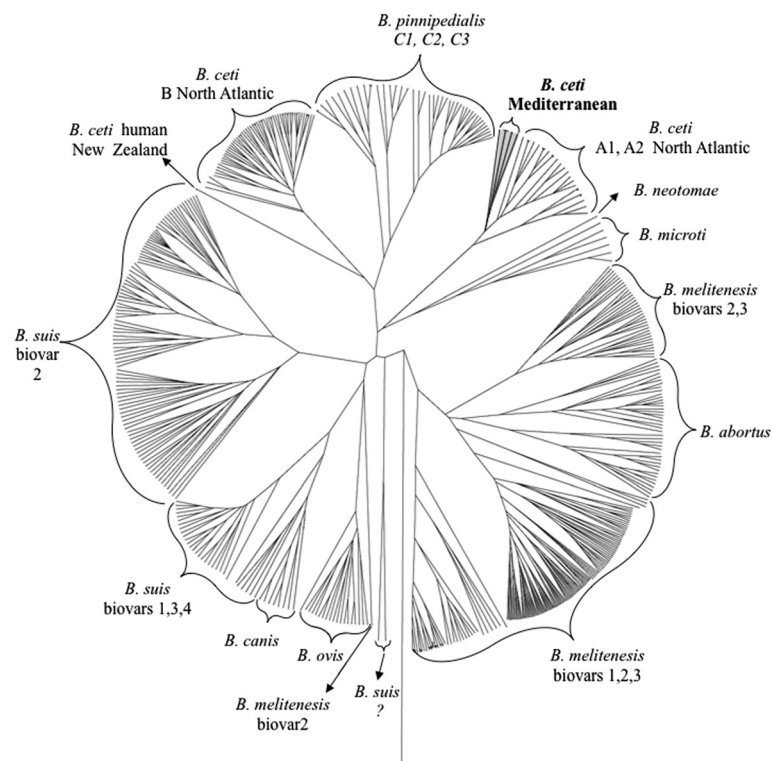


Figure 7 *Brucella* phylogenetic tree based on MLVA16 analysis of all the various *Brucella* species and strains (<http://mlva.upsud.fr/brucella/>). Notice that the three Western (Spanish) and the Eastern (Italian) *B. ceti* Mediterranean strains cluster together in a single group (shaded in gray) in a clade close to the Atlantic isolates (A1 and A2), all belonging to ST 26.

infections of the brain have been unambiguously linked with epizootic disease and deaths of Mediterranean striped dolphins in the past [17,38].

As a corollary to these findings, it appears mandatory to establish adequate differential diagnoses for the aetiological agents that may affect disoriented live-stranded dolphins. This is becoming even more relevant when a number of cetaceans in the Mediterranean Sea show a rapid decline [39], mainly due to by-catch, the presence of pollutant contamination, and the pressure of infectious diseases that threaten the health of free-ranging cetaceans [40]. Brucellosis, as a contagious disease, can be an additional factor hampering the conservation efforts of cetaceans at local and global scale. Our study confirms the relevance of *B. ceti* as a cetacean pathogen in the Mediterranean, the severity of pathological signs in stranded *S. coeruleoalba* dolphins, and gives insight on the phylogenetic structure of these *B. ceti* Mediterranean isolates. The Mediterranean *B. ceti* strains isolated so far form a distinct phylogenetic cluster, close to that of *B. ceti* strains isolated from dolphins inhabiting the Atlantic Ocean. In addition to seriously compromising the wellbeing of marine mammals, the *B. ceti* strains possess all the current molecular virulence

determinants and therefore, are potential pathogens for other animals, including humans [5].

Conclusions

B. ceti has been isolated for the first time from the Spanish Mediterranean Sea, expanding the known range of this species. Neurobrucellosis with non-suppurative meningoencephalomyelitis in a striped dolphin and spondylitis in a bottlenose dolphin were the main clinicopathological features of these cases. In a third case, *B. ceti* was isolated from the spleen of a striped dolphin. The *omp2b* haplotype was common for all three *B. ceti* Mediterranean isolates and multiplex characterization showed that they were similar to the Atlantic dolphin type *B. ceti* B14/94 strain. Following MLVA16 analysis the three *B. ceti* Spanish Mediterranean strains clustered together in a distinct clade with the two reported *B. ceti* isolated in the Mediterranean Italian littorals, and close to the Atlantic A1 dolphin cluster. This taxonomical position was supported by protein and fatty acid analyses. Collecting appropriate samples for testing for *Brucella* has to be included in necropsy protocols in stranded dolphins.

Ethics statement

The work did not include experimental procedures. Handling of live and dead cetaceans (species included in CITES 2 list) was done with official governmental permission. Medical treatments on live dolphins were applied following established procedures for these species.

Additional files

Additional file 1: Table S1. MLVA-16 genetic profiles for the three *B. ceti* isolates*.

Additional file 2: Table S2. Protein molecular weights peaks of the different *Brucella* strains determined by MALDI-TOF.

Additional file 3: Table S3. Fatty acids determined by gas liquid chromatography of different *Brucella* strains.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

FA provided veterinary treatment to live-stranded dolphins. MIA, LP, and MD performed necropsies and histopathologic examinations. LP performed immunostaining and RT-PCR for CeMV. JMB and PMM performed bacterial isolation and species identification. MB and RGB performed immunostaining for *Brucella*. NRV confirmed bacteriological identification of isolates received from Spain and performed MLVA analysis. CGV, CCD and ECO designed, analysed and performed lipidomics and molecular analysis of isolates and reference strains. EM designed, performed and analysed proteomics data, and integrated all results from taxonomic studies. All authors helped to draft specific parts of the manuscript. MD, EM and JMB were responsible of the design of the study and assembled the final draft of the manuscript. All authors approved the final version of the manuscript.

Acknowledgements

We thank Departament de Medi Natural from the Government of Catalonia for economical support. We thank Bruno Lomonte, Instituto Clodomiro Picado, University of Costa Rica for his assistance in the proteomic analysis. Research at the CITA is funded by INIA (RTA2011-00103-00-00), MICIN (AGL2011-30453-C04), and consolidated group A14 from Gobierno de Aragón. This work was partially funded by grants FIDA-2014 UNA, FS-CONARE UNA/UCR. We wish to acknowledge M.J. de Miguel, S. Serrano, M. Uriarte, B. Pérez and A. Neira for the excellent technical assistance.

Author details

¹Departament de Sanitat i Anatomia Animal, Facultat de Veterinària, Universitat Autònoma de Barcelona (UAB), and Servei de Diagnòstic de Patologia Veterinària, Facultat de Veterinària, Universitat Autònoma de Barcelona (UAB), 08193, Bellaterra, Barcelona, Spain. ²Programa de Investigación en Enfermedades Tropicales, Escuela de Medicina Veterinaria, Universidad Nacional, Heredia, Costa Rica. ³CITA Gobierno de Aragón, 50059 Zaragoza, Spain. ⁴Centre Fundació para la Conservació y Recuperación de Animales Marinos (CRAM), El Prat de Llobregat, Barcelona, Spain. ⁵Centro de Investigación en Enfermedades Tropicales, Facultad de Microbiología, Universidad de Costa Rica, San José, Costa Rica. ⁶Ministerio de Agricultura y Ganadería, Laboratorio de Patología, Heredia, Costa Rica. ⁷Instituto Clodomiro Picado, Universidad de Costa Rica, San José, Costa Rica. ⁸Centre de Recerca en Sanitat Animal (CRESA), UAB-IRTA, 08193, Bellaterra, Cerdanyola del Vallès, Barcelona, Spain.

Received: 2 March 2014 Accepted: 28 August 2014

Published online: 17 September 2014

References

1. Ewalt DR, Payeur JB, Martin BM, Cummins DR, Miller WG: **Characteristics of a *Brucella* species from a bottlenose dolphin (*Tursiops truncatus*).** *J Vet Diagn Invest* 1994, **6**:448–452.

2. Foster G, Osterman BS, Godfroid J, Jacques I, Cloeckaert A: ***Brucella ceti* sp. nov. and *Brucella pinnipedialis* sp. nov. for *Brucella* strains with cetaceans and seals as their preferred hosts.** *Int J Syst Evol Microbiol* 2007, **57**:2688–2693.
3. Foster G, Jahans KL, Reid RJ, Ross HM: **Isolation of *Brucella* species from cetaceans, seals and an otter.** *Vet Rec* 1996, **138**:583–586.
4. Nymo IH, Tryland M, Godfroid J: **A review of *Brucella* infection in marine mammals, with special emphasis on *Brucella pinnipedialis* in the hooded seal (*Cystophora cristata*).** *Vet Res* 2011, **42**:93.
5. Guzmán-Verrí C, González-Barrientos R, Hernández-Mora G, Morales J-A, Baquero-Calvo E, Chaves-Olarte E, Moreno E: ***Brucella ceti* and Brucellosis in Cetaceans.** *Front Cell Infect Microbiol* 2012, **2**:3.
6. Whatmore AM, Dawson CE, Groussaud P, Koylass MS, King AC, Shankster SJ, Sohn AH, Probert WS, McDonald WL: **Marine mammal *Brucella* genotype associated with zoonotic infection.** *Emerg Infect Dis* 2008, **14**:517–518.
7. González L, Patterson IA, Reid RJ, Foster G, Barbera M, Blasco JM, Kennedy S, Howie FE, Godfroid J, MacMillan AP, Schock A, Buxton D: **Chronic meningoencephalitis associated with *Brucella* sp. Infection in live-stranded striped dolphins (*Stenella coeruleoalba*).** *J Comp Pathol* 2002, **126**:147–152.
8. Muñoz PM, García-Castrillo C, López-García P, González-Cueli JC, De Miguel MJ, Marín CM, Barberán M, Blasco JM: **Isolation of *Brucella* species from a live-stranded striped dolphin (*Stenella coeruleoalba*) in Spain.** *Vet Rec* 2006, **158**:450–451.
9. Davison NJ, Cranwell MP, Perrett LL, Dawson CE, Deaville R, Stubberfield EJ, Jarvis DS, Jepson PD: **Meningoencephalitis associated with *Brucella* species in a live-stranded striped dolphin (*Stenella coeruleoalba*) in south-west England.** *Vet Rec* 2009, **165**:86–89.
10. Foster G, MacMillan AP, Godfroid J, Howie F, Ross HM, Cloeckaert A, Reid RJ, Brew S, Patterson IAP: **A review of *Brucella* sp. infection of sea mammals with particular emphasis on isolates from Scotland.** *Vet Microbiol* 2002, **90**:563–580.
11. Alba P, Terracciano G, Franco A, Lorenzetti S, Cocumelli C, Fichi G, Eleni C, Zygmunt MS, Cloeckaert A, Battisti A: **The presence of *Brucella ceti* ST26 in a striped dolphin (*Stenella coeruleoalba*) with meningoencephalitis from the Mediterranean Sea.** *Vet Microbiol* 2013, **164**:158–163.
12. González-Barrientos R, Morales JA, Hernández-Mora G, Barquero-Calvo E, Guzmán-Verrí C, Chaves-Olarte E, Moreno E: **Pathology of striped dolphins (*Stenella coeruleoalba*) infected with *Brucella ceti*.** *J Comp Pathol* 2010, **142**:347–352.
13. Hernández-Mora G, González-Barrientos R, Morales J-A, Chaves-Olarte E, Guzmán-Verrí C, Barquero-Calvo E, Baquero-Calvo E, De-Miguel M-J, Marín C-M, Blasco J-M, Moreno E: **Neurobrucellosis in stranded dolphins, Costa Rica.** *Emerg Infect Dis* 2008, **14**:1430–1433.
14. Van Bresselem MF, Van Waerebeek K, Raga JA, Godfroid J, Brew SD, MacMillan AP: **Serological evidence of *Brucella* species infection in odontocetes from the south Pacific and the Mediterranean.** *Vet Rec* 2001, **148**:657–661.
15. Garofolo G, Zilli K, Troiano P, Petrella A, Marotta F, Di Serafino G, Ancora M, Di Giannatale E: ***Brucella ceti* from two striped dolphins stranded on Apulia coastline, Italy.** *J Med Microbiol* 2013, **63**:325–329.
16. OIE: **Bovine Brucellosis. Manual of Diagnostic Tests and Vaccines for Terrestrial Animals 2013 2013, Chapter 2.4.3.**
17. Soto S, González R, Alegre F, González B, Medina P, Raga JA, Marco A, Domingo M: **Epizootic of dolphin morbillivirus on the Catalanian Mediterranean coast in 2007.** *Vet Rec* 2011, **169**:101–102.
18. Farrell ID: **The development of a new selective medium for the isolation of *Brucella abortus* from contaminated sources.** *Res Vet Sci* 1974, **16**:280–286.
19. De Miguel MJ, Marín CM, Muñoz PM, Dieste L, Grilló MJ, Blasco JM: **Development of a selective culture medium for primary isolation of the main *Brucella* species.** *J Clin Microbiol* 2011, **49**:1458–1463.
20. Alton GG, Jones LM, Angus RD, Verger JM: **Techniques for the Brucellosis Laboratory.** Paris France: INRA; 1988.
21. Mayer-Scholl A, Draeger A, Göllner C, Scholz HC, Nöckler K: **Advancement of a multiplex PCR for the differentiation of all currently described *Brucella* species.** *J Microbiol Methods* 2010, **80**:112–114.
22. Cloeckaert A, Verger JM, Grayon M, Paquet JY, Garin-Bastuji B, Foster G, Godfroid J: **Classification of *Brucella* spp. isolated from marine mammals by DNA polymorphism at the omp2 locus.** *Microbes Infect* 2001, **3**:729–738.
23. Le Flèche P, Jacques I, Grayon M, Dahouk AI S, Bouchon P, Denoëuf F, Nöckler K, Neubauer H, Guilloteau LA, Vergnaud G: **Evaluation and selection of tandem repeat loci for a *Brucella* MLVA typing assay.** *BMC Microbiol* 2006, **6**:9.

24. Tanaka S, Suto T, Isayama Y, Azuma R, Hatakeyama H: **Chemo-taxonomical studies on fatty acids of *Brucella* species.** *Ann Sclavo* 1977, **19**:67–82.
25. Dahouk Al S, Flèche PL, Nöckler K, Jacques I, Grayon M, Scholz HC, Tomaso H, Vergnaud G, Neubauer H: **Evaluation of *Brucella* MLVA typing for human brucellosis.** *J Microbiol Methods* 2007, **69**:137–145.
26. Maquart M, Le Flèche P, Foster G, Tryland M, Ramisse F, Djonje B, Dahouk Al S, Jacques I, Neubauer H, Walravens K, Godfroid J, Cloeckert A, Vergnaud G: **MLVA-16 typing of 295 marine mammal *Brucella* isolates from different animal and geographic origins identifies 7 major groups within *Brucella ceti* and *Brucella pinnipedialis*.** *BMC Microbiol* 2009, **9**:145.
27. *MLVA-NET for Brucella.* <http://mlva.u-psud.fr/brucella/>.
28. Moreno E, Moriyón I: **The Genus *Brucella*.** In *The Prokaryotes*, Volume 5. Edited by Dworkin M, Falkow S, Rosenberg E, Schleifer K-H, Stackebrandt E. New York: Springer Verlag; 2006:315–456.
29. Isidoro-Ayza M, Perez L, Cabanes FJ, Castella G, Andres M, Vidal E, Domingo M: **Central nervous system mucormycosis caused by *Cunninghamella Bertholletiae* in a bottlenose dolphin (*Tursiops truncatus*).** *J Wildl Dis* 2014, **50**:634–638.
30. Soto S, Alba A, Ganges L, Vidal E, Raga J, Alegre F, González B, Medina P, Zorrilla I, Martínez J, Marco A, Pérez M, Pérez B, Mesas A, Valverde R, Domingo M: **Post-epizootic chronic dolphin morbillivirus infection in Mediterranean striped dolphins *Stenella coeruleoalba*.** *Dis Aquat Org* 2011, **96**:187–194.
31. Ferreira L, Vega Castaño S, Sánchez-Juanes F, González-Cabrero S, Menegotto F, Orduña-Domingo A, González-Buitrago JM, Muñoz-Bellido JL: **Identification of *Brucella* by MALDI-TOF mass spectrometry. Fast and reliable identification from agar plates and blood cultures.** *PLoS One* 2010, **5**:e14235.
32. Brower A, Lucero N, Okwumabua O, Groussaud P, Gopaul KK, Whatmore AM, Cravero SL, Trangoni MD: **Newly identified variability in *Brucella canis* fatty-acid content is associated with geographical origin.** *Epidemiol Infect* 2013, **141**:852–858.
33. Voorhees KJ, Jensen KR, McAlpin CR, Rees JC, Cody R, Ubukata M, Cox CR: **Modified MALDI MS fatty acid profiling for bacterial identification.** *J Mass Spectrom* 2013, **48**:850–855.
34. Natoli A, Birkun A, Aguilar A, Lopez A, Hoelzel AR: **Habitat structure and the dispersal of male and female bottlenose dolphins (*Tursiops truncatus*).** *Proc Biol Sci* 2005, **272**:1217–1226.
35. Azzolin M, Papale E, Lammers MO, Gannier A, Giacoma C: **Geographic variation of whistles of the striped dolphin (*Stenella coeruleoalba*) within the Mediterranean Sea.** *J Acoust Soc Am* 2013, **134**:694–705.
36. Larsen AK, Nymo IH, Boysen P, Tryland M, Godfroid J: **Entry and elimination of marine mammal *Brucella* spp. by hooded seal (*Cystophora cristata*) alveolar macrophages in vitro.** *PLoS One* 2013, **8**:e70186.
37. Dagleish MP, BARLEY J, Howie FE, Reid RJ, Herman J, Foster G: **Isolation of *Brucella* species from a diseased atlanto-occipital joint of an Atlantic white-sided dolphin (*Lagenorhynchus acutus*).** *Vet Rec* 2007, **160**:876–878.
38. Domingo M, Visa J, Pumarola M, Marco AJ, Ferrer L, Rabanal R, Kennedy S: **Pathologic and immunocytochemical studies of morbillivirus infection in striped dolphins (*Stenella coeruleoalba*).** *Vet Pathol* 1992, **29**:1–10.
39. Maynou F, Sbrana M, Sartor P, Maravelias C, Kavadis S, Damalas D, Cartes JE, Osio G: **Estimating trends of population decline in long-lived marine species in the Mediterranean Sea based on fishers' perceptions.** *PLoS One* 2011, **6**:e21818.
40. Di Guardo G, Mazzariol S, Fernández A: **Biologically threatened dolphins and whales.** *Environ Microbiol* 2011, **13**:2833–2834.

doi:10.1186/s12917-014-0206-7

Cite this article as: Isidoro-Ayza et al.: *Brucella ceti* infection in dolphins from the Western Mediterranean sea. *BMC Veterinary Research* 2014 10:206.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

