

Low FOXJ2 expression is associated with unfavorable postoperative prognosis of patients with epithelial ovarian cancer

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Abstract

The forkhead box (FOX) family is a large and diverse group of transcription factors. Forkhead box J2 (FOXJ2) is a member of the FOX family that is aberrantly expressed in a variety of cancers. However, its role in epithelial ovarian cancer (EOC) remains elusive. The purpose of this study was to evaluate the prognostic value of FOXJ2 expression in patients with epithelial ovarian cancer.

The current study retrospectively included 151 patients with EOC from January 2013 to September 2016. FOXJ2 expression was analyzed by immunohistochemistry based on tissue microarrays. Then, the prognostic value of FOXJ2 expression and clinical outcomes were evaluated by Kaplan–Meier and cox regression analysis.

Low FOXJ2 expression was associated with high International Federation of Gynecology and Obstetrics (FIGO) stage. Kaplan-Meier curves showed that high FOXJ2 expression was associated with improved median overall survival (OS, 57.9 vs 31.9 months; *P*=.037) and longer median progression-free survival (PFS, 31.8 vs 18.1 months; *P*=.012). Univariate analysis demonstrated that FOXJ2 expression was significantly correlated with OS and PFS in patients with epithelial ovarian cancer. Multivariate analysis revealed FOXJ2 expression as an independent prognostic factor of progression-free survival of epithelial ovarian cancer patients. Low FOXJ2 expression is a novel adverse prognostic factor of clinical outcome in epithelial ovarian cancer.

Abbreviations: Forkhead box, FOX = Forkhead box, FOXJ2 = Forkhead box J2, PBS = phosphate buffered saline, PDS = primary debulking surgery, TMAs = tissue microarrays, WHO = World Health Organization.

Keywords: epithelial ovarian cancer, forkhead box J2, overall survival, prognostic factor, progression-free survival

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This study was approved by the ethics committee of Nantong Tumor Hospital. The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

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The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

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1. Introduction

Ovarian cancer, one of the most common malignancies affecting the female reproductive system, has the highest mortality rate among malignant gynecological tumors.^[1,2] The World Health Organization (WHO) estimates that 225,500 individuals are diagnosed with ovarian cancer yearly, with 140,200 patients succumbing to the malignancy; therefore, ovarian cancer is the 7th commonest and the 8th deadliest cancer among women around the world.^[3,4] Ovarian cancer stages range between I and VI; only approximately 13% of serous ovarian carcinoma cases are detected at stage I or II, with most individuals diagnosed at the stage of distant metastasis.^[5] Despite complete remission upon initial treatment, 60% of individuals with advanced stage ovarian cancer show relapse within 5 years.^[6,7] Unfortunately, the pathogenesis of ovarian cancer remains unclear. Thus, it is crucial to investigate the molecular regulatory mechanisms related to the malignant behaviors of epithelial ovarian cancer and identify new therapeutic targets.

Forkhead box (FOX) family factors are transcription factors that share an evolutionarily conserved DNA-binding domain, termed the "fork-head" or "winged-helix" domain.^[8] Currently, mounting evidence suggests that FOX family members are abnormally expressed in many cancers, and contribute to a variety of cellular processes, such as proliferation, differentiation, adhesion, migration, and invasion.^[9–11] Forkhead box J2 (FOXJ2), a member of the FOX family, is widely distributed in different organs and tissues, from the fetus to adults.^[12] Meanwhile, the expression patterns of FOXJ2 have been reported to be aberrant in a variety of cancers, including breast cancer, extra-hepatic cholangiocarcinoma, nasopharyngeal car-

cinoma, glioma, and non-small cell lung cancer.^[13–17] FOXJ2 was shown to positively regulate cell cycle progression and induce tumorigenesis.^[18,19] However, FOXJ2 was described as a tumor suppressor in liver cancer,^[20] indicating that its exact function in cancer deserves further investigation.

Specifically, the role of FOXJ2 in the development of epithelial ovarian cancer remains unknown.

Therefore, this study aimed to assess the prognostic value of FOXJ2 expression in patients with epithelial ovarian cancer. The results could advance our understanding of the molecular basis of EOC development.

2. Methods

2.1. Patients and specimens

This was a retrospective study. A total of 151 tumor specimens from EOC patients administered surgical treatment in Nantong Tumor Hospital from January 2013 to September 2016 were enrolled in our study. Tumor specimens were obtained by surgery and selected for tissue microarrays (TMAs). Inclusion criteria were:

- 1. pathological diagnosis of epithelial ovarian cancer confirmed by 2 experienced pathologists according to the 2014 WHO classification of ovarian tumors;^[21]
- 2. available formalin fixed paraffin embedded specimen of the tumor mass ($\geq 1 \text{ cm}^3$);
- underwent surgical treatment by primary debulking surgery (PDS) or interval debulking surgery after neoadjuvant chemotherapy (NACT+IDS);
- 4. complete clinical and follow-up data.

Exclusion criteria were:

- 1. other previous malignant tumors;
- 2. samples with over 80% necrotic or hemorrhagic area;
- 3. missing follow-up or clinic data. This study was approved by the ethics committee of Nantong Tumor Hospital.

Informed consent was waived by the committee because of the retrospective nature of the study.

2.2. Data collection

Overall survival was defined as time from cancer diagnosis to death or the date of last contact. Progression-free survival was defined as time from cancer diagnosis to recurrence or progression.^[22] Patients were followed up every 2 to 3 months during the first 2 years after the end of the initial treatment and every 4 to 6 months thereafter. Censoring occurred on January 12, 2020. Chemosensitivity was identified by a time interval of ≥ 6 months between chemotherapy completion and the detection of recurrence. Chemoresistance was defined as disease progression during adjuvant chemotherapy or within a time interval of < 6 months between chemotherapy completion and the detection of recurrence.^[23]

2.3. Tissue microarray construction and immunohistochemistry

TMAs were obtained with formalin-fixed, paraffin embedded surgical specimens. All samples were assessed histologically by hematoxylin and eosin staining, and representative tumor areas were marked on the paraffin blocks away from necrotic and hemorrhagic regions. Tissue cylinders of 2 mm in diameter containing tumor tissues were punched out from the selected area of each tissue block and transferred into a TMA block using a TMA instrument.

Sections from TMA blocks were sectioned at 4 µm. Paraffin embedded sections were dewaxed in xylene and rehydrated in graded ethanol. After washing with phosphate buffered saline (PBS) for 3 times, citrate buffer (pH=6) was used to restore the product in a pressure cooker at high pressure. The tissue sections were rinsed with PBS and soaked in 3% H₂O₂ for 10 minutes to block endogenous peroxidase. The samples were washed with PBS, and incubated with rabbit anti-FOXJ2 polyclonal primary antibodies (Abcam, ab22857, 1:100) at room temperature for 2 hours. After rinsing with PBS, the slides were incubated with secondary antibodies for 30 minutes, washed with PBS, developed with DAB solution for about 3 minutes, counterstained with hematoxylin, dehydrated, and mounted with resin mount. Two independent pathologists evaluated FOXJ2 staining in TMAs, based on a semi-quantitative H-score ranging from 0 to 300, derived from the multiplication of staining intensity (0, negative; 1, weak staining; 2, moderate staining; 3, strong staining) and distribution (0-100%). In brief, H-score was derived as 3x percentage of strongly stained positive cells + 2 \times percentage of moderately stained positive cells + $1 \times$ percentage of weakly stained positive cells, with a range of 0 to 300.^[24,25]

2.4. Statistical analysis

Statistical analysis were performed with SPSS 23.0 (IBM, Armonk, NY). The Chi-Squared test was performed to evaluate the associations of FOXJ2 expression with clinicopathological parameters. The Kaplan– Meier method and log- rank test were applied to analyze differences in survival rates between groups. The Cox proportional hazard regression model was used for univariate and multivariate analyses of prognosis. P < .05 was considered statistically significant.

3. Results

3.1. FOXJ2 expression and associations with clinicopathological characteristics

A total of 151 epithelial ovarian cancer patients were included in the analysis. Patient characteristics were shown in Table 1. The median age was 58 years (range 20-76). There were 63 (41.72%) patients in the NACT+IDS subgroup, and 88(58.28%) cases in the PDS group. FOXJ2 expression levels were not compared between tumor and adjacent normal tissue samples, because it is usually hard to define tumor boundaries, which is one of the characteristics of ovarian cancer. Hence, we first evaluated FOXJ2 expression by immunohistochemistry in tumor specimens from 151 epithelial ovarian cancer patients. FOXJ2 expression was detected in tumor cell nuclei and cytoplasm. Immunohistochemical scores (H-scores) of the tumor tissues differed among specimens. Representative photographs under light microscope showing differential FOXI2 immunohistochemical staining intensities in epithelial ovarian cancer tissues were shown in Figure 1. The average measured H-score was 104.5 (range 0-300). The cut-off point for the high/low expression subgroups was set at 100.0. Consequently, a total of 58 (38.4%) patients

Table 1			
Clinical characteristics of the patients.			
Parameter	N		
Age, years			
Median (range)	58 (20-		
Menopause			
M -	0.4		

Menopause		
No	34	22.52%
Yes	117	77.48%
FIGO stage		
1-11	37	24.50%
III-IV	114	75.50%
Histological subtype		
Serous	114	75.50%
Non-serous	37	24.50%
Histological grade		
Low-grade	25	16.56%
High-grade	126	83.44%
Ascites (moderate to large volume)		
No	74	49.01%
Yes	77	50.99%
CA125 (U/ml)		
<500	61	40.40%
≥500	90	59.60%
Treatment		
PDS	88	58.28%
NACT+IDS	63	41.72%
Residual disease		
No	86	56.95%
Yes	65	43.05%
Chemosensitivity		
Sensitive	107	70.86%
Resistant	44	29.14%
Recurrence and progression		
No	39	25.83%
Yes	112	74.17%

CA125 = cancer antigen 125, PDS = primary debulking surgery, NACT+IDS = interval debulking

surgery after neoadjuvant chemotherapy.

58 (20-76)

Table 2

%

Correlations of FOXJ2 expression and clinicopathological characteristics of ovarian cancer patients (n=151).

	FOXJ2 ex		
Parameter	High (n=58)	Low (n=93)	P value
Age, years			.887
< 58	28	46	
≥58	30	47	
Menopause			.981
No	13	21	
Yes	45	72	
FIGO stage			.002
-	22	15	
III-IV	36	78	
Histological subtype			.637
Serous	45	69	
Non-serous	13	24	
Histological grade			.126
Low-grade	13	12	
High-grade	45	81	
Ascites (moderate to large volume)			.598
No	30	44	
Yes	28	49	
CA125 (U/ml)			.381
<500	26	35	
≥500	32	58	
Treatment			<.001
PDS	45	43	
NACT+IDS	13	50	
Residual disease			.180
No	37	49	
Yes	21	44	
Chemosensitivity			.151
Sensitive	45	62	
Resistant	13	31	
Recurrence and progression			.011
No	19	20	
Yes	39	73	

CA125 = cancer antigen 125, PDS = primary debulking surgery, NACT+IDS = interval debulking surgery after neoadjuvant chemotherapy.

P value for high vs low FOXJ2 expression groups.



negative staining

weak staining

moderate staining

strong staining

Figure 1. Representative micrographs depicting different FOXJ2 expression levels, as assessed by immunohistochemical (IHC) staining of epithelial ovarian cancer tissue samples. Magnifications are 40× (A) and 100× (B).



were classified into the FOXJ2 high expression subgroup and the low expression subgroup included 93 (61.6%) individuals.

Table 2 presented the associations of FOXJ2 expression with clinicopathological characteristics in EOC patients. Patients with low FOXJ2 expression was associated with advanced FIGO stage (P=.002) and tended to receive PDS treatment (P<.001). The expression of FOXJ2 had no significant correlations with other clinicopathological characteristics (P > .05).

3.2. Correlations between FOXJ2 expression and prognosis of EOC patients

The median follow-up was 64.2 (3.3–80.8) months. Median OS and PFS in all patients were 41.1 months and 20.5 months, respectively. The 3-year overall survival and progression-free survival rates in all patients were 55.0% and 35.8%, respectively. Kaplan–Meier survival analysis was performed to assess OS and PFS according to FOXJ2 expression levels. The high FOXJ2 expression subgroup showed longer median OS (57.9 vs 31.9 months, P=.037; Fig. 2A) and longer median PFS (31.8 vs 18.1 months, P=.012; Figure 2B) compared with the low FOXJ2 expression subgroup.

We further performed subgroup analysis according to FIGO stage (I–II or III–IV) (Fig. 3). When the analysis was restricted to the FIGO III-IV subgroup, low FOXJ2 expression correlated with decreased OS and PFS (P=.028 and P=.005, respectively; Fig. 3B, 3D). However, in the FIGO I-II subgroup, the difference was not significant in OS or PFS (P=.526 and P=.592, respectively; Fig. 3A, 3C).

3.3. Univariate and multivariate Cox proportional hazard analysis

Univariate analysis was performed for OS and PFS to estimate the clinical significance of FOXJ2 expression in epithelial ovarian cancer (Tables 3 and 4). The results showed that FIGO stage, histological grade, CA125 levels, therapeutic approach, residual disease, chemosensitivity, and FOXJ2 expression were significantly correlated with OS. In addition, FIGO stage, histological

grade, CA125 levels, therapeutic approach, residual disease, moderate to large amounts of ascitic fluid and FOXJ2 expression were significantly correlated with PFS.

Next, significant factors (P < .05) in univariate analysis were included into multivariate analysis. In the multivariate Cox regression analysis, FIGO stage (HR = 2.025, 95% CI = 1.127– 3.605, P=.014), histological grade (HR = 3.872, 95% CI = 1.730–8.670, P=.001), residual disease (HR = 4.933, 95% CI = 3.055–7.967, P < .001) and chemosensitivity (HR = 2.993, 95% CI = 1.823–4.916, P < .001) were independent prognostic factors of OS (Table 3). Furthermore, FIGO stage (HR = 2.214, 95% CI = 1.220–4.018, P = .009), histological grade (HR = 2.849, 95% CI = 1.407–5.770, P = .004), residual disease (HR = 4.046, 95% CI = 2.700–6.062, P < .001), FOXJ2 expression (HR = 1.850, 95%CI 1.331–2.408; P = .025) were independent prognostic factors of PFS (Table 4). Overall, it illustrated that FOXJ2 expression was an independent prognostic factor of PFS, but not OS, in EOC patients.

In the PDS and NACT+IDS subgroups, Cox regression analysis of factors potentially predicting OS and PFS were shown in Tables 5 and 6. Univariate analysis showed that FOXJ2 expression was significantly correlated with OS and PFS in both the PDS and NACT+IDS subgroups. But multivariate analysis showed that FOXJ2 expression (HR=1.762, 95%CI=1.358–2.048, P=.010) was an independent prognostic factor of PFS only in the PDS subgroup, but not NACT+IDS group.

4. Discussion

In this study, we assessed FOXJ2 expression levels in EOC specimens and determined the association between FOXJ2 expression and prognosis in epithelial ovarian cancer. The results demonstrated that FOXJ2 expression was negatively correlated to FIGO stage. Moreover, low FOXJ2 expression was an independent prognostic factor of progression-free survival in epithelial ovarian cancer.

Previous evidence suggests that FOXJ2 actively participated in tumor development and metastasis. It was demonstrated that



Figure 3. Kaplan-Meier curves of OS (A and B) and PFS(C and D) based on FOXJ2 expression in the FIGO I-II (A and C) and FIGO III-IV (B and D) subgroups.

FOXJ2 inhibited metastasis in human breast cancer^[13] and glioma^[16] by regulating EMT key markers, including E-cadherin and vimentin. Qiang et al identified that FOXJ2 expression was abnormally downregulated in extrahepatic cholangiocarcinoma and its overexpression could markedly inhibit cell proliferation, migration and invasion in vitro, verifying FOXJ2 as a tumor suppressor.^[14] In addition, FOXJ2 could also inhibit the proliferation of human hepatocellular carcinoma cells, and reduced expression of FOXJ2 was significantly associated with the poor prognosis of patients with human hepatocellular carcinoma.^[20] The present study revealed that low FOXJ2 expression was associated with advanced FIGO stage by assessing 151 epithelial ovarian cancer cases. Besides, FOXJ2 expression was significantly different regarding treatment approaches. Tumor specimens from patients in NACT+IDS subgroups had significantly lower FOXJ2 expression than those in PDS subgroups. Compared with those in PDS subgroup, patients in the NACT+IDS subgroup were mainly advanced FIGO stage III-IV cases. Therefore, FOXJ2 might act as a tumor suppressor in ovarian cancer. Patients with low FOXJ2 expression tended to progress to a higher stage and received NACT+IDS treatment. Another explanation might be the neoadjuvant chemotherapy (Taxol and carboplatin) administered to the NACT+IDS group. FOXJ2 expression might also be involved in the interaction with chemotherapeutic drugs. Cell biology experiments are required to further assess its interaction with chemotherapeutic drugs or its original role in EOC.

The prognostic significance of FOXJ2 varies with the type of malignancy. Studies mentioned above proved that FOXJ2 could suppress carcinogenesis. Nonetheless, Shan et al found that downregulation of FOXJ2 decreased the cell population in the S phase with enhanced G1 cycle arrest in nasopharyngeal carcinoma CNE-2 cells, and confirmed that patients with FOXJ2 overexpression had shorter survival in nasopharyngeal carcinoma.^[15] In the current study, Kaplan–Meier survival analysis revealed that the high-FOXJ2 expression group showed longer median OS and PFS compared with the low-FOXJ2 expression group. Similar findings were obtained in the FIGO III-IV subgroup in stratified analysis. In the FIGO I-II subgroup, the difference was not significant, which may be related to the small

Table 3

Univariate and multivariate cox regression analyses of factors potentially predicting OS.

	Univariate		Multivariate	
	HR (95% CI)	Р	HR (95% CI)	Р
Age, years				
<58	Ref.			
≥58	1.247 (0.830-1.875)	.287		
Menopause	· · · ·			
No	Ref.			
Yes	1.442 (0.852-2.438)	.173		
FIGO stage				
I-II	Ref.		Ref.	
III-IV	2.577 (1.479-4.490)	.001	2.025 (1.127-3.605)	.014
Histological subtype				
Serous	Ref.			
Non-serous	0.941 (0.587-1.507)	.800		
Histological grade				
Low-grade	Ref.		Ref.	
High-grade	3.705 (1.786-7.684)	<.001	3.872 (1.730-8.670)	.001
Ascites (moderate to large volume)				
No	Ref.			
Yes	1.461 (0.970-2.200)	.069		
CA125 (U/ml)				
<500	Ref.			
≥500	2.000 (1.285-3.113)	.002		
Treatment				
PDS	Ref.			
NACT+IDS	2.243 (1.485-3.387)	<.001		
Residual disease				
No	Ref.		Ref.	
Yes	6.375 (4.124–9.855)	<.001	4.933 (3.055-7.967)	<.001
Chemosensitivity				
Sensitive	Ref.			
Resistant	5.084 (3.324-7.776)	<.001	2.993 (1.823-4.916)	<.001
FOXJ2 expression				
High (>100)	Ref.			
Low (\leq 100)	1.577 (1.024-2.429)	.039	1.351 (1.058-2.146)	.202

Ref. = reference, CA125 = cancer antigen 125, PDS = primary debulking surgery, NACT+IDS = interval debulking surgery after neoadjuvant chemotherapy, OS = overall survival HR = hazard ratio, CI = confidence interval.

Table 4

Univariate and multivariate cox regression analyses of factors potentially predicting PFS.

$\begin{tabular}{ c c c c c } \hline HR (95% Cl) P & HR (95% Cl) P \\ \hline S58 & 1.140 (0.785-1.655) .493 \\ \hline Menopause & Ref. \\ \hline No & Ref. \\ \hline Yes & 1.523 (0.947-2.450) .0.83 \\ \hline FIGO stage & Ref. \\ III-V & 3.280 (1.945-5.530) <0.001 & 2.214 (1.220-4.018) .0.09 \\ \hline HI & Ref. & Ref. \\ III-V & 3.280 (1.945-5.530) &0.999 \\ \hline Strous & Ref. \\ Non-serous & 0.683 (0.434-1.074) .0.999 \\ \hline Strous & Ref. & Ref. \\ Illoygrade & Ref. & Ref. \\ Illoygrade & Ref. & Ref. \\ Illoygrade & 3.490 (1.812-6.720) &0.001 & 2.849 (1.407-5.770) .0.004 \\ \hline Ascites (moderate to large volume) & Ref. \\ \hline Yes & 1.660 (1.138-2.420) & .009 \\ \hline Strous & Ref. & Ref. \\ \hline Yes & 1.660 (1.138-2.420) & .009 \\ \hline Fox Jone & Ref. & Ref. \\ \hline Strous & Ref. & Ref. \\ \hline Yes & 1.660 (1.138-2.467) & .017 \\ \hline Restitual disease & Ref. & Ref. \\ \hline Yes & 1.633 (1.094-2.467) & .017 \\ \hline Fox Jone & Ref. & Ref. \\ \hline Yes & 1.633 (1.094-2.467) & .017 \\ \hline FOX J2 expression & Ref. & Ref. \\ \hline Yes & 4.266 (2.896-6.284) & <.001 & 4.046 (2.700-6.062) & <.001 \\ \hline FOX J2 expression & Ref. & Ref. \\ \hline Help (1.00) & Ref. & Ref. \\ \hline Yes & 1.000 & 1.962 (1.331-2.408) & .025 \\ \hline \end{tabular}$		Univariate		Multivariate	
Age, years Ref. <58 1.40 (0.785-1.655) .493 Meropause 1.140 (0.785-1.655) .493 No Ref.		HR (95% CI)	Р	HR (95% CI)	Р
$^{-}$ S8 Ref. ≥58 1.140 (0.785–1.655) .493 Menopause Ref. No Ref. Yes 1.523 (0.947–2.450) .083 FIGO stage Ref. Ref. II-I Ref. .009 Hirt/ 3.280 (1.945–5.530) <.001	Age, years				
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FIGO stage Ref. Ref. Ref. .009 HI-V 3.280 (1.945–5.530) <.001	Yes	1.523 (0.947-2.450)	.083		
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$\begin{array}{c c c c c c c } & 3.280 \ (1.945-5.530) & <.001 & 2.214 \ (1.220-4.018) & .009 \\ \hline Histological subtype & & & & & & & & & & & & & & & & & & &$	I-II	Ref.		Ref.	
Histological subtype Serous Ref. Non-serous 0.683 (0.434-1.074) .099 Histological grade Low-grade Ref. Ref. High-grade 3.490 (1.812-6.720) <.001 2.849 (1.407-5.770) .004 Ascites (moderate to large volume) No Ref. Yes 1.660 (1.138-2.420) .009 CA125 (U/m) <500 Ref. ≥500 1.689 (1.140-2.502) .009 Treatment PDS Ref. NACT+IDS 1.643 (1.094-2.467) .017 Residual disease No Ref. Ref. Yes 4.266 (2.896-6.284) <.001 4.046 (2.700-6.062) <.001 FOXJ2 expression High (>100) Ref. Ref. Yes (1.385-2.808) .012 1.850 (1.331-2.408) .025	III-IV	3.280 (1.945-5.530)	<.001	2.214 (1.220-4.018)	.009
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CA125 (U/ml) <500	Yes	1.660 (1.138-2.420)	.009		
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Treatment Ref. PDS Ref. NACT+IDS 1.643 (1.094–2.467) .017 Residual disease .017 No Ref. Ref. Yes 4.266 (2.896–6.284) <.001	≥500	1.689 (1.140-2.502)	.009		
PDS Ref. NACT+IDS 1.643 (1.094-2.467) .017 Residual disease .017 No Ref. Ref. Yes 4.266 (2.896-6.284) <.001	Treatment				
NACT+IDS 1.643 (1.094-2.467) .017 Residual disease No Ref. Ref. Yes 4.266 (2.896-6.284) <.001	PDS	Ref.			
Residual disease Ref. Ref. No Ref. Ref. Yes 4.266 (2.896–6.284) <.001	NACT+IDS	1.643 (1.094-2.467)	.017		
No Ref. Ref. Yes 4.266 (2.896–6.284) <.001	Residual disease	, ,			
Yes 4.266 (2.896-6.284) <.001 4.046 (2.700-6.062) <.001 F0XJ2 expression	No	Ref.		Ref.	
F0XJ2 expression High (>100) Ref. Ref. Low (≤ 100) 1.962 (1.358-2.808) .012 1.850 (1.331-2.408) .025	Yes	4.266 (2.896-6.284)	<.001	4.046 (2.700-6.062)	<.001
High (>100) Ref. Ref. Low (≤ 100) 1.962 (1.358-2.808) .012 1.850 (1.331-2.408) .025	FOXJ2 expression			х , ,	
Low (≤ 100) 1.962 (1.358-2.808) .012 1.850 (1.331-2.408) .025	High (>100)	Ref.		Ref.	
	Low (< 100)	1.962 (1.358-2.808)	.012	1.850 (1.331-2.408)	.025

Ref. = reference, CA125 = cancer antigen 125; PDS = primary debulking surgery, NACT+IDS = interval debulking surgery after neoadjuvant chemotherapy, OS = overall survival, HR = hazard ratio, CI = confidence interval.

Table 5

Cox regression analyses of factors potentially predicting OS in PDS and NACT+IDS subgroups.

	PDS Univariate ([*] Multivariate)		NACT+IDS Univariate ([*] Multivariate)	
	HR (95% CI)	Р	HR (95% CI)	Р
Age, years				
<58	Ref.		Ref.	
≥58	1.248 (0.686-2.272)	.468	1.097 (0.830-1.875)	.745
Menopause				
No	Ref.		Ref.	
Yes	1.259 (0.604-2.625)	.540	1.546 (0.726-3.294)	.258
FIGO stage				
I-II	Ref.		NA	
III-IV	1.880 (1.001-3.532)	.046		
Histological subtype				
Serous	Ref.		Ref.	
Non-serous	1.542 (0.841-2.287)	.162	0.651 (0.257-1.650)	.366
Histological grade				
Low-grade	Ref.	*	Ref.	
High-grade	3.159 (1.323-7.543)	.010	2.511 (0.632-7.068)	.023
Ascites (moderate to large volume)				
No	Ref.		Ref.	
Yes	0.960 (0.507-1.818)	.901	1.318 (0.689-2.520)	.404
CA125 (U/ml)				
<500	Ref.		Ref.	
≥500	1.842 (0.998-3.403)	.047	1.534 (0.782-3.009)	.213
Residual disease				
No	Ref.	*	Ref.	*
Yes	6.872 (3.595–13.133)	<.001	6.514 (3.481–12.191)	<.001
Chemosensitivity				
Sensitive	Ref.	*	Ref.	*
Resistant	6.052 (3.239–11.307)	<.001	3.815 (2.106-6.910)	<.001
FOXJ2 expression				
High (>100)	Ref.		Ref.	
Low (\leq 100)	1.874 (1.122–3.283)	.046	1.675 (1.092–2.746)	.040

Ref. = reference, CA125 = cancer antigen 125, PDS = primary debulking surgery, NACT+IDS = interval debulking surgery after neoadjuvant chemotherapy, OS = overall survival, HR = hazard ratio, CI = confidence interval.

Table 6

Cox regression analyses of factors potentially predicting PFS in PDS and NACT+IDS subgroups.

	PDS Univariate ([®] Multivariate)		NACT+IDS Univariate ([*] Multivariate)	
	HR (95% CI)	Р	HR (95% CI)	Р
Age, years				
<58	Ref.		Ref.	
≥58	1.088 (0.640-1.851)	.756	1.085 (0.636-1.850)	.766
Menopause				
No	Ref.		Ref.	
Yes	1.498 (0.773-2.904)	.231	0.977 (0.525-1.819)	.941
FIGO stage				
-	Ref.		NA	
III-IV	2.692 (1.508-4.804)	.003*		
Histological subtype				
Serous	Ref.		Ref.	
Non-serous	1.026 (0.588-1.790)	.927	0.920 (0.656-1.064)	.670
Histological grade				
Low-grade	Ref.		Ref.	
High-grade	3.430 (1.541-7.638)	.001*	1.685 (0.580-5.994)	.035
Ascites (moderate to large volume)				
No	Ref.		Ref.	
Yes	1.375 (0.796-2.375)	.254	1.670 (1.083-2.204)	.056
CA125 (U/ml)				
<500	Ref.		Ref.	
≥500	1.508 (0.884-2.572)	.132	1.529 (1.052-2.221)	.089
Residual disease				
No	Ref.		Ref.	
Yes	4.276 (2.477-7.383)	<.001*	4.973 (2.742-9.020)	<.001
FOXJ2 expression				
High (>100)	Ref.	*	Ref.	
Low (\leq 100)	1.987 (1.369-2.510)	.010	1.762 (1.358-2.408)	.027

Ref. = reference, CA125 = cancer antigen 125, PDS = primary debulking surgery, NACT+IDS = interval debulking surgery after neoadjuvant chemotherapy, OS = overall survival, HR = hazard ratio, CI = confidence interval

sample size. Univariate analysis showed that FOXJ2 expression was significantly correlated with OS and PFS in patients with epithelial ovarian cancer. Similar results were obtained in both the PDS and NACT+IDS subgroups. Multivariate analysis showed that FOXJ2 expression was an independent prognostic factor of progression-free survival in patients with epithelial ovarian cancer, as well as the PDS subgroup. Overall, these findings demonstrated the clinical significance of FOXJ2 expression, which independently predicts progression-free survival in epithelial ovarian cancer.

This study has limitations. It was a single center retrospective analysis, with a limited sample size. In addition, no external validation was performed. Even though this study found the association between FOXJ2 and the prognosis of EOC patients, the underlying pathophysiological mechanisms and causal relationships still remain unclear. Therefore, further studies are needed to investigate the role and mechanism of FOXJ2 at the ovarian cancer cell level.

In conclusion, the present report firstly demonstrated that low FOXJ2 expression is associated with unfavorable prognosis in epithelial ovarian cancer, and FOXJ2 independently predicts EOC progression-free survival. Further studies are required to validate these findings.

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Author contributions

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References

- Momenimovahed Z, Tiznobaik A, Taheri S, et al. Ovarian cancer in the world: epidemiology and risk factors. Int J Womens Health 2019; 11:287–99.
- [2] Torre LA, Trabert B, DeSantis CE, et al. Ovarian cancer statistics, 2018. CA Cancer J Clin 2018;68:284–96.
- [3] Ferlay J, Soerjomataram I, Dikshit R, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBO-CAN 2012. Int J Cancer 2015;136:E359–386.

- [4] Bray F, Ferlay J, Soerjomataram I, et al. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2018;68:394–424.
- [5] Narod S. Can advanced-stage ovarian cancer be cured? Nat Rev Clin Oncol 2016;13:255–61.
- [6] Vidal F, Guerby P, Luyckx M, et al. Are early relapses in advanced-stage ovarian cancer doomed to a poor prognosis? PLoS One 2016;11: e0147787.
- [7] Gui T, Cao D, Yang J, et al. Tumor heterogeneity has important consequences for personalized medicine in ovarian cancer. Histol Histopathol 2015;30:173–81.
- [8] Kaufmann E, Knöchel W. Five years on the wings of fork head. Mech Dev 1996;57:3–20.
- [9] Yu C, Chen L, Yie L, et al. Targeting FoxM1 inhibits proliferation, invasion and migration of nasopharyngeal carcinoma through the epithelial-to-mesenchymal transition pathway. Oncol Rep 2015;33: 2402–10.
- [10] Ou-Yang L, Xiao SJ, Liu P, et al. Forkhead box C1 induces epithelialmesenchymal transition and is a potential therapeutic target in nasopharyngeal carcinoma. Mol Med Rep 2015;12:8003–9.
- [11] Zhou Z, Zhang L, Xie B, et al. FOXC2 promotes chemoresistance in nasopharyngeal carcinomas via induction of epithelial mesenchymal transition. Cancer Lett 2015;363:137–45.
- [12] Pérez-Sánchez C, Gómez-Ferrería MA, de La Fuente CA, et al. FHX, a novel fork head factor with a dual DNA binding specificity. J Biol Chem 2000;275:12909–16.
- [13] Wang Y, Yang S, Ni Q, et al. Overexpression of forkhead box J2 can decrease the migration of breast cancer cells. J Cell Biochem 2012; 113:2729–37.
- [14] Qiang Y, Wang F, Yan S, et al. Abnormal expression of Forkhead Box J2 (FOXJ2) suppresses migration and invasion in extrahepatic cholangiocarcinoma and is associated with prognosis. Int J Oncol 2015;46:2449–58.
- [15] Shan Y, Chang T, Shi S, et al. Foxj2 overexpression is associated with poor prognosis, progression, and metastasis in nasopharyngeal carcinoma. Onco Targets Ther 2017;10:3733–41.
- [16] Qiu X, Ji B, Yang L, et al. The role of FoxJ2 in the migration of human glioma cells. Pathol Res Pract 2015;211:389–97.
- [17] Yang Q, Cao X, Tao G, et al. Effects of FOXJ2 on TGF-(1-induced epithelial-mesenchymal transition through Notch signaling pathway in non-small lung cancer. Cell Biol Int 2017;41:79–83.
- [18] Kehn K, Berro R, Alhaj A, et al. Functional consequences of cyclin D1/ BRCA1 interaction in breast cancer cells. Oncogene 2007;26:5060–9.
- [19] Bach DH, Long NP, Luu TT, et al. The dominant role of forkhead box proteins in cancer. Int J Mol Sci 2018;19:3279.
- [20] Zhang Z, Meng G, Wang L, et al. The prognostic role and reduced expression of FOXJ2 in human hepatocellular carcinoma. Mol Med Rep 2016;14:254–62.
- [21] Hauptmann S, Friedrich K, Redline R, et al. Ovarian borderline tumors in the 2014 WHO classification: evolving concepts and diagnostic criteria. Virchows Arch 2017;470:125–42.
- [22] Rauh-Hain JA, Melamed A, Wright A, et al. Overall survival following neoadjuvant chemotherapy vs primary cytoreductive surgery in women with epithelial ovarian cancer: analysis of the national cancer database. JAMA Oncol 2017;3:76–82.
- [23] Davis A, Tinker AV, Friedlander M. Platinum resistant ovarian cancer: what is it, who to treat and how to measure benefit? Gynecol Oncol 2014;133:624–31.
- [24] Fuhrich DG, Lessey BA, Savaris RF. Comparison of HSCORE assessment of endometrial beta3 integrin subunit expression with digital HSCORE using computerized image analysis (ImageJ). Anal Quant Cytopathol Histpathol 2013;35:210–6.
- [25] Gülşen Gürgen S, Yücel AT, Umur N, et al. Immunohistochemical changes after metoclopramide administration in rat brain cells. Proceedings 2018;2018:1545.