

Research Article

The Effect of Probiotics, Prebiotics, and Synbiotics on CD4 Counts in HIV-Infected Patients: A Systematic Review and Meta-Analysis

Yuan-Sheng Fu ¹, Qin-Shu Chu ¹, Akililu Alemu Ashuro ¹, Dong-Sheng Di ¹,
Qi Zhang ¹, Xue-Mei Liu ², and Yin-Guang Fan ¹

¹Department of Epidemiology and Biostatistics, School of Public Health, Anhui Medical University, 81 Meishan Road, Hefei, Anhui 230032, China

²Liuzhou Center for Disease Control and Prevention, 1 Tanzhongxi Road, Liuzhou, Guangxi Zhuang Autonomous Region 545000, China

Correspondence should be addressed to Yin-Guang Fan; fanyingguang@163.com

Received 7 July 2020; Revised 23 October 2020; Accepted 13 November 2020; Published 27 November 2020

Academic Editor: Washington L. C. dos Santos

Copyright © 2020 Yuan-Sheng Fu et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Background. Probiotics as a potential adjuvant therapy may improve the restoration of the intestinal CD4⁺ T-cell population in HIV-infected patients, whereas findings from clinical trials are inconsistent. This systematic review and meta-analysis of randomized controlled trials (RCTs) was performed to quantify the effects of probiotic, prebiotic, and synbiotic supplementation on CD4 counts in HIV-infected patients. **Methods.** We searched PubMed, Embase, Web of Science, Scopus, and the Cochrane Central Register of Controlled Trials for relevant articles published up to March 20, 2020. Two authors independently performed the study selection, data extraction, and risk of bias assessment. Data were pooled by using the random effects model, and weighted mean difference (WMD) was considered the summary effect size. Publication bias was evaluated by a funnel plot and Egger's test. **Results.** The search strategy identified 1712 citations. After screening, a total of 16 RCTs with 19 trials were included in the meta-analysis. Pooling of the extracted data indicated no significant difference between the probiotics/prebiotics/synbiotics and placebo groups on CD4 counts (WMD = 3.86, 95% confidence interval (CI) -24.72 to 32.45, $P = 0.791$). In subgroup analysis, a significant increase in CD4 counts was found in the study with high risk of bias (WMD = 188, 95% CI 108.74 to 227.26, $P \leq 0.001$). Egger's test showed no evidence of significant publication bias ($P = 0.936$). **Conclusions.** In summary, the evidence for the efficacy of probiotics, prebiotics, and synbiotics in improving HIV-infected patients' CD4 counts as presented in currently published RCTs is insufficient. Therefore, further comprehensive studies are needed to reveal the exact effect of probiotics, prebiotics, and synbiotics on CD4⁺ cell counts.

1. Introduction

Individuals living with human immunodeficiency virus (HIV) are characterized by progressive CD4⁺ T-cell depletion and immunodeficiency [1]. HIV infection alters gut microbial ecology [2], and a huge gastrointestinal (GI) pathology is observed even during primary infection. HIV enteropathy includes pronounced gut-associated CD4⁺ T-cell loss and an impaired gastrointestinal (GI) epithelial barrier [3–5]. These detrimental changes presumably result in microbial translo-

cation and a loss of gut homeostasis [1, 6, 7], which in turn leads to chronic immune activation and disease progression [8, 9]. In addition, the efficacy of antiretroviral treatment in the GI tract seems to be poor, resulting in insufficient reconstitution of CD4⁺ T cells and incomplete viral suppression [10–12]. In view of the key role of decreasing bacterial translocation and proinflammatory cytokine production in the maintenance of gut homeostasis, new therapies aimed at restoring the integrity of the epithelial and gut-associated lymphoid tissue (GALT) through oral prebiotics, probiotics,

or synbiotics, as well as improving chronic immune activation, are promising new strategies to alleviate disease progression of HIV patients.

Probiotics are defined as “live microorganisms which, when administered in adequate amounts, confer a health benefit on the host” [13] and have an effect on the immunological response. They mainly stimulate the secretion of polymeric IgA, avoid the overgrowth and translocation of bacteria, and promote the development of regulatory T (Treg) cells through the production of anti-inflammatory cytokines [14–17]. Related to probiotics are prebiotics, indigestible food ingredients, generally oligosaccharides, that improve host health by selectively stimulating the growth of beneficial bacteria in the colon, such as *Bifidobacteria* and *Lactobacilli* [18, 19]. Prebiotics can increase the production of short-chain fatty acids (SCFAs), thereby reducing inflammation [20]. A study in mice also showed that prebiotics had an immunostimulatory effect on the induced site [21]. Synbiotics are products that combine prebiotics and probiotics, with a potentially synergistic action. Given the evidence of the beneficial effects of probiotic, prebiotic, and synbiotic consumption during the course of different viral infections and noninfectious diseases [22–25], a growing body of studies try to prove that the use of probiotics, prebiotics, and synbiotics may be able to help preserve the immune function of HIV patients and consequently prevent the depletion of CD4⁺ T cells. However, the results are inconsistent across different studies [26–29]. Therefore, we conducted a systematic review and meta-analysis of available RCTs to evaluate the effect of probiotics, prebiotics, and synbiotics on CD4 counts in HIV patients.

2. Materials and Methods

2.1. Search Strategy. This systematic review and meta-analysis was conducted in accordance with the guidelines of the Cochrane Handbook [30] and was reported in compliance with the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) statement [31]. We searched PubMed, Embase, Web of Science, Scopus, and the Cochrane Central Register of Controlled Trials for studies published before March 20, 2020. Studies were searched using the following search terms: (Probiotic OR Prebiotic OR Synbiotic OR *Lactobacillus* OR *Bifidobacterium* OR *Saccharomyces* OR “*Streptococcus thermophiles*” OR “fermented milk” OR “*Escherichia coli*”) AND (HIV/AIDS OR HIV OR AIDS OR “Human Immunodeficiency Virus” OR “Acquired Immunodeficiency Syndrome”) AND (Random OR Randomized OR “Randomized controlled trial” OR “controlled clinical trial” OR “randomized studies”). No restrictions were placed on the language and date. In addition, the references of the included articles were also screened to find other relevant publications.

2.2. Study Selection. Studies were included with the following criteria: (1) RCTs with parallel or cross-over design, (2) studies conducted in HIV-1-infected adults over 18 years of age, (3) intervention using probiotics, prebiotics, or synbiotics, (4) comparison with placebo or control groups, and (5)

CD4 counts as a primary or secondary outcome. Exclusion criteria were as follows: (1) nonrandomized clinical trials; (2) uncontrolled studies; (3) studies conducted in children or pregnant women; (4) letters, conference abstracts, case reports, reviews, or observational studies; or (5) studies not clearly reporting CD4 counts before or after the intervention. All studies were independently assessed by two authors, and any disagreement was resolved by a third researcher.

2.3. Data Extraction and Quality Assessment. The following data were extracted: first author’s name, year of publication, study design, country of study, sample size, age and gender of participants, details of interventions (including strain, dosage, and duration of intervention), intake of antiretroviral drugs or not, and the main results on the interested outcomes. For the missing data, the authors were contacted through e-mails to get relevant data. The methodological quality of included studies was evaluated by using the Cochrane Collaboration’s risk of bias tool [32]. The following domains were assessed: random sequence generation, allocation concealment, blinding of participants and personnel, blinding of outcome assessment, incomplete outcome data, selective reporting, and other bias. The risk of bias for each domain was judged as low, high, or unclear according to the Cochrane Handbook for Systematic Reviews. Any disagreements during the processes of data extraction and quality assessment were resolved by discussion. When consensus was not reached, a third investigator worked as an arbitrator.

2.4. Data Synthesis and Analysis. The mean difference (MD) and standard deviation (SD) of CD4 counts between the probiotics/prebiotics/synbiotics and control groups were used to estimate the pooled effects. For the trials that provided more than one interval results, the last intervention results were included in the analysis. And weighted mean difference (WMD) with 95% confidence interval (CI) was considered the summary effect size. Heterogeneity was assessed by Cochran’s Q test and *I*-square (I^2) statistic, and heterogeneity with an I^2 value > 50% or $P < 0.1$ was considered significant [33]. To account for heterogeneity between articles, a random effects model was applied in this meta-analysis. The subgroup analysis was also carried out according to the type of intervention, intake of antiretroviral drugs or not, duration of intervention, income of the country, and risk of bias assessment. Furthermore, a funnel plot and Egger’s linear regression were used to evaluate the potential publication bias. Meta-analysis was performed using Stata software version 14.0 (Stata Corp., College Station, TX, USA) and RevMan version 5.3 (Cochrane Collaboration, Oxford, UK). A two-tailed $P < 0.05$ was considered to be significant.

3. Results

A total of 1712 relevant articles were identified by searching the initial online databases. After duplicates were removed, the remaining 1247 studies were screened by title and abstract, 1182 of which were excluded, as they did not meet the eligibility criteria. The full text of the remaining 65 records was retrieved, and 16 studies (19 trials) that fulfilled

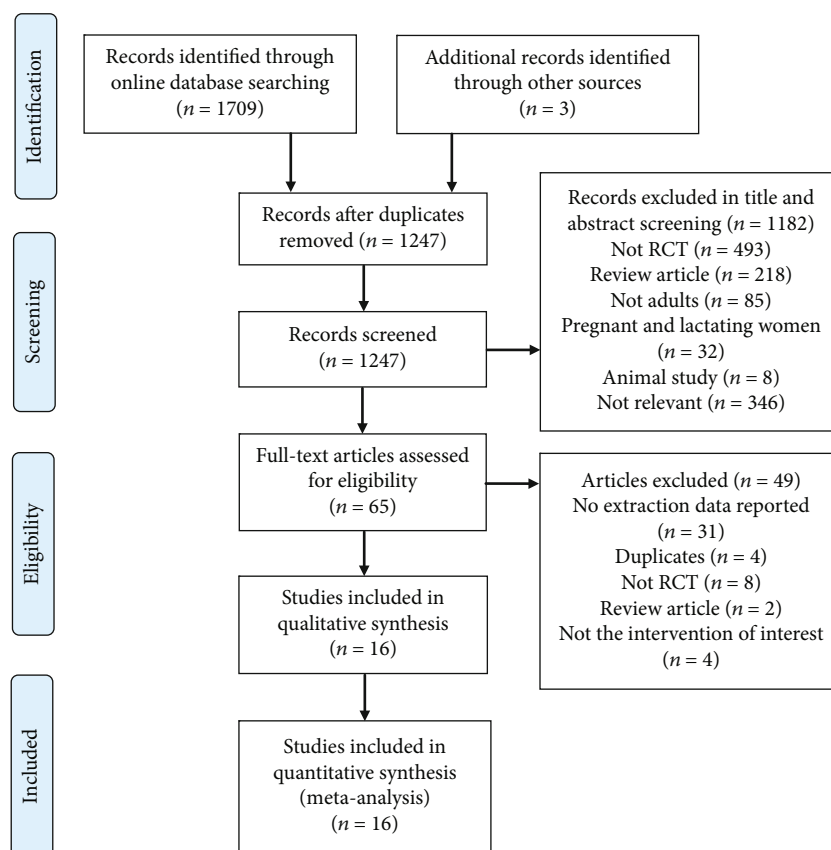


FIGURE 1: Flowchart of study selection.

the inclusion criteria were included in the systematic review and meta-analysis [2, 26–29, 34–44]. The process of study selection and reasons for exclusion are presented in Figure 1.

3.1. Characteristics of the Included Studies. The majority of the studies were randomized, double-blind, placebo-controlled trials except one [40] randomized, nonblinded, placebo-controlled trial; one [42] randomized, triple-blind, placebo-controlled trial; and one [39] randomized, double-blind, cross-over placebo-controlled trial. These studies were published from 1998 to 2020, with sample sizes ranging from 10 to 340 individuals. The duration of intervention varied from 15 days to 52 weeks. Twelve trials administered probiotics [2, 26, 28, 34–36, 38–40, 42–44], while four trials administered prebiotics [27, 29, 44] and three trials administered synbiotics [37, 41, 44]. All of the included clinical trials were with two-arm parallel design except two studies [29, 44] which were with three-arm and four-arm parallel design. The three-arm and four-arm parallel design studies were considered two and three trials. The characteristics of the enrolled studies are summarized in Table 1.

3.2. Risk of Bias Assessment. The risk of bias of the included studies is presented in Figure 2. Among the 19 trials, seven [26, 28, 29, 34, 37, 38] were judged to have a low risk of bias; eleven [2, 27, 35, 36, 39, 41–44] were categorized as having unclear risk and one [40] as having high risk of bias. All the included trials achieved adequate random sequence

generation and blinding of outcome assessment. Seven [2, 27, 39–43] studies provided no description of allocation concealment procedure, and six [27, 36, 41, 44] studies were rated to have unclear risk of selective reporting bias. Attrition bias was found in one study [35] due to loss of participants during the study period. All of the trials except for one [40] had a high risk of bias in blindness of participants and key study personnel.

3.3. Meta-Analysis: Main Results. In total, 16 RCTs with 19 treatment arms were included in the meta-analysis. Due to the relatively high heterogeneity among the included studies ($I^2 = 55.7\%$, $P = 0.002$), a random effects model was selected for quantitative synthesis. Overall, the pooled results indicated no significant difference after probiotic, prebiotic, and synbiotic supplementation in comparison with the placebo controls on CD4 counts (WMD = 3.86, 95% CI: –24.72 to 32.45, $P = 0.791$). The forest plot of the meta-analysis is shown in Figure 3.

3.4. Subgroup Analysis. Because of the existence of heterogeneity, subgroup analysis was conducted based on the type of intervention (probiotics vs. prebiotics vs. synbiotics), duration of intervention (<30 vs. ≥30 days), intake of antiretroviral drugs or not (yes vs. no), income of country (high vs. low and middle), and risk of bias assessment (low vs. unclear vs. high). The result of subgroup analysis for trials with high risk of bias showed a significant increase in the CD4 counts

TABLE 1: Characteristics of the included randomized controlled trials.

Study (year)	Study design	Country	Sample size	Age (years)	Sex	Duration	ARV	Intervention (strain and daily dose)	Main outcome measures
Wolf et al. (1998)	Randomized, double-blind, placebo-controlled trial	USA	35	23 to 50	M (95%)	21 d	Not on ARV	<i>Lactobacillus reuteri</i> (10^{10} cfu/day)	Serum chemistry, hematology, immune profile, urinalysis, physical examination
Heiser et al. (2004)	Randomized, controlled trial	USA	35	42.6 ± 7.4	M (100%)	12 wk	All on ARV	<i>Acidophilus</i> and <i>Bifidobacteria</i> (1.2 g/d) and soluble fiber (Proctor & Gamble, Cincinnati, 11 g/d)	Diarrhea, CD4 count, HIV RNA
Anukam et al. (2008)	Randomized, triple-blind, placebo-controlled trial	Nigeria	23	18 to 44	F (100%)	15 d	Not on ARV	Probiotic yogurt containing <i>Lactobacillus rhamnosus</i> GR-1 and <i>Lactobacillus reuteri</i> RC-14 (2.5×10^9 cfu/day)	Hematologic profiles, CD4 count, QoL
Gori et al. (2011a)	Randomized, double-blind, placebo-controlled trial	Italy	31	38.3 ± 9.5	M (66%)	12 wk	ARV naive	Short chain galactooligosaccharides/long-chain fructooligosaccharides/pectin hydrolysate-derived acidic oligosaccharides (15 g/d)	Gut microbiota composition, immunological markers, LPS, sCD14, NK cell activity
Gori et al. (2011b)	Randomized, double-blind, placebo-controlled trial	Italy	33	38.3 ± 9.5	M (76%)	12 wk	ARV naive	Short-chain galactooligosaccharides/long-chain fructooligosaccharides/pectin hydrolysate-derived acidic oligosaccharides (30 g/d)	Gut microbiota composition, immunological markers, LPS, sCD14, NK cell activity
Hummelen et al. (2011)	Randomized, double-blind, placebo-controlled trial	Tanzania	53	NA	F (100%)	25 wk	Not on ARV	<i>Lactobacillus rhamnosus</i> GR-1 and <i>Lactobacillus reuteri</i> RC-14 (2×10^9 cfu/day)	CD4 count, immune markers (IgG, IgE, IFN- γ , and IL-10)
Hummelen et al. (2011)	Randomized, double-blind, placebo-controlled trial	Tanzania	111	NA	F (86%)	4 wk	ARV naive	<i>Lactobacillus rhamnosus</i> GR-1 (12.5×10^{10} cfu/day) and micronutrients	CD4 count, hematology indicators
Hemsworth et al. (2012)	Randomized, double-blind, cross-over controlled trial	Canada	42	47.6 ± 9.3	M (75%)	30 d	All on ARV	Yogurt containing micronutrients and <i>Lactobacillus rhamnosus</i> CAN-1 (minutes 10^9 cfu/mL)	Immunologic parameters, nutritional and biochemical parameters
Schunter et al. (2012)	Randomized, double-blind, placebo-controlled trial	USA	27	47.5	M (100%)	4 wk	All on ARV	A symbiotic consists of 4 strains of probiotic bacteria (10^{10} each) plus 4 nondigestible, fermentable dietary fibers (2.5 g each)	Bacterial translocation, CD4 ⁺ T-cells, CD8 ⁺ T-cells, CRP, sCD14
Gonzalez-Hernandez et al. (2012a)	Randomized, double-blind, placebo-controlled trial	Mexico	10	18 to 65	M (90%)	16 wk	ARV naive	<i>Lactobacillus rhamnosus</i> HN001 plus <i>Bifidobacterium lactis</i> Bi-07 at 10^9 cfu/mL	Safety, QoL, CD4 count, cytokine level

TABLE 1: Continued.

Study (year)	Study design	Country	Sample size	Age (years)	Sex	Duration	ARV	Intervention (strain and daily dose)	Main outcome measures
Gonzalez-Hernandez et al. (2012b)	Randomized, double-blind, placebo-controlled trial	Mexico	10	18 to 65	M (90%)	16 wk	ARV naive	10 g fructooligosaccharides (FOS)	Safety, QoL, CD4 count, cytokine level
Gonzalez-Hernandez et al. (2012c)	Randomized, double-blind, placebo-controlled trial	Mexico	10	18 to 65	M (100%)	16 wk	ARV naive	(<i>Lactobacillus rhamnosus</i> HN001 plus <i>Bifidobacterium lactis</i> Bi-07 at 10 ⁹ cfu/mL)+10 g FOS	Safety, QoL, CD4 count, cytokine level
Cahn et al. (2013)	Randomized, double-blind, placebo-controlled trial	Italy, Netherlands, UK, Thailand, US, Brazil, Argentina, Australia	340	39.6	M (82%)	52 wk	Not on ARV	Oligosaccharides (short-chain GOS, long-chain FOS, and pectin-derived AOS) and micronutrients	CD4 count, plasma viral load, safety, and tolerability
Yang et al. (2014)	Randomized, double-blind, placebo-controlled trial	USA	17	49.6 ± 8.7	M (94%)	90 d	All on ARV	<i>Bacillus coagulans</i> GBI-30, 6086 (2 × 10 ⁹ cfu/day)	CD4 count, CD4 percentage, proinflammatory blood biomarkers
Stiksrud et al. (2015)	Randomized, double-blind, placebo-controlled trial	Norway, Sweden	24	50.8	M (67%)	8 wk	All on ARV	Fermented skimmed milk supplemented with <i>Lactobacillus rhamnosus</i> GG (10 ⁸ cfu/mL), <i>Bifidobacterium animalis</i> subsp. <i>Lactis</i> B-12 (10 ⁸ cfu/mL), and <i>Lactobacillus acidophilus</i> La-5 (10 ⁷ cfu/mL)	CD4 count, CD4/CD8 ratio, soluble inflammation markers, D-dimer, LPS, sCD14
Villar-Garcia et al. (2015)	Randomized, double-blind, placebo-controlled trial	Spain	44	47.5	M (84%)	12 wk	All on ARV	<i>Saccharomyces boulardii</i> (2 capsules 3 times a day or 6 × 10 ⁷ living bacteria)	Microbial translocation and inflammation markers, immunological and clinical data
Adriana et al. (2016)	Randomized, double-blind, placebo-controlled trial	USA	73	51	M (86%)	22 wk	All on ARV	Probiotic Visbiome Extra Strength	sCD14, IL-6, CD4 count, CD4/CD8 ratio, sCD163
Serrano-Villar et al. (2019)	Randomized, double-blind, placebo-controlled trial	Spain	59	38	M (92%)	48 wk	ARV naive	PMT25341 (a mixture of prebiotics, probiotics, oligonutrients, essential amino acids, omega-3 fatty acids)	CD4 count, CD4/CD8 ratio, markers of T-cell activation, bacterial translocation, inflammation
Tenore et al. (2020)	Randomized, double-blind, placebo-controlled trial	Brazil	48	44.5	M (90%)	12 wk	All on ARV	<i>Lactobacillus casei</i> Shirota	CD4 count, CD4/CD8 ratio, levels of CD4 ⁺ and CD8 ⁺ T-cell activation, sCD14

NA: not available; ARV: antiretroviral; F: female; M: male; d: day; wk: week; NK: natural killer; LPS: lipopolysaccharide; IFN-γ: interferon-γ; QoL: quality of life; IL-10: interleukin-10; CRP: C-reactive protein.

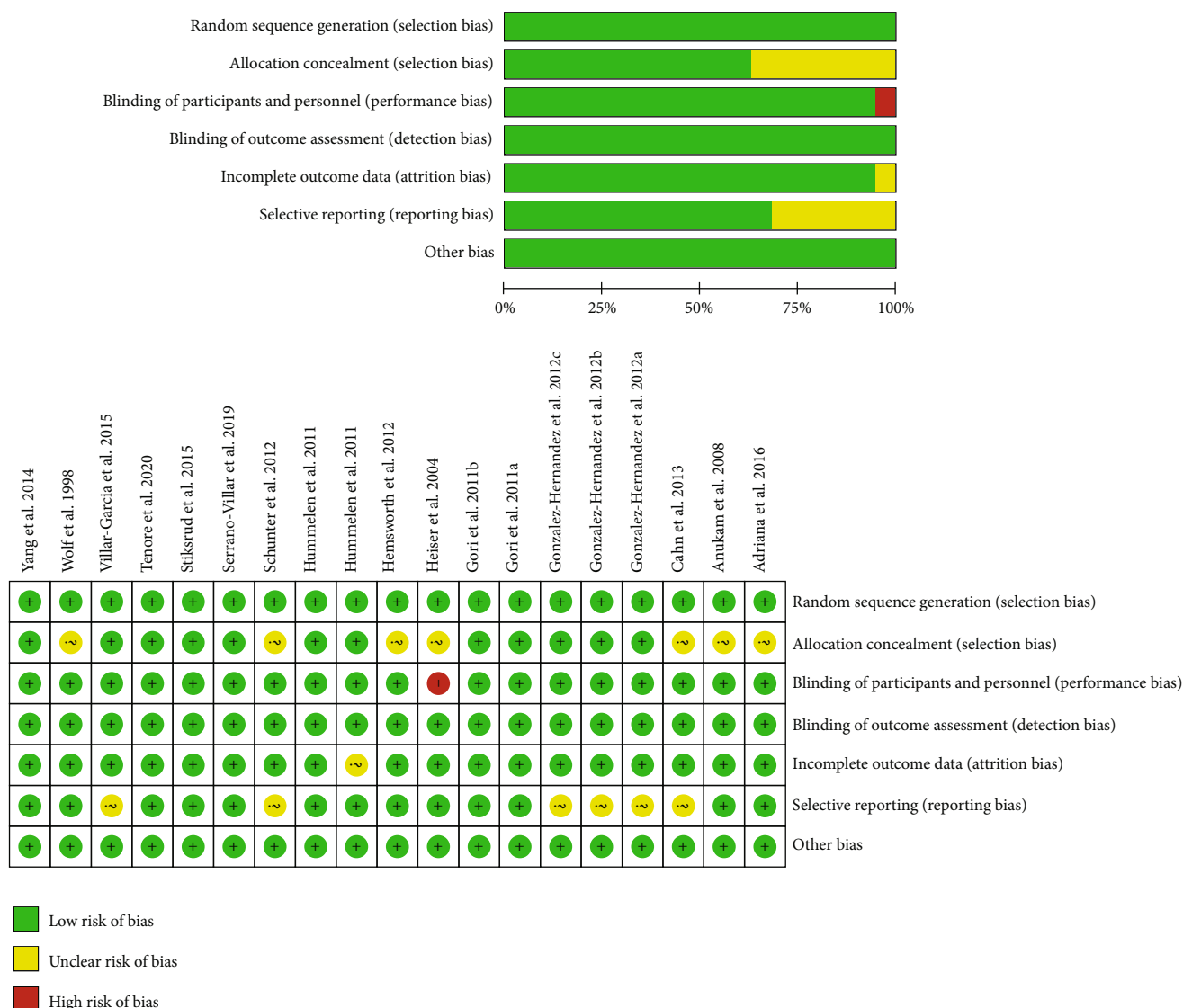


FIGURE 2: Risk of bias and its summary for the included trials.

compared to that with low and unclear risk of bias (WMD = 188; 95% CI, 108.74, 227.26; $P \leq 0.001$). However, the other subgroup analysis revealed that none of the subgroups achieved statistical significance. A summary of the results of subgroup analysis is shown in Table 2.

3.5. Sensitivity Analysis and Publication Bias. Sensitivity analysis was carried out by removing studies one by one to test the reliability of the results of meta-analysis. The results in Figure 4 showed that no matter which study was omitted, the overall statistical significance does not change. In addition, excluding these studies that provide more than one interval results does not change the significance of the findings (WMD = 4.28, 95% CI: -30.88 to 39.44, $P = 0.81$). Publication bias was assessed by a funnel plot and the result of Egger’s test. Visual inspection of the funnel plot showed that RCTs are symmetrically scattered around the null vertical line, suggesting no bias (Figure 5). Egger’s regression

intercept test confirmed that there was no significant publication bias ($P = 0.936$).

4. Discussion

In this study, we reviewed and performed a systematic review and meta-analysis to assess the effect of probiotic, prebiotic, and synbiotic supplementation on CD4 counts in HIV-infected patients. The results of our meta-analysis show that these interventions did not cause any significant change on the CD4 counts. In subgroup analysis, a significant increase in CD4 counts was found in studies with high risk of bias. However, subgroup analysis based on the type of intervention, intake of antiretroviral drugs or not, duration of intervention, and the income of the country of the included studies revealed no significant findings. Egger’s test showed that the potential risk of publication bias is low, and sensitivity analysis supports the reliability of the results.

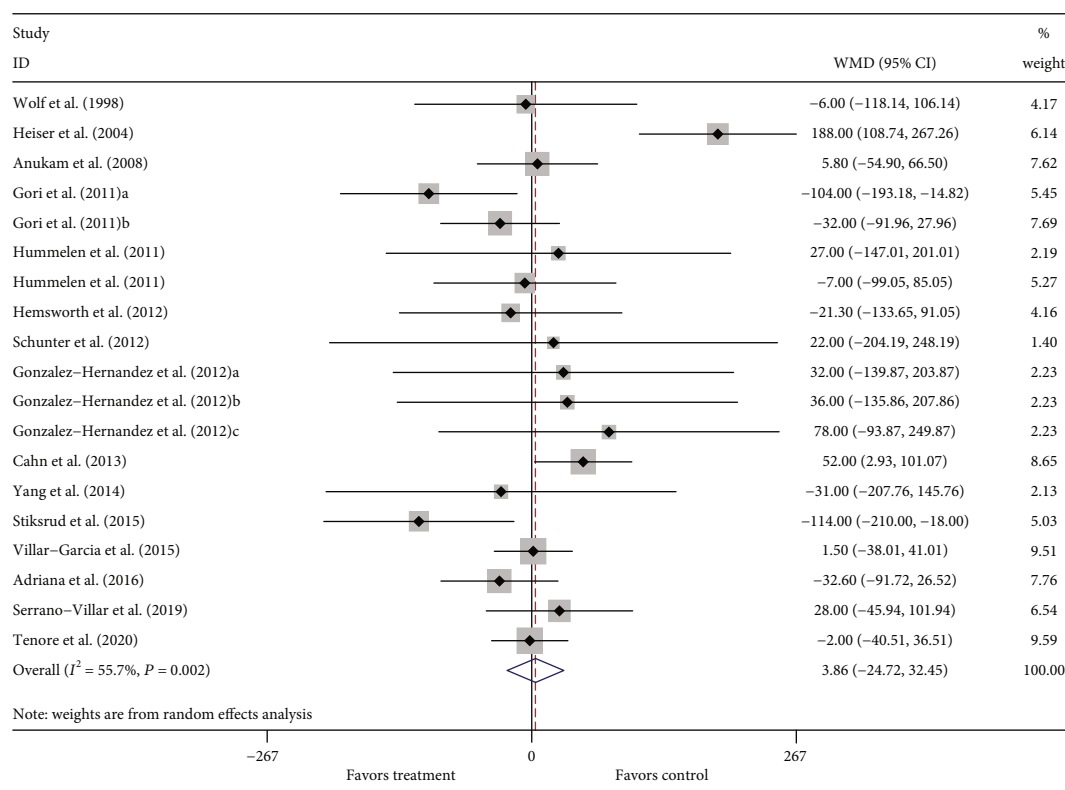


FIGURE 3: Forest plot of the effect of probiotic, prebiotic, and synbiotic supplementation on CD4 counts. The square in the figure represents the effect of the study, and the size of the square represents the weight of the study. The horizontal line represents the confidence interval of the effect value. The diamond in the figure represents the pooled effect. WMD: weighted mean difference; CI: confidence interval.

TABLE 2: Summary of subgroup analysis.

Subgroup	No. of trials	WMD	95% CI	P	Weight	I^2 (%)	P for heterogeneity	P for subgroup difference
Intervention type								
Probiotics	12	4.23	(-33.02, 41.47)	0.824	65.80	61.6	0.003	0.60
Prebiotics	4	-13.80	(-87.87, 60.28)	0.715	24.30	72.2	0.013	
Synbiotics	3	34.67	(-30.38, 99.72)	0.296	10.17	0.0	0.866	
Duration								
<30 days	4	1.44	(-43.81, 46.68)	0.950	18.46	0.0	0.992	0.91
≥30 days	15	4.57	(-30.48, 39.62)	0.798	81.54	65.5	≤0.001	
Intake of ARV								
Yes	9	6.94	(-40.38, 54.26)	0.774	52.25	72.3	≤0.001	0.83
No	10	0.81	(-32.18, 33.79)	0.962	47.45	23.7	0.225	
Income of country								
High	4	-1.33	(-44.16, 41.50)	0.952	68.62	72.1	≤0.001	0.83
Low and middle	15	4.37	(-24.52, 33.26)	0.767	31.38	0.0	0.979	
Risk of bias assessment								
Low	7	-29.01	(-65.70, 7.69)	0.121	41.70	38.7	0.134	≤0.001
Unclear	11	9.82	(-12.97, 32.60)	0.399	52.16	0.0	0.800	
High	1	188	(108.74, 227.26)	≤0.001	6.14	NA	NA	

WMD: weighted mean difference; CI: confidence interval; NA: not available; ARV: antiretroviral.

These findings are counterintuitive because they appear to be inconsistent with some previous studies [17, 27, 45]. HIV infection dramatically alters the intestinal environ-

ment, leading to significant changes in the structural and functional characteristics of the intestinal tract, including microbial translocation and gut inflammation [46-49].

placebo groups in improving CD4 counts. One possible explanation is that in our meta-analysis, the absolute CD4⁺ T cell was reported as a predictor of immune status and disease progression and to be used in quantitative synthesis; however, the CD4⁺ percentage of total T cells as a strong independent predictor of immune status and disease progression [51] may be a more appropriate indicator for comparison. In addition, probiotics are not pharmaceutical substances. Probiotics can be administered as single strains or combination compounds, but different strains produce varied effects and how the single strains interact when co-administered was unclear. Moreover, the dose-response curves of most strains have not been described [52]. In summary, the heterogeneity of probiotic application and the limitations in medicine have hampered the scientific quality of clinical research on probiotics.

Though the interesting outcome in this review is CD4 counts, several included studies [28, 29, 36, 38, 39, 41] also reported related results, namely, gut inflammation and microbial translocation levels, both of them were known to be associated with the progression and prognosis of HIV infection [8–10]. It should be mentioned that many of the trials reported only an improvement in one or two markers of inflammation, while there was no significant difference in the rest of the analysis. Similarly, very few of the studies [2, 26, 28, 37, 41, 43] have evaluated the level of immune markers such as CD8 counts and the CD4/CD8 ratio, which has been considered a prognostic parameter of non-AIDS morbidity [53, 54]. In summary, since the specific mechanism of probiotics in the gut repair is not clear, tracking these outcomes with CD4 counts may yield new and interesting findings, which may provide a broader perspective on the therapeutic potential of probiotics, prebiotics, and synbiotics in HIV patients.

Subgroup analysis revealed that risk of bias assessment may be the source of heterogeneity. However, other subgroup analysis failed to explain the heterogeneity between studies. Also, it should be mentioned that the effects of prebiotic, probiotic, and synbiotic intervention on CD4 counts were statistically significant in trials with high risk of bias. Since only one study was included in the subgroup with high risk of bias and the quality of the study was relatively low due to the failure of blind implementation, the results were hampered with uncertainty.

Our results are different from previously published systematic reviews [55, 56]. Of note, two of the included studies reported improvement of CD4 counts among those receiving probiotic supplements [38, 42]; however, after analysis according to the data inclusion criteria of our meta-analysis, the result showed no significant difference. In addition, compared to previous reviews, our analysis specifically focused on RCTs and adult patients (≥ 18 years), and we performed a more comprehensive analysis on available evidences that may potentially be involved in the efficacy of probiotic administration on CD4 counts in HIV-infected patients. Our meta-analysis also included the updated references that have not been analyzed in other meta-analysis [26, 27, 37, 43, 57]. These reasons may cause our findings to be inconsistent with other reviews. This meta-analysis has some limita-

tions. First, most of the included trials had relatively small sample sizes, which may lead to an underestimation of the intervention effect; therefore, large-scale trials are warranted. Second, heterogeneity exists between studies in regard to applied probiotic strain(s) and dosage. Therefore, future studies with more high-quality trials are recommended to determine the ideal number and combination of species or strains and their ideal dose for use in probiotic supplements. Third, different formations of administration (yogurt, milk, and capsule) were used in the included trials. Though in vitro analysis of the activity of the probiotics from yogurt or capsules did not differ [34], there may be discrepancies in the survival and colonization of probiotic strains in the intestinal tract.

5. Conclusion

In summary, the results of this meta-analysis suggest that the evidence for the efficacy of probiotics, prebiotics, and synbiotics in improving HIV-infected patients' CD4 counts from current RCTs is insufficient. The promotion of these interventions for the benefit of HIV-infected patients in clinical subjects should be implemented only when more valid evidence in this area is obtained. Future clinical studies with a well design and large sample size are needed to further elucidate probiotics, prebiotics, and synbiotics' mechanisms of action, safety profile, and clinical potential, in both support immune system reconstitution and longer-term health outcomes on HIV-infected patients.

Data Availability

The data is available upon request.

Conflicts of Interest

The authors declare no conflict of interest.

Authors' Contributions

Y.S.F., Q.S.C., and Y.G.F. designed the study. Y.S.F., Q.S.C., and X.M.L. searched databases and performed the selection of studies. Y.S.F., A.A.A., and Q.Z. analyzed the data and wrote the manuscript. Y.G.F. and D.S.D. critically evaluated the review and commented on it. The final version was confirmed by all the authors for submission. Yuan-Sheng Fu and Qin-Shu Chu contributed equally to this work.

Acknowledgments

This study was supported by Grants for Scientific Research of BSKY (xj201526) from Anhui Medical University and the program of technical support for Liuzhou City (K2020043) authorized by the Chinese National Center for AIDS/STD Control and Prevention.

Supplementary Materials

Detailed search strategy. Figure Legends (*Supplementary materials*)

References

- [1] J. W. Mellors, A. Muñoz, J. V. Giorgi et al., "Plasma viral load and CD4+ lymphocytes as prognostic markers of HIV-1 infection," *Annals of Internal Medicine*, vol. 126, no. 12, pp. 946–954, 1997.
- [2] B. W. Wolf, K. B. Wheeler, D. G. Ataya, and K. A. Garleb, "Safety and tolerance of Lactobacillus reuteri supplementation to a population infected with the human immunodeficiency virus," *Food and chemical toxicology : an international journal published for the British Industrial Biological Research Association*, vol. 36, no. 12, pp. 1085–1094, 1998.
- [3] J. M. Brenchley, T. W. Schacker, L. E. Ruff et al., "CD4+ T cell depletion during all stages of HIV disease occurs predominantly in the gastrointestinal tract," *The Journal of Experimental Medicine*, vol. 200, no. 6, pp. 749–759, 2004.
- [4] S. Mehandru, M. A. Poles, K. Tenner-Racz et al., "Primary HIV-1 infection is associated with preferential depletion of CD4+ T lymphocytes from effector sites in the gastrointestinal tract," *The Journal of Experimental Medicine*, vol. 200, no. 6, pp. 761–770, 2004.
- [5] A. Nazli, O. Chan, W. N. Dobson-Belaire et al., "Exposure to HIV-1 directly impairs mucosal epithelial barrier integrity allowing microbial translocation," *PLoS Pathogens*, vol. 6, no. 4, 2010.
- [6] J. M. Brenchley, D. A. Price, T. W. Schacker et al., "Microbial translocation is a cause of systemic immune activation in chronic HIV infection," *Nature Medicine*, vol. 12, no. 12, pp. 1365–1371, 2006.
- [7] A. Gori, C. Tincati, G. Rizzardini et al., "Early impairment of gut function and gut flora supporting a role for alteration of gastrointestinal mucosa in human immunodeficiency virus pathogenesis," *Journal of Clinical Microbiology*, vol. 46, no. 2, pp. 757–758, 2008.
- [8] G. Marchetti, A. Cozzi-Lepri, E. Merlini et al., "Microbial translocation predicts disease progression of HIV-infected antiretroviral-naïve patients with high CD4+ cell count," *AIDS*, vol. 25, no. 11, pp. 1385–1394, 2011.
- [9] S. Reus, J. Portilla, J. Sánchez-Payá et al., "Low-Level HIV Viremia Is Associated With Microbial Translocation and Inflammation," *JAIDS Journal of Acquired Immune Deficiency Syndromes*, vol. 62, no. 2, pp. 129–134, 2013.
- [10] G. Marchetti, G. M. Bellistri, E. Borghi et al., "Microbial translocation is associated with sustained failure in CD4+ T-cell reconstitution in HIV-infected patients on long-term highly active antiretroviral therapy," *AIDS*, vol. 22, no. 15, pp. 2035–2038, 2008.
- [11] M. Massanella, E. Negredo, N. Pérez-Álvarez et al., "CD4 T-cell hyperactivation and susceptibility to cell death determine poor CD4 T-cell recovery during suppressive HAART," *AIDS*, vol. 24, no. 7, pp. 959–968, 2010.
- [12] E. Merlini, F. Bai, G. M. Bellistri, C. Tincati, A. d'Arminio Monforte, and G. Marchetti, "Evidence for polymicrobial flora translocating in peripheral blood of HIV-infected patients with poor immune response to antiretroviral therapy," *PLoS One*, vol. 6, no. 4, 2011.
- [13] C. Hill, F. Guarner, G. Reid et al., "Expert consensus document. The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic," *Gastroenterology & hepatology*, vol. 11, no. 8, pp. 506–514, 2014.
- [14] A. C. Senok, A. Y. Ismaeel, and G. A. Botta, "Probiotics: facts and myths," *Clinical Microbiology and Infection : The Official Publication of the European Society of Clinical Microbiology and Infectious Diseases*, vol. 11, no. 12, pp. 958–966, 2005.
- [15] S. Parvez, K. A. Malik, S. Ah Kang, and H. Y. Kim, "Probiotics and their fermented food products are beneficial for health," *Journal of Applied Microbiology*, vol. 100, no. 6, pp. 1171–1185, 2006.
- [16] R. B. Sartor, "Microbial influences in inflammatory bowel diseases," *Gastroenterology*, vol. 134, no. 2, pp. 577–594, 2008.
- [17] L. Trois, E. M. Cardoso, and E. Miura, "Use of probiotics in HIV-infected children: a randomized double-blind controlled study," *Journal of Tropical Pediatrics*, vol. 54, no. 1, pp. 19–24, 2008.
- [18] G. R. Gibson and M. B. Roberfroid, "Dietary modulation of the human colonic microbiota: introducing the concept of prebiotics," *The Journal of Nutrition*, vol. 125, no. 6, pp. 1401–1412, 1995.
- [19] G. R. Gibson, R. Hutkins, M. E. Sanders et al., "Expert consensus document: the International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of prebiotics," *Gastroenterology & hepatology*, vol. 14, no. 8, pp. 491–502, 2017.
- [20] R. J. F. Felizardo, I. K. M. Watanabe, P. Dardi, L. V. Rossoni, and N. O. S. Câmara, "The interplay among gut microbiota, hypertension and kidney diseases: the role of short-chain fatty acids," *Pharmacological Research*, vol. 141, pp. 366–377, 2019.
- [21] N. Manhart, A. Spittler, H. Bergmeister, M. Mittlböck, and E. Roth, "Influence of fructooligosaccharides on Peyer's patch lymphocyte numbers in healthy and endotoxemic mice," *Nutrition*, vol. 19, no. 7-8, pp. 657–660, 2003.
- [22] X. Xia, J. Chen, J. Xia et al., "Role of probiotics in the treatment of minimal hepatic encephalopathy in patients with HBV-induced liver cirrhosis," *The Journal of International Medical Research*, vol. 46, no. 9, pp. 3596–3604, 2018.
- [23] S. R. Sarkar and S. Banerjee, "Gut microbiota in neurodegenerative disorders," *Journal of Neuroimmunology*, vol. 328, pp. 98–104, 2019.
- [24] E. Palma, N. Recine, L. Domenici, M. Giorgini, A. Pierangeli, and P. B. Panici, "Long-term Lactobacillus rhamnosus BMX 54 application to restore a balanced vaginal ecosystem: a promising solution against HPV-infection," *BMC Infectious Diseases*, vol. 18, no. 1, p. 13, 2018.
- [25] S. K. Sarin, A. Pande, and B. Schnabl, "Microbiome as a therapeutic target in alcohol-related liver disease," *Journal of Hepatology*, vol. 70, no. 2, pp. 260–272, 2019.
- [26] S. de Barros Tenore, V. I. Avelino-Silva, P. R. Costa et al., "Immune effects of Lactobacillus casei Shirota in treated HIV-infected patients with poor CD4+ T-cell recovery," *AIDS*, vol. 34, no. 3, pp. 381–389, 2020.
- [27] P. Cahn, K. Ruxrungtham, B. Gazzard et al., "The immunomodulatory nutritional intervention NR100157 reduced CD4 + T-cell decline and immune activation: a 1-year multicenter randomized controlled double-blind trial in HIV-infected persons not receiving antiretroviral therapy (the BITE study)," *Clinical Infectious Diseases : An Official Publication of the Infectious Diseases Society of America*, vol. 57, no. 1, pp. 139–146, 2013.
- [28] B. Stiksrud, P. Nowak, F. C. Nwosu et al., "Reduced Levels of D-dimer and Changes in Gut Microbiota Composition After Probiotic Intervention in HIV-Infected Individuals on Stable

- ART," *JAIDS Journal of Acquired Immune Deficiency Syndromes*, vol. 70, no. 4, pp. 329–337, 2015.
- [29] A. Gori, G. Rizzardini, B. van't Land et al., "Specific prebiotics modulate gut microbiota and immune activation in HAART-naive HIV-infected adults: results of the "COPA" pilot randomized trial," *Mucosal Immunology*, vol. 4, no. 5, pp. 554–563, 2011.
- [30] J. P. H. S. V. Fellow and J. J. Deeks, *Selecting Studies and Collecting Data*, John Wiley & Sons, Ltd, 2008.
- [31] D. Moher, "Preferred Reporting Items for Systematic Reviews and Meta-Analyses: The PRISMA Statement," *Annals of Internal Medicine*, vol. 151, no. 4, pp. 264–9, W64, 2009.
- [32] J. P. T. Higgins, D. G. Altman, P. C. Gotzsche et al., "The Cochrane Collaboration's tool for assessing risk of bias in randomised trials," *BMJ*, vol. 343, no. oct18 2, p. d5928, 2011.
- [33] J. P. T. Higgins and S. G. Thompson, "Quantifying heterogeneity in a meta-analysis," *Statistics in Medicine*, vol. 21, no. 11, pp. 1539–1558, 2002.
- [34] R. Hummelen, J. Chandalucha, N. L. Butamanya et al., "Effect of 25 weeks probiotic supplementation on immune function of HIV patients," *Gut Microbes*, vol. 2, no. 2, pp. 80–85, 2014.
- [35] R. Hummelen, J. Hemsworth, J. Chandalucha et al., "Effect of micronutrient and probiotic fortified yogurt on immune-function of anti-retroviral therapy naive HIV patients," *Nutrients*, vol. 3, no. 10, pp. 897–909, 2011.
- [36] J. Villar-García, J. J. Hernández, R. Güerri-Fernández et al., "Effect of Probiotics (*Saccharomyces boulardii*) on Microbial Translocation and Inflammation in HIV-Treated Patients," *JAIDS Journal of Acquired Immune Deficiency Syndromes*, vol. 68, no. 3, pp. 256–263, 2015.
- [37] S. Serrano-Villar, M. de Lagarde, J. Vázquez-Castellanos et al., "Effects of immunonutrition in advanced human immunodeficiency virus disease: a randomized placebo-controlled clinical trial (Promaltia Study)," *Clinical Infectious Diseases : An Official Publication of the Infectious Diseases Society of America*, vol. 68, no. 1, pp. 120–130, 2019.
- [38] O. O. Yang, T. Kelesidis, R. Cordova, and H. Khanlou, "Immunomodulation of antiretroviral drug-suppressed chronic HIV-1 infection in an oral probiotic double-blind placebo-controlled trial," *AIDS Research and Human Retroviruses*, vol. 30, no. 10, pp. 988–995, 2014.
- [39] J. C. Hemsworth, S. Hekmat, and G. Reid, "Micronutrient supplemented probiotic yogurt for HIV-infected adults taking HAART in London, Canada," *Gut Microbes*, vol. 3, no. 5, pp. 414–419, 2014.
- [40] C. R. Heiser, J. A. Ernst, J. T. Barrett, N. French, M. Schutz, and M. P. Dube, "Probiotics, Soluble Fiber, and L-Glutamine (GLN) Reduce Nelfinavir (NFV) or Lopinavir/Ritonavir (LPV/r)-related Diarrhea," *Journal of the International Association of Physicians in AIDS Care*, vol. 3, no. 4, pp. 121–129, 2016.
- [41] M. Schunter, H. Chu, T. L. Hayes et al., "Randomized pilot trial of a synbiotic dietary supplement in chronic HIV-1 infection," *BMC Complementary and Alternative Medicine*, vol. 12, no. 1, 2012.
- [42] K. C. Anukam, E. O. Osazuwa, H. B. Osadolor, A. W. Bruce, and G. Reid, "Yogurt containing probiotic *Lactobacillus rhamnosus* GR-1 and *L. reuteri* RC-14 helps resolve moderate diarrhea and increases CD4 count in HIV/AIDS patients," *Journal of Clinical Gastroenterology*, vol. 42, no. 3, pp. 239–243, 2008.
- [43] A. Andrade and E. T. Overton, *Effects of the probiotic Visbiome Extra Strength on gut microbiome & immune activation markers*, 2016, <https://clinicaltrials.gov/show/NCT02706717>.
- [44] L. A. González-Hernández, L. F. Jave-Suarez, M. Fafutis-Morris et al., "Synbiotic therapy decreases microbial translocation and inflammation and improves immunological status in HIV-infected patients: a double-blind randomized controlled pilot trial," *Nutrition Journal*, vol. 11, no. 1, 2012.
- [45] S. L. Irvine, R. Hummelen, S. Hekmat, C. W. N. Looman, J. D. F. Habbema, and G. Reid, "Probiotic yogurt consumption is associated with an increase of CD4 count among people living with HIV/AIDS," *Journal of Clinical Gastroenterology*, vol. 44, no. 9, pp. e201–e205, 2010.
- [46] M. Mahy, J. Stover, K. Stanecki, R. Stoneburner, and J. M. Tassie, "Estimating the impact of antiretroviral therapy: regional and global estimates of life-years gained among adults," *Sexually transmitted infections*, vol. 86, Suppl 2, pp. -ii67–ii71, 2010.
- [47] J. T. Salas and T. L. Chang, "Microbiome in human immunodeficiency virus infection," *Clinics in Laboratory Medicine*, vol. 34, no. 4, pp. 733–745, 2014.
- [48] X. Dagenais-Lussier, A. Mouna, J. P. Routy et al., "Current topics in HIV-1 pathogenesis: the emergence of deregulated immuno-metabolism in HIV-infected subjects," *Cytokine & Growth Factor Reviews*, vol. 26, no. 6, pp. 603–613, 2015.
- [49] R. Ponte, V. Mehraj, P. Ghali, A. Couëdel-Courteille, R. Cheynier, and J. P. Routy, "Reversing gut damage in HIV infection: using non-human primate models to instruct clinical research," *eBioMedicine*, vol. 4, pp. 40–49, 2016.
- [50] S. Cunningham-Rundles, S. Ahrné, R. Johann-Liang et al., "Effect of probiotic bacteria on microbial host defense, growth, and immune function in human immunodeficiency virus type-1 infection," *Nutrients*, vol. 3, no. 12, pp. 1042–1070, 2011.
- [51] D. M. Moore, R. S. Hogg, B. Yip, K. Craib, E. Wood, and J. S. G. Montaner, "CD4 percentage is an independent predictor of survival in patients starting antiretroviral therapy with absolute CD4 cell counts between 200 and 350 cells/ μ L," *HIV Medicine*, vol. 7, no. 6, pp. 383–388, 2006.
- [52] A. C. Ouwehand, "A review of dose-responses of probiotics in human studies," *Beneficial Microbes*, vol. 8, no. 2, pp. 143–151, 2017.
- [53] J. L. Castilho, B. E. Shepherd, J. Koethe et al., "CD4+/CD8+ ratio, age, and risk of serious noncommunicable diseases in HIV-infected adults on antiretroviral therapy," *AIDS*, vol. 30, no. 6, pp. 899–908, 2016.
- [54] S. Serrano-Villar, T. Sainz, S. A. Lee et al., "HIV-infected individuals with low CD4/CD8 ratio despite effective antiretroviral therapy exhibit altered T cell subsets, heightened CD8+ T cell activation, and increased risk of non-AIDS morbidity and mortality," *PLoS Pathogens*, vol. 10, no. 5, 2014.
- [55] H. Miller, R. Ferris, and B. R. Phelps, "The effect of probiotics on CD4 counts among people living with HIV: a systematic review," *Beneficial Microbes*, vol. 7, no. 3, pp. 345–351, 2016.
- [56] G. M. Carter, A. Esmaeili, H. Shah et al., "Probiotics in human immunodeficiency virus infection: a systematic review and evidence synthesis of benefits and risks," *Open forum infectious diseases*, vol. 3, no. 4, 2016.
- [57] J. C. Hemsworth, S. Hekmat, and G. Reid, "Micronutrient supplemented probiotic yogurt for HIV-infected adults taking HAART in London, Canada," *Gut Microbes*, vol. 3, no. 5, pp. 414–419, 2014.