

Plasma Melatonin Levels in Relation to the Light-Dark Cycle and Parental Background in Domestic Pigs

By *H. Andersson*

Department of Clinical Chemistry, Centre of Reproductive Biology in Uppsala, Swedish University of Agricultural Sciences, Uppsala, Sweden.

Andersson H: Plasma melatonin levels in relation to the light-dark cycle and parental background in domestic pigs. Acta vet. scand. 2001, 42, 287-294. – To study porcine melatonin secretion in a stable environment 3 daytime (10.00 - 15.00) and 3 nighttime (22.00 - 03.00) plasma samples were collected by jugular venipuncture from 15 gilts, 16 sows, 3 boars and 48 piglets (24 females and 24 males from 8 litters) and analysed for melatonin content. Nighttime melatonin concentrations were higher than daytime melatonin concentrations ($p < 0.001$), whereas no effect of sampling order could be discerned. The 3 adult Hampshire boars had higher melatonin concentrations during the day and the night, than the 31 adult Yorkshire females ($p < 0.05$). There was no clear difference between gilts and sows in plasma melatonin. The gilts from one of the litters had higher plasma melatonin concentrations than the gilts in 3 other litters ($p < 0.05$). Among the 48 piglets, the increase of nocturnal melatonin secretion differed between litters ($p < 0.01$), whereas the influence of father was not quite significant ($p = 0.12$). No difference in daytime melatonin concentrations between litters could be found, and there was no difference in melatonin levels between the male and female piglets. In conclusion, this study demonstrates that domestic pigs express a nocturnal increase of melatonin secretion in a standard stable environment. For some animals the amplitude of nighttime melatonin secretion was very low, although always higher than the daytime base levels. Furthermore, the levels of nighttime melatonin secretion differed between litters, which suggests a genetic background.

genetic.

Introduction

The circadian rhythm of pineal melatonin, with an increased secretion during the night and low concentrations during the day, is mediating photoperiodic information to the neuroendocrine reproductive system in many non-tropical seasonal breeding mammals (*Bartness & Goldman 1989*).

The domestic pig breeds continuously, although seasonal variations in reproduction, with reduced fertility during late summer and autumn, have been demonstrated from many parts of the

world (*Claus & Weiler 1985, Love et al. 1993, and Peltoniemi et al. 1999*). The period of seasonal infertility coincides with the anestrus period of the European wild boar (*Sus scrofa*) (*Mauget 1982*). Seasonal change in photoperiod has been suggested as an important factor causing these fertility problems, and artificial photoperiod has been shown to influence the timing of puberty in both gilts (*Paterson & Pearce 1990*) and boars (*Andersson et al. 1998*).

In the domestic pig, the reports of the existence of a typical circadian rhythm of peripheral melatonin have been contradictory, with only few studies reporting melatonin profiles that consistently change according to the light-dark phases (Paterson *et al.* 1992a, Andersson *et al.* 2000). Originally, no melatonin rhythm was found under short or long photoperiods (Reiter *et al.* 1987, McConnell & Ellendorff 1987, Minton *et al.* 1989), but day-night differences could be demonstrated in at least some animals in an equatorial photoperiod (McConnell & Ellendorff 1987, Minton & Cash 1990). Thereafter, several discrepant studies have been published (e.g. Diekman *et al.* 1992, Green *et al.* 1996 and 1999, Diekman & Green 1997, Bollinger *et al.* 1997, Bubenik *et al.* 2000), and the deviations of the results have been explained by variations of light intensity (Griffith & Minton 1992), by the great pig-to-pig variability (Green *et al.* 1996, Bollinger *et al.* 1997) and by inadequate assay methods (Klupiec *et al.* 1997, Andersson *et al.* 2000).

The amplitude of the nocturnal melatonin secretion in pigs appears to be lower than in most studied mammalian species (Andersson *et al.* 2000). If only a minor increase in melatonin secretion during the dark-phase is sufficient for a photoperiodic response on the reproductive system is not known.

The aim of this study was to investigate if parental background influence porcine melatonin in the light environment of a pig stable, and if sampling by jugular venipuncture can be used for evaluating individual melatonin profiles.

Materials and methods

Animals and photoperiod

Female Yorkshire pigs, 15 gilts from 5 litters and 16 sows, and 3 Hampshire boars were bled during winter (November-February) at 60°N (6-9 h of light). In August at 60°N (15-16 h of

light), 48 crossbred (YxH) piglets, 24 females and 24 males (10-14 weeks of age), offspring of four gilts, 4 sows and 2 boars from the winter bleeding, were bled. The animals were kept in standard stable management with windows and additional light (light bulbs) during working hours (8:00-16:00). Daytime light intensity varied depending on weather conditions between 150-300 lux, with occasional higher intensities. Overall nighttime light conditions were very low for the gilts and piglets (<5 lux). The sows and boars had low-intensity night illumination (light bulbs) creating a nighttime light intensity between 5-10 lux.

Plasma sampling

Three daytime samples and 3 nighttime samples were collected by jugular venipuncture into heparinised tubes between 10:00-15:00 and 22:00-03:00, respectively, from each animal, with approximately hourly intervals. Nighttime light intensity varied somewhat depending on lunar phase and weather conditions, such as cloudiness and snow. To facilitate sampling during the night, dim red light and a small flashlight were used. Thus, it is not possible to exactly say which light intensity the animals were exposed to at each moment of sampling, although any direct light exposure of the pigs' eyes was avoided at all times. After collection the samples were centrifuged and stored at -20°C until analysed for melatonin content.

Melatonin assay

Plasma melatonin was analysed by radio immunoassay (Bhhlmann Laboratories AG, Schönenbuch, Switzerland). Before assay, 1 ml portions of controls and samples were extracted twice in 4.5 ml of diethyl ether. The tubes were then shaken for 1 min and put into a freezing bath. The supernatant was decanted and the solvent removed by evaporation to dryness in a 37°C water bath, whereupon the residue was

dissolved in 1 ml of incubation buffer. Duplicate aliquots (400 μ l) of standards, extracted controls and extracted plasma samples were pipetted into the tubes, followed by 100 μ l of anti-melatonin antiserum (Kennaway G280; caprine against melatonin conjugated to bovine thyroglobulin, see Vaughan, 1993), and 100 μ l of the 125 I-melatonin tracer. The tubes were then incubated for 20 h (\pm 4 h) at 2-8°C. While stirring the second anti-body, 100 μ l of the suspension was added to the tubes, after which they were incubated at 2-8°C. After 15 min 1 ml of cold, distilled water was added to the tubes, which were then centrifuged at 2-8°C. After 15 min the supernatant was removed and the radioactivity of the tubes was counted in a gamma counter for 2 min. Serial dilutions of pig plasma containing high concentrations of melatonin produced displacement curves parallel to the standard curve. The intra-assay and inter-assay coefficients of variations for 20 assays, were 13.1% and 8.2% (2.4 pg/ml), and 8.4% and 8.0% (19.5 pg/ml), respectively, and the sensitivity of the assay was 0.3 pg/ml (intercept of maximal binding - 2 S.D.). Using reversed-phase column extraction, the manufacturer calculated the minimal detectable concentration to be 0.3 pg/ml. The specificity of the assay has been evaluated by Bhlmann Laboratories AG and all measured compounds show less than 0.05% cross-reactivity. Selected samples were reanalysed on a later occasion, in order to ensure assay repeatability.

Statistics

Statistical analyses were performed by analysis of variance by MIXED procedures (*SAS Institute Inc.* 1997) and least square means option was used to compare different means. Melatonin levels from the winter bleeding were tested for variance of time-of-day (day versus night), sampling order within time-of-day, sex and age within sex with individual animal as

Table 1. Daytime (10:00-15:00) and nighttime (22:00-03:00) plasma melatonin concentrations (least square means \pm s.e.m.). (N=82)

	Day	Night	P-value
Melatonin (pg/ml)	2.7 \pm 0.8	14.4 \pm 0.8	p<0.001

random effect. The melatonin concentrations of the gilts from the winter bleeding were further analysed in a model with time-of-day, sampling order within time-of-day and mother (litter) as fixed effects (effect of fathers could not be considered as it partly overlapped with litter) and individual animal as random effect. Melatonin levels from the summer bleeding were initially analysed in a model with time-of-day, sampling order within time-of-day, sex, father, mother (litter) within father as fixed effects and individual animal within father as random effect. As no significant variation was associated with sampling order within time-of-day, sex or father, melatonin from the piglets were reanalysed in a model with time-of-day, mother(litter) and the interaction between mother(litter) and time-of-day as fixed effects and individual animal within father as random effect. Melatonin concentrations from both sampling occasions were analysed for the effects of time-of-day, sampling order within time-of-day, sex and age within sex as fixed effects and individual animal as random effect.

Results

Nighttime melatonin concentrations were higher than daytime melatonin concentrations (Table 1), whereas no effect of sampling order could be discerned at either bleeding occasion. The adults and the young animals were bled at different times of the year. When wild and domestic pigs were compared in 4 seasons, the melatonin rhythm was entrained by the photoperiod of the season whereas no effect of sea-

Table 2. Daytime (10:00-15:00) and nighttime (22:00-03:00) plasma melatonin concentrations (least square means \pm s.e.m.) in gilts from different litters.

	Litter 1 (n=4)	Litter 2 (n=2)	Litter 3 (n=4)	Litter 4 (n=3)	Litter 5 (n=2)
<i>Melatonin (pg/ml)</i>					
Day	2.6 ^a \pm 2.7	1.9 ^a \pm 3.8	0.6 ^a \pm 2.7	0.9 ^a \pm 3.1	2.5 ^a \pm 3.8
Night	23.2 ^a \pm 2.7	8.7 ^b \pm 3.8	6.4 ^b \pm 2.7	10.3 ^b \pm 0.9	15.0 ^{ab} \pm 3.8

^{a,b}Values within a row with no superscript in common differ significantly ($p < 0.05$)

son on melatonin levels could be found (*Tast et al.* 2001). There was no difference in melatonin levels between adult and young animals in this study.

Adults

The 3 adult Hampshire boars had higher nighttime (23.5 ± 2.9 pg/ml; least square mean \pm s.e.m.) and daytime (9.8 ± 2.9 pg/ml) melatonin concentrations than the 31 Yorkshire sows and gilts (night: 14.1 ± 0.9 pg/ml, day: 3.3 ± 0.9 pg/ml) ($p < 0.05$). There was no clear difference between gilts and sows in melatonin levels. In spite of the low numbers of animals per litter, the gilts from one of the litters had higher plasma melatonin concentrations than the gilts in 3 other litters (Table 2).

Piglets

Among the 48 piglets, the effect of father was not quite significant ($p = 0.12$) and there was no difference in melatonin concentrations between the male and female piglets. There was an interaction between time-of-day and litter (mother) ($p < 0.01$) as nighttime but not daytime plasma melatonin concentrations differed between litters (Fig. 1).

Discussion

In a pig stable environment, domestic pigs showed a nocturnal increase in plasma melatonin secretion. Nighttime plasma melatonin levels differed between litters, which indicates

that the great individual variations in the amplitude of nocturnal melatonin secretion, observed in this species (e.g. *Andersson et al.* 2000, *Tast et al.* 2001) has a genetic background.

Jugular venipuncture, which is a commonly used bleeding method in pigs, requires restraining of the animal. The stress that is associated with being restrained leads to increase of heart rate, catecholamine, cortisol and β -endorphin levels etc. (*Roozen et al.* 1995). Some of these stress reaction, such as plasma cortisol concentrations, can be expected to have been increasing during the bleeding period, yet no differences in plasma melatonin level between first, second and last time of sampling could be discerned, indicating that the stress and handling as such during the bleeding did not disturb the melatonin measurements. As all animals showed a higher average nighttime melatonin concentration than daytime level, and there was a high individual variation in nighttime melatonin levels, this indicates that plasma samples collected by jugular venipuncture can serve as a basis for evaluating melatonin profiles from a large number of animals. However, occasional high melatonin concentrations were observed during the day. Since plasma sampled by indwelling jugular catheters revealed only low to undetectable daytime melatonin concentrations, using the same assay (*Andersson et al.* 2000, *Tast et al.* 2001), the random higher measurements in this study possibly were caused by a cross reaction with some factor(s), which may

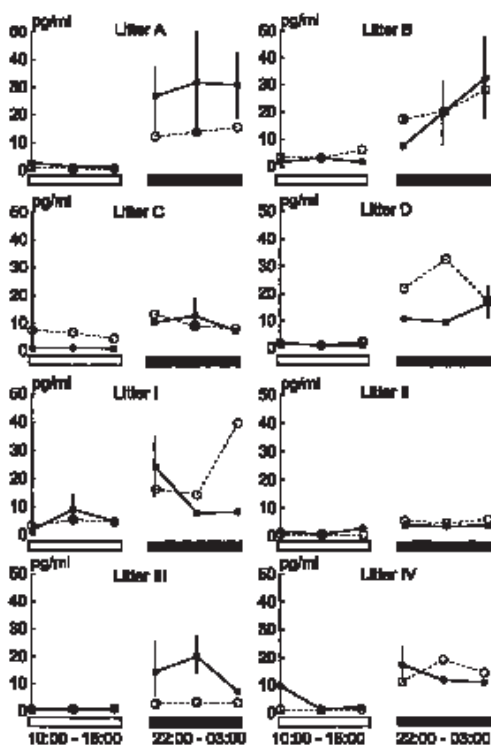


Figure 1. Daytime (10:00-15:00; white horizontal bars) and nighttime (22:00-03:00; dark horizontal bars) plasma melatonin concentrations (mean \pm sem) in piglets from different litters ($n=48$, six piglets per litter, 3 males and 3 females). The mothers' melatonin levels are marked with open circles (A-D are sows and I-IV are gilts).

have entered the blood sample as the needle passes through the epidermis and subcutaneous layers at the time of venipuncture. Irrespective of cause, this emphasises the importance to use multiple sampling in order to correctly evaluate the individual melatonin profiles, when jugular venipuncture is applied.

The 3 adult Hampshire boars in this study showed higher plasma melatonin concentrations than the adult females, although a clear nighttime increase in melatonin secretion was

observed in both sexes. Daytime melatonin concentrations consistently elevated above the detection limit were only observed for the 3 adult boars (not among the male piglets). Although higher pineal concentrations of melatonin have been observed in male compared to female Siberian (also called Djungarian) hamsters (*Phodopus sungorus*; Niklowitz *et al.* 1996), interpretation of results from so few animals must be made with caution, especially since the gender in this case overlapped with the breed. Extra-pineal melatonin is synthesised in e.g. the gastrointestinal tract, but its contribution to circulating melatonin levels is controversial (Heuther 1993). Though the melatonin levels of the boars over all were significantly higher than the plasma concentrations of the adult females, there was no sex difference in the extent of the night-time melatonin increase. Therefore, the possible sex differences in melatonin concentrations probably have no importance for the role of melatonin as an endocrine signal of darkness. However, increased diurnal levels of the main urinary melatonin metabolite (6-sulfatoxymelatonin) have been observed among Siberian/Djungarian hamsters that are reproductively unresponsive to photoperiod (Niehaus & Lerchl 1998).

Although the melatonin rhythm in sheep is highly repeatable within the individual (Chemineau *et al.* 1996), the amplitude of nocturnal melatonin shows high inter-individual variability (Malpoux *et al.* 1987), which is caused by a genetic variability in the synthesis of pineal melatonin (Zarazaga *et al.* 1998a and Zarazaga *et al.* 1998b). In contrast to an earlier study (Andersson *et al.* 2000), there was no significant effect of fathers in this study. Therefore, it can only be suggested that inter-individual variability in night-time melatonin concentrations reflects a genetic variation. Differences in night-time melatonin seemed to be depending on the

sibling group among the gilts, although the number of gilts per sibling group was low (2-4 animals per litter). But, since the same influence of litter was seen among the piglets (6 animals per litter), the variation in amplitude of night-time melatonin secretion between sibling-groups could be confirmed. The offspring used in this study had spent their short lives in an almost identical environment. Furthermore, the older piglets were no longer kept together with their litter mates at the time of the bleeding, but were mixed with piglets from other litters according to sex. Thus, the social group did not overlap with the sibling group among these piglets. Age and weight of the piglets overlapped with litter, as a result of the study design.

In lambs a melatonin pattern that reflects the light-dark cycle is present already at 3 weeks of age and the amplitude of nighttime melatonin secretion increases between 6 and 27 weeks of age (Claypool *et al.* 1989). In contrast, in female rhesus monkeys the nighttime amplitude of melatonin secretion decreases during pubertal development (Wilson & Gordon 1988). Among the piglets, however, there was no clear trend of an increase or decrease of the amplitude of night-time melatonin concentrations with age, as both the highest and the lowest average night-time melatonin concentrations were found among the older (and heavier) piglets. Together, this supports the hypothesis that the differences in melatonin pattern between litters observed in this study, probably is a result of the genetically determined capacity for pineal melatonin synthesis which has been described in sheep (Zarazaga *et al.* 1998a).

As seasonal infertility is a management problem for the pig producers, it was important to see whether a night-time increase in melatonin secretion was observed in a conventional pig stable environment. This study showed increased melatonin secretion during the dark

hours as is the case in other animals (Reiter 1993). The nocturnal increase in pigs is relatively low compare to many other studied species, but the average nighttime melatonin concentration was always higher than the average day-time concentration for each individual animal. Studies on the effects of photoperiod or exogenous melatonin administration on pig reproduction have shown varied results (e.g. Kretzling *et al.* 1983, Lee *et al.* 1987 and Paterson *et al.* 1992b). Whether the low nocturnal secretion of melatonin observed among some sibling groups influences the response to photoperiod or melatonin is not possible to state, since no reproductive parameters were measured in this study. However, a circadian rhythm in melatonin, with a clear elevation during the dark phase, is required for transferring photoperiodic information in all seasonal breeding mammals (Reiter 1993).

In conclusion, this study demonstrates that domestic pigs of different ages, breeds and sex show a night-time elevation of melatonin secretion in a pig stable environment. Although always higher than the daytime base levels, the increase in melatonin secretion during the night is small in some animals. Furthermore, the amplitude of the nighttime melatonin secretion differed between litters, which suggests a genetic background

Acknowledgement

The author wish to thank the Department of Animal Breeding and Genetics, SLU for the use of their breeding herd, Eva Norling, Ulf Hermansson and Carola Jansson for help with the collection of blood samples and all the rest of the staff at Funbo-Lövsta for taking such good care of the animals, Karin Burvall is thanked for all the hard work with the melatonin assay.

References

Andersson H, Lillpers K, Rydhmer L, Forsberg M:
Influence of light environment and photoperiod

- on plasma melatonin and cortisol profiles in young domestic boars, comparing two commercial melatonin assays. *Domest. Anim. Endocrinol.* 2000, *19*, 261-274.
- Andersson H, Wallgren M, Rydhmer L, Lundström K, Andersson K, Forsberg M*: Photoperiodic effects on pubertal maturation of spermatogenesis, pituitary responsiveness to exogenous GnRH, and expression of boar taint in crossbred boars. *Anim. Reprod. Sci.* 1998, *54*, 121-137.
- Bartness TM, Goldman BD*: Mammalian pineal melatonin: a clock for all seasons. *Experientia*, 1989, *45*, 939-945.
- Bollinger AL, Wilson ME, Pusateri AE, Green ML, Martin TG, Diekman MA*: Lack of a nocturnal rise in serum concentrations of melatonin as gilts attain puberty. *J. Anim. Sci.* 1997, *75*, 1885-1892.
- Bubenik GA, Pang SF, Cockshut JR, Smith PS, Grovum LW, Friendship, RM, Hacker RR*: Circadian variation of portal, arterial and venous blood levels of melatonin in pigs and its relationship to food intake and sleep. *J. Pineal. Res.* 2000, *28*, 9-15.
- Chemineau P, Beltrán de Heredia I, Daveau A, Bodin L*: High repeatability of the amplitude and duration of the nyctohemeral rhythm of the plasma melatonin concentrations in Ile-de-France ewes. *J. Pineal. Res.* 1996, *21*, 1-6.
- Claus R, Weiler U*: Influence of light and photoperiodicity on pig prolificacy. *J. Reprod. Fert. Suppl.* 1985, *33*, 185-197.
- Claypool LE, Wood R-I, Yellon SM., Foster DL*: The ontogeny of melatonin secretion in the lamb. *Endocrinology*, 1989, *124*, 2135-2143.
- Diekman MA, Brandt KE, Green ML, Clapper JA, Mayaler JR*: Lack of a nocturnal rise of serum melatonin in prepubertal gilts. *Domest. Anim. Endocrinol.* 1992, *9*, 161-167.
- Diekman MA, Green ML*: Serum concentrations of melatonin in prepubertal gilts exposed to artificial lighting and sunlight. *Theriogenology* 1997, *47*, 923-928.
- Green ML, Clapper JA, Andres CJ, Diekman MA*: Serum melatonin in prepubertal gilts exposed to either constant or stepwise biweekly alteration in scotophase. *Domest. Anim. Endocrinol.* 1996, *13*, 307-323.
- Green ML, Clapper JA, Diekman M*: Serum concentrations of melatonin during scotophase and photophase in 3, 4, 5 and 6 months old gilts and barrows. *Anim. Reprod. Sci.* 1999, *57*, 99-110.
- Griffith MK, Minton JE*: Effect of light intensity on circadian profiles of melatonin, prolactin, ACTH, and cortisol in pigs. *J. Anim. Sci.* 1992, *70*, 492-498.
- Heuther G*: The contribution of extrapineal sites of melatonin synthesis to circulating melatonin levels in higher vertebrates. *Experientia* 1993, *49*, 665-670.
- Klupiec CEG, Love RJ, Kennaway DJ*: Clarifying plasma melatonin profiles in domestic pigs: A critical and comparative evaluation of two radio immunoassay systems. *J. Pineal. Res.* 1997, *22*, 65-74.
- Krealing RR, Rampacek GB, Mabry JW, Cunningham FL, Pinkert CA*: Serum concentrations of pituitary and adrenal hormones in female pigs exposed to two photoperiods. *J. Anim. Sci.* 1983, *57*, 1243-1250.
- Lee K-H, Diekman MA, Moss GE, Allrich RD*: Pituitary gonadotropins, hypothalamic gonadotropin-releasing hormone and testicular traits of boars exposed to natural or supplemental lighting during pubertal development. *Biol. Reprod.* 1987, *36*, 1164-1169.
- Love RJ, Evans G, Klupiec C*: Seasonal effects on fertility in gilts and sows. *J. Reprod. Fert. Suppl.* 1993, *48*, 191-206.
- McConnell SJ, Ellendorff F*: Absence of nocturnal plasma melatonin surge under long and short artificial photoperiods in the domestic sow. *J. Pineal. Res.* 1987, *4*, 201-210.
- Malpoux B, Robinson JE, Brown MB, Karsch FJ*: Reproductive refractoriness of the ewe to inductive photoperiod is not caused by inappropriate secretion of melatonin. *Biol. Reprod.* 1987, *36*, 1333-1341.
- Mauget R*: Seasonality of reproduction in the wild boar. In: DJA Cole and GR Foxcroft (Editors), *Control of Pig Reproduction*, Butterworths, London 1982, 509-526.
- Minton JE, Cash WC*: Effect of cranial sympathectomy on circadian rhythms of cortisol, adrenocorticotrophic hormone and melatonin in boars. *J. Anim. Sci.* 1990, *68*, 4277-4284.
- Minton JE, Davies DL, Stevenson SJ*: Contribution of the photoperiod to circadian variations in serum cortisol and melatonin in boars. *Domest. Anim. Endocrinol.* 1989, *6*, 177-181.
- Niehaus M, Lerchl A*: Urinary 6-sulfoxy melatonin profiles in male Djungarian hamsters (*Phodopus sungorus*) responding and not responding to short-day photoperiods: Possible role of elevated daytime levels. *J. Pineal Res.* 1998, *25*, 167-171.

- Niklowitz P, Böckers TM, Lerchl A: Afternoon injections of melatonin in the Djungarian hamster *Phodopus sungorus*: Long lasting sex-specific effects and influence of acute treatment on the endogenous pineal melatonin rhythm. *J Pineal Res.* 1996, *21*, 231-238.
- Paterson AM, Martin GB, Foldes A, Maxwell CA, Pearce GP: Concentrations of plasma melatonin and luteinizing hormone in domestic gilts reared under artificial long and short days. *J. Reprod. Fertil.* 1992a, *94*, 85-95.
- Paterson AM, Maxwell CA, Foldes A: Seasonal inhibition of puberty in domestic gilts is overcome by melatonin administered orally, but not by implant. *J. Reprod. Fertil.* 1992b, *94*, 97-105.
- Paterson AM, Pearce GP: Attainment of puberty in domestic gilts reared under long-day or short-day artificial light regimens. *Anim. Reprod. Sci.* 1990, *23*, 135-144
- Peltoniemi OAT, Love RJ, Heinonen M, Tuvionen V, Saloniemi H: Seasonal and management effects on fertility of the sow: a descriptive study. *Anim Reprod. Sci.* 1999, *55*, 47-61.
- Reiter RJ: The melatonin rhythm: both a clock and a calendar. *Experientia* 1993, *49*, 654-664.
- Reiter RJ, Britt JH, Armstrong JD: Absence of a nocturnal rise in either norepinephrine, N-acetyltransferase, hydroxyindole-O-methyltransferase or melatonin in the pineal gland of the domestic pig kept under natural environment photoperiods. *Neurosci. Lett.* 1987, *81*, 171-176.
- Roosen AWM, Tsuma VT, Magnusson U: Effects on short-term stress on plasma concentrations of catecholamines, b-endorphins, and cortisol in gilts. *Am. J. Vet. Res.* 1995, *56*, 1225-1227.
- SAS Institute Inc.: The SAS system for Windows. 1997, Cary, NC, USA.
- Tast A, Halli O, Ahlström S, Andersson H, Love R.J, Peltoniemi OAT: Seasonal alterations in circadian melatonin rhythms in European wild boar and domestic gilt. *J. Pineal. Res.* 2001, *30*, 43-49.
- Vaughan GM: New sensitive serum melatonin radio immunoassay employing the Kennaway G280 antibody: Syrian hamster morning adrenergic response. *J. Pineal. Res.* 1993, *15*, 88-103.
- Wilson ME, Gordon TP: Nocturnal changes in serum melatonin during female puberty in rhesus monkeys: a longitudinal study. *J. Endocrinol.* 1988, *121*, 553-562.
- Zarazaga L, Malpoux B, Bodin L, Chemineau P: The large variability in melatonin blood levels in ewes is under strong genetic influence. *Am. J. Physiol.* 1998a, *274*, E607-610.
- Zarazaga L, Malpoux B, Guillaume D, Bodin L, Chemineau P: Genetic variability in melatonin concentrations in ewes originates in its synthesis, not in its catabolism. *Am. J. Physiol.* 1998b, *274*, E1086-1090.

Sammanfattning

Melatoninnivåer i plasma hos tamsvin i relation till ljus-mörker och härkomst.

För att studera melatoninutsöndring hos grisar i stallmiljö, samlades 3 dagsprover (10.00-15.00) och 3 nattprover (22.00-03.00) plasma med hjälp av venpunktion från 15 gyltor, 16 suggor, 3 galtar och 48 kulingar (24 honor och 24 hanar från 8 kullar) och analyserades på melatonininnehåll. Melatoninkoncentrationerna under natten var högre än under dagen ($p < 0,001$), men ingen effekt av provtagningsordning kunde ses. De 3 galtarna hade högre melatoninnivåer än de 31 gyltorna och suggorna, både under dag och natt, medan det inte fanns någon skillnad mellan gyltor och suggor. Fyra gyltor från samma kull hade högre melatoninnivåer under natten än gyltorna från 3 andra kullar ($p < 0,05$). Bland de 48 kulingarna var det skillnad mellan kullarna i melatoninnivå under natten ($p < 0,01$), medan effekten av fäder inte var riktigt signifikant ($p = 0,12$). Det fanns ingen skillnad i dagsnivåer mellan kullarna och ingen skillnad mellan hanar och honor. Sammantaget visar denna studie att grisar i stallmiljö har en ökad melatoninutsöndring under natten. Hos somliga djur var amplituden i melatoninutsöndring under natten liten, men alltid större än under dagen. Vidare, så skiljde sig amplituden av melatoninutsöndring under natten mellan kullarna, vilket tyder på en genetisk variation.

(Received September 5, 2000, accepted January 31, 2001).

Reprints may be obtained from: Department of Clinical Chemistry, PO Box 7038, S-750 07 Uppsala, Sweden. E-mail: Hakan.Andersson@klke.slu.se, tel.: +46-18-671614, fax: +46-18-309565.

Present address: MCR Human Reproductive Sciences Unit, Centre for Reproductive Biology, 37 Chalmers Street, Edinburgh, EH3 9ET, UK. E-mail: h.andersson@hrsv.mrc.ac.uk, tel: +44 (01) 131 229 2575, fax: +44 (01) 131 228 5571.