



# Decrypting the Possible Mechanistic Role of Fenofibrate in Alzheimer's Disease and Type 2 Diabetes: The Truth and Mystery

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### **ABSTRACT**

Alzheimer's disease (AD) is a neurodegenerative disease caused by the progressive deposition of extracellular amyloid beta (A $\beta$ ) and intracellular neurofibrillary tangles (NFTs). Of note, metabolic disorders such as insulin resistance (IR) and type 2 diabetes (T2D) are associated with the development of brain IR and associated neurodegeneration. In addition, AD neuropathology and linked cognitive impairment accelerate the development of peripheral IR and the progression of T2D. Therefore, there is a bidirectional relationship between T2D and AD. It has been demonstrated that AD and T2D induce dysregulation of peroxisome proliferator-activated receptor alpha (PPAR- $\alpha$ ) leading to the central and peripheral metabolic disturbances. Hence, dysregulated PPAR- $\alpha$  could be a shared mechanism in both AD and T2D, and restoration of PPAR- $\alpha$  signalling by PPAR- $\alpha$  agonist fenofibrate (FN) may alleviate T2D and AD. Therefore, this review aims to shed light on the potential involvement of PPAR- $\alpha$  in T2D and AD, and how FN could be effective in the management of AD. FN seems to be effective in both AD and T2D by dual neuroprotective and antidiabetic effects that can mitigate AD neuropathology and T2D-related complications by modulating various cellular processes and inflammatory signalling pathways. In conclusion, FN could be a possible candidate in the management of AD and T2D by modulating different signalling pathways involved in the pathogenesis of these conditions.

# 1 | Introduction

Alzheimer's disease (AD) is a neurodegenerative disease caused by the progressive deposition of extracellular amyloid beta  $(A\beta)$ 

and intracellular neurofibrillary tangles (NFTs) [1, 2]. Multiple processes, including oxidative stress, autophagy, inflammation, cholinergic dysfunction and mitochondrial dysfunctions are involved in AD neuropathology [3, 4] (Figure 1).

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# Amyloid β plaque Neurofibrillary tangle Neuron loss and degeneration Cholinergic Insufficiency Autophagy Alzheimer's patient

**FIGURE 1** | Pathophysiology of AD: Progressive accumulation of the extracellular amyloid beta  $(A\beta)$  and intracellular neurofibrillary tangles (NFTs) induce neurodegeneration with subsequent cholinergic dysfunction and glutamatergic overactivity. These neuropathological changes also inhibit neuronal autophagy and trigger the development of neuroinflammation.

It has been revealed that Aβ plaques are crucial in the development of AD [5]. When plaque sequestration ability is diminished, soluble Aβ can spread into extracellular causing severe synaptic dysfunction and neuronal injury [5]. Similarly, Aβ oligomers affect cell membrane ion channels and receptors to activate both intracellular and extracellular neurotoxicity [5, 6]. These pathological changes stimulate an extreme imbalance between inhibitory and excitatory neurotransmitters with the progress of hyperexcitability [6]. It has been shown that imbalance of the excitatory/inhibitory axis was correlated with the severity of Aβ deposition in AD patients compared to healthy controls [7]. Therefore, Aβ-induced neuronal damage and the progression of increased AD are facilitated by synaptic dysfunction and the increase of excitatory/inhibitory imbalance. Importantly, AD represents 70% of all dementia types, and the majority of AD cases are late-onset AD, meaning they develop sporadically beyond the age of 65 [8]. However, 5% of AD is of genetic type caused by mutations in the amyloid precursor protein (APP) genes and presented as an early-onset AD [9, 10].

Mounting evidence supports the link between type 2 diabetes (T2D) and the onset of AD [10, 11]. Notably, brain insulin resistance (IR) and inhibition of neuronal insulin receptors are commonly associated with the advancement of T2D [11]. These changes reduce brain glucose metabolism, increase  $A\beta_{1-42}$  production, impair  $A\beta$  clearance and trigger the deposition of aberrant  $A\beta$  plaque [12, 13]. Accordingly, it has been reported that T2D patients have a 50%–75% chance for developing AD, while people with AD have a greater chance for developing T2D [14]

(Figure 2). According to the in vitro study, high blood glucose level causes apoptosis in rat embryonic cortical neurons. High blood glucose level induces the phosphorylation of tau protein and the formation of A $\beta$  [14]. In addition, a study conducted on mice with diabetes found that they are more susceptible to tau protein hyperphosphorylation [14]. These in vivo and in vitro outcomes point to a possible bidirectional link between T2D and AD risk.

Furthermore, impairment of brain insulin signalling and brain IR is associated with the development of mild cognitive impairment (MCI) [15]. However, insulin treatment did not reduce brain IR [15]. Conversely, insulin-sensitising medications that overcome IR may be able to more successfully reduce brain IR and AD neuropathology. Belonging to the biguanide group, metformin is an insulin-sensitising molecule that is used as a crucial therapeutic treatment for T2D by regulating hepatic gluconeogenesis and glycogenolysis [16]. The fundamental cause of peripheral and brain IR may be related to the dysregulation of the peroxisome proliferatoractivated receptor alpha (PPAR-α) which regulates insulin sensitivity. Furthermore, PPAR-α has been demonstrated to be significantly dysregulated in both T2D [17] and AD [18]. It has been established that augmentation of peripheral and central PPAR-α by PPAR-α agonist fenofibrate (FN) can attenuate the metabolic derangement in T2D and reduce the progression of AD [19, 20]. However, the underlying mechanism for the neuroprotective effect of FN against AD is not fully elucidated. Therefore, this review has been amended to discuss the possible role of PPAR- $\alpha$  agonist FN in AD and T2D.

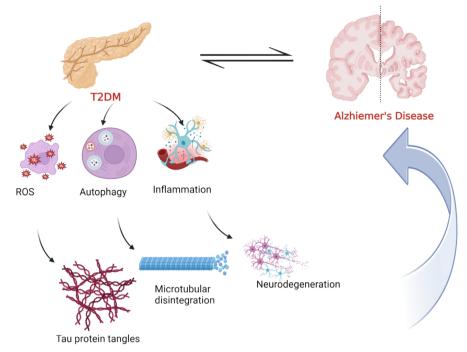


FIGURE 2 | The relationship between AD risk and T2D: T2D inhibits autophagy, and activates the release of reactive oxygen species (ROS) and pro-inflammatory cytokines leading to the formation of tau protein tangles and microtubular disintegration as well as neurodegeneration.

### 2 | The Link Between T2D and AD

T2D is a metabolic disease characterised by insulin insensitivity and dysfunctional  $\beta$  cells in the pancreas, which results in hyperglycaemia and IR [21]. Of note, 90% of diabetes is T2D which often affects persons over the age of 40 [22]. Obesity, hypertension and dyslipidaemia are among the most cardiometabolic diseases associated with the development of T2D [16]. T2D patients may be asymptomatic and may be present with long-term complications such as nephropathy, retinopathy and other microvascular complications [23]. However, the classical clinical features of T2D are polyuria, polydipsia, polyphagia and progressive weight loss [24].

T2D develops due to the progressive pancreatic  $\beta$ -cell dysfunction [25]. Notably, amyloid formation in the pancreatic  $\beta$  cells leads to progressive pancreatic  $\beta$ -cell death, pancreatic  $\beta$ -cell dysfunction and the development of overt T2D advance [26]. Islet amyloid polypeptide (IAPP) which is commonly referred to as an amylin, is the primary component of pancreatic  $\beta$ -cell amyloids, and is mostly cleaved by pancreatic neprilysin (NEP) [26]. Importantly, progressive deposition of IAPP in the pancreatic  $\beta$  cells is associated with pancreatic  $\beta$ -cell toxicity and dysfunctions [26]. Pancreatic  $\beta$ -cell dysfunction is prevented by NEP which cleaves and eliminates extracellular pancreatic IAPP. Therefore, inhibition of the pancreatic IAPP by activating NEP seems to be protective against the development of T2D [27].

Furthermore, the aggregation of  $A\beta$  in the brain facilitates the development of AD, whereas the accumulation of IAPP in the pancreas induces the development of T2D [28]. Pre-pro-IAPP is the precursor to IAPP, which is a 37-amino acid peptide [28]. Notably, pancreatic  $\beta$  cells cosecrete insulin into the bloodstream together with IAPP. As IAPP builds up over time, oligomers and

fibrils develop, and amyloid deposits are subsequently observed in T2D [28].

Moreover, IAPP-induced pancreatic β cytotoxicity may be exacerbated by human IAPP processing inefficiency brought on by abnormalities in autophagy and proteasome activity [29]. Consequently, restoring IAPP proteostasis could be an effective approach to treat and prevent T2D [30]. Consequently, the pathophysiology of brain IR and cognitive dysfunction have similar characteristics in both T2D and AD. As a result, AD is categorised as type 3 diabetes (T3D) [31]. T3D refers to the metabolic syndrome that may lead to abnormalities linked to progressive brain IR with consequent impairment of central insulin signalling processes, accumulation of neurotoxins, neuronal stress and resulting in a course of neurodegeneration [31]. Many studies have specified that impaired hippocampus insulin signalling distorts memory and other executive functions, attributing to the decline of insulin signalling and concomitant development of IR [32, 33].

Furthermore, brain and peripheral IR lead to hyperglycaemia, which participates in the development of T2D-linked comorbidities, such as obesity and dyslipidaemia [31]. Subjects with hyperglycaemia in AD present with neuronal loss, formation of plaques and tangles and reduced neurogenesis. Inflammation seems to play an essential role in the development of IR in AD and T2D. Alterations in glucose metabolism result from changes in the expression of the insulin receptor substrates 1 and 2 (IRS-1 and IRS-2), and seem to be mediated by several inflammatory pathways being present in both pathologies [32, 33].

Although there are some similarities in the IR of AD and T2D, brain and peripheral IR also have their distinct features. Failure to activate IRS-1 is the hallmark of AD, while inhibition of IRS-2

is the main feature of T2D. Inflammation mediates the alterations in glucose metabolism in AD and T2D. Targeting inflammation and insulin receptors may be a successful strategy to prevent and ameliorate T2D and AD symptoms [26, 27].

These verdicts highlighted that T2D and AD share several metabolic defects, such as IR, impaired glucose metabolism and mitochondrial defects. Prominently, T2D considerably increases the risk of cognitive decline and dementia, particularly AD. Besides, patients with AD are more susceptible to T2D and the possibility of linkage between the processes responsible for the loss of brain cells and  $\beta$  cells in these diseases [26–28].

# 3 | Pharmacology of FN

FN is a chlorobenzophenone derivative drug used in the management of hypertriglyceridaemia, mixed dyslipidaemia and diabetic retinopathy and can reduce primary and secondary cardiovascular events [34, 35]. In 1974, FN was mostly synthesised from clofibrate in France under the name precetofen, which the WHO no-proprietary guideline, renamed it as a FN [36]. FN is a lipid-modifying drug mainly used in patients with elevated triglyceride levels. FN exerts its action via the activation of PPAR- $\alpha$ . FN is a useful option for patients with primary combined dyslipidaemias or secondary dyslipidaemias, such as those associated with T2D and metabolic syndrome [36]. By activating PPAR- $\alpha$ , FN modulates the expression and the functional activity of lipoprotein lipase and apolipoprotein CIII resulting in lipolysis and the removal of triglycerides from plasma [37, 38].

Apolipoproteins AI and AII are expressed more when FN is present, which lowers LDL and VLDL levels while raising HDL [37, 38]. Furthermore, regardless of glycaemic management, FN reduces the incidence of amputation by 37% in T2D patients [34, 35]. FN is susceptible to pharmacological interactions with immunosuppressive medications, statins, warfarin and bile acid sequestrants [37, 38]. The most frequent side effects of FN are headache, arthralgia, myalgia and kidney stones. Patients with hypothyroidism, hypersensitivity, liver illness, gallbladder disorders and renal impairment should not use FN [38, 39].

# 4 | Role of FN in T2D and AD

### 4.1 | FN in T2D

FN reduces cardiovascular events in T2D patients proposing the possible role of FN in preventing cardiovascular complications by reducing atherogenic dyslipidaemia [19]. FN is recommended in T2D patients with dyslipidaemia which develops due to dysregulation of peripheral insulin signalling [39]. FN also improves lipid profile in T2D patients with poor glycaemic control by shifting low-density lipoprotein (LDL) towards nonatherogenic LDL [40]. A systematic review revealed that early use of FN in T2D patients prevents endothelial dysfunction and can attenuate other microvascular dysfunctions [41]. FN also has anti-inflammatory effects and reduces cardiometabolic disorders in patients with metabolic syndrome [42]. A cohort study involving 242 patients with metabolic syndrome with or without T2D randomised to FN, atorvastatin and placebo illustrated that FN

group experienced more glucose homeostasis than others [43]. This finding suggests that FN effectively attenuates metabolic syndrome mediated by prediabetes. Likewise, FN improves impaired glucose tolerance, including impaired fasting glucose and impaired glucose tolerance in patients with metabolic disorders [43]. Therefore, FN could be an adjuvant therapeutic modality in controlling glucose homeostasis in T2D patients. However, intensive blood pressure control and prolonged use of FN in T2D patients may increase the risk of kidney dysfunction, as evident by increased serum creatinine [44]. Different studies implicated FN in the deterioration of kidney function by increasing serum creatinine levels [45, 46]. Evidence from clinical studies illustrated that FN use within weeks increases serum creatinine levels [47]. It has been reported that prolonged use of 200 mg/day of FN increases creatinine production and serum creatinine level without any effect on the eGFR [44]. Nevertheless, this effect is only observed in patients with underlying kidney diseases or when used with drugs that alter kidney haemodynamics, such as renin-angiotensin (RAS) inhibitors [47]. The mechanism underlying the harmful effects of FN on renal function may involve the inhibition of vasodilator prostaglandin, and glomerular pressure and reduce creatinine clearance [47]. However, FN is effective in managing hypertensive glomerulosclerosis and diabetic nephropathy [47].

Regarding the potential role of FN in the regulation of glucose homeostasis, Holm et al. [42], confirmed that FN improves glucose homeostasis in mice by increasing sulfatide and long-chain sphingolipids, which improve pancreatic islet insulin sensitivity. Moreover, FN improves blood glucose homeostasis by attenuating inflammation and apoptosis in the pancreatic  $\beta$  cells [36]. An experimental study demonstrated that administration of FN in mice increases the size of sympathetic neurons and tyrosine hydroxylase activity in the pancreatic islet cells through enhancement of pancreatic β function [42]. Remarkably, FN can reverse autoimmune response in nonobese diabetic mice by augmenting the level of sulfatide and long-chain sphingolipids in the pancreatic islet with subsequent improvement of pancreatic insulin sensitivity [48]. Different experimental studies revealed that sulfatide and long-chain sphingolipids improve pancreatic β cells [49, 50]. Sulfatide and long-chain sphingolipids are regarded as immune modulators and regulate insulin secretion from pancreatic  $\beta$  cells [51]. Furthermore, FN protects against liver lipogenesis and hepatic IR by inhibiting the development of endoplasmic reticulum (ER) stress [52].

Moreover, PPAR- $\alpha$  is essential in the expression of hepatokines which involve the regulation of nutrition in mice [53]. Notably, hepatokines which are hepatic-derived proteins improve glucose homeostasis [54]. Sparc-related modular calcium-binding protein 1 (SMOC1) regulates glucose homeostasis in mice and might be a therapeutic target in managing T2D [54]. Circulating SMOC1 is positively correlated with insulin sensitivity [55]. Hepatokine fibroblast growth factor 21 (FGF21) is improved by long-term FN therapy, and this raises insulin sensitivity in T2D [56]. As a result, FGF21 appears to be a possible target for T2D therapy [57]. Furthermore, T2D and the development of the metabolic syndrome are predicted by FGF21 resistance [58]. It has been documented that patients with impaired glucose tolerance and T2D have increased levels of circulating FGF21 as a compensatory mechanism [59].

These findings suggest that FN mitigates IR, improves pancreatic  $\beta$  cells and could be a therapeutic strategy in the management of T2D (Figure 3).

### 4.2 | FN in AD

The distribution of brain PPAR-α was initially recognised by Warden and Truitt [20] by immunofluorescence. PPAR- $\alpha$  is expressed in all cell types in mice and humans [20]. PPAR- $\alpha$ promotes the expression of N-methyl-D-aspartate (NMDA) receptors, and adenosine monophosphate protein activated (AMPA) receptor-associated glutamate receptor, which improves synaptic plasticity via cyclic response element binding protein (CREBP) [60]. PPAR-α regulates glutamate neuronal homeostasis by increasing the expression of glutamate transporters in astrocytes [61]. Glutamate transporters in astrocytes are downregulated by the presence of Aß in AD, leading to postsynaptic overstimulation and neurotoxicity [61]. Originally, it has been established that FN through activation of PPAR-α inhibits beta-site amyloid precursor protein enzyme 1 (BACE-1), thereby reducing soluble APPB and  $A\beta_{42}$  release [62]. However, FN has no inhibitory effects on the presenilin-1 (PS-1) and APP [62]. Thus, FN affects the amyloidogenic process and may reduce AD neuropathology. FN is regarded as an inverse  $\gamma$ -secretase modulator which increases the generation of  $A\beta_{1-42}$  [63]. Also, FN enhances the clearance of  $A\beta_{1-42}$  [63]. Nonetheless, FN, through modulation of  $\gamma$ -secretase, promotes the production of highly aggregated  $A\beta_{42}$  and reduces  $A\beta_{38}$  production [64]. In this sense, FN and selective cyclooxygenase (COX-2) inhibitors, coxibs, were considered as diverse compounds that induce AD development in animal model studies [64]. Even though  $A\beta$  is the primary pathology of AD in preclinical studies, most clinical trials failed to confirm this hypothesis [18]. Therefore, the intervention of early risk factors involved in AD neuropathology such as apolipoprotein E4 (ApoE) by PPAR- $\alpha$  activators may reduce AD risk [18].

Notably, PPAR-α is intricate in various brain disorders, including depression, schizophrenia, sleep disorders, stroke and epilepsy, through modulation of hippocampal brainderived neurotrophic factor (BDNF), endocannabinoid and cholinergic signalling [18]. In addition, PPAR- $\alpha$  agonists such as Wy14643 improve cognitive function and prevent scopolamine-induced cognitive dysfunction in mice [65]. A previous study by de la Monte observed that PPAR-α was downregulated in AD brains [66], suggesting an involvement of PPAR- $\alpha$  in AD neuropathology. Notably, endogenous A $\beta$ production is augmented in PPAR-α knockout mice [67], suggesting a protective role of PPAR agonists against AD neuropathology. Besides, PPAR-α agonists have been reported to produce beneficial effects in many preclinical AD models [68, 69]. PPAR- $\alpha$  agonist gemfibrozil attenuates A $\beta$ -induced neurotoxicity and burden with a reduction of astrogliosis and microgliosis in the hippocampus and cortex of mice [68]. Likewise, cinnamic acid, a potent PPAR-α activator, inhibits cognitive impairment in the AD mouse model [68]. Cinnamic acid upregulates transcription factor EB (TFEB) and stimulates lysosomal biogenesis. Moreover, cinnamic acid reduced cerebral amyloid plaque burden and improved memory via PPARα [68]. Therefore, stimulation of lysosomal biogenesis by cinnamic acid may have therapeutic implications for the treatment of AD and other lysosomal disorders originating

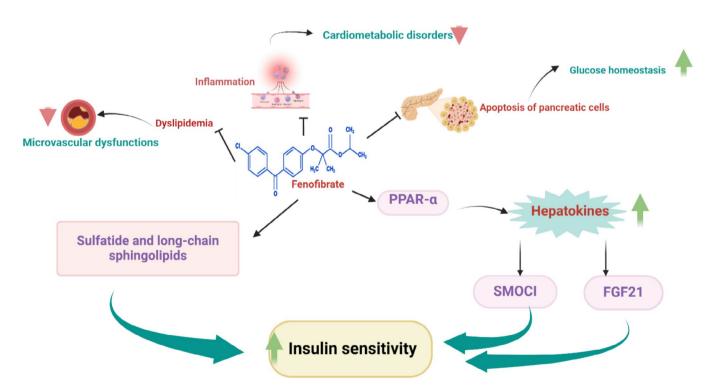


FIGURE 3 | The role of FN in T2D: FN by activating PPAR- $\alpha$  promotes the release of hepatokines which increase the expression of fibroblast growth factor 21 (FGF21) and Sparc-related modular calcium-binding protein 1 (SMOC1) that increase insulin sensitivity. In addition, FN by increasing the release of sulfatide and long-chain sphingolipids augments insulin sensitivity.

from accumulation of toxic protein aggregates. In particular, PPAR-α agonists, Wy14643, and gemfibrozil enhance the cognitive abilities of AD mice models by augmentation of autophagosome biogenesis [70]. Gemfibrozil and Wy14643 induce autophagy in human microglia cells and U251 human glioma cells stably expressing the human APP and this effect is PPARα-dependent. Administration of PPARα agonists decreases amyloid pathology and reverses memory deficits and anxiety symptoms in APP-transgenic mice. There is a reduced level of soluble  $A\beta$  and insoluble  $A\beta$  in hippocampus and cortex tissues from APP transgenic mice after treatment with either gemfibrozil or Wy14643, which promoted the recruitment of microglia and astrocytes to the vicinity of Aß plaques and enhanced autophagy [70]. These results indicated that PPARα is an important factor regulating autophagy in the clearance of  $A\beta$  and suggested gemfibrozil be assessed as a possible treatment for AD. A clinical trial conducted in 2020 that evaluated the possible protective effect of gemfibrozil on AD neuropathology in subjects with normal or mild cognitive impairments illustrated that gemfibrozil improves cognitive function [18]. Gemfibrozil has been shown to increase mir-107, which is involved in the regulation of BACE-1 [71]. In AD patients, mir-107 is reduced, causing activation of BACE-1 and progressive A\beta production and advancement of AD neuropathology [71]. Interestingly, a disintegrin and metalloproteinase domain-containing protein 10 (ADAM10), commonly known as α-secretase, is activated by gemfibrozil and other PPAR-α agonists, triggering the neuroprotective nonamyloidogenic pathway [67]. Hippocampal integrity and synaptic plasticity are enhanced by the neuroprotective soluble  $APP\alpha$ that is produced when  $\alpha$ -secretase cleaves APP [72]. Thus, PPAR-α agonists can decrease AD neuropathology by shifting APP proteolysis towards the nonamyloidogenic pathway. In addition, activation of PPAR- $\alpha$  by other agents such as low dose of aspirin and statins induces a significant reduction of Aβ-induced neurotoxicity and AD neuropathology [73, 74]. Of note, aspirin is one of the most widely used medications in the world, upregulates TFEB and increases lysosomal biogenesis in brain cells. Excitingly, aspirin induced the activation of PPAR-α and stimulated the transcription of TFEB gene via PPARα. Oral administration of low-dose aspirin decreased amyloid plaque pathology in both male and female transgenic mice in a PPARα-dependent fashion [70]. The inhibitors of HMG-CoA reductase and cholesterol-lowering drugs, statins, were found to excite the expression of neurotrophins in brain cells independent of the mevalonate pathway. Statins serve as ligands of PPARα and that Leu331 and Tyr 334 residues of PPARα are important for statin binding. Statins increase neurotrophins through PPARα-mediated transcriptional activation of CREB signalling. Accordingly, simvastatin increases CREB and brain-derived neurotrophic factor (BDNF) in the hippocampus of mice [71]. Furthermore, metformin inhibits DPP4 circulating level and increases glucagon-like peptide 1 (GLP-1) via PPARα and is independent of AMPK in mice. Thus, metformin exerts glucoregulatory actions through modulation of the incretin axis [75]. It has been shown that myocardial lipid accumulation, oxidative stress, apoptosis and cardiac remodelling and dysfunction induced in T2D by low streptozotocin doses and high-fat diets were considerably reversed by GLP-1 agonists and analogues treatments for 8 weeks in mice [76]. GLP-1 protects cardiac function by inhibiting the

ROCK/PPAR $\alpha$  pathway, thereby ameliorating lipotoxicity in diabetic cardiomyopathy [76]. Both metformin and GLP-1 agonists and analogues have neuroprotective effects against the development and progression of AD [77, 78].

Remarkably, PPAR-α agonists improve synaptic plasticity in male mice only due to higher expression of brain PPAR-α in males [79]. PPARα mediates the improvement of hippocampal synaptic plasticity upon nuclear retinoid X receptor (RXR) activation in a transgenic mouse model with cognitive deficits. This improvement results from an increase in GluA1 subunit expression of the AMPA receptor, eliciting an AMPA response at the excitatory synapses. Associated with a two times higher PPARα expression in males than in females, thus; male, but not female, PPARα null mutants display impaired hippocampal long-term potentiation [79]. Moreover, PPARα knockdown in the hippocampus of cognition-impaired mice compromises the beneficial effects of RXR activation on synaptic plasticity only in males. Furthermore, selective PPARa activation with pemafibrate improves synaptic plasticity in male cognition-impaired mice, but not in females [79]. Interestingly, hormones are known to influence the expression of PPARα in a sex-specific manner because gonadectomy of male rats decreases PPAR $\alpha$  expression levels. Oestrogens are known to improve synaptic plasticity, and behaviour is affected in ovariectomised female rats [80]. Thus, PPAR-α agonist FN enhances synaptic plasticity in male but not in female mice [79], suggesting a sexual dimorphism in response to PPAR-α agonists in AD management, and this may explain a higher AD risk in women [81]. Of interest, both oestrogen and androgen improve PPAR-α expression [82, 83]. Oestrogen replacement therapy in menopausal women improves cognitive function and reduces AD risk by increasing the expression of neuronal PPAR- $\alpha$  [84].

These mouse models do not fully reflect all pathological changes observed in patients and translating synaptic plasticity changes in mice with cognitive deficits in humans is challenging. However, a PPAR $\alpha$  agonist bexarotene improves cognition in mouse models, and in a patient with mild AD [85].

Moreover, PPAR-α agonist FN regulates the endogenous inhibitor of glutamate kynurenic acid, which is derived from L-tryptophan and involved in memory impairment [86, 87]. PPAR-α agonist gemfibrozil improves glutaminergic neurotransmission and associated memory function by inhibiting the synthesis of kynurenic acid [88, 89]. Also, PPAR- $\alpha$  increases the expression of nicotinic acetylcholine receptor alpha 7 (nA-ChR- $\alpha$ 7) [90], which is downregulated in AD [91]. Notably, nAChR-α7 is highly expressed in the cerebral cortex and hippocampus, mainly in the presynaptic and postsynaptic neurons [91]. In AD, nAChR- $\alpha$ 7 is highly dysregulated by A $\beta$ , which is disseminated on the cell membrane via the internalisation of these receptors [91]. The interaction between nAChR- $\alpha$ 7 and Aβ leads to inhibition and stimulation at different brain regions causing abnormal neurotransmission in AD model [92]. However, the main effect of A $\beta$  is inhibitory on the nAChR- $\alpha$ 7 since nAChR- $\alpha 7$  agonists improve cognitive function in animals treated by  $A\beta$  [93].

In addition, PPAR- $\alpha$  signalling phosphatidylinositol 3 kinase (PI3K) inhibits the A $\beta$  production [94]. PI3K pathway has a

neuroprotective effect against AD neuropathology. Notably, nAChR- $\alpha$ 7 through activation of the PI3K pathway inhibits A $\beta$ -induced neurotoxicity [95]. Interestingly, the universal PPAR activator GFT1803 inhibits production and increases the clearance of A $\beta$ , though PPAR- $\alpha$  agonists are more effective in reducing A $\beta$  production [96].

Moreover, PPAR- $\alpha$  regulates various cellular processes, including oxidative stress, mitochondrial dysfunction and neurotransmission dysfunction, mainly in the amygdala and prefrontal cortex [97]. Therefore, PPAR- $\alpha$  agonists may be recommended in the treatment of neurodegenerative disorders, including AD. PPAR- $\alpha$  agonist FN has been reported to be effective in different neurodegenerative disorders including PD, MS and amyotrophic lateral sclerosis (ALS) [97]. For example, PPAR- $\alpha$  agonist FN reduces cognitive dysfunction in preclinical and PD patients [98]. Of interest, FN reduces depressive symptoms through the modulation of neurosteroids [99] that are common in different neurodegenerative disorders, including AD.

Taken together, PPAR- $\alpha$  agonist FN seems to be effective in both AD and T2D. However, the underlying mechanistic role of FN in these conditions needs to be elucidated according to cellular and molecular pathways (Figure 4).

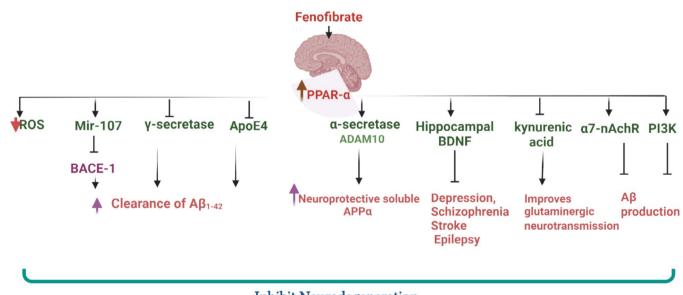
### 5 | Mechanistic Role of FN in AD and T2D

### 5.1 | Immunoinflammatory Response

It has been shown that abnormal immunoinflammatory response could be the possible mechanism linking T2D and AD [100]. IR and the pathophysiology of T2D are considerably influenced by systemic inflammation. In general, microglia respond to peripheral inflammation in an adaptable manner.

After exposing mice to IL-1β and TNFα systemically, the hippocampus exhibits the production of cytokines and chemokines [25, 101-103]. It has been illustrated that immune system stimulation in mice causes brain pathologies similar to AD, which include the accumulation of APP and its proteolytic fragments as well as changes in tau phosphorylation [104]. Higher peripheral amounts of IL-6, TNF-α, IL-1β, TGF-β, IL-12 and IL-8 are linked with AD [105]. Patients with AD who are in the early stages of the disease have activated peripheral immune cells in circulation. Peripheral inflammation and cognitive dysfunction have also been related in a number of studies. Peripheral inflammatory indicators, for instance, have been seen in AD patients [106]. Increased peripheral TNF- $\alpha$  and IL-1 $\beta$  levels have been linked to an increased risk of AD [106]. It has been demonstrated that proinflammatory cytokines can cross the BBB [107]. Gene expression analysis has demonstrated that brain-specific inflammatory responses are triggered by peripheral inflammation [107]. An increase in central inflammation is also likely to result from harm to the BBB and subsequent immune cell infiltration. It is unclear whether cerebral inflammation develops from peripheral inflammation or the other way around [106, 107]. These findings highlighted that peripheral and central immunoinflammatory responses are intricate in the development and progression of both AD and T2D.

By modifying the immunological response of T cells, FN can reduce the autoimmune response in mice with experimental Sjogren syndrome [108]. Mice lacking in PPAR- $\alpha$  exhibited aberrant immunological reactions to the inflammatory mediators such as leukotrienes and prostaglandins [22, 109–113]. PPAR- $\alpha$  ligands reduce the expression of adhesion molecules, cyclooxygenase-2 (COX-2) and IL-6 [114]. By upregulating the production of NF- $\kappa$ B inhibitor (I $\kappa$ B $\alpha$ ), PPAR- $\alpha$  ligands suppress the expression of NF- $\kappa$ B [115]. It has been shown that the PPAR- $\alpha$  ligand WY14643 prevents IgG from interacting with myelin



Inhibit Neurodegeneration Decrease risk of AD

**FIGURE 4** | Role of FN in AD: FN attenuates AD neuropathology by activating PPAR- $\alpha$  which through modulation of many signalling pathways and neurotrophic factors can inhibit the production and augment the clearance of A $\beta$ . Also, FN activates the neuroprotective nonamyloidogenic pathway by activating  $\alpha$ -secretase. For detail see the text above.

oligodendrocytes in mice. FN and other PPAR- $\alpha$  ligands reduce inflammation by promoting the release of anti-inflammatory cytokines [115, 116]. Remarkably, PPAR- $\alpha$  ligands increase the production of Th2 cytokines [117]. FN is able to decrease Th17 differentiation compared to other PPAR- $\alpha$  agonists such as bezafibrate. FN inhibits the expression of STAT3 and IL-21, both of which are necessary for Th17 differentiation [23, 98, 118–121]. A case–control study revealed that prolonged use of FN lowers inflammatory markers, such as CRP and IL-6 in patients with dyslipidaemia [122].

Notably, COX-2 upregulation is associated with vascular inflammation and impairment of vasomotor response in T2D [123]. Targeting of COX-2 by specific inhibitors attenuates the development of long-term complications such as diabetic neuropathy [124]. Similarly, IL-6 serum level is augmented in obese T2D patients compared to healthy controls [125]. In addition, NF-kB is upregulated and associated with T2D complications in both animals and humans [126]. Therefore, Th2 cytokine production is impaired in T2D patients leading to abnormal immune response and immunoinflammatory response [127]. Overactivated NFκB is engaged with the development of synaptic dysfunction in diabetic encephalopathy [128]. Consequently, an excess of inflammation coupled with an aberrant immune response may represent a potential connection between AD and T2D [129]. In addition, COX-2 expression is elevated in AD and linked with the development of neuroinflammation, thus; inhibiting COX-2 may be helpful in the treatment of AD [130]. Compared to healthy controls, AD patients had higher serum levels of IL-6 [131]. Dysregulation of the anti-inflammatory Th2 immune response may be the cause of this aberrant immunoinflammatory response in AD patients. Furthermore, the pathophysiology of both AD and T2D involves the nod-like receptor pyrin 3 receptor (NLRP3) inflammasome [132] which is involved in the pathophysiology of both AD and T2D [133, 134]. The maturation of IL-1β and IL-18 as well as the activation of caspase-1 are mediated by the NLRP3 inflammasome [133, 134]. NF-κB is one of the several triggers that activate the expression of the NLRP3 inflammasome [135–139]. The pathophysiology of neuroinflammation and neurodegeneration are intricately linked to the activation of NLRP3 inflammasome [133]. Additionally, NLRP3 inflammasome is amplified and associated with the severity of AD [134]. Within activated microglia, the NLRP3 inflammasome stimulates the production and release of IL-1β and IL-18. The pathophysiology of AD is linked to abnormal activation of the NLRP3 inflammasome, as demonstrated by preclinical and clinical results [134]. Elevations of IL-1 $\beta$  CSF levels in patients with severe AD indicate an overactivation of the NLRP3 inflammasome [134]. AD severity can be attenuated by using certain inhibitors to target the NLRP3 inflammasome [134]. Importantly, FN inhibits the production of NLRP3 inflammasome and NFκB, thus; it has a strong anti-inflammatory effect against the development of lung inflammation [140]. By blocking the NF-κB signalling pathway, FN prevents retinal damage and injury of the retinal blood barrier [141]. Additionally, by suppressing the expression of NLRP3 inflammasome, FN can decrease the severity of diabetic retinopathy [141].

Furthermore, innate immune sensors known as toll-like receptors (TLRs) alert the immune system to the presence of exogenous pathogens [100]. When TLR is activated, the

proinflammatory cytokines are released, and the adaptive immune system is activated to get rid of invasive pathogens [100]. TLR is capable of identifying danger signals that result from tissue damage and inflammation. TLRs play a critical role in AD neuropathology and are abundantly expressed by immune cells in the CNS [142]. TLR agonists contribute to the exaggeration of detrimental inflammatory reactions. Since PPAR-α activation inhibits TLR expression through many pathways, it has been proven that PPAR-α agonists and TLRs have reciprocal interactions [143]. FN suppresses CD14 expression which raises TLR expression and causes the release of pro-inflammatory cytokines [144]. Furthermore, FN suppresses the expression of the MyD88-TLR4 signalling pathway and the release of IL-12 [145]. TLRs are greatly upregulated in T2D, contributing to the advancement of inflammatory diseases and associated diabetic complications [146]. TLRs may, therefore, be a possible connection between AD and T2D. Thus, by blocking TLRs and their effectors, the PPAR-α agonist FN can suppress the first immunological response in AD neuropathology.

Therefore, FN through modulation of the immunoinflammatory signalling pathway can mitigate the pathogenesis of T2D and AD neuropathology.

### 5.2 | Neuroinflammation

The development of various neurodegenerative diseases is linked with the development of neuroinflammation [147]. Neuroinflammation is created by the activation of T and B lymphocytes in the CNS [148]. For instance, synaptopathy caused by neuroinflammation during the early stages of MS can occur independently of the demyelination process, which could account for cognitive failure in MS patients [149]. Exaggerated immunological disruption and the progression of neuroinflammation in the late stages of MS contribute to the pathophysiology of the disease [149, 150]. Patients with AD have reduced cholinergic activity, which controls immune cell response and activity [151]. When immune cells' levels of acetylcholine are lowered, the proinflammatory cytokines are released leading to the development of neuroinflammation [151, 152]. Thus, reducing neuroinflammation may be a useful therapeutic approach to reduce the neuropathology associated with AD. A low-grade chronic inflammatory disease accelerates the development of neuroinflammation in T2D [153]. Inhibition of the proinflammatory cytokines and the inflammatory signalling pathway has been shown in an experimental study to reduce the development of neuroinflammation in diabetic mice [154]. Hyperglycaemiainduced BBB injury promotes the transport of inflammatory cells and the development of neuroinflammation in T2D [155].

Various preclinical studies have demonstrated the protective effect of FN against the onset and progression of neuroinflammation [156, 157]. In rats with severe brain damage, FN reduces neuroinflammation by inhibiting the development of oxidative stress [134]. By preventing oxidative stress, neuroinflammation and mitochondrial dysfunction, the risk of neurodegeneration is reduced. Importantly, FN exerts a neuroprotective impact against the onset and progression of MS by suppressing oxidative stress, neuroinflammation and mitochondrial dysfunction [157]. The suppression of inflammatory signalling pathways as

well as antioxidant and anti-inflammatory actions are the fundamental mechanisms by which FN contributes to the inhibition of neuroinflammation [156, 157]. FN inhibits neuroinflammation by activating neurons nicotinic cholinergic receptors [158]. These results suggested that FN may modify neuroinflammatory responses to lower the aetiology of AD and T2D (Figure 5).

### 5.3 | Oxidative Stress

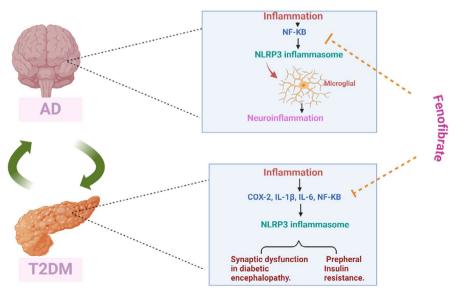
Oxidative stress plays a crucial role in the development of AD by increasing neuronal injury and apoptosis [16, 138, 159, 160]. It has been revealed that reactive oxygen species (ROS) which facilitate the activation of microglia can induce neuronal injury and apoptosis [161]. In AD, inflammatory responses can trigger the development of oxidative stress in the activated macrophages and microglia, resulting in neuronal injury [162]. Of interest, oxidative stress and the generation of ROS contribute to the progression of inflammation in AD [161]. As a result, oxidative stress and inflammation are positively activated in a vicious cycle in AD. According to these results, oxidative stress may intensify inflammatory responses, increase neuronal damage and accelerate the progression of AD neuropathology [161]. Consequently, using of antioxidants may prevent AD development and progression. Interestingly, antioxidants may be advantageous for human AD by reducing oxidative stress and associated inflammatory changes [163]. For example, the antioxidant alpha lipoic acid has been shown in preclinical and clinical trials to attenuate brain atrophy and enhance the clinical course of AD [164]. Similarly, oxidative stress is increased in T2D due to IR, redox imbalance and hyperglycaemia [165]. Furthermore, ER stress induced by hyperglycaemia boosts the production of ROS, which damages pancreatic  $\beta$  cell function and causes hyperglycaemia in a positive feedback loop [165]. Moreover, ROS production is increased by IR-induced lipotoxicity, which leads to pancreatic  $\beta$ -cell dysfunction [165]. NADPH oxidase activity and ROS were shown to be greater in the monocytes of T2D patients compared to controls [166]. Thus, oxidative stress is implicated in the aetiology and consequences of T2D.

Furthermore, PPAR- $\alpha$  agonists have been demonstrated to have strong antioxidant properties and may be able to treat a variety of neurodegenerative diseases including AD [167]. In AD mouse models, lipid peroxidation, oxidative stress and inflammation are inhibited by the PPAR- $\alpha$  agonist GW7647 [167]. In addition, FN increases antioxidant capacity and reduces oxidative damage brought on by hyperglycaemia [168]. Similarly, a PPAR- $\alpha$  agonist clofibrate, reduces neuroinflammation and oxidative stress in AD rat model [169]. Furthermore, lipid peroxidation, oxidative stress and inflammation are inhibited by the PPAR- $\alpha$  agonist GW7647 in T2D model [170].

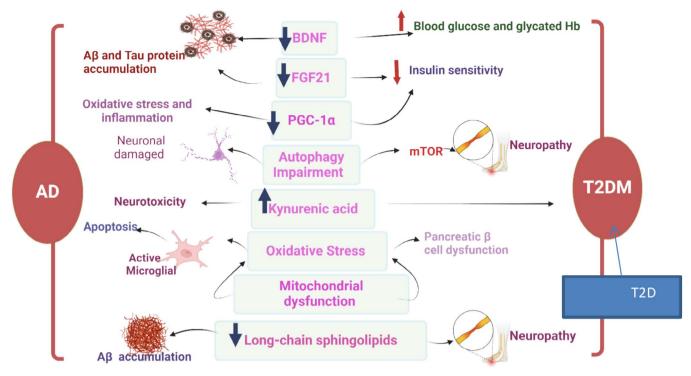
Thus, FN can reduce the pathophysiology of AD and T2D by suppressing oxidative stress and enhancing endogenous antioxidant capacity (Figure 6).

# 5.4 | Mitochondrial Dysfunction

Mitochondrial dysfunction plays a crucial role in the degeneration of neurons and axons in AD [171, 172]. The primary mechanism behind the development of mitochondrial dysfunction in AD is the impairment of mitochondrial permeability transition pores by Ca<sup>2+</sup> dyshomeostasis and ROS [171, 173]. In AD, mitochondrial failure is thought to be a significant initiator of programmed neuronal death [174]. Serum lactate and uric acid are thought to be possible indicators of mitochondrial malfunction [175]. The aetiology of T2D also includes mitochondrial dysfunction [176]. Mitochondrial dysfunction is caused by changes in mitochondrial DNA due to ageing, obesity and dyslipidaemia [176]. Mitochondrial dysfunction is correlated with the emergence of IR, ER stress and ROS release [177]. Consistently, mitochondrial oxidative stress



**FIGURE 5** | Role of inflammation in AD and T2D: FN inhibits the expression of inflammatory signalling pathways such as NF- $\kappa$ B and NLRP3 inflammasome, suppresses the release of proinflammatory cytokines and inflammatory molecules which implicated in the development of neuroinflammation in both AD and T2D.



**FIGURE 6** | The crosstalk between AD and T2D: Many signalling pathways, cellular pathways and neurotrophic factors are dysregulated in both T2D and AD. For detail see the text.

in the peripheral lymphocytes is higher in T2D patients compared to healthy controls [178].

Therefore, mitigating mitochondrial dysfunction could be a mechanistic approach to halt the progression of both T2D and AD. PPAR- $\alpha$  agonists play a fundamental role in regulating mitochondrial function in diabetic patients [179]. Interestingly, FN improves insulin sensitivity by enhancing mitochondrial  $\beta$ -oxidation [180]. Additionally, FN inhibits mitochondrial dysfunction in burn patients [179]. Moreover, FN promotes neurogenesis by modulating mitochondrial biogenesis in the experimental ischemic reperfusion injury [181]. The protective effect of FN against mitochondrial dysfunction is achieved by increasing the expression of mitochondrial uncoupling protein 2, which shields mitochondria from oxidative stress by reducing the production of ROS [181]. Consequently, FN shows potential efficacy in combating AD and T2D by suppressing mitochondrial dysfunction.

# 5.5 | Autophagy

Autophagy is an essential process within cells that eliminate harmful misfolded proteins and damaged organelles. Impaired neuronal autophagy triggers the development and progression of neurodegeneration [182–185]. In addition, autophagy regulates both adaptive and innate immunity, and any anomaly in autophagy can lead to an aberrant immunological response [186]. The survival and homeostasis of neurons mostly rely on basal neuronal autophagy. Malfunctioning autophagy plays a role in the development of AD [187]. Administering an early blockade of the mechanistic target of rapamycin (mTOR) a negative regulator of autophagy effectively decreases AD

neuropathology in mice. Nevertheless, the suppression of mTOR during the later stage of AD worsens the neuropathology of the disease [188]. The process of autophagy can have both beneficial and harmful effects [188]. For instance, enhancing autophagy in T and B cells stimulates the progression of neuroinflammation, while suppressing autophagy in this context could potentially be a beneficial approach in the treatment of AD [188]. Surprisingly, restoration of basal neuronal autophagy function in late AD impedes the progression of disease severity [188, 189]. The autophagy process is also dysregulated in T2D as it increased in T2D compared to healthy controls [190]. The autophagy process in T2D is upregulated as a compensatory mechanism in response to IR, hyperglycaemia and proinflammatory cytokines [191]. Therefore, inhibition of the autophagy process and its regulator mTOR may attenuate T2D-mediated complications [192]. However, an updated study illustrated that autophagy comprises T2D and ageing. Loss of autophagy promotes the development and progression of diabetic complications. Thus, activation of autophagy by miRNA may be a novel therapeutic option in the management of T2D and related complications [193].

These findings suggest a controversy regarding the potential role of autophagy in AD and T2D. Targeting of neuronal autophagy is crucial for understanding the neuropathology of AD. FN enhances the autophagy process, according to multiple studies [194, 195]. For instance, FN increases SIRT1 expression and autophagy function to mitigate myocardial damage in diabetic rats [194]. Additionally, FN inhibits acute kidney injury by controlling the autophagy process by inducing the expression of AMPK [99]. Both SIRT1 and AMPK activate autophagy process [196, 197]. Therefore, FN may slow the development of AD and T2D by activating autophagy process.

# 5.6 | Brain-Derived Neurotrophic Factor (BDNF)

BDNF is a growth factor produced and released from glial cells and neurons. BDNF controls synaptic plasticity, neurotransmission and neuronal development and maturation [198-203]. BDNF acts by activating tropomyosin receptor kinase B (TrkB) receptor which is widely expressed in the CNS. BDNF has a neuroprotective effect against many neurodegenerative diseases including AD [204]. Neuroinflammation, tau protein phosphorylation, neuronal death and Aß inhibit BDNF signalling. BDNF/TrkB induces PI3K and improves synaptic function by modifying intracellular glutamate and Ca<sup>2+</sup> neurotransmission [204]. Notably, Aβ accumulation interferes with BDNF/TrkB signalling which is extremely dysregulated in AD [153, 204]. BDNF and pro-BDNF are reduced in the human brains in early AD neuropathology [205]. Ng et al. [206] found that BDNF serum levels were reduced in AD patients compared to the healthy controls. Similarly, CSF BDNF levels were lower in AD patients compared to the healthy controls [207]. Furthermore, the severity of cognitive impairment is correlated with low serum levels of BDNF [208].

On the other hand, BDNF regulates insulin sensitivity and glucose homeostasis [209]. A case–control study showed that hyperglycaemia blocks BDNF output from the brain in T2D and healthy controls [209]. In addition, BDNF is expressed in many no-neuronal tissues, including pancreatic  $\alpha$  cells and cardiomyocytes [210]. Platelets are the main source of peripheral BDNF in T2D [210]. BDNF reduces blood glucose and glycated Hb by central inhibiting of food intake and through the peripheral effect by impeding glucagon release from the pancreatic  $\alpha$  cells [211, 212]. Mutation of BDNF/TrkB signalling induces the development of obesity and IR [213]. Also, the reduction of circulating BDNF is correlated with a reduction in insulin sensitivity [214]. These results highlighted that BDNF is decreased in T2D and augmentation of BDNF may reduce IR and T2D-related complications.

Furthermore, FN reduces the impact of hippocampal ischemia-reperfusion and enhances cognitive performance in rats [215]. The neuroprotective effect of FN is mediated by upregulating the expression of BDNF [216]. Additionally, PPAR- $\alpha$  agonists increase the expression of hippocampus BDNF, which reduces the progression of neurodegeneration in neurodegenerative diseases [216]. BDNF controls the activity of microglia and prevents microglia-induced neurodegeneration [217]. Specifically, naturally occurring phytochemicals such as curcumin, cannabinoids and genistein have been found to inhibit the progression of neurodegenerative disorders by upregulating the expression of BDNF [218]. Moreover, researches have been demonstrated that PPAR- $\alpha$  agonists can enhance the treatment of T2D by increasing the activity of BDNF [219].

These data indicate that PPAR- $\alpha$  agonist FN can decrease the development of AD neuropathology and T2D pathogenesis by enhancing the production of BDNF.

### 5.7 | Fibroblast Growth Factor (FGF21)

FGF21 is a metabolic hormone also known as hepatokine that is produced and secreted by hepatocytes [220]. FGF21 regulates lipid and glucose metabolism by inducing fatty acid oxidation and

gluconeogenesis, respectively, and it is abundantly expressed in the pancreatic islets and adipocytes [220, 221]. FGF21 regulates PPAR- $\alpha$  expression in a feedback loop to improve insulin sensitivity [221-224]. FGF21 has been shown by Chen et al. [225], to reduce the onset and progression of certain neurodegenerative diseases, such as AD. Results from studies conducted in vitro and in vivo demonstrated that FGF21 by upregulating protein phosphatase 2 A (PP2A) has a neuroprotective effect against oxidative stress and apoptosis caused by A\Beta and tau protein [225]. Defective and aberrant autophagy triggers the release of FGF21 as a compensatory mechanism to modulate the autophagy process in AD [150]. Larson and Gill [226] demonstrated that FGF21 signalling was dysregulated in both AD and cardiometabolic disorders such as T2D, suggesting that FGF21 could be a possible link between AD and T2D. It has been shown that long-term FN treatment improves FGF21, which enhances insulin sensitivity in T2D [56]. Therefore, FGF21 appears to be a potential therapeutic target for T2D [57]. Furthermore, the development of T2D and the metabolic syndrome are predicted by the development of FGF21 resistance [58]. There have been reports that T2D patients and impaired glucose tolerance had elevated levels of circulating FGF21 as a compensatory strategy [59]. Interestingly, a case-control study showed that FGF21 serum levels were shown to be lower in T2D patients compared to healthy controls [227]. Furthermore, the use of FN in those patients increased FGF21 levels in T2D patients. These findings pointed out that FGF21 is reduced in both AD and T2D patients. Therefore, increasing FGF21 by FN may improve these conditions or at least attenuate AD risk in T2D patients.

Moreover, AD and T2D are highly prevalent ageing-related diseases associated with significant morbidity and mortality [228-230]. Some findings in human and animal models have linked T2D to AD-type dementia. Despite epidemiological associations between T2D and cognitive impairment, the inter-relational mechanisms could be related to the metabolic derangement. T2D is associated with cardiovascular and metabolic disorders which augment AD neuropathology. Therefore, there is an overlap between the pathologic mechanisms of T2D and AD [228-230]. The nexus of cardiometabolic risk factors, collectively referred to as the metabolic syndrome, has insulin resistance and hypertriglyceridaemia at its centre, and has been associated with the development of major types of dementia, including AD [231]. Triglyceride-rich lipoprotein is involved in the development of AD in T2D patients by inducing the expression of brain ApoE4 [231]. Notably, FN is effective in treating hypertriglyceridaemia mainly in T2D patients [34] thereby FN can attenuate T2D-induced AD by regulating lipid profile.

# 5.8 | Long-Chain Sphingolipids

Long-chain sphingolipids are involved in the control of pancreatic  $\beta$ -cell function and regulate insulin secretion [232]. Indeed, decreased long-chain sphingolipid levels are linked with the progression of diabetic nephropathy [233]. An experimental study revealed that the ablation of very long-chain sphingolipids induces the development of IR [234]. In AD, long-chain sphingolipids are reduced and promote A $\beta$  deposition and neuronal injury in AD [235, 236]. Long-chain sphingolipids are diminished in AD in relation to the healthy controls [235, 236]; therefore, long-chain sphingolipid could

be a biomarker for AD neuropathology. In addition, sulfatide regulates pancreatic  $\beta$ -cell function and neuronal activity. Therefore, the reduction of sulfatide occurs in the pathophysiology of AD and T2D [237, 238]. Of note, FN can reverse autoimmune response in nonobese diabetic mice by augmentation level of sulfatide and long-chain sphingolipids in the pancreatic islet with subsequent improvement of pancreatic insulin sensitivity [48]. Different experimental studies revealed that sulfatide and long-chain sphingolipids improve pancreatic β cells [49, 50]. Sulfatide and long-chain sphingolipids are regarded as immune modulators and regulate insulin secretion from pancreatic  $\beta$  cells [51]. Moreover, FN protects against liver lipogenesis and hepatic IR by inhibiting the development of ER stress [52]. Consistently, sulfatide serum level is reduced in AD [237] and T2D [238]. These observations indicated that FN, through modulation of sulfatide and long-chain sphingolipids, improves the pathogenesis of both T2DM and AD.

# 5.9 | Kynurenic Acid

Research has demonstrated that the presence of mild inflammation and long-term stress significantly raises the likelihood of prediabetes progressing into T2D [239]. It has been proposed that kynurenic acid, which is derived from tryptophan, is upregulated and stimulates the development of T2D by inducing the development of IR [239]. Therefore, kynurenic acid is augmented in T2D patients in relation to the controls. However, exercise regulates IR and energy expenditure through modulation levels of kynurenic acid [240]. It has been noted that the baseline level of kynurenic acid was increased in obese patients when they develop T2D [241], suggesting that a higher kynurenic acid serum level is correlated with T2D risk. Furthermore, kynurenic acid, which is produced from astrocytes, is also involved in AD neuropathology [242]. Increased brain kynurenic acid is regarded as a response to the inflammatory reactions in AD and other neurodegenerative ailments [242]. Notably, CSF kynurenic acid level is increased in AD cases in relation to healthy controls [242]. However, serum and erythrocytes kynurenic acid levels are reduced in AD due to an increase in kynurenine aminotransferase [243]. Remarkably, kynurenic acid is neuroprotective at low levels and neurotoxic at higher levels [244]; thus, it is regarded as a double-sword edge in AD. Moreover, PPAR-α agonist FN regulates the endogenous inhibitor of glutamate kynurenic acid [86]. PPAR-α agonist gemfibrozil improves glutaminergic neurotransmission and associated memory function by inhibiting the synthesis of kynurenic acid [88, 89]. Therefore, PPAR- $\alpha$  agonist FN improves both AD and T2D through the modulation of the synthesis and release of kynurenic acid.

### 5.10 | PPAR- $\gamma$ Coactivator 1-Alpha (PGC-1 $\alpha$ )

PGC-1 $\alpha$  is the master regulator of mitochondrial biogenesis by controlling gluconeogenesis, energy metabolism, upregulating of autophagy and the unfolded protein response [245]. PGC-1 $\alpha$  controls cholesterol homeostasis, blood pressure, insulin sensitivity and obesity [246]. Because PGC-1 $\alpha$  increases insulin sensitivity, there is a correlation between lower PGC-1 $\alpha$  expression and risk of IR [247]. A case–control research revealed a link between PGC-1 $\alpha$  gene polymorphism and the onset of T2D [248].

However, a prior investigation did not find a connection between T2D and PGC- $1\alpha$  gene polymorphism [249].

Furthermore, PGC-1α expression is highly reduced and negatively associated with A\beta accumulation and amyloid plaque in AD model [250]. Of interest, deficiency of PGC-1 $\alpha$  is linked with hyperactivity and behavioural changes in mice [251]. PGC-1α has a neuroprotective role through anti-inflammatory and antioxidant effects [252]. Moreover, PGC-1α triggers the expression of neuroprotective receptors like PPAR-α and estrogenic receptors [251, 252]. Suwa et al. [253], illustrated that the insulin-sensitising drug metformin promotes the expression of PGC-1α. Metformin has been revealed to be effective against AD neuropathology through the activation of PGC-1α signalling [254]. In addition, PPAR- $\alpha$  agonist promotes peripheral and central PGC- $1\alpha$  expression [255]. Notably, FN, through activation of PGC-1α signalling can reduce lipotoxicity in mice with diabetic nephropathy [256] and neuropathy [257]. Therefore, FN, through activation of PGC-1α signalling, can have a dual role in mitigating T2D and AD neuropathology.

Taken together, FN has dual neuroprotective and antidiabetic effects that can mitigate AD neuropathology and T2D-related complications through the modulation of various cellular processes and inflammatory signalling pathways. However, the present review has several limitations as the molecular and genetic mechanisms linking AD and T2D were not completely discussed. In addition, the reciprocal relationships between AD and T2D in relation to the effect of FN were focused on the immunoinflammatory relationship with little discussion on the metabolic pathway which connects AD and T2D. Despite these limitations, this review sheds light and gives a clue to the potential beneficial effect of FN against the development and progression of AD in T2D. To verify this concept, additional preclinical trials and prospective studies are recommended in this regard.

### 6 | Conclusion

Evidence from epidemiological findings showed an association between T2D and AD. Due to the progression of brain IR and dysregulation of neuronal insulin receptors, T2D and IR enhance the risk of AD. These pathological modifications include aberrant deposition of Aß plaques, reduced brain glucose metabolism, increased generation of  $A\beta_{1\text{--}42}$  and impairment of  $A\beta$ clearance. Notably, PPAR-α is highly dysregulated in T2D and AD. A PPAR-α agonist FN which is mainly recommended in T2D patients with dyslipidaemia can improve lipid profile in T2D patients. Thus, FN is an effective agent in the attenuation of metabolic syndrome mediated by prediabetes. Similarly, FN improves impaired glucose tolerance through activation of PPAR-α which inhibits BACE-1, thereby reducing soluble APP $\beta$  and A $\beta_{42}$  release and enhancing  $A\beta_{1,42}$  clearance. In spite of the fact that  $A\beta$  is the primary pathology of AD; however, most clinical trials failed to confirm this hypothesis. Consequently, the intervention of early risk factors involved in AD neuropathology by PPAR-α activators may reduce AD risk. Remarkably, PPAR-α agonists induce a nonamyloidogenic pathway via activation of  $\alpha$ -secretase, which is also called ADAM10, leading to decreased AD neuropathology by shifting APP proteolysis towards nonamyloidogenic Aβ. Thus, PPAR- $\alpha$  agonist FN seems to be effective in both AD and

T2D; however, the underlying mechanistic role of FN in these conditions is not fully elucidated. It has been shown that FN via modulation of the immunoinflammatory signalling pathway can mitigate the pathogenesis of T2D and AD neuropathology. Furthermore, FN mitigates various cellular processes, including mitochondrial dysfunction, IR, oxidative stress, autophagy, immunoinflammatory disorders and ER stress, which are involved in the pathogenesis of T2D and AD neuropathology. Likewise, FN improves different mediators and signalling pathways that are intricate in the pathophysiology of both T2D and AD.

Taken together, FN has dual neuroprotective and antidiabetic effects that can mitigate AD neuropathology and T2D-related complications through the modulation of various cellular processes and inflammatory signalling pathways.

### **Author Contributions**

Mansour A. Alsaleem: writing – review and editing (equal). Hayder M. Al-Kuraishy: conceptualization (equal), conceptualization (equal), validation (equal), validation (equal). Ali I. Al-Gareeb: resources (equal), validation (equal), visualization (equal). Maha M. Abdel-Fattah: writing – review and editing (equal). Mohammed Alrouji: writing – review and editing (equal). Nasser A. Al-Harchan: writing – review and editing (equal). Mubarak Alruwaili: writing – original draft (equal). Marios Papadakis: funding acquisition (lead), writing – original draft (equal). Athanasios Alexiou: supervision (equal), visualization (equal), writing – original draft (equal). Gaber El-Saber Batiha: writing – review and editing (equal).

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### **Ethics Statement**

The authors have nothing to report.

### Consent

The authors have nothing to report.

# **Conflicts of Interest**

The authors declare no conflicts of interest.

### **Data Availability Statement**

All data are available in the manuscript.

### References

- 1. R. J. Castellani, R. K. Rolston, and M. A. Smith, "Alzheimer Disease," *Disease-a-Month: DM* 56, no. 9 (2010): 484–546.
- 2. J. L. Cummings and G. Cole, "Alzheimer Disease," *Journal of the American Medical Association* 287, no. 18 (2002): 2335–2338.
- 3. L. G. Y. Apostolova, "Alzheimer Disease," Continuum: Lifelong Learning in Neurology 22, no. 2 (2016): 419–434.
- 4. A. Serrano-Pozo, M. P. Frosch, E. Masliah, and B. T. Hyman, "Neuropathological Alterations in Alzheimer Disease," *Cold Spring Harbor Perspectives in Medicine* 1, no. 1 (2011): a006189.

- 5. D. M. Holtzman, E. Mandelkow, and D. Selkoe, "Alzheimer Disease in 2020," *Cold Spring Harbor Perspectives in Medicine* 2, no. 11 (2012): a011585.
- 6. T. D. Bird, "Alzheimer Disease Overview" (2018), GeneReviews®.
- 7. C. Mount and C. Downton, "Alzheimer Disease: Progress or Profit?," *Nature Medicine* 12, no. 7 (2006): 780–784.
- 8. S. Weis, M. Sonnberger, A. Dunzinger, et al., "Neurodegenerative Diseases: Alzheimer Disease (AD)," In *Imaging Brain Diseases*, Eds. S. Weis, M. Sonnberger, A. Dunzinger, et al., (Vienna, Austria: Springer, 2019), 897–931.
- 9. R. E. Tanzi, "The Genetics of Alzheimer Disease," *Cold Spring Harbor Perspectives in Medicine* 2, no. 10 (2012): a006296.
- 10. L. A. Profenno, A. P. Porsteinsson, and S. V. Faraone, "Meta-Analysis of Alzheimer's Disease Risk With Obesity, Diabetes, and Related Disorders," *Biological Psychiatry* 67, no. 6 (2010): 505–512.
- 11. S. C. Correia, R. X. Santos, G. Perry, X. Zhu, P. I. Moreira, and M. A. Smith, "Insulin-Resistant Brain State: The Culprit in Sporadic Alzheimer's Disease?," *Ageing Research Reviews* 10, no. 2 (2011): 264–273.
- 12. A. M. Koenig, D. Mechanic-Hamilton, S. X. Xie, et al., "Effects of the Insulin Sensitizer Metformin in Alzheimer Disease: Pilot Data From a Randomized Placebo-Controlled Crossover Study," *Alzheimer Disease and Associated Disorders* 31, no. 2 (2017): 107–113.
- 13. M. Barbagallo and L. J. Dominguez, "Type 2 Diabetes Mellitus and Alzheimer's Disease," World Journal of Diabetes 5, no. 6 (2014): 889–893.
- 14. B. Kim, C. Backus, S. S. Oh, and E. L. Feldman, "Hyperglycemia-Induced Tau Cleavage In Vitro and In Vivo: A Possible Link Between Diabetes and Alzheimer's Disease," *Journal of Alzheimer's Disease* 34, no. 3 (2013): 727–739.
- 15. J. K. Morris, E. D. Vidoni, J. D. Mahnken, et al., "Cognitively Impaired Elderly Exhibit Insulin Resistance and No Memory Improvement With Infused Insulin," *Neurobiology of Aging* 39 (2016): 19–24.
- 16. H. M. Al-Kuraishy, O. M. Sami, N. R. Hussain, and A. I. Al-Gareeb, "Metformin and/or Vildagliptin Mitigate Type II Diabetes Mellitus Induced-Oxidative Stress: The Intriguing Effect," *Journal of Advanced Pharmaceutical Technology & Research* 11, no. 3 (2020): 142–147.
- 17. H. Kim, M. Haluzik, Z. Asghar, et al., "Peroxisome Proliferator–Activated Receptor- $\alpha$  Agonist Treatment in a Transgenic Model of Type 2 Diabetes Reverses the Lipotoxic State and Improves Glucose Homeostasis," *Diabetes* 52, no. 7 (2003): 1770–1778.
- 18. F. Sáez-Orellana, J.-N. Octave, and N. Pierrot, "Alzheimer's Disease, a Lipid Story: Involvement of Peroxisome Proliferator-Activated Receptor  $\alpha$ ," *Cells* 9, no. 5 (2020): 1215.
- 19. G. M. Keating and K. F. Croom, "Fenofibrate: A Review of Its Use in Primary Dyslipidaemia, the Metabolic Syndrome and Type 2 Diabetes Mellitus," *Drugs* 67 (2007): 121–153.
- 20. A. Warden, J. Truitt, M. Merriman, et al., "Localization of PPAR Isotypes in the Adult Mouse and Human Brain," *Scientific Reports* 6, no. 1 (2016): 27618.
- 21. H. M. Al-Kuraishy, A. I. Al-Gareeb, M. Alblihed, S. G. Guerreiro, N. Cruz-Martins, and G. E. Batiha, "COVID-19 in Relation to Hyperglycemia and Diabetes Mellitus," *Frontiers in Cardiovascular Medicine* 8 (2021): 644095.
- 22. H. M. Al-Kuraishy, N. R. Hussian, M. S. Al-Naimi, A. I. Al-Gareeb, F. Al-Mamorri, and A. K. Al-Buhadily, "The Potential Role of Pancreatic γ-Aminobutyric Acid (GABA) in Diabetes Mellitus: A Critical Reappraisal," *International Journal of Preventive Medicine* 12 (2021): 19
- 23. H. A. Rasheed, H. M. Al-Kuraishy, A. I. Al-Gareeb, N. R. Hussien, and M. S. Al-Nami, "Effects of Diabetic Pharmacotherapy on Prolactin

- Hormone in Patients With Type 2 Diabetes Mellitus: Bane or Boon," *Journal of Advanced Pharmaceutical Technology & Research* 10, no. 4 (2019): 163–168.
- 24. N. R. Hussien, M. S. Al-Naimi, H. A. Rasheed, H. M. Al-Kuraishy, and A. I. Al-Gareeb, "Sulfonylurea and Neuroprotection: The Bright Side of the Moon," *Journal of Advanced Pharmaceutical Technology & Research* 9, no. 4 (2018): 120–123.
- 25. H. M. Al-Kuraishy, A. I. Al-Gareeb, H. J. Waheed, and T. J. Al-Maiahy, "Differential Effect of Metformin and/or Glyburide on Apelin Serum Levels in Patients With Type 2 Diabetes Mellitus: Concepts and Clinical Practice," *Journal of Advanced Pharmaceutical Technology & Research* 9, no. 3 (2018): 80–86.
- 26. H. Guan, K. M. Chow, R. Shah, C. J. Rhodes, and L. B. Hersh, "Degradation of Islet Amyloid Polypeptide by Neprilysin," *Diabetologia* 55 (2012): 2989–2998.
- 27. T. T. Nguyen, Q. T. H. Ta, T. K. O. Nguyen, T. T. D. Nguyen, and V. van Giau, "Type 3 Diabetes and Its Role Implications in Alzheimer's Disease," *International Journal of Molecular Sciences* 21, no. 9 (2020): 3165.
- 28. D. Milardi, E. Gazit, S. E. Radford, et al., "Proteostasis of Islet Amyloid Polypeptide: A Molecular Perspective of Risk Factors and Protective Strategies for Type II Diabetes," *Chemical Reviews* 121, no. 3 (2021): 1845–1893.
- 29. R. G. Bennett, F. G. Hamel, and W. C. Duckworth, "An Insulin-Degrading Enzyme Inhibitor Decreases Amylin Degradation, Increases Amylin-Induced Cytotoxicity, and Increases Amyloid Formation in Insulinoma Cell Cultures," *Diabetes* 52, no. 9 (2003): 2315–2320.
- 30. S. Zraika, R. L. Hull, J. Udayasankar, et al., "Identification of the Amyloid-Degrading Enzyme Neprilysin in Mouse Islets and Potential Role in Islet Amyloidogenesis," *Diabetes* 56, no. 2 (2007): 304–310.
- 31. P. Westermark, U. Engström, K. H. Johnson, G. T. Westermark, and C. Betsholtz, "Islet Amyloid Polypeptide: Pinpointing Amino Acid Residues Linked to Amyloid Fibril Formation," *Proceedings of the National Academy of Sciences of the United States of America* 87, no. 13 (1990): 5036–5040.
- 32. L. S. Ferreira, C. S. Fernandes, M. N. N. Vieira, and F. G. De Felice, "Insulin Resistance in Alzheimer's Disease," *Frontiers in Neuroscience* 12 (2018): 830.
- 33. V. Ormazabal, S. Nair, O. Elfeky, C. Aguayo, C. Salomon, and F. A. Zuñiga, "Association Between Insulin Resistance and the Development of Cardiovascular Disease," *Cardiovascular Diabetology* 17 (2018): 1–14.
- 34. J. P. Ferreira, F. Vasques-Nóvoa, D. Ferrão, et al., "Fenofibrate and Heart Failure Outcomes in Patients With Type 2 Diabetes: Analysis From ACCORD," *Diabetes Care* 45, no. 7 (2022): 1584–1591.
- 35. A. A. Abulaban, H. M. Al-kuraishy, A. I. Al-Gareeb, et al., "Role of Fenofibrate in Multiple Sclerosis," *European Journal of Medical Research* 29, no. 1 (2024): 113.
- 36. F. Lalloyer and B. Staels, "Fibrates, Glitazones, and Peroxisome Proliferator-Activated Receptors," *Arteriosclerosis, Thrombosis, and Vascular Biology* 30, no. 5 (2010): 894–899.
- 37. S. S. Alkhayyat, H. M. Al-kuraishy, A. I. Al-Gareeb, et al., "Fenofibrate for COVID-19 and Related Complications as an Approach to Improve Treatment Outcomes: The Missed Key for Holy Grail," *Inflammation Research* 71, no. 10 (2022): 1159–1167.
- 38. H. A. Rasheed, N. R. Hussien, M. S. Al-Naimi, H. M. Al-Kuraishy, and A. I. Al-Gareeb, "Fenofibrate and *Crataegus oxyacantha* Is an Effectual Combo for Mixed Dyslipidemia," *Biomedical and Biotechnology Research Journal* 4, no. 3 (2020): 259–265.
- 39. M. L. Morieri, H. S. Shah, J. Sjaarda, et al., "PPARA Polymorphism Influences the Cardiovascular Benefit of Fenofibrate in Type 2 Diabetes: Findings From ACCORD-Lipid," *Diabetes* 69, no. 4 (2020): 771–783.

- 40. C. Tan, L. S. Chew, E. S. Tai, et al., "Benefits of Micronised Fenofibrate in Type 2 Diabetes Mellitus Subjects With Good Glycemic Control," *Atherosclerosis* 154, no. 2 (2001): 469–474.
- 41. L. Czupryniak, S. R. Joshi, J. A. Gogtay, and M. Lopez, "Effect of Micronized Fenofibrate on Microvascular Complications of Type 2 Diabetes: A Systematic Review," *Expert Opinion on Pharmacotherapy* 17, no. 11 (2016): 1463–1473.
- 42. L. J. Holm, M. Haupt-Jorgensen, J. D. Giacobini, J. P. Hasselby, M. Bilgin, and K. Buschard, "Fenofibrate Increases Very-Long-Chain Sphingolipids and Improves Blood Glucose Homeostasis in NOD Mice," *Diabetologia* 62 (2019): 2262–2272.
- 43. R. Krysiak, A. Gdula-Dymek, R. Bachowski, and B. Okopień, "Pleiotropic Effects of Atorvastatin and Fenofibrate in Metabolic Syndrome and Different Types of Pre-Diabetes," *Diabetes Care* 33, no. 10 (2010): 2266–2270.
- 44. A. K. Mottl, J. B. Buse, F. Ismail-Beigi, et al., "Long-Term Effects of Intensive Glycemic and Blood Pressure Control and Fenofibrate Use on Kidney Outcomes," *Clinical Journal of the American Society of Nephrology* 13, no. 11 (2018): 1693–1702.
- 45. R. L. Attridge, C. R. Frei, L. Ryan, J. Koeller, and W. D. Linn, "Fenofibrate-Associated Nephrotoxicity: A Review of Current Evidence," *American Journal of Health-System Pharmacy* 70, no. 14 (2013): 1219–1225.
- 46. J.-C. Ansquer, R. N. Dalton, E. Caussé, D. Crimet, K. le Malicot, and C. Foucher, "Effect of Fenofibrate on Kidney Function: A 6-Week Randomized Crossover Trial in Healthy People," *American Journal of Kidney Diseases* 51, no. 6 (2008): 904–913.
- 47. M. S. Kostapanos, M. Florentin, and M. S. Elisaf, "Fenofibrate and the Kidney: An Overview," *European Journal of Clinical Investigation* 43, no. 5 (2013): 522–531.
- 48. K. Buschard, L. J. Holm, and U. Feldt-Rasmussen, "Insulin Independence in Newly Diagnosed Type 1 Diabetes Patient Following Fenofibrate Treatment," *Case Reports in Medicine* 2020, no. 1 (2020): 6865190.
- 49. E. Boslem, P. J. Meikle, and T. J. Biden, "Roles of Ceramide and Sphingolipids in Pancreatic  $\beta$ -Cell Function and Dysfunction," *Islets* 4, no. 3 (2012): 177–187.
- 50. T. E. Fox and M. Kester Eds., "Therapeutic Strategies for Diabetes and Complications: A Role for Sphingolipids?," in *Sphingolipids as Signaling and Regulatory Molecules* (New York, NY: Springer, 2010), 206–216.
- 51. L. Subramanian, H. Blumenfeld, R. Tohn, et al., "NKT Cells Stimulated by Long Fatty Acyl Chain Sulfatides Significantly Reduces the Incidence of Type 1 Diabetes in Nonobese Diabetic Mice," *PLoS One* 7, no. 5 (2012): e37771.
- 52. D. Zhang, S. Niu, Y. Ma, et al., "Fenofibrate Improves Insulin Resistance and Hepatic Steatosis and Regulates the Let-7/SERCA2b Axis in High-Fat Diet-Induced Non-Alcoholic Fatty Liver Disease Mice," *Frontiers in Pharmacology* 12 (2022): 770652.
- 53. S. Smati, M. Régnier, T. Fougeray, et al., "Regulation of Hepatokine Gene Expression in Response to Fasting and Feeding: Influence of PPAR- $\alpha$  and Insulin-Dependent Signalling in Hepatocytes," *Diabetes & Metabolism* 46, no. 2 (2020): 129–136.
- 54. S. Crunkhorn, "Hepatokine Improves Glucose Control," *Nature Reviews Drug Discovery* 19, no. 11 (2020): 756–757.
- 55. M. K. Montgomery, J. Bayliss, C. Devereux, et al., "SMOC1 Is a Glucose-Responsive Hepatokine and Therapeutic Target for Glycemic Control," *Science Translational Medicine* 12, no. 559 (2020): eaaz8048.
- 56. K. L. Ong, K. A. Rye, R. O'Connell, et al., "Long-Term Fenofibrate Therapy Increases Fibroblast Growth Factor 21 and Retinol-Binding Protein 4 in Subjects With Type 2 Diabetes," *Journal of Clinical Endocrinology & Metabolism* 97, no. 12 (2012): 4701–4708.

- 57. W. Y. So and P. S. Leung, "Fibroblast Growth Factor 21 as an Emerging Therapeutic Target for Type 2 Diabetes Mellitus," *Medicinal Research Reviews* 36, no. 4 (2016): 672–704.
- 58. T. Bobbert, F. Schwarz, A. Fischer-Rosinsky, et al., "Fibroblast Growth Factor 21 Predicts the Metabolic Syndrome and Type 2 Diabetes in Caucasians," *Diabetes Care* 36, no. 1 (2013): 145–149.
- 59. A. O. Chavez, M. Molina-Carrion, M. A. Abdul-Ghani, F. Folli, R. A. DeFronzo, and D. Tripathy, "Circulating Fibroblast Growth Factor-21 Is Elevated in Impaired Glucose Tolerance and Type 2 Diabetes and Correlates With Muscle and Hepatic Insulin Resistance," *Diabetes Care* 32, no. 8 (2009): 1542–1546.
- 60. A. Ferrigno, C. Berardo, L. G. di Pasqua, et al., "The Selective Blockade of Metabotropic Glutamate Receptor-5 Attenuates Fat Accumulation in an In Vitro Model of Benign Steatosis," *European Journal of Histochemistry* 64, no. 4 (2020): 3175.
- 61. H.-T. Huang, C. K. Liao, W. T. Chiu, and S. F. Tzeng, "Ligands of Peroxisome Proliferator-Activated Receptor-Alpha Promote Glutamate Transporter-1 Endocytosis in Astrocytes," *International Journal of Biochemistry & Cell Biology* 86 (2017): 42–53.
- 62. H. Zhang, Y. Gao, P. F. Qiao, F. L. Zhao, and Y. Yan, "Fenofibrate Reduces Amyloidogenic Processing of APP in APP/PS1 Transgenic Mice via PPAR-α/PI3-K Pathway," *International Journal of Developmental Neuroscience* 38 (2014): 223–231.
- 63. S. O. Abdul-Hay, P. Edirisinghe, and G. R. Thatcher, "Selective Modulation of Amyloid-β Peptide Degradation by Flurbiprofen, Fenofibrate, and Related Compounds Regulates Aβ Levels," *Journal of Neurochemistry* 111, no. 3 (2009): 683–695.
- 64. T. Kukar, M. P. Murphy, J. L. Eriksen, et al., "Diverse Compounds Mimic Alzheimer Disease–Causing Mutations by Augmenting A $\beta$ 42 Production," *Nature Medicine* 11, no. 5 (2005): 545–550.
- 65. H. Xu, Z. You, Z. Wu, L. Zhou, J. Shen, and Z. Gu, "WY14643 Attenuates the Scopolamine-Induced Memory Impairments in Mice," *Neurochemical Research* 41 (2016): 2868–2879.
- 66. S. M. de la Monte and J. R. Wands, "Molecular Indices of Oxidative Stress and Mitochondrial Dysfunction Occur Early and Often Progress With Severity of Alzheimer's Disease," *Journal of Alzheimer's Disease* 9, no. 2 (2006): 167–181.
- 67. G. T. Corbett, F. J. Gonzalez, and K. Pahan, "Activation of Peroxisome Proliferator-Activated Receptor  $\alpha$  Stimulates ADAM10-Mediated Proteolysis of APP," *Proceedings of the National Academy of Sciences of the United States of America* 112, no. 27 (2015): 8445–8450.
- 68. S. Chandra, A. Roy, M. Jana, and K. Pahan, "Cinnamic Acid Activates PPAR $\alpha$  to Stimulate Lysosomal Biogenesis and Lower Amyloid Plaque Pathology in an Alzheimer's Disease Mouse Model," Neurobiology of Disease 124 (2019): 379–395.
- 69. H. Oakley, S. L. Cole, S. Logan, et al., "Intraneuronal  $\beta$ -Amyloid Aggregates, Neurodegeneration, and Neuron Loss in Transgenic Mice With Five Familial Alzheimer's Disease Mutations: Potential Factors in Amyloid Plaque Formation," *Journal of Neuroscience* 26, no. 40 (2006): 10129–10140.
- 70. R. Luo, L. Y. Su, G. Li, et al., "Activation of PPARA-Mediated Autophagy Reduces Alzheimer Disease-Like Pathology and Cognitive Decline in a Murine Model," *Autophagy* 16, no. 1 (2020): 52–69.
- 71. J. Krol, I. Loedige, and W. Filipowicz, "The Widespread Regulation of microRNA Biogenesis, Function and Decay," *Nature Reviews Genetics* 11, no. 9 (2010): 597–610.
- 72. X.-Z. Yuan, S. Sun, C. C. Tan, J. T. Yu, and L. Tan, "The Role of ADAM10 in Alzheimer's Disease," *Journal of Alzheimer's Disease* 58, no. 2 (2017): 303–322.
- 73. S. Chandra, M. Jana, and K. Pahan, "Aspirin Induces Lysosomal Biogenesis and Attenuates Amyloid Plaque Pathology in a Mouse Model

- of Alzheimer's Disease via PPARα," *Journal of Neuroscience* 38, no. 30 (2018): 6682–6699.
- 74. A. Roy, M. Jana, M. Kundu, et al., "HMG-CoA Reductase Inhibitors Bind to PPAR $\alpha$  to Upregulate Neurotrophin Expression in the Brain and Improve Memory in Mice," *Cell Metabolism* 22, no. 2 (2015): 253–265.
- 75. A. Maida, B. J. Lamont, X. Cao, and D. J. Drucker, "Metformin Regulates the Incretin Receptor Axis via a Pathway Dependent on Peroxisome Proliferator-Activated Receptor- $\alpha$  in Mice," *Diabetologia* 54 (2011): 339–349.
- 76. L. Wu, K. Wang, W. Wang, et al., "Glucagon-Like Peptide-1 Ameliorates Cardiac Lipotoxicity in Diabetic Cardiomyopathy via the PPAR α Pathway," *Aging Cell* 17, no. 4 (2018): e12763.
- 77. M. Sanati, S. Aminyavari, A. R. Afshari, and A. Sahebkar, "Mechanistic Insight Into the Role of Metformin in Alzheimer's Disease," *Life Sciences* 291 (2022): 120299.
- 78. H. Du, X. Meng, Y. Yao, and J. Xu, "The Mechanism and Efficacy of GLP-1 Receptor Agonists in the Treatment of Alzheimer's Disease," *Frontiers in Endocrinology* 13 (2022): 1033479.
- 79. N. Pierrot, L. Ris, I. C. Stancu, et al., "Sex-Regulated Gene Dosage Effect of PPAR $\alpha$  on Synaptic Plasticity," *Life Science Alliance* 2, no. 2 (2019): e201800262.
- 80. M.-A. Arevalo, I. Azcoitia, I. Gonzalez-Burgos, and L. M. Garcia-Segura, "Signaling Mechanisms Mediating the Regulation of Synaptic Plasticity and Memory by Estradiol," *Hormones and Behavior* 74 (2015): 19–27.
- 81. B. C. Riedel, P. M. Thompson, and R. D. Brinton, "Age, APOE and Sex: Triad of Risk of Alzheimer's Disease," *Journal of Steroid Biochemistry and Molecular Biology* 160 (2016): 134–147.
- 82. D. L. Korol, E. L. Malin, K. A. Borden, R. A. Busby, and J. Couper-Leo, "Shifts in Preferred Learning Strategy Across the Estrous Cycle in Female Rats," *Hormones and Behavior* 45, no. 5 (2004): 330–338.
- 83. M. A. Zhang, D. Rego, M. Moshkova, et al., "Peroxisome Proliferator-Activated Receptor (PPAR)  $\alpha$  and  $\gamma$  Regulate IFN $\gamma$  and IL-17A Production by Human T Cells in a Sex-Specific Way," *Proceedings of the National Academy of Sciences of the United States of America* 109, no. 24 (2012): 9505–9510.
- 84. H. Joffe, J. E. Hall, S. Gruber, et al., "Estrogen Therapy Selectively Enhances Prefrontal Cognitive Processes: A Randomized, Double-Blind, Placebo-Controlled Study With Functional Magnetic Resonance Imaging in Perimenopausal and Recently Postmenopausal Women," *Menopause* 13, no. 3 (2006): 411–422.
- 85. N. Pierrot, R. Lhommel, L. Quenon, et al., "Targretin Improves Cognitive and Biological Markers in a Patient With Alzheimer's Disease," *Journal of Alzheimer's Disease* 49, no. 2 (2015): 271–276.
- 86. A. Herrstedt, M. L. Bay, C. Simonsen, et al., "Exercise-Mediated Improvement of Depression in Patients With Gastro-Esophageal Junction Cancer Is Linked to Kynurenine Metabolism," *Acta Oncologica* 58, no. 5 (2019): 579–587.
- 87. Z. Al-Qahtani, H. M. Al-Kuraishy, N. H. Ali, Y. H. A. Elewa, and G. E. Batiha, "Kynurenine Pathway in Type 2 Diabetes: Role of Metformin," *Drug Development Research* 85, no. 5 (2024): e22243.
- 88. I. Zakrocka, W. Turski, and T. Kocki, "Clonidine Decreases Kynurenic Acid Production in Rat Brain Cortex In Vitro—A Novel Antihypertensive Mechanism of Action?," *Journal of Pre-Clinical and Clinical Research* 10, no. 1 (2016): 57–59.
- 89. G. S. Deora, S. Kantham, S. Chan, et al., "Multifunctional Analogs of Kynurenic Acid for the Treatment of Alzheimer's Disease: Synthesis, Pharmacology, and Molecular Modeling Studies," *ACS Chemical Neuroscience* 8, no. 12 (2017): 2667–2675.
- 90. G. Donvito, D. Bagdas, W. Toma, et al., "The Interaction Between Alpha 7 Nicotinic Acetylcholine Receptor and Nuclear Peroxisome

- Proliferator-Activated Receptor- $\alpha$  Represents a New Antinociceptive Signaling Pathway in Mice," *Experimental Neurology* 295 (2017): 194–201.
- 91. K.-G. Ma and Y.-H. Qian, "Alpha 7 Nicotinic Acetylcholine Receptor and Its Effects on Alzheimer's Disease," *Neuropeptides* 73 (2019): 96–106.
- 92. H. R. Parri, C. M. Hernandez, and K. T. Dineley, "Research Update: Alpha7 Nicotinic Acetylcholine Receptor Mechanisms in Alzheimer's Disease," *Biochemical Pharmacology* 82, no. 8 (2011): 931–942.
- 93. S. Sadigh-Eteghad, J. Mahmoudi, S. Babri, and M. Talebi, "Effect of Alpha-7 Nicotinic Acetylcholine Receptor Activation on Beta-Amyloid Induced Recognition Memory Impairment. Possible Role of Neurovascular Function," *Acta Cirurgica Brasileira* 30, no. 11 (2015): 736–742.
- 94. M. Jana, A. Jana, X. Liu, S. Ghosh, and K. Pahan, "Involvement of Phosphatidylinositol 3-Kinase-Mediated Up-Regulation of IκBα in Anti-Inflammatory Effect of Gemfibrozil in Microglia," *Journal of Immunology* 179, no. 6 (2007): 4142–4152.
- 95. T. Kihara, S. Shimohama, H. Sawada, et al., " $\alpha$ 7 Nicotinic Receptor Transduces Signals to Phosphatidylinositol 3-Kinase to Block A  $\beta$ -Amyloid-Induced Neurotoxicity," *Journal of Biological Chemistry* 276, no. 17 (2001): 13541–13546.
- 96. S. Chandra and K. Pahan, "Gemfibrozil, A Lipid-Lowering Drug, Attenuates Amyloid Plaque Pathology and Enhances Memory via PPARα," *Alzheimer's & Dementia* 14, no. 7S\_Part\_31 (2018): P1655–P1656.
- 97. N. Viswakarma, Y. Jia, L. Bai, et al., "Coactivators in PPAR-Regulated Gene Expression," *PPAR Research* 2010, no. 1 (2010): 250126.
- 98. D. Uppalapati, N. R. das, R. P. Gangwal, M. V. Damre, A. T. Sangamwar, and S. S. Sharma, "Neuroprotective Potential of Peroxisome Proliferator Activated Receptor- $\alpha$  Agonist in Cognitive Impairment in Parkinson's Disease: Behavioral, Biochemical, and PBPK Profile," *PPAR Research* 2014, no. 1 (2014): 753587.
- 99. K. E. Nisbett and G. Pinna, "Emerging Therapeutic Role of PPAR $-\alpha$  in Cognition and Emotions," *Frontiers in Pharmacology* 9 (2018): 998.
- 100. P. Behzadi, H. A. García-Perdomo, and T. M. Karpiński, "Toll-Like Receptors: General Molecular and Structural Biology," *Journal of Immunology Research* 2021, no. 1 (2021): 9914854.
- 101. D. T. Skelly, E. Hennessy, M. A. Dansereau, and C. Cunningham, "Correction: A Systematic Analysis of the Peripheral and CNS Effects of Systemic LPS, IL-1 $\beta$ , TNF- $\alpha$  and IL-6 Challenges in C57BL/6 Mice," *PLoS One* 8, no. 12 (2013): e69123.
- 102. H. M. Al-Kuraishy, O. M. Al-Fakhrany, E. Elekhnawy, et al., "Traditional Herbs Against COVID-19: Back to Old Weapons to Combat the New Pandemic," *European Journal of Medical Research* 27, no. 1 (2022): 186.
- 103. M. Moubarak, K. I. Kasozi, H. F. Hetta, et al., "The Rise of SARS-CoV-2 Variants and the Role of Convalescent Plasma Therapy for Management of Infections," *Life* 11, no. 8 (2021): 734.
- 104. D. Krstic, A. Madhusudan, J. Doehner, et al., "Systemic Immune Challenges Trigger and Drive Alzheimer-Like Neuropathology in Mice," *Journal of Neuroinflammation* 9 (2012): 1–24.
- 105. W. Swardfager, K. Lanctôt, L. Rothenburg, A. Wong, J. Cappell, and N. Herrmann, "A Meta-Analysis of Cytokines in Alzheimer's Disease," *Biological Psychiatry* 68, no. 10 (2010): 930–941.
- 106. P. Bermejo, S. Martín-Aragón, J. Benedí, et al., "Differences of Peripheral Inflammatory Markers Between Mild Cognitive Impairment and Alzheimer's Disease," *Immunology Letters* 117, no. 2 (2008): 198–202.
- 107. H. M. Al-Kuraishy, A. I. Al-Gareeb, G. Mostafa-Hedeab, et al., "Effects of  $\beta$ -Blockers on the Sympathetic and Cytokines Storms in Covid-19," *Frontiers in Immunology* 12 (2021): 749291.
- 108. X. Guo, W. Dang, N. Li, et al., "PPAR-α Agonist Fenofibrate Ameliorates Sjögren Syndrome-Like Dacryoadenitis by Modulating

- Th1/Th17 and Treg Cell Responses in NOD Mice," *Investigative Ophthalmology & Visual Science* 63, no. 6 (2022): 12.
- 109. Y. Yang, A. R. Gocke, A. Lovett-Racke, P. D. Drew, and M. K. Racke, "PPAR Alpha Regulation of the Immune Response and Autoimmune Encephalomyelitis," *PPAR Research* 2008, no. 1 (2008): 546753.
- 110. H. M. Al-Kuraishy, A. I. Al-Gareeb, K. J. Alzahrani, N. Cruz-Martins, and G. E. Batiha, "The Potential Role of Neopterin in Covid-19: A New Perspective," *Molecular and Cellular Biochemistry* 476 (2021): 4161–4166.
- 111. G. E.-S. Batiha, A. Gari, N. Elshony, et al., "Hypertension and Its Management in COVID-19 Patients: The Assorted View," *International Journal of Cardiology Cardiovascular Risk and Prevention* 11 (2021): 200121.
- 112. H. M. Al-Kuraishy, A. I. Al-Gareeb, S. Qusti, E. M. Alshammari, F. O. Atanu, and G. E. Batiha, "Arginine Vasopressin and Pathophysiology of COVID-19: An Innovative Perspective," *Biomedicine & Pharmacotherapy* 143 (2021): 112193.
- 113. H. M. Al-Kuraishy, N. R. Hussien, M. S. Al-Naimi, A. K. Al-Buhadily, A. I. Al-Gareeb, and C. Lungnier, "Is Ivermectin-Azithromycin Combination the Next Step for COVID-19?," *Biomedical and Biotechnology Research Journal* 4, no. Suppl 1 (2020): S101–S103.
- 114. J. Yu, E. Ip, A. dela Peña, et al., "COX-2 Induction in Mice With Experimental Nutritional Steatohepatitis: Role as Pro-Inflammatory Mediator," *Hepatology* 43, no. 4 (2006): 826–836.
- 115. D. Huang, Q. Zhao, H. Liu, Y. Guo, and H. Xu, "PPAR- $\alpha$  Agonist WY-14643 Inhibits LPS-Induced Inflammation in Synovial Fibroblasts via NF-kB Pathway," *Journal of Molecular Neuroscience* 59 (2016): 544–553.
- 116. G. M. Gallucci, B. Alsuwayt, A. M. Auclair, J. L. Boyer, D. N. Assis, and N. S. Ghonem, "Fenofibrate Downregulates NF- $\kappa$ B Signaling to Inhibit Pro-Inflammatory Cytokine Secretion in Human THP-1 Macrophages and During Primary Biliary Cholangitis," *Inflammation* 45, no. 6 (2022): 2570–2581.
- 117. S. Bahrambeigi, M. Molaparast, F. Sohrabi, et al., "Targeting PPAR Ligands as Possible Approaches for Metabolic Reprogramming of T Cells in Cancer Immunotherapy," *Immunology Letters* 220 (2020): 32–37.
- 118. H. M. Al-Kuraishy, A. I. Al-Gareeb, S. M. Abdullah, N. Cruz-Martins, and G. E. Batiha, "Case Report: Hyperbilirubinemia in Gilbert Syndrome Attenuates Covid-19-Induced Metabolic Disturbances," *Frontiers in Cardiovascular Medicine* 8 (2021): 642181.
- 119. H. M. Al-Kuraishy and A. I. Al-Gareeb, "Effect of Orlistat Alone or in Combination With Garcinia Cambogia on Visceral Adiposity Index in Obese Patients," *Journal of Intercultural Ethnopharmacology* 5, no. 4 (2016): 408–414.
- 120. H. M. Al-Kuraishy, A. I. Al-Gareeb, M. S. Al-Niemi, et al., "The Prospective Effect of Allopurinol on the Oxidative Stress Index and Endothelial Dysfunction in Covid-19," *Inflammation* 45, no. 4 (2022): 1651–1667.
- 121. D. Hasan Khudhair, A. I. Al-Gareeb, H. M. Al-Kuraishy, et al., "Combination of Vitamin C and Curcumin Safeguards Against Methotrexate-Induced Acute Liver Injury in Mice by Synergistic Antioxidant Effects," *Frontiers in Medicine* 9 (2022): 866343.
- 122. S. Aslibekyan, E. K. Kabagambe, M. R. Irvin, et al., "A Genome-Wide Association Study of Inflammatory Biomarker Changes in Response to Fenofibrate Treatment in the Genetics of Lipid Lowering Drug and Diet Network," *Pharmacogenetics and Genomics* 22, no. 3 (2012): 191–197.
- 123. Z. Bagi, N. Erdei, Z. Papp, I. Edes, and A. Koller, "Up-Regulation of Vascular Cyclooxygenase-2 in Diabetes Mellitus," *Pharmacological Reports* 58 (2006): 52.
- 124. R. Pop-Busui, A. P. Kellogg, and H. T. Cheng, "Cyclooxygenase-2 Pathway as a Potential Therapeutic Target in Diabetic Peripheral Neuropathy," *Current Drug Targets* 9, no. 1 (2008): 68–76.

- 125. K. F. Rodrigues, N. T. Pietrani, A. A. Bosco, F. M. F. Campos, V. C. Sandrim, and K. B. Gomes, "IL-6, TNF- $\alpha$ , and IL-10 Levels/ Polymorphisms and Their Association With Type 2 Diabetes Mellitus and Obesity in Brazilian Individuals," *Archives of Endocrinology and Metabolism* 61 (2017): 438–446.
- 126. S. Patel and D. Santani, "Role of NF-κB in the Pathogenesis of Diabetes and Its Associated Complications," *Pharmacological Reports* 61, no. 4 (2009): 595–603.
- 127. M. Rachmiel, O. Bloch, T. Bistritzer, et al., "TH1/TH2 Cytokine Balance in Patients With Both Type 1 Diabetes Mellitus and Asthma," *Cytokine* 34, no. 3–4 (2006): 170–176.
- 128. T. Xu, J. Liu, X. R. Li, et al., "The mTOR/NF-кВ Pathway Mediates Neuroinflammation and Synaptic Plasticity in Diabetic Encephalopathy," *Molecular Neurobiology* 58 (2021): 3848–3862.
- 129. I. Granic, A. M. Dolga, I. M. Nijholt, G. van Dijk, and U. L. M. Eisel, "Inflammation and NF-κB in Alzheimer's Disease and Diabetes," *Journal of Alzheimer's Disease* 16, no. 4 (2009): 809–821.
- 130. C. H. Trepanier and N. W. Milgram, "Neuroinflammation in Alzheimer's Disease: Are NSAIDs and Selective COX-2 Inhibitors the Next Line of Therapy?," *Journal of Alzheimer's Disease* 21, no. 4 (2010): 1089–1099.
- 131. I. M. Cojocaru, M. Cojocaru, G. Miu, and V. Sapira, "Study of Interleukin-6 Production in Alzheimer's Disease," *Romanian Journal of Internal Medicine* 49, no. 1 (2011): 55–58.
- 132. J. M. Rubio-Perez and J. M. Morillas-Ruiz, "A Review: Inflammatory Process in Alzheimer's Disease, Role of Cytokines," *Scientific World Journal* 2012, no. 1 (2012): 756357.
- 133. M.-S. Tan, J. T. Yu, T. Jiang, X. C. Zhu, and L. Tan, "The NLRP3 Inflammasome in Alzheimer's Disease," *Molecular Neurobiology* 48 (2013): 875–882.
- 134. I. M. Gora, A. Ciechanowska, and P. Ladyzynski, "NLRP3 Inflammasome at the Interface of Inflammation, Endothelial Dysfunction, and Type 2 Diabetes," *Cells* 10, no. 2 (2021): 314.
- 135. H. M. Al-Kuraishy, A. I. Al-Gareeb, M. S. Al-Niemi, A. K. Al-Buhadily, N. A. Al-Harchan, and C. Lugnier, "COVID-19 and Phosphodiesterase Enzyme Type 5 Inhibitors," *Journal of Microscopy and Ultrastructure* 8, no. 4 (2020): 141–145.
- 136. H. M. Al-Kuraishy, T. J. L-Maiahy, A. I. Al-Gareeb, R. A. Musa, and Z. H. Ali, "COVID-19 Pneumonia in an Iraqi Pregnant Woman With Preterm Delivery," *Asian Pacific Journal of Reproduction* 9, no. 3 (2020): 156–158.
- 137. H. M. Alkuraishy, A. I. Al-Gareeb, and H. J. Waheed, "Lipoprotein-Associated Phospholipase A2 Is Linked With Poor Cardio-Metabolic Profile in Patients With Ischemic Stroke: A Study of Effects of Statins," *Journal of Neurosciences in Rural Practice* 9, no. 4 (2018): 496–503.
- 138. M. Alorabi, S. Cavalu, H. M. Al-kuraishy, et al., "Pentoxifylline and Berberine Mitigate Diclofenac-Induced Acute Nephrotoxicity in Male Rats via Modulation of Inflammation and Oxidative Stress," *Biomedicine & Pharmacotherapy* 152 (2022): 113225.
- 139. H. M. Al-Kuraishy, A. I. Al-Gareeb, N. Qusty, A. Alexiou, and G. E. Batiha, "Impact of Sitagliptin on Non-Diabetic Covid-19 Patients," *Current Molecular Pharmacology* 15, no. 4 (2022): 683–692.
- 140. C. Delayre-Orthez, J. Becker, I. Guenon, et al., "PPAR $\alpha$  Downregulates Airway Inflammation Induced by Lipopolysaccharide in the Mouse," *Respiratory Research* 6 (2005): 1–10.
- 141. A. Mandala, A. Armstrong, B. Girresch, et al., "Fenofibrate Prevents Iron Induced Activation of Canonical Wnt/ $\beta$ -Catenin and Oxidative Stress Signaling in the Retina," *NPJ Aging and Mechanisms of Disease* 6, no. 1 (2020): 12.

- 142. M. Calvo-Rodriguez, C. García-Rodríguez, C. Villalobos, and L. Núñez, "Role of Toll Like Receptor 4 in Alzheimer's Disease," *Frontiers in Immunology* 11 (2020): 1588.
- 143. N. Dana, S. Haghjooy Javanmard, and G. Vaseghi, "The Effect of Fenofibrate, a PPAR $\alpha$  Activator on Toll-Like Receptor-4 Signal Transduction in Melanoma Both In Vitro and In Vivo," *Clinical and Translational Oncology* 22 (2020): 486–494.
- 144. M. Wang, Y. Yan, Z. Zhang, et al., "Programmed PPAR- $\alpha$  Downregulation Induces Inflammaging by Suppressing Fatty Acid Catabolism in Monocytes," *Iscience* 24, no. 7 (2021): 102766.
- 145. T. Behl, E. Sharma, A. Sehgal, et al., "Expatiating the Molecular Approaches of HMGB1 in Diabetes Mellitus: Highlighting Signalling Pathways via RAGE and TLRs," *Molecular Biology Reports* 48 (2021): 1869–1881.
- 146. Z. Sepehri, Z. Kiani, A. A. Nasiri, and F. Kohan, "Toll-Like Receptor 2 and Type 2 Diabetes," *Cellular & Molecular Biology Letters* 21 (2016): 1–9.
- 147. M. Lyman, D. G. Lloyd, X. Ji, M. P. Vizcaychipi, and D. Ma, "Neuroinflammation: The Role and Consequences," *Neuroscience Research* 79 (2014): 1–12.
- 148. J. Stephenson, E. Nutma, P. van der Valk, and S. Amor, "Inflammation in CNS Neurodegenerative Diseases," *Immunology* 154, no. 2 (2018): 204–219.
- 149. I. Bjelobaba, D. Savic, and I. Lavrnja, "Multiple Sclerosis and Neuroinflammation: The Overview of Current and Prospective Therapies," *Current Pharmaceutical Design* 23, no. 5 (2017): 693–730.
- 150. L. Yang and J. Nao, "Focus on Alzheimer's Disease: The Role of Fibroblast Growth Factor 21 and Autophagy," *Neuroscience* 511 (2023): 13–28
- 151. E. D. AlFadly, P. A. Elzahhar, A. Tramarin, et al., "Tackling Neuroinflammation and Cholinergic Deficit in Alzheimer's Disease: Multi-Target Inhibitors of Cholinesterases, Cyclooxygenase-2 and 15-Lipoxygenase," *European Journal of Medicinal Chemistry* 167 (2019): 161–186.
- 152. M. Cox, C. Bassi, M. E. Saunders, et al., "Beyond Neurotransmission: Acetylcholine in Immunity and Inflammation," *Journal of Internal Medicine* 287, no. 2 (2020): 120–133.
- 153. S. Asslih, O. Damri, and G. Agam, "Neuroinflammation as a Common Denominator of Complex Diseases (Cancer, Diabetes Type 2, and Neuropsychiatric Disorders)," *International Journal of Molecular Sciences* 22, no. 11 (2021): 6138.
- 154. X. Chen, A. C. Famurewa, J. Tang, O. O. Olatunde, and O. J. Olatunji, "Hyperoside Attenuates Neuroinflammation, Cognitive Impairment and Oxidative Stress via Suppressing TNF- $\alpha$ /NF- $\alpha$ B/Caspase-3 Signaling in Type 2 Diabetes Rats," *Nutritional Neuroscience* 25, no. 8 (2022): 1774–1784.
- 155. S. Rom, V. Zuluaga-Ramirez, S. Gajghate, et al., "Hyperglycemia-Driven Neuroinflammation Compromises BBB Leading to Memory Loss in Both Diabetes Mellitus (DM) Type 1 and Type 2 Mouse Models," *Molecular Neurobiology* 56 (2019): 1883–1896.
- 156. X. R. Chen, V. C. Besson, B. Palmier, Y. Garcia, M. Plotkine, and C. Marchand-Leroux, "Neurological Recovery-Promoting, Anti-Inflammatory, and Anti-Oxidative Effects Afforded by Fenofibrate, a PPAR Alpha Agonist, in Traumatic Brain Injury," *Journal of Neurotrauma* 24, no. 7 (2007): 1119–1131.
- 157. M. A. Esmaeili, S. Yadav, R. K. Gupta, et al., "Preferential PPAR- $\alpha$  Activation Reduces Neuroinflammation, and Blocks Neurodegeneration In Vivo," *Human Molecular Genetics* 25, no. 2 (2016): 317–327.
- 158. M. Melis, S. Scheggi, G. Carta, et al., "PPAR $\alpha$  Regulates Cholinergic-Driven Activity of Midbrain Dopamine Neurons via a

- Novel Mechanism Involving α7 Nicotinic Acetylcholine Receptors," *Journal of Neuroscience* 33, no. 14 (2013): 6203–6211.
- 159. H. M. Al-Kuraishy, A. I. Al-Gareeb, and T. J. Al-Maiahy, "Concept and Connotation of Oxidative Stress in Preeclampsia," *Journal of Laboratory Physicians* 10, no. 3 (2018): 276–282.
- 160. H. M. Al-Kuraishy, A. I. Al-Gareeb, and M. S. Al-Nami, "Vinpocetine Improves Oxidative Stress and Pro-Inflammatory Mediators in Acute Kidney Injury," *International Journal of Preventive Medicine* 10, no. 1 (2019): 142.
- 161. Y. Cheng, Y. Song, H. Chen, et al., "Ferroptosis Mediated by Lipid Reactive Oxygen Species: A Possible Causal Link of Neuroinflammation to Neurological Disorders," *Oxidative Medicine and Cellular Longevity* 2021, no. 1 (2021): 5005136.
- 162. D. A. Butterfield and B. Halliwell, "Oxidative Stress, Dysfunctional Glucose Metabolism and Alzheimer Disease," *Nature Reviews Neuroscience* 20, no. 3 (2019): 148–160.
- 163. G. Veurink, G. Perry, and S. K. Singh, "Role of Antioxidants and a Nutrient Rich Diet in Alzheimer's Disease," *Open Biology* 10, no. 6 (2020): 200084.
- 164. D. Kaur, T. Behl, A. Sehgal, et al., "Decrypting the Potential Role of  $\alpha$ -Lipoic Acid in Alzheimer's Disease," *Life Sciences* 284 (2021): 119899.
- 165. E. Burgos-Morón, Z. Abad-Jiménez, A. Martínez de Marañón, et al., "Relationship Between Oxidative Stress, ER Stress, and Inflammation in Type 2 Diabetes: The Battle Continues," *Journal of Clinical Medicine* 8, no. 9 (2019): 1385.
- 166. X. Huang, M. Sun, D. Li, et al., "Augmented NADPH Oxidase Activity and p22phox Expression in Monocytes Underlie Oxidative Stress of Patients With Type 2 Diabetes Mellitus," *Diabetes Research and Clinical Practice* 91, no. 3 (2011): 371–380.
- 167. X.-X. Qu, J. H. He, Z. Q. Cui, T. Yang, and X. H. Sun, "PPAR- $\alpha$  Agonist GW7647 Protects Against Oxidative Stress and Iron Deposit via GPx4 in a Transgenic Mouse Model of Alzheimer's Diseases," *ACS Chemical Neuroscience* 13, no. 2 (2021): 207–216.
- 168. Q. Liu, X. Zhang, R. Cheng, J. X. Ma, J. Yi, and J. Li, "Salutary Effect of Fenofibrate on Type 1 Diabetic Retinopathy via Inhibiting Oxidative Stress–Mediated Wnt/ $\beta$ -Catenin Pathway Activation," *Cell and Tissue Research* 376 (2019): 165–177.
- 169. A. A. Oyagbemi, O. E. Adebiyi, K. O. Adigun, et al., "Clofibrate, a PPAR- $\alpha$  Agonist, Abrogates Sodium Fluoride-Induced Neuroinflammation, Oxidative Stress, and Motor Incoordination via Modulation of GFAP/Iba-1/Anti-Calbindin Signaling Pathways," *Environmental Toxicology* 35, no. 2 (2020): 242–253.
- 170. D. M. Muoio, J. M. Way, C. J. Tanner, et al., "Peroxisome Proliferator-Activated Receptor- $\alpha$  Regulates Fatty Acid Utilization in Primary Human Skeletal Muscle Cells," *Diabetes* 51, no. 4 (2002): 901–909.
- 171. W. Wang, F. Zhao, X. Ma, G. Perry, and X. Zhu, "Mitochondria Dysfunction in the Pathogenesis of Alzheimer's Disease: Recent Advances," *Molecular Neurodegeneration* 15 (2020): 1–22.
- 172. G. E.-S. Batiha, H. M. Al-Kuraishy, A. I. Al-Gareeb, and E. Elekhnawy, "SIRT1 Pathway in Parkinson's Disease: A Faraway Snapshot but So Close," *Inflammopharmacology* 31, no. 1 (2023): 37–56.
- 173. Z. K. Alkaç, F. Ahmet Korkak, G. Dağoğlu, C. Akdeniz İncili, B. Dağoğlu Hark, and S. Tanyıldızı, "Puerarin Mitigates Oxidative Injuries, Opening of Mitochondrial Permeability Transition Pores and Pathological Damage Associated With Liver and Kidney in *Xanthium strumarium*-Intoxicated Rats," *Toxicon* 213 (2022): 13–22.
- 174. A. B. Reiss, S. Ahmed, C. Dayaramani, et al., "The Role of Mitochondrial Dysfunction in Alzheimer's Disease: A Potential Pathway to Treatment," *Experimental Gerontology* 164 (2022): 111828.
- 175. M. Ghareghani, S. Dokoohaki, A. Ghanbari, et al., "Melatonin Exacerbates Acute Experimental Autoimmune Encephalomyelitis

- by Enhancing the Serum Levels of Lactate: A Potential Biomarker of Multiple Sclerosis Progression," *Clinical and Experimental Pharmacology and Physiology* 44, no. 1 (2017): 52–61.
- 176. M. V. Pinti, G. K. Fink, Q. A. Hathaway, A. J. Durr, A. Kunovac, and J. M. Hollander, "Mitochondrial Dysfunction in Type 2 Diabetes Mellitus: An Organ-Based Analysis," *American Journal of Physiology. Endocrinology and Metabolism* 316 (2019): E268–E285.
- 177. K. Roszczyc-Owsiejczuk and P. Zabielski, "Sphingolipids as a Culprit of Mitochondrial Dysfunction in Insulin Resistance and Type 2 Diabetes," *Frontiers in Endocrinology* 12 (2021): 635175.
- 178. S. Khan, G. V. Raghuram, A. Bhargava, et al., "Role and Clinical Significance of Lymphocyte Mitochondrial Dysfunction in Type 2 Diabetes Mellitus," *Translational Research* 158, no. 6 (2011): 344–359.
- 179. I. E. Elijah, E. Børsheim, D. M. Maybauer, C. C. Finnerty, D. N. Herndon, and M. O. Maybauer, "Role of the PPAR-α Agonist Fenofibrate in Severe Pediatric Burn," *Burns* 38, no. 4 (2012): 481–486.
- 180. S. Jeong and M. Yoon, "Fenofibrate Inhibits Adipocyte Hypertrophy and Insulin Resistance by Activating Adipose PPAR $\alpha$  in High Fat Diet-Induced Obese Mice," *Experimental & Molecular Medicine* 41, no. 6 (2009): 397–405.
- 181. S. Ramanan, M. Kooshki, W. Zhao, F. C. Hsu, D. R. Riddle, and M. E. Robbins, "The PPARα Agonist Fenofibrate Preserves Hippocampal Neurogenesis and Inhibits Microglial Activation After Whole-Brain Irradiation," *International Journal of Radiation Oncology, Biology, Physics* 75, no. 3 (2009): 870–877.
- 182. N. H. Ali, H. M. Al-Kuraishy, A. I. Al-Gareeb, et al., "Autophagy and Autophagy Signaling in Epilepsy: Possible Role of Autophagy Activator," *Molecular Medicine* 29, no. 1 (2023): 142.
- 183. H. M. Al-Kuraishy, G. M. Sulaiman, M. S. Jabir, et al., "Defective Autophagy and Autophagy Activators in Myasthenia Gravis: A Rare Entity and Unusual Scenario," *Autophagy* 20 (2024): 1473–1482.
- 184. H. M. Al-Kuraishy, M. S. Jabir, A. I. Al-Gareeb, D. J. Klionsky, and A. K. Albuhadily, "Dysregulation of Pancreatic  $\beta$ -Cell Autophagy and the Risk of Type 2 Diabetes," *Autophagy* 20, no. 11 (2024): 2361–2372.
- 185. M.S.Jabir, M.K.A.Mohammed, S.Albukhaty, et al., "Functionalized SWCNTs@ ag-TiO2 Nanocomposites Induce ROS-Mediated Apoptosis and Autophagy in Liver Cancer Cells," *Nanotechnology Reviews* 12, no. 1 (2023): 20230127.
- 186. Y. Aman, T. Schmauck-Medina, M. Hansen, et al., "Autophagy in Healthy Aging and Disease," *Nature Aging* 1, no. 8 (2021): 634–650.
- 187. P. H. Reddy and D. M. Oliver, "Amyloid Beta and Phosphorylated Tau-Induced Defective Autophagy and Mitophagy in Alzheimer's Disease," *Cells* 8, no. 5 (2019): 488.
- 188. J. M. Carosi and T. J. Sargeant, "Rapamycin and Alzheimer Disease: A Hypothesis for the Effective Use of Rapamycin for Treatment of Neurodegenerative Disease," *Autophagy* 19, no. 8 (2023): 2386–2390.
- 189. Z. Long, J. Chen, Y. Zhao, et al., "Dynamic Changes of Autophagic Flux Induced by Abeta in the Brain of Postmortem Alzheimer's Disease Patients, Animal Models and Cell Models," *Aging* 12, no. 11 (2020): 10912–10930.
- 190. J. Kosacka, M. Kern, N. Klöting, et al., "Autophagy in Adipose Tissue of Patients With Obesity and Type 2 Diabetes," *Molecular and Cellular Endocrinology* 409 (2015): 21–32.
- 191. H. Watada and Y. Fujitani, "Minireview: Autophagy in Pancreatic  $\beta\text{-Cells}$  and Its Implication in Diabetes," *Molecular Endocrinology* 29, no. 3 (2015): 338–348.
- 192. D. Bhattacharya, M. Mukhopadhyay, M. Bhattacharyya, and P. Karmakar, "Is Autophagy Associated With Diabetes Mellitus and Its Complications? A Review," *EXCLI Journal* 17 (2018): 709–720.
- 193. A. Sehrawat, J. Mishra, S. S. Mastana, et al., "Dysregulated Autophagy: A Key Player in the Pathophysiology of Type 2 Diabetes

- and Its Complications," *Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease* 1869, no. 4 (2023): 166666.
- 194. J. Zhang, Y. Cheng, J. Gu, et al., "Fenofibrate Increases Cardiac Autophagy via FGF21/SIRT1 and Prevents Fibrosis and Inflammation in the Hearts of Type 1 Diabetic Mice," *Clinical Science* 130, no. 8 (2016): 625–641.
- 195. M. Sohn, K. Kim, M. J. Uddin, et al., "Delayed Treatment With Fenofibrate Protects Against High-Fat Diet-Induced Kidney Injury in Mice: The Possible Role of AMPK Autophagy," *American Journal of Physiology. Renal Physiology* 312, no. 2 (2017): F323–F334.
- 196. Y. Liu, Y. Xu, J. Zhu, et al., "Metformin Prevents Progression of Experimental Pulmonary Hypertension via Inhibition of Autophagy and Activation of Adenosine Monophosphate-Activated Protein Kinase," *Journal of Vascular Research* 56, no. 3 (2019): 117–128.
- 197. Y. Wu, X. Li, J. X. Zhu, et al., "Resveratrol-Activated AMPK/SIRT1/ Autophagy in Cellular Models of Parkinson's Disease," *Neurosignals* 19, no. 3 (2011): 163–174.
- 198. L. Gao, Y. Zhang, K. Sterling, and W. Song, "Brain-Derived Neurotrophic Factor in Alzheimer's Disease and Its Pharmaceutical Potential," *Translational Neurodegeneration* 11, no. 1 (2022): 4.
- 199. N. H. Ali, H. M. Al-Kuraishy, A. I. Al-Gareeb, et al., "BDNF/TrkB Activators in Parkinson's Disease: A New Therapeutic Strategy," *Journal of Cellular and Molecular Medicine* 28, no. 10 (2024): e18368.
- 200. N. H. Ali, H. M. Al-Kuraishy, A. I. Al-Gareeb, S. A. Alnaaim, H. M. Saad, and G. E. S. Batiha, "The Molecular Pathway of p75 Neurotrophin Receptor (p75NTR) in Parkinson's Disease: The Way of New Inroads," *Molecular Neurobiology* 61, no. 5 (2024): 2469–2480.
- 201. N. H. Ali, H. M. Al-Kuraishy, A. I. Al-Gareeb, et al., "Role of Brain Renin–Angiotensin System in Depression: A New Perspective," *CNS Neuroscience & Therapeutics* 30, no. 4 (2024): e14525.
- 202. A. Turkistani, H. M. Al-Kuraishy, A. I. Al-Gareeb, et al., "The Functional and Molecular Roles of p75 Neurotrophin Receptor (p75NTR) in Epilepsy," *Journal of Central Nervous System Disease* 16 (2024): 11795735241247810.
- 203. H. M. Al-Kuraishy, A. I. Al-Gareeb, Y. H. A. Elewa, et al., "Parkinson's Disease Risk and Hyperhomocysteinemia: The Possible Link," *Cellular and Molecular Neurobiology* 43, no. 6 (2023): 2743–2759.
- 204. C.-C. Lin and T.-L. Huang, "Brain-Derived Neurotrophic Factor and Mental Disorders," *Biomedical Journal* 43, no. 2 (2020): 134–142.
- 205. S. Peng, J. Wuu, E. J. Mufson, and M. Fahnestock, "Precursor Form of Brain-Derived Neurotrophic Factor and Mature Brain-Derived Neurotrophic Factor Are Decreased in the Pre-Clinical Stages of Alzheimer's Disease," *Journal of Neurochemistry* 93, no. 6 (2005): 1412–1421.
- 206. T. K. S. Ng, C. S. H. Ho, W. W. S. Tam, E. H. Kua, and R. C. M. Ho, "Decreased Serum Brain-Derived Neurotrophic Factor (BDNF) Levels in Patients With Alzheimer's Disease (AD): A Systematic Review and Meta-Analysis," *International Journal of Molecular Sciences* 20, no. 2 (2019): 257.
- 207. O.V. Forlenza, B.S. Diniz, A. L. Teixeira, et al., "Lower Cerebrospinal Fluid Concentration of Brain-Derived Neurotrophic Factor Predicts Progression From Mild Cognitive Impairment to Alzheimer's Disease," *Neuromolecular Medicine* 17 (2015): 326–332.
- 208. G. Weinstein, A. S. Beiser, S. H. Choi, et al., "Serum Brain-Derived Neurotrophic Factor and the Risk for Dementia: The Framingham Heart Study," *JAMA Neurology* 71, no. 1 (2014): 55–61.
- 209. K. Krabbe, A. R. Nielsen, R. Krogh-Madsen, et al., "Brain-Derived Neurotrophic Factor (BDNF) and Type 2 Diabetes," *Diabetologia* 50 (2007): 431–438.
- 210. C. Eyileten, A. Kaplon-Cieslicka, D. Mirowska-Guzel, L. Malek, and M. Postula, "Antidiabetic Effect of Brain-Derived Neurotrophic

- Factor and Its Association With Inflammation in Type 2 Diabetes Mellitus," *Journal of Diabetes Research* 2017, no. 1 (2017): 2823671.
- 211. T. Nonomura, A. Tsuchida, M. Ono-Kishino, T. Nakagawa, M. Taiji, and H. Noguchia, "Brain-Derived Neurotrophic Factor Regulates Energy Expenditure Through the Central Nervous System in Obese Diabetic Mice," *Journal of Diabetes Research* 2, no. 3 (2001): 201–209.
- 212. O. Hanyu, K. Yamatani, T. Ikarashi, et al., "Brain-Derived Neurotrophic Factor Modulates Glucagon Secretion From Pancreatic Alpha Cells: Its Contribution to Glucose Metabolism," *Diabetes, Obesity and Metabolism* 5, no. 1 (2003): 27–37.
- 213. G. S. Yeo, C. C. Connie Hung, J. Rochford, et al., "A de Novo Mutation Affecting Human TrkB Associated With Severe Obesity and Developmental Delay," *Nature Neuroscience* 7, no. 11 (2004): 1187–1189.
- 214. M. Karczewska-Kupczewska, I. Kowalska, A. Nikołajuk, et al., "Circulating Brain-Derived Neurotrophic Factor Concentration Is Downregulated by Intralipid/Heparin Infusion or High-Fat Meal in Young Healthy Male Subjects," *Diabetes Care* 35, no. 2 (2012): 358–362.
- 215. O. K. Idowu, S. O. Ajayi, O. O. Oluyomi, N. M. Atobatele, and A. A. Okesina, "Effect of Fenofibrate on Hippocampal Insulin Resistance and Cognitive Function in a Rat Model of Type 2 Diabetes Mellitus," *Journal of Krishna Institute of Medical Sciences* 10, no. 1 (2021): 75.
- 216. E. M. Ali, "Brain Derived Neurotrophic Factor Has a Role in the Antidepressant Effect of Atorvastatin and Fenofibrate in Wistar Rats," *Records of Pharmaceutical and Biomedical Sciences* 6, no. 3 (2022): 179–194.
- 217. N. Prowse and S. Hayley, "Microglia and BDNF at the Crossroads of Stressor Related Disorders: Towards a Unique Trophic Phenotype," *Neuroscience & Biobehavioral Reviews* 131 (2021): 135–163.
- 218. A. Sharma and H.-J. Lee, "Role of Phytoconstituents as PPAR Agonists: Implications for Neurodegenerative Disorders," *Biomedicine* 9, no. 12 (2021): 1914.
- 219. B. S. K. Chua, "The Structural Selectivity and Various Ligands of PPAR Gamma" (2023).
- 220. S. A. Kliewer and D. J. Mangelsdorf, "Fibroblast Growth Factor 21: From Pharmacology to Physiology," *American Journal of Clinical Nutrition* 91, no. 1 (2010): 254S–257S.
- 221. H. Staiger, M. Keuper, L. Berti, M. Hrabě de Angelis, and H. U. Häring, "Fibroblast Growth Factor 21—Metabolic Role in Mice and Men," *Endocrine Reviews* 38, no. 5 (2017): 468–488.
- 222. H. M. Al-Kuraishy, G. E. Batiha, H. Faidah, A. I. Al-Gareeb, H. M. Saad, and J. Simal-Gandara, "Pirfenidone and Post-Covid-19 Pulmonary Fibrosis: Invoked Again for Realistic Goals," *Inflammopharmacology* 30, no. 6 (2022): 2017–2026.
- 223. H. M. Al-Kuraishy, A. I. Al-Gareeb, H. M. Saad, and G. E. Batiha, "The Potential Therapeutic Effect of Statins in Multiple Sclerosis: Beneficial or Detrimental Effects," *Inflammopharmacology* 31, no. 4 (2023): 1671–1682.
- 224. H. M. Al-Kuraishy, A. I. Al-Gareeb, T. J. Al-Maiahy, A. Alexiou, N. Mukerjee, and G. E. Batiha, "An Insight Into the Placental Growth Factor (PlGf)/Angii Axis in Covid-19: A Detrimental Intersection," *Biotechnology and Genetic Engineering Reviews* 40, no. 4 (2024): 3326–3345.
- 225. S. Chen, S. T. Chen, Y. Sun, et al., "Fibroblast Growth Factor 21 Ameliorates Neurodegeneration in Rat and Cellular Models of Alzheimer's Disease," *Redox Biology* 22 (2019): 101133.
- 226. S. C. Larsson and D. Gill, "Genetic Evidence Supporting Fibroblast Growth Factor 21 Signalling as a Pharmacological Target for Cardiometabolic Outcomes and Alzheimer's Disease," *Nutrients* 13, no. 5 (2021): 1504.
- 227. M. Mraz, M. Bartlova, Z. Lacinova, et al., "Serum Concentrations and Tissue Expression of a Novel Endocrine Regulator Fibroblast

- Growth Factor-21 in Patients With Type 2 Diabetes and Obesity," Clinical Endocrinology 71, no. 3 (2009): 369–375.
- 228. Y. Chornenkyy, W. X. Wang, A. Wei, and P. T. Nelson, "Alzheimer's Disease and Type 2 Diabetes Mellitus Are Distinct Diseases With Potential Overlapping Metabolic Dysfunction Upstream of Observed Cognitive Decline," *Brain Pathology* 29, no. 1 (2019): 3–17.
- 229. H. M. Al-Kuraishy, M. S. Jabir, A. K. Albuhadily, A. I. Al-Gareeb, and M. F. Rafeeq, "The Link Between Alzheimer Disease and Metabolic Syndrome: A Mutual Relationship and Long Rigorous Investigation," *Ageing Research Reviews* 91 (2023): 102084.
- 230. M. S. Al-Naimi, H. A. Rasheed, H. M. Al-Kuraishy, and A. I. Al-Gareeb, "Berberine Attenuates Olanzapine Induced-Metabolic Syndrome," *Journal of the Pakistan Medical Association* 69, no. Suppl 3 (2019): S88–S92.
- 231. G. F. Watts and J. C. Mamo, "Hypertriglyceridemia and Alzheimer Disease: Opening the Mind to New Therapeutic Opportunities," *Clinical Chemistry* 67, no. 1 (2021): 6–8.
- 232. K. Griess, M. Rieck, N. Müller, et al., "Sphingolipid Subtypes Differentially Control Proinsulin Processing and Systemic Glucose Homeostasis," *Nature Cell Biology* 25, no. 1 (2023): 20–29.
- 233. R. L. Klein, S. M. Hammad, N. L. Baker, et al., "Decreased Plasma Levels of Select Very Long Chain Ceramide Species Are Associated With the Development of Nephropathy in Type 1 Diabetes," *Metabolism* 63, no. 10 (2014): 1287–1295.
- 234. J. W. Park, W. J. Park, Y. Kuperman, S. Boura-Halfon, Y. Pewzner-Jung, and A. H. Futerman, "Ablation of Very Long Acyl Chain Sphingolipids Causes Hepatic Insulin Resistance in Mice Due to Altered Detergent-Resistant Membranes," *Hepatology* 57, no. 2 (2013): 525–532.
- 235. G. van Echten-Deckert and J. Walter, "Sphingolipids: Critical Players in Alzheimer's Disease," *Progress in Lipid Research* 51, no. 4 (2012): 378–393.
- 236. S. M. Crivelli, C. Giovagnoni, L. Visseren, et al., "Sphingolipids in Alzheimer's Disease, How Can We Target Them?," *Advanced Drug Delivery Reviews* 159 (2020): 214–231.
- 237. X. Han, D. M. Holtzman, D. W. McKeel, Jr., J. Kelley, and J. C. Morris, "Substantial Sulfatide Deficiency and Ceramide Elevation in Very Early Alzheimer's Disease: Potential Role in Disease Pathogenesis," *Journal of Neurochemistry* 82, no. 4 (2002): 809–818.
- 238. K. Buschard, M. Blomqvist, T. Osterbye, and P. Fredman, "Involvement of Sulfatide in Beta Cells and Type 1 and Type 2 Diabetes," *Diabetologia* 48 (2005): 1957–1962.
- 239. G. F. Oxenkrug, "Increased Plasma Levels of Xanthurenic and Kynurenic Acids in Type 2 Diabetes," *Molecular Neurobiology* 52 (2015): 805–810
- 240. D. Zhen, J. Liu, X. D. Zhang, and Z. Song, "Kynurenic Acid Acts as a Signaling Molecule Regulating Energy Expenditure and Is Closely Associated With Metabolic Diseases," *Frontiers in Endocrinology* 13 (2022): 847611.
- 241. E. Yu, C. Papandreou, M. Ruiz-Canela, et al., "Association of Tryptophan Metabolites With Incident Type 2 Diabetes in the PREDIMED Trial: A Case-Cohort Study," *Clinical Chemistry* 64, no. 8 (2018): 1211–1220.
- 242. M. González-Sánchez, J. Jiménez, A. Narváez, et al., "Kynurenic Acid Levels Are Increased in the CSF of Alzheimer's Disease Patients," *Biomolecules* 10, no. 4 (2020): 571.
- 243. Z. Hartai, A. Juhász, Á. Rimanóczy, et al., "Decreased Serum and Red Blood Cell Kynurenic Acid Levels in Alzheimer's Disease," *Neurochemistry International* 50, no. 2 (2007): 308–313.
- 244. A. Ostapiuk and E. M. Urbanska, "Kynurenic Acid in Neurodegenerative Disorders—Unique Neuroprotection or Double-Edged Sword?," CNS Neuroscience & Therapeutics 28, no. 1 (2022): 19–35.

- 245. J. Zhao, L. Li, Z. Pei, et al., "Peroxisome Proliferator Activated Receptor (PPAR)- $\gamma$  Co-Activator 1- $\alpha$  and Hypoxia Induced Factor-1 $\alpha$  Mediate Neuro-and Vascular Protection by Hypoxic Preconditioning In Vitro," *Brain Research* 1447 (2012): 1–8.
- 246. V. J. de Oliveira Bristot, A. C. de Bem Alves, L. R. Cardoso, D. da Luz Scheffer, and A. S. Aguiar, Jr., "The Role of PGC- $1\alpha$ /UCP2 Signaling in the Beneficial Effects of Physical Exercise on the Brain," *Frontiers in Neuroscience* 13 (2019): 292.
- 247. A. P. Russell, "PGC- $1\alpha$  and Exercise: Important Partners in Combating Insulin Resistance," *Current Diabetes Reviews* 1, no. 2 (2005): 175–181.
- 248. K. S. Vimaleswaran, V. Radha, S. Ghosh, et al., "Peroxisome Proliferator-Activated Receptor- $\gamma$  Co-Activator- $1\alpha$  (PGC- $1\alpha$ ) Gene Polymorphisms and Their Relationship to Type 2 Diabetes in Asian Indians," *Diabetic Medicine* 22, no. 11 (2005): 1516–1521.
- 249. T. Nelson, T. Fingerlin, L. Moss, M. Barmada, R. Ferrell, and J. Norris, "The Peroxisome Proliferator-Activated Receptor Gamma Coactivator-1 Alpha Gene (PGC-1 $\alpha$ ) Is Not Associated With Type 2 Diabetes Mellitus or Body Mass Index Among Hispanic and Non Hispanic Whites From Colorado," *Experimental and Clinical Endocrinology & Diabetes* 115, no. 4 (2007): 268–275.
- 250. W. Qin, V. Haroutunian, P. Katsel, et al., "PGC-1α Expression Decreases in the Alzheimer Disease Brain as a Function of Dementia," *Archives of Neurology* 66, no. 3 (2009): 352–361.
- 251. T. Wenz, "Mitochondria and PGC-1α in Aging and Age-Associated Diseases," *Journal of Aging Research* 2011, no. 1 (2011): 810619.
- 252. J. Lv, S. Jiang, Z. Yang, et al., "PGC-1 $\alpha$  Sparks the Fire of Neuroprotection Against Neurodegenerative Disorders," *Ageing Research Reviews* 44 (2018): 8–21.
- 253. M. Suwa, T. Egashira, H. Nakano, H. Sasaki, and S. Kumagai, "Metformin Increases the PGC-1α Protein and Oxidative Enzyme Activities Possibly via AMPK Phosphorylation in Skeletal Muscle In Vivo," *Journal of Applied Physiology* 101, no. 6 (2006): 1685–1692.
- 254. X. Xu, Y. Sun, X. Cen, et al., "Metformin Activates Chaperone-Mediated Autophagy and Improves Disease Pathologies in an Alzheimer Disease Mouse Model," *Protein & Cell* 12, no. 10 (2021): 769–787.
- 255. G. Z. Liu, T. T. Hou, Y. Yuan, et al., "Fenofibrate Inhibits Atrial Metabolic Remodelling in Atrial Fibrillation Through PPAR- $\alpha$ /Sirtuin 1/PGC-1 $\alpha$  Pathway," *British Journal of Pharmacology* 173, no. 6 (2016): 1095–1109.
- 256. Y. A. Hong, J. H. Lim, M. Y. Kim, et al., "Fenofibrate Improves Renal Lipotoxicity Through Activation of AMPK-PGC-1 $\alpha$  in Db/Db Mice," *PLoS One* 9, no. 5 (2014): e96147.
- 257. Y. R. Cho, J. H. Lim, M. Y. Kim, et al., "Therapeutic Effects of Fenofibrate on Diabetic Peripheral Neuropathy by Improving Endothelial and Neural Survival in Db/Db Mice," *PLoS One* 9, no. 1 (2014): e83204.