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Fanconi anemia and dyskeratosis congenita/telomere biology disorders: Two inherited bone marrow failure syndromes with genomic instability

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Inherited bone marrow failure syndromes (IBMFS) are a complex and heterogeneous group of genetic diseases. To date, at least 13 IBMFS have been characterized. Their pathophysiology is associated with germline pathogenic variants in genes that affect hematopoiesis. A couple of these diseases also have genomic instability, Fanconi anemia due to DNA damage repair deficiency and dyskeratosis congenita/telomere biology disorders as a result of an alteration in telomere maintenance. Patients can have extramedullary manifestations, including cancer and functional or structural physical abnormalities. Furthermore, the phenotypic spectrum varies from cryptic features to patients with significantly evident manifestations. These diseases require a high index of suspicion and should be considered in any patient with abnormal hematopoiesis, even if extramedullary manifestations are not evident. This review describes the disrupted cellular processes that lead to the affected maintenance of the genome structure, contrasting the dysmorphological and oncological phenotypes of Fanconi anemia and dyskeratosis congenita/telomere biology disorders. Through a dysmorphological analysis, we describe the phenotypic features that allow to make the differential diagnosis and the early identification of patients, even before the onset of hematological or oncological manifestations. From the oncological perspective, we analyzed the spectrum and risks of cancers in patients and carriers.

KEYWORDS

Fanconi anemia (FA)/BRCA pathway, dyskeratosis congenita (DC), telomere biology disorders, genomic instability, PHENOS, VACTERL-H, GINPOD, cancer risk

1 Introduction

The inherited bone marrow failure (BMF) syndromes (IBMFS) are a group of genetic and hereditary diseases characterized by common childhood onset exhaustion of the hematopoietic stem and progenitor cells (HSPCs) pool, a high frequency of extra-medullary phenotypes, and variable degree of cancer risk (1). BMF at any age should prompt a consideration of a IBMFS diagnosis (2). The two most frequent and probably best characterized syndromes of this group are Fanconi anemia (FA) that results from germline pathogenic variants (PVs) in genes of the FA/Breast Cancer Susceptibility (BRCA) DNA repair pathway and dyskeratosis congenita (DC) in which telomere maintenance genes are affected (3, 4). Recently, the designation telomere biology disorders (TBDs) have gained relevance for describing the spectrum of phenotypes associated with the telomere maintenance defects, including DC (3, 4). Although FA and DC/TBD are recognizable different syndromes, they may share a frequent initial clinical presentation consisting of primary refractory pancytopenia, megaloblastosis, cutaneous dyschromia, and growth retardation. Before cellular phenotype testing and molecular investigations were possible, early literature proposed that DC was a variant of FA (5), implying that historically FA and DC/TBD have been considered differential diagnoses to each other. Currently, the DC/TBD Diagnosis and Management Guidelines consider performing chromosome breakage analysis to rule out FA in the work-up for BMF evaluation.

In this review, we present a thorough analysis contrasting the phenotype of FA and DC/TBD from molecular and cytogenetic findings to dysmorphological and oncological aspects, making a one-to-one comparison between both conditions. We make special emphasis in contrasting their physical characteristics and highlighting the differences that could help in the differential diagnosis.

2 Pathophysiology

2.1 FA/BRCA DNA repair pathway

The *FANC* genes protein products collaborate in the socalled FA/BRCA pathway (6). This is a biochemical ensemble that maintains genome integrity by protecting the DNA replication forks during the S phase of the cell cycle and by promoting DNA repair (7). DNA replication is a timely process

triggered by the activation of multiple origin replication sites in the lineal eukaryotic chromosomes, starting multiple replication forks that will coordinately replicate the DNA molecule (8). DNA replication is achieved by DNA polymerases that move along with the replication fork machinery, incorporating nucleotides into a nascent daughter DNA strand (9). The DNA replication process is not seamless, and addition of nucleotides by the replication fork machinery can be interrupted by diverse structures occurring in the template DNA molecule (10). These structures include damaged DNA bases, DNA-protein complexes, DNA-RNA hybrids (R-loops), and certain DNA structures inherent to the complementary nature of the DNA bases, such as G quadraplexes (10, 11). All these events have the capacity to dampen or slow down the progression of the replication forks and may retard the timely replication of the entire DNA molecule.

The better known function of the FA/BRCA pathway is the removal of interstrand cross-links (ICLs) in the DNA molecule (7) (Figure 1). ICLs are structures covalently binding both DNA strands that interfere with DNA replication and genetic transcription by impeding the separation of the two DNA strands (10). ICLs can have an endogenous origin: Normal metabolic transactions of the cell generate by-products, which include aldehydes, reactive oxygen species, and nitrous acid, with the capacity to react with the DNA molecule (12, 13). They can also have an exogenous origin, ICLs inducing drugs are a cornerstone of cancer chemotherapy, and it is precisely the replicative damage that they generate what leads to apoptosis activation in cancer cells. Frequently used agents include platinums and nitrogen mustards (10).

The FA/BRCA pathway comprises a series of coordinated actions that allow (1) the recognition of the ICLs (2), the removal of the ICLs with the generation of a double-strand break (DSBs) as a DNA repair intermediate; and (3) the repair of the DSB by homologous recombination (HR) (6, 7) (Figure 1).

ICLs are recognized during S phase, mainly by the UHRF1 protein which, together with an anchor complex composed of Fanconi Anemia Complementation Group M (FANCM) and FAAP24, ensure the recruitment of the FA core complex and the FANCD2-I heterodimer to the appropriate location in the chromatin (14). The positioning of these complexes constitutes the upstream portion of the FA/BRCA pathway (15–18). The FA core complex is a large protein assembly with E3 ubiquitin ligase enzymatic activity, integrated of at least 10 proteins (FANCA, FANCB, FANCC, FANCE, FANCE, FANCG, FANCC, FANCE, FANCG, FANCC, FANCE, FANCE, FANCC, FANCE, FANCE, FANCC, FANCE, FANCE, FANCC, FANCE, FANCE



FIGURE 1

The Fanconi anemia pathway detects and repairs DNA interstrand cross-links. (A) An ICL covalently holds together the two complementary DNA strands. ICLs arrest the progression of the replication fork and are recognized by FANCM and its associated proteins. (B) The FANCM anchor complex recruits the FA core complex to the ICL site. The FA core complex in turn ubiquitylates the FANCD2-I complex. (C) Ubiquitylated FANCD2-I complex forms a filamentous array that clamps the stalled DNA replication fork and protects it from nucleolytic degradation. The FANCP/SLX4 is a DNA endonucleases toolbox that will unhook the ICL site by cleaving the surrounding DNA. (D) After ICLs unhooking two DNA repair intermediates appear, a DSB that will be repaired by the downstream FA/BRCA proteins using HR and a DNA adduct that will be processed with the help of TLS DNA polymerases. (E) The DSB will initially undergo a short resection mediated by exonuclease activity of the MRN complex. (F) Extensive DSB resection will be performed by the EXO1 endonuclease, and the produced DNA overhang will be covered and protected by RPA1 protein subunits. (G) The HR mechanism will start with the exchange of RPA1 by subunits of the recombinase RAD51. This exchange will be assisted by homologous recombination mediators including FANC/PALB2, FANCDI/BRCA2, and FANCS/BRCA1. (H) RAD51- coated DNA overhangs will function as recombinase filaments that will search for sequence homology. (I) Strand invasion will be followed by sequence detection, copy and synthesis of the sequence of interest, Holliday junctions resolution, and ligation of the phosphate DNA backbone. DSBs, double-strand breaks; FA, Fanconi anemia; HR, homologous recombination; ICLs, interstrand cross-links; TLS, translesion synthesis.

and FAAP20) (17). The FA core complex recruits the E2 conjugating enzyme UBE2T/FANCT to monoubiquitylate the FANCD2-I complex (Ub-FANCD2I) (19, 20). Ub-FANCD2I is being long recognized as essential for the recruitment of proteins involved in the DNA repair downstream activities of the FA/ BRCA pathway (21), as FANCD2-I forms nuclear DNA repair foci co-localize with DNA repair factors (22). However, recent structural studies have shown that the recruitment of additional DNA repair factors to the sites of damage occurs independently of Ub-FANCD2-I and, indeed, Ub-FANCD2-I forms a nucleoprotein array that clamps around stalled replication forks. This Ub-FANCD2-I clamp may, instead of recruiting DNA repair factors, protect the DNA replication fork by preventing the activity of DNA nucleases, such as MRE11 and DNA2, that would resect and destabilize the DNA replication fork (23-25). Importantly, cells from patients with FA with PVs in components of the FA core complex do not have the capacity to ubiquitylate the FANCD2-I complex and cannot make these critical foci that maintain genome stability (17).

FANCP/SLX4 is a scaffolding protein that allows the engagement of multiple DNA endonucleases: MUS81, SLX1, and FANCQ/XPF/ERCC4 (26, 27). FANCP/SLX4 is thought to be recruited by Ub-FANCD2-I, but recent reports have also shown that FANCP/SLX4 can directly recognize stalled replication forks and can be recruited to the site of damage independently from Ub-FANCD2 (28, 29). The endonucleases coordinated by FANCP/SLX4 would cleave the phosphobackbone of the double stranded DNA, cut the DNA strand contiguous to the ICLs, and generate a DNA adduct and a DSB (30). These two secondary DNA lesions need to be further funneled into appropriate downstream pathways for repair. REV1 and REV3 are low-processivity translesion synthesis DNA polymerases with the capacity to bypass the DNA adduct generated after ICL incision and ensure the continuity of the DNA replication process (31), whereas HR, a high-fidelity DSBs repair pathway, takes charge of the DSB generated after ICL incision (7). Switching off the FA/BRCA pathway is also relevant, and this happens when the USP1-UAF1 complex deubiquitylates the FANCD2-I complex, adding an additional layer of regulation to the FA/BRCA pathway (17).

The FA/BRCA pathway blocks the activity of alternative lowfidelity DNA repair pathways, including the non-homologous end-joining (NHEJ) pathway, and promotes the DSB repair by HR (32). This ensures fidelity in the repair of the DSB generated after ICL cleavage. In this high-fidelity pathway, the ends of the DSB are processed by the DNA exonucleases CtIP and the MRN (MRE11-RAD50-NBS1) complex, initially performing a short resection, followed by an extended resection that is mediated by EXO1 (33). This generates a 3' single-stranded DNA (ssDNA) overhang that is coated by replication protein A1 (RPA1), a protein that prevents formation of secondary structures in the ssDNA due to auto-complementarity (34). Eventually, RPA1 is evicted from the ssDNA and is substituted by FANCR/RAD51,

forming a recombination filament that performs homology search in the sister chromatid (35). The recombination filament invades the homologous sequence aided by the FANCD1/ BRCA2, FANCN/PALB2, FANCO/RAD51C, FANCJ/BRIP1/ BACH1, and FANCU/XRCC1 proteins, when it detects enough homology, it copies the sequence of interest aided by DNA polymerases (7, 21). Of note, cells from patients with FA who have PVs in downstream components of the FA/BRCA pathway can ubiquitylate FANCD2-I foci in the presence of ICLs (36). These cells can detect the ICLs and perform the initial ICLs unhooking; however, they are not able to perform the accurate repair of DSB by HR. Patients with PVs in the downstream FANC genes are even more rare than patients with PVs in the upstream FA/BRCA pathway or FANCD2-I complex (37-39). This may probably be due to early lethality since lack of HR proteins will affect the repair not only of DSB generated during ICL processing but also the repair of direct DSB, thus affecting the tolerability to more DNA damaging agents and processes.

2.2 The telomere maintenance system

The linear chromosomes found in eukaryotic cells need protection of their ends from nuclease degradation and require special structures and mechanisms for this task. The telomere is a complex structure located at the ends of the linear chromosomes that accomplishes this protective role. Telomeres are composed of repetitive DNA sequences coated by specialized proteins, ribonucleoprotein complexes, and longnoncoding RNA (40–42) (Figure 2).

The telomeric DNA sequence is highly conserved among vertebrates; it is composed of the repetitive hexameric sequence TTAGGG (43). In human chromosomes, it can extend for up to 5 kb (44), whereas, in mice, it may go up to 100 kb (45). The architecture of the telomeric DNA is special; it is composed of a region of double-stranded DNA that is several kilobases in length, where the TTAGGG hexamer and its complementary sequence AATCCC are repeated. Eventually, the AATCCC rich strand is discontinued and only the single-stranded 3' tail known as the Goverhang continues (enriched in the TTAGGG sequence) (41). The G-overhang invades the preceding double-stranded region generating a lariat-like structure known as the telomere loop or *tloop* (42). Importantly, the telomeric length of the chromosomes is progressively shortened with every cell division and serves as a "molecular clock" of the proliferative life span of primary cells (46). Telomeric lengths among chromosomes of the same cell, tissue, and organism can be highly heterogeneous (47). Of upmost relevance, critically short telomeres are known to trigger the entrance of the cell to senescence, also known as the Hayflick limit of proliferative life span, a mechanism that controls the tissues life span and regeneration (46).

At least three different protein complexes with different and specific functions have been associated with the telomeres



(Figure 2). First, the shelterin complex, a six-subunit protein complex that binds directly to the telomeric repeats and protects chromosome ends by inhibiting the activation of the DNA damage response (DDR) (explained below) (48). The human shelterin complex includes six proteins: telomere repeat binding factor 1 (TRF1), TRF2, repressor/activator protein 1 (RAP1), TRF1-interacting nuclear protein 2 (TIN2), TIN2-interacting protein 1 (TPP1), and protection of telomeres 1 (POT1) (40, 41, 48).

The second telomere-associated complex elongates the TTAGGG telomeric sequences through reverse transcriptase activity. This ribonucleoprotein complex is a telomerase that synthesizes new telomere sequences onto chromosome ends (49). The telomerase holoenzyme has two components: Telomerase Reverse Transcriptase (TERT), the core telomerase protein, which contains the telomerase reverse transcriptase domain; and Telomerase RNA Component (TERC), the RNA component, which provides the template for telomeric sequence synthesis (49). The human telomerase is assembled in the Cajal bodies (50).

The telomerase complex relies on other proteins for its assembly and forms a ribonucleoprotein complex with several accessory proteins, among which dyskerin is the best characterized among mammalian telomerase complexes and the most frequently affected gene (DKC1) in patients with DC/ TBD (49, 51). Three other smaller accessory proteins accompany dyskerin, namely, NHP2, NOP10, and GAR1 (49, 52). Dyskerin associates with TERC, the RNA component of the complex, through its binding to an H/ACA box structural motif within TERC, which is essential for TERC stability and telomerase function (49, 53). Telomerase uses its intrinsic RNA template to synthesize telomeric DNA repeats and has the capacity to add ~60 nucleotides per telomere each cell cycle. The telomerase complex is recruited by the shelterin complex (41). Importantly, telomerase is more abundant in germline cells and is absent or less abundant in somatic cells (41, 54).

A third complex is the CST complex, composed of three proteins: CTC1, STN1, and TEN1. The CST complex aids in terminating the telomeric extension performed by the telomerase complex. CST complex binds to the newly extended single-stranded DNA and facilitates the recruitment of DNA polymerase alpha-primase (Pol α -primase) (41, 55). Pol α -primase fills in the C-strand (AATCCC), complementary to the recently extended TTAGGG overhang, thus helping convert part of the recently extended tail into double stranded DNA, leaving the overhang length to ~50–300 nucleotides (55).

A less-well characterized component of the telomeric complex is the long non-coding telomeric repeat-containing RNA (TERRA), which is transcribed by the RNA polymerase II from intrachromosomal telomeric repeats. Although TERRA function is not fully understood, it may work as a scaffold for telomeric proteins (mentioned below) (56).

2.3 Defects in both the FA/BRCA pathway and the telomere maintenance system exacerbate the cellular DNA damage response

The FA/BRCA pathway and the telomere maintenance system promote genome stability (6, 48). Importantly, both are linked, in opposing ways, to the DDR that monitors the integrity of the genome. On the one hand, the presence of DNA damage promotes activation of the cell cycle checkpoints, halts cell cycle progression, and allows the FA/BRCA pathway to perform repairing activities (7). On the other hand, the relationship between the telomere associated proteins and the DDR is the opposite; the components of the shelterin complex inhibit the activation of the DDR; this prevents the telomeric ends to be recognized as unrepaired DSB and joined with other DSB (57) (Figure 3).

In this context, the main DDR orchestrators are the members of the PIKK family of kinases, namely, ATM (ataxia telangiectasia mutated), ATR (ataxia telangiectasia and Rad3-related), and DNA-PKCs. ATM is known to signal DSB, whereas ATR recognizes stalled replication forks and RPA1-coated single-stranded DNA (58, 59). The best known ATM target is the checkpoint kinase 2 (CHK2) (60), whereas for ATR, it is CHK1 (61). The ATM–CHK2 axis, as well as the ATR–CHK1 axis, phosphorylates p53, a master transcription factor that activates the expression of proteins that block cell cycle progression (62). In this sense, the presence of DNA damage, in the form of ICLs and DSBs, promotes p53 activity to block cell cycle progression and allowing DNA repair to occur. In contrast, if the DNA damage is excessive, p53 activity will promote cell death by apoptosis (63) (Figure 3).



DSBs and deprotected telomeres trigger similar DNA damage signaling. The members of the PIKK family of kinases are activated by DNA damage. Both DSBs, generated either by exogenous or endogenous sources, and deprotected telomeres are recognized as DNA damage that must be repaired. Excessive DNA damage leads to activation of p53, the guardian of the genome. p53 is a transcription factor with the capacity to control the activation of cell cycle arrest genes when DNA damage is repairable; however, unsurmountable amounts of DNA damage will lead the cell into apoptosis, also in a p53-dependent manner. p53-mediated apoptosis in response to DNA damage may be a common mechanism leading to HSPCs demise in Fanconi anemia and DC/TBD. DSBs, double-strand breaks; DC/TBD, dyskeratosis congenita/Telomere biology disorders; HSPCs, hematopoietic stem and progenitor cells; PIKK, phosphatidylinositol-3 kinase-related kinases.

ATM establishes an activation loop with the MRN complex (64). The MRN complex has an endonuclease activity that resects DSB and generates ssDNA that will be coated by RPA1 subunits (65); this will subsequently activate ATR and reinforce the checkpoint signaling (59). In the context of the FA/BRCA pathway, ATR will be involved in the initial recognition of ICLs, will phosphorylate FANCM (66, 67), as well as Fanconi Anemia Complementation Group I (FANCI) (23, 68) for promoting the activation of the FA pathway.

At the telomeres, the story is the opposite, the illegitimate repair of telomeric ends is actively inhibited (48). Unleashing the DNA repair activity at the telomeres can have catastrophic consequences, including the formation of end-to-end chromosome fusions, resulting in dicentric chromosomes that will conflict the chromosome segregation process during mitosis (69–71). This can lead to breakage–fusion–bridge (BFB) cycles and chromothripsis, a single event of thousands of genomic breakages followed by chromosomal rearrangements clustered in a confined genomic region in one or a few chromosomes (72). These alterations generate great amounts of structural chromosomal damage (explained in detail in the next section). This repressive activity lies on the components of the shelterin complex that actively inhibit the PIKK kinases. More specifically, TRF2 inhibits ATM and POT1 inhibits ATR (73).

In the absence of TRF2, ATM phosphorylates its targets CHK2 and p53. The later activates the recruitment of DNA damage repair proteins at the unprotected telomeres forming telomere dysfunction-induced foci (74). These foci are characterized by the presence of yH2AX and recruitment of 53BP1 (p53-binding protein 1), a well-known NHEJ protein (74). TRF2 also blocks the dimerization of the Ku complex, thereby preventing the activation of NHEJ (75, 76). Therefore, TRF2 is a multipurpose protein that inhibits the checkpoint activation and NHEJ at the chromosome ends, whereas POT1 inhibits alt-EJ of DSB (77).

Two mechanisms have been proposed to explain how TRF2 inhibits ATM. The first involves the *t-loop* formation; this lariat-like structure is stimulated by TRF2 and helps hiding the telomere ends from being detected by MRN (78, 79). The second prevents further ATM activation and avoids the DNA ends to be processed by the nucleolytic activity of MRN. TRF2 might also inhibit ATM directly by inhibiting its kinase activity (80).

POT1 is an ssDNA-binding protein specially located at the telomeric ends (81). POT1 binds directly to the telomeric ssDNA and, with the help of the heterogeneous nuclear ribonucleoprotein A1 and TERRA, displaces RPA, avoiding its recruitment, preventing the accumulation of RPA-coated ssDNA (Figure 2), the critical substrate needed for ATR activation (81). POT1 depletion leads also to the formation of telomere dysfunction-induced foci in an ATR-dependent manner. ATR can also be inhibited by TRF1; this happens during telomere replication with the help of TPP1 and POT1 (82).

When the FA pathway is nonfunctional or the telomere maintenance system fails, there will be checkpoint activation and the subsequent activation of p53. Of note, activation of p53 has been observed in HSPCs from patients with FA (83) and human embryonic stem cells with PVs in genes relevant for DC/TBD (84). Altogether, this suggests that p53 is the main executor of the cell death program in both diseases, leading to BMF by depletion of the HSPCs pool. Abrogation of p53 rescues the proliferation capacity of FA cells; however, losing p53 promotes the division of cells with great amounts of DNA damage and potentially to malignancy (83, 84). Recently, overexpression of MYC, a proto-oncogen, was observed in HSPCs from patients with FA, presumably as a counterbalancing mechanism against the p53-mediated cell death. MYC, however, produces great levels of replication stress and DNA damage that eventually might reinforce p53 activation, in what seems to be a vicious cycle (85). MYC activation has not been studied in DC/TBD, but MYC may also be a force allowing the survival of cells deficient in telomere protection.

2.4 Chromosomal instability in FA and DC/TBD

2.4.1. Fanconi anemia

The FA/BRCA pathway is essential for the repair of ICLs (4, 6). Having a non-functional FA/BRCA pathway has several consequences; one of them is that ICL-derived DSB will not be repaired error free by HR (4). For this reason, FA cells are highly sensitive to endogenous and exogenous agents that generate ICLs (7, 75). Some examples are as follows: cisplatin and mitomycin C (MMC), drugs that are used in the treatment of cancer, and also diepoxybutane (DEB) (12, 86).

FA cells, therefore, rely on low-fidelity alternative DNA repair pathways, including the canonical NHEJ (4, 87) and potentially the alternative microhomology-mediated endjoining pathway (MMEJ). Both pathways are error prone, and their excessive use increases the frequency of chromosomal abnormalities, mainly of the structural type.

The processing and repair of DSBs are influenced by several factors, including the cell cycle phase and the number of DSBs simultaneously present in the same cell (88, 89).

In an FA cell, the HR pathway is deficient and the number of DSBs is high. Under these conditions, FA cells recur to error prone end-joining repair mechanisms that operate throughout the entire cell cycle. The emergency posed by the presence of several DSBs may be solved with the activity of alternative end-joining pathways, which intrinsically are template-independent and need a minimal homology for joining free DNA ends (Table 1) (90, 91). However, the potentially indiscriminate ligation of DNA that ends from different chromosomes may lead to gross chromosomal abnormalities (4). When metaphase spreads from patients with FA are analyzed, chromatid breaks

	Speed	Template dependence	Homology usage	End resection	Cell cycle phase	Accuracy
Homologous Recombination	Slow	Dependent	>100 base pairs	Yes	S, G2	Highly accurate
Microhomology Mediated End-Joining	Fast	Independent	2-20 base pairs	Yes	G1, S, and G2	Error prone
Non homologous End-Joining	Fast	Independent	0-4 base pairs	No	G1, S, and G2	Error prone

TABLE 1 Characteristics of the major double-strand breaks repair pathways.

References (90, 91).

can be observed, which is the evidence of unrepaired DSBs, as well as radial figures and other types of structural chromosomal aberrations, resulting from the erroneous ligations of DSBs coming generally from chromatids of non-homologous chromosomes (Figure 4) (92–94).

Radial figures originate in S/G2 phases, when two chromatids become available from two replicated chromosomes. Unrepaired DSBs lead to chromatid breaks and, when rejoined by the error-prone DNA repair pathways, radial figures are produced after the ligation of chromatids from different chromosomes. The abnormal processing and repair of the ICLs might generate multiple types of complex structural chromosomal abnormalities, including translocations, deletions, or dicentric chromosomes. Abnormal segregation of dicentric chromosomes during mitosis may also be a source of complex rearrangements.

Translocations, deletions, and duplications may arise when the two centromeres of a dicentric chromosome are pulled to different mitotic poles during anaphase (Figure 5). These structural aberrations are frequently accompanied by numerical abnormalities, such as aneuploidies and endoreduplications. Endoreduplication occurs when a cell enters a new round of DNA replication in the absence of cytokinesis, causing a numerical anomaly leading to the appearance of "double" chromosomes in metaphase spreads when the cells with the endorreduplicated material reach mitosis (95).

The presence of chromosomal instability (CI) is used for FA diagnosis by comparing the frequency of chromosomal abnormalities in the subject sample after ICLs induced damage, to those produced in a negative control (a healthy donor) and, ideally, a positive control (a FA cell line) (92, 94, 96). The presence of radial figures and an ~10-fold increase in the frequency of chromosomal abnormalities indicates a positive diagnosis for FA (92, 93). Importantly, the genomic instability that characterizes FA can generate revertant mutants that recover the DNA repair capacity. Up to 30% of patients with FA may be hematologic mosaics based on normalization of their blood counts. Nevertheless, two studies that intentionally looked for somatic reversion found that only 15% of patients



FIGURE 4

Chromosomal aberrations in Fanconi anemia. (A) Exposure of FA cells to ICL-inducing agents increases the number of structural chromosome aberrations and the number of aberrations per cell. Arrows show radial figures, and arrowheads show chromatid breaks; both are characteristic chromosomal aberrations found in FA cells. (B) Representation of two replicated non-homologous chromosomes with their sister chromatids linked by cohesins and each with a DSB in only one chromatid. Two types of radial figures may arise when the DSBs in two different replicated chromosomes are mis-rejoined. Tetraradial 1 results from the end joining of two chromosome proximal segments. Mitotic segregation of tetraradial 1 produces daughter cells with a dicentric chromosome and an acentric fragment (originated by the ligation of the two distal segments). Tetraradial 2 results from the joining of one proximal and one distal segment of two different chromosomes. Mitotic segregation of tetraradial 2 results in daughter cells with translocations. DSBs, double-strand breaks; FA, Fanconi anemia; ICLs, interstrand cross-links.



Chromosomal consequences of telomere attrition. (A) Telomere attrition produces telomere fusions that originate dicentric chromosomes. (B) Segregation of the dicentric when the two centromeres are pulled by opposite poles of the mitotic spindle is the main source of chromosome and cell abnormalities, because the dicentric may induce a breakage–fusion–bridge cycle. If the chromatin of the dicentric bridge is enzymatically fragmented or if the dicentric breaks and a micronucleus is formed, gross structural rearrangements, such as chromothripsis, may arise in the daughter cells. Alternatively, when chromatin bridges are not resolved, interruption of mitosis and cytokinesis, may lead to polyploid cells and endorreduplicated chromosomes in the next mitosis. Until now, chromothripsis has not been detected in cells from patients with DC/ TBD; however, anaphase bridges, fragmentation of a single chromosome, which is the precedent of chromothripsis, endoreduplications, and polyploidy have been found in DC cells.

developed mosaicism (97, 98). A recent review of FA mosaic publications with emphasis on blood count normalization alerted of a possible publication bias and described that some degree of mosaicism is present in around 15% of FA cases, but the incidence of PV reversion occurs in fewer than 5% (99). Therefore, when the frequency of chromosome aberrations and the number of aberrant cells shows intermediate values compared with a positive control, a mosaic must be suspected. In these cases, the FA diagnosis must be confirmed in skin fibroblasts, for which revertant mosaicism has not been described and should retain the MMC/DEB sensitivity (99).

Chromosomal damage can be lethal, and cell death may be activated by accumulation of gross chromosomal alterations or by mitosis blockage; however, non-lethal DNA damage may create a vicious cycle of non-clonal chromosomal aberrations and CI, rendering a heterogeneous cell population from which natural selection might pick a successful clone that can evolve into cancer (4).

2.4.2 DC/Telomere biology disorders

Telomeres are structures that protect the chromosome ends from erosion and illegitimate fusion with other chromosomes (91). Telomerase and telomere-stabilizing proteins maintain the normal length and function of telomeres in stem cells of some tissues, such as germ cells, HSPCs, activated T-cells, monocytes, skin, and intestinal lining; therefore, the lifelong maintenance and correct function of these cells depend on the telomerase activity (100).

DC/TBD patients have an excessive shortening of telomeric repeats, below the first percentile for age, and the telomeres become uncapped (101). Cells detect this as a DSB and trigger the DDR (102, 103). Erroneous rejoining of DSBs may result in telomere-telomere fusions between chromosomes, provoking structural chromosomal aberrations, such as dicentric or multicentric chromosomes.

During mitosis, a dicentric chromosome may be captured by microtubules from opposite poles of the mitotic spindle, causing anaphase/telophase bridges; this affects the proper completion of mitosis and promotes tetraploidization and aneuploidy. Dicentric bridges may persist through mitosis and cytokinesis, inducing the chromosome BFB cycles that directly affect chromosomal segregation. It has been proposed that resolution of the chromatin bridges implies the formation of ssDNA segments and formation of micronuclei containing whole chromosomes or chromosome fragments. These anomalies might lead to chromothripsis, yet the latter has only been observed *in vitro* (72).

Telomere shortening in DC/TBD has been widely documented (3, 104). However, the formation of dicentrics and their consequent CI remain controversial due to contradictory results. On several occasions, the presence of structural and numerical alterations, both spontaneous and induced, has been intentionally sought (98, 105-107). Most of these reports have been negative when the spontaneous chromosomal aberrations and the MMC/DEB-induced DNA damage have been interrogated (98, 105-107). However, other studies report positive results, with elevated percentage of spontaneous structural alterations, including chromatid and chromosomal breaks (108, 109), as well as DNA damage induced by Xradiation, 4-nitro-quinoline-1-oxide (4NQO), and bleomycin (107, 110, 111). Still, Dokal et al. did not find hypersensitivity to DEB, MMC, 4NQO, bleomycin, or gamma radiation in fibroblasts from patients with DC/TBD (112).

The different findings on the cytogenetics of DC/TBD may be related to the type of cells studied. For example, a single report showed no spontaneous chromosomal aberrations in peripheral blood lymphocytes, but those anomalies were found in skin fibroblasts from the same patient (102). The site of the biopsy from which the cells are cultured can also influence the cytogenetic findings, since they can represent tissues in a precancerous state, for example, when the aberrations are studied from bone marrow that may be in the way of a neoplastic process or fibroblasts from cutaneous lesions (102, 103). The age of the patients has also been proposed as a source of variation, since older members of the same family display chromosomal aberration in skin fibroblasts, whereas the younger ones did not (112). Of note, dicentric chromosomes, expected in DC/TBD due to increased telomeres stickiness, could not appear spontaneously.

Interestingly, some cytogenetic studies have found nonclassical chromosome abnormalities, such as pulverization (fragmentation of the entire chromatin) and fragmentation of individual chromosomes, whereas the rest of the chromosomes in the same metaphase spread appear normal (109, 113). Other genomic events found in DC/TBD cells include micronuclei, lagging chromosomes, and anaphase bridges (113), as well as endoreduplications and polyploidy (114). These types of cytogenetic findings may be important in the light of recent discoveries, according to which the BFB cycle can cause cell death (as pulverized mitosis) or very irregular mitosis that could survive if cytokinesis is interrupted (endoreduplication or polyploidy). BFB can also produce micronuclei, which can contain lagging chromosomes or fragments resulting from the breakage of dicentric chromosomes; the latter could finally cause fragmentation of a single chromosome, or the chromatin bridge may be enzymatically fragmented (TREX1-mediated) and result in chromothripsis (Figure 5) (40, 72).

All these data indicate that even though FA and DC/TBD share mechanisms that result in genomic instability and

ultimately BMF, the cytogenetic features are evidently different. In FA, the presence of CI, both spontaneously and induced by MMC and DEB, is observed in practically all the cell types that have been studied from patients with FA. This characteristic is the hallmark that allows the cytogenetic diagnosis. In addition, CI is expected as a consequence of failure of the FA/BRCA pathway. In contrast, in DC/TBD, the presence of CI has not been definitively documented, even though telomeric attrition is well recognized and important in the diagnosis of DC/TBD. Finding only sporadically dicentric in cells from patients with DC/TBD is a fact that does not have a convincing explanation. It is possible that the difficult handling of dicentrics during cell division generates immediate cell death and, in cases in which the cell survives, the consequences at the chromosomal level could possibly be non-classical chromosomal abnormalities and chromothripsis, which conventional cytogenetic methods cannot detect. Molecular methodologies that can study the whole genome, such as whole genome sequencing and M-FISH, could help in the evaluation of chromosome instability in DC/TBD.

3 Dysmorphological and oncological phenotype of patients with FA and DC/TBD

The unifying characteristic of patients with DC/TBD is that they have short telomeres, yet the phenotypic presentation can be particularly broad as illustrated in Figure 6 (3, 103). DC is the archetypal TBD. It was initially characterized by the so-called DC triad that consists of reticular skin pigmentation, nail dystrophy, and oral leukoplakia (115). Although, these features are not universal nor do they exist concomitantly in all patients with DC/TBD, but they are individually found in more than 80% of patients (116). The presence of the three classic triad components has been reported in 46% of patients with DC/ TBD (115). Other consistent but less frequent physical abnormalities are pulmonary and liver fibrosis (116-118). On the severe side of clinical presentation, the DC/TBD subtypes known as Hoyeraal-Hreidarsson (HHS) and Revesz syndromes (RS) have an early presentation of BMF accompanied by neurological impairment. Specifically, HHS is associated with neurological and immunological abnormalities as well as intrauterine growth restriction (IUGR) (119-121), whereas RS includes retinopathy, neurological anomalies, and IUGR (3, 122).

DC/TBD have important genetic heterogeneity; at least 17 germline PVs in telomere biology genes have been associated with DC/TBD phenotype (Table 2) (3, 7, 118, 123–130). Autosomal dominant, autosomal recessive, and X-linked inheritance patterns have been described (3, 118) (Table 2).



small Head, Eyes, Neurological, Otological, Short stature) acronyms. In DC/TBD, the physical phenotype can be informed by the GINPOD (GastroIntestinal [including liver and esophageal], Neurological, Pulmonary, Ocular, and Dermatological [including skin, nails, and leukoplakia]) acronym. GINPOD acronym includes the anomalies described in at least 20% of patients with DC/TBD. **(B)** Patients with FA have facial dysmorphism that integrate a recognizable FA facies: It includes small eyes and palpebral fissures, ptosis, and a pointed chin and can be complemented by ear abnormalities. In DC/TBD, the more striking facial findings are dacryostenosis and limited mouth opening. **(C)** At the nervous system level, patients with FA can have pituitary malformations and posterior fossa cysts, whereas patients with the HHS and RS, severe subtypes of DC/TBD, have calcifications. Both disorders may have corpus callosum abnormalities as well as cerebellum and cerebral atrophy. **(D)** Erythroplakia and leukoplakia are the more frequent oral findings in both FA and DC/TBD; tongue leukoplakia is more frequent in DC/TBD have salate leukoplakia is more frequently found in FA. Dental phenotype is contrasting as FA has microdontia and oligodontia and DC/TBD has taurodontia. Features exclusive to FA are in green, those exclusive to DC/TBD are in red, and those shared by both disorders are in black. DC/TBD, dyskeratosis congenita/Telomere biology disorders; EA, esophageal atresia; FA, Fanconi anemia; HHS, Hoyeraal-Hreidarsson; RS, Revesz syndromes; TEF, Tracheoesophageal fistula.

Genetic heterogeneity is also remarkable in FA. PVs in at least 22 genes that work together in the FA/BRCA pathway have been identified in patients with FA (131) (Table 2). The majority of genes have an autosomal recessive inheritance pattern, except for *FANCB*, which is X-linked and *FANCR/RAD51*, which is autosomal dominant (6, 132) (Table 2).

The physical phenotype in patients with FA is variable and multisystemic as depicted in Figure 6 (37, 133). Classical abnormalities include those described in the VACTERL-H (Vertebral, Anal, Cardiac, Tracheo-esophageal fistula, Esophageal atresia, Renal, upper Limb, and Hydrocephalus) association (134, 135). From 5% to 30% of patients with FA meet the criteria for VACTERL-H (\geq 3 out of 8 features) (37, 134, 135). However, the clinician should suspect FA even if the patient has less than two VACTERL-H anomalies, especially if radial ray and kidney malformations coexist (136). Other common abnormalities are grouped in the PHENOS acronym (skin Pigmentation, small Head, small Eyes, Nervous system,

 TABLE 2 Genes, inheritance patterns, pathophysiology, and diagnostic strategies.

		oni Anemia	DC/TBD ^s			
Genes associated with the phenotype	Gene	OMIM number	Proportion attributed to pathogenic variants in each gene	Gene	OMIM number	Proportion attributed to pathogenic variants in each gene
	1. FANCA	607139	64%	1. $ACD^{\ddot{Y}}$	609377	<1%
	2. FANCB	300515	2%	2. CTC1	613129	1-3%
	3. FANCC	613899	10%	3. DCLRE1B/ SNM1B/Apollo	609683	<1%
	4. FANCD1/ BRCA2 ^{œ,±}	600185	2%	4. DKC1/NAP57 ^Ÿ	300126	20-25%
	5. FANCD2	613984	3%	5. NAF1	617868	<1%
	6. FANCE	613976	1%	6. NHP2/NOLA3 ^Ÿ	606470	<1%
	7. FANCF	613897	2%	7. NOP10/NOLA3	606471	<1%
	8. FANCG/ XRCC9	602956	8%	8. PARN ^Ÿ	604212	<1%
	9. FANCI	611360	1%	9. POT1	606478	<1%
	10. FANCJ/ BRIP1 ^œ	605882	2%	10. RPA1	179835	<1%
	11. FANCL	608111	<1%	11. $RTEL1^{\ddot{Y}}$	608833	2-8%
	12. FANCM	609644	<1%	12. STN1/OBFC1	613128	<1%
	13. FANCN/ PALB2 ^{œ,±}	610355	2%	13. TERC/hTRz	602322	5–10%
	14. FANCO/ RAD51C ^œ	602774	<1%	14. $TERT^{\dot{Y}}$	187270	1–7%
	15. FANCP/SLX4	613278	2%	15. $TINF2^{\ell}$	604319	12-20%
	16. FANCQ/ ERCC4/XPF	133520	<1%	16. WRAP53/ TCAB1 ^Ÿ	612661	<1%
	17. FANCR/ RAD51	179617	<1%	17. ZCCHC8	616381	<1%
	18. FANCS/ BRCA1 ^œ	113705	<1%	Unknown		20%
	19. FANCT/ UBE2T	610538	<1%			
	20. FANCU/ XRCC2	600375	<1%			
	21. FANCV/ MAD2L2/REV7	604094	<1%			
	22. FANCW/ RFWD3	614151	<1%			
Inheritance patterns			•			•
	Autose	omal recessiv	• • • • • • • • • • • • • • • • • • •	Autosomal don	ninant: DCI TERC/hTF	RE1B/SNM1B/Apollo, NAF1, RPA1, R, TINF2, and ZCCHC8
	A	utosomal do	minant: FANCR/RAD51			

(Continued)

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	Fanconi Anemia	DC/TBD [§]
	X-linked: FANCB	Autosomal dominant or autosomal recessive: ACD, PARN, RTEL1, and TERT
		• Autosomal recessive: CTC1, NHP2/NOLA3, NOP10/NOLA2, POT1, STN1/OBFC1, WRAP53/TCAB1 •
		X-linked: DKC1/NAP57
Pathophysiology	DNA rep	air Telomere maintenance
Diagnostic strategies	•	•
	Chromosome breakage analysis (gold standard) using DEB or MMC •	Telomere length (gold standard): southern blot, flow-FISH, or qPCR •
	Genotyping of FANC genes	Genotyping of genes associated with DC/TBD phenotype

^{ec}Well-established cancer predisposition genes when they are in heterozygosis. [±] Genotypes associated with embryonal tumors (Wilms tumor, neuroblastoma, and brain tumors [most often medulloblastomas]) and acute myeloid leukemia at an early age. [§]Some other genes have been associated with telomere maintenance but there is insufficient evidence to consider them as DC/TBD genes. [£]Pathogenic variants in this gen are associated with Revesz syndrome. ^ŶPathogenic variants in this gen are associated with Hoyeraal-Hreidarsson syndrome. DC/TBD, dyskeratosis congenita/telomere biology disorders; DEB, diepoxybutane; MMC, mitomycin C; OMIM, Online Mendelian Inheritance in Man. (3, 7, 118, 123–130).

Otology, and Short stature) (135). Overall, the most frequent anomalies are as follows: short stature, upper limb structural abnormalities, pigmentary skin changes, renal malformations, and microcephaly (37). Both acronyms can trigger an early suspicion of the disease, even before the onset of the hematooncological phenotype (136).

3.1 Neurological findings

Retrospective studies have shown that nervous system abnormalities are frequently found in both FA and DC/TBD (37, 117, 133, 137, 138). Structural neurological anomalies were reported in 40% of patients with FA, whereas 10–50% of patients with DC/TBD had these findings (37, 117, 133, 137, 138).

3.1.1 Structural abnormalities

Case reports of patients with FA have described a wide range of neurological anomalies such as microcephaly, cerebral/ cerebellum hypoplasia, small or abnormal pituitary gland, abnormalities of the corpus callosum, malformations of posterior fossa, hydrocephalus, and malignancies (36, 139– 143). The identification of central nervous system (CNS) abnormalities increases when brain imaging is used. Brain magnetic resonance imaging is important for the thorough description of patients with FA, since almost 60% of patients have abnormal imaging (144, 145). When comparing brain images of patients with FA and age and sex-matched control population without intracranial pathology, it was found that 61% had at least one CNS or skull-based abnormality (144, 145). The most frequent structural CNS anomalies are localized in the midline, such as pituitary hypoplasia (46%), and in posterior fossa consisting of cysts, vermis, and ponto-cerbellar hypoplasia (32%) (144, 145). However, when the imaging studies were performed in patients with either neurological or endocrine manifestations, CNS abnormalities were found in 90% (146). In this group, the most frequent structural anomalies were posterior fossa abnormalities that required neurosurgery (146).

Genotype–phenotype association has been evaluated in patients with FA regarding CNS anomalies, microcephaly, and hydrocephalus. Structural CNS anomalies have been associated with PVs in ID2 complex genes, microcephaly with PVs in *FANCD1*, *FANCD2*, *FANCJ*, ID2 complex, downstream genes, and hydrocephalus with *FANCB* genotype (37).

The frequency of structural abnormalities in DC/TBD as discovered by systematic evaluation through brain magnetic resonance imaging is 57% (147). The brain structure that is predominantly affected is the cerebellum; the more frequent structural abnormality is cerebellum hypoplasia or atrophy, seen in 39% of patients with DC/TBD. Other frequent findings are cerebral atrophy (18%) and midline alterations such as corpus callosum abnormalities (16%) and small pons (11%) (147). The number of neuroimaging abnormalities has been found to be correlated with shorter telomeres, XL and AR inheritance patterns, and severe BMF (147). The same analysis for patients with DC/TBD showed much higher frequencies than in general population of *corpus callosum* anomalies, prominent *cisterna magna*, white matter lesions, and cavum *septum pellucidum*, and *cavum vergae* variants (147).

3.1.2 Functional abnormalities

The information concerning functional neurological abnormalities in FA is scarce; a systematic approach to describe the functional neurologic phenotype in FA is missing. The available information comes from transversal case series analysis in which 32.5% of functional neurological involvement has been estimated. The identified abnormalities include intellectual disability, epilepsy, attention deficit hyperactivity disorder, and migraine (145, 146).

Any functional neurological disorder in DC/TBD has been described in around 50% of patients (147). Neuromotor findings include abnormal gait, dysarthria, incoordination, and ataxia (147). These types of alterations have been associated with shorter telomeres, HHS or RS, XL or AR inheritance patterns, and severe BMF (147). Some psychiatric manifestations have also been found in patients with DC/TBD (depression, anxiety or panic attacks, autism, and bipolar disorders) (147).

Most persons with DC/TBD have no significant developmental delay or intellectual disability, but the severe phenotypes HHS and RS do. HHS neurological phenotype is associated with cerebellum hypoplasia, microcephaly, and neurodevelopmental disorder (119–121), whereas RS includes bilateral exudative retinopathy, intracranial calcifications, and neurodevelopmental disorder (3, 122).

3.2 Ocular findings

Retrospective analyses have described that 11–20% of patients with FA have ophthalmological abnormalities (37, 133). Published cases have reported small eyes (148–150), strabismus (151, 152), epicanthal folds (153), ptosis, cataracts (154, 155), glaucoma (154, 156, 157), myopia (152), astigmatism, and retinopathy (154, 155, 158–160). Ocular alterations are also frequently described in DC/TBD (133), but their nature is different from what is seen in FA.

3.2.1 Structural abnormalities

Intentional review of ocular characteristics identifies at least one abnormally small dimension of the eye structure in 95% of patients with FA (151). Cross-sectional studies with systematic evaluation have found microcornea in 46-100%, microphthalmia in 69-100%, small palpebral fissures in 75-94%, ptosis in 28-88%, hypotelorism in 25-43%, cataracts in 14-30%, limbal neovascularization in 26-28%, epicanthal folds in 0-9%, strabismus in 7.5%, and posterior embryotoxon in 5% of assessed patients (151, 161, 162). Compared with age- and gender-matched normal values, patients with FA have abnormally diminished: interpupillary distance, inner canthal distance, outer canthal distance, palpebral fissure length, palpebral fissure width, and corneal diameter (151, 161, 162). The interpupillary distance, outer canthal distance, and corneal diameter are smaller compared with other IBMFS (151). Specifically, the palpebral fissure length was significantly smaller than in DC/TBD (151). This is an interesting finding, since the short palpebral fissure is considered a part of the eye involvement in the PHENOS acronym (135) and seems to be specific of FA. The limbal neovascularization and cataract were more common in patients who underwent hematopoietic cell transplantation (HCT) (161). An association between small eyes and a *FANCD2* genotype has been found by a single publication (37).

Optic nerve hypoplasia, characterized by a decreased number of optic nerve axons (18), can manifest as small disc and cup areas in patients with FA (161, 162). The identification of nerve hypoplasia during eye fundus examination should prompt neuroimaging studies, since this ophthalmologic feature may be associated with midline cerebral structural defects, such as the absence of the *septum pellucidum*, agenesis of the *corpus callosum*, cerebral hemisphere abnormalities, or pituitary gland anomalies (163).

Around 30% of patients with DC/TBD have eye anomalies according to retrospective studies (116). Ocular size is not a main feature, only small outer canthal distance and palpebral fissure length have been reported (151). The most common findings in DC/TBD are nasolacrimal duct obstruction in 29% (164–166), retinopathy in 21%, trichiasis (165, 167), and entropion in 7% (151, 167). The spectrum of ocular anomalies also includes conjunctivitis (167), ectropion (168), corneal scarring (167), cataracts (169, 170), and retinal vasculopathy (171–174).

3.2.2 Functional abnormalities

Regarding visual functions, 88% of evaluated patients with FA have abnormalities including motion problems (162). No systematic evaluation of functional alterations in DC/TBD have been performed.

Considering the high frequency of ocular abnormalities both in FA and DC/TBD, integral ophthalmic assessment at the time of diagnosis is important to procure patients with the benefit of prompt and specific interventions. Due to their frequencies, small ocular parameters should bring to mind an FA diagnosis and nasolacrimal duct obstruction a DC/TBD one.

An important consideration is that, besides ocular manifestations that are part of the primary diagnosis, patients with FA and DC/TBD are subject to receive treatments that affect the eye. In particular, steroids and HCT regimens are associated with an increased risk of glaucoma, cataract, corneal ulcers, cytomegalovirus retinitis, fungal endophthalmitis, and retinopathy (175, 176). A higher prevalence of these findings has been found in patients after HCT (162). Regular ophthalmological follow-up is important after HCT for early intervention to avoid complications.

3.3 Otological findings

The prevalence of otological problems in patients with FA varies from 10% to 20% according to literature reviews (1, 2).

Hearing impairments are more frequent in FA compared with other IBMFS (177).

3.3.1 Structural abnormalities

Structural ear anomalies of any kind have been found in more than half of patients after an intentional evaluation (177). As it appears to be the rule in FA, the structural abnormality range is wide. The following alterations have been described: small tympanic membrane in 62%, malformed malleus in 33-57%, bony islands at tympanic membrane in 48%, pinna malformations (including microtia) in 10-35%, abnormal course or prominence of the chorda tympani nerve in 34%, narrow external auditory canal in 32%, narrow internal auditory canal in 6%, and atresia of the external auditory canal in 3% (177, 178).

In sharp contrast, in DC/TBD structural, ear abnormalities have not been reported despite the intentional study of otologic features has been done (177).

3.3.2 Hearing impairment

Hearing loss has been found in 36–55% patients with FA, with a high proportion of bilateral and asymmetric impairment (3-6). The most common type of hearing loss in FA is the conductive type, followed by sensorineural and, last, the mixed pattern (3-5). This translates into mild and moderate degrees of hearing impairment (177-180), particularly decreased hearing has been associated with the presence of structural ear anomalies (3). Assessment through the speech perception in noise test showed that up to 53% of evaluated patients had hearing impairment (178).

An analysis that sought associations of hearing loss with physical anomalies in FA found that 100% of patients with radial bone malformation had some hearing impairment; this association was statistically significant compared with patients without radius anomaly (177), which led to speculate that a developmental relationship between the radius and ear could exist (3). This is why otological findings, particularly associated with radial ray malformations and/or skin pigmentation changes, should prompt an evaluation for FA.

Genotype-phenotype association has been established between structural or functional otological anomalies and *FANCB* and *FANCD2* genotypes (37, 181).

Although hearing loss has not been commonly described in DC/TBD (117, 133, 138), intentional otological evaluation identified that 6% of patients have hearing problems, which is double than in healthy population through epidemiological studies (177, 182). In the general population, a link between telomere attrition and age-related hearing loss has been demonstrated (183). However, another cross-sectional study showed that telomere length was not associated with hearing acuity in healthy children or their midlife parents (184).

The genetic instability associated with FA and the shorter telomere length of DC/TBD have been associated with premature aging, which hypothetically might lead to progressive sensorineural hearing loss (177).

Structural or functional otological findings, or a combination of both, are common in patients with FA, and less frequent but seen as part of the DC/TBD spectrum. Formal hearing assessment, including otolaryngology evaluation, is recommended as part of the management of patients with FA and DC/TBD (177). The identification of otological abnormalities may lead to prompt intervention to reduce language complications through adequate rehabilitation interventions.

3.4 Oral findings

The oral cavity can be easily disregarded during physical examination, yet it can frequently hold key features to integrate a diagnosis. In patients with DC/TBD, some sort of dental alteration was found in 23% (185). On their part, almost half of patients with FA have been reported to have at least one mucosal lesion (186), and alterations in the number and shape of teeth have also been found (187, 188). Moreover, both for FA and DC/TBD, the extreme risk of cancer in this location requires diligent oral inspection (189).

3.4.1 Dental organs

3.4.1.1 Developmental abnormalities

Although developmental abnormalities in dental organs have been reported in both DC/TBD and FA, the spectrums are different. In DC/TBD, the reported anatomic differences are very prevalent as a decreased root/crown ratio has been identified in up to 75% of patients and mild taurodontia in 60% (190). On the other hand, the dental phenotype in FA has a wider spectrum, even though it is less constant: Microdontia has been reported in 44% (187), variations in teeth numbers have been found with dental agenesia identified in over 25% (187, 188), and supernumerary teeth in less than 10% (187).

3.4.1.2 Decay, periodontal disease, and gingivitis

Common oral alterations such as caries and periodontal disease have also been found in patients with DC/TBD and FA (190); it is hard to control the multiple factors associated with these disorders in order to conclusively define if they belong in the phenotype of these uncommon syndromes. Nevertheless, these features have been described in both of these IBMFS. According to the DC registry, 17% of patients have extensive dental caries or loss of dental pieces (117); this has also been found in 35% of patients with FA (187). Gingivitis appears to be an even more prevalent feature in FA; it has been identified in

nearly 50% (188, 191). Dental hygiene and access to stomatologic care are undoubtedly fundamental factors in the development of these type of oral manifestations.

3.4.2 Lesions of the oral mucosa

Lesions of the oral mucosa are frequent in both of these syndromes: Transient blisters and erosion are early manifestations of DC/TBD (192), and mucosal lesions have been found in up to 42% of patients with FA (186). A recent extensive analysis of 279 patients with FA revealed that they have frequent synchronic and metachronic oral lesions (193).

3.4.2.1 Leukoplakia

The WHO's working expert group for potentially malignant oral disorders has conserved the leukoplakia definition as "a predominantly white plaque of questionable risk having excluded (other) known disorders or disorders that carry no increased risk for cancer" (194). This is a cardinal diagnostic feature in DC/ TBD that is part of the classical clinical triad used for diagnosis suspicion; it has been reported in 78% of patients from the DC registry (117) and can be as frequent as 96% in certain literature reviews (185). The location of leukoplakia in DC/TBD patients is mainly found in the dorsal tongue followed by buccal mucosa (190). It has been hypothesized that the leukoplakia seen in patients with DC/TBD are hyperkeratotic scars that result from genetically mediated epithelial atrophy, pigmentation, and scarring (192). Leukoplakia is not exclusive of DC/TBD; after an intentional review of oral manifestations in FA, some authors have gone to hypothesize that this is an underreported feature as it was found in 12% of patients, located predominantly in the palate (186). Meanwhile, in the general population, leukoplakia has been found in less than 5% with a predominant location in the commissure or the buccal mucosa (195).

3.4.2.2 Erythroid lesions

The second most frequent oral lesion in both DC/TBD and FA is erythroplakia (190, 193). In DC/TBD, the erythematous patches are more frequently found in the occlusal plane (190). These lesions, by their own, do not seem to be related to malignancy, but in the largest analysis of oral lesions in patients with FA, it was found that lesions that combined erythroplakia with erosion were highly correlated with a positive brush biopsy for malignancy (193) and are the ones that should be more closely followed up.

3.5 Gastrointestinal findings

The spectrum of gastrointestinal (GI) system alterations found in patients with FA an DC/TBD does not overlap, but they can equally impact the patient's quality of life. In FA, GI alterations are mainly malformations that will usually require surgery (37, 133); meanwhile, in patients with DC/TBD, GI features are not congenital but appear later in life.

3.5.1 Liver abnormalities

The GI organs can be affected in multiple ways in patients with DC/TBD. One of the largest cohorts of patients with DC/ TBD described hepatic manifestations in 40% of patients (196). The range of hepatic involvement includes mild findings, such as hepatomegaly, inflammation, hemosiderosis, and fibrosis, to more severe manifestations such as cirrhosis and portal hypertension (115, 197, 198). Imaging evaluation has showed an increase in hepatic echogenicity in 39%, hepatomegaly in 26%, and nodular contour suggestive of cirrhosis in 25% of patients (196). Severe symptoms are more common in children and have been found in 16% of patients with DC/TBD. A study of 69 individuals from five unrelated families showed intrafamily variability in which penetrance had an important role and phenotype variability with a broad spectrum of liver disorders that included both inflammatory and fibrotic findings (197). Liver involvement and/or isolated liver fibrosis can also be a sole manifestation in patients with DC/TBD, a presentation more commonly associated with PVs in TERT rather than TERC or TINF2 (196, 199). Although the specific pathophysiology has not been identified, some reports associate liver abnormalities with vascular anomalies (200).

Liver alterations in patients with FA are usually secondary to androgen therapy for BMF; they consist of hepatomegaly, increase in hepatic echogenicity, biliary duct dilation, and elevated liver enzymes (201). However, liver alterations have been described in 5/44 patients with FA in the absence of androgen therapy or another risk factors. The liver alterations found were very severe chronic cytolysis pattern, significant biological liver abnormalities, minor cytolysis pattern, hepatomegaly, and adenomas (201).

3.5.2 Digestive tract abnormalities

The tubular part of the GI can also be affected in DC/TBD; esophageal stenosis has been identified in 10% of the patients; other alterations such as Schatzki's ring, atrophic gastritis, esophageal intraepithelial lymphocytosis, inflammatory changes, and mild fibrosis of gastric lamina propria have been reported in lower frequencies (202). Although small and large bowels are not commonly affected, there are reports of pancolitis, epithelial sloughing, and increased apoptosis. Upper tract GI hemorrhage is an unusual manifestation that can be severe and deadly if not treated promptly. A multicenter review reported 16 patients with DC/TBD who presented with recurrent GI hemorrhage unresponsive to conventional management (203). Although no specific cause for the GI hemorrhage was identified, a higher risk for it has been established for patients with the presence of telangiectatic superficial vessels in the stomach and small intestine (203). PVs in the gene *TINF2* were more frequently detected in patients who had GI hemorrhage (203). Patients with PVs in this gene were overrepresented among patients who had pulmonary arteriovenous malformations (PAVMs) (204), which could suggest that a similar vascular lesion might be found in their GI tracts. Other nonspecific GI symptoms reported in patients with DC/TBD include early satiety, nausea, abdominal pain, solid dysphagia, failure to thrive, and watery or bloody diarrhea. Besides, DC/TBD, very short telomere length has been evaluated as a biomarker for hepatic involvement. Around 70% of the subjects with hepatic involvement have shorter telomeres (196).

In FA, anal and esophageal malformations, the GI anomalies included in the VACTERL-H association have a fairly low frequency of 5% (133, 205). According to Alter and Rosenberg's literature review, of those patients with FA who have more than three VACTERL-H features, the Anal, Renal, Limb (ARL) combination is the third most frequent, and, overall, 34% of patients with FA and VACTERL-H have a GI malformation. The combination of both anal and esophageal malformations in a single patient with FA and VACTERL-H is infrequent as only 3% will have both (206). Other GI malformations reported in FA are atresia of jejunum, annular pancreas, exocrine pancreatic insufficiency, and intestinal malrotation (207). GI problems as a single feature of FA is rare. There are two case reports of patients diagnosed with FA after the recognition of esophageal atresia type III in the neonatal period (208). Regarding genotype-phenotype association, PVs in FANCB have been associated with anal anomalies, tracheo-esophageal fistula, and esophageal/duodenal atresia whereas FANCN only with anal malformations (37).

GI symptoms in patients with FA or TDC/BD with or without history of HCT or androgen therapy deserve evaluation. This could prevent severe complications such as GI bleeding, cancer, or liver dysfunction. An integral GI evaluation that includes radiological and/or biochemical studies is required in patients with DC/TBD in the interest of diagnosing any of the described liver anomalies (196, 209).

3.6 Cardiopulmonary findings

Both FA and DC/TBD have cardiopulmonary manifestations, yet each condition has preferent trophism for an organ. The heart is frequently affected in FA, and the lungs are usually spared whereas, in DC/TBD, the opposite is seen. Moreover, the type and pathophysiologic mechanisms leading to these features are particular to each condition.

3.6.1 Cardiac abnormalities

The frequency of cardiovascular manifestations in patients with FA differs according to the type of study that reports it; it ranges from 13% according to a sound literature review to up to 55% if a systematic heart evaluation is performed (37, 206). The discrepancy on the reported data could be attributed to the fact that some cardiovascular anomalies are asymptomatic and consequently underdiagnosed. Cardiac structural abnormalities are frequently the third feature for a VACTERL-H diagnosis in patients with FA (37). The CRL (cardiac, renal, and limb) combination, with or without other features, was found in 54% of patients with FA (59/108) (37). There is a wide spectrum of structural heart defects reported in FA, mostly diagnosed at birth (37, 206). The most common cardiac defects documented are septal defects, but others such patent ductus arteriosus, coarctation, situs inversus, truncus arteriosus, bicuspid aorta, and a left persistent superior vena cava have also been reported (207, 210). An association between cardiac defects and a *FANCB* and *FANCI* genotypes has been found by a single publication (37).

In sharp contrast, only 1–4% of patients with DC/TBD are born with a congenital heart defect; the spectrum essentially consists of ventricular and atrial septal defects and dilated cardiomyopathy (103, 198). Congenital heart defects are one of the most common malformations in the population, presenting in around 1% of the population; it is not clear if DC/TBD have an intrinsic increased risk for congenital heart anomalies or if those who present them have a concurring disease of multifactorial etiology (211, 212).

3.6.2 Pulmonary abnormalities

Pulmonary fibrosis is the best known complication in patients with DC/TBD, with a prevalence of 20% and is usually diagnosed beginning at the fourth decade of life (138, 213, 214). The characteristic radiological findings are diffuse interstitial markings (215). A cohort of 72 patients with idiopathic pulmonary fibrosis showed that this anomaly can occur as the first manifestation in patients with short telomeres and even in the absence of hematological findings (216). In a systematic review of patients with DC/TBD and pulmonary fibrosis, the early onset of this alteration (before 40 years of age) was associated with more severe and earlier onset of BMF (214). The same analysis found an association between PVs in TINF2 gene and an early onset pulmonary fibrosis and BMF (214). Other pulmonary anomalies associated with DC/TBD are typical and atypical interstitial pneumonia, bronchiectasis, cysts, bronchiolitis obliterans, chronic hypersensitivity pneumonitis, and emphysema, which have been reported with a lower frequency in two small cohorts of patients with DC/TBD (217, 218).

Pulmonary disease is the second most common cause of death after BMF in patients with DC/TBD (213). Yet, in patients with PVs in genes with an AD pattern of inheritance (38.7%), pulmonary complications are the main cause of mortality (198). PAVMs have been reported in 3% of patients with classic DC (204, 219). A study of 13 unrelated patients with DC/TBD showed a median age of 15 years old for PAVMs diagnosis;

46% (6/13) of them had germline PVs in *TINF2* (204). The observed and reported frequencies are low, but this could result from failure to recognize PAVMs in virtue of shared symptoms with pulmonary fibrosis (103, 204). Further investigation is needed to establish the etiology of PAVMs in order to determine whether they are a consequence of telomere dysfunction or if they are associated with HCT, since they are a common finding in patients who undergo this procedure (198, 204). It is interesting that the phenotype of patients with DC/TBD due to PVs in *TINF2* has important vascular features such as PAVMs, GI hemorrhage, and in the exudative retinopathy found in RS.

In FA pulmonary abnormalities, either malformations or auto-immune process have been reported as coincidental findings; these are not fibrotic changes like the ones seen in DC/TBD. There are reports of two patients with FA who had a pulmonary arterio-venous fistula and single reports of pulmonary alveolar proteinosis, pulmonary glial heterotopia, and interstitial lung disease (220–223). Nevertheless, further investigation is needed to understand whether these pulmonary manifestations could be the direct consequence or FA and/or treatment for its complications.

Upon diagnosis, a complete cardiovascular examination in patients with FA is warranted due to the high incidence of structural heart defects; no further follow-up is needed if malformations are not detected. Meanwhile, pulmonary findings in patients with DC/TBD are only expected in symptomatic patients who will need follow-up with pulmonary function test and/or computed tomography scan to monitor disease progression.

3.7 Genitourinary findings

3.7.1 Kidney and urinary tract abnormalities

The location of urinary anomalies is different between patients with DC/TBD and FA. Patients with FA usually have malformations of the upper urinary tract (224, 225), whereas fibrotic alterations of the lower urinary tract are found in patients with DC/TBD.

Alterations in the urinary system are seen in 22–50% of patients with FA (37, 207, 224, 226, 227). The combination of renal plus limb abnormalities (RL) can be seen in up to 20% of patients with FA, in those who happen to also have a VACTERL-H diagnosis, the frequency goes up to 94% (1, 6). This highlights the importance of renal evaluation when FA is suspected. The spectrum of urinary anomalies is heterogenous; it includes renal hypoplasia or aplasia, dysplastic or ectopic kidneys, crossed fused ectopia of kidneys, double ureters, cysts, and horseshoe kidney (224, 228). These renal anomalies can frequently be accompanied by urinary tract infections, which are key indicators of a renal malformation particularly if present frequently and in early childhood (224). Hydronephrosis of

unknown etiology has been reported in three cases with FA (36, 229, 230).

In a comprehensive literature review that included 380 probands, patients with null genotypes, irrespective of the gene and those with PVs in *FANCB* and *FANCJ*, had a positive association with renal malformations (37).

Urinary tract anomalies in patients with DC/TBD most commonly manifest in the lower tract and are diagnosed later in life once associated symptoms develop (117). Urethral stricture or stenosis is reported in at least 5% of patients, but it can increase up to 10% in men (198). Bleeding or acute urinary retention syndrome is a sign consistent with the suspicion of a urethral stenosis (231). Although most urinary anomalies in patients with DC/TBD are age dependent, there are case reports of patients with malformations such as renal agenesis (232), horseshoe kidney, bilateral renal cysts, and duplication of renal collecting system and ureters (198). Renal structural abnormalities are common congenital malformations in the general population that occur in one of 1,000 births; this apparently isolated renal malformations reports might be coincidental findings (21).

3.7.2 Genitalia abnormalities

Genital abnormalities are not as common in patients with FA. Men present genital anomalies in 12–16% (37, 227) with a lower frequency in women, but this could be due to an underdiagnosis since the internal nature of the female tract requires imaging or specialized maneuvers for a thorough evaluation (37). Male patients can have poor genital development with micro-penis, hypospadias, and cryptorchidism or even major malformations such as complete transposition of penis and scrotum (207, 226). Uterine malformations such as bicornate uterus and labial fusion have also been reported (207, 233).

Reported genitalia manifestations in DC/TBD are phimosis, hypospadias, penile leukoplakia as well as undescended testes and in fewer cases hypogonadism (103, 198, 234). Women have also presented vaginal atrophy and leukoplakia (103).

Although the genitourinary system can be affected on both patients with FA and DC/TBD, the major difference is in the part of the genitourinary system that is affected. The upper urinary tract corresponds to FA, and the lower urinary tract corresponds to DC/TBD. In addition, the time of diagnosis differs, because the anomalies associated with DC/TBD tend to be progressive and identified later in life, whereas those associated with FA are congenital malformations and could be detectable from birth.

3.8 Dermatological findings

Dermatologic findings are the cornerstone of the classic DC triad. Reticulate hyperpigmentation and nail dysplasia are individually present in nearly 90% of patients with a DC/TBD

(116). The dermatological phenotype may not be present at the time of diagnosis but, as many DC/TBD features, insidiously develops or becomes more pronounced over time. The skin pigmentation usually appears between the ages of 5 and 10 years, and nail dystrophia can appear in the first months of life, although is usually present toward adolescence (235). Skin pigmentation changes are most commonly found in sunexposed areas (neck, trunk, and both extremities); they consist of reticulate hyperpigmentation, which can be associated with hypopigmentation, atrophy, poikiloderma, and telangiectasias (116-118). Nails appear dystrophic with longitudinal ridging and are usually more severely affected in the hands than the feet (236, 237). Other cutaneous manifestations include hair involvement, such as premature hair graving or loss and even alopecia, palmoplantar hyperkeratosis, and eczema (213, 235-242).

Skin findings in FA are so frequent that they have a place in the PHENOS acronym. Retrospective studies have found skin anomalies in around 40% of patients (37, 133). The most common alterations are pigmentary changes such as café-aulait macules, generalized hyperpigmentation, and hypopigmented macules (37, 133). Cross-sectional studies that intentionally looked for skin changes in patients with FA have found at least one pigmentary anomaly in 82-97% of evaluated patients (243-245). The most frequent finding was café-au-lait macules, with a mean of 7.5 per patient; they were distributed across the whole body (243). Hypopigmented macules, seen in 46% of patients, were the second most common anomaly (243). Recently, freckle-like macules were noted in 24.7%; they were characteristically located at flexing areas that included lateral neck, popliteal fossa, and axillary, inguinal, and suprapubic creases (243). Other less common findings were confluent flexural hyperpigmentation and diffuse macules (243). The combination of hypopigmented along with hyperpigmented macules was the most frequently identified at 45.2% (243, 244). No associations were found between skin findings and radiation exposure, cutaneous graft-versus-host disease, or HCT. However, increasing age was associated with an increase in the odds of freckle-like macules (243). Sweet syndrome has been recurrently described in patients with FA (246-249). This neutrophilic dermatosis characterized by painful violaceous plaques and nodules with dermal neutrophilic infiltrate can occur as a paraneoplastic syndrome (250); therefore, in patients with FA, it should alert of the possible development of an oncological process. According to a comprehensive literature review, a higher proportion of patients with PVs in FANCD1 and FANCD2 had at least one skin anomaly compared with all other genes (37).

Patients with FA or DC/TBD who undergo an HCT can have long-term skin findings (251); however, these anomalies are different from those due to the disease.

Awareness of the high frequency of dermatological abnormalities presentation in patients with FA or DC/TBD is

essential to increase diagnostic suspicion and favor early diagnoses.

3.9 Skeletal findings

These anomalies represent an area of sharp contrast between FA and DC/TBD. Skeletal abnormalities are found in over half of patients with FA, whereas these alterations are only incidentally described in DC/TBD (3, 118).

3.9.1 Upper limb abnormalities

The characteristic upper limb malformations linked to FA are radial ray anomalies (radius, thumb, and/or thenar alterations), which are part of the VACTERL-H acronym (135, 206, 252–255). Classically, FA has been associated with severe and, sometimes, bilateral upper limbs anomalies (253), which is why a FA diagnosis may be overlooked in patients with mild anomalies, such a thenar hypoplasia.

Around 50% of cases with FA have some kind of hand or arm anomalies (133, 256), but 40% are described as radial ray anomalies (37). Retrospective analyses have found the following radial ray malformations: thumb hypoplasia in 32%, preaxial polydactyly (including triphalangeal thumb) in 19%, thumb aplasia in 12%, absent or hypoplastic radii in 8-16%, and hypoplastic thenar eminence in 3% (256). All cases with an abnormality of the radius had a concomitant thumb anomaly (37, 257, 258), which is contrasting with thrombocytopenia and absent radii syndrome where thumb not severely affected (139). Upper limb alterations were more frequently mild and bilateral than those severe and unilateral (253, 257, 258). Actually, bilateral hand anomalies had a 4.6 odds ratio for positive DEB test result when contrasted to unilateral involvement (258). Other type of hand anomalies including clinodactyly, brachymesophalanges, and brachydactyly have been reported (37, 256).

The relevance of radial ray abnormalities as a pivot sign for suspicion of FA is indisputable; children with a limb anomaly were diagnosed two years younger compared with those without, and the severity of the thumb hypoplasia was inversely associated with the age at diagnosis (257). When a patient with radial ray alteration is being evaluated, clinicians should consider that the more severe the radial ray defect is, the more possible a syndromic condition is (253, 259); also, it must be kept in mind that patients with FA may have very discreet upper limb alterations that can be missed by an inexperienced appraisal.

A study determined that incidence of FA among children with congenital thumb anomalies was 7% (258). The same analysis showed that patients diagnosed with FA represent 1% of the patients with any type of congenital thumb anomalies and 3% of those with thumb hypoplasia (258). Most patients tested for FA had other physical abnormalities, and all of those who had a positive FA test result had café-au-lait macules (258).

3.9.2 Vertebrae and other skeletal abnormalities

The spectrum of these type of anomalies includes hemivertebrae, hypoplastic vertebrae, vertebral fusion (Klippel-Feil syndrome), spina bifida, coccygeal aplasia, scoliosis, kyphosis, and atlantoaxial instability (37, 133, 260). A recent prospective study published that vertebrae anomalies are found in almost 50% of patients after intentional evaluation (245), which is in sharp contrast with the 4% described in the literature (37, 133). Some reports have depicted less frequent anomalies, such as rib hypoplasia, Sprengel deformity, and hip dysplasia (149, 225, 261–264).

In patients with FA and VACTERL-H, 27% had the VL combination that includes vertebral and upper limb alterations (206). Orthopedic surgeons should be attentive, as they may come across patients with FA in their everyday practice. They are in an advantageous position to identify patients with FA before the onset of hematologic manifestations, which is essential for an appropriate treatment and surveillance.

Upper limb malformations were more frequent in patients with PVs in *FANCB* and *FANCD2* (37). *FANCD2* genotype was also associated with lower limb anomalies, specifically hip involvement, whereas *FANCB* was associated with vertebral defects (37, 149).

3.10 Growth and endocrine findings

The endocrine system has not been studied in patients with DC/TBD; although there are some anecdotal cases in the literature, hormone deficiencies seem uncommon in these patients (3). In contrast, almost 80% of patients with FA have been shown to have an endocrine abnormality; this striking frequency of endocrine affection has been proposed to be the consequence of affected secretory cells due to DNA damage caused by excessive reactive oxygen species (265). HCT, a fundamental therapy for BMF both in patients with FA and DC/TBD can contribute in endocrine conditions such as diabetes, dyslipidemia, hypogonadism, thyroid, adrenal, and pituitary dysfunction (266).

3.10.1 Short stature, intrauterine growth retardation, and microcephaly

Comprehensive knowledge of the clinical presentation of DC/TBD comes from analysis of features from patients who participate in the DC registry (117). Data from 118 male patients with a clinical diagnosis of DC/TBD showed that short stature is part of the phenotype in almost 20% of patients, whereas intrauterine growth retardation and microcephaly was documented in just over 7% and 6%, respectively (117). Short stature does not appear to be due to growth hormone deficiency, since there is a single report of a child with growth hormone

deficiency in whom brain magnetic resonance imaging was normal (267). Analysis of other cohorts has confirmed the frequency of short stature but found lower frequencies of IUGR and microcephaly (185). Precisely, these two features are found in over 96% of patients attained by the severe variant of DC/TBD known as HHS (268).

Meanwhile, growth deficiency is such an important piece of the FA phenotype that two features have been integrated into the PHENOS acronym (P: skin Pigmentation; H: small Head; E: small Eyes, N: Nervous system, O: Otology, S: Short stature), which recapitulates some of the physical manifestations of FA (37). The data from different cohorts have documented short stature in 46–60% of patients with FA (265, 269, 270), whereas microcephaly has been documented in 33% (270). This phenotype has been further delineated to specify that about half of patients are small for gestational age and that only 25% of them will reach normal stature (271). Moreover, in FA, concomitant endocrinopathies are frequent. The combination of growth hormone deficiency (reported in 12% of patients with FA) and hypothyroidism will result in shorter stature in these subjects (265).

According to a genotype-phenotype association analysis, short stature was more frequent in patients with null PVs and those with PVs in downstream genes, whereas microcephaly was associated with null genotypes irrespective of the gene and patients with PVs in *FANCD1*, *FANCD2*, *FANCJ*, ID2 complex, and downstream genes (37).

3.10.2 Other endocrine abnormalities

Thyroid function is frequently compromised in patients with FA; hypothyroidism was found in up to 61% of children and 37% of adults (265). Primary hypothyroidism is predominant, although some patients have central hypothyroidism secondary to hypothalamic dysfunction (271). Meanwhile, in DC/TBD, hypothyroidism has only been reported in two isolated cases (272, 273).

Glucose metabolism has also been found to be disturbed in patients with FA: Impaired fasting glucose or glucose intolerance was documented in 24% of patients whereas diabetes mellitus in 10% (269) and insulin resistance in over 40% (269, 270). Meanwhile, in DC/TBD, such metabolic alterations do not appear to be frequent (274). This finding is intriguing, because several cross-sectional studies have shown that patients with type 2 diabetes mellitus had telomeres significantly shorter than control subjects (275-277). These studies have also found a significant inverse relationship between oxidative damage and telomere length (275–277). In mice with short telomeres, the β -cell function, insulin secretion, and glucose metabolism are impaired recapitulating the early stages of human type 2 diabetes mellitus (278). Nevertheless, to our knowledge, this has not been systematically evaluated in patients with DC/TBD.

3.11 Fertility and pregnancy findings

There are fertility issues in both DC/TBD and FA, although, like endocrine abnormalities, they are more frequent in patients with FA. According to the DC registry, hypogonadism is found in approximately 6% of patients (117). Meanwhile, this is a more pervasive feature in FA where both men and women are similarly affected: Hypogenitalism characterized by small testes and phallus has been reported in 64% of male patients with FA, and over 75% of female patients have premature ovarian failure (271).

Ovarian reserve of patients with FA has been intentionally explored by measuring anti-Mullerian hormone (AMH) (279). Ovarian compromise appears to be a component of the FA phenotype; it is present from prepuberal stages and is not a complication derived from HCT, since both transplanted and non-transplanted patients show low AMH levels (279, 280). This finding has also been replicated in DC/TBD patients (median AMH level 0.55 ng/ml), but AMH levels were not as low as what was found in patients with FA (median 0.05 ng/ml) (14, 15). These low AMH levels are in accordance with expected fertility issues.

Pregnancies in patients with FA are rare; less than 50 pregnancies, regardless of transplant status, have been reported in the literature (4). Reproductive complications in women with FA include decreased fertility, preeclampsia, increased premature births and C-sections, and declined maternal hematopoiesis during pregnancy (281). Reproductive and pregnancy outcomes in women with DC/TBD have recently been explored in a group of 26 women older than 16 years who had a total of 80 pregnancies (282). Reproductive complications in this group were similar to those reported in patients with FA, with decreased fertility as 23% of patients required some sort of fertility treatment, a high rate of fertility loss and recurrent pregnancy loss, as well as more frequent preeclampsia, preterm births and a higher C-section delivery rate. Moreover, hematological abnormalities during pregnancy or delivery occurred in a quarter of pregnancies (282).

3.12 Immunologic findings

FA and DC/TBD share the common defining feature of BMF. In most patients with the classical presentation of either condition, the impairment of the white blood cell lineage translates as subtle immunologic abnormalities. However, there are seldom cases, particularly those belonging to the HHS subtype of DC/TBD, for whom severe immune manifestations are the rule.

There are a few studies that describe the immune phenotype of patients with FA. In a large cohort of patients with FA with advanced BMF, immunologic data showed hypogammaglobulinemia, reduced absolute lymphocyte counts, and increased serum TGF- β and Il-6 (283). The National Cancer Institute (NCI) IBMFS cohort looked at the data from patients belonging to the whole spectrum of severity of the BMF; this well-controlled study corroborated that the immunologic phenotype of patients with FA consisted of decreased immunoglobulins and absolute lymphocyte counts as well as detailed that B and NK deficiencies occur at a younger age whereas T-cell deficiencies later in life (284). Moreover, by meticulously comparing the data from patients and their unaffected relatives, the study revealed that abnormal immune functions in FA are more prevalent in adults than in children (284). Although cytokine dysregulation with over production of TNF- α and IFN- γ have been proposed to contribute to an altered hematopoiesis in patients with FA (285, 286), these finding were not corroborated in the NCI cohort (284).

Regarding DC/TBD, the clinical presentation of the immune phenotype is wide. It is a cornerstone of HHS, in which severe combined immunodeficiency attains all patients (287). In the NCI cohort study of immunological features of patients with DC/TBD, the absolute lymphocyte counts and their subset numbers were reduced in children and in the lower range for adults; IgA tended to be increased in adults but no other alteration of immunoglobulins was documented (284). In contrast to what was seen in FA, the immune deficiency was more important in children than in adults; this difference may be due to the fact that one-third of DC/TBD patients in this study had an HHS phenotype, which presents in pediatric age (284). Although severe combined immunodeficiency is a feature of HHS, not all patients with DC/TBD will have such a severe immune outcome; common variable immunodeficiency has also been reported as the initial presentation in DC/TBD (288).

3.13 Oncological findings

3.13.1 FA and DC/TBD as cancer predisposition syndromes

Cancer predisposition genes are defined as those genes where germline PVs and likely pathogenic variants (LPVs) confer high or moderate risks of cancer (greater than twofold relative risks) (289). FA and DC/TBD are known to be cancer predisposition syndromes. The cumulative incidence of cancer is close to 20% by age 65 years in both diseases (189, 219, 290). An important consequence of the hematological management improvement is that cancer has become a frequent cause of death in patients with FA and DC/TBD. The shared and contrasting oncological phenotype for both diseases is summarized in Figure 7.

3.13.2 Cancer in FA

Patients with FA have a much greater risk of developing specific types of cancer than general population. FA confers a



700-fold higher risk of developing acute myeloid leukemia (AML) and head and neck squamous cell carcinoma (HNSCC) compared with non-FA population (291, 292). Moreover, the median age of diagnosis of these neoplasms is significantly younger than that of sporadic cancer (293). Patients with FA are also highly susceptible to squamous cell cancer (SCC) in other locations, such as cervix, anus, and vulva (294). Patients with FA have a 3,000-fold higher risk of vulvar SCC than the general population. In addition, SCCs in FA are usually more aggressive and often diagnosed in more advanced stages of the disease (290).

Patients with biallelic PVs in FANCD1/BRCA2 and FANCN/ PALB2 have very high risks of malignancies, and they can develop embryonal tumors which are not seen in patients with FA who have PVs in other FANC genes. The embryonal cancers reported in these patients are as follows: Wilms tumor, neuroblastoma, and brain tumors (most often medulloblastomas) (262). A recent analysis of 71 patients with PVs in FANCD1/BRCA2 and 16 patients with FANCN/PALB2 genotype confirmed this association (295). This publication also showed that patients with these genotypes can have multiple cancers and an earlier age of the tumor presentation (295). Physicians treating patients with cancer should be alert of the *FANCD1/BRCA2* and *FANCN/PALB2* associated malignancies and consider the possibility of FA diagnosis. Wilms tumor, neuroblastoma, and brain cancer could appear before the hematological manifestations. Importantly, the combination of these embryonal tumors, or the association with AML, is a strong indicator for FA diagnosis. Patients with biallelic PVs in *FANCD1/BRCA2* or *FANCN/PALB2* should have cancer surveillance at very young age, particularly focused on the associated cancers.

3.13.2.1 Heterozygous PVs in FA and the development of cancer

A major concern is whether the risk of cancer is increased in heterozygous individuals. Five *FANC* genes are well-established cancer predisposition genes when they are in heterozygosis. These genes are as follows: *FANCD1/BRCA2*, *FANCJ/BRIP1*, *FANCN/PALB2*, *FANCO/RAD51C*, and *FANCS/BRCA1* (7).

It is unclear whether heterozygous PVs of the other *FANC* genes can confer cancer predisposition. McReynolds et al., and Berwick et al., found that the heterozygous of PVs in other *FANC* genes do not have an increased risk of cancer (296, 297).

FANC gene	Cancer risk in heterozygotes	Risk of breast cancer	Risk of ovarian cancer	Other types of cancer	References
FANCA	Possible	Insufficient data Strength of evidence: limited	Unknown	Unknown	(298, 299)
FANCB	Unknown	Unknown	Unknown	Unknown	
FANCC	Possible	Insufficient data Strength of evidence: limited	Unknown	Insufficient data: early-onset pancreatic cancer Strength of evidence: limited	(7, 297)
FANCD1/ BRCA2	Yes	Absolute risk: >60% Strength of evidence: Very strong (with predisposition to ER+ disease)	Absolute risk: 13–29% Strength of evidence: Very strong	Pancreatic cancer Absolute risk: 5–10% Strength of evidence: Very strong Prostate cancer and melanoma Absolute risk: Unknown or insufficient evidence	(300)
FANCD2	Possible	Unknown	Unknown	Insufficient data: T cell acute lymphoblastic leukemia, testicular seminoma, HNSCC Strength of evidence: limited	(293, 301)
FANCE	Possible	Unknown	Unknown	Insufficient data: HNSCC Strength of evidence: limited	(293)
FANCF	Possible	Unknown	Unknown	Insufficient data: HNSCC Strength of evidence: limited	(302)
FANCG/ XRCC9	Possible	Unknown	Unknown	Insufficient data: early-onset pancreatic cancer, HNSCC Strength of evidence: limited	(7, 302, 303)
FANCI	Possible	Unknown	Unknown	Insufficient data: uterine serous papillary carcinoma, well-differentiated SCC of tongue Strength of evidence: limited	(304)
FANCJ/ BRIP1	Yes	Absolute risk: Insufficient data to define Strength of evidence: Limited; potential increase in female breast cancer (including triple negative)	Variant is estimated to be 5.8–12% though lifetime risk of developing ovarian cancer	Unknown	(305, 306)
FANCL	Possible	Unknown	Unknown	Insufficient data: HNSCC Strength of evidence: limited	(293)
FANCM	Possible	Absolute risk: Insufficient data to define Strength of evidence: strong (with predisposition to triple-negative and early-onset breast cancer)	Unknown	Unknown	(307, 308)
FANCN/ PALB2	Yes	Absolute risk: 41–60% Strength of evidence: Strong (with overrepresentation of triple-negative disease	Absolute risk: 3–5% Strength of evidence: Strong	Pancreatic cancer Absolute risk: 5–10% Strength of evidence: Limited Other cancers Unknown or insufficient evidence	(309)
FANCO/ RAD51C	Yes	Absolute risk: 15–40% Strength of evidence: Strong for ER/PR- negative breast cancer	Unknown	Absolute risk: >10% Strength of evidence: Strong	(310)
FANCP/ SLX4	Unknown	Unknown	Unknown	Unknown	
FANCQ/ ERCC4/XPF	Unknown	Unknown	Unknown	Unknown	
FANCR/ RAD51	Unknown	Absolute risk: 15–40% Strength of evidence: Strong for ER/PR- negative breast cancer	Unknown	Absolute risk: >10% Strength of evidence: Strong	
FANCS/ BRCA1	Yes	Absolute risk: >60% Strength of evidence: Very strong (with predisposition to triple-negative disease)	Absolute risk: 39–58% Strength of evidence: Very strong	Pancreatic cancer Absolute risk: ≤5% Strength of evidence: Strong	(300)

TABLE 3 Heterozygous Pathogenic Variants in FANC Genes and Cancer Development.

(Continued)

FANC gene	Cancer risk in heterozygotes	Risk of breast cancer	Risk of ovarian cancer	Other types of cancer	References
				Prostate cancer: Insufficient evidence	
FANCT/ UBE2T	Possible	Insufficient data Strength of evidence: limited	Unknown	Unknown	(311)
FANCU/ XRCC2	Possible	Insufficient data Strength of evidence: limited	Unknown	Unknown	(304)
FANCV/ MAD2L2/ REV7	Unknown	Unknown	Unknown	Unknown	
FANCW/ RFWD3	Unknown	Unknown	Unknown	Unknown	

TABLE 3 Continued

The strength of evidence categories is based on the NCCN guidelines. Very strong: Prospective cohort studies in a population-based setting have demonstrated risk. Strong: Traditional case-control studies or more than three case-control studies including those with cases ascertained by commercial laboratories or those without controls from the same population. Traditional case-control study: A retrospective study that compares patients with a disease or specific outcome (cases) with patients without the disease or outcome (controls). Limited: Small sample size or case series. None.

ERs, estrogen receptors; HNSCC, head and neck squamous cell carcinoma; NCCN, National Comprehensive Cancer Network; PR, progesterone receptor; SCC, squamous cell carcinoma.

However, some studies suggest that the *FANC* genes could increase the risk of developing cancer in heterozygous individuals. In Table 3, we summarized the evidence related to risk of cancer for each *FANC* gene (7, 293, 297–311).

The detection of PVs in these cancer predisposition genes is important, since genetic counseling and several cancer surveillance measures can be offered to carriers. Chemoprevention and surgeries such as double mastectomy and risk-reduction salpingo-oophorectomy are options (123). Patients with PVs in FANCD1/BRCA2, FANCS/BRCA1, and FANCN/PALB2 are candidates to treatment with poly(ADPribose) polymerase (PARP) inhibitors. One of the more recent studies is the Olympya trial, where patients with HER2-negative early breast cancer and germline PVs or LPVs in FANCD1/ BRCA2 and FANCS/BRCA1 are treated with adjuvant Olaparib (312). Treated group had a significantly longer survival free of invasive or distant disease than placebo (312). PARP inhibitors were demonstrated to be effective in cancers with HR deficiency phenotype beyond PVs in FANCD1/BRCA2 or FANCS/ BRCA1 (313).

3.13.3 Cancer in DC

The most frequent malignancies present in patients with DC/TBD are HNSCC, AML, non-Hodgkin lymphoma, and anal SCC (219).

The first study that quantified the risk of cancer in patients with DC/TBD was the NCI IBMFS prospective cohort. According to this study, after 15 years of follow-up, patients with DC/TBD had an 11-fold higher risk of developing cancer than the general population, specifically, a 195-fold higher risk of leukemia and a 1,154-fold higher risk of tongue cancer (189, 219). Other reported malignancies were esophageal, rectal, endometroid, and cervical cancers. It is difficult to establish a genotype-phenotype association in DC/TBD. However, some data associate *DKC1*, *TERC*, *TERT*, *TINF2*, and *WRAP53* genes with a higher risk of cancer (198). Patients who developed cancer from the DC/TBD NCI cohort presented different patterns of inheritance: 48.1% had AD-non*TINF2* disease, 29.6% had AR/XLR disease, 3.7% had *TINF2*-associated disease, and 18.7% had an unknown mode of inheritance (198). Regarding the type of cancer, 71.4% of the leukemias and 40% of the solid tumors were developed by patients with AD disease and 35% of the solid tumors were developed by patients with AR/XLR disease. Five patients (three *TINF2*-associated diseases, two AR/XLR) developed a solid malignancy after HCT (198).

There is a lack of information regarding the risk of cancer in heterozygous carriers of PVs in genes associated with DC/TBD phenotype. While PVs in the TERT are highly penetrant and rare, more frequent and low penetrant single nucleotide polymorphisms (SNPs) have been linked to TBD, although they are not considered cancer predisposition syndromes. Most SNP carriers in the general population remain disease free. However, SNPs in TERT have been associated with cancer (314). For example, four TERT heterozygous variants (p.A243V, p.T726M, p.A1062T, and p.V1090M) were found in 3.3% of patients diagnosed with hepatocellular carcinoma associated with cirrhosis (315, 316). Moreover, the p.A1062T variant reported a threefold higher allele frequency in patients with AML and has also been found in higher frequency in cirrhotic patients (316). The p.A279T variant reported a fivefold higher allele frequency in patients with esophageal carcinoma (317).

4 Conclusions

FA and DC/TBD are IBMFS with genetic instability and cancer predisposition. A similar hematooncological presentation must compel the clinician to consider these two diseases as

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differential diagnoses during evaluation. Specialized testing such as chromosomal breakage and telomere length may be a common practice in BMF clinics in many developed countries but, worldwide, this is not the norm. Despite technological advances and the decreasing cost of DNA sequencing, there are significative disparities in access to genomic medicine, and diagnosis through high-throughput methods is available only to a small fraction of the world's population. This is why, in countries where access to specialized tests is limited if not impossible, clinical assessment is king. The one-to-one comparison presented in this review may be used as guidance to integrate a clinical suspicion. Both disorders affect multiple systems and have pleiotropic manifestations. By and large, intentional evaluation results in a higher detection rate of specific features, particularly those that do not give symptomatology but need special tests or maneuvers to uncover them. It is important to keep in mind that there are age-dependent features that might need to be reevaluated later in the life of these patients. Their hematological and oncological manifestations overlap importantly. However, despite both IBMFS having extramedullary features, a discerning analysis of their type and time frame of appearance can clinically distinguish the two disorders. This distinction may permit hand-tailored follow-up to intentionally identify and care for features specific to each diagnosis as well as better guide the confirming test to request. Moreover, a precise diagnosis can further impact on the family as a whole, as this may allow the discussion of theoretical recurrence risks.

The genetic pathway disrupted in each of these conditions, hints as to why, despite presenting features in the same organs, their type and spectrum are distinct. In patients with FA, the failure in DNA repair appears to influence the availability of stem cells during morphogenesis. Also, DNA repair is necessary to maintain the pool of pluripotent stem cells during embryogenesis (318), possibly affecting migrating patterns (224). The latter may explain why many of the features included in VACTERL-H and PHENOS acronyms belong to the malformation category. Since equivocal formation occurs during embryogenesis, physical abnormalities found in FA are usually present before birth. They might not be detected immediately unless they represent a critical danger to the patient's life, such as severe cardiac malformations, esophageal atresia, or anal malformations.

In comparison, physical findings in DC/TBD have an insidious evolution and are related to a scarring phenotype that results from fibrosis. We propose that frequent anomalies seen in at least 20% of patients with DC/TBD could be grouped in the GINPOD acronym (GastroIntestinal [including liver and esophageal], Neurological, Pulmonary, Ocular, and Dermatological [including skin, nails, and leukoplakia]). Telomere stability is altered in DC/TBD; this has a particular impact in tissues that depend on continuous replacement like

epithelia. The maintenance of an adequate number of cells is dependent in telomere length; therefore, telomere attrition severely limits the cell proliferation and tissue regeneration capacities, leading to a replacement of healthy tissue by fibrotic one (46). Telomere metabolism disruption can also lead to a population of cells with genomic damage progress to cancer.

The cytogenetic phenotype of patients with FA is constant throughout life and is present in all cellular types; these qualities have made the cytogenetic test the gold standard for FA diagnosis. On the contrary, in patients with DC/TBD, having short telomeres does not translate into a consistent cytogenetic phenotype that can be used for diagnosis. The characteristic DC/ TBD telomeric shortening, present since birth and evidenced by measuring telomere length, can progress with age and may become evident later in life. This progression could be related with the anticipation seen in some families with DC/TBD (319, 320). In accordance with this, higher frequencies of chromosomal aberrations have been observed in older members than in younger members of the same family (116). Further analysis of the DC/TBD cytogenetic phenotype, using next generation sequencing, FISH-banding, and M-FISH, could help elucidate the chromosome consequences of the telomere attrition.

Last but not least, discovery of genes associated with the FA and DC/TBDs phenotypes remains active. There are a few known patients with a cytogenetic diagnosis of FA (positive chromosome breakage test) in whom the genotype causing it has not been identified yet despite genetic analysis of known FANC genes. This incomplete genotype issue is also true for patients with telomeres below the first percentile for age. PVs in genes long-time known to protect the genome's integrity, like dominant PVs in RPA1, have recently been recognized as a cause of DC/TBDs (321, 322). This issue is expected to be solved by more powerful and more readily available genotyping techniques.

Author contributions

MF-R: Conceptualization, Data curation, Formal analysis, Supervision, Writing: Review and Editing. BG: Conceptualization, Data curation, Formal analysis, Supervision, Writing: Review and Editing. PL-A: Data curation, Formal analysis, Writing: Review and Editing. Rt'H: Data curation, Formal analysis, Writing: Review and Editing. TW-O: Data curation, Formal analysis, Supervision, Writing: Review and Editing. SF: Conceptualization, Data curation, Formal analysis, Supervision, Fundingacquisition, Writing: Review and Editing. AR: Conceptualization, Data curation, Formal analysis, Supervision, Funding acquisition, Writing: Review and Editing. All authors contributed to the article and approved the submitted version.

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References

1. Liesveld J. Bone marrow failure syndromes. Best Pract Res Clin Haematol (2021) 34(2). doi: 10.1016/j.beha.2021.101288

2. Groarke EM, Young NS, Calvo KR. Distinguishing constitutional from acquired bone marrow failure in the hematology clinic. *Best Pract Res Clin Haematol* (2021) 34(2):101275. doi: 10.1016/j.beha.2021.101275

3. Savage SA, Niewisch MR. Dyskeratosis Congenita and Related Telomere Biology Disorders. (2009) [updated 2022 Mar 31]. In: MP Adam, DB Everman, GM Mirzaa, RA Pagon, SE Wallace, LJH Bean, et al, editors. GeneReviews[®] [Internet]. Seattle (WA): University of Washington, Seattle; 1993–2022.

4. García-de-Teresa B, Rodríguez A, Frias S. Chromosome instability in fanconi anemia: From breaks to phenotypic consequences. *Genes (Basel)* (2020) 11 (12):1528. doi: 10.3390/genes11121528

5. Steier W, van Voolen GA, Selmanowitz VJ. Dyskeratosis congenita: relationship to fanconi's anemia. *Blood* (1972) 39(4). doi: 10.1182/blood.V39.4.510.510

6. Rodríguez A, D'Andrea A. Fanconi anemia pathway. *Curr Biol* (2017) 27(18): R986–8. doi: 10.1016/j.cub.2017.07.043

7. Niraj J, Färkkilä A, D'andrea AD. The fanconi anemia pathway in cancer. Annu Rev Cancer Biol (2019) 3:457–78. doi: 10.1146/annurev-cancerbio-030617-

8. Fragkos M, Ganier O, Coulombe P, Méchali M. DNA Replication origin activation in space and time. *Nat Rev Mol Cell Biol* (2015) 16(6):360-74. doi: 10.1038/nrm4002

9. Johansson E, Dixon N. Replicative DNA polymerases. Cold Spring Harb Perspect Biol (2013) 5(6):a012799. doi: 10.1101/cshperspect.a012799

10. Deans AJ, West SC. DNA Interstrand crosslink repair and cancer. Nat Rev Cancer (2011) 11(7):467–80. doi: 10.1038/nrc3088

11. Murat P, Balasubramanian S. Existence and consequences of G-quadruplex structures in DNA. Curr Opin Genet Dev (2014) 25:22–9. doi: 10.1016/j.gde.2013.10.012

12. Garaycoechea JI, Crossan GP, Langevin F, Mulderrig L, Louzada S, Yang F, et al. Alcohol and endogenous aldehydes damage chromosomes and mutate stem cells. *Nature* (2018) 553(7687):171–7. doi: 10.1038/nature25154

13. Voulgaridou GP, Anestopoulos I, Franco R, Panayiotidis MI, Pappa A. DNA Damage induced by endogenous aldehydes: Current state of knowledge. *Mutat Res* (2011) 711(1-2):13-27. doi: 10.1016/j.mrfmmm.2011.03.006

14. Deans AJ, West SC. FANCM connects the genome instability disorders bloom's syndrome and fanconi anemia. *Mol Cell* (2009) 36(6):943–53. doi: 10.1016/j.molcel.2009.12.006

15. Gari K, Décaillet C, Delannoy M, Wu L, Constantinou A. Remodeling of DNA replication structures by the branch point translocase FANCM. *Proc Natl Acad Sci U.S.A.* (2008) 105(42):16107–12. doi: 10.1073/pnas.0804777105

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16. Gari K, Décaillet C, Stasiak AZ, Stasiak A, Constantinou A. The fanconi anemia protein FANCM can promote branch migration of holliday junctions and replication forks. *Mol Cell* (2008) 29(1):141–8. doi: 10.1016/j.molcel.2007.11.032

17. van Twest S, Murphy VJ, Hodson C, Tan W, Swuec P, O'Rourke JJ, et al. Mechanism of ubiquitination and deubiquitination in the fanconi anemia pathway. *Mol Cell* (2017) 65(2):247–59. doi: 10.1016/j.molcel.2016.11.005

18. Liang CC, Zhan B, Yoshikawa Y, Haas W, Gygi SP, Cohn MA. UHRF1 is a sensor for DNA interstrand crosslinks and recruits FANCD2 to initiate the fanconi anemia pathway. *Cell Rep* (2015) 10(12):1947–56. doi: 10.1016/j.celrep.2015.02.053

19. Machida YJ, Machida Y, Chen Y, Gurtan AM, Kupfer GM, D'Andrea AD, et al. UBE2T is the E2 in the fanconi anemia pathway and undergoes negative autoregulation. *Mol Cell* (2006) 23(4):589–96. doi: 10.1016/j.molcel.2006.06.024

20. Rickman KA, Lach FP, Abhyankar A, Donovan FX, Sanborn EM, Kennedy JA, et al. Deficiency of UBE2T, the E2 ubiquitin ligase necessary for FANCD2 and FANCI ubiquitination, causes FA-T subtype of fanconi anemia. *Cell Rep* (2015) 12 (1):35–41. doi: 10.1016/j.celrep.2015.06.014

21. Nakanishi K, Yang YG, Pierce AJ, Taniguchi T, Digweed M, D'Andrea AD, et al. Human fanconi anemia monoubiquitination pathway promotes homologous DNA repair. *Proc Natl Acad Sci U.S.A.* (2005) 102(4):1110–5. doi: 10.1073/pnas.0407796102

22. Bogliolo M, Lyakhovich A, Callén E, Castellà M, Cappelli E, Ramírez MJ, et al. Histone H2AX and fanconi anemia FANCD2 function in the same pathway to maintain chromosome stability. *EMBO J* (2007) 26(5):1340–51. doi: 10.1038/ sj.emboj.7601574

23. Tan W, van Twest S, Murphy VJ, Deans AJ. ATR-mediated FANCI phosphorylation regulates both ubiquitination and deubiquitination of FANCD2. *Front Cell Dev Biol* (2020) 8. doi: 10.3389/fcell.2020.00002

24. Alcón P, Shakeel S, Chen ZA, Rappsilber J, Patel KJ, Passmore LA. FANCD2-FANCI is a clamp stabilized on DNA by monoubiquitination of FANCD2 during DNA repair. *Nat Struct Mol Biol* (2020) 27(3):240–8. doi: 10.1038/s41594-020-0380-1

25. Wang R, Wang S, Dhar A, Peralta C, Pavletich NP. DNA Clamp function of the monoubiquitinated fanconi anaemia ID complex. *Nature* (2020) 580 (7802):278–82. doi: 10.1038/s41586-020-2110-6

26. Crossan GP, van der Weyden L, Rosado IV, Langevin F, Gaillard PHL, McIntyre RE, et al. Disruption of mouse Slx4, a regulator of structure-specific nucleases, phenocopies fanconi anemia. *Nat Genet* (2011) 43(2):147-52. doi: 10.1038/ng.752

27. Kim Y, Lach FP, Desetty R, Hanenberg H, Auerbach AD, Smogorzewska A. Mutations of the SLX4 gene in fanconi anemia. *Nat Genet* (2011) 43(2):142-6. doi: 10.1038/ng.750

28. Lachaud C, Castor D, Hain K, Muñoz I, Wilson J, MacArtney TJ, et al. Distinct functional roles for the two SLX4 ubiquitin-binding UBZ domains mutated in fanconi anemia. *J Cell Sci* (2014) 127(Pt 13):2811–7. doi: 10.1242/jcs.146167

29. Ishimoto R, Tsuzuki Y, Matsumura T, Kurashige S, Enokitani K, Narimatsu K, et al. SLX4-XPF mediates DNA damage responses to replication stress induced by DNA-protein interactions. *J Cell Biol* (2021) 220(1):e202003148. doi: 10.1083/jcb.202003148

30. Klein Douwel D, Boonen RACM, Long DT, Szypowska AA, Räschle M, Walter JC, et al. XPF-ERCC1 acts in unhooking DNA interstrand crosslinks in cooperation with FANCD2 and FANCP/SLX4. *Mol Cell* (2014) 54(3):460–71. doi: 10.1016/i.molcel.2014.03.015

31. Chang DJ, Cimprich KA. DNA Damage tolerance: When it's OK to make mistakes. *Nat Chem Biol* (2009) 5(2):82–90. doi: 10.1038/nchembio.139

32. Pace P, Mosedale G, Hodskinson MR, Rosado IV, Sivasubramaniam M, Patel KJ. Ku70 corrupts DNA repair in the absence of the fanconi anemia pathway. *Science* (2010) 329(5988):219–23. doi: 10.1126/science.1192277

33. Ceccaldi R, Rondinelli B, D'Andrea AD. Repair pathway choices and consequences at the double-strand break. *Trends Cell Biol* (2016) 26(1):52-64. doi: 10.1016/j.tcb.2015.07.009

34. Golub EI, Gupta RC, Haaf T, Wold MS, Radding CM. Interaction of human rad51 recombination protein with single-stranded DNA binding protein, RPA. *Nucleic Acids Res* (1998) 26(23):5388–93. doi: 10.1093/nar/26.23.5388

35. Bhattacharyya A, Ear US, Koller BH, Weichselbaum RR, Bishop DK. The breast cancer susceptibility gene BRCA1 is required for subnuclear assembly of Rad51 and survival following treatment with the DNA cross-linking agent cisplatin. *J Biol Chem* (2000) 275(31):23899–903. doi: 10.1074/jbc.C000276200

36. Vaz F, Hanenberg H, Schuster B, Barker K, Wiek C, Erven V, et al. Mutation of the RAD51C gene in a fanconi anemia-like disorder. *Nat Genet* (2010) 42 (5):406–9. doi: 10.1038/ng.570

37. Fiesco-Roa MO, Giri N, McReynolds LJ, Best AF, Alter BP. Genotypephenotype associations in fanconi anemia: A literature review. *Blood Rev* (2019) 37:100589. doi: 10.1016/j.blre.2019.100589

38. Park J, Chung NG, Chae H, Kim M, Lee S, Kim Y, et al. FANCA and FANCG are the major fanconi anemia genes in the Korean population. *Clin Genet* (2013) 84(3):271–5. doi: 10.1111/cge.12042

39. Mori M, Hira A, Yoshida K, Muramatsu H, Okuno Y, Shiraishi Y, et al. Pathogenic mutations identified by a multimodality approach in 117 Japanese fanconi anemia patients. *Haematologica* (2019) 104(10):1962–73. doi: 10.3324/haematol.2018.207241

40. Maciejowski J, de Lange T. Telomeres in cancer: Tumour suppression and genome instability. *Nat Rev Mol Cell Biol* (2017) 18(3):175–86. doi: 10.1038/nrm. 2016.171

41. Lim CJ, Cech TR. Shaping human telomeres: from shelterin and CST complexes to telomeric chromatin organization. *Nat Rev Mol Cell Biol* (2021) 22 (4):283–98. doi: 10.1038/s41580-021-00328-y

42. Amiard S, Doudeau M, Pinte S, Poulet A, Lenain C, Faivre-Moskalenko C, et al. A topological mechanism for TRF2-enhanced strand invasion. *Nat Struct Mol Biol* (2007) 14(2):147–54. doi: 10.1038/nsmb1192

43. Ingles ED, Deakin JE. Telomeres, species differences, and unusual telomeres in vertebrates: Presenting challenges and opportunities to understanding telomere dynamics. *AIMS Genet* (2016) 3(1):1–24. doi: 10.3934/genet.2016.1.1

44. Moyzis RK, Buckingham JM, Cram LS, Dani M, Deaven LL, Jones MD, et al. A highly conserved repetitive DNA sequence, (TTAGGG)n, present at the telomeres of human chromosomes. *Proc Natl Acad Sci U.S.A.* (1988) 85(18). doi: 10.1073/pnas.85.18.6622

45. Greider CW. Telomerase and telomere-length regulation: lessons from small eukaryotes to mammals. *Cold Spring Harb Symp Quant Biol* (1993) 58:719-23. doi: 10.1101/SQB.1993.058.01.079

46. Hayflick L. How and why we age. *Exp Gerontol* (1998) 33(7–8):639–53. doi: 10.1016/S0531-5565(98)00023-0

47. Londoño-Vallejo JA. Telomere length heterogeneity and chromosome instability. *Cancer Lett* (2004) 212(2):135-44. doi: 10.1016/j.canlet.2004.05.008

48. de Lange T. Shelterin: The protein complex that shapes and safeguards human telomeres. *Genes Dev* (2005) 19(18):2100-10. doi: 10.1101/gad.1346005

49. Armanios M, Blackburn EH. The telomere syndromes. Nat Rev Genet (2012) 13(10):693-704. doi: 10.1038/nrg3246

50. Podlevsky JD, Chen JJL. It all comes together at the ends: Telomerase structure, function, and biogenesis. *Mutat Res* (2012) 730(1–2):3–11. doi: 10.1016/j.mrfmmm.2011.11.002

51. Mitchell JR, Wood E, Collins K. A telomerase component is defective in the human disease dyskeratosis congenita. *Nature* (1999) 402(6761):551–5. doi: 10.1038/990141

52. Meier UT. The many facets of H/ACA ribonucleoproteins. Chromosoma (2005) 114(1):1-14. doi: 10.1007/s00412-005-0333-9

53. Chen JL, Blasco MA, Greider CW. Secondary structure of vertebrate telomerase RNA. *Cell* (2000) 100(5):503–14. doi: 10.1016/S0092-8674(00)80687-X

54. Wright WE, Piatyszek MA, Rainey WE, Byrd W, Shay JW. Telomerase activity in human germline and embryonic tissues and cells. *Dev Genet* (1996) 18 (2):173–9. doi: 10.1002/(SICI)1520-6408(1996)18:2<173::AID-DVG10>3.0. CO;2-3

55. Lazzerini-Denchi E, Sfeir A. Stop pulling my strings - what telomeres taught us about the DNA damage response. *Nat Rev Mol Cell Biol* (2016) 17(6):364–78. doi: 10.1038/nrm.2016.43

56. Diman A, Decottignies A. Genomic origin and nuclear localization of TERRA telomeric repeat-containing RNA: from darkness to dawn. *FEBS J* (2018) 285(8):1389–1398. doi: 10.1111/febs.14363

57. Guo X, Deng Y, Lin Y, Cosme-Blanco W, Chan S, He H, et al. Dysfunctional telomeres activate an ATM-ATR-dependent DNA damage response to suppress tumorigenesis. *EMBO J* (2007) 26(22):4709-19. doi: 10.1038/sj.emboj.7601893

58. Burger K, Ketley RF, Gullerova M. Beyond the trinity of ATM, ATR, and DNA-PK: Multiple kinases shape the DNA damage response in concert with RNA metabolism. *Front Mol Biosci* (2019) 6. doi: 10.3389/fmolb.2019.00061

59. Fedak EA, Adler FR, Abegglen LM, Schiffman JD. ATM And ATR activation through crosstalk between DNA damage response pathways. *Bull Math Biol* (2021) 83(4):38. doi: 10.1007/s11538-021-00868-6

60. Matsuoka S, Huang M, Elledge SJ. Linkage of ATM to cell cycle regulation by the Chk2 protein kinase. *Science* (1998) 282(5395):1893–7. doi: 10.1126/ science.282.5395.1893

61. Zhao H, Piwnica-Worms H. ATR-mediated checkpoint pathways regulate phosphorylation and activation of human Chk1. *Mol Cell Biol* (2001) 21(13):4129–39. doi: 10.1128/MCB.21.13.4129-4139.2001

62. Smith HL, Southgate H, Tweddle DA, Curtin NJ. DNA Damage checkpoint kinases in cancer. *Expert Rev Mol Med* (2020) 22:e2. doi: 10.1017/erm.2020.3

63. Taylor WR, Stark GR. Regulation of the G2/M transition by p53. Oncogene (2001) 20(15):1803-15. doi: 10.1038/sj.onc.1204252

64. Uziel T, Lerenthal Y, Moyal L, Andegeko Y, Mittelman L, Shiloh Y. Requirement of the MRN complex for ATM activation by DNA damage. *EMBO J* (2003) 22(20):5612–21. doi: 10.1093/emboj/cdg541

65. Nimonkar AV, Genschel J, Kinoshita E, Polaczek P, Campbell JL, Wyman C, et al. BLM-DNA2-RPA-MRN and EXO1-BLM-RPA-MRN constitute two DNA end resection machineries for human DNA break repair. *Genes Dev* (2011) 25(4):350–62. doi: 10.1101/gad.2003811

66. Singh TR, Ali AM, Paramasivam M, Pradhan A, Wahengbam K, Seidman MM, et al. ATR-dependent phosphorylation of FANCM at serine 1045 is essential for FANCM functions. *Cancer Res* (2013) 73(14):4300–10. doi: 10.1158/0008-5472.CAN-12-3976

67. Collis SJ, Ciccia A, Deans AJ, Horejsí Z, Martin JS, Maslen SL, et al. FANCM and FAAP24 function in ATR-mediated checkpoint signaling independently of the fanconi anemia core complex. *Mol Cell* (2008) 32(3):313–24. doi: 10.1016/j.molcel.2008.10.014

68. Ishiai M, Kitao H, Smogorzewska A, Tomida J, Kinomura A, Uchida E, et al. FANCI phosphorylation functions as a molecular switch to turn on the fanconi anemia pathway. *Nat Struct Mol Biol* (2008) 15(11):1138–46. doi: 10.1038/ nsmb.1504

69. Attwooll CL, Akpinar M, Petrini JHJ. The mre11 complex and the response to dysfunctional telomeres. *Mol Cell Biol* (2009) 29(20):5540–51. doi: 10.1128/MCB.00479-09

70. Dimitrova N, de Lange T. Cell cycle-dependent role of MRN at dysfunctional telomeres: ATM signaling-dependent induction of nonhomologous end joining (NHEJ) in G1 and resection-mediated inhibition of NHEJ in G2. *Mol Cell Biol* (2009) 29(20):5552–63. doi: 10.1128/MCB.00476-09

71. Badie S, Carlos AR, Folio C, Okamoto K, Bouwman P, Jonkers J, et al. BRCA1 and CtIP promote alternative non-homologous end-joining at uncapped telomeres. *EMBO J* (2015) 34(3):410–24. doi: 10.15252/embj.201488947

72. Maciejowski J, Li Y, Bosco N, Campbell PJ, de Lange T. Chromothripsis and kataegis induced by telomere crisis. *Cell* (2015) 163(7):1641–54. doi: 10.1016/j.cell.2015.11.054

73. Denchi EL, de Lange T. Protection of telomeres through independent control of ATM and ATR by TRF2 and POT1. *Nature* (2007) 448(7157):1068-71. doi: 10.1038/nature06065

74. Takai H, Smogorzewska A, de Lange T. DNA Damage foci at dysfunctional telomeres. Curr Biol (2003) 13(17):1549–56. doi: 10.1016/S0960-9822(03)00542-6

75. Hsu HL, Gilley D, Blackburn EH, Chen DJ. Ku Is associated with the telomere in mammals. *Proc Natl Acad Sci U.S.A.* (1999) 96(22):12454–8. doi: 10.1073/pnas.96.22.12454

76. Ribes-Zamora A, Indiviglio SM, Mihalek I, Williams CL, Bertuch AA. TRF2 interaction with Ku heterotetramerization interface gives insight into c-NHEJ prevention at human telomeres. *Cell Rep* (2013) 5(1):194–206. doi: 10.1016/j.celrep.2013.08.040

77. Rai R, Zheng H, He H, Luo Y, Multani A, Carpenter PB, et al. The function of classical and alternative non-homologous end-joining pathways in the fusion of dysfunctional telomeres. *EMBO J* (2010) 29(15):2598–610. doi: 10.1038/emboj.2010.142

 Schmutz I, Timashev L, Xie W, Patel DJ, de Lange T. TRF2 binds branched DNA to safeguard telomere integrity. *Nat Struct Mol Biol* (2017) 24(9):734–742. doi: 10.1038/nsmb.3451

79. Timashev LA, de Lange T. Characterization of t-loop formation by TRF2. Nucleus (2020) 11(1):164–177. doi: 10.1080/19491034.2020.1783782

80. Karlseder J, Hoke K, Mirzoeva OK, Bakkenist C, Kastan MB, Petrini JHJ, et al. The telomeric protein TRF2 binds the ATM kinase and can inhibit the ATM-dependent DNA damage response. *PloS Biol* (2004) 2(8):E240. doi: 10.1371/journal.pbio.0020240

81. Flynn RL, Centore RC, O'Sullivan RJ, Rai R, Tse A, Songyang Z, et al. TERRA and hnRNPA1 orchestrate an RPA-to-POT1 switch on telomeric single-stranded DNA. *Nature* (2011) 471(7339):532–6. doi: 10.1038/nature09772

82. Zimmermann M, Kibe T, Kabir S, de Lange T. TRF1 negotiates TTAGGG repeat-associated replication problems by recruiting the BLM helicase and the TPP1/POT1 repressor of ATR signaling. *Genes Dev* (2014) 28(22):2477–91. doi: 10.1101/gad.251611.114

83. Ceccaldi R, Parmar K, Mouly E, Delord M, Kim JM, Regairaz M, et al. Bone marrow failure in fanconi anemia is triggered by an exacerbated p53/p21 DNA damage response that impairs hematopoietic stem and progenitor cells. *Cell Stem Cell* (2012) 11(1):36–49. doi: 10.1016/j.stem.2012.05.013

84. Fok WC, Niero EL de O, Dege C, Brenner KA, Sturgeon CM, Batista LFZ. p53 mediates failure of human definitive hematopoiesis in dyskeratosis congenita. *Stem Cell Rep* (2017) 9(2):409–418. doi: 10.1016/j.stemcr.2017.06.015

85. Rodríguez A, Zhang K, Färkkilä A, Filiatrault J, Yang C, Velázquez M, et al. MYC promotes bone marrow stem cell dysfunction in fanconi anemia. *Cell Stem Cell* (2021) 28(1):33–47.e8. doi: 10.1016/j.stem.2020.09.004

86. Schärer OD. DNA Interstrand crosslinks: Natural and drug-induced DNA adducts that induce unique cellular responses. *ChemBioChem* (2005) 6(1):27-32. doi: 10.1002/cbic.200400287

87. Cai MY, Dunn CE, Chen W, Kochupurakkal BS, Nguyen H, Moreau LA, et al. Cooperation of the ATM and fanconi Anemia/BRCA pathways in doublestrand break end resection. *Cell Rep* (2020) 30(7):2402–2415.e5. doi: 10.1016/ j.celrep.2020.01.052

88. Symington LS, Gautier J. Double-strand break end resection and repair pathway choice. *Annu Rev Genet* (2011) 45:247–71. doi: 10.1146/annurev-genet-110410-132435

89. Delacôte F, Lopez BS. Importance of the cell cycle phase for the choice of the appropriate DSB repair pathway, for genome stability maintenance: The trans-s double-strand break repair model. *Cell Cycle* (2008) 7(1):33–8. doi: 10.4161/cc.7.1.5149

90. Scully R, Panday A, Elango R, Willis NA. DNA Double-strand break repairpathway choice in somatic mammalian cells. *Nat Rev Mol Cell Biol* (2019) 20 (11):698–714. doi: 10.1038/s41580-019-0152-0

91. Pannunzio NR, Watanabe G, Lieber MR. Nonhomologous DNA endjoining for repair of DNA double-strand breaks. *J Biol Chem* (2018) 293 (27):10512–10523. doi: 10.1074/jbc.TM117.000374

92. Molina B, Ramos S, Frias S. Fanconi anemia, part 1. cytogenetic diagnosis. Acta Pediatr Mex (2022) 43(2):102-28. doi: 10.18233/APM43No2pp102-1282393

93. Auerbach AD. Diagnosis of fanconi anemia by diepoxybutane analysis. Curr Protoc Hum Genet (2015) 2015:8.7.1–8.7.17. doi: 10.1002/0471142905.hg0807s85

94. Esmer C, Sánchez S, Ramos S, Molina B, Frias S, Carnevale A. DEB test for fanconi anemia detection in patients with atypical phenotypes. *Am J Med Genet* (2004) 124 A(1):35–9. doi: 10.1002/ajmg.a.20327

95. Kubbies M, Schindler D, Hoehn H, Schinzel A, Rabinovitch PS. Endogenous blockage and delay of the chromosome cycle despite normal recruitment and growth phase explain poor proliferation and frequent edomitosis in fanconi anemia cells. *Am J Hum Genet* (1985) 37(5):1022–30.

96. Auerbach AD. Fanconi anemia diagnosis and the diepoxybutane (DEB) test. *Exp Hematol* (1993) 21(6):731–3.

97. Soulier J, Leblanc T, Larghero J, Dastot H, Shimamura A, Guardiola P, et al. Detection of somatic mosaicism and classification of fanconi anemia patients by analysis of the FA/BRCA pathway. *Blood* (2005) 105(3):1329–36. doi: 10.1182/blood-2004-05-1852

98. Fargo JH, Rochowski A, Giri N, Savage SA, Olson SB, Alter BP. Comparison of chromosome breakage in non-mosaic and mosaic patients with fanconi anemia,

relatives, and patients with other inherited bone marrow failure syndromes. *Cytogenetic Genome Res* (2014) 144(1):15–27. doi: 10.1159/000366251

99. Nicoletti E, Rao G, Bueren JA, Río P, Navarro S, Surrallés J, et al. Mosaicism in fanconi anemia: Concise review and evaluation of published cases with focus on clinical course of blood count normalization. *Ann Hematol* (2020) 99(5):913–24. doi: 10.1007/s00277-020-03954-2

100. Feldstein J, Fernandez Garcia S. The diagnosis and treatment of dyskeratosis congenita: A review. *J Blood Med* (2014) 21(5):157–67. doi: 10.2147/JBM.S47437

101. Alter BP, Baerlocher GM, Savage SA, Chanock SJ, Weksler BB, Willner JP, et al. Very short telomere length by flow fluorescence *in situ* hybridization identifies patients with dyskeratosis congenita. *Blood* (2007) 110(5):1439–47. doi: 10.1182/ blood-2007-02-075598

102. Park M. Overview of inherited bone marrow failure syndromes. *Blood Res* (2022) 57(S1):49-54. doi: 10.5045/br.2022.2022012

103. Niewisch MR, Savage SA. An update on the biology and management of dyskeratosis congenita and related telomere biology disorders. *Expert Rev Hematol* (2019) 12(12):1037–52. doi: 10.1080/17474086.2019.1662720

104. Alter BP, Giri N, Savage SA, Rosenberg PS. Telomere length in inherited bone marrow failure syndromes. *Haematologica* (2015) 100(1):49–54. doi: 10.3324/ haematol.2014.114389

105. Sirinavin C, Trowbridge AA. Dyskeratosis congenita: Clinical features and genetic aspects. report of a family and review of the literature. *J Med Genet* (1975) 12(4):339–54. doi: 10.1136/jmg.12.4.339

106. Shahidi NT. Fanconi anemia, dyskeratosis congenita, and WT syndrome. Am J Med Genet Suppl (1987) 3:263-78. doi: 10.1002/ajmg.1320280531

107. Schneider A, Mayer U, Gebhart E, Harms D, Gromball J, Glöckel U, et al. Clastogen-induced fragility may differentiate pancytopenia of congenital dyskeratosis from fanconi anaemia. *Eur J Pediatr* (1988) 148(1):37–9. doi: 10.1007/BF00441811

108. Morrison JG. Dyskeratosis congenita: Two extremes. S Afr Med J (1974) 48 (6):223–5.

109. Aguilar-Martinhz A, Lautre-Ecenarro MJ, Urbina-Gonzalez F, Cristobal-Gil MC, Guerra-Rodriguez P, Garcia-Perez A. Cytogenetic abnormalities in dyskeratosis congenita-report of five cases. *Clin Exp Dermatol* (1988) 13(2):100– 4. doi: 10.1111/j.1365-2230.1988.tb00668.x

110. DeBauche DM, Pai GS, Stanley WS. Enhanced G2 chromatid radiosensitivity in dyskeratosis congenita fibroblasts. *Am J Hum Genet* (1990) 46 (2):350–7.

111. Pai GS, Yan Y, DeBauche DM, Stanley WS, Paul SR. Bleomycin hypersensitivity in dyskeratosis congenita fibroblasts, lymphocytes, and transformed lymphoblasts. *Cytogenetic Genome Res* (1989) 52(3–4):186–9. doi: 10.1159/000132874

112. Dokal I, Bungey J, Williamson P, Oscier D, Hows J, Luzzatto L. Dyskeratosis congenita fibroblasts are abnormal and have unbalanced chromosomal rearrangements. *Blood* (1992) 80(12):3090-6. doi: 10.1182/blood.V80.12.3090.bloodjournal80123090

113. Kehrer H, Krone W, Schindler D, Kaufmann R, Schrezenmeier H. Cytogenetic studies of skin fibroblast cultures from a karyotypically normal female with dyskeratosis congenita. *Clin Genet* (1992) 41(3):129–34. doi: 10.1111/j.1399-0004.1992.tb03648.x

114. Gutman A, Frumkin A, Adam A, Bloch-Shtacher N, Rozenszajn LA. X-Linked dyskeratosis congenita with pancytopenia. *Arch Dermatol* (1978) 114 (11):1667–71. doi: 10.1001/archderm.1978.01640230041011

115. Womer R, Clark JE, Wood P, Sabio H, Kelly TE. Dyskeratosis congenita: Two examples of this multisystem disorder. *Pediatrics* (1983) 71(4):603–9. doi: 10.1542/peds.71.4.603

116. Dokal I. Dyskeratosis congenita. Hematol Am Soc Hematol Educ Program (2011) 2011:480-6. doi: 10.1182/asheducation-2011.1.480

117. Dokal I. Dyskeratosis congenita in all its forms. Br J Haematol (2000) 110 (4):768–79. doi: 10.1046/j.1365-2141.2000.02109.x

118. AlSabbagh MM. Dyskeratosis congenita: A literature review. JDDG: J der Deutschen Dermatologischen Gesellschaft (2020) 18(9):943–967. doi: 10.1111/ ddg.14268

119. Zhang M, Cao Y, Wu H, Li H. Brain imaging features of children with hoyeraal-hreidarsson syndrome. *Brain Behav* (2021) 11(5):e02079. doi: 10.1002/brb3.2079

120. Berthet F, Tuchschmid P, Boltshauser E, Seger RA, Aalfs CM, van den Berg H, et al. The hoyeraal-hreidarsson syndrome: Don't forget the associated immunodeficiency. *Eur J Pediatr* (1995) 154(12):998. doi: 10.1007/BF01958649

121. Nespoli L, Lascari C, Maccario R, Nosetti L, Broggi U, Locatelli F, et al. The hoyeraal-hreidarsson syndrome: The presentation of the seventh case. *Eur J Pediatr* (1997) 156(10):818–20.

122. Karremann M, Neumaier-Probst E, Schlichtenbrede F, Beier F, Brümmendorf TH, Cremer FW, et al. Revesz syndrome revisited. Orphanet J Rare Dis (2020) 15(1):299. doi: 10.1186/s13023-020-01553-y

123. Wegman-Ostrosky T, Savage SA. The genomics of inherited bone marrow failure: from mechanism to the clinic. *Br J Haematol* (2017) 177(4):526–42. doi: 10.1111/bjh.14535

124. Shtessel L, Ahmed S. Telomere dysfunction in human bone marrow failure syndromes. *Nucleus* (2011) 2(1):24–9. doi: 10.4161/nucl.2.1.13993

125. le Guen T, Jullien L, Touzot F, Schertzer M, Gaillard L, Perderiset M, et al. Human RTEL1 deficiency causes hoyeraal-hreidarsson syndrome with short telomeres and genome instability. *Hum Mol Genet* (2013) 22(16):3239–49. doi: 10.1093/hmg/ddt178

126. Marrone A, Walne A, Tamary H, Masunari Y, Kirwan M, Beswick R, et al. Telomerase reverse-transcriptase homozygous mutations in autosomal recessive dyskeratosis congenita and hoyeraal-hreidarsson syndrome. *Blood* (2007) 110 (13):4198–205. doi: 10.1182/blood-2006-12-062851

127. Benyelles M, O'Donohue MF, Kermasson L, Lainey E, Borie R, Lagresle-Peyrou C, et al. NHP2 deficiency impairs rRNA biogenesis and causes pulmonary fibrosis and hoyeraal-hreidarsson syndrome. *Hum Mol Genet* (2020) 29(6):907–22. doi: 10.1093/hmg/ddaa011

128. Mehta PA, Ebens C. Fanconi anemia. In: *GeneReviews® [Internet]*. Seattle (WA): University of Washington, Seattle; Initial Posting: February 14, 2002; Last Update: June 3, 2021

129. The Rockefeller University, 2004-2022, *Fanconi anemia Mutation Database, LOVD v.3.0.* (New York:The Rockefeller University) (2022). Available at: https://www2.rockefeller.edu/fanconi

130. McKusick-nathans institute of genetic medicine JHUS of m. OMIM[®] database (2022). Available at: https://www.omim.org/.

131. Knies K, Inano S, Ramírez MJ, Ishiai M, Surrallés J, Takata M, et al. Biallelic mutations in the ubiquitin ligase RFWD3 cause fanconi anemia. *J Clin Invest* (2017) 127(8):3013–3027. doi: 10.1172/JCI92069

132. Wang AT, Smogorzewska A. SnapShot: Fanconi anemia and associated proteins. Cell (2015) 160(1-2):354-354.e1. doi: 10.1016/j.cell.2014.12.031

133. Shimamura A, Alter BP. Pathophysiology and management of inherited bone marrow failure syndromes. *Blood Rev* (2010) 24(3):101–22. doi: 10.1016/ j.blre.2010.03.002

134. Faivre L, Portnoï MF, Pals G, Stoppa-Lyonnet D, le Merrer M, Thauvin-Robinet C, et al. Should chromosome breakage studies be performed in patients with VACTERL association? *Am J Med Genet* (2005) 137 A(1):55–8. doi: 10.1002/ajmg.a.30853

135. Alter BP, Giri N. Thinking of VACTERL-h? rule out fanconi anemia according to PHENOS. *Am J Med Genetics Part A* (2016) 170(6):1520-4. doi: 10.1002/ajmg.a.37637

136. Fiesco-Roa M, Gómez-Moreno P, Espinosa-Curiel R, Rodriguez A, Frías S. Dysmorphology as a clinical tool for an early diagnosis of fanconi anemia. *Acta Pediatr Mex* (2022) 43(2):129–40. doi: 10.18233/APM43No2pp129-1402402

137. Rackley S, Pao M, Seratti GF, Giri N, Rasimas JJ, Alter BP, et al. Neuropsychiatric conditions among patients with dyskeratosis congenita: A link with telomere biology? *Psychosomatics* (2012) 53(3):230–5. doi: 10.1016/j.psym.2011.09.003

138. Vulliamy T, Dokal I. Dyskeratosis congenita. Semin Hematol (2006) 43 (3):157-66. doi: 10.1053/j.seminhematol.2006.04.001

139. Fiesco Roa M, Monsivais Orozco A, Rodriguez A, Frías S, Garcia-de Teresa B. Síndromes de falla medular hereditarios: Etiología, fisiopatología, diagnóstico y tratamiento. *Acta Pediátrica México* (2021) 42(4):192–207. doi: 10.18233/ APM42No4pp192-2072110

140. Ruud E, Wesenberg F. Microcephalus, medulloblastoma and excessive toxicity from chemotherapy: An unusual presentation of fanconi anaemia. *Acta Paediatric* (2007) 90(5):580–3. doi: 10.1111/j.1651-2227.2001.tb00803.x

141. Berrebi D, Lebras MN, Belarbi N, Couturier J, Fattet S, Faye A, et al. Bilateral adrenal neuroblastoma and nephroblastoma occurring synchronously in a child with fanconi's anemia and VACTERL syndrome. *J Pediatr Surg* (2006) 41(1): e11–4. doi: 10.1016/j.jpedsurg.2005.10.087

142. Ameziane N, May P, Haitjema A, van de Vrugt HJ, van Rossum-Fikkert SE, Ristic D, et al. A novel fanconi anaemia subtype associated with a dominantnegative mutation in RAD51. *Nat Commun* (2015) 18(6):8829. doi: 10.1038/ ncomms9829

143. Sherafat-Kazemzadeh R, Mehta SN, Care MM, Kim MO, Williams DA, Rose SR, et al. Small pituitary size in children with fanconi anemia. *Pediatr Blood Cancer* (2007) 49(2):166–70. doi: 10.1002/pbc.21148

144. Johnson-Tesch BA, Gawande RS, Zhang L, MacMillan ML, Nascene DR. Fanconi anemia: Correlating central nervous system malformations and genetic

complementation groups. *Pediatr Radiol* (2017) 47(7):868–76. doi: 10.1007/s00247-017-3817-x

145. Aksu T, Gümrük F, Bayhan T, Coşkun Ç, Oğuz KK, Unal S. Central nervous system lesions in fanconi anemia: Experience from a research center for fanconi anemia patients. *Pediatr Blood Cancer* (2020) 67(12):e28722. doi: 10.1002/pbc.28722

146. Stivaros SM, Alston R, Wright NB, Chandler K, Bonney D, Wynn RF, et al. Central nervous system abnormalities in fanconi anaemia: Patterns and frequency on magnetic resonance imaging. *Br J Radiol* (2015) 88(1056):20150088. doi: 10.1259/bjr.20150088

147. Bhala S, Best AF, Giri N, Alter BP, Pao M, Gropman A, et al. CNS manifestations in patients with telomere biology disorders. *Neurol Genet* (2019) 5 (6):370. doi: 10.1212/NXG.000000000000370

148. Yoshimasu T, Tanaka R, Suenobu S, Yagasaki H, Yoshino H, Ueda T, et al. Prompt and durable hematopoietic reconstitution by unrelated cord blood transplantation in a child with Fanconi anemia. *Bone Marrow Transplant* (2001) 27(7):767–9. doi: 10.1038/sj.bmt.1702876

149. Kalb R, Neveling K, Hoehn H, Schneider H, Linka Y, Batish SD, et al. Hypomorphic mutations in the gene encoding a key fanconi anemia protein, FANCD2, sustain a significant group of FA-D2 patients with severe phenotype. *Am J Hum Genet* (2007) 80(5):895–910. doi: 10.1086/517616

150. Savage SA, Ballew BJ, Giri N, Chandrasekharappa SC, Ameziane N, de Winter J, et al. Novel FANCI mutations in fanconi anemia with VACTERL association. *Am J Med Genet Part A* (2016) 170A(2):386–391. doi: 10.1002/ajmg.a.37461

151. Tsilou ET, Giri N, Weinstein S, Mueller C, Savage SA, Alter BP. Ocular and orbital manifestations of the inherited bone marrow failure syndromes: Fanconi anemia and dyskeratosis congenita. *Ophthalmology* (2010) 117(3):615–22. doi: 10.1016/j.ophtha.2009.08.023

152. Takenaka S, Kuroda Y, Ohta S, Mizuno Y, Hiwatari M, Miyatake S, et al. Japanese Patient with RAD51 -associated fanconi anemia. *Am J Med Genet Part A* (2019) 179(6):900–902. doi: 10.1002/ajmg.a.61130

153. Pignatti CB, Bianchi E, Polito E. Fanconi's anemia in infancy. report of a case diagnosed in the pre-anemic stage. *Helv Paediatr Acta* (1977) 32(4-5):413-8.

154. Yahia SB, Touffahi SA, Zeghidi H, Zaouali S, Khairallah M. Ocular neovascularization in a patient with fanconi anemia. *Can J Ophthalmol* (2006) 41(6):778–9. doi: 10.3129/i06-078

155. Bahar I. Weinberger d, kramer m, axer-siegel r. retinal vasculopathy in fanconi anemia: A case report. *Retina* (2005) 25(6):799-800. doi: 10.1097/00006982-200509000-00023

156. Elgohary MA, Lim SK, Siriwardena D, Moore AT, Wormald RP. Increased crystalline lens thickness and phacomorphic glaucoma in patients with fanconi anemia. *J Cataract Refractive Surg* (2006) 32(10):1771–4. doi: 10.1016/j.jcrs.2006.04.036

157. Aslan D, Ozdogan S, Onol M, Kaya Z, Gursel T. An unusual ocular manifestation in fanconi anemia: Congenital glaucoma. *Am J Hematol* (2005) 78 (1):64–6. doi: 10.1002/ajh.20232

158. Denny M, Haug SJ, Cunningham ET, Jumper JM. Fanconi anemia presenting as bilateral diffuse retinal occlusive vasculopathy. *RETINAL cases Brief Rep* (2016) 10(2):171-4. doi: 10.1097/ICB.00000000000219

159. Martín-Sanz R, Peña D, López-Miguel A, Coco-Martín MB, González-García H, Álvarez-Guisasola FJ, et al. Coats disease in a patient with fanconi anemia: A case report. *Eur J Ophthalmol* (2015) 25(2):182–3. doi: 10.5301/ejo.5000517

160. Chen MS, Chang KP, Lin KH, Tsai CL, Ho TC. Retinopathy in a patient with fanconi anemia and vitamin B12 deficiency. *Eye* (2012) 26(2):331. doi: 10.1038/eye.2011.276

161. Graf CM, Nichele S, Siviero RB, Loth G, Trennepohl JP, Zinher MT, et al. Ocular manifestations in patients with fanconi anemia: A single-center experience including 106 patients. *J Pediatr* (2022) 242:228–234. doi: 10.1016/j.jpeds.2021.11.013

162. Törnquist AL, Martin L, Winiarski J, Fahnehjelm KT. Ocular manifestations and visual functions in patients with fanconi anaemia. *Acta Ophthalmol* (2014) 92(2):171-8. doi: 10.1111/aos.12132

163. Kaur S, Jain S, Sodhi H, Rastogi A, Kamlesh. Optic nerve hypoplasia. Oman J Ophthalmol (2013) 6(2):77–82. doi: 10.4103/0974-620X.116622

164. Nazir S, Sayani N, Phillips PH. Retinal hemorrhages in a patient with dyskeratosis congenita. J Am Assoc Pediatr Ophthalmol Strabismus (2008) 12 (4):415–7. doi: 10.1016/j.jaapos.2008.01.016

165. Zagórski Z, Biziorek B, Rakowska E, Jedrzejewski D. Zinsser-Engman-Cole-Syndrom (Dyskeratosis congenita) mit schwerem sicca-syndrom, panuveitis und hornhautperforation - eine Kasuistik12. *Klinische Monatsblätter für Augenheilkunde* (2001) 218(6):455–8. doi: 10.1055/s-2001-16262

166. Escutia Muñoz B, Marí Ruiz JI, Sánchez Motilla JM, Sánchez Carazo JL, Aliaga Boniche A, Esparcia Navarro A. Alteraciones cutáneas, atresia de conductos lacrimales y pancitopenia en hombre joven. *Rev Clínica Española* (2001) 201 (9):545–7. doi: 10.1016/S0014-2565(01)70907-1 167. Merchant A, Zhao TZ, Foster CS. Chronic keratoconjunctivitis associated with congenital dyskeratosis and erythrokeratodermia variablis. *Ophthalmology* (1998) 105(7):1286–91. doi: 10.1016/S0161-6420(98)97035-X

168. Çakmak SK, Gönül M, Kılıç A, Gül Ü, Koçak O, Demiriz M. A case of dyskeratosis congenita with chiari 1 malformation, absence of inferior vena cava, webbed neck, and low posterior hair neck. *Int J Dermatol* (2008) 47(4):377–9. doi: 10.1111/j.1365-4632.2008.03525.x

169. Balogh E, Chandler JC, Varga M, Tahoun M, Menyhárd DK, Schay G, et al. Pseudouridylation defect due to DKC1 and NOP10 mutations causes nephrotic syndrome with cataracts, hearing impairment, and enterocolitis. *Proc Natl Acad Sci* (2020) 117(26):15137–15147. doi: 10.1016/j.ekir.2020.02.476

170. Menon V, Kumar A, Verma L. Zinsser-Cole-Engman syndrome (dyskeratosis congenita) with cataract-a rare association. *Jpn J Ophthalmol* (1986) 30(2):192-6.

171. Finzi A, Morara M, Pichi F, Veronese C, Ciardella AP. Vitreous hemorrhage secondary to retinal vasculopathy in a patient with dyskeratosis congenita. *Int Ophthalmol* (2014) 34(4):923–6. doi: 10.1007/s10792-013-9867-7

172. Johnson CA, Hatfield M, Pulido JS. Retinal vasculopathy in a family with autosomal dominant dyskeratosis congenita. *Ophthalmic Genet* (2009) 30(4):181–4. doi: 10.3109/13816810903148012

173. Teixeira LF. Bilateral retinal vasculopathy in a patient with dyskeratosis congenita. *Arch Ophthalmol* (2008) 126(1):134-5. doi: 10.1001/archophthalmol.2007.4

174. Vaz-Pereira S, Pacheco PA, Gandhi S, Kulasekararaj AG, Marsh JC, Pal B, et al. Bilateral retinal vasculopathy associated with autosomal dominant dyskeratosis congenita. *Eur J Ophthalmol* (2013) 23(5):772–5. doi: 10.5301/ejo.5000297

175. Tabbara KF, Al-Ghamdi A, Al-Mohareb F, Ayas M, Chaudhri N, Al-Sharif F, et al. Ocular findings after allogeneic hematopoietic stem cell transplantation. *Ophthalmology* (2009) 116(9):1624–9. doi: 10.1016/j.ophtha.2009.04.054

176. Teär Fahnehjelm K, Törnquist AL, Olsson M, Bäckström I, Andersson Grönlund M, Winiarski J. Cataract after allogeneic hematopoietic stem cell transplantation in childhood. *Acta Paediatrica* (2016) 105(1):82–9. doi: 10.1111/ apa.13173

177. Kalejaiye A, Giri N, Brewer CC, Zalewski CK, King KA, Adams CD, et al. Otologic manifestations of fanconi anemia and other inherited bone marrow failure syndromes. *Pediatr Blood Cancer* (2016) 63(12):2139–2145. doi: 10.1002/pbc.26155

178. Verheij E, Oomen KPQ, Smetsers SE, van Zanten GA, Speleman L. Hearing loss and speech perception in noise difficulties in fanconi anemia. *Laryngoscope* (2017) 127(10):2358-2361. doi: 10.1002/lary.26480

179. José Vale M, José Dinis M, Bini-Antunes M, Porto B, Barbot J, Coutinho MB. Audiologic abnormalities of fanconi anaemia. *Acta Oto-Laryngol* (2008) 128 (9):992–6. doi: 10.1080/00016480701793750

180. Santos F, Selesnick SH, Glasgold RA. Otologic manifestations of fanconi anemia. *Otol Neurotol* (2002) 23(6):873–5. doi: 10.1097/00129492-200211000-00011

181. Karempelis P, Greenlund L, Gruhl R, Dorrity J, Wagner J, Roby B. Etiologies of hearing loss in fanconi anemia. *Int J Pediatr Otorhinolaryngol* (2022) 155:111068. doi: 10.1016/j.ijporl.2022.111068

182. Mehra S, Eavey RD, Keamy DG. The epidemiology of hearing impairment in the united states: Newborns, children, and adolescents. *Otolaryngol-Head Neck Surg* (2009) 140(4):461–72. doi: 10.1016/j.otohns.2008.12.022

183. Liu H, Luo H, Yang T, Wu H, Chen D. Association of leukocyte telomere length and the risk of age-related hearing impairment in Chinese Hans. *Sci Rep* (2017) 7(1):10106. doi: 10.1038/s41598-017-10680-9

184. Wang J, Nguyen MT, Sung V, Grobler A, Burgner D, Saffery R, et al. Associations between telomere length and hearing status in mid-childhood and midlife. *Ear Hearing* (2019) 40(5):1256-1259. doi: 10.1097/AUD.0000000000000705

185. Li F, Li W, Qiao X, Xie X. Clinical features of dyskeratosis congenita in mainland China: Case reports and literature review. *Int J Hematol* (2019) 109 (3):328–335. doi: 10.1007/s12185-018-02582-x

186. Grein Cavalcanti L, Lyko KF, Araújo RLF, Amenábar JM, Bonfim C, Torres-Pereira CC. Oral leukoplakia in patients with fanconi anaemia without hematopoietic stem cell transplantation. *Pediatr Blood Cancer* (2015) 62(6):1024–6. doi: 10.1002/pbc.25417

187. Tekcicek M, Tavil B, Cakar A, Pinar A, Unal S, Gumruk F. Oral and dental findings in children with fanconi anemia. *Pediatr Dent* (2007) 29 (3):248-52.

188. de Araujo MR, de Oliveira Ribas M, Koubik ACGA, Mattioli T, de Lima AAS, França BHS. Fanconi's anemia: Clinical and radiographic oral manifestations. *Oral Diseases* (2007) 13(3):291–5. doi: 10.1111/j.1601-0825.2006.01282.x 189. Alter BP, Giri N, Savage SA, Rosenberg PS. Cancer in the national cancer institute inherited bone marrow failure syndrome cohort after fifteen years of follow-up. *Haematologica* (2018) 103(1):30–9. doi: 10.3324/haematol.2017.178111

190. Atkinson JC, Harvey KE, Domingo DL, Trujillo MI, Guadagnini JP, Gollins S, et al. Oral and dental phenotype of dyskeratosis congenita. *Oral Dis* (2008) 14(5):419-27. doi: 10.1111/j.1601-0825.2007.01394.x

191. Açikgöz A, Ozden FO, Fisgin T, Açikgöz G, Duru F, Yarali N, et al. Oral and dental findings in fanconi's anemia. *Pediatr Hematol Oncol* (2005) 22(6):531–9. doi: 10.1080/08880010591002413

192. Nico MMS, Hammerschmidt M, Lourenço SV. Oral mucosal manifestations in some genodermatoses: Correlation with cutaneous lesions. *Eur J Dermatol* (2013) 23(5):581–91. doi: 10.1684/ejd.2013.2073

193. Velleuer E, Dietrich R, Pomjanski N, de Santana Almeida Araujo IK, Silva de Araujo BE, Sroka I, et al. Diagnostic accuracy of brush biopsy-based cytology for the early detection of oral cancer and precursors in fanconi anemia. *Cancer Cytopathol* (2020) 128(6):403–413. doi: 10.1002/cncy.22249

194. Warnakulasuriya S, Kujan O, Aguirre-Urizar JM, Bagan JV, González-Moles MÁ, Kerr AR, et al. Oral potentially malignant disorders: A consensus report from an international seminar on nomenclature and classification, convened by the WHO collaborating centre for oral cancer. *Oral Dis* (2021) 27(8):1862–1880. doi: 10.1111/odi.13704

195. Axéll T. Occurrence of leukoplakia and some other oral white lesions among 20,333 adult Swedish people. *Community Dent Oral Epidemiol* (1987) 15 (1):46–51. doi: 10.1111/j.1600-0528.1987.tb00479.x

196. Kapuria D, Ben-Yakov G, Ortolano R, Cho MH, Kalchiem-Dekel O, Takyar V, et al. The spectrum of hepatic involvement in patients with telomere disease. *Hepatology* (2019) 69(6):2579-85. doi: 10.1002/hep.30578

197. Calado RT, Regal JA, Kleiner DE, Schrump DS, Peterson NR, Pons V, et al. A spectrum of severe familial liver disorders associate with telomerase mutations. *PloS One* (2009) 4(11):e7926. doi: 10.1371/journal.pone.0007926

198. Niewisch MR, Giri N, McReynolds LJ, Alsaggaf R, Bhala S, Alter BP, et al. Disease progression and clinical outcomes in telomere biology disorders. *Blood* (2022) 139(12):1807–1819. doi: 10.1182/blood.2021013523

199. Calado RT, Brudno J, Mehta P, Kovacs JJ, Wu C, Zago MA, et al. Constitutional telomerase mutations are genetic risk factors for cirrhosis. *Hepatology* (2011) 53(5):1600-7. doi: 10.1002/hep.24173

200. Olson TS, Chan ES, Paessler ME, Sullivan KE, Frantz CN, Russo P, et al. Liver failure due to hepatic angiosarcoma in an adolescent with dyskeratosis congenita. *J Pediatr Hematol/Oncol* (2014) 36(4):312-5. doi: 10.1097/MPH.0b013e318286d4d4

201. Masserot-Lureau C, Adoui N, Degos F, de Bazelaire C, Soulier J, Chevret S, et al. Incidence of liver abnormalities in fanconi anemia patients. *Am J Hematol* (2012) 87:547–9. doi: 10.1002/ajh.23153

202. Jonassaint NL, Guo N, Califano JA, Montgomery EA, Armanios M. The gastrointestinal manifestations of telomere-mediated disease. *Aging Cell* (2013) 12 (2):319–23. doi: 10.1111/acel.12041

203. Himes RW, Chiou EH, Queliza K, Shouval DS, Somech R, Agarwal S, et al. Gastrointestinal hemorrhage: A manifestation of the telomere biology disorders. *J Pediatric* (2021) 230:55–61.e4. doi: 10.1016/j.jpeds.2020.09.038

204. Khincha PP, Bertuch AA, Agarwal S, Townsley DM, Young NS, Keel S, et al. Pulmonary arteriovenous malformations: An uncharacterised phenotype of dyskeratosis congenita and related telomere biology disorders. *Eur Respir J* (2017) 49(1):1601640. doi: 10.1183/13993003.01640-2016

205. Steinberg-Shemer O, Goldberg TA, Yacobovich J, Levin C, Koren A, Revel-Vilk S, et al. Erratum: Characterization and genotype-phenotype correlation of patients with fanconi anemia in a multi-ethnic population. *Haematologica* (2020) 105(7):1825–34. doi: 10.3324/haematol.2019.222877

206. Alter BP, Rosenberg PS. VACTERL-h association and fanconi anemia. Mol Syndromol (2013) 4(1-2):87-93. doi: 10.1159/000346035

207. Jercan C, Bică AM, Marcu AD, Radu LE, Şerbănică A, Asan M, et al. Clinical and hematological aspects in pediatric patients diagnosed with fanconi anemia – a single-center experience. *Pediatru.ro* (2021) 4(64):22. doi: 10.26416/ Pedi.64.4.2021.5783

208. Perel Y, Butenandt O, Carrere A, Saura R, Fayon M, Lamireau T, et al. Oesophageal atresia, VACTERL association: Fanconi's anaemia related spectrum of anomalies. *Arch Dis Child* (1998) 78:375–6. doi: 10.1136/adc.78.4.375

209. Savage SA, Bertuch AA. The genetics and clinical manifestations of telomere biology disorders. *Genet Med* (2010) 12(12):753-64. doi: 10.1097/GIM.0b013e3181f415b5

210. Anemia Research Fund F. Fanconi anemia clinical care Guidelines_5th edition 2020_web. (Eugene Oregon: Fanconi anemia Research Fund) (2020)

211. Hoffman JIE, Kaplan S. The incidence of congenital heart disease. J Am Coll Cardiol (2002) 39(12):1890–900.

212. Reller MD, Strickland MJ, Riehle-Colarusso T, Mahle WT, Correa A. Prevalence of congenital heart defects in metropolitan Atlanta, 1998-2005. J Pediatric (2008) 153(6):807–13. doi: 10.1016/j.jpeds.2008.05.059

213. Knight S, Vulliamy T, Copplestone A, Gluckman E, Mason P, Dokal I. Dyskeratosis congenita (DC) registry: Identification of new features of DC. *Br J Haematol* (1998) 103(4):990–6. doi: 10.1046/j.1365-2141.1998.01103.x

214. Wang P, Xu Z. Pulmonary fibrosis in dyskeratosis congenita: A case report with a PRISMA-compliant systematic review. *BMC Pulm Med* (2021) 21(1):279. doi: 10.1186/s12890-021-01645-w

215. Savage SA, Alter BP. Dyskeratosis congenita. Hematol Oncol Clin North Am (2009) 23(2):215-31. doi: 10.1016/j.hoc.2009.01.003

216. Armanios MY, Chen JJL, Cogan JD, Alder JK, Ingersoll RG, Markin C, et al. Telomerase mutations in families with idiopathic pulmonary fibrosis. *N Engl J Med* (2007) 356(13):1317–26. doi: 10.1056/NEJMoa066157

217. Diaz De Leon A, Cronkhite JT, Yilmaz C, Brewington C, Wang R, Xing C, et al. Subclinical lung disease, macrocytosis, and premature graying in kindreds with telomerase (TERT) mutations. *Chest* (2011) 140(3):753–63. doi: 10.1378/ chest.10-2865

218. Wang P, Xu Z. Pulmonary fibrosis in dyskeratosis congenita: A case report with a PRISMA-compliant systematic review. *BMC Pulmonary Med* (2021) 21 (1):279. doi: 10.1186/s12890-021-01645-w

219. Alter BP, Giri N, Savage SA, Peters JA, Loud JT, Leathwood L, et al. Malignancies and survival patterns in the national cancer institute inherited bone marrow failure syndromes cohort study. *Br J Haematol* (2010) 150(2):179–88. doi: 10.1111/j.1365-2141.2010.08212.x

220. Rubinstein WS, Wenger SL, Hoffman RM, Auerbach AD, Mulvihill JJ. Interstitial lung disease in an adult with fanconi anemia: Clues to the pathogenesis. *J Med Genet* (1997) 69:315–9. doi: 10.1002/(SICI)1096-8628(19970331)69:3<315:: AID-AJMG18>3.0.CO;2-U

221. Steens RD, Summers QA, Tarala RA. Pulmonary alveolar proteinosis in association with fanconi's anemia and psoriasis; A possible common pathogenetic mechanism. *Chest* (1992) 102(2):637–8. doi: 10.1378/chest.102.2.637

222. Eldar M, Shoenfeld Y, Zaizov R, Fogel R, Asherov J, Liban E, et al. Pulmonary alveolar proteinosis associated with fanconi's anemia. *Respiration* (1979) 38:77–9. doi: 10.1159/000194077

223. Samarakoon L, Ranawaka N, Rodrigo C, Constantine GR, Goonarathne L. Fanconi anaemia with bilateral diffuse pulmonary arterio venous fistulae: A case report. *BMC Blood Disord* (2012) 12:1. doi: 10.1186/1471-2326-12-1

224. Sathyanarayana V, Lee B, Wright NB, Santos R, Bonney D, Wynn R, et al. Patterns and frequency of renal abnormalities in fanconi anaemia: Implications for long-term management. *Pediatr Nephrol* (2018) 33(9):1547–51. doi: 10.1007/s00467-018-3952-0

225. de Kerviler E, Guermazi A, Zagdanski AM, Gluckman E, Frija J. *Pictorial review the clinical and radiological features of fanconi's anaemia.* Available at: http://www.idealibrary.com.

226. Glanz A, Fraser FC. Spectrum of anomalies in fanconi anaemia*. J Med Genet (1982) 19:412-6. doi: 10.1136/jmg.19.6.412

227. Faivre L, Guardiola P, Lewis C, Dokal I, Ebell W, Zatterale A, et al. Association of complementation group and mutation type with clinical outcome in fanconi anemia. *Blood* (2000) 96(13):4064–70. doi: 10.1182/blood.V96.13.4064

228. da Cunha MM, Barreto FC, Nichele S, Trennepohl J, Ribeiro L, Loth G, et al. Kidney complications in 107 fanconi anemia patients submitted to hematopoietic cell transplantation. *Eur J Pediatric* (2022) 181(2):715–23. doi: 10.1007/s00431-021-04263-0

229. Alter BP, Tenner MS. Brain tumors in patients with fanconi's anemia. Arch Pediatr Adolesc Med (1994) 148(6):661-3. doi: 10.1001/archpedi.1994. 02170060115027

230. Jung M, Ramanagoudr-Bhojappa R, van Twest S, Rosti RO, Murphy V, Tan W, et al. Association of clinical severity with FANCB variant type in fanconi anemia. *Blood* (2020) 135(18):1588–1602. doi: 10.1182/blood.2019003249

231. Olsen TG, Peck GL, Lovegrove RH. Acute urinary tract obstruction in dyskeratosis congenita. J Am Acad Dermatol (1981) 4(5):556–60. doi: 10.1016/S0190-9622(81)70056-2

232. Balci S, Engiz O, Erekul A, Gozdasoglu S, Vulliamy T. An atypical form of dyskeratosis congenita with renal agenesis and no mutation in DKC1, TERC and TERT genes. J Eur Acad Dermatol Venereol (2009) 23(5):607–8. doi: 10.1111/j.1468-3083.2008.02985.x

233. Kesici S, Ünal Ş, Kuşkonmaz B, Aytaç S, Çetin M, Gümrük F. Fanconi anemia: A single center experience of a large cohort. *Turkish J Pediatric* (2019) 61 (4):477–84. doi: 10.24953/turkjped.2019.04.002

234. Kirwan M, Dokal I. Dyskeratosis congenita: A genetic disorder of many faces. Clin Genet (2008) 73:103–12. doi: 10.1111/j.1399-0004.2007.00923.x

235. Bakar Ö, Işik U, Canpolat C, Alanay Y. Hoyeraal-hreidarsson syndrome: An extremely rare dyskeratosis congenita phenotype. *Pediatr Dermatol* (2015) 32 (6):e263–6. doi: 10.1111/pde.12693

236. Coelho JD, Lestre S, Kay T, Lopes MJP, Fiadeiro T, Apetato M. Dyskeratosis congenita-two siblings with a new missense mutation in the DKC1 gene. *Pediatr Dermatol* (2011) 28(4):464–6. doi: 10.1111/j.1525-1470.2010.01299.x

237. Reimann C, Kloeckener-Gruissem B, Niemeyer CM, Vanscheidt W. Late manifestation of dyskeratosis congenita presenting as chronic dermal ulcer in a 37year-old man. J Eur Acad Dermatol Venereol (2008) 22(7):897–8. doi: 10.1111/ j.1468-3083.2007.02530.x

238. Koruyucu M, Barlak P, Seymen F. Oral and dental findings of dyskeratosis congenita. *Case Rep Dent* (2014) 2014:454128. doi: 10.1155/2014/454128

239. Vulliamy T, Beswick R, Kirwan M, Marrone A, Digweed M, Walne A, et al. Mutations in the telomerase component NHP2 cause the premature ageing syndrome dyskeratosis congenita. *Proc Natl Acad Sci U.S.A.* (2008) 105 (23):8073–8. doi: 10.1073/pnas.0800042105

240. Amin Guldmann S, Byrjalsen A, Shaker S, Elberling J. A new pathogenic variant of the RTEL1 gene and dyskeratosis congenita: A dermatological view. *Acta Derm Venereol* (2022) 102:adv00710. doi: 10.2340/actadv.v102.919

241. Quintana-Castanedo L, Feito-Rodríguez M, Pérez-Conde I, Maseda-Pedrero R, Chiloeches-Fernández C, de Lucas Laguna R. Chronic sole ulcer in a child with dyskeratosis congenita: An atypical wound successfully treated with punch grafting. *Pediatr Dermatol* (2020) 37(6):1131–1134. doi: 10.1111/pde.14316

242. Kanegane H, Kasahara Y, Okamura J, Hongo T, Tanaka R, Nomura K, et al. Identification of DKC1 gene mutations in Japanese patients with X-linked dyskeratosis congenita. *Br J Haematol* (2005) 129(3):432–4. doi: 10.1111/j.1365-2141.2005.05473.x

243. Ruggiero JL, Dodds M, Freese R, Polcari IC, Maguiness S, Hook KP, et al. Cutaneous findings in fanconi anemia. *J Am Acad Dermatol* (2021) 85(5):1253–58. doi: 10.1016/j.jaad.2020.08.047

244. Feben C, Kromberg J, Wainwright R, Stones D, Sutton C, Poole J, et al. Phenotypic consequences in black south African fanconi anemia patients homozygous for a founder mutation. *Genet Med* (2014) 16(5):400–6. doi: 10.1038/gim.2013.159

245. Altintas B, Giri N, McReynolds LJ, Best A, Alter BP. Genotype-phenotype and outcome associations in patients with fanconi anemia: The national cancer institute cohort. *Haematologica* (2022). doi: 10.3324/haematol.2021.279981

246. Chatham-Stephens K, Devere T, Guzman-Cottrill J, Kurre P. Metachronous manifestations of sweet's syndrome in a neutropenic patient with fanconi anemia. *Pediatr Blood Cancer* (2008) 51(1):128-30. doi: 10.1002/pbc.21538

247. McDermott MB, Corbally MT, O'Marcaigh AS. Extracutaneous sweet syndrome involving the gastrointestinal tract in a patient with fanconi anemia. *J Pediatr Hematol Oncol* (2001) 23(1):59–62. doi: 10.1097/00043426-200101000-00015

248. Cerejeira A, Amoedo P, Coelho AR, Silva R, Pedrosa A, Nogueira A, et al. Recurrent sweet syndrome presenting in a figurate pattern in a patient with fanconi anemia. *Int J Dermatol* (2022) 61(5):e171–e173. doi: 10.1111/ijd.15848

249. Giulino L, Guinan EC, Gillio AP, Drachtman RA, Teruya-Feldstein J, Boulad F. Sweet syndrome in patients with fanconi anaemia: Association with extracutaneous manifestations and progression of haematological disease. *Br J Haematol* (2011) 154(2):278–81. doi: 10.1111/j.1365-2141.2011.08604.x

250. Cohen PR. Sweet's syndrome-a comprehensive review of an acute febrile neutrophilic dermatosis. Orphanet J Rare Dis (2007) 2:34. doi: 10.1186/1750-1172-2-34

251. Akdogan N, Ersoy-Evans S, Visal Okur F, Kuskonmaz B, Uckan Cetinkaya D. Long-term skin findings in patients with fanconi anemia after allogeneic hematopoietic stem cell transplantation. *Bone Marrow Transplant* (2021) 56 (11):2866–2868. doi: 10.1038/s41409-021-01441-4

252. James MA, McCarroll HR, Manske PR. The spectrum of radial longitudinal deficiency: A modified classification. *J Handb Surg Am* (1999) 24(6):1145–55. doi: 10.1053/jhsu.1999.1145

253. Forman M, Canizares MF, Bohn D, James MA, Samora J, Steinman S, et al. Association of radial longitudinal deficiency and thumb hypoplasia: An update using the CoULD registry. *J Bone Joint Surg Am* (2020) 102(20):1815–1822. doi: 10.2106/JBJS.20.00281

254. Porteous MEM, Cross I, Burn J. VACTERL with hydrocephalus: One end of the fanconi anemia spectrum of anomalies? *Am J Med Genet* (1992) 43(6):1032–4. doi: 10.1002/ajmg.1320430624

255. Solomon BD, Bear KA, Kimonis V, de Klein A, Scott DA, Shaw-Smith C, et al. Clinical geneticists' views of VACTERL/VATER association. *Am J Med Genet Part A* (2012) 158A(12):3087–100. doi: 10.1002/ajmg.a.35638

256. Alter BP. Arm anomalies and bone marrow failure may go hand in hand. J Handb Surg Am (1992) 17(3):566–71. doi: 10.1016/0363-5023(92)90372-V

257. Bourke G, Wilks D, Kinsey S, Feltbower RG, Giri N, Alter BP. The incidence and spectrum of congenital hand differences in patients with fanconi anaemia: Analysis of 48 patients. *J Handb Surg Eur* (2022) 47(7):711–15. doi: 10.1177/17531934221087521

258. Webb ML, Rosen H, Taghinia A, McCarty ER, Cerrato F, Upton J, et al. Incidence of fanconi anemia in children with congenital thumb anomalies referred for diepoxybutane testing. *J Handb Surg Am* (2011) 36(6):1052–7. doi: 10.1016/j.jhsa.2011.02.018

259. Goldfarb CA, Wall L, Manske PR. Radial longitudinal deficiency: The incidence of associated medical and musculoskeletal conditions. *J Handb Surg Am* (2006) 31(7):1176–82. doi: 10.1016/j.jhsa.2006.05.012

260. McGaughran J. Klippel-feil anomaly in fanconi anemia. *Clin Dysmorphol* (2003) 12(3):197. doi: 10.1097/01.mcd.0000077560.66911.1f

261. Reyes P, García-de Teresa B, Juárez U, Pérez-Villatoro F, Fiesco-Roa MO, Rodríguez A, et al. Fanconi anemia patients from an indigenous community in Mexico carry a new founder pathogenic variant in FANCG. *Int J Mol Sci* (2022) 23 (4):2334. doi: 10.3390/ijms23042334

262. Alter BP, Rosenberg PS, Brody LC. Clinical and molecular features associated with biallelic mutations in FANCD1/BRCA2. J Med Genet (2007) 44 (1):1–9. doi: 10.1136/jmg.2006.043257

263. Colle I, Laureys G, Raevens S, Libbrecht L, Leroy JG, Reyntjens K, et al. Living related liver transplantation in an adult patient with hepatocellular adenoma and carcinoma 13 years after bone marrow transplantation for fanconi anemia: A case report. *Hepatol Res* (2013) 43(9):991–8. doi: 10.1111/hepr.12043

264. Bottega R, Nicchia E, Cappelli E, Ravera S, de Rocco D, Faleschini M, et al. Hypomorphic FANCA mutations correlate with mild mitochondrial and clinical phenotype in fanconi anemia. *Haematologica* (2018) 103(3):417–426. doi: 10.3324/ haematol.2017.176131

265. Rose SR, Myers KC, Rutter MM, Mueller R, Khoury JC, Mehta PA, et al. Endocrine phenotype of children and adults with fanconi anemia. *Pediatr Blood Cancer* (2012) 59(4):690–6. doi: 10.1002/pbc.24095

266. Gunasekaran U, Agarwal N, Jagasia MH, Jagasia SM. Endocrine complications in long-term survivors after allogeneic stem cell transplant. *Semin Hematol* (2012) 49(1):66–72. doi: 10.1053/j.seminhematol.2011.10.010

267. Kossiva L, Vartzelis G, Harisi M, Logotheti I, Garoufi A. Too short stature, too many stigmata. *BMJ Case Rep* (2010) bcr0620103087. doi: 10.1136/bcr.06.2010.3087

268. Glousker G, Touzot F, Revy P, Tzfati Y, Savage SA. Unraveling the pathogenesis of hoyeraal-hreidarsson syndrome, a complex telomere biology disorder. *Br J Haematol* (2015) 170(4):457–71. doi: 10.1111/bjh.13442

269. Giri N, Batista DL, Alter BP, Stratakis CA. Endocrine abnormalities in patients with fanconi anemia. *J Clin Endocrinol Metab* (2007) 92(7):2624–31. doi: 10.1210/jc.2007-0135

270. Dillon B, Feben C, Segal D, du Plessis J, Reynders D, Wainwright R, et al. Endocrine profiling in patients with fanconi anemia, homozygous for a FANCG founder mutation. *Mol Genet Genomic Med* (2020) 8(8):e1351. doi: 10.1002/mg3.1351

271. Petryk A, Kanakatti Shankar R, Giri N, Hollenberg AN, Rutter MM, Nathan B, et al. Endocrine disorders in fanconi anemia: Recommendations for screening and treatment. *J Clin Endocrinol Metab* (2015) 100(3):803–11. doi: 10.1210/jc.2014-4357

272. Noda-Yamashita T, Kajihara I, Kidou M, Ihn H. Dyskeratosis congenita associated with congenital hypothyroidism. *J Dermatol* (2018) 45(4):e76–e77. doi: 10.1111/1346-8138.14109

273. Kutbay NO, Yurekli BS, Erdemir Z, Karaca E, Unal I, Yaman B, et al. A case of dyskeratosis congenita associated with hypothyroidism and hypogonadism. *Hormones (Athens)* (2016) 15(2):297–299. doi: 10.14310/horm.2002.1655

274. Savage S, Cook E. Dyskeratosis congenita and telomere biology disorders: Diagnosis and management guidelines (2015). Available at: https://teamtelomere.org/wp-content/uploads/2018/07/DC-TBD-Diagnosis-And-Management-Guidelines.pdf.

275. Sampson MJ, Winterbone MS, Hughes JC, Dozio N, Hughes DA. Monocyte telomere shortening and oxidative DNA damage in type 2 diabetes. *Diabetes Care* (2006) 29(2):283–9. doi: 10.2337/diacare.29.02.06.dc05-1715

276. Adaikalakoteswari A, Balasubramanyam M, Mohan V. Telomere shortening occurs in Asian Indian type 2 diabetic patients. *Diabetic Med* (2005) 22(9):1151-6. doi: 10.1111/j.1464-5491.2005.01574.x

277. Salpea KD, Talmud PJ, Cooper JA, Maubaret CG, Stephens JW, Abelak K, et al. Association of telomere length with type 2 diabetes, oxidative stress and UCP2 gene variation. *Atherosclerosis* (2010) 209(1):42–50. doi: 10.1016/j.atherosclerosis.2009.09.070

278. Guo N, Parry EM, Li LS, Kembou F, Lauder N, Hussain MA, et al. Short telomeres compromise β -cell signaling and survival. *PloS One* (2011) 6(3):e17858. doi: 10.1371/journal.pone.0017858

279. Sklavos MM, Giri N, Stratton P, Alter BP, Pinto LA. Anti-müllerian hormone deficiency in females with fanconi anemia. *J Clin Endocrinol Metab* (2014) 99(5):1608–14. doi: 10.1210/jc.2013-3559

280. Sklavos MM, Stratton P, Giri N, Alter BP, Savage SA, Pinto LA. Reduced serum levels of anti-müllerian hormone in females with inherited bone marrow failure syndromes. *J Clin Endocrinol Metab* (2015) 100(2):e197–203. doi: 10.1210/jc.2014-2838

281. Giri N, Reed HD, Stratton P, Savage SA, Alter BP. Pregnancy outcomes in mothers of offspring with inherited bone marrow failure syndromes. *Pediatr Blood Cancer* (2018) 65(1). doi: 10.1002/pbc.26757

282. Giri N, Alter BP, Savage SA, Stratton P. Gynaecological and reproductive health of women with telomere biology disorders. *Br J Haematol* (2021) 193 (6):1238–1246. doi: 10.1111/bjh.17545

283. Korthof ET, Svahn J, de Latour RP, Terranova P, Moins-Teisserenc H, Socié G, et al. Immunological profile of fanconi anemia: A multicentric retrospective analysis of 61 patients. *Am J Hematol* (2013) 88(6):472-6. doi: 10.1002/ajh.23435

284. Giri N, Alter BP, Penrose K, Falk RT, Pan Y, Savage SA, et al. Immune status of patients with inherited bone marrow failure syndromes. *Am J Hematol* (2015) 90(8):702–8. doi: 10.1002/ajh.24046

285. Dufour C, Corcione A, Svahn J, Haupt R, Poggi V, Béka'ssy AN, et al. TNFalpha and IFN-gamma are overexpressed in the bone marrow of fanconi anemia patients and TNF-alpha suppresses erythropoiesis *in vitro*. *Blood* (2003) 102 (6):2053–9. doi: 10.1182/blood-2003-01-0114

286. Rosselli F, Sanceau J, Gluckman E, Wietzerbin J, Moustacchi E. Abnormal lymphokine production: A novel feature of the genetic disease fanconi anemia. II. *in vitro* and *in vivo* spontaneous overproduction of tumor necrosis factor alpha. *Blood* (1994) 83(5):1216–25. doi: 10.1182/blood.V83. 5.1216.bloodjournal8351216

287. Sznajer Y, Baumann C, David A, Journel H, Lacombe D, Perel Y, et al. Further delineation of the congenital form of X-linked dyskeratosis congenita (Hoyeraal-hreidarsson syndrome). *Eur J Pediatr* (2003) 162(12):863–7. doi: 10.1007/s00431-003-1317-5

288. Allenspach EJ, Bellodi C, Jeong D, Kopmar N, Nakamura T, Ochs HD, et al. Common variable immunodeficiency as the initial presentation of dyskeratosis congenita. J Allergy Clin Immunol (2013) 132(1):223–6. doi: 10.1016/j.jaci.2012.11.052

289. Rahman N. Realizing the promise of cancer predisposition genes. *Nature* (2014) 505(7483):302-8. doi: 10.1038/nature12981

290. Altintas B, Giri N, McReynolds LJ, Alter BP. Genotype-phenotype associations in patients with fanconi anemia: National cancer institute cohort. *Blood* (2020) 136(Supplement 1):4–5. doi: 10.1182/blood-2020-137200

291. Alter BP. Fanconi anemia and the development of leukemia. Best Pract Research: Clin Haematol (2014) 27:214–21. doi: 10.1016/j.beha.2014.10.002

292. Rosenberg PS, Greene MH, Alter BP. Cancer incidence in persons with fanconi anemia. *Blood* (2003) 101(3):822-6. doi: 10.1182/blood-2002-05-1498

293. Chandrasekharappa SC, Chinn SB, Donovan FX, Chowdhury NI, Kamat A, Adeyemo AA, et al. Assessing the spectrum of germline variation in fanconi anemia genes among patients with head and neck carcinoma before age 50. *Cancer* (2017) 123(20):3943–54. doi: 10.1002/cncr.30802

294. Dias AR, Silva MC de A, Carvalho FM, Carvalho H de A, Diz M del PE, Baracat CE, et al. Fanconi anemia and vaginal squamous cell carcinoma. *Rare Tumors* (2012) 4(1):11–2. doi: 10.4081/rt.2012.e4

295. McReynolds LJ, Biswas K, Giri N, Sharan SK, Alter BP. Genotype-cancer association in patients with fanconi anemia due to pathogenic variants in FANCD1 (BRCA2) or FANCN (PALB2). *Cancer Genet* (2021) 258–259:101–109. doi: 10.1016/j.cancergen.2021.10.001

296. McReynolds LJ, Giri N, Leathwood L, Risch MO, Carr AG, Alter BP. Risk of cancer in heterozygous relatives of patients with fanconi anemia. *Genet Med* (2022) 24(1):245–250. doi: 10.1016/j.gim.2021.08.013

297. Berwick M, Satagopan JM, Ben-Porat L, Carlson A, Mah K, Henry R, et al. Genetic heterogeneity among fanconi anemia heterozygotes and risk of cancer. *Cancer Res* (2007) 67(19):9591-6. doi: 10.1158/0008-5472.CAN-07-1501

298. Abbasi S, Rasouli M. A rare FANCA gene variation as a breast cancer susceptibility allele in an Iranian population. *Mol Med Rep* (2017) 15(6):3983-3988. doi: 10.3892/mmr.2017.6489

299. Solyom S, Winqvist R, Nikkilä J, Rapakko K, Hirvikoski P, Kokkonen H, et al. Screening for large genomic rearrangements in the FANCA gene reveals extensive deletion in a Finnish breast cancer family. *Cancer Lett* (2011) 302(2):113–8. doi: 10.1016/j.canlet.2010.12.020

300. Chen S, Parmigiani G. Meta-analysis of BRCA1 and BRCA2 penetrance. J Clin Oncol (2007) 25(11):1329–33. doi: 10.1200/JCO.2006.09.1066

301. Smetsers S, Muter J, Bristow C, Patel L, Chandler K, Bonney D, et al. Heterozygote FANCD2 mutations associated with childhood T cell ALL and

testicular seminoma. Familial Cancer (2012) 11(4):661–5. doi: 10.1007/s10689-012-9553-3

302. Türke C, Horn S, Petto C, Labudde D, Lauer G, Wittenburg G. Loss of heterozygosity in FANCG, FANCF and BRIP1 from head and neck squamous cell carcinoma of the oral cavity. *Int J Oncol* (2017) 50(6):2207–2220. doi: 10.3892/ijo.2017.3974

303. Rogers CD, van der Heijden MS, Brune K, Yeo CJ, Hruban RH, Kern SE, et al. The genetics of FANCC and FANCG in familial pancreatic cancer. *Cancer Biol Ther* (2004) 3(2):167–9. doi: 10.4161/cbt.3.2.609

304. Chan SH, Ni Y, Li ST, Teo JX, Ishak NDB, Lim WK, et al. Spectrum of germline mutations within fanconi anemia-associated genes across populations of varying ancestry. *JNCI Cancer Spectrum* (2021) 5(1):pkaa117. doi: 10.1093/jncics/pkaa117

305. Domchek SM, Robson ME. Update on genetic testing in gynecologic cancer. *J Clin Oncol* (2019) 37(27):2501-2509. doi: 10.1200/ JCO.19.00363

306. Hu C, Polley EC, Yadav S, Lilyquist J, Shimelis H, Na J, et al. The contribution of germline predisposition gene mutations to clinical subtypes of invasive breast cancer from a clinical genetic testing cohort. *JNCI: J Natl Cancer Inst* (2020) 112(12):1231–1241. doi: 10.1093/jnci/djaa023

307. Neidhardt G, Hauke J, Ramser J, Groß E, Gehrig A, Müller CR, et al. Association between loss-of-Function mutations within the FANCM gene and early-onset familial breast cancer. *JAMA Oncol* (2017) 3(9):1245–1248. doi: 10.1001/jamaoncol.2016.5592

308. Figlioli G, Bogliolo M, Catucci I, Caleca L, Lasheras SV, Pujol R, et al. The FANCM:p.Arg658* truncating variant is associated with risk of triplenegative breast cancer. *NPJ Breast Cancer* (2019) 5:38. doi: 10.1038/s41523-019-0127-5

309. Yang X, Leslie G, Doroszuk A, Schneider S, Allen J, Decker B, et al. Cancer risks associated with germline PALB2 pathogenic variants: An international study of 524 families. *J Clin Oncol* (2020) 38(7):674-685. doi: 10.1200/JCO.19.01907

310. Pelttari LM, Heikkinen T, Thompson D, Kallioniemi A, Schleutker J, Holli K, et al. RAD51C is a susceptibility gene for ovarian cancer. *Hum Mol Genet* (2011) 20(16):3278–88. doi: 10.1093/hmg/ddr229

311. Virts EL, Jankowska A, Mackay C, Glaas MF, Wiek C, Kelich SL, et al. AluY-mediated germline deletion, duplication and somatic stem cell reversion in UBE2T defines a new subtype of fanconi anemia. *Hum Mol Genet* (2015) 24 (18):5093–108. doi: 10.1093/hmg/ddv227

312. Tutt ANJ, Garber JE, Kaufman B, Viale G, Fumagalli D, Rastogi P, et al. Adjuvant olaparib for patients with BRCA1- or BRCA2-mutated breast cancer. N Engl J Med (2021) 384(25):2394–2405. doi: 10.1056/NEJM0a2105215

313. Keung MYT, Wu Y, Vadgama JV. PARP inhibitors as a therapeutic agent for homologous recombination deficiency in breast cancers. *J Clin Med* (2019) 8 (4):435. doi: 10.3390/jcm8040435

314. Kachuri L, Amos CI, McKay JD, Johansson M, Vineis P, Bueno-de-Mesquita HB, et al. Fine mapping of chromosome 5p15.33 based on a targeted deep sequencing and high density genotyping identifies novel lung cancer susceptibility loci. *Carcinogenesis* (2016) 37(1):96–105. doi: 10.1093/carcin/bgv165

315. Xu J, Trudeau MA, Sandford AJ, Wong JMY. Evaluations of short telomere risk-associated single nucleotide polymorphisms in telomerase reverse transcriptase gene on telomere length maintenance. *bioRxiv* (2019) 552133. doi: 10.1101/552133

316. Donaires FS, Scatena NF, Alves-Paiva RM, Podlevsky JD, Logeswaran D, Santana BA, et al. Telomere biology and telomerase mutations in cirrhotic patients with hepatocellular carcinoma. *PloS One* (2017) 12(8):e0183287. doi: 10.1371/journal.pone.0183287

317. Zhang Y, Calado R, Rao M, Hong JA, Meeker AK, Dumitriu B, et al. Telomerase variant A279T induces telomere dysfunction and inhibits noncanonical telomerase activity in esophageal carcinomas. *PloS One* (2014) 9(7): e101010. doi: 10.1371/journal.pone.0101010

318. Bakker ST, de Winter JP, Riele H. Learning from a paradox: recent insights into fanconi anaemia through studying mouse models. *Dis Models Mech* (2013) 6 (1):40–7. doi: 10.1242/dmm.009795

319. Marrone A, Walne A, Dokal I. Dyskeratosis congenita: Telomerase, telomeres and anticipation. *Curr Opin Genet Dev* (2005) 15(3):249-57. doi: 10.1016/j.gde.2005.04.004

320. Vulliamy T, Marrone A, Szydlo R, Walne A, Mason PJ, Dokal I. Disease anticipation is associated with progressive telomere shortening in families with dyskeratosis congenita due to mutations in TERC. *Nat* (2004) 36(5):447–9. doi: 10.1038/ng1346

321. Sharma R, Sahoo SS, Honda M, Granger SL, Goodings C, Sanchez L, et al. Gain-of-function mutations in RPA1 cause a syndrome with short telomeres and somatic genetic rescue. *Blood* (2022) 139(7):1039-51. doi: 10.1182/blood.2021011980

322. Savage SA. Telomere biology disorders gain a family member. *Blood* (2022) 139(7):957–9. doi: 10.1182/blood.2021014533