

CASE REPORT

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Leukemoid reaction and autocrine growth of bladder cancer induced by paraneoplastic production of granulocyte colony-stimulating factor – a potential neoplastic marker: a case report and review of the literature

Anup Kasi Loknath Kumar^{1*}, Megha Teeka Satyan¹, Jeffrey Holzbeierlein², Mohen Mirza² and Peter Van Veldhuizen¹

Abstract

Introduction: Granulocyte colony-stimulating factor produced by nonhematopoietic malignant cells is able to induce a leukemoid reaction by excessive stimulation of leukocyte production. Expression of granulocyte colony-stimulating factor and its functional receptors have been confirmed in bladder cancer cells. *In vitro* studies have demonstrated that granulocyte colony-stimulating factor/receptor exhibits a high affinity binding and this biological axis increases proliferation of the carcinoma. Urothelial carcinoma of the bladder is rarely associated with a leukemoid reaction and autocrine growth induced by paraneoplastic production of granulocyte colony-stimulating factor. In the world literature, there have been less than 35 cases reported in the last 35 years. The clinicopathological aspects, biology, prognosis and management of granulocyte colony-stimulating factor-secreting bladder cancers are poorly understood.

Case presentation: A 39-year-old Caucasian woman with an invasive high-grade urothelial carcinoma presented with hematuria and low-grade fevers. Laboratory tests revealed an elevated white blood cell count and absolute neutrophil count and an elevated 24-hour urine protein. Upon further evaluation she was found to have locally advanced high-grade urothelial carcinoma without nodal or distant metastasis. Her serum granulocyte colony-stimulating factor level was 10 times the normal limit. This led to the diagnosis of a paraneoplastic leukemoid reaction. Her white blood cell count immediately normalized after cystectomy but increased in concordance with recurrence of her disease. Unfortunately, she rapidly progressed and expired within 10 months from the time of first diagnosis.

Conclusions: This is one of the few cases reported that illustrates the existence of a distinct and highly aggressive subtype of bladder cancer which secretes granulocyte colony-stimulating factor. Patients presenting with a leukemoid reaction should be tested for granulocyte colony-stimulating factor/receptor biological axis. Moreover, granulocyte colony-stimulating factor could be a potential neoplastic marker as it can follow the clinical course of the underlying tumor and thus be useful for monitoring its evolution. Neoadjuvant chemotherapy should be considered in these patients due to the aggressive nature of these tumors. With a better understanding of the biology, this autocrine growth signal could be a potential target for therapy in future.

Keywords: Bladder cancer, Granulocyte colony-stimulating factor, Leukemoid reaction

* Correspondence: anupdoc@gmail.com

¹Division of Hematology and Oncology, University of Kansas Medical Center, Kansas City, Kansas, USA

Full list of author information is available at the end of the article

Introduction

Paraneoplastic leukemoid reaction is a type of paraneoplastic syndrome in which there is an elevation in the white blood cell count, predominantly neutrophils. Granulocyte colony-stimulating factor (G-CSF) secreted by nonhematopoietic malignant cells has been reported to be able to induce a leukemoid reaction through excessive stimulation of leukocyte production [1-3]. Urothelial carcinoma of the bladder is rarely associated with a leukemoid reaction, with less than 30 cases reported over the past 30 years, some of which tried to elucidate the role of G-CSF [1,3-7]. A previous study reported that 9.2% of bladder tumors are associated with high levels of G-CSF [8]. In addition, expression of G-CSF and functional G-CSF receptors (G-CSFR) has been demonstrated in bladder cancer cells [9]. *In vitro* studies have demonstrated that G-CSF/G-CSFR exhibit high affinity [1] binding and this biological axis increases proliferation in bladder cancer cells [1,9-11]. This autocrine mechanism of growth may be associated with aggressive tumor growth and adverse clinical outcomes [1,12]. Here, we present a rare case of a leukemoid reaction and autocrine growth of bladder cancer induced by paraneoplastic production of G-CSF. We review the literature on the main clinicopathological aspects of this important, but rare, condition and review the biology of G-CSF in bladder cancer and its implications for diagnosis, management, prognosis and future research.

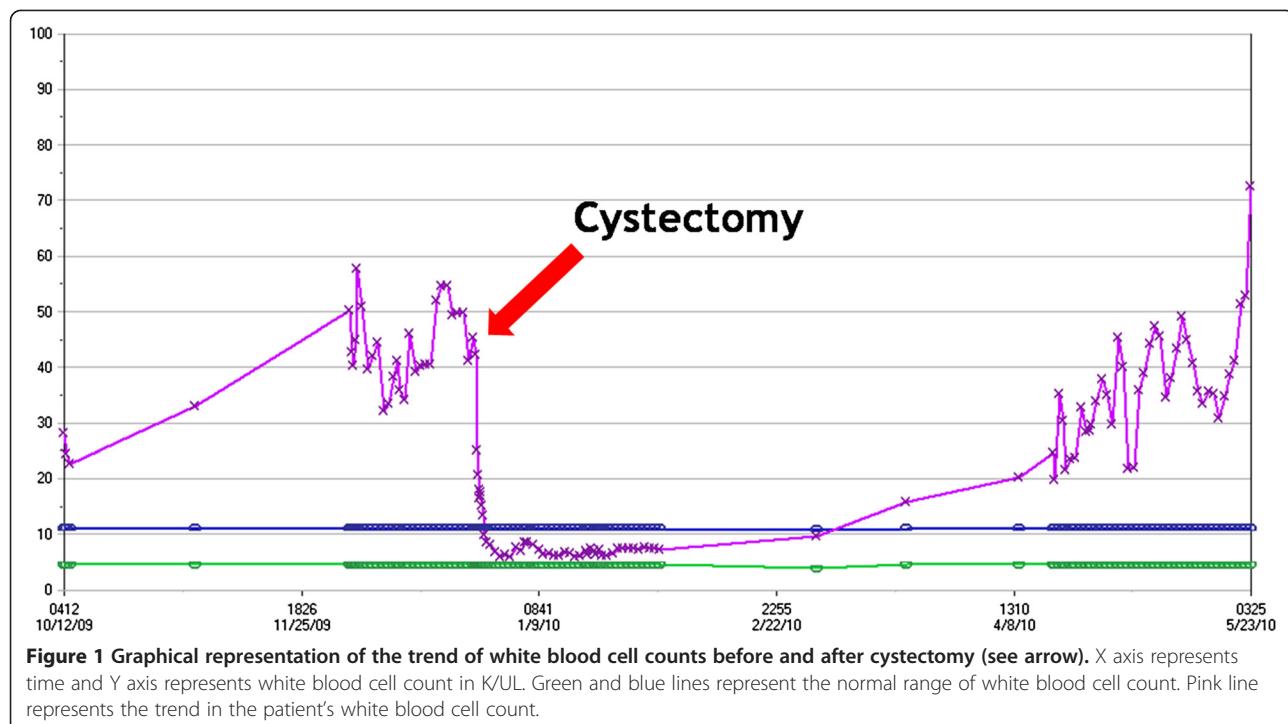
Case presentation

A 39-year-old non-cigarette-smoking Caucasian woman with hypertension, type 2 diabetes, diabetic retinopathy and neuropathy was diagnosed with muscle invasive high-grade urothelial carcinoma with squamous and glandular differentiation and necrosis by transurethral resection of her bladder tumor 4 months prior to presentation to our hospital. Three weeks prior to presentation in our clinic she had noted gross hematuria, daily low-grade fevers, night sweats and weight gain of 32kg. A physical examination revealed that her temperature was 39°C (102.2°F), pulse 110 per minute, respiratory rate 20 per minute, and blood pressure 150/83mmHg. Her examination was only remarkable for progressive anasarca. Laboratory studies revealed elevated white blood cell (WBC) count and absolute neutrophil count (ANC; peak WBC 57.8K/UL, peak ANC 43.24K/UL), leukocyte alkaline phosphatase score 295, erythrocyte sedimentation rate (ESR) >140mm/hour, C-reactive protein (CRP) 29.5mg/dL, and antineutrophil antibody (ANA) titer 320. She also had nephrotic range proteinuria with 24-hour urine protein of 14.65g and a serum creatinine of 3.27mg/dL (Table 1 and Figure 1). On admission, her urine grew greater than 100,000 *Aerococcus urinae* for which she was treated with a course of ceftriaxone. Repeat urine and blood cultures were

Table 1 Comparison of laboratory values before and after cystectomy

Laboratory values	Before cystectomy	After cystectomy
Total white blood cell (K/UL)	Peak 57.8	Peak 6.8
Absolute neutrophil count (K/UL)	Peak 43.24	Peak 5.05
Leukocyte alkaline phosphatase score	295	-
Erythrocyte sedimentation rate (mm/hour)	>140	45
C-reactive protein (mg/dL)	29.5	6.89
Antineutrophilic antibody	320	<80
24-hour urine protein (g)	14.65	5.25
Serum creatinine (mg/dL)	3.27	1.30

negative but she had continued elevation of her WBC count. The results of anti-double stranded deoxyribonucleic acid (DNA) antibody, rheumatic factor, anti-SSA, anti-SSB, anti-glomerular basement membrane, myeloperoxidase, proteinase 3, cytoplasmic antineutrophil cytoplasmic antibody, perinuclear antineutrophil cytoplasmic antibody, hepatitis panel, and human immunodeficiency virus antibody tests were negative. Serum protein electrophoresis, creatine phosphokinase, quantitative immunoglobulin assay, complement levels and chest radiograph were all normal. Hence, symptoms, signs and laboratory studies were negative for an infectious etiology (including a tagged WBC scan) and autoimmune/rheumatological disease. A bone scan and positron emission tomography scan ruled out metastasis (Figure 2). A computed tomography (CT) scan showed a large nodular bladder mass (Figure 3). The fevers and night sweats were attributable to her malignancy and her weight gain/anasarca was related to her nephrotic syndrome. After her medical condition was optimized, she underwent a radical cystectomy, bilateral pelvic lymph node dissection, and ileal conduit urinary diversion along with total abdominal hysterectomy/bilateral salpingo-oophorectomy. Immediately after removal of her bladder tumor, her presenting symptoms and laboratory values improved remarkably. Fevers and night sweats completely resolved. Her WBC count was 6.8K/UL, ANC 5.05K/UL, ESR 45mm/hour, CRP 6.89mg/dL, ANA <80, 24-hours urine proteins of 5.25g, and serum creatinine 1.30mg/dL (Table 1). Histopathology revealed poorly differentiated high-grade transitional cell carcinoma invading the entire bladder wall and the myometrium of the lower uterine segment along with vascular invasion without metastasis. Her pathologic staging was pT4a,N0,M0 (Figure 4). In addition, to further evaluate her nephrotic syndrome, a renal ultrasound-guided biopsy was done. Her kidneys were of normal size. The biopsy showed diabetic nephropathy with extensive nodular glomeruli sclerosis, moderate interstitial fibrosis and mild focal



chronic inflammation, mild tubular atrophy, and severe arteriolar hyalinosis. She responded to hemodialysis and her serum creatinine trended down to 1.30mg/dL with resolution of anasarca.

The patient refused adjuvant chemotherapy and returned 3 months following her cystectomy with an elevated WBC (peak WBC 45.3K/UL) and serum creatinine 1.90. A CT scan showed multiple pelvic masses with abdominal lymphadenopathy and a large right pelvic mass in close approximation to the area of the sigmoid colon causing obstruction. Her serum G-CSF level was 406.6pg/mL (normal <39.1pg/mL) confirming the diagnosis of paraneoplastic leukemoid reaction. She underwent a diverting colostomy to relieve symptoms of bowel obstruction. However due to poor performance status, renal failure and the development of a colovaginal fistula, she was placed on palliative care and expired soon thereafter.

Discussion

This rare case of urothelial carcinoma of the bladder had several unusual features including young age at presentation, marked leukemoid reaction, elevated G-CSF levels (ten times the normal limit), nephrotic range proteinuria and very rapid progression of disease to death within 6 months of the onset of the leukemoid reaction. Urothelial carcinoma in this case showed squamous differentiation which has been reported to have elevated G-CSF levels [4-6]. A possible mechanism for development of leukemoid reaction and rapid metastatic spread of the tumor is that the high-grade urothelial carcinoma harbored

both paraneoplastic production of G-CSF and expression of functional G-CSFR. This may have stimulated the bone marrow with increased myelopoiesis and leukemoid reaction. The stimulation of tumor growth may have been due to the expression of functional G-CSFR by the neoplastic cells, creating a positive feedback cycle. This mechanism has been demonstrated in *in vitro* studies [1,9,11]. Consistent with this mechanism, the above unusual features normalized after removal of the tumor (Table 1 and Figure 1). Similar post-surgical normalization of the leukemoid reaction and G-CSF has been previously observed in bladder cancer [5,13]. Three months later, with recurrence of tumor, there was elevated WBC and G-CSF. Hence, G-CSF could be used to follow the evolution of cancer and as a potential future prognostic marker. Of note, this case also exhibited nephrotic range proteinuria which improved after tumor removal, thereby clinically seemed to be paraneoplastic, but biopsy showed features of diabetic nephropathy. Minimal-change glomerulopathy associated with urothelial carcinoma has been rarely reported in the literature [14,15]. However, this may have been masked by her pre-existing diabetic nephropathy.

In the present case, as the tumor recurred and grew in her pelvic cavity, the concentration of serum G-CSF was elevated with an associated increase in her WBC count. These findings support the hypothesis that the bladder cancer cells producing G-CSF may grow more rapidly or that the G-CSF production exhibited by the tumor cells may promote the malignant progression. We thus undertook a comprehensive review of all studies and similar

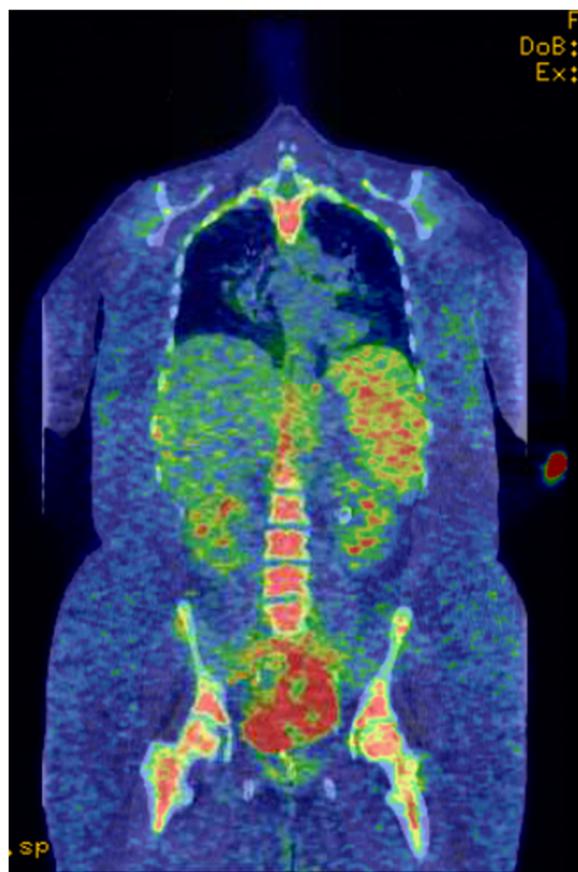


Figure 2 Positron emission tomography scan showing metabolic activity only in the bladder, prior to cystectomy. There is no evidence of regional or distant metastasis. There is slight increased activity in the skeletal system which is probably secondary to a marrow stenting agent.

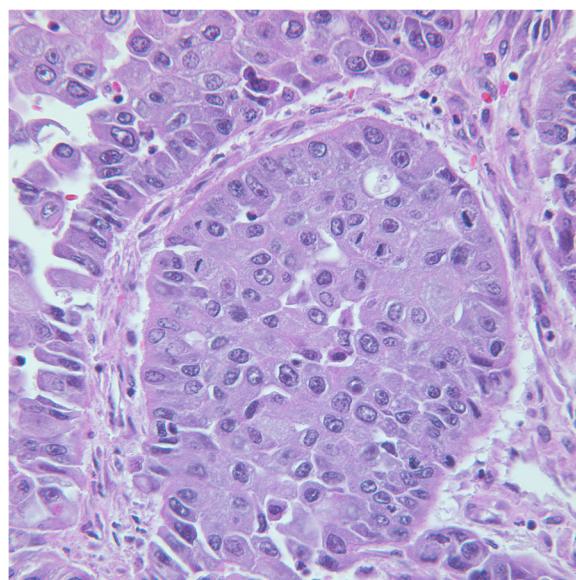


Figure 4 Histological examination of the cystectomy specimen showing poorly differentiated high-grade transitional cell carcinoma. (Hematoxylin and eosin section, original magnification $\times 220$).

cases reported to date to explore the above hypothesis. It is also probable that the patient had micrometastatic disease, undetectable by currently existing diagnostic modalities, prior to surgery. Rapid development of metastases within 3 months after surgery may have been invigorated by G-CSF secretion and activation of the G-CSF/G-CSFR biological axis stimulating tumor growth.

A review by Turalic *et al.* showed that the average age of these patients was 70.1 years and the male to female ratio was 3:2. The WBC was as high as 181,800/mL (average 66,758/mL) and there was an average G-CSF level of 203pg/mL with a mean survival of 5.9 months following surgical therapy in these patients. However, those who received adjuvant chemotherapy reported survivals of >20 months [4]. An observational study in Japanese patients by Mizutani *et al.* in 1995 showed that 9.2% of bladder tumors had elevated G-CSF levels (mean G-CSF of 328), which positively correlated with an increase in grade and progression of stage of cancer, more so in patients with distant metastasis. Furthermore, patients with bladder cancer without elevated G-CSF had better disease-specific survival rates at 5 years follow-up when compared with those with an elevated level of serum G-CSF [8].

The mechanism responsible for overexpression of G-CSF in bladder cancer is yet to be clearly elucidated. Possibilities include rearrangement of the G-CSF gene occurring within one of the alleles [16] and intrinsic activation of nuclear factors that work on the promoter region of the G-CSF gene [17]. Treatment with phorbol ester, interleukin-1 beta, or interferon gamma increased



Figure 3 Computed tomography scan of the pelvis showing large nodular bladder mass with heterogeneous enhancement, prior to cystectomy. Exchange are most extensive along the right bladder wall though is also seen along the left bladder wall and fundus.

the level of G-CSF, granulocyte-macrophage colony-stimulating factor, and macrophage colony-stimulating factor in an *in vitro* bladder cancer cell line study. This capability to respond to various stimuli implies that numerous regulatory pathways may be involved in the production of cytokines [18]. Although the mechanism of G-CSF gene overexpression requires further investigation, Chakraborty *et al.* conducted a series of *in vitro* studies which have shown that G-CSFR specifically signals for beta-1 integrin expression, adhesion and invasion of bladder cancer cells which could promote metastasis [19]. Furthermore, they demonstrated that the G-CSF/G-CSFR biological axis facilitated survival and growth of bladder cancer cells and stimulated STAT3-dependent survivin expression [11]. The histogenesis of transitional cell carcinoma of the bladder is uncertain, although different theories have been proposed. Some authors have suggested that a metaplastic phenomenon with various degrees of differentiation may be responsible for the malignant transitional cell G-CSF production [20]. In addition, this theory is supported further by the remarkable potential of the transitional epithelium and transitional cell carcinoma to differentiate into several lines [21]. The frequent finding of both squamous and glandular differentiation has long been observed in transitional cell carcinoma. Furthermore, the presence of neuroendocrine (small cell) differentiation has also been reported [22]. Hematopoietic differentiation of transitional cell carcinoma, resulting in the acquirement of G-CSF production and G-CSFR expression, is another possibility supported by our observations. Lastly, G-CSF may be released from the foci of tumor necrosis [7]. Hence, G-CSF could be a potential screening tool to predict aggressive cancer and hence pursue an early aggressive treatment modality.

G-CSF administration has been in vogue as an adjunct to the management of granulocytopenias, either with or without prior chemotherapy [23-26]. Subsequent studies have further confirmed the efficacy of G-CSF in conjunction with systemic chemotherapy for patients with advanced bladder cancer [27,28]. A few studies recommend G-CSF administration to enhance the tumor-specific cytotoxicity of chemotherapy [29,30], to allow dose intensification of chemotherapy [31-33] with low myelotoxicity and mucosal toxicity levels [31]. In contrast, Perez *et al.* reported a case of rapid clinical deterioration and leukemoid reaction after treatment of urothelial carcinoma with G-CSF along with chemotherapy [7]. Recombinant G-CSF may have both direct and indirect stimulatory effects on the growth of bladder cancer cells *in vitro*. It may stimulate the growth of the residual tumor cells after chemotherapy while it is being used clinically to combat myelosuppression induced by anti-tumor chemotherapy in bladder cancer [34]. Due to controversial reports, at this time, we can only advocate

that all bladder cancers should be tested for the expression of G-CSFR before using G-CSF, because there is the possibility of growth stimulation by G-CSF if they have G-CSFR on their cell surfaces. With a better understanding of the biology, this autocrine growth signal could be a potential target for therapy in future.

Conclusions

On the basis of this case experience and review of literature we conclude that there is a distinct and highly aggressive subtype of bladder cancer which secretes G-CSF and expresses G-CSFR inducing an autocrine growth cycle. Hence such patients presenting with leukemoid reaction should be tested for G-CSF/G-CSFR biological axis. Moreover, G-CSF could be a potential neoplastic marker as it can follow the clinical course of the underlying tumor and thus be useful for monitoring its evolution. Neoadjuvant chemotherapy should be considered as the aggressive nature of these tumors appears to overwhelm these patients before they receive adjuvant treatments. With a better understanding of the biology, this autocrine growth signal could be a potential target for therapy in future.

We are currently planning to conduct a preliminary study to identify patients with such bladder cancer, not only to estimate its prevalence but also to assess the variation in G-CSF levels in response to various therapies (surgery, chemotherapy, radiotherapy). In addition we will evaluate G-CSF as a prognostic marker and as a tool to predict a more aggressive biological phenotype of invasive bladder cancer. Future studies will be necessary to determine the regulatory effects of G-CSF on bladder cancer cells.

Consent

Written informed consent was obtained from the patient's next of kin for publication of this case report and any accompanying images. A copy of the written consent is available for review by the Editor-in-Chief of this journal.

Abbreviations

ANA: Antineutrophilic antibody; ANC: Absolute neutrophil count; CRP: C-reactive protein; CT: Computed tomography; ESR: Erythrocyte sedimentation rate; G-CSF: Granulocyte colony-stimulating factor; G-CSFR: Granulocyte colony-stimulating factor receptor; WBC: white blood cell.

Competing interests

The authors declare that we have no competing interests.

Authors' contributions

AKLK drafted the manuscript and prepared the figures. PW, MM and JH reviewed and amended the manuscript. All authors read and approved the final manuscript.

Acknowledgements

The authors would like to thank Dr Ivan Damjanov, Pathologist at University of Kansas Medical Center, Kansas for providing biopsy descriptions and picture (Figure 4).

Author details

¹Division of Hematology and Oncology, University of Kansas Medical Center, Kansas City, Kansas, USA. ²Department of Urology, University of Kansas Medical Center, Kansas City, Kansas, USA.

Received: 20 November 2013 Accepted: 3 March 2014

Published: 13 May 2014

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doi:10.1186/1752-1947-8-147

Cite this article as: Kasi Loknath Kumar et al.: Leukemoid reaction and autocrine growth of bladder cancer induced by paraneoplastic production of granulocyte colony-stimulating factor – a potential neoplastic marker: a case report and review of the literature. *Journal of Medical Case Reports* 2014 **8**:147.