

## Identifying Signatures of Selection Related to Comb Development

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The aim of this study was to identify genes involved in comb development to provide insights into the molecular mechanism of chickens' comb formation. Fixation index ( $F_{ST}$ ) and average number of base differences ( $\pi$ ) of males with large and small combs were calculated based on whole-genome resequencing data. Chromosome regions with larger  $F_{ST}$  values and smaller  $\pi$  were considered candidate selection regions. Through further annotation of gene functions and pathways, we sought to screen possible selected genes associated with comb development. By screening whole genome resequencing data,  $F_{ST}$  and  $\pi$  were calculated using a 40 Kb sliding window strategy and eight regions were identified. Quantitative trait loci (QTL; *FOX1* gene) related to comb length were found on chromosome 1. QTL (*GLPIR*, *BTBD9*, *MIR6633*, and *MDGAI* genes) related to comb weight were found on chromosome 3. QTL (*ALDH1A1*, *TMCI*, and *ANXA1* genes) associated with comb area were found on the Z chromosome. Nineteen genes, Wnt signaling pathway and neuroactive ligand-receptor interaction signaling pathway directly or indirectly related to comb growth and development were found through functional annotation and GO analysis. Among the selected genes *LYN*, *GLPIR*, *FOX1*, *TBK1*, *STRAP*, *ST6GALNAC*, and Wnt signaling pathways were related to immunity. *MDGAI*, *BTBD9*, *MTSSI*, *SrGAPs*, and neuroactive ligand receptor interaction signaling pathways related to neural function were screened. *ALDH1A1*, *ANXA1*, *THBS*, *HIF-1 $\alpha$* , and *ACTN1* genes were related to heat dissipation. Among the selected genes *FOX1*, *MDGAI*, and *ANXA1* associated with immunity, neurological function, and heat dissipation function coincided with genes affecting the length, weight, and area of the comb. Comprehensive analysis suggested that comb development was due to multiple genes and signaling pathways.

**Key words:** candidate genes, comb, genetic differentiation, selection signatures, whole-genome resequencing

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### Introduction

Many avenues for selection in poultry breeding provide insights into the future selection of elite progeny. Among these, the comb is an important and reliable parameter for selection within a variety or breed. A longer redder comb in red jungle fowl males is preferentially chosen by females (Johnsen and Zuk, 1996). Comb color and size in grouse are indicators of T-cell-mediated immune responses (Mougeot, 2008) and the comb is generally an indicator of health status, disease resistance, and nutritional status in flocks of red

jungle fowl (Zuk *et al.*, 1990; Johnson *et al.*, 1993; Mukhtar and Khan, 2012). Also, the comb is associated with reproductive performance and bone mass (Wright *et al.*, 2008). Comb size and comb weight are associated with total egg production, egg number, and mean egg weight in chickens (Rydmel, 2010). In China, comb development is an important factor associated with consumer acceptance and price of live and frozen chickens. In breeding programs, it is common to eliminate the chickens with smaller combs and select those with larger combs. Therefore, there is a strong natural and artificial selection for comb size.

Selection leads to specific changes in the patterns of variation among selected loci and in the neutral loci linked to them. These genomic footprints of selection, known as selection signatures, and can be used to identify loci that have been subjected to selection. Based on genome-wide resequencing data, a large number of selected segments associated with domestication, fitness, and importance have been reported. Among these studies, research subjects include chicken (Elferink *et al.*, 2012; Guo *et al.*, 2016; Zhang *et al.*,

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2017; Lillie *et al.*, 2018).

Wright's fixation index ( $F_{ST}$ ) was initially developed by Lewontin and Krakauer (1973) to scan for signatures of selection. Negative selection usually decreases  $F_{ST}$ , while positive selection causes it to increase (Barreiro *et al.*, 2008). Genomic regions under different selection or selection pressure eventually develop obvious genetic differences. Genomic regions with high levels of genetic differentiation can be identified by comparing  $F_{ST}$  values between distinct populations (Vitalis *et al.*, 2001; Guo *et al.*, 2016; Lillie *et al.*, 2018).

Nucleotide variation, in the form of  $\pi$  (Tajima, 1983), can be used to quantify the degree of genetic differentiation among populations based on differences in allele frequencies. Both  $F_{ST}$  and  $\pi$  statistics are useful for the detection of selection signatures (Lillie *et al.*, 2018).

This study was designed to compare male chicken with large and small combs. Based on whole-genome resequencing data and a public database, the selection signal for comb development was detected by comparing  $F_{ST}$  and  $\pi$  values between two groups. By comparing the genetic differentiation between two groups we expect to identify the genes related to comb development. This research could be useful in understanding the underlying mechanisms implicated in comb development and would contribute to more efficient breeding of chickens with larger combs.

## Materials and Methods

### Ethics Statement

All animal experiments were approved by the Animal Care and Use Committee at the Institute of Poultry Science, Chinese Academy of Agricultural Science (Approval ID: S20160605) and conducted at the institute. All of the experiments followed relevant guidelines and regulations set by the Ministry of Agriculture of the People's Republic of China.

### Sampling

All males used in the present study were sourced from a Partridge Shank chicken breeding line with single comb in Poultry Science, Chinese Academy of Agricultural Science. All chickens were produced under the same conditions and raised using a standardized feeding method with free access to water. To obtain males with different comb sizes at 9 weeks of age for sequencing, comb areas and heights of 200 males were measured using the software for automatic measurement of comb development parameters (registration number: 2015SR071422). The means and standard devia-

tions of comb areas and comb heights was  $2291.90 \pm 481.30 \text{ mm}^2$  and  $23.06 \pm 6.61 \text{ mm}$ , respectively.

Based on those results, 22 males with bigger combs (BC) and 19 males with smaller combs (SC) were selected for further analysis. The thresholds for the BC group were comb areas of more than  $2000 \text{ mm}^2$  and comb height of more than 34 mm, while those for the SC group were comb areas less than  $1600 \text{ mm}^2$  and comb height of less than 20 mm (Table 1).

### Re-sequencing of Chicken Samples, Mapping and SNP Calling

Blood samples were obtained from the wing vein. Genomic DNA was extracted from blood samples using the standard phenol/chloroform method. From total population ( $n=200$ ), the bigger comb population (BC;  $n=22$ ) and the smaller comb population (SC;  $n=19$ ) were extracted before pooling the DNA of each population. Two pooled samples (BC and SC) were separately sequenced using a next-generation sequencing technique on the Illumina HiSeq 2000 platform. A total of 140.4 Gb of pair-end reads of 100 bp were obtained. The following quality control criteria were implemented by removing 1) any reads with  $\geq 10\%$  unidentified nucleotides (N); 2) any reads with  $>10$  nt aligned to the adapter sequence, allowing  $\leq 10\%$  mismatches; 3) any reads with  $> 50\%$  bases having phred quality  $< 5$ ; 4) putative PCR duplicates generated by PCR amplification in the library construction process. A total of 133.6 Gb high quality paired-end reads was mapped to the chicken genome (*Gallus gallus*-5.0, <http://hgdownload.soe.ucsc.edu/downloads.html#chicken>) using BWA (Li and Durbin, 2009). Depth of reads of BC and SC populations were  $55.41 \times$  and  $53.2 \times$ , respectively, and  $> 90\%$  coverage of the chicken genome was retained for single nucleotide polymorphism (SNP) calling via the Genome Analysis Toolkit (GATK; McKenna *et al.*, 2010). NGS data has been submitted to SRA in NCBI (SRR10303810 and SRR10303811). The BioProject accession number is PRJNA576318 and the Biosample accession number is SAMN12990250.

### Analysis of Selection Signatures

Two population genetic indices were estimated from the sequence data. Firstly, nucleotide diversity ( $\pi$ ) was used to measure the degree of polymorphism within the population. Secondly,  $F_{ST}$  was determined as a measure of population differentiation due to genetic structure. In our analyses,  $\pi$  was calculated across 40,000 bp windows for each population pool using Popoolation (v1.2.2; Kofler *et al.*, 2011a) with the following parameter set: window size of 10 K, step size of 5

Table 1. Comb area and circumference of sampled chickens

Sample	BC group		SC group	
	means	SD	means	SD
Comb areas ( $\text{mm}^2$ )	2736.9	218.95	1846.89	166.22
Comb heights (mm)	40.34	3.63	19.65	1.75

Note: BC: Big comb; SC: Small comb

K, minimum allele count of 2, minimum base quality of 20, minimum coverage of 10 and maximum coverage of 200, and the log value of  $\pi$  ratios was calculated.  $F_{ST}$  was calculated using PoPoolation2 (v1.201; Kofler *et al.*, 2011b), with same parameter set as for  $\pi$ . Samtools (Li *et al.*, 2009) was used to generate input files for PoPoolation2.  $F_{ST}$  was calculated using the standard equation according to the principles of population genetics (Hartl and Clark, 2007).  $F_{ST} = \frac{P_{i\_total} - P_{i\_within}}{P_{i\_within}}$ , where  $P_{i\_within} = \frac{(P_{i\_population1} + P_{i\_population2})}{2}$ , and  $P_i = 1 - fA^2 - fT^2 - fC^2 - fG^2$  with  $fN$  being the frequency of nucleotide N (i.e. A, T, C or G),  $P_{i\_total}$  is the total  $P_i$  for which allele frequencies in both populations are averages and  $P_i$  is calculated as above. The  $F_{ST}$  values were Z-transformed as follows:  $Z(F_{ST}) = \frac{F_{ST} - \mu F_{ST}}{\sigma F_{ST}}$ , where  $\mu F_{ST}$  is the mean  $F_{ST}$ , and  $\sigma F_{ST}$  is the standard deviation of  $F_{ST}$  (Karlsson *et al.*, 2007). Putatively selected regions were located in fully overlapping windows with an extremely low  $\pi$  value (top 5% level) and extremely high  $Z(F_{ST})$  values (top 5% level).

#### Candidate Gene Annotation for Selected Regions, GO Analysis, and Pathway Analysis

We annotated genes in selected genomic regions using the chicken genome as a reference via the UCSC genome browser (<http://genome.ucsc.edu/>). The genes putatively under selection were submitted to David (<http://david.ncifcrf.gov/>) for enrichment analysis of the Gene Ontology (GO) and KEGG pathways. In addition, the chicken quantitative trait loci (QTL) database (<https://www.animalgenome.org/cgi-bin/QTLdb/GG/index>) was systematically tested, and analyzed to detect whether comb height, comb area, comb weight, comb length, early maturity, body weight growth, and other traits were enriched in selected regions.

## Results

### Candidate Selected Region

The average  $Z(F_{ST})$  was 2.202 with a standard deviation of 0.614. It can be seen from the  $Z(F_{ST})$  that the populations with large and small sized combs exhibited genetic differentiations in some chromosomal regions, with the most significant interval located on chromosome 15 (see Fig. 1a). The genomic nucleotide diversity ( $\pi$ ) of the chicken population was determined using the same size sliding window strategy. The average  $\pi$  was 1.533, and the standard deviation was 0.414. Whether these intervals were related to

comb size was determined by comparing the overlapping areas between the smaller interval of  $\pi$  and the larger interval of  $Z(F_{ST})$  (Fig. 1b). In order to select the extreme candidate selected region, the overlapping region of the interval of  $Z(F_{ST}) \geq 5$  and the interval of 200 with the smallest negative value of  $\pi$  was selected, resulting in eight candidate regions, including 19 genes.

### Enrichment of Related Trait QTL in Candidate Selected Regions

To verify that the candidate regions were associated with comb length, comb thickness, comb area, and comb weight, the known QTL contained in the eight candidate selected regions were searched for in the Chicken QTL database (<https://www.animalgenome.org>). QTL (*FOX1* gene) related to the length of the comb were found on chromosome 1, QTL (*GLP1R*, *BTBD9*, *MIR6633*, and *MDGA1* genes) related to comb weight were found on chromosome 3, and QTL (*ALDH1A1*, *TMC1*, and *ANXA1* genes) associated with the area of the comb were found on the Z chromosome (Table 2). These results indicated that the QTL associated with the comb development was enriched in the candidate region and also suggested that the method for detecting the selection signal was reliable.

### GO Enrichment Analysis and Pathway Analysis of Candidate Genes

A total of 26 genes were identified by genetic annotation of candidate regions using the online UCSC genome browser tool (Fig. 1). To clarify the function of these genes, PANTHER was used for functional annotation and GO analysis. Twenty genes were directly or indirectly related to comb growth and development (Table 3). Most of these genes are involved in cell growth and development or important basic signaling pathways. For example, *LYN*, *SRGAP1*, *THBS1*, *WWC1*, and *MTSSI* are involved in cell migration and receptor binding. *MDGA1*, *ST6GALNAC5*, *PLA2G4A*, *ADAM10*, *ZDHHC21*, *ACO1* are involved in Golgi action. The growth and development of the comb is inseparable from cell migration, receptor binding, and the role of the Golgi apparatus. The main function of the Golgi apparatus is to process, classify, and package proteins synthesized by the endoplasmic reticulum, which are then sent to specific parts of the cell or secreted outside the cell.

GO enrichment analysis showed that *LYN*, *WNT9A*, *WWOX*, and *CSNK2A* were involved in Wnt signaling path-

Table 2. Eight regions of selection

Chromosome	Gene	Region	$\pi$	Z ( $F_{ST}$ )	Comb QTL
1	<i>FOXO1</i>	170581722-170644997	1.1705	1.6818	Comb length
3	<i>GLP1R</i>	29377290-29456632	1.1655	3.6490	Comb weight
3	<i>BTBD9</i>	29594123-29704099	1.3132	1.9223	Comb weight
3	<i>MIR6633</i>	80493162-80493272	1.4758	2.2259	Comb weight
3	<i>MDGA1</i>	30023079-30150365	1.1434	1.7914	Comb weight
Z	<i>ALDH1A1</i>	35894470-35947479	1.8474	2.1396	Comb area
Z	<i>TMC1</i>	35808065-35866858	1.9209	2.1256	Comb area
Z	<i>ANXA1</i>	36026282-36040351	2.2242	2.0813	Comb area

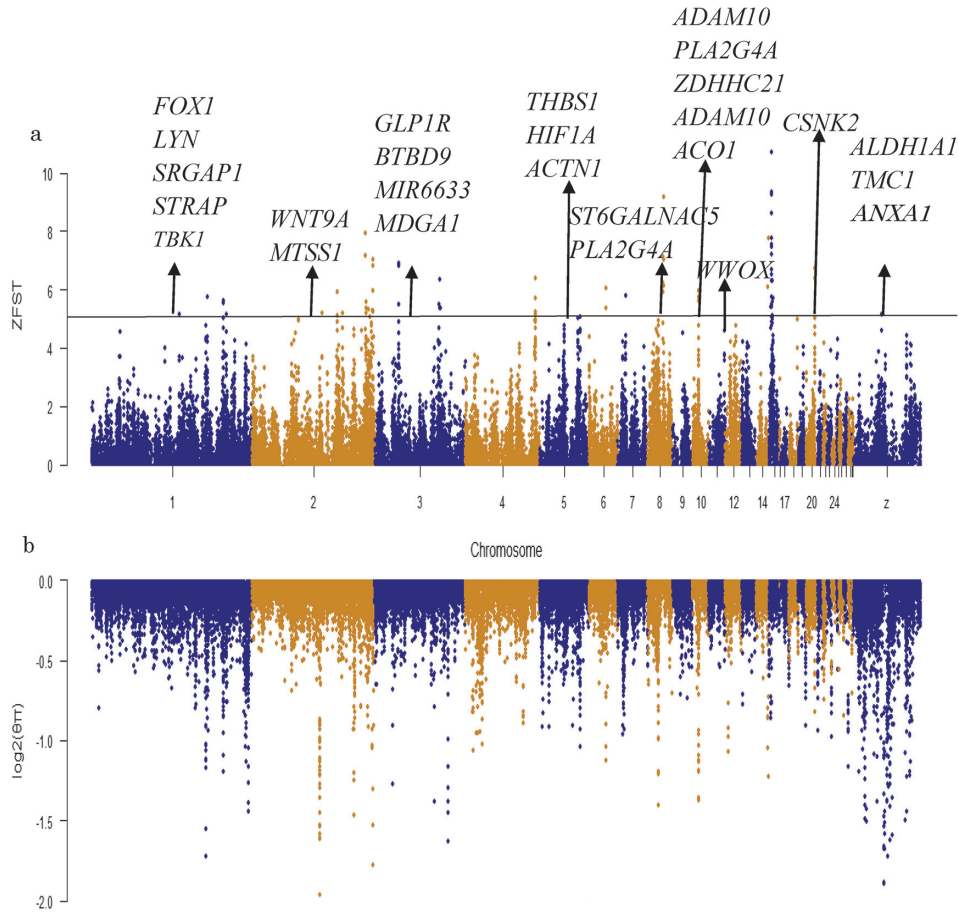


Fig. 1. **The distribution of Z ( $F_{ST}$ ) and  $\pi$  on chromosome.** Note: a. The positive end of the Z ( $F_{ST}$ ) plotted on chromosomes. A dashed horizontal line indicates the cut-off Z ( $F_{ST}$ )=5. b. The negative end of  $\pi$  plotted on chromosomes. A dashed horizontal line indicates the cut-off for 200 smallest  $\pi$ .

way. The Wnt signaling pathway is a complex network of protein action that, while commonly functional in embryonic development and cancer, is also involved in the normal physiological processes of adult animals. The opening or closing of this pathway controls the expression of a large number of growth and metabolism-related genes (Xu *et al.*, 2015).

The KEGG pathway analysis of these candidate genes was performed using David 6.7, and the neuroactive ligand-receptor interaction (gga04080) signaling pathway was enriched. Five genes, *AGTR1*, *TACR3*, *TRPV1*, *PTH1R*, and *GLP1R*, participate in this signal path.

### Discussion

A total of eight selection regions were identified in this study, including a number of QTL affecting comb length, area, and weight. QTL (*FOX1* gene) related to comb length were found on chromosome 1, QTLs (*GLP1R*, *BTBD9*, *MIR6633*, *MDGAI* genes) related to comb weight were

found on chromosome 3, and QTLs associated with comb area (*ALDH1A1*, *TMC1*, *ANXA1* genes) were found on the Z chromosome. Functional annotation and GO analysis were performed, and 19 genes were directly or indirectly related to comb development.

Among the selected genes *LYN*, *GLP1R*, *FOX1*, *TBK1*, *STRAP*, *ST6GALNAC*, and Wnt signaling pathways were related to immunity. *MDGAI*, *BTBD9*, *MTSSI*, *SrGAPs*, and neuroactive ligand receptor interaction signaling pathways related to neural function were screened. Genes related to heat dissipation function (*ALDH1A1*, *ANXA1*, *THBS*, *HIF-1 $\alpha$* , and *ACTN1*) were included in the selected regions. Among these 19 genes, some genes related to immunity and heat dissipation function, *FOXO1*, *MDGAI*, and *ANXA1*, coincide with genes affecting comb weight and area.

GO analysis showed that *LYN*, *WNT9A*, *WWOX*, and *CSNK2A* are involved in the Wnt signaling pathway. Its members have high homology and play important roles in regulating embryonic development, participating in cell pro-



Table 3. **GO term enrichment for gene related to comb development**

Chromosome	Gene	Biological process
1	<i>LYN, FOXO1</i>	cell migration (GO:0016477), receptor binding (GO:0005103), Golgi apparatus (GO:0005798)
1	<i>SRGAP1</i>	cell migration (GO:0016478)
1	<i>STRAP</i>	negative regulation of epithelial cell migration (GO:0010633), receptor binding (GO:0005104)
1	<i>TBK1</i>	phosphoprotein binding (GO:0051219),
2	<i>WNT9A</i>	Wnt signaling pathway (GO:0016056), iris morphogenesis (GO:0061073)
2	<i>MTSS1</i>	receptor binding (GO:0005102)
3	<i>MDGAI</i>	Golgi apparatus (GO:0005800)
5	<i>THBS1</i>	cell migration (GO:0016477), peptide cross-linking (GO:0018150)
5	<i>HIF1A</i>	iris morphogenesis (GO:0061072)
5	<i>ACTN1</i>	phosphoprotein binding (GO:0005794)
8	<i>ST6GALNAC5</i>	Golgi apparatus (GO:0005794)
8	<i>PLA2G4A</i>	Golgi apparatus (GO:0005794)
10	<i>ZDHHC21, ADAM10, ACO1</i>	Golgi apparatus (GO:0005794)
11	<i>WWOX</i>	Wnt signaling pathway (GO:0016057)
20	<i>CSNK2A</i>	Wnt signaling pathway (GO:0016055)
Z	<i>ANXA1</i>	peptide cross-linking (GO:0018149)

liferation and differentiation, as well as migration and polarity.

Among the genes involved in the Wnt signaling pathway, tyrosine protein kinase (*LYN*) may be involved in the body's immunity. The expression of this gene is regulated by the methylation of the promoter CpG site. The high methylation of this gene is important for maintaining normal cell growth and proliferation. *LYN* has an indispensable role in immunoglobulin-mediated signaling, particularly in establishing B cell tolerance (Hibbs *et al.*, 1995).

As a transcriptional regulator, *FOXO1* plays an important regulatory role in animal growth and development through transcriptional regulation and signal transcription. *FOXO1*, as a potential factor, has an important relationship with skeletal muscle differentiation and myotube formation in myoblast differentiation (Yang *et al.*, 2008). Through genome-wide association analysis, four SNPs in the upstream and downstream flanking regions of the chicken *FOXO1* gene were significantly correlated with growth traits such as body weight and average daily gain (Wang, 2012). In our study, the *FOXO1* gene was identified to have a potential role in comb length QTL on chromosome 1.

The comb structure is divided into three layers: epidermis, dermis, and central layer. The epidermis is divided into stratum corneum and germinal layer. Merck's cells are contained in the germinal layer. Merkel cells establish prominent connections with nerve endings and play a key role in sensing external stimuli (Hitchcock *et al.*, 2014). In the epidermis, collagen fiber bundles, adipose tissue, many larger arteries, and some nerves are arranged vertically. Our researches showed that MAM domain glycosylphosphatidylinositol anchor 1 (*MDGAI*) and neuroactive ligand receptor

interaction signaling pathways were related to neural function. *MDGAI* genes were identified to have a potential role in comb weight.

*MDGAI*, a unique cell surface glycoprotein, is similar to Ig-containing cell adhesion molecules that influence neuronal migration and process outgrowth. *MDGAI* acts cell autonomously to control the migration of *MDGAI*-expressing superficial layer cortical neurons (Takeuchi and Dennis, 2006). The *MDGAI* gene affects the development of the nervous system by changing the neuronal migration pathway and participates in the etiology and pathogenesis of schizophrenia (Gangwar *et al.*, 2017).

The KEGG pathway analysis enriched the neuroactive ligand-receptor interaction signaling pathway, and the five genes (*AGTR1*, *TACR3*, *TRPV1*, *PTH1R*, and *GLPIR*) observed here were involved in this signaling pathway. The neuroactive ligand-receptor interaction signaling pathway is a collection of all receptors and ligands on the plasma membrane that are associated with intracellular and extracellular signaling pathways. Therefore, this signaling pathway is interesting because there are not only differentially expressed genes in this pathway, but are also physiologically the most closely related to neurological function.

The structure of the comb and the blood vessels distributed in the comb play an important role in the heat dissipation process of the chicken. The larger the comb area, the better the heat dissipation in a warmer climate, while small combs are beneficial in a cooler climate. Our research showed that the genes (*ALDH1A1* and *ANXA1*) selected for the comb area are involved in heat dissipation. *ALDH1A1* is increased in obese patients, negatively regulating their heat production. *ALDH1A1* ping cells are called *Aldh1a14* thermogenic fat

cells. In vivo and in vitro experiments, all *ALDH1A1* enzyme activities are coordinated with retinoic acid (RA) production (Reichert *et al.*, 2011). Annexin A1 (also known as Annexin I, *ANXA1*) is a widely-functioning calcium and phospholipid binding protein, which participates in many cell activities such as anti-inflammatory reaction, cell differentiation and proliferation, cell death signal regulation, and apoptotic cell phagocytosis (Perretti and Solito, 2004; Sun *et al.*, 2016), inhibiting cell migration to the site of inflammation (Damazo *et al.*, 2011).

### Conclusions

Based on genome-wide resequencing data, combined with population genetic differentiation index  $F_{ST}$  and population genomic nucleotide diversity ( $\pi$ ) for system selection signal detection, a total of eight selection regions were identified. QTL (*FOX1* gene) related to the length of the comb were found on chromosome 1. QTL (*GLP1R*, *BTBD9*, *MIR6633*, *MDGA1* genes) related to comb weight were found on chromosome 3. QTL (*ALDH1A1*, *TMC1*, *ANXA1* genes) associated with the area of the comb were found on the Z chromosome. Nineteen genes directly or indirectly related to comb growth and development were found through functional annotation and GO analysis. Comb development is affected by multiple genes and signaling pathways. Among the selected genes *FOX1*, *MDGA1*, and *ANXA1* associated with immunity, neurological function, and heat dissipation function coincided with genes affecting the length, weight, and area of the comb.

### Conflicts of Interest

We certify that there is no conflict of interest with any financial and personal relationships with other people or organization regarding the material discussed in the manuscript.

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### References

- Barreiro LB, Laval G, Quach H, Patin E and Quintana-Murci L. Natural selection has driven population differentiation in modern humans. *Nature Genetics*, 40: 340–345. 2008.
- Damazo AS, SampBio AL, Nakata CM, Flower RJ, Perretti M and Oliani SM. Endogenous annexin A1 counter-regulates bleomycin-induced lung fibrosis. *BMC Immunology*, 12: 1–13. 2011.
- Elferink MG, Megens HJ, Vereijken A, Hu XX, Crooijmans RPMA and Groenen MAM. Signatures of selection in the genomes of commercial and non-commercial chicken breeds. *PLoS ONE*, 7: e32720. 2012.
- Gangwar SP, Zhong XY, Seshadrinathan S, Chen H, Machius M and Rudenko G. Molecular mechanism of MDGA1: regulation of neuroligin 2: neuroligin trans-synaptic bridges. *Neuron*, 94: 1132–1141. 2017.
- Guo X, Fang Q, Ma CD, Zhou BY, Wan Y and Jiang RS. Whole-genome resequencing of Xishuangbanna fighting chicken to identify signatures of selection. *Genetics Selection Evolution*, 48: 62. 2016.
- Hartl DL and Clark AG. Principles of population genetics. 4th ed. Sunderland: Sinauer Associates. 2007.
- Hibbs ML, Tarlinton DM, Armes J, Armes J, Grail D, Hodgson G, Maglitto R, Stacker SA and Ashley RD. Multiple defects in the immune system of Lyn-deficient mice, culminating in autoimmune disease. *Cell*, 83: 301–311. 1995.
- Hitchcock IS, Genever PG and Cahusac PMB. Essential components for a glutamatergic synapse between Merkel cell and nerve terminal in rats. *Neuroscience Letters*, 362: 196–199. 2014.
- Johnson K, Thornhill R, Ligon JD and Zuk M. The direction of mothers' and daughters' preferences and the heritability of male ornaments in red jungle fowl (*Gallus gallus*). *Behavioral Ecology*, 4: 254–259. 1993.
- Johnsen TS and Zuk M. Repeatability of mate choice in female red jungle fowl. *Behavioral Ecology*, 7: 243–246. 1996.
- Karlsson EK, Baranowska I, Wade CM, Salmon Hillbertz NHC, Zody MC, Anderson N, Biagi TM, Patterson N, Pielberg GR, Kulbokas EJ, Comstock KE, Keller ET, Mesirov JP, H von Euler, Kampe O, Hedhammar A, Lander ES, Andersson R, Andersson L and Lindblad-Toh K. Efficient mapping of mendelian traits in dogs through genome-wide association. *Nature Genetics*, 39: 1321–1328. 2007.
- Kofler R, Pandey RV and Schlotterer C. PoPoolation2: Identifying differentiation between populations using sequencing of pooled DNA samples (Pool-Seq). *Bioinformatics*, 27: 3435–3436. 2011a.
- Kofler R, Orozco-terWenge I P, Maio N De, Pandey RV, Nolte V, Futschik A, Kosiol C and Schlotterer C. PoPoolation: A toolbox for population genetic analysis of next generation sequencing data from pooled individuals. *PLoS ONE*. 6.doi: 10.1371/journal.pone.0015925. 2011b.
- Lewontin RC and Krakauer J. Distribution of gene frequency as a test of the theory of the selective neutrality of polymorphisms. *Genetics*, 74: 175–195. 1973.
- Li H and Durbin R. Fast and accurate short read alignment with Burrows–Wheeler transform. *Bioinformatics*, 25: 1754–1760. 2009.
- Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R and 1000 Genome Project Data Processing Subgroup. The Sequence Alignment/Map format and SAMtools. *Bioinformatics*, 25: 2078–2079. 2009.
- Lillie M, Sheng Z Y, Honaker C F, Andersson L, Siegel P B and Carlborg O. Genomic signatures of 60 years of bidirectional selection for 8-week body weight in chickens. *Poultry Science*, 97: 781–790. 2018.
- McKenna A, Hanna M, Banks E, Sivachenko A, Cibulskis K, Kernytsky A, Garimella K, Altshuler D, Gabriel S, Daly M and DePristo MA. The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. *Genome research*, 20: 1297–1303. 2010.
- Mougeot F. Ornamental comb colour predicts T-cell-mediated immunity in male red grouse *Lagopus lagopus scoticus*. *Naturwissenschaften*, 95: 125–132. 2008.
- Mukhtar N and Khan SH. Comb: An important reliable visual ornamental trait for selection in chickens. *World's Poultry*

- Science Journal, 68: 425–434. 2012.
- Pavlidis P, Zivkovic D, Stamatakis A and Alachiotis N. SweeD: likelihood-based detection of selective sweeps in thousands of genomes. *Molecular Biology and Evolution*, 30, 2224–2234. 2013.
- Perretti M and Solito E. Annexin 1 and neutrophil apoptosis. *Biochemical Society Transactions*, 32: 507–510. 2004.
- Reichert B, Yasmeeen R, Jeyakumar SM, Yang FP, Thomou T, Alder H, Duester G, Maiseyue A, Mihai G, Harrison EH, Rajagopalan S, Kirkland JL and Ziouzenkova O. Concerted action of aldehyde dehydrogenases influences depot specific fat formation. *Molecular Endocrinology*, 25: 799–809. 2011.
- Rydmeel S. Honesty of female ornaments in *Gallus gallus*. Thesis. Linköping University Swenden. 2010.
- Sun X L, Fang H, Zhang HW, Zhen YF, Yang Y and Li YK. Expression and significance of EphA7 and *ANXA1* in papillary thyroid carcinomas. *Journal of Clinical and Experimental Pathology*, 32: 552–555. 2016.
- Takeuchi A and Dennis DMO'Leary. Radial Migration of Superficial Layer Cortical Neurons Controlled by Novel Ig Cell Adhesion Molecule *MDGAI*. *Journal of Neuroscience*, 26: 4460–4464. 2006.
- Tajima F. Evolutionary relationship of DNA sequences in finite populations. *Genetics*, 105: 437–460. 1983.
- Vitalis R, Dawson K and Boursot P. Interpretation of variation across marker loci as evidence of selection. *Genetics*, 158: 181–1823. 2001.
- Wang SB. Effects of SNPs in the flanking region of chicken *FOXO1* gene on its transcriptional relation. Thesis. South China Agricultural University. 2012.
- Wright D, Kerje S, Brandstrom H, Schutz K, Kindmark A, Andersson L, Jensen P and Pizzari T. The genetic architecture of a female sexual ornament. *Evolution*, 62: 86–98. 2008.
- Xu Y, Yang Z, Yuan H, Li Z, Li Y, Liu Q and Chen J. PCD10 inhibits cell proliferation of multiple myeloma via the negative relation of the Wnt/BCL-9 signaling pathway. *PLoSOne*, 34: 747–754. 2015.
- Yang YJ, Bai L, Pang WJ and Yang GS. Comparison of *FOXO1* and *MyoD* mRNA Expression in muscle tissues among different pig breeds. *Chinese Journal of Biochemistry and Molecular Biology*, 24: 257–261. 2008.
- Zhang MM, Yang L, Su ZC, Zhu MZ, Li WT, Wu KL and Deng XM. Genome-wide scan and analysis of positive selective signatures in dwarf brown-egg layers and silky fowl chickens. *Poultry Science*, 96: 4158–4171. 2017.
- Zuk M, Thornhill R and Ligon JD. Parasites and mate choice in red jungle fowl. *American Zoologist*, 30: 235–244. 1990.