Candida albicans and candidalysin in inflammatory disorders and cancer

Jemima Ho, p Giorgio Camilli, James S. Griffiths, Jonathan P. Richardson, Nessim Kichik and Julian R. Naglik Centre for Host-Microbiome Interactions, Faculty of Dentistry, Oral & Craniofacial Sciences, King's College London, London, UK

doi:10.1111/imm.13255

Received 30 June 2020; revised 5 August 2020; accepted 14 August 2020. *Correspondence: Jemima Ho, Centre for Host-Microbiome Interactions, Faculty of Dentistry, Oral & Craniofacial Sciences, King's College London, London SE1 1UL, UK. Email: Jemima.ho@kcl.ac.uk Senior author: Julian Naglik

Summary

As our understanding of mycology progresses, the impact of fungal microbes on human health has become increasingly evident. *Candida albicans* is a common commensal fungus that gives rise to local and systemic infections, particularly in immunocompromised patients where it can result in mortality. However, *C. albicans* has also been quietly linked with a variety of inflammatory disorders, to which it has traditionally been considered incidental; recent studies may now provide new aspects of these relationships for further consideration. This review provides a novel perspective on the impact of *C. albicans* and its peptide toxin, candidalysin, on human health, exploring their contributions to pathology within a variety of diseases.

Keywords: Candida albicans; candidalysin; gut barrier; IL-17; mucosal disease.

Introduction

Candida albicans is a prevalent fungus that comprises part of the healthy human microbiota. Within such microbial communities, C. albicans often exists as a harmless commensal yeast in low-to-moderate numbers, likely kept in check by competing microbes and host immunity. Its ability to shift from commensal to infectious pathogen is of particular interest to the clinical understanding of Candida infections and remains incompletely understood. Current evidence suggests that pathogenic switching is primarily a consequence of immune compromise brought about by a variety of factors including microbial environment,¹ immune-suppressive drug treatment and pre-existing infection or disease.^{2–4} Indeed, immunocompromised patients are particularly susceptible and exhibit mucosal candidiasis of enhanced severity and frequency, with potential to progress to systemic candidaemia. This represents a significant clinical burden, with ~ 2 000 000 infections in HIV + patients and 700 000 total systemic infections recorded in 2017.5

In addition to conditions that occur as a result of persistent or severe *Candida* infection, such as oral and vulvovaginal candidiasis (VVC) or systemic candidaemia, an increasing number of seemingly unrelated diseases have also been reported to show association with *Candida* infection. Elevated incidence of candidiasis has been linked with periodontitis,^{6–9} inflammatory bowel disease (IBD),^{10,11} and skin^{12–14} and respiratory disorders,^{15–18} among others; however, the causal relationship in such circumstances remains unclear. Whilst weakened immunity occurring as a result of disease may certainly favour growth of opportunistic fungi, a role for *C. albicans* in perpetuating ongoing disease and promoting acute or chronic pathology may also warrant consideration, particularly in the context of its secreted toxin, candidalysin.

Candidalysin is a recently described cytolytic peptide exclusively secreted by pathogenic hyphal forms of *C. albicans.*¹⁹ Interestingly, candidalysin plays an important role in triggering innate antifungal immunity during infection,^{20,21} which is largely governed by neutrophil and interleukin (IL)-17 responses.^{22–25} This review will examine the variety of diseases associated with *C. albicans* infections and assess the role of this fungus and its toxin, candidalysin, in disease development and associated pathology.

Abbreviations: AD, atopic dermatitis; AMPs, antimicrobial peptides; CNS, central nervous system; ECM, extracellular matrix; EGFR, epidermal growth factor receptor; EMT, epithelial–mesenchymal transition; IBD, inflammatory bowel disease; IL-17, Interleukin; MMPs, matrix metalloproteinases; MS, multiple sclerosis; SAFS, severe asthma with fungal sensitization; VVC, vulvovaginal candidiasis

IL-17-mediated disorders

The IL-17 signalling pathway possesses critical roles in immune regulation, and aberrant function results in a range of diseases that share a common feature of chronic inflammatory-induced pathology. This is best defined by IL-17A, the most studied IL-17 family member, which has been shown to contribute to dermatitis,^{26,27} psoriasis,²⁸ IBD,^{29,30} arthritis,³¹ multiple sclerosis (MS),³² periodontal disease^{6,7} and systemic lupus erythematosus,³³ among other inflammatory diseases. Though much is still unknown about the mechanisms of IL-17-mediated pathology, its potent induction of antimicrobial peptides (AMPs), proinflammatory cytokines and downstream neutrophil recruitment^{34–37} is thought to contribute. Of particular interest is the central role of IL-17A in orchestration of antifungal defences. IL-17A and associated effector molecules and cells are potently induced in response to C. albicans infection, 3^{3E-41} with candidalysin accounting for robust induction of early and innate 'natural' Th17 cell-derived IL-17A.²⁰

Notably, elevated incidence or sensitization to C. albicans is often observed in a specific group of IL-17-mediated pathologies localized at mucosal and epithelial surfaces. These include periodontitis, $^{6-9}$ atopic dermatitis (AD), 14,42 psoriasis,^{43,44} IBD,^{10,11} mycotic keratitis⁴⁵ and severe asthma with fungal sensitization (SAFS).^{17,18} Moreover, evidence of improved disease outcomes following antifungal measures has been observed and suggests a role for C. albicans in contributing to disease pathology. Examples include tamoxifen-induced C. albicans inhibition to reduce severity of periodontitis in women,⁴⁶ as well as fluconazole treatment⁴⁷ or faecal microbiota transplantation (inhibiting *C. albicans* burdens)⁴⁸ to rescue ulcerative colitis symptoms in mice. There are also limited reports of improved C. albicans-related respiratory diseases following fluconazole administration^{15,49} (though Aspergillus species have been better studied in airways disease 50,51). It is likely that other examples could be found by investing more research into C. albicans in this context.

Furthermore, candidalysin can directly induce IL- 1β ,^{21,52} IL- $36^{53,54}$ and the NLRP3 inflammasome,^{55,56} central proinflammatory components known to significantly contribute to IL-17-mediated diseases.^{57–61} Together, these studies suggest a role and potential mechanisms for *C. albicans* in contributing to the elevated immune response and pathology observed in IL-17-mediated inflammatory diseases. However, the relationships are complex. Therapeutic blockade of IL-17, whilst beneficial in MS,⁶² arthritis⁶³ and psoriasis,⁶⁴ does not improve AD⁶⁸ and was shown to result in exacerbation of existing^{65,66} and even de novo⁶⁷ IBD pathology, as well as increased incidence of *Candida* infections.²⁹ Greater understanding is thus required to fully delineate the complex mechanisms underlying IL-17-mediated diseases and how *Candida* or indeed other fungal species may impact pathology.

A significant component of successful IL-17-mediated antifungal response is potent neutrophil recruitment and activation,^{39,69} which, at an early stage in infection, can be triggered by the presence of candidalysin.²⁰ Interestingly, whilst a robust neutrophil response functions to resolve $\operatorname{oral}^{21,22}$ and central nervous system (CNS)²⁵ C. albicans infections, neutrophils are found to drive pathology of VVC,^{52,70} Candida keratitis (CaK),⁷¹ systemic candidaemia⁷² and C. albicans-associated cystic fibrosis, with the latter arising from neutrophil-induced degradation of chitinase, suppressing host ability to protect against chitin containing C. albicans.⁷³ These studies suggest a delicate balance and complexity of antifungal neutrophil responses, which are likely dependent on multiple components. Continued research may determine the underlying factors leading to pathology in this context.

Barrier integrity and disease

Another component often compromised in disease and particularly in infection is the epithelium. Its integrity and function as a selectively permeable barrier are dependent on cohesive contacts between neighbouring epithelial cells. This is largely provided by E-cadherin, a transmembrane glycoprotein that forms binding pairs with that of neighbouring cells, which, clustering together at adherens junctions, are supported by the cytoskeleton to form a tight belt across the epithelium that fastens cells together.⁷⁴ In addition to structural defences, a healthy epithelium provides front-line induction of innate immune responses through release of alarmins, AMPs and immune cell chemokines, all of which are additionally compromised upon loss of barrier integrity. Notably, periodontal,⁷⁵⁻⁷⁷ gut⁷⁸⁻⁸⁰ and skin⁸¹⁻⁸³ disorders are commonly associated with the loss of E-cadherin, resulting in enhanced barrier permeability and inflammatory pathology, with restoration of E-cadherin showing improvement in disease outcomes.84

Candida albicans infection has also been shown to diminish E-cadherin expression in both *in vivo* and *in vitro* infection models,^{85–87} with candidalysin highlighted as a direct contributor to epithelial damage, loss of barrier integrity and subsequent translocation of *C. albicans* across the intestinal epithelia.⁸⁸ As the gut is considered the main site of *C. albicans* entry into the bloodstream,⁸⁹ where it may lead to systemic candidaemia and mortality, the pathological impact of *Candida* and candidalysin at this organ appears highly significant. The ability of *C. albicans* to diminish E-cadherin expression and other cell adhesion proteins such as occludin and desmoglein-2,⁹⁰ and diminish barrier integrity, may be of particular interest in the context of IBD, oral and skin disorders.

Recently, alcoholic hepatitis was found to be another potential example of the pathological impact of C. albicans in breaching gut barriers.^{91,92} The gut-liver axis describes the relationship and role of the gut microbiome in shaping healthy liver metabolism. This connection is supported by the close anatomical proximity between the gut and liver, as well as a specialized portal circulation, which permits enhanced permeability and interaction between gutderived substances and liver-resident cells. Gut-derived bacterial components and endotoxins in particular have long been implicated in driving alcoholic liver disease.93,94 Recent studies now show a similarly significant role for C. albicans. Upon infection, ligation of C. albicans β-glucans to host dectin-1 receptors on liver-resident macrophages (kupffer cells), results in inflammatory IL-1ß release and enhanced ethanol-induced liver disease in mice.91 A second, non-dectin-1-mediated but candidalvsin-induced mechanism also drives elevated hepatic damage, steatosis and mortality in ethanol-fed C. albicans-infected mice.92 The authors additionally observe that alcoholic hepatitis patients carry elevated levels of the candidalysin-encoding gene, ECE1, when compared to healthy controls. These data identify two distinct C. albicans mechanisms, each independently promoting alcoholic hepatitis and highlights this pathogen as a new and considerable factor for the development of alcoholic liver disease.

Dysregulated growth signalling

The ability of *C. albicans* to activate the epidermal growth factor receptor (EGFR) may also contribute to *C. albicans*-associated comorbidities. The EGFR is a transmembrane

protein with a broad range of functions controlling various cell proliferative and maintenance roles, in addition to immune induction. It is often found highly dysregulated, via overexpression or constitutive activation, in a select group of cancers including head and neck, breast, lung, colon and vulvovaginal cancers.⁹⁵ Interestingly, the majority of these EGFR-associated cancers are located at sites where *C. albicans* commonly infects, with reports providing contrasting evidence both for and against elevated incidences of candidiasis in these patients. Whilst immune suppression resulting from anticancer therapy may indeed play a role, a long-standing debate on the ability of *C. albicans* to potentiate oncogenic disease exists, primarily in oral cancer. Recent studies may now provide additional aspects for consideration.⁹⁶

C. albicans infection potently activates the EGFR. Upon cell adhesion, EGFR is bound and activated by the fungal cell wall protein Als3p, which initiates endocytosis of the fungus, providing an entry mechanism into host cells.⁹⁷ Additionally, candidalysin can indirectly activate EGFR through a complex mechanism involving matrix metalloproteinases (MMPs) and EGFR ligands, resulting in downstream immune activation.²¹ Notably, MMPs⁹⁸ and EGFR ligands⁹⁵ are each independently implicated in a number of cancers. Other observations suggesting contribution to cancer development include the ability for C. albicans to activate epithelial MAPK⁹⁹ and ERK signalling pathways, which are associated with growth and proliferation; loss of E-cadherin and occludin,⁹⁰ observed in epithelial-mesenchymal transition (EMT); activation of angiogenesis¹⁰⁰ and pro-angiogenic factors;^{101,102} and the ability of Candida to enhance production of known carcinogenic

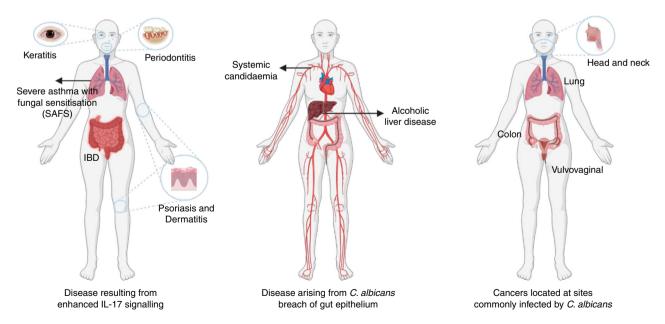


Figure 1. Candida albicans potential contribution to disease. Potential C. albicans mechanisms of contributing to disease include potent induction of IL-17 signalling, breach of gut epithelial barriers and activation of multiple cancer-associated factors.

molecules such as nitrosamines^{103,104} and acetaldehyde.^{105,106} However, clinical and *in vivo* evidence substantiating a direct causal or potentiating role for *C. albicans* in cancer is particularly limited. As such, the association here remains ambiguous.

Activation of MMPs is also observed in oral disease,¹⁰⁷ resulting in breakdown of gingival and periodontal ligament collagens, tissue remodelling, inflammation and uncontrolled extracellular matrix (ECM) turnover, also associated with cancer.^{108,109} Investigation into potential links with *C. albicans* would be of great interest given the known associations of this fungus with oral disease, its ability to signal through²¹ and induce MMPs,¹¹⁰ and MMP activation being a demonstrated mechanism for disease utilized by other oral pathogens.^{111,112}

Conclusions

As we increase our understanding of *C. albicans* induced pathophysiology, the potential for infection to contribute to several comorbidities becomes increasingly apparent. Its natural distribution throughout the body and ability to activate events highly linked with disease may be of significant consequence. Induction of IL-17-mediated signalling, breach of epithelial barriers and activation of cancer-associated factors provide the most convincing examples of its ability to contribute to disease (summarized in Fig. 1), though greater understanding is required to fully delineate its role in these instances, as well as others, yet unknown. Further research into the association of *C. albicans* with these diseases shall undoubtedly shed light on new mechanisms of disease development, which may shift perceptions of this under-investigated microbe and its influence on human health.

Acknowledgements

J.H. conceptualised and wrote the paper; G.C. created the schematic diagram; J.G., J.P.R., N.K. and J.R.N. edited the paper. This work was supported by grants from the Well-come Trust (214229 Z 18 Z), Biotechnology & Biological Sciences Research Council (BB/N014677/1), National Institutes of Health (R37-DE022550), King's Health Partners Challenge Fund (R170501), the Rosetrees Trust (M680), and the NIH Research at Guys and St. Thomas's NHS Foundation Trust and the King's College London Biomedical Research Centre (IS-BRC-1215-20006).

Disclosures

The authors declare no conflict of interest.

Data availability statement

Not applicable. No data were generated in the making of this article.

References

- 1 Mallick EM, Bennett RJ. Sensing of the microbial neighborhood by Candida albicans. PLoS Pathog 2013; 9:e1003661.
- 2 Festekjian A, Neely M. Incidence and predictors of invasive candidiasis associated with candidaemia in children. *Mycoses* 2011; **54**:146–53.
- 3 Leroy O, Gangneux JP, Montravers P, Mira JP, Gouin F, Sollet JP, et al. Epidemiology, management, and risk factors for death of invasive Candida infections in critical care: a multicenter, prospective, observational study in France (2005–2006). Crit Care Med 2009; 37:1612–8.
- 4 Toda M, Williams SR, Berkow EL, Farley MM, Harrison LH, Bonner L, et al. Population-based active surveillance for culture-confirmed candidemia - Four Sites, United States, 2012–2016. MMWR Surveill Summ 2019; 68:1–15.
- 5 Bongomin F, Gago S, Oladele RO, Denning DW. Global and multi-national prevalence of fungal diseases—estimate precision. J Fungi 2017; 3:57.
- 6 Dutzan N, Abusleme L, Bridgeman H, Greenwell-Wild T, Zangerle-Murray T, Fife ME, et al. On-going mechanical damage from mastication drives homeostatic Th17 Cell Responses at the oral barrier. *Immunity* 2017; 46:133–47.
- 7 Eskan MA, Jotwani R, Abe T, Chmelar J, Lim JH, Liang S, et al. The leukocyte integrin antagonist Del-1 inhibits IL-17-mediated inflammatory bone loss. Nat Immunol 2012; 13:465–73.
- 8 Canabarro A, Valle C, Farias MR, Santos FB, Lazera M, Wanke B. Association of subgingival colonization of *Candida albicans* and other yeasts with severity of chronic periodontitis. *J Periodontal Res* 2013; **48**:428–32.
- 9 De-La-Torre J, Quindós G, Marcos-Arias C, Marichalar-Mendia X, Gainza ML, Eraso E, et al. Oral Candida colonization in patients with chronic periodontitis. Is there any relationship? Rev Iberoam Micol 2018; 35:134–9.
- 10 Sokol H, Leducq V, Aschard H, Pham HP, Jegou S, Landman C, et al. Fungal microbiota dysbiosis in IBD. Gut 2017; 66:1039–48.
- 11 Yazdani R, Moazzami B, Madani SP, Behniafard N, Azizi G, Aflatoonian M, et al. Candidiasis associated with very early onset inflammatory bowel disease: first IL10RB deficient case from the National Iranian Registry and review of the literature. Clin Immunol 2019; 205:35–42.
- 12 Park CO, Fu X, Jiang X, Pan Y, Teague JE, Collins N, et al. Staged development of long-lived T-cell receptor αβ TH17 resident memory T-cell population to Candida albicans after skin infection. J Allergy Clin Immunol 2018; 142:647–62.
- 13 Savolainen J, Lintu P, Kosonen J, Kortekangas-Savolainen O, Viander M, Pène J, et al. Pityrosporum and Candida specific and non-specific humoral, cellular and cytokine responses in atopic dermatitis patients. Clin Exp Allergy 2001; 31:125–34.
- 14 Savolainen J, Lammintausta K, Kalimo K, Viander M. Candida albicans and atopic dermatitis. Clin Exp Allergy 1993; 23:332–9.
- 15 Shao TY, Ang WXG, Jiang TT, Huang FS, Andersen H, Kinder JM, et al. Commensal Candida albicans positively calibrates systemic Th17 immunological responses. Cell Host Microbe 2019; 25:404–17.e6.
- 16 Bacher P, Hohnstein T, Beerbaum E, Röcker M, Blango MG, Kaufmann S, et al. Human anti-fungal Th17 immunity and pathology rely on cross-reactivity against Candida albicans. Cell 2019; 176:1340–55.e15.
- 17 Khosravi AR, Bandghorai AN, Moazzeni M, Shokri H, Mansouri P, Mahmoudi M. Evaluation of *Candida albicans* allergens reactive with specific IgE in asthma and atopic eczema patients. *Mycoses* 2009; 52:326–33.
- 18 Masaki K, Fukunaga K, Matsusaka M, Kabata H, Tanosaki T, Mochimaru T, et al. Characteristics of severe asthma with fungal sensitization. Ann Allergy Asthma Immunol 2017; 119:253–7.
- 19 Moyes DL, Wilson D, Richardson JP, Mogavero S, Tang SX, Wernecke J, et al. Candidalysin is a fungal peptide toxin critical for mucosal infection. Nature 2016; 532:64–8.
- 20 Verma AH, Richardson JP, Zhou C, Coleman BM, Moyes DL, Ho J, et al. Oral epithelial cells orchestrate innate type 17 responses to *Candida albicans* through the virulence factor candidalysin. *Sci Immunol* 2017;2:eaam8834.
- 21 Ho J, Yang X, Nikou S-A, Kichik N, Donkin A, Ponde NO, et al. Candidalysin activates innate epithelial immune responses via epidermal growth factor receptor. Nat Commun 2019; 10:2297.
- 22 Conti HR, Shen F, Nayyar N, Stocum E, Sun JN, Lindemann MJ, et al. Th17 cells and IL-17 receptor signaling are essential for mucosal host defense against oral candidiasis. J Exp Med 2009; 206:299–311.
- 23 Swidergall M, Khalaji M, Solis NV, Moyes DL, Drummond RA, Hube B, et al. Candidalysin is required for neutrophil recruitment and virulence during systemic Candida albicans infection. J Infect Dis 2019; 220:1477–88.
- 24 Aggor FEY, Break TJ, Trevejo-Nuñez G, Whibley N, Coleman BM, Bailey RD, et al. Oral epithelial IL-22/STAT3 signaling licenses IL-17-mediated immunity to oral mucosal candidiasis. *Sci Immunol* 2020; 5:eaba0570.
- 25 Drummond RA, Swamydas M, Oikonomou V, Zhai B, Dambuza IM, Schaefer BC, et al. CARD9 + microglia promote antifungal immunity via IL-1β- and CXCL1-mediated neutrophil recruitment. Nat Immunol 2019; 20:559–70.

- 26 Leonardi S, Cuppari C, Manti S, Filippelli M, Parisi GF, Borgia F, et al. Serum interleukin 17, interleukin 23, and interleukin 10 values in children with atopic eczema/ dermatitis syndrome (AEDS): Association with clinical severity and phenotype. Allergy Asthma Proc 2015; 36:74–81.
- 27 Koga C, Kabashima K, Shiraishi N, Kobayashi M, Tokura Y. Possible pathogenic role of Th17 cells for atopic dermatitis. J Invest Dermatol 2008; 128:2625–30.
- 28 Langley RG, Elewski BE, Lebwohl M, Reich K, Griffiths CEM, Papp K, et al. Secukinumab in plaque psoriasis - results of two phase 3 trials. N Engl J Med 2014; 371:326–38.
- 29 Fujino S, Andoh A, Bamba S, Ogawa A, Hata K, Araki Y, et al. Increased expression of interleukin 17 in inflammatory bowel disease. Gut 2003; 52:65–70.
- 30 Moschen AR, Tilg H, Raine T. IL-12, IL-23 and IL-17 in IBD: immunobiology and therapeutic targeting. Nat Rev Gastroenterol Hepatol 2019; 16:185–96.
- 31 Lubberts E. The IL-23-IL-17 axis in inflammatory arthritis. Nat Rev Rheumatol 2015; 11:415–29.
- 32 Bühler U, Fleischer V, Luessi F, Rezk A, Belikan P, Graetz C, et al. Role of IL-17-producing lymphocytes in severity of multiple sclerosis upon natalizumab treatment. *Mult Scler* 2017; 23:567–76.
- 33 Koga T, Ichinose K, Tsokos GC. T cells and IL-17 in lupus nephritis. Clin Immunol 2017; 185:95–9.
- 34 Liang SC, Tan XY, Luxenberg DP, Karim R, Dunussi-Joannopoulos K, Collins M, et al. Interleukin (IL)-22 and IL-17 are coexpressed by Th17 cells and cooperatively enhance expression of antimicrobial peptides. J Exp Med 2006; 203:2271–9.
- 35 Amatya N, Garg AV, Gaffen SL. IL-17 Signaling: the Yin and the Yang. Trends Immunol 2017; 38:310–22.
- 36 Blauvelt A, Chiricozzi A. The Immunologic Role of IL-17 in psoriasis and psoriatic arthritis pathogenesis. *Clin Rev Allergy Immunol* 2018; 55:379–90.
- 37 Coffelt SB, Kersten K, Doornebal CW, Weiden J, Vrijland K, Hau CS, et al. IL-17producing γδ T cells and neutrophils conspire to promote breast cancer metastasis. *Nature* 2015; **522**:345–8.
- 38 Huppler AR, Whibley N, Woolford CA, Childs EE, He J, Biswas PS, et al. A Candida albicans strain expressing mammalian interleukin-17A results in early control of fungal growth during disseminated infection. *Infect Immun* 2015; 83:3684–92.
- 39 Huppler AR, Conti HR, Hernández-Santos N, Darville T, Biswas PS, Gaffen SL. Role of Neutrophils in IL-17–dependent immunity to mucosal Candidiasis. J Immunol 2014; 192:1745–52.
- 40 Bär E, Whitney PG, Moor K, ReiseSousa C, LeibundGut-Landmann S. IL-17 regulates systemic fungal immunity by controlling the functional competence of NK Cells. *Immunity* 2014; 40:117–27.
- 41 Whibley N, Tritto E, Traggiai E, Kolbinger F, Moulin P, Brees D, et al. Antibody blockade of IL-17 family cytokines in immunity to acute murine oral mucosal candidiasis. J Leukoc Biol 2016; 99:1153–64.
- 42 Savolainen J, Kosonen J, Lintu P, Viander M, Pène J, Kalimo K, et al. Candida albicans mannan- and protein-induced humoral, cellular and cytokine responses in atopic dermatitis patients. Clin Exp Allergy 1999; 29:824–31.
- 43 Pietrzak A, Grywalska E, Socha M, Roliński J, Franciszkiewicz-Pietrzak K, Rudnicka L, et al. Prevalence and possible role of Candida species in patients with psoriasis: a systematic review and meta-analysis. *Mediators Inflamm* 2018; 2018:824–831.
- 44 Waldman A, Gilhar A, Duek I, Berdicevsky I. Incidence of *Candida* in psoriasis a study on the fungal flora of psoriatic patients. *Mycoses* 2001; 44:77–81.
- 45 Zou Y, Zhang H, Li H, Chen H, Song W, Wang Y. Strain-dependent production of interleukin-17/interferon-γ and matrix remodeling-associated genes in experimental *Candida albicans* keratitis. *Mol Vis* 2012; 18:1215–25.
- 46 Muthular M, Bálsamo F, Passero P, Jewtuchowicz V, Miozza V, Villalba MB, et al. Effects of tamoxifen on periodontal disease and *Candida albicans* of patients with breast cancer and other pathologies. *Future Microbiol* 2019; 14:129–37.
- 47 Leonardi I, Li X, Semon A, Li D, Doron I, Putzel G, et al. CX3CR1+, mononuclear phagocytes control immunity to intestinal fungi. Science 2018; 359:232–6.
- 48 Leonardi I, Paramsothy S, Doron I, Semon A, Kaakoush NO, Clemente JC, et al. Fungal trans-kingdom dynamics linked to responsiveness to fecal microbiota transplantation (FMT) therapy in ulcerative colitis. Cell Host Microbe 2020; 27:823–9.e3.
- 49 Wardhana DEA. A patient with allergic bronchopulmonary mycosis caused by Aspergillus fumigatus and Candida albicans. Acta Med Indones 2012; 44:317–23.
- 50 Mirsadraee M, Dehghan S, Ghaffari S, Mirsadraee N. Long-term effect of antifungal therapy for the treatment of severe resistant asthma: an active comparator clinical trial. Curr Med Mycol 2019; 5:1–7.
- 51 Pasqualotto AC, Powell G, Niven R, Denning DW. The effects of antifungal therapy on severe asthma with fungal sensitization and allergic bronchopulmonary aspergillosis. *Respirology* 2009; 14:1121–7.
- 52 Richardson JP, Willems HME, Moyes DL, Shoaie S, Barker KS, Tan SL, et al. Candidalysin drives epithelial signaling, neutrophil recruitment, and immunopathology at the vaginal mucosa. *Infect Immun* 2017; 86:e00645-17.

- 53 Verma AH, Zafar H, Ponde NO, Hepworth OW, Sihra D, Aggor FEY, et al. IL-36 and IL-1/IL-17 Drive immunity to oral candidiasis via parallel mechanisms. J Immunol 2018; 201:627–34.
- 54 Braegelmann J, Braegelmann C, Bieber T, Wenzel J. Candida induces the expression of IL-36γ in human keratinocytes: implications for a pathogen-driven exacerbation of psoriasis?. J Eur Acad Dermatol Venereol 2018; 32:e403–e406.
- 55 Kasper L, König A, Koenig P-A, Gresnigt MS, Westman J, Drummond RA, et al. The fungal peptide toxin candidalysin activates the NLRP3 inflammasome and causes cytolysis in mononuclear phagocytes. Nat Commun 2018; 9:4260.
- 56 Rogiers O, Frising UC, Kucharíková S, Jabra-Rizk MA, van Loo G, Van Dijck P, et al. Candidalysin crucially contributes to nlrp3 inflammasome activation by *Candida albicans* hyphae. *MBio* 2019; **10**: e02221-18.
- 57 Fonseca-Camarillo G, Furuzawa-Carballeda J, Iturriaga-Goyon E, Yamamoto-Furusho JK. Differential expression of IL-36 family members and IL-38 by immune and non-immune cells in patients with active inflammatory bowel disease. *Biomed Res Int* 2018; **2018**:5140691.
- 58 Scheibe K, Kersten C, Schmied A, Vieth M, Primbs T, Carlé B, et al. Inhibiting interleukin 36 receptor signaling reduces fibrosis in mice with chronic intestinal inflammation. Gastroenterology 2019; 156:1082–97.e11.
- 59 Wang W, Yu X, Wu C, Jin H. Il-36γ inhibits differentiation and induces inflammation of keratinocyte via Wnt signaling pathway in psoriasis. *Int J Med Sci* 2017; 14:1002–7.
- 60 Jang HY, Koo JH, Lee SM, Park BH. Atopic dermatitis-like skin lesions are suppressed in fat-1 transgenic mice through the inhibition of inflammasomes. *Exp Mol Med* 2018; 50:1–9.
- 61 Yang BY, Cheng YG, Liu Y, Liu Y, Tan JY, Guan W, et al. Datura Metel L. Ameliorates imiquimod-induced psoriasis-like dermatitis and inhibits inflammatory cytokines production through TLR7/8-MyD88-NF-κB-NLRP3 inflammasome pathway. Molecules 2019; 24:2157.
- 62 Lückel C, Raifer H, Campos Carrascosa L, Guralnik A, Zhang Y, et al. IL-17+ CD8+ T cell suppression by dimethyl fumarate associates with clinical response in multiple sclerosis. Nat Commun 2019; 10.5722
- 63 Mease PJ, McInnes IB, Kirkham B, Kavanaugh A, Rahman P, Van Der Heijde D, et al. Secukinumab inhibition of interleukin-17A in patients with psoriatic arthritis. N Engl J Med 2015; 373:1329–39.
- 64 Krueger JG, Wharton KA, Schlitt T, Suprun M, Torene RI, Jiang X, et al. IL-17A inhibition by secukinumab induces early clinical, histopathologic, and molecular resolution of psoriasis. J Allergy Clin Immunol 2019; 144:750–63.
- 65 Hohenberger M, Cardwell LA, Oussedik E, Feldman SR. Interleukin-17 inhibition: role in psoriasis and inflammatory bowel disease. J Dermatol Treat 2018; 29:13–8.
- 66 Hueber W, Sands BE, Lewitzky S, Vandemeulebroecke M, Reinisch W, Higgins PDR, et al. Secukinumab, a human anti-IL-17A monoclonal antibody, for moderate to severe Crohn's disease: Unexpected results of a randomised, double-blind placebo-controlled trial. Gut 2012; 61:1693–700.
- 67 Smith MK, Pai J, Panaccione R, Beck P, Ferraz JG, Jijon H. Crohn's-like disease in a patient exposed to anti-Interleukin-17 blockade (Ixekizumab) for the treatment of chronic plaque psoriasis: a case report. *BMC Gastroenterol* 2019; **19**:162.
- 68 Ungar B, Pavel AB, Li R, Kimmel G, Nia J, Hashim P, et al. Phase 2 Randomized, Double-blind Study of IL-17-Targeting with Secukinumab in atopic dermatitis. J Allergy Clin Immunol 2020; 6749:30684–9.
- 69 Conti HR, Peterson AC, Brane L, Huppler AR, Hernández-Santos N, Whibley N, et al. Oral-resident natural Th17 cells and γδ T cells control opportunistic Candida albicans infections. J Exp Med 2014; 211:2075–84.
- 70 Yano J, Lilly E, Barousse M, Fidel PL. Epithelial Cell-Derived S100 Calcium-binding proteins as key mediators in the hallmark acute neutrophil response during *Candida* vaginitis. *Infect Immun* 2010; **78**:5126–37.
- 71 Zhang H, Li H, Li Y, Zou Y, Dong X, Song W, et al. IL-17 plays a central role in initiating experimental *Candida albicans* infection in mouse corneas. Eur J Immunol 2013; 43:2671–82.
- 72 Del Fresno C, Saz-Leal P, Enamorado M, Wculek SK, Martínez-Cano S, Blanco-Menéndez N, et al. DNGR-1 in dendritic cells limits tissue damage by dampening neutrophil recruitment. Science 2018; 362:351–6.
- 73 Hector A, Chotirmall SH, Lavelle GM, Mirković B, Horan D, Eichler L, et al. Chitinase activation in patients with fungus-associated cystic fibrosis lung disease. J Allergy Clin Immunol 2016; 138:1183–9.e4.
- 74 Guillot C, Lecuit T. Mechanics of epithelial tissue homeostasis and morphogenesis. *Science* 2013; 340:1185–9.
- 75 Lee G, Kim HJ, Kim HM. RhoA-JNK regulates the E-cadherin junctions of human gingival epithelial cells. J Dent Res 2016; 95:284–91.
- 76 Abdulkareem AA, Shelton RM, Landini G, Cooper PR, Milward MR. Potential role of periodontal pathogens in compromising epithelial barrier function by inducing epithelial-mesenchymal transition. J Periodontal Res 2018; 53:565–74.

- 77 Arun R, Hemalatha R, Arun KV, Kumar TSS. E-cadherin and CD1a expression in gingival epithelium in periodontal health, disease and post-treatment. *Indian J Dent Res* 2010; 21:396–401.
- 78 Schneider MR, Dahlhoff M, Horst D, Hirschi B, Trülzsch K, Müller-Höcker J, et al. A key role for E-cadherin in intestinal homeostasis and paneth cell maturation. PLoS One 2010; 5:e14325.
- 79 Muise AM, Walters TD, Glowacka WK, Griffiths AM, Ngan BY, Lan H, et al. Polymorphisms in E-cadherin (CDH1) result in a mislocalised cytoplasmic protein that is associated with Crohn's disease. Gut 2009; 58:1121–7.
- 80 Mehta S, Nijhuis A, Kumagai T, Lindsay J, Silver A. Defects in the adherens junction complex (E-cadherin/ β-catenin) in inflammatory bowel disease. *Cell Tissue Res* 2015; 360:749–60.
- 81 Xie G, Ao X, Lin T, Zhou G, Wang M, Wang H, et al. E-Cadherin–mediated cell contact controls the epidermal damage response in radiation dermatitis. J Invest Dermatol 2017; 137:1731–9.
- 82 Nakai K, Yoneda K, Hosokawa Y, Moriue T, Presland RB, Fallon PG, et al. Reduced expression of epidermal growth factor receptor, E-cadherin and occludin in the skin of flaky tail mice is due to filaggrin and loricrin deficiencies. Am J Pathol. 2012; 181:969–77.
- 83 Nelson AM, Cong Z, Gettle SL, Longenecker AL, Kidacki M, Kirby JS, et al. E-cadherin and p120ctn protein expression are lost in hidradenitis suppurativa lesions. Exp Dermatol 2019; 28:867–71.
- 84 Terciolo C, Dobric A, Ouaissi M, Siret C, Breuzard G, Silvy F, et al. Saccharomyces boulardii CNCM 1-745 restores intestinal barrier integrity by regulation of E-cadherin recycling. J Crohns Colitis 2017; 11:999–1010.
- 85 Rouabhia M, Semlali A, Audoy J, Chmielewski W. Antagonistic effect of *Candida albi*cans and IFNγ on E-cadherin expression and production by human primary gingival epithelial cells. *Cell Immunol* 2012; 280:61–7.
- 86 Xu H, Sobue T, Bertolini M, Thompson A, Dongari-Bagtzoglou A. Streptococcus oralis and Candida albicans synergistically activate μ-Calpain to degrade E-cadherin from oral epithelial junctions. J Infect Dis 2016; 214:925–34.
- 87 Qu S, Chen L, Tian H, Wang Z, Wang F, Wang L, et al. Effect of perillaldehyde on prophylaxis and treatment of vaginal candidiasis in a murine model. Front Microbiol 2019; 10.1466
- 88 Allert S, Förster TM, Svensson C-M, Richardson JP, Pawlik T, Hebecker B, et al. Candida albicans-induced epithelial damage mediates translocation through intestinal barriers. MBio 2018; 9:e00915-18.
- 89 Miranda LN, van der Heijden IM, Costa SF, Sousa API, Sienra RA, Gobara S, et al. Candida colonisation as a source for candidaemia. J Hosp Infect 2009; 72:9–16.
- 90 Frank CF, Hostetter MK. Cleavage of E-cadherin: a mechanism for disruption of the intestinal epithelial barrier by *Candida albicans. Transl Res* 2007; 149:211–22.
- 91 Yang AM, Inamine T, Hochrath K, Chen P, Wang L, Llorente C, et al. Intestinal fungi contribute to development of alcoholic liver disease. J Clin Invest 2017; 127:2829–41.
- 92 Chu H, Duan Y, Lang S, Jiang L, Wang Y, Llorente C, et al. The Candida albicans exotoxin candidalysin promotes alcohol-associated liver disease. J Hepatol 2020; 72:391–400.
- 93 Szabo G. Gut-liver axis in alcoholic liver disease. Gastroenterology 2015; 148:30-6.
- 94 Schnabl B, Brenner DA. Interactions between the intestinal microbiome and liver diseases. Gastroenterology 2014; 146:1513–24.

- 95 Sigismund S, Avanzato D, Lanzetti L. Emerging functions of the EGFR in cancer. Mol Oncol 2018;12:3–20.
- 96 Engku Nasrullah Satiman EAF, Ahmad H, Ramzi AB, Abdul Wahab R, Kaderi MA, Wan Harun WHA, et al. The role of *Candida albicans* candidalysin ECE1 gene in oral carcinogenesis. J Oral Pathol Med 2020; jop.13014.
- 97 Zhu W, Phan QT, Boontheung P, Solis NV, Loo JA, Filler SG. EGFR and HER2 receptor kinase signaling mediate epithelial cell invasion by *Candida albicans* during oropharyngeal infection. *Proc Natl Acad Sci USA* 2012; 109:14194–9.
- 98 Vandenbroucke RE, Libert C. Is there new hope for therapeutic matrix metalloproteinase inhibition? Nat Rev Drug Discovery 2014; 13:904–27.
- 99 Román E, Correia I, Prieto D, Alonso R, Pla J. The HOG MAPK pathway in Candida albicans: more than an osmosensing pathway. Int Microbiol 2020; 23:23–9.
- 100 Ashman RB, Papadimitriou JM. Endothelial cell proliferation associated with lesions of murine systemic candidiasis. *Infect Immun* 1994; 62:5151–3.
- 101 Barker KS, Park H, Phan QT, Xu L, Homayouni R, Rogers PD, et al. Transcriptome profile of the vascular endothelial cell response to *Candida albicans. J Infect Dis* 2008; 198:193–202.
- 102 Vellanki S, Huh EY, Saville SP, Lee SC. Candida albicans morphology-dependent host fgf-2 response as a potential therapeutic target. J Fungi 2019; 5:22.
- 103 Krogh P. The role of yeasts in oral cancer by means of endogenous nitrosation. Acta Odontol Scand 1990; 48:85–8.
- 104 Krogh P, Hald B, Holmstrup P. Possible mycological etiology of oral mucosal cancer: catalytic potential of infecting *Candida albicans* and other yeasts in production of Nnitrosobenzylmethylamine. *Carcinogenesis* 1987; 8:1543–8.
- 105 Alnuaimi A, Ramdzan A, Wiesenfeld D, O'Brien-Simpson N, Kolev S, Reynolds E, et al. Candida virulence and ethanol-derived acetaldehyde production in oral cancer and non-cancer subjects. Oral Dis 2016; 22:805–14.
- 106 Gainza-Cirauqui ML, Nieminen MT, Novak Frazer L, Aguirre-Urizar JM, Moragues MD, Rautemaa R. Production of carcinogenic acetaldehyde by *Candida albicans* from patients with potentially malignant oral mucosal disorders. *J Oral Pathol Med* 2013; 42:243–9.
- 107 Sorsa T, Tjäderhane L, Salo T. Matrix metalloproteinases (MMPs) in oral diseases. Oral Dis 2004; 10:311–8.
- 108 Gürsoy UK, Könönen E, Tervahartiala T, Gürsoy M, Pitkänen J, Torvi P, et al. Molecular forms and fragments of salivary MMP-8 in relation to periodontitis. J Clin Periodontol 2018; 45:1421–8.
- 109 Franco C, Patricia HR, Timo S, Claudia B, Marcela H. Matrix metalloproteinases as regulators of periodontal inflammation. Int J Mol Sci 2017; 18:440.
- 110 Yuan X, Mitchell BM, Wilhelmus KR. Expression of matrix metalloproteinases during experimental *Candida albicans* keratitis. *Investig Ophthalmol Vis Sci* 2009; 50:737–42.
- 111 Potempa J, Banbula A, Travis J. Role of bacterial proteinases in matrix destruction and modulation of host responses. *Periodontol 2000* 2000; 24:153–92.
- 112 Ding Y, Haapasalo M, Kerosuo E, Lounatmaa K, Kotiranta A, Sorsa T. Release and activation of human neutrophil matrix metallo- and serine proteinases during phagocytosis of *Fusobacterium nucleatum*, *Porphyromonas gingivalis* and *Treponema denticola*. J Clin Periodontol 1997; 24:237–48.