

# Whole-Genome Sequencing Reveals a New Genospecies of *Methylobacterium* sp. GXS13, Isolated from *Vitis vinifera* L. Xylem Sap

Wan Xin Lai,<sup>a,b</sup> Han Ming Gan,<sup>a,b</sup> André O. Hudson,<sup>c</sup> Michael A. Savka<sup>c</sup>

School of Science<sup>a</sup> and Monash University Malaysia Genomics Facility,<sup>b</sup> Monash University Malaysia, Jalan Lagoon Selatan, Selangor, Malaysia; The Thomas H. Gosnell School of Life Sciences, College of Science, Rochester Institute of Technology, Rochester, New York, USA<sup>c</sup>

**The whole-genome sequence of a new genospecies of *Methylobacterium* sp., named GXS13 and isolated from grapevine xylem sap, is reported and demonstrates potential for methylotrophy, cytokinin synthesis, and cell wall modification. In addition, biosynthetic gene clusters were identified for cupriachelin, carotenoid, and acyl-homoserine lactone using the antiSMASH server.**

Received 8 December 2015 Accepted 11 December 2015 Published 4 February 2016

**Citation** Lai WX, Gan HM, Hudson AO, Savka MA. 2016. Whole-genome sequencing reveals a new genospecies of *Methylobacterium* sp. GXS13, isolated from *Vitis vinifera* L. xylem sap. *Genome Announc* 4(1):e01695-15. doi:10.1128/genomeA.01695-15.

**Copyright** © 2016 Lai et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Michael A. Savka, [massbi@rit.edu](mailto:massbi@rit.edu).

Members of the genus *Methylobacterium* are fastidious Gram-negative rods known for their methylotrophic metabolism, i.e., ability to utilize C1 compounds as their sole carbon and energy source. *Methylobacterium* species have been isolated from soil, leaf surface, grape xylem fluid, water, diseased tissue, and biochemical reagents (1–3).

Some *Methylobacterium* spp. are plant associated as during growth plants emit substantial amounts of methanol through the stomata (4). In some cases, *Methylobacterium* species also exhibited a symbiotic relationship with their plant host through the production of cytokinin that stimulates seed germination and plant development. Previously, we characterized and sequenced the whole genome of a grapevine xylem isolate, *Methylobacterium* sp. GXF4, leading to the identification of two *luxIR* homologs implicated in cell-to-cell communication and a unique  $\beta$ -galactosidase gene.

In the work reported here, we performed low-coverage whole-genome sequencing on morphologically dissimilar pink-pigmented bacterial isolates from the same grapevine xylem fluid and identified a genomically distinct isolate, strain GXS13, based on *in silico* genome-genome hybridization against *Methylobacterium* sp. GXF4 (5, 6).

Genomic DNA (gDNA) was extracted from strain GXS13 grown on potato dextrose agar medium for 5 days and prepped using the NEBNext Ultra DNA library prep kit (New England Biolabs, Ipswich, MA). The library was quantified and subsequently sequenced on the Illumina MiSeq (Illumina, San Diego, CA) at the Monash University Malaysia Genomics Facility. The raw reads were adapter trimmed with Trimmomatic 0.33 (7) and assembled using Spades 3.5.0 (8). The assembly contains 112 contigs with a total genome size of 5,805,293 bp ( $N_{50}$  of 109,000 bp; GC content of 68.50%; 85 $\times$  coverage). Initial taxonomy assignment was performed using SpecI (9) and subsequently refined based on average nucleotide identity (ANI) analysis with JSpecies. Genome annotation based on PGAAP (10) led to the identifica-

tion of 5,159 open reading frames (ORFs), 50 tRNAs, and 12 rRNAs.

SpecI assigned strain GXF13 to the genus *Methylobacterium*. A similarity search against *Methylobacterium* type strain gene sequences indicated that strain GXS13 is closely related to *Methylobacterium mesophilicum* DSM 1708T (16S rRNA and *gyrB* gene identities of 99.58% and 92.65%, respectively). ANI analysis shows that strain GXS13 has the highest score of 89.89% (as of October 2015) to *Methylobacterium* sp. GXF4 (6). Methanol oxidation genes were identified at contig 2 (*mxafJGIRSACK-LDEHB*), contig 3 (*mxqQE*), contig 1 (*mxbMD*), contig15 (*pqqA*), contig 1 (*pqqBCDE*), and contig 10 (*pqqFG*) (6, 11). Additionally, strain GXS13 also carries the *miaA* gene (contig 88) implicated in tRNA-mediated cytokinin synthesis via the isoprenylation of specific adenine in some tRNA (12, 13). Comparison of the strain GXS13 genome with publicly available *Methylobacterium* genomes identified a unique gene in strain GXS13 coding for CAZY family GT34 glycosyltransferase in contig 21 involved in the formation of a glycosidic bond between plant cell wall components, xyloglucans, and heteromannans (14–16). In addition, using the antibiotics and secondary metabolite analysis shell (antiSMASH) server (17, 18), cupriachelin, carotenoid, and acyl-homoserine lactone gene clusters were identified. The genomic potential for cytokinin synthesis and plant cell wall modification suggests the positive role of strain GXS13 in the growth and development of its host, grapevine.

**Nucleotide sequence accession numbers.** The nucleotide sequences have been deposited at DDBJ/EMBL/GenBank under accession number [LKKO00000000](https://www.ncbi.nlm.nih.gov/nuccore/LKKO00000000). The BioProject number is PRJNA297388 and the BioSample number is SAMN04123207.

## ACKNOWLEDGMENTS

M.A.S., A.O.H., and H.M.G. acknowledge the College of Science (COS) and the Thomas H. Gosnell School of Life Sciences (GSOLS) at the Rochester Institute of Technology (RIT) for ongoing support. H.M.G. and M.A.S. also acknowledge a Dean's FEAD grant from the COS at RIT. In

addition, we acknowledge support from the Monash University Malaysia Tropical Medicine and Biology Multidisciplinary Platform.

## REFERENCES

- Rice EW, Reasoner DJ, Johnson CH, DeMaria LA. 2000. Monitoring for methylobacteria in water systems. *J Clin Microbiol* 38:4296–4297.
- Sanders JW, Martin JW, Hooke M, Hooke J. 2000. *Methylobacterium mesophilicum* infection: case report and literature review of an unusual opportunistic pathogen. *Clin Infect Dis* 30:936–938. <http://dx.doi.org/10.1086/313815>.
- Salter SJ, Cox MJ, Turek EM, Calus ST, Cookson WO, Moffatt MF, Turner P, Parkhill J, Loman NJ, Walker AW. 2014. Reagent and laboratory contamination can critically impact sequence-based microbiome analyses. *BMC Biol* 12:87. <http://dx.doi.org/10.1186/s12915-014-0087-z>.
- Nemecek-Marshall M, MacDonald RC, Franzen JJ, Wojciechowski CL, Fall R. 1995. Methanol emission from leaves (enzymatic detection of gas-phase methanol and relation of methanol fluxes to stomatal conductance and leaf development). *Plant Physiol* 108:1359–1368.
- Richter M, Rosselló-Móra R. 2009. Shifting the genomic gold standard for the prokaryotic species definition. *Proc Natl Acad Sci USA* 106:19126–19131. <http://dx.doi.org/10.1073/pnas.0906412106>.
- Gan HM, Chew TH, Hudson AO, Savka MA. 2012. Genome sequence of *Methylobacterium* sp. strain gxf4, a xylem-associated bacterium isolated from *Vitis vinifera* L. grapevine. *J Bacteriol* 194:5157–5158. <http://dx.doi.org/10.1128/JB.01201-12>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <http://dx.doi.org/10.1093/bioinformatics/btu170>.
- Nurk S, Bankevich A, Antipov D, Gurevich AA, Korobeynikov A, Lapidus A, Prjibelski AD, Pyshkin A, Sirotkin A, Sirotkin Y, Stepanauskas R, Clingenpeel SR, Woyke T, McLean JS, Lasken R, Tesler G, Alekseyev MA, Pevzner PA. 2013. Assembling single-cell genomes and mini-metagenomes from chimeric MDA products. *J Comput Biol* 20:714–737. <http://dx.doi.org/10.1089/cmb.2013.0084>.
- Mende DR, Sunagawa S, Zeller G, Bork P. 2013. Accurate and universal delineation of prokaryotic species. *Nat Methods* 10:881–884. <http://dx.doi.org/10.1038/nmeth.2575>.
- Angiuoli SV, Gussman A, Klimke W, Cochrane G, Field D, Garrity G, Kodira CD, Kyrpides N, Madupu R, Markowitz V, Tatusova T, Thomson N, White O. 2008. Toward an online repository of standard operating procedures (SOPs) for (meta)genomic annotation. *Omics* 12:137–141. <http://dx.doi.org/10.1089/omi.2008.0017>.
- Chistoserdova L, Chen SW, Lapidus A, Lidstrom ME. 2003. Methylo-trophy in *Methylobacterium extorquens* AM1 from a genomic point of view. *J Bacteriol* 185:2980–2987. <http://dx.doi.org/10.1128/JB.185.10.2980-2987.2003>.
- Koenig RL, Morris RO, Polacco JC. 2002. tRNA is the source of low-level trans-zeatin production in *Methylobacterium* spp. *J Bacteriol* 184:1832–1842. <http://dx.doi.org/10.1128/JB.184.7.1832-1842.2002>.
- Gray MW. 1996. RNA editing in plant organelles: a fertile field. *Proc Natl Acad Sci USA* 93:8157–8159. <http://dx.doi.org/10.1073/pnas.93.16.8157>.
- Williams GJ, Thorson JS. 2009. Natural product glycosyltransferases: properties and applications. *Adv Enzymol Relat Areas Mol Biol* 76:55–119.
- Wang D, Qu Z, Adelson DL, Zhu JK, Timmis JN. 2014. Transcription of nuclear organellar DNA in a model plant system. *Genome Biol Evol* 6:1327–1334. <http://dx.doi.org/10.1093/gbe/evu111>.
- Levasseur A, Drula E, Lombard V, Coutinho PM, Henrissat B. 2013. Expansion of the enzymatic repertoire of the CAZy database to integrate auxiliary redox enzymes. *Biotechnol Biofuels* 6:41. <http://dx.doi.org/10.1186/1754-6834-6-41>.
- Blin K, Medema MH, Kazempour D, Fischbach MA, Breitling R, Takano E, Weber T. 2013. antiSMASH 2.0—a versatile platform for genome mining of secondary metabolite producers. *Nucleic Acids Res* 41:W204–W212. <http://dx.doi.org/10.1093/nar/gkt449>.
- Medema MH, Blin K, Cimermancic P, de Jager V, Zakrzewski P, Fischbach MA, Weber T, Takano E, Breitling R. 2011. antiSMASH: rapid identification, annotation and analysis of secondary metabolite Biosynthesis gene clusters in bacterial and fungal genome sequences. *Nucleic Acids Res* 39:W339–W346. <http://dx.doi.org/10.1093/nar/gkr466>.