



Complete Genome Sequences for Two *Talaromyces marneffei* Clinical Isolates from Northern and Southern Vietnam

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ABSTRACT *Talaromyces marneffei* is a thermally dimorphic fungus endemic in China and Southeast Asia that causes fatal infections in immunocompromised individuals, particularly in patients with advanced HIV disease. Here, we report the complete genome sequences of two clinical isolates from northern and southern Vietnam.

The thermally dimorphic fungus *Talaromyces* (formerly *Penicillium*) *marneffei* causes fatal infections in immunocompromised individuals. In Vietnam, Thailand, and southern China, where *T. marneffei* is highly endemic, talaromycosis is a leading opportunistic infection and cause of death in HIV-infected individuals. The on-treatment mortality rates in HIV-infected and non-HIV-infected individuals approach 30% and 50%, respectively (1–3).

Two isolates of *T. marneffei* were collected from patients enrolled in an antifungal clinical trial in Vietnam (4). As prior multilocus sequence typing (MLST) analysis suggested a geographic substructure of *T. marneffei* in Southeast Asia (5), we selected one isolate from northern Vietnam (11CN-20-091) and one isolate from southern Vietnam (11CN-03-130) for genome sequencing. The isolates were cultured in Sabouraud dextrose agar (SDA) in the yeast form at 37°C for 5 days. DNA was prepared using the MasterPure yeast DNA purification kit (Epicentre). Oxford Nanopore libraries were constructed using the 1D ligation kit (catalog number SQK-LSK109) and loaded on a FLO-MIN106D flow cell for each sample on a GridIon instrument. Base calling was performed using Albacore v2.3.4 for 11CN-20-091 and using MinKNOW v3.1.20 for 11CN-03-130. A total coverage of 170× was generated for 11CN-03-130, and 152× coverage was generated for 11CN-20-091 (Table 1). The reads for each sample were assembled using Canu v1.5 (6), with the parameters “genomeSize=29000000” and “correctedErrorRate=0.075.” Next, the nanopore reads were aligned to the Canu assembly with minimap2 v2.9r720 (7), with the parameter “-ax map-ont,” and each assembly was polished with Nanopolish v0.11.0.

Two sets of Illumina data were used for error correction. One library for each sample was constructed using the DNA Flex Illumina protocol and sequenced on an iSeq instrument to generate paired 250-base reads (Table 1). A second library for each sample was constructed by MacroGen using the Nextera kit and sequenced on a HiSeq 4000 instrument to generate paired 101-base reads (Table 1). The assemblies were polished with all Illumina data using three rounds of alignment with BWA-MEM v0.7.7-r411 (8) and Pilon v1.23 (9) correction. Alignment of the assembly of 11CN-03-130 to that of 11CN-20-091 using Nucmer v3.1 (10) identified candidate rearrangement events. Nanopore and Illumina read alignments were visually inspected across these junctions in IGV v2.1.5 (11); where this revealed misassemblies, contigs were manually broken, correctly joined, and polished as described above. Contig alignments suggested that two contig joins could be made in 11CN-03-130, and one join could be

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TABLE 1 *Talaromyces* genome statistics

Genome statistic	Data for <i>Talaromyces</i> isolate:	
	11CN-03-130	11CN-20-091
No. of Nanopore reads	1,331,235	943,888
Nanopore coverage (×)	175	157
No. of Illumina reads (Flex library)	5,009,866	3,604,362
No. of Illumina reads (Nextera library)	13,936,480	18,321,844
Illumina coverage (both libraries) (×)	110	82
No. of contigs	9	8
Maximum contig length (bp)	6,376,262	6,464,005
Contig N_{50} (bp)	3,743,714	3,704,010
Total contig length (bp)	28,216,733	28,198,338
Assembly GC content (%)	46.79	46.76
No. of protein-coding genes	10,025	9,994
BUSCO (%)	97.7	97.40

made in 11CN-20-091; all junctions were validated and polished as described above. One intrachromosomal rearrangement supported by read alignments was identified between these two isolates.

The assembly of 11CN-03-130 consists of nine contigs which have an N_{50} value of 3.74 Mb and a total length of 28.22 Mb. Of the 8 largest contigs, 7 have telomeric repeats (TTAGG[GA]) at both ends; contig000003 has telomeric repeats at the end, and the start consists of rRNA gene repeats. The smallest contig (82.9 kb) consists of ~11 tandem copies of rRNA gene repeat units and is likely linked to the start of contig000003. The assembly of 11CN-20-091 consists of 8 contigs that have telomeric repeats at both ends, an N_{50} value of 3.70 Mb, and a total length of 28.20 Mb. The GC content of both assemblies is 46.8%. The total size is similar to those previously noted for draft assemblies of *T. marneffei* isolates ATCC 18225 (28.6 Mb) (12) and PM1 (28.9 Mb) (13).

Gene structures were predicted using transcriptome sequencing (RNA-seq) data from yeast and mycelia (14). RNA-seq reads were aligned to each assembly using STAR v2.7 (15), with the parameter “-alignIntronMax 10000,” and alignments were input to BRAKER v1.7 (16). A total of 10,025 genes were predicted for 11CN-03-130, and 9,994 genes were predicted for 11CN-20-091. BUSCO v3 (17) identified 97.7% and 97.4% of the peizomycotina_odb9 gene set in the 11CN-03-130 and 11CN-20-091 gene sets, respectively.

Data availability. The sequence, assembly, and annotation reported here are available in GenBank under BioProject accession number [PRJNA522919](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA522919). Raw sequence reads have been deposited in the NCBI Sequence Read Archive for 11CN-03-130 (Oxford Nanopore data, accession number [SRR8592562](https://www.ncbi.nlm.nih.gov/sra/SRR8592562); Illumina iSeq data, accession number [SRR8784960](https://www.ncbi.nlm.nih.gov/sra/SRR8784960); and Illumina HiSeq 4000 data, accession number [SRR10359552](https://www.ncbi.nlm.nih.gov/sra/SRR10359552)) and for 11CN-20-091 (Oxford Nanopore data, accession number [SRR8592561](https://www.ncbi.nlm.nih.gov/sra/SRR8592561); Illumina iSeq data, accession number [SRR8784959](https://www.ncbi.nlm.nih.gov/sra/SRR8784959); and Illumina HiSeq 4000 data, accession number [SRR10359551](https://www.ncbi.nlm.nih.gov/sra/SRR10359551)). Annotated assemblies are deposited under GenBank accession number [WINJ000000000](https://www.ncbi.nlm.nih.gov/nuccore/WINJ000000000) for 11CN-03-130 and under accession numbers [CP045653](https://www.ncbi.nlm.nih.gov/nuccore/CP045653) to [CP045660](https://www.ncbi.nlm.nih.gov/nuccore/CP045660) for 11CN-20-091.

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REFERENCES

1. Le T, Wolbers M, Chi NH, Quang VM, Chinh NT, Lan NPH, Lam PS, Kozal MJ, Shikuma CM, Day JN, Farrar J. 2011. Epidemiology, seasonality, and predictors of outcome of AIDS-associated *Penicillium marneffei* infection in Ho Chi Minh City, Viet Nam. *Clin Infect Dis* 52:945–952. <https://doi.org/10.1093/cid/cir028>.
2. Hu Y, Zhang J, Li X, Yang Y, Zhang Y, Ma J, Xi L. 2013. *Penicillium marneffei*

- infection: an emerging disease in mainland China. *Mycopathologia* 175: 57–67. <https://doi.org/10.1007/s11046-012-9577-0>.
3. Kawila R, Chaiwarith R, Supparatpinyo K. 2013. Clinical and laboratory characteristics of *penicilliosis marneffei* among patients with and without HIV infection in Northern Thailand: a retrospective study. *BMC Infect Dis* 13:464. <https://doi.org/10.1186/1471-2334-13-464>.
 4. Le T, Kinh NV, Cuc NTK, Tung NLN, Lam NT, Thuy PTT, Cuong DD, Phuc PTH, Vinh VH, Hanh DTH, Tam VV, Thanh NT, Thuy TP, Hang NT, Long HB, Nhan HT, Wertheim HFL, Merson L, Shikuma C, Day JN, Chau NVV, Farrar J, Thwaites G, Wolbers M, IVAP Investigators. 2017. A trial of itraconazole or amphotericin B for HIV-associated talaromycosis. *N Engl J Med* 376: 2329–2340. <https://doi.org/10.1056/NEJMoa1613306>.
 5. Henk DA, Shahar-Golan R, Devi KR, Boyce KJ, Zhan N, Fedorova ND, Nierman WC, Hsueh P-R, Yuen K-Y, Sieu TPM, Kinh NV, Wertheim H, Baker SG, Day JN, Vanittanakom N, Bignell EM, Andrianopoulos A, Fisher MC. 2012. Clonality despite sex: the evolution of host-associated sexual neighborhoods in the pathogenic fungus *Penicillium marneffei*. *PLoS Pathog* 8:e1002851. <https://doi.org/10.1371/journal.ppat.1002851>.
 6. Koren S, Walenz BP, Berlin K, Miller JR, Bergman NH, Phillippy AM. 2017. Canu: scalable and accurate long-read assembly via adaptive k-mer weighting and repeat separation. *Genome Res* 27:722–736. <https://doi.org/10.1101/gr.215087.116>.
 7. Li H. 2018. Minimap2: pairwise alignment for nucleotide sequences. *Bioinformatics* 34:3094–3100. <https://doi.org/10.1093/bioinformatics/bty191>.
 8. Li H. 2013. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. *arXiv arXiv:1303.3997*. <https://arxiv.org/abs/1303.3997>.
 9. Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. *PLoS One* 9:e112963. <https://doi.org/10.1371/journal.pone.0112963>.
 10. Kurtz S, Phillippy A, Delcher AL, Smoot M, Shumway M, Antonescu C, Salzberg SL. 2004. Versatile and open software for comparing large genomes. *Genome Biol* 5:R12. <https://doi.org/10.1186/gb-2004-5-2-r12>.
 11. Robinson JT, Thorvaldsdóttir H, Winckler W, Guttman M, Lander ES, Getz G, Mesirov JP. 2011. Integrative genomics viewer. *Nat Biotechnol* 29: 24–26. <https://doi.org/10.1038/nbt.1754>.
 12. Nierman WC, Fedorova-Abrams ND, Andrianopoulos A. 2015. Genome sequence of the AIDS-associated pathogen *Penicillium marneffei* (ATCC 18224) and its near taxonomic relative *Talaromyces stipitatus* (ATCC 10500). *Genome Announc* 3:e01559-14. <https://doi.org/10.1128/genomeA.01559-14>.
 13. Woo PCY, Lau SKP, Liu B, Cai JJ, Chong KTK, Tse H, Kao RYT, Chan C-M, Chow W-N, Yuen K-Y. 2011. Draft genome sequence of *Penicillium marneffei* strain PM1. *Eukaryot Cell* 10:1740–1741. <https://doi.org/10.1128/EC.05255-11>.
 14. Yang E, Chow W-N, Wang G, Woo PCY, Lau SKP, Yuen K-Y, Lin X, Cai JJ. 2014. Signature gene expression reveals novel clues to the molecular mechanisms of dimorphic transition in *Penicillium marneffei*. *PLoS Genet* 10:e1004662. <https://doi.org/10.1371/journal.pgen.1004662>.
 15. Dobin A, Davis CA, Schlesinger F, Drenkow J, Zaleski C, Jha S, Batut P, Chaisson M, Gingeras TR. 2013. STAR: ultrafast universal RNA-seq aligner. *Bioinformatics* 29:15–21. <https://doi.org/10.1093/bioinformatics/bts635>.
 16. Hoff KJ, Lange S, Lomsadze A, Borodovsky M, Stanke M. 2016. BRAKER1: unsupervised RNA-seq-based genome annotation with GeneMark-ET and AUGUSTUS. *Bioinformatics* 32:767–769. <https://doi.org/10.1093/bioinformatics/btv661>.
 17. Simão FA, Waterhouse RM, Ioannidis P, Kriventseva EV, Zdobnov EM. 2015. BUSCO: assessing genome assembly and annotation completeness with single-copy orthologs. *Bioinformatics* 31:3210–3212. <https://doi.org/10.1093/bioinformatics/btv351>.