www.aginganddisease.org

Review

### Kita-Kyushu Lung Cancer Antigen-1 (KK-LC-1): A Promising Cancer Testis Antigen

### Rui Bai<sup>1</sup>, Cheng Yuan<sup>2\*</sup>

<sup>1</sup>Department of Radiation and Medical Oncology, Zhongnan Hospital of Wuhan University, Wuhan, Hubei, China. <sup>2</sup>Department of Gynecological Oncology, Zhongnan Hospital, Wuhan University, Wuhan, China.

[Received October 28, 2021; Revised December 6, 2021; Accepted December 7, 2021]

ABSTRACT: Cancer has always been a huge problem in the field of human health, and its early diagnosis and treatment are the key to solving this problem. Cancer testis antigens (CTAs) are a family of multifunctional proteins that are specifically expressed in male spermatozoa and tumor cells but not in healthy somatic cells. Studies have found that CTAs are involved in the occurrence and development of tumors, and some CTAs trigger immunogenicity, which suggests a possibility of tumor immunotherapy. The differential expression and function of CTAs in normal tissues and tumor cells can promote the screening of tumor markers and the development of new immunotherapies. This article introduces the expression of Kita-Kyushu lung cancer antigen-1 (KK-LC-1), a new member of the CTA family, in different types of tumors and its role in immunotherapy.

Key words: KK-LC-1, Cancer testis antigen, Immunotherapy, Carcinogenesis, Immune target

### 1. Introduction

Cancer testis antigens (CTAs) are a class of proteins encoded by 276 genes that are only expressed in the testes, not in any other normal tissues [1-3]. Approximately half of the CTA genes are encoded on the X chromosome, so they are called CT-X genes [4, 5]. These CTAs belong to the typical testicular restriction type and are more immunogenic than non-CT-X genes located on autosomes [6].

CTAs were first reported in 1977 and have been found to be abnormally expressed in a variety of cancers, including melanoma, head and neck cancer, lung cancer, liver cancer, gastric cancer, ovarian cancer and breast cancer [7-13]. CTAs are not only closely related to the stemness of tumor cells, tumorigenicity, mobility, metastasis and the drug resistance of cancer cells [3, 1425], but they also show high tumor specificity and sensitivity [26-30]. Because of these characteristics, CTAs are considered to be tumor-specific markers that can be used in cancer diagnosis and prognostication, and possibly as targets for cancer treatment [31-33].

The use of autologous typing is a turning point in the search for tumor antigens. Autologous typing is a method that uses autoantibodies and T cells of cancer patients to identify tumor cells and normal cells to determine whether the patient has a tumor-specific T cell response and antibodies [34]. Using this method, a series of tumor antigens were found, including viral oncoproteins, mutant proteins, fusion proteins, overexpressed proteins, differentiation proteins and cancer testis antigens.

The first CTA was cloned by van der Bruggen and others in 1991 [35], and then BAGE [36] and GAGE [37] were discovered one after another. In 1995, Sahin U's

\*Correspondence should be addressed to: Dr. Cheng Yuan, Department of Gynecological Oncology, Zhongnan Hospital, Wuhan University, Wuhan, China. Wuhan, Hubei 430071, China. E-mail: 2018203030071@whu.edu.cn.

**Copyright:** © 2021 Bai R & Yuan C. This is an open-access article distributed under the terms of the <u>Creative Commons Attribution</u> <u>License</u>, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

team improved the identification of CTA through serological analysis of a recombinant cDNA expression library (SEREX) and used autologous patient serum to screen a phage display library derived from tumor cDNA [38]. This technology contributes to the discovery of a variety of tumor antigens, including NY-ESO-1, the most successful tumor immunotherapy target thus far [39]. Subsequently, to reflect the expression specificity of such tumor-associated antigens, CHEN et al. introduced the term "cancer/testis antigen", namely, CTA [40].

At present, the CTA database (www.cta.lncc.br/ index.php) includes more than 200 genes, and the number is still increasing. These tumor antigens mainly include mutated genes and overexpressed genes that are ubiquitous in tissues, as well as genes that are not expressed in normal adult cells but are expressed in cancer cells. At present, more than 730 CTAs have been identified, but it is still uncertain whether some CTAs identified by expression data are immunogenic [41-43]. In this article, we will introduce the characteristics and therapeutic prospects of Kita-Kyushu lung cancer antigen-1 (KK-LC-1), a newly discovered member of the CTA family.

KK-LC-1, whose full name is Kita-Kyushu lung cancer antigen-1, also known as CT83 or cxorf61, was discovered by Takashi Fukuyama's team in 2006[44]. Takashi Fukuyama derived lung adenocarcinoma cell lines from the tumors of lung cancer patients, stimulated the regional lymph node cells of the patients, induced cytotoxic T lymphocyte (CTL) clones, and then identified new antigen coding genes by screening a cDNA library from allogeneic lung cancer cell lines.

KK-LC-1 is located on chromosome Xq22 and consists of 556bp. It is not expressed in normal tissues except the testis but is highly expressed in lung cancer, gastric cancer and breast cancer. At present, it has been reported that KK-LC-1 plays a role in the immune response as a new antigen, but the structure and function of this gene are not fully understood. How this gene plays a role in diseases remains to be further studied.

## 2. Expression and significance of KK-LC-1 in different tumors

KK-LC-1 is abnormally expressed in different types of cancers, including lung cancer, gastric cancer, breast cancer and liver cancer [44-47], but the biological function and potential mechanism of KK-LC-1 in cancer are still unclear.

### 2.1 Gastric cancer

In 2015, Masahiko Watanabe's team found that KK-LC-1 was highly expressed in gastric cancer, and the expression

was higher than that of other CTAs, such as MAGE-A1, MAGE-A3 and NY-ESO-1 [48]. In 2017, Masahiko Watanabe's team found that the expression of KK-LC-1 in stage I gastric cancer was very high, suggesting that KK-LC-1 can be used as a potential marker for the early diagnosis and treatment of gastric cancer [49]. In the same year, Noritada Kobayashi's team found that KK-LC-1 was highly expressed in gastric cancer caused by Helicobacter pylori infection, suggesting that Helicobacter pylori infection may induce the expression of specific CTAs [45]. In 2020, Yoshihito Takahashi's team found that the expression of KK-LC-1 in gastric cancer was related to Helicobacter pylori infection and atrophy [50]. A combination of improved ABCD methods (serological detection of Helicobacter pylori (HP) antibodies and the pepsinogen (PG) method for risk stratification of gastric cancer) and KK-LC-1 detection may improve the accuracy of diagnosis. To better detect the expression of KK-LC-1, in 2019, Noritada Kobayashi's team synthesized a new antibody, Kmab34B3, which can successfully detect KK-LC-1 in gastric cancer cells and tissues [51].

#### 2.2 Liver cancer

By examining 60 pairs of liver cancer and paracancerous tissues, the researchers determined that KK-LC-1 is highly expressed in liver cancer and is closely related to the prognosis of liver cancer [47]. A series of phenotypic experiments were carried out by knocking down or overexpressing KK-LC-1. KK-LC-1 can promote the proliferation, invasion and migration of hepatocellular carcinoma cells. An animal model further verified the tumor-promoting effect of KK-LC-1 in vivo. In terms of the mechanism, the researchers found that KK-LC-1 can regulate the expression of the Notch1 intracellular domain (NICD1) and Notch1 effector Hes1. In vivo experiments also verified the correlation between KK-LC-1 and NICD1. Further exploration found that KK-LC-1, through interaction with presenilin-1, activates the Notch1 signaling pathway and then it plays a role in promoting the growth, migration and invasion of liver cancer, thus triggering the tumorigenesis of liver cancer. Presenilin-1 is the catalytic subunit of endosynthesis that can catalyze the Notch1 gene. These results suggest that high levels of KK-LC-1 may be an independent predictor of poor survival in liver cancer patients [47].

#### 2.3 Breast cancer

In 2018, Masahiko Watanabe's team analyzed the surgical specimens of 51 patients with breast cancer and found that KK-LC-1 could be detected in triple-negative breast cancer cases and all tumors without estrogen receptor

expression, and its expression level in tumor tissues was significantly higher than that in paracancerous tissues. KK-LC-1 can be used as a marker for the clinical diagnosis and immunotherapy of breast cancer [46].

### 2.4 Lung cancer

In 2018, a team from a university in China constructed a CTA map of stage III lung cancers requiring surgery through a comprehensive analysis of 10 kinds of CTAs of NSCLC [52]. The purpose of this study was to find the most appropriate CTA indicators to assist in decision-making. At the same time, the CTA maps also had therapeutic potential for TCR-T cell treatment.

Takeshi Hanagiri's team found that the positive rate of KK-LC-1 was 30.9% by examining clinical samples of surgical resection of non-small-cell lung cancer (NSCLC). The decreased expression of class I molecules indicates a poor prognosis among patients with positive CTAs and is an important obstacle to tumor antigen immunotherapy. This provides a new idea for the immunotherapy of CTAs, and the future synergistic therapy of CTAs and HLA may be a new breakthrough [53].

# 3. Application of targeted KK-LC-1 in tumor immunotherapy

In view of the high immunogenicity and tumor specificity of CTAs, carcinogenic CTAs are sensitive targets for cancer immunotherapy [54-64]. In recent decades, several targeted immunotherapies for carcinogenic CTAs have been developed, and these immunotherapies have been tested in preclinical and early clinical environments. At present, most clinical studies have focused on the treatment of melanoma-associated antigen A (MAGEA) and New York esophageal squamous cell carcinoma-1 (NY-ESO-1), including cancer vaccines targeting CTAs to prevent tumor occurrence and development, and monoclonal antibodies against CTAs and CAR-T designed based on CTAs [65-73]. Due to the late discovery of KK-LC-1, the reported treatments include vaccines, photodynamic therapy combined with new photosensitizers and TCR-T therapy (Fig. 1).



**Figure 1. Targeting KK-LC-1 for effective cancer immunotherapy.** The strategies for effective cancer immunotherapy focused on KK-LC-1 include vaccines, photodynamic therapy, and TCR-T cells. The mechanism of CTA reactivation may be DNA methylation and histone modification.

### 3.1 Vaccine

The vaccine designed for DNA of MAGE-A in 2018 has shown anticancer properties in a number of clinical trials[74]. A clinical trial published in Nature in 2020 showed that liposome RNA (RNA-LPX) vaccines based on MAGEA, and two other kinds of CTAs could induce strong cellular immunity [75]. Because KK-LC-1 is also

antigen-specific, we suspect that it is possible to synthesize a vaccine against KK-LC-1. A new paper in 2021 reported the design of a multiepitope vaccine against MAGEA3, MAGEA4, NY-ESO-1 and KK-LC-1 using reverse vaccinology for the first time [76]. The main idea is to draw the epitope map according to the CTA sequence, estimate the population coverage (PC) of the CD4+ and CD8+ epitopes, and then connect the candidate linear B cell (BL), CD4+ and CD8+ epitopes into a multiepitope structure (Mvax) using the flagellin domain as an adjuvant. After successful construction, the tertiary structure of Mvax was modeled and verified, and the antigenicity and cross-reactivity of Mvax was tested. It is critical that all epitopes contained in the vaccine dock with their human leukocyte antigen (HLA) conjugates to better act on the antigenic fragments presented by HLA. Finally, the researchers successfully designed a multiepitope vaccine targeting CTAs in NSCLC. Computer evaluation showed that Mvax has antigenicity, immunogenicity, stability and safety and is expected to be used in in vitro and in vivo studies.

### 3.2 Photodynamic therapy

Photodynamic therapy (PDT) is a noninvasive and highly selective tumor therapy, but its therapeutic effect has been limited by skin phototoxicity for a long time. Therefore, to overcome this disadvantage, it is best to selectively deliver photosensitizers to tumor cells with the help of specific antibodies against tumor-associated antigens. Some researchers developed and identified a new mouse monoclonal antibody (CT83 MAb7G4) against human CT83, which can effectively combine with the new photosensitizer gallium (III)5,10,15-tris (ethoxycarbonyl) corrole (1-Ga) to form the antibody-photosensitizer complex 7G4-1-Ga [77]. Enzyme-linked immunosorbent assays (ELISA), flow cytometry and cytotoxicity assays showed that 7G4-1-GA had high specificity for CT83. In addition, 7G4-1-Ga has a stronger cytotoxic effect on human tumor cells expressing CT83 than 1-Ga in vitro. These results suggest that a combination of anti-CT83 monoclonal antibody and antibody-conjugated photosensitizer 1-GA may have good application prospects in tumors with high expression of CT83.

### 3.3 TCR-T

TCR-T is used to extract the  $\alpha$  and  $\beta$  chain genes encoding TCR from effector T cells induced by tumor antigens. It is introduced to mature T cells by genetic engineering technology, and then transfused back into patients who lack tumor antigen specific response T cells, so that recipient T cells express antigen specific TCR and exert the function of effector T cells. At present, the cancer

testis antigen NY-ESO-1 is mostly being studied in clinical research [78]. The expression of cancer testis antigen is very low in normal tissues, and the probability of off-target effects is low. It is an ideal target antigen for adoptive cellular immunotherapy [79]. NY-ESO-1 TCR-T therapy was used in 20 patients with multiple myeloma, the clinical response rate was 80%, and TCR was continuously expressed in vivo for more than two years [21]. Eighteen patients with metastatic synovial sarcoma and 20 patients with melanoma were treated with TCR-T, and the clinical response rates were 61% and 55%, respectively [3]. Given that KK-LC-1 and NY-ESO-1 have many similar features, we suspect that TCR-T can also be designed for KK-LC-1.

In 2019, Bridget Marcinkowski et al. designed TCR-T presented by HLA-A\*01:01, targeting KK-LC-152-60[80]. The researchers tested whether T cells transduced with KK-LC-1 TCR (KK-LC-1TCR-Ts) could recognize tumor cell lines expressing KK-LC-1 and HLA-A\*01:01 in vitro. In the overnight coculture test, TCR-T cells recognized cell lines expressing target antigens and HLAlimiting elements and released interferon- $\gamma$ . To evaluate whether KK-LC-1 TCR-T cells can mediate the antitumor response in vivo, researchers constructed a mouse xenograft model. The results showed that the model with low expression of KK-LC-1 on the surface of tumor cells was prone to relapse, and the model with high expression of KK-LC-1 could make the tumor regress after a single intravenous injection of KK-LC-1 TCR-Ts. These results suggest that KK-LC-1 TCR-Ts can prevent tumor progression both in vitro and in vivo.

### 4. Potential regulatory mechanism of KK-LC-1

CTAs are immunogenic proteins, so they can trigger cellular immunity and humoral immunity. Given that their expression in adult somatic tissues is greatly restricted and has immunogenic potential, CTAs are considered good candidate targets for cancer immunotherapy. CTA-based treatments include antibodies, vaccines and anti-CTA chimeric antigen receptor-modified T cells (CAR-Ts). Although several CTA-targeted therapies have achieved encouraging results in preclinical and early clinical trials, the anticancer effect of CTA-targeted immunotherapy is not ideal, so these immunotherapies have not yet been used in the clinic as first-line anticancer drugs [81-83].

Although immunotherapy for CTAs is theoretically feasible, it is difficult to implement immunotherapy due to the low and local expression of CTAs in tumors [84, 85]. Therefore, we urgently need to explore the transcriptional regulation mechanism of CTAs in tumor cells to promote the expression of CTAs. It has been reported that the reactivation of CTAs is primarily due to changes in DNA methylation in the genome [5, 20, 86,

87]. Bisulfite sequencing showed that compared with the corresponding normal tissues, the promoter region of CTAs in many tumors was hypomethylated, resulting in increased protein expression [88, 89]. There are abundant CpG islands in the promoter region of the CTA genes, which is an important reason why these genes are sensitive to methylation regulation.

Weber et al. proposed for the first time that the treatment of tumor cells with dexitabine, an inhibitor of DNA methyltransferase, can reactivate the expression of MAGEA1 [90]. This suggests that demethylation of genomic DNA can activate CTA expression. Later, it was gradually found that DNA demethylation could reactivate the expression of NY-ESO-1, KK-LC-1 and other CTAs genes [66, 91-96]. Researchers found that the DNA methylation level is negatively correlated with the expression level of KK-LC-1. Experiments with 5-aza-2'-deoxycytidine (5-aza-dC), methylation-specific PCR (MSP), and bisulfite sequencing PCR (BSP) also confirmed this conclusion.

In addition to DNA demethylation, which is considered to be the main factor in the activation of CTAs, histone acetylation also contributes to the transcriptional activation of CTAs and enhances the activation of tumor testis antigen genes by DNMTi [97]. Inhibition of HDAC alone could not activate the transcription of CTAs, but its combination with a DNA methyltransferase inhibitor could significantly enhance the transcription of CTAs. These strategies can effectively promote the expression of CTAs to enhance their immunogenicity, thus improving the response to T cell-based therapy [98].

Although epigenetic mechanisms play an important role in regulating the expression of tumor testis antigen genes, there is growing evidence that other nonepigenetic mechanisms also play a key role. At present, these mechanisms are not well understood, and the mechanisms currently recognized include sequence-specific transcription factors, signal transduction and activated tyrosine kinases. For example, two Ets binding sites are involved in the transcriptional activation of the MAGE1 gene [99-101], tyrosine kinase affects the methylation and expression of the MAGE gene promoter [102-107], cAMP increases the expression of MAGE-A11 in prostate cells [108], SP1 increases the expression of NY-ESO-1 in lung cancer cells [109], and there is a loop regulation between p53 and tumor testis antigen [110]. BORIS itself is a member of the CTA family [111-114], which is reactivated by hypomethylation in cancer. It can bind to SET1A H3K4, a methyltransferase associated with histone transcriptional activity modification, and then regulate the expression of other members of the CTA family [115]. The mechanism of epigenetic reactivation of KK-LC-1 expression remains to be further studied.

### 5. Discussion

At present, the exploration of KK-LC-1 is based on clinical research. To further understand the mechanism of KK-LC-1, more basic research needs to be carried out. The starting point can be carried out from the following two aspects:

There are antibodies available against NY-ESO-1, PRAME, and CT45. In 2019, Seth M. Pollack's team used the antibody LV305 against NY-ESO-1 for the first time in the clinic, and the results showed good safety and antigen-specific responses [116]. From AS15 in 2015 to Pr20 in 2017, antibodies against PRAME have been well verified in animal models [117, 118], and the new anti-PRAME monoclonal antibody developed in 2021 is more powerful and can recognize folded proteins on the surface of cell membranes [119]. In 2018, a report published in the journal Cell showed that CT45-derived HLA-I peptides can activate patient-derived cytotoxic T cells to kill and inhibit tumor progression [120]. Since KK-LC-1 belongs to the CTA family, we believe that it is possible to design antibodies against KK-LC-1 in the future.

In addition, an increasing number of studies have focused on a combination of targeted therapies against CTAs and immune checkpoint inhibitors[54, 121, 122]. In 2016, Lindy G Durrant's team found that the long-term survival rate of mice was significantly improved after treatment with a combination of the NY-ESO-1 vaccine SCIB2 and anti-PD-1/CTLA-4, suggesting the feasibility of combination therapy [123]. In the future, SCIB2 can be used in the clinic. For patients with a low tumor mutation load, SCIB2 alone may be effective, but for patients with a high tumor mutation load, the effect of combined therapy may be more significant. In 2019, Mikio Oka's team found that patients with non-small-cell lung cancer treated with NY-ESO-1 and XAGE1 monoclonal antibodies combined with anti-PD-1 had better efficacy and longer survival, and the antibody titer was positively correlated with the efficacy of anti-PD-1 therapy [124]. Therefore, we believe that NY-ESO-1 and XAGE1 antibodies are markers for predicting the efficacy of anti-PD-1 in patients with NSCLC and can be included in clinical trials of immune checkpoint inhibitors as stratification factors in the future. In 2020, Baoen Shan's team found that although esophageal cancer expressed MAGE-A11, the expression level was not high, thus limiting the effectiveness of immunotherapy. MAGE-A11-derived CTLs can kill esophageal cancer cells expressing MAGE-A11 but have almost no killing effect on MAGE-A11-negative tumor cells [29]. At the same time, researchers found that PD-L1 can affect the antitumor function of CTLs. Therefore, they think a combination of DNA methyltransferase inhibitors and PDL1 inhibitors may be an effective approach. The results of cellular and animal experiments also confirmed that combination therapy could produce a specific cellular immune response to esophageal tumors, a promising strategy for clinical applications.

However, there is currently no literature reporting on a combination of KK-LC-1 and immune checkpoint inhibitors, and the simultaneous use of both may be a valuable method for clinical applications in the future. Concluding remarks.

In summary, KK-LC-1 is a new target for immunotherapy and may become a valuable tumorrelated marker in the future. Immunotherapy and combination therapy against KK-LC-1 may create new opportunities for cancer treatment. In the future, these new treatment strategies may result in new breakthroughs in cancer immunotherapy.

### **Conflict of Interests**

All of authors have declared that no competing interest exists.

### References

- [1] Holden S, Bernard O, Artzt K, Whitmore WF, Bennett D (1977). Human and mouse embryonal carcinoma cells in culture share an embryonic antigen (F9). Nature, 270:518-520.
- [2] Hogan B, Fellous M, Avner P, Jacob F (1977). Isolation of a human teratoma cell line which expresses F9 antigen. Nature, 270:515-518.
- [3] Gibbs ZA, Whitehurst AW (2018). Emerging Contributions of Cancer/Testis Antigens to Neoplastic Behaviors. Trends Cancer, 4:701-712.
- Fratta E, Coral S, Covre A, Parisi G, Colizzi F, Danielli R, et al. (2011). The biology of cancer testis antigens: putative function, regulation and therapeutic potential. Mol Oncol, 5:164-182.
- [5] Simpson AJG, Caballero OL, Jungbluth A, Chen Y-T, Old LJ (2005). Cancer/testis antigens, gametogenesis and cancer. Nat Rev Cancer, 5:615-625.
- [6] Hofmann O, Caballero OL, Stevenson BJ, Chen Y-T, Cohen T, Chua R, et al. (2008). Genome-wide analysis of cancer/testis gene expression. Proc Natl Acad Sci U S A, 105:20422-20427.
- [7] Rooney MS, Shukla SA, Wu CJ, Getz G, Hacohen N (2015). Molecular and genetic properties of tumors associated with local immune cytolytic activity. Cell, 160:48-61.
- [8] Wu SC, Münger K (2021). Role and Clinical Utility of Cancer/Testis Antigens in Head and Neck Squamous Cell Carcinoma. Cancers (Basel), 13.
- [9] Lam RA, Tien TZ, Joseph CR, Lim JX, Thike AA, Iqbal J, et al. (2021). Cancer-Testis Antigens in Triple-Negative Breast Cancer: Role and Potential Utility in Clinical Practice. Cancers (Basel), 13.
- [10] Zhu Y, Zhu X, Tang C, Guan X, Zhang W (2021).

Progress and challenges of immunotherapy in triplenegative breast cancer. Biochim Biophys Acta Rev Cancer, 1876:188593.

- [11] Lu H, Zhang Q, He S, Liu S, Xie Z, Li X, et al. (2021). Reduction-Sensitive Fluorinated-Pt(IV) Universal Transfection Nanoplatform Facilitating CT45-Targeted CRISPR/dCas9 Activation for Synergistic and Individualized Treatment of Ovarian Cancer. Small, 17:e2102494.
- [12] Fujiya K, Terashima M, Ohshima K, Aizawa D, Sugino T, Serizawa M, et al. (2021). MAGEA10 expression is a predictive marker of early hepatic recurrence after curative gastrectomy for gastric and gastroesophageal junction cancer. Gastric Cancer, 24:341-351.
- [13] Craig AJ, Garcia-Lezana T, Ruiz de Galarreta M, Villacorta-Martin C, Kozlova EG, Martins-Filho SN, et al. (2021). Transcriptomic characterization of cancer-testis antigens identifies MAGEA3 as a driver of tumor progression in hepatocellular carcinoma. PLoS Genet, 17:e1009589.
- [14] DeWolf WC, Lange PH, Einarson ME, Yunis EJ (1979). HLA and testicular cancer. Nature, 277:216-217.
- [15] Whitfield WG, González C, Sánchez-Herrero E, Glover DM (1989). Transcripts of one of two Drosophila cyclin genes become localized in pole cells during embryogenesis. Nature, 338:337-340.
- [16] Gliki G, Ebnet K, Aurrand-Lions M, Imhof BA, Adams RH (2004). Spermatid differentiation requires the assembly of a cell polarity complex downstream of junctional adhesion molecule-C. Nature, 431:320-324.
- [17] Chen Y-T, Chadburn A, Lee P, Hsu M, Ritter E, Chiu A, et al. (2010). Expression of cancer testis antigen CT45 in classical Hodgkin lymphoma and other B-cell lymphomas. Proc Natl Acad Sci U S A, 107:3093-3098.
- [18] Scudellari M (2011). A ballsy search for cancer targets. Nat Med, 17:916-918.
- [19] Yamada R, Takahashi A, Torigoe T, Morita R, Tamura Y, Tsukahara T, et al. (2013). Preferential expression of cancer/testis genes in cancer stem-like cells: proposal of a novel sub-category, cancer/testis/stem gene. Tissue Antigens, 81:428-434.
- [20] Whitehurst AW (2014). Cause and consequence of cancer/testis antigen activation in cancer. Annu Rev Pharmacol Toxicol, 54:251-272.
- [21] Smith ZD, Shi J, Gu H, Donaghey J, Clement K, Cacchiarelli D, et al. (2017). Epigenetic restriction of extraembryonic lineages mirrors the somatic transition to cancer. Nature, 549:543-547.
- [22] Wen M, Ren H, Zhang S, Li T, Zhang J, Ren P (2021). CT45A1 promotes the metastasis of osteosarcoma cells in vitro and in vivo through β-catenin. Cell Death Dis, 12:650.
- [23] Cheng C-C, Wooten J, Gibbs ZA, McGlynn K, Mishra P, Whitehurst AW (2020). Sperm-specific COX6B2 enhances oxidative phosphorylation, proliferation, and survival in human lung adenocarcinoma. Elife, 9.
  [24] Lazar I, Fabre B, Feng Y, Khateb A, Turko P, Martinez

Gomez JM, et al. (2020). SPANX Control of Lamin A/C Modulates Nuclear Architecture and Promotes Melanoma Growth. Mol Cancer Res, 18:1560-1573.

- [25] Yeon M, Kim Y, Pathak D, Kwon E, Kim DY, Jeong MS, et al. (2021). The CAGE-MiR-181b-5p-S1PR1 Axis Regulates Anticancer Drug Resistance and Autophagy in Gastric Cancer Cells. Front Cell Dev Biol, 9:666387.
- [26] Gjerstorff MF, Andersen MH, Ditzel HJ (2015). Oncogenic cancer/testis antigens: prime candidates for immunotherapy. Oncotarget, 6:15772-15787.
- [27] Mahmoud AM (2018). Cancer testis antigens as immunogenic and oncogenic targets in breast cancer. Immunotherapy, 10:769-778.
- [28] Wei X, Chen F, Xin K, Wang Q, Yu L, Liu B, et al. (2019). Cancer-Testis Antigen Peptide Vaccine for Cancer Immunotherapy: Progress and Prospects. Transl Oncol, 12:733-738.
- [29] Wu Y, Sang M, Liu F, Zhang J, Li W, Li Z, et al. (2020). Epigenetic modulation combined with PD-1/PD-L1 blockade enhances immunotherapy based on MAGE-A11 antigen-specific CD8+T cells against esophageal carcinoma. Carcinogenesis, 41:894-903.
- [30] Daiko H, Marafioti T, Fujiwara T, Shirakawa Y, Nakatsura T, Kato K, et al. (2020). Exploratory openlabel clinical study to determine the S-588410 cancer peptide vaccine-induced tumor-infiltrating lymphocytes and changes in the tumor microenvironment in esophageal cancer patients. Cancer Immunol Immunother, 69:2247-2257.
- [31] da Silva VL, Fonseca AF, Fonseca M, da Silva TE, Coelho AC, Kroll JE, et al. (2017). Genome-wide identification of cancer/testis genes and their association with prognosis in a pan-cancer analysis. Oncotarget, 8:92966-92977.
- [32] Cui Z, Chen Y, Hu M, Lin Y, Zhang S, Kong L, et al. (2020). Diagnostic and prognostic value of the cancertestis antigen lactate dehydrogenase C4 in breast cancer. Clin Chim Acta, 503:203-209.
- [33] Hou Z, Liang X, Wang X, Zhou Z, Shi G (2020). Myeloid-derived suppressor cells infiltration in nonsmall-cell lung cancer tumor and MAGE-A4 and NY-ESO-1 expression. Oncol Lett, 19:3982-3992.
- [34] Knuth A, Danowski B, Oettgen HF, Old LJ (1984). Tcell-mediated cytotoxicity against autologous malignant melanoma: analysis with interleukin 2dependent T-cell cultures. Proc Natl Acad Sci U S A, 81:3511-3515.
- [35] van der Bruggen P, Traversari C, Chomez P, Lurquin C, De Plaen E, Van den Eynde B, et al. (1991). A gene encoding an antigen recognized by cytolytic T lymphocytes on a human melanoma. Science, 254:1643-1647.
- [36] Boël P, Wildmann C, Sensi ML, Brasseur R, Renauld JC, Coulie P, et al. (1995). BAGE: a new gene encoding an antigen recognized on human melanomas by cytolytic T lymphocytes. Immunity, 2:167-175.
- [37] Van den Eynde B, Peeters O, De Backer O, Gaugler B, Lucas S, Boon T (1995). A new family of genes coding for an antigen recognized by autologous cytolytic T

lymphocytes on a human melanoma. J Exp Med, 182:689-698.

- [38] Sahin U, Türeci O, Schmitt H, Cochlovius B, Johannes T, Schmits R, et al. (1995). Human neoplasms elicit multiple specific immune responses in the autologous host. Proc Natl Acad Sci U S A, 92:11810-11813.
- [39] Thomas R, Al-Khadairi G, Roelands J, Hendrickx W, Dermime S, Bedognetti D, et al. (2018). NY-ESO-1 Based Immunotherapy of Cancer: Current Perspectives. Front Immunol, 9:947.
- [40] Chen YT, Scanlan MJ, Sahin U, Türeci O, Gure AO, Tsang S, et al. (1997). A testicular antigen aberrantly expressed in human cancers detected by autologous antibody screening. Proc Natl Acad Sci U S A, 94:1914-1918.
- [41] Maxfield KE, Taus PJ, Corcoran K, Wooten J, Macion J, Zhou Y, et al. (2015). Comprehensive functional characterization of cancer-testis antigens defines obligate participation in multiple hallmarks of cancer. Nat Commun, 6:8840.
- [42] Li B, Li T, Pignon J-C, Wang B, Wang J, Shukla SA, et al. (2016). Landscape of tumor-infiltrating T cell repertoire of human cancers. Nat Genet, 48:725-732.
- [43] Bassani-Sternberg M, Bräunlein E, Klar R, Engleitner T, Sinitcyn P, Audehm S, et al. (2016). Direct identification of clinically relevant neoepitopes presented on native human melanoma tissue by mass spectrometry. Nat Commun, 7:13404.
- [44] Fukuyama T, Hanagiri T, Takenoyama M, Ichiki Y, Mizukami M, So T, et al. (2006). Identification of a new cancer/germline gene, KK-LC-1, encoding an antigen recognized by autologous CTL induced on human lung adenocarcinoma. Cancer Res, 66:4922-4928.
- [45] Fukuyama T, Futawatari N, Ichiki Y, Shida A, Yamazaki T, Nishi Y, et al. (2017). Correlation Between Expression of the Cancer/Testis Antigen KK-LC-1 and Infection in Gastric Cancer. In Vivo, 31:403-407.
- [46] Kondo Y, Fukuyama T, Yamamura R, Futawatari N, Ichiki Y, Tanaka Y, et al. (2018). Detection of KK-LC-1 Protein, a Cancer/Testis Antigen, in Patients with Breast Cancer. Anticancer Res, 38:5923-5928.
- [47] Chen Z, Zuo X, Pu L, Zhang Y, Han G, Zhang L, et al. (2019). Hypomethylation-mediated activation of cancer/testis antigen KK-LC-1 facilitates hepatocellular carcinoma progression through activating the Notch1/Hes1 signalling. Cell Prolif, 52:e12581.
- [48] Shida A, Futawatari N, Fukuyama T, Ichiki Y, Takahashi Y, Nishi Y, et al. (2015). Frequent High Expression of Kita-Kyushu Lung Cancer Antigen-1 (KK-LC-1) in Gastric Cancer. Anticancer Res, 35:3575-3579.
- [49] Futawatari N, Fukuyama T, Yamamura R, Shida A, Takahashi Y, Nishi Y, et al. (2017). Early gastric cancer frequently has high expression of KK-LC-1, a cancertestis antigen. World J Gastroenterol, 23:8200-8206.
- [50] Shida A, Fukuyama T, Futawatari N, Ohmiya H, Ichiki Y, Yamashita T, et al. (2020). Cancer/testis antigen,

Kita-Kyushu lung cancer antigen-1 and ABCD stratification for diagnosing gastric cancers. World J Gastroenterol, 26:424-432.

- [51] Takahashi Y, Fukuyama T, Futawatari N, Ichiki Y, Ohmiya H, Chuman M, et al. (2019). Expression of Kita-Kyushu Lung Cancer Antigen-1 as Detected by a Novel Monoclonal Antibody in Gastric Cancer. Anticancer Res, 39:6259-6263.
- [52] Jin S, Cao S, Grigorev A, Li J, Meng Q, Wang C, et al. (2018). Establishment of cancer/testis antigen profiling based on clinicopathological characteristics in resected pathological stage III non-small cell lung cancer. Cancer management and research, 10:2031-2046.
- [53] Hanagiri T, Shigematsu Y, Shinohara S, Takenaka M, Oka S, Chikaishi Y, et al. (2013). Clinical significance of expression of cancer/testis antigen and downregulation of HLA class-I in patients with stage I nonsmall cell lung cancer. Anticancer Res, 33:2123-2128.
- [54] Slingluff CL, Zarour HM, Tawbi HA-H, Kirkwood JM, Postow MA, Friedlander P, et al. (2021). A phase 1 study of NY-ESO-1 vaccine + anti-CTLA4 antibody Ipilimumab (IPI) in patients with unresectable or metastatic melanoma. Oncoimmunology, 10:1898105.
- [55] Jay A, Reitz D, Namekawa SH, Heyer W-D (2021). Cancer testis antigens and genomic instability: More than immunology. DNA Repair (Amst), 108:103214.
- [56] Naik A, Thomas R, Al-Khadairi G, Bacha R, Hendrickx W, Decock J (2021). Cancer testis antigen PRAME: An anti-cancer target with immunomodulatory potential. J Cell Mol Med, 25:10376-10388.
- [57] Almshayakhchi R, Nagarajan D, Vadakekolathu J, Guinn B-A, Reeder S, Brentville V, et al. (2021). A Novel HAGE/WT1-ImmunoBody Vaccine Combination Enhances Anti-Tumour Responses When Compared to Either Vaccine Alone. Front Oncol, 11:636977.
- [58] Bassan D, Gozlan YM, Sharbi-Yunger A, Tzehoval E, Greenstein E, Bitan L, et al. (2021). Avidity optimization of a MAGE-A1-specific TCR with somatic hypermutation. Eur J Immunol, 51:1505-1518.
- [59] Stanojevic M, Hont AB, Geiger A, O'Brien S, Ulrey R, Grant M, et al. (2021). Identification of novel HLArestricted preferentially expressed antigen in melanoma peptides to facilitate off-the-shelf tumorassociated antigen-specific T-cell therapies. Cytotherapy, 23:694-703.
- [60] Davari K, Holland T, Prassmayer L, Longinotti G, Ganley KP, Pechilis LJ, et al. (2021). Development of a CD8 co-receptor independent T-cell receptor specific for tumor-associated antigen MAGE-A4 for next generation T-cell-based immunotherapy. J Immunother Cancer, 9.
- [61] Wang W-L, Gokgoz N, Samman B, Andrulis IL, Wunder JS, Demicco EG (2021). RNA expression profiling reveals PRAME, a potential immunotherapy target, is frequently expressed in solitary fibrous tumors. Mod Pathol, 34:951-960.
- [62] Zhang Y, Yu X, Liu Q, Gong H, Chen A-A, Zheng H,

et al. (2021). SAGE1: a Potential Target Antigen for Lung Cancer T-Cell Immunotherapy. Mol Cancer Ther, 20:2302-2313.

- [63] Merhi M, Raza A, Inchakalody VP, Siveen KS, Kumar D, Sahir F, et al. (2020). Persistent anti-NY-ESO-1specific T cells and expression of differential biomarkers in a patient with metastatic gastric cancer benefiting from combined radioimmunotherapy treatment: a case report. J Immunother Cancer, 8.
- [64] Tsang YH, Mills GB (2020). The roles of variants in pancreatic cancer development and their potential impact on cancer immunotherapy. Autophagy, 16:1923-1924.
- [65] Chawla SP, Van Tine BA, Pollack SM, Ganjoo KN, Elias AD, Riedel RF, et al. (2021). Phase II Randomized Study of CMB305 and Atezolizumab Compared With Atezolizumab Alone in Soft-Tissue Sarcomas Expressing NY-ESO-1. J Clin Oncol:JCO2003452.
- [66] Holmberg-Thydén S, Dufva IH, Gang AO, Breinholt MF, Schejbel L, Andersen MK, et al. (2021). Epigenetic therapy in combination with a multiepitope cancer vaccine targeting shared tumor antigens for high-risk myelodysplastic syndrome - a phase I clinical trial. Cancer Immunol Immunother.
- [67] Ishikawa T, Kageyama S, Miyahara Y, Okayama T, Kokura S, Wang L, et al. (2021). Safety and antibody immune response of CHP-NY-ESO-1 vaccine combined with poly-ICLC in advanced or recurrent esophageal cancer patients. Cancer Immunol Immunother, 70:3081-3091.
- [68] Yang Y, Guo X, Hu B, He P, Jiang X, Wang Z, et al. (2021). Generated SecPen\_NY-ESO-1\_ubiquitinpulsed dendritic cell cancer vaccine elicits stronger and specific T cell immune responses. Acta Pharm Sin B, 11:476-487.
- [69] Vasileiou S, Lulla PD, Tzannou I, Watanabe A, Kuvalekar M, Callejas WL, et al. (2021). T-Cell Therapy for Lymphoma Using Nonengineered Multiantigen-Targeted T Cells Is Safe and Produces Durable Clinical Effects. J Clin Oncol, 39:1415-1425.
- [70] Rochigneux P, Chanez B, De Rauglaudre B, Mitry E, Chabannon C, Gilabert M (2021). Adoptive Cell Therapy in Hepatocellular Carcinoma: Biological Rationale and First Results in Early Phase Clinical Trials. Cancers (Basel), 13.
- [71] Przybyla A, Lehmann AA, Zhang T, Mackiewicz J, Galus Ł, Kirchenbaum GA, et al. (2021). Functional T Cell Reactivity to Melanocyte Antigens Is Lost during the Progression of Malignant Melanoma, but Is Restored by Immunization. Cancers (Basel), 13.
- [72] Somaiah N, Chawla SP, Block MS, Morris JC, Do K, Kim JW, et al. (2020). A Phase 1b Study Evaluating the Safety, Tolerability, and Immunogenicity of CMB305, a Lentiviral-Based Prime-Boost Vaccine Regimen, in Patients with Locally Advanced, Relapsed, or Metastatic Cancer Expressing NY-ESO-1. Oncoimmunology, 9:1847846.
- [73] Zhang H, Sun M, Wang J, Zeng B, Cao X, Han Y, et al. (2021). Identification of NY-ESO-1 Specific

Murine T Cell Receptors With Distinct Recognition Pattern for Tumor Immunotherapy. Front Immunol, 12:644520.

- [74] Duperret EK, Liu S, Paik M, Trautz A, Stoltz R, Liu X, et al. (2018). A Designer Cross-reactive DNA Immunotherapeutic Vaccine that Targets Multiple MAGE-A Family Members Simultaneously for Cancer Therapy. Clin Cancer Res, 24:6015-6027.
- [75] Sahin U, Oehm P, Derhovanessian E, Jabulowsky RA, Vormehr M, Gold M, et al. (2020). An RNA vaccine drives immunity in checkpoint-inhibitor-treated melanoma. Nature, 585:107-112.
- [76] Herrera LRM (2021). Reverse Vaccinology Approach in Constructing a Multi-Epitope Vaccine Against Cancer-Testis Antigens Expressed in Non-Small Cell Lung Cancer. Asian Pac J Cancer Prev, 22:1495-1506.
- [77] Ye Z, Liang Y, Ma Y, Lin B, Cao L, Wang B, et al. (2018). Targeted photodynamic therapy of cancer using a novel gallium (III) tris (ethoxycarbonyl) corrole conjugated-mAb directed against cancer/testis antigens 83. Cancer Med.
- [78] Gupta A, Nuber N, Esslinger C, Wittenbrink M, Treder M, Landshammer A, et al. (2013). A novel humanderived antibody against NY-ESO-1 improves the efficacy of chemotherapy. Cancer Immun, 13:3.
- [79] Raskin S, Van Pelt S, Toner K, Balakrishnan PB, Dave H, Bollard CM, et al. (2021). Novel TCR-like CAR-T cells targeting an HLA\*0201-restricted SSX2 epitope display strong activity against acute myeloid leukemia. Mol Ther Methods Clin Dev, 23:296-306.
- [80] Marcinkowski B, Stevanović S, Helman SR, Norberg SM, Serna C, Jin B, et al. (2019). Cancer targeting by TCR gene-engineered T cells directed against Kita-Kyushu Lung Cancer Antigen-1. J Immunother Cancer, 7:229.
- [81] Rijensky NM, Blondheim Shraga NR, Barnea E, Peled N, Rosenbaum E, Popovtzer A, et al. (2020). Identification of Tumor Antigens in the HLA Peptidome of Patient-derived Xenograft Tumors in Mouse. Mol Cell Proteomics, 19:1360-1374.
- [82] Suri A, Jagadish N, Saini S, Gupta N (2015). Targeting cancer testis antigens for biomarkers and immunotherapy in colorectal cancer: Current status and challenges. World J Gastrointest Oncol, 7:492-502.
- [83] Li Y, Li J, Wang Y, Zhang Y, Chu J, Sun C, et al. (2017). Roles of cancer/testis antigens (CTAs) in breast cancer. Cancer Lett, 399:64-73.
- [84] Chen M, Ni J, Chang H-C, Lin C-Y, Muyan M, Yeh S (2009). CCDC62/ERAP75 functions as a coactivator to enhance estrogen receptor beta-mediated transactivation and target gene expression in prostate cancer cells. Carcinogenesis, 30:841-850.
- [85] Jungbluth AA, Stockert E, Chen YT, Kolb D, Iversen K, Coplan K, et al. (2000). Monoclonal antibody MA454 reveals a heterogeneous expression pattern of MAGE-1 antigen in formalin-fixed paraffin embedded lung tumours. Br J Cancer, 83:493-497.
- [86] Woloszynska-Read A, Mhawech-Fauceglia P, Yu J, Odunsi K, Karpf AR (2008). Intertumor and

intratumor NY-ESO-1 expression heterogeneity is associated with promoter-specific and global DNA methylation status in ovarian cancer. Clin Cancer Res, 14:3283-3290.

- [87] Ehrlich M (2002). DNA methylation in cancer: too much, but also too little. Oncogene, 21:5400-5413.
- [88] Glazer CA, Smith IM, Ochs MF, Begum S, Westra W, Chang SS, et al. (2009). Integrative discovery of epigenetically derepressed cancer testis antigens in NSCLC. PLoS One, 4:e8189.
- [89] Smith IM, Glazer CA, Mithani SK, Ochs MF, Sun W, Bhan S, et al. (2009). Coordinated activation of candidate proto-oncogenes and cancer testes antigens via promoter demethylation in head and neck cancer and lung cancer. PLoS One, 4:e4961.
- [90] Weber J, Salgaller M, Samid D, Johnson B, Herlyn M, Lassam N, et al. (1994). Expression of the MAGE-1 tumor antigen is up-regulated by the demethylating agent 5-aza-2'-deoxycytidine. Cancer Res, 54:1766-1771.
- [91] Griffiths EA, Srivastava P, Matsuzaki J, Brumberger Z, Wang ES, Kocent J, et al. (2018). NY-ESO-1 Vaccination in Combination with Decitabine Induces Antigen-Specific T-lymphocyte Responses in Patients with Myelodysplastic Syndrome. Clin Cancer Res, 24:1019-1029.
- [92] Klar AS, Gopinadh J, Kleber S, Wadle A, Renner C (2015). Treatment with 5-Aza-2'-Deoxycytidine Induces Expression of NY-ESO-1 and Facilitates Cytotoxic T Lymphocyte-Mediated Tumor Cell Killing. PLoS One, 10:e0139221.
- [93] Rao M, Chinnasamy N, Hong JA, Zhang Y, Zhang M, Xi S, et al. (2011). Inhibition of histone lysine methylation enhances cancer-testis antigen expression in lung cancer cells: implications for adoptive immunotherapy of cancer. Cancer Res, 71:4192-4204.
- [94] Xiao W-H, Sanren G-W, Zhu J-H, Li Q-W, Kang H-R, Wang R-L, et al. (2010). Effect of 5-aza-2'deoxycytidine on immune-associated proteins in exosomes from hepatoma. World J Gastroenterol, 16:2371-2377.
- [95] Makani VKK, Mendonza JJ, Edathara PM, Yerramsetty S, Pal Bhadra M (2021). BORIS/CTCFL expression activates the TGFβ signaling cascade and induces Drp1 mediated mitochondrial fission in neuroblastoma. Free Radic Biol Med, 176:62-72.
- [96] Kosaka A, Yajima Y, Hatayama M, Ikuta K, Sasaki T, Hirai N, et al. (2021). A stealth antigen SPESP1, which is epigenetically silenced in tumors, is a suitable target for cancer immunotherapy. Cancer Sci, 112:2705-2713.
- [97] Nin DS, Wujanto C, Tan TZ, Lim D, Damen JMA, Wu K-Y, et al. (2021). GAGE mediates radio resistance in cervical cancers via the regulation of chromatin accessibility. Cell Rep, 36:109621.
- [98] Akers SN, Odunsi K, Karpf AR (2010). Regulation of cancer germline antigen gene expression: implications for cancer immunotherapy. Future Oncol, 6:717-732.
- [99] De Smet C, Courtois SJ, Faraoni I, Lurquin C, Szikora JP, De Backer O, et al. (1995). Involvement of two Ets

binding sites in the transcriptional activation of the MAGE1 gene. Immunogenetics, 42:282-290.

- [100] Maier H, Colbert J, Fitzsimmons D, Clark DR, Hagman J (2003). Activation of the early B-cellspecific mb-1 (Ig-alpha) gene by Pax-5 is dependent on an unmethylated Ets binding site. Mol Cell Biol, 23:1946-1960.
- [101] Serrano A, García A, Abril E, Garrido F, Ruiz-Cabello F (1996). Methylated CpG points identified within MAGE-1 promoter are involved in gene repression. Int J Cancer, 68:464-470.
- [102] Heinrich MC, Blanke CD, Druker BJ, Corless CL (2002). Inhibition of KIT tyrosine kinase activity: a novel molecular approach to the treatment of KITpositive malignancies. J Clin Oncol, 20:1692-1703.
- [103] Hoei-Hansen CE, Kraggerud SM, Abeler VM, Kaern J, Rajpert-De Meyts E, Lothe RA (2007). Ovarian dysgerminomas are characterised by frequent KIT mutations and abundant expression of pluripotency markers. Mol Cancer, 6:12.
- [104] Yang B, Wu J, Maddodi N, Ma Y, Setaluri V, Longley BJ (2007). Epigenetic control of MAGE gene expression by the KIT tyrosine kinase. J Invest Dermatol, 127:2123-2128.
- [105] Zhu X, Asa SL, Ezzat S (2008). Fibroblast growth factor 2 and estrogen control the balance of histone 3 modifications targeting MAGE-A3 in pituitary neoplasia. Clin Cancer Res, 14:1984-1996.
- [106] Monte M, Simonatto M, Peche LY, Bublik DR, Gobessi S, Pierotti MA, et al. (2006). MAGE-A tumor antigens target p53 transactivation function through histone deacetylase recruitment and confer resistance to chemotherapeutic agents. Proc Natl Acad Sci U S A, 103:11160-11165.
- [107] Yang B, O'Herrin SM, Wu J, Reagan-Shaw S, Ma Y, Bhat KMR, et al. (2007). MAGE-A, mMage-b, and MAGE-C proteins form complexes with KAP1 and suppress p53-dependent apoptosis in MAGE-positive cell lines. Cancer Res, 67:9954-9962.
- [108] Bai S, Grossman G, Yuan L, Lessey BA, French FS, Young SL, et al. (2008). Hormone control and expression of androgen receptor coregulator MAGE-11 in human endometrium during the window of receptivity to embryo implantation. Mol Hum Reprod, 14:107-116.
- [109] Kang Y, Hong JA, Chen GA, Nguyen DM, Schrump DS (2007). Dynamic transcriptional regulatory complexes including BORIS, CTCF and Sp1 modulate NY-ESO-1 expression in lung cancer cells. Oncogene, 26:4394-4403.
- [110] Renaud S, Pugacheva EM, Delgado MD, Braunschweig R, Abdullaev Z, Loukinov D, et al. (2007). Expression of the CTCF-paralogous cancertestis gene, brother of the regulator of imprinted sites (BORIS), is regulated by three alternative promoters modulated by CpG methylation and by CTCF and p53 transcription factors. Nucleic Acids Res, 35:7372-7388.
- [111] Klenova EM, Morse HC, Ohlsson R, Lobanenkov VV (2002). The novel BORIS + CTCF gene family is

uniquely involved in the epigenetics of normal biology and cancer. Semin Cancer Biol, 12:399-414.

- [112] Loukinov DI, Pugacheva E, Vatolin S, Pack SD, Moon H, Chernukhin I, et al. (2002). BORIS, a novel male germ-line-specific protein associated with epigenetic reprogramming events, shares the same 11-zinc-finger domain with CTCF, the insulator protein involved in reading imprinting marks in the soma. Proc Natl Acad Sci U S A, 99:6806-6811.
- [113] Vatolin S, Abdullaev Z, Pack SD, Flanagan PT, Custer M, Loukinov DI, et al. (2005). Conditional expression of the CTCF-paralogous transcriptional factor BORIS in normal cells results in demethylation and derepression of MAGE-A1 and reactivation of other cancer-testis genes. Cancer Res, 65:7751-7762.
- [114] Hong JA, Kang Y, Abdullaev Z, Flanagan PT, Pack SD, Fischette MR, et al. (2005). Reciprocal binding of CTCF and BORIS to the NY-ESO-1 promoter coincides with derepression of this cancer-testis gene in lung cancer cells. Cancer Res, 65:7763-7774.
- [115] Nguyen P, Bar-Sela G, Sun L, Bisht KS, Cui H, Kohn E, et al. (2008). BAT3 and SET1A form a complex with CTCFL/BORIS to modulate H3K4 histone dimethylation and gene expression. Mol Cell Biol, 28:6720-6729.
- [116] Somaiah N, Block MS, Kim JW, Shapiro GI, Do KT, Hwu P, et al. (2019). First-in-Class, First-in-Human Study Evaluating LV305, a Dendritic-Cell Tropic Lentiviral Vector, in Sarcoma and Other Solid Tumors Expressing NY-ESO-1. Clin Cancer Res, 25:5808-5817.
- [117] Gérard C, Baudson N, Ory T, Segal L, Louahed J (2015). A Comprehensive Preclinical Model Evaluating the Recombinant PRAME Antigen Combined With the AS15 Immunostimulant to Fight Against PRAME-expressing Tumors. J Immunother, 38:311-320.
- [118] Chang AY, Dao T, Gejman RS, Jarvis CA, Scott A, Dubrovsky L, et al. (2017). A therapeutic T cell receptor mimic antibody target tumor-associated PRAME peptide/HLA-I antigens. J Clin Invest, 127:2705-2718.
- [119] Sivaccumar JP, Leonardi A, Iaccarino E, Corvino G, Sanguigno L, Chambery A, et al. (2021). Development of a New Highly Selective Monoclonal Antibody against Preferentially Expressed Antigen in Melanoma (PRAME) and Identification of the Target Epitope by Bio-Layer Interferometry. Int J Mol Sci, 22.
- [120] Coscia F, Lengyel E, Duraiswamy J, Ashcroft B, Bassani-Sternberg M, Wierer M, et al. (2018). Multilevel Proteomics Identifies CT45 as a Chemosensitivity Mediator and Immunotherapy Target in Ovarian Cancer. Cell, 175.
- [121] Martinez M, Kim S, St Jean N, O'Brien S, Lian L, Sun J, et al. (2021). Addition of anti-TIM3 or anti-TIGIT Antibodies to anti-PD1 Blockade Augments Human T cell Adoptive Cell Transfer. Oncoimmunology, 10:1873607.
- [122] Zhang M, Hong JA, Kunst TF, Bond CD, Kenney CM, Warga CL, et al. (2021). Randomized phase II trial of

a first-in-human cancer cell lysate vaccine in patients with thoracic malignancies. Transl Lung Cancer Res, 10:3079-3092.

[123] Xue W, Metheringham RL, Brentville VA, Gunn B, Symonds P, Yagita H, et al. (2016). SCIB2, an antibody DNA vaccine encoding NY-ESO-1 epitopes, induces potent antitumor immunity which is further enhanced by checkpoint blockade. Oncoimmunology, 5:e1169353.

[124] Ohue Y, Kurose K, Karasaki T, Isobe M, Yamaoka T, Futami J, et al. (2019). Serum Antibody Against NY-ESO-1 and XAGE1 Antigens Potentially Predicts Clinical Responses to Anti-Programmed Cell Death-1 Therapy in NSCLC. J Thorac Oncol, 14:2071-2083.