

RESEARCH ARTICLE

Joint Mapping and Allele Mining of the Rolled Leaf Trait in Rice (*Oryza sativa* L.)

Qiang Zhang^{1,2}, Tianqing Zheng², Long Hoang^{2,3,4}, Chunchao Wang², Nafisah^{2,3}, Charles Joseph^{2,3}, Wenzhong Zhang^{1*}, Jianlong Xu^{2,3,4*}, Zhikang Li^{2,4}

1 Shenyang Agricultural University, 120 Dongling Road, Shenyang 110866, China, **2** Institute of Crop Sciences/National Key Facility for Crop Gene Resources and Genetic Improvement, Chinese Academy of Agricultural Sciences, 12 South Zhong-Guan-Cun Street, Beijing 100081, China, **3** Agricultural Genomics Institute at Shenzhen, Chinese Academy of Agricultural Sciences, Shenzhen 518120, China, **4** Shenzhen Institute of Breeding & Innovation, Chinese Academy of Agricultural Sciences, Shenzhen 518120, China

☯ These authors contributed equally to this work.

✉ Current address: Institute of Agricultural Sciences for Southern Vietnam, 121 Nguyen Binh Khiem, District 1, Ho Chi Minh City, Vietnam

✉ Current address: Indonesian Center for Rice Research, Jalan Raya 9, Sukamandi, Subang, West Java, Indonesia, 41256

✉ Current address: Department of Botany, University of Dar es Salaam, Box 35091, Dar es Salaam, Tanzania

* xujlcaas@126.com (JLX); zwzhong@126.com (WZZ)



OPEN ACCESS

Citation: Zhang Q, Zheng T, Hoang L, Wang C, Nafisah, Joseph C, et al. (2016) Joint Mapping and Allele Mining of the Rolled Leaf Trait in Rice (*Oryza sativa* L.). PLoS ONE 11(7): e0158246. doi:10.1371/journal.pone.0158246

Editor: Istvan Rajcan, University of Guelph, CANADA

Received: March 1, 2016

Accepted: June 12, 2016

Published: July 21, 2016

Copyright: © 2016 Zhang et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its Supporting Information file.

Funding: A subprogram of the Key Special Project on Molecular Design Breeding for Rice to TQZ (2016YFD0101800) from the Chinese Ministry of Science & Technology, the "863" Key Project to JLX (2014AA10A601) from the Chinese Ministry of Science & Technology, and the "Green Super Rice" project to ZKL (OPPGD1393) from the Bill and Melinda Gates Foundation. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Abstract

The rolled leaf trait, long considered to be a key component of plant architecture, represents an important target trait for improving plant architecture at the population level. We therefore performed linkage mapping using a set of 262 highly variable RILs from two rice cultivars (Minghui 63 and 02428) with minor differences in leaf rolling index (LRI) in conjunction with GWAS mapping of a random subset of the 1127 germplasms from the 3K Rice Genomes Project (3K Rice). A total of seven main-effect loci were found to underlie the transgressive segregation of progenies from parents with minor differences in LRI. Five of these loci were previously identified and two (*qRI7b* and *qRI9b*) are newly reported with additional evidence from GWAS mapping for *qRI7b*. A total of 18 QTLs were identified by GWAS, including four newly identified QTLs. Six QTLs were confirmed by linkage mapping with the above RIL population, and 83.3% were found to be consistent with previously reported loci based on comparative mapping. We also performed allele mining with representative SNPs and identified the elite germplasms for the improvement of rolled leaf trait. Most favorable alleles at the detected loci were contributed by various 3K Rice germplasms. By a re-scanning of the candidate region with more saturated SNP markers, we dissected the region harboring *gRI4-2* into three subregions, in which the average effect on LRI was 3.5% with a range from 2.4 to 4.1% in the third subregion, suggesting the presence of a new locus or loci within this region. The representative SNPs for favorable alleles in the reliable QTLs which were consistently identified in both bi-parental mapping and GWAS, such as *qRI4*, *qRI5*, *qRI6*, *qRI7a*, and *qRI7b* will be useful for future molecular breeding programs for ideal plant type in rice.

Competing Interests: The authors have declared that no competing interests exist.

Introduction

The rolled (V-shaped or curled) leaf trait has long been considered by experienced breeders to be a key trait for ideal plant type not only for *indica* hybrid rice breeding [1] but also for inbred *japonica* cultivar development in Northern China [2]. Extremely rolled leaves often lead to reduced rates of photosynthesis and apoplastic transport ability [3] and even reduced light use efficiency [4]. At the individual level, rolled leaves are not always directly associated with yield component traits in certain crosses, such as MH63 × 02428 [5], let alone the unfavorable traits including dwarf and/or narrow leaves and/or smaller panicles often occur in conjunction with rolled leaves in most artificial mutants, except for a few natural mutants such as *rl(t)* [6]. Nonetheless, moderately rolled leaves can improve photosynthetic efficiency in certain cultivars [7,8] and thus contribute to economic and grain quality traits [6,9]. In addition, the rolled leaf trait is thought to contribute to lodging resistance and ventilation, which are strongly associated with disease resistance, especially to fungal diseases, at the population level [10]. Moreover, cultivars with moderately rolled leaves are suitable for cultivation at relatively high density [11].

To date, no fewer than 70 genes/QTLs for the rolled leaf trait have been mapped or cloned throughout the genome. Most studies of the rolled leaf trait have involved the use of parents with significant phenotypic differences. Unlike the extremely rolled leaf phenotype, moderately rolled leaves or leaves with various degrees of rolling, especially inside-rolled (adaxial rolled) leaves, would be a useful trait to target in breeding. Uncovering hidden diversity in the progenies of parents with minor phenotypic differences in target traits is also important for dissecting complex traits including resistance to various biotic and abiotic stresses [12, 13]. Whether the mechanisms underlying hidden diversity also function for relatively simple traits such as rolled leaves remains unclear.

Currently, mining favorable alleles is an important component of plant breeding [14]. However, traditional QTL detection by linkage mapping is usually performed using populations with limited parental variation [15]. Performing GWAS offers opportunities to overcome this shortcoming. If GWAS is performed jointly with linkage mapping, the relatively high false positive rate of GWAS is largely constrained, and the efficiency of QTL mapping is much improved, as demonstrated in maize [16]. This technique has been successfully employed in rice to help dissect complex biotic stress traits, such as rice black-streaked dwarf disease resistance, as well as relatively simple genetic traits including rice leaf stripe disease resistance [17]. The simultaneous exploration of natural variations would be highly useful for rice breeding.

Here, we utilized a traditional recombinant inbred line (RIL) population derived from two parents with minor differences in leaf rolling index (LRI) for linkage mapping, along with a germplasm panel from the 3K Rice [18], for joint mapping of loci affecting the rolled leaf trait and for mining favorable alleles. The results of this study will greatly facilitate molecular breeding of rice cultivars with ideal plant type in the future.

Materials and Methods

Plant materials

Minghui 63 (MH63), the male parent of the widely cultivated hybrid *indica* rice variety Shanyou63, which is distributed over a wide area spanning more than 21 longitudes and 20 latitudes in China [19], was crossed with 02428. This typical *japonica* line, with a neutral allele at the major locus *S*⁵, controlling hybrid sterility in most inter-subspecies crosses, as well as tolerance to low CO₂ stress, was isolated from mutant progenies from a cross between two landraces, Pang-Xie-Gu and Ji-Bang-Dao [20]. The F₁ hybrids of MH63 × 02428 were then

consecutively selfed until the F₈ generation to produce a set of 262 recombinant inbred lines (RILs) [21].

A germplasm panel of 1,129 accessions (S1 Table) randomly chosen from the 3K Rice Genomes Project [18] was adopted in this study to mine favorable alleles and to confirm the results of QTL mapping.

Planting and phenotyping

All of the above plant materials were transplanted in the field at a spacing of 13.2 cm between individuals and 25 cm between rows, with a final planting density of approximately 18,000 individuals per 667 m², at the Experimental Station of the Institute of Crop Sciences, Chinese Academy of Agricultural Sciences (ICS, CAAS) at Beijing (40.2°N, 116.2°E) and Sanya (18.3°N, 109.3°E) of Hainan province, as well as the Experimental Station of the Institute of Agricultural Genomics, Chinese Academy of Agricultural Sciences at Shenzhen (22.6°N, E114.5°E) of Guangdong province. The RILs, the two parents, and the germplasm panel were planted in a random complete block design with two replications.

Phenotyping of rolled leaf traits was performed as previously described [22]. The top two leaves of three main tillers per individual plant were measured for leaf width (LW) and distance between leaf boarders (LN) at the widest part of each leaf. At least five individuals per line were measured for the RILs, the two parents, and the germplasms panel. The LRI was calculated using the following formula: $LRI (\%) = (LW - LN) / LW \times 100$.

Genotyping and mapping

Genomic DNA from MH63, 02428, and the RILs in the F₈ generation was isolated using a DNeasy mini Kit (Qiagen), and the genotypes of the RILs were determined based on SNPs generated from whole genome sequencing with an Illumina Genome Analyzer IIX as described previously [23].

Minghui 63 (MH63) and 02428 were submitted to whole genome re-sequencing, and a total of 5,336,108,154 and 5,562,905,674 bp sequences were obtained, respectively. Alignment analysis was carried out using the MSU6.1 assembly of the Nipponbare sequence as the reference genome. A total of 5,062,106,567 bp and 5,278,080,725 bp of consistent sequences were obtained for MH63 and 02428, covering 96.57% and 94.03% of the whole genome, respectively. Single nucleotide polymorphisms (SNPs) were then identified based on these two consistent sequences to obtain an SNP dataset. A total of 48,498, 42,124, and 36,410 SNP loci were found between MH63 and 02428 with supporting evidence from more than three, four, and five reads, respectively. Since a new version of Nipponbare assembly has been available after the accomplishment of this step, we later re-mapped all the reads to the Os-Nipponbare-Reference-IRGSP-1.0 [24]. All the following works were carried out based on this new version of reference genome.

A total of 384 SNPs that are evenly distributed along the genome were used to design an Illumina SNP chip [25] for genotyping of all 262 RILs using their parents and the F₁ populations as controls to build up a frame map. The frame map was constructed with IciMapping, version 3.3 [26]. Further mapping was carried via RAD sequencing [27] of each RIL as well as the two parents. Ultimately, a total of 58,936 qualified SNP consisting of 4,568 chromosome bins were identified and integrated into the frame map, with an average distance of 77 kb between adjacent markers.

The germplasm panel was re-sequenced with an average depth of more than 10X [18]. The reads were mapped to the reference genome of Nipponbare, and 14M high-quality SNPs were identified [18]. Based on these 14M markers, 2.9M SNPs related to potential protein-coding

areas were carefully selected for further development of the 50k microarray chips. In order to build an SNP set for primary association studies in which the locations of the SNPs were independent of the SNPs chosen for microarray chip design, 27,921 SNPs were selected from the 2.9M SNPs by choosing one SNP per 100 counts.

Data analysis

The ICIM mapping module from the V.3.3 package of QTL IciMapping [26] was used to detect the main-effect QTLs underlying the rolled leaf trait in the RILs. The default setting of LOD 2.5 was adopted as the threshold for identifying a putative locus.

Comparative mapping was carried out against a reference sequence map, GRAMENE annotation sequence map 2009 [28], to compare the QTLs detected in this study with previously reported QTLs or genes known to be associated with the rolled leaf trait in rice.

The basic scenario of compressed mixed linear model [29] implemented in the Genomic Association and Prediction Integrated Tool (GAPIT) Version 2 [30] was adopted for association analysis between QTL-flanking markers and LRI for the germplasm panel. To minimize the possible effects of population structure, the parameter of Model.selection in GAPIT was set as TRUE. Under this condition, a forward model selection by the Bayesian information criterion (BIC) was conducted to determine the optimal parameters of principal components for the LRI data. A relatively stringent threshold was adopted to identify significant correlation between the SNP and LRI with a $-\text{LOG}_{10}(\text{P})$ value of 5.0. To minimize to the possibility of type II errors in QTL detection [31], a relatively low threshold of $-\text{LOG}_{10}(\text{P}) = 2.5$ was also adopted with supporting evidence from linkage mapping or comparative mapping.

The allelic effects were estimated by setting the Major.allele.zero = TRUE in GAPIT Version 2 to identify the donors of favorable alleles and their effects on LRI.

Results

Distribution of LRI in the MH63 × 02428 RIL population and the germplasm panel

As shown in Fig 1A, the RIL population exhibited a similar pattern of distribution of LRI traits throughout the three environments i.e., Beijing (BJ), Shenzhen (SZ), and Hainan (HN). LRI appeared to be relatively stable in all three environments. On the other hand, even though the LRI trait did not significantly differ between the two parents (both were less than 10%), highly transgressive variations were still available in the progenies (ranging from 0–90%). However, in the germplasm panel, the variation was slightly smaller, with the LRI ranging from 0–70%, as shown in Fig 1B.

Linkage mapping of main-effect QTLs controlling the rolled leaf trait

A total of seven main-effect QTLs (*qRI4*, *qRI5*, *qRI6*, *qRI7a*, *qRI7b*, *qRI9a*, and *qRI9b*) affecting LRI were detected on chromosomes 4, 5, 6, 7, and 9 by linkage mapping in the MH63 × 02428 RIL population across the three environments (Table 1, Fig 2). Among these, four QTLs (*qRI4*, *qRI5*, *qRI6*, and *qRI9b*) were stably expressed across all three environments, *qRI7a* and *qRI7b* were significant in only two environments (HN & SZ or SZ & BJ), and *qRI9a* was specifically expressed at Sanya of Hainan. Although the locus effects varied in different environments, the direction of gene effects on the LRI remained consistent. Among these loci, the alleles at *qRI4*, *qRI5*, *qRI7b*, and *qRI9a* from the *japonica* parent, 02428, increased the LRI, while the 02428 alleles at the three other loci (*qRI6*, *qRI7a*, and *qRI9b*) reduced the LRI in all three environments.

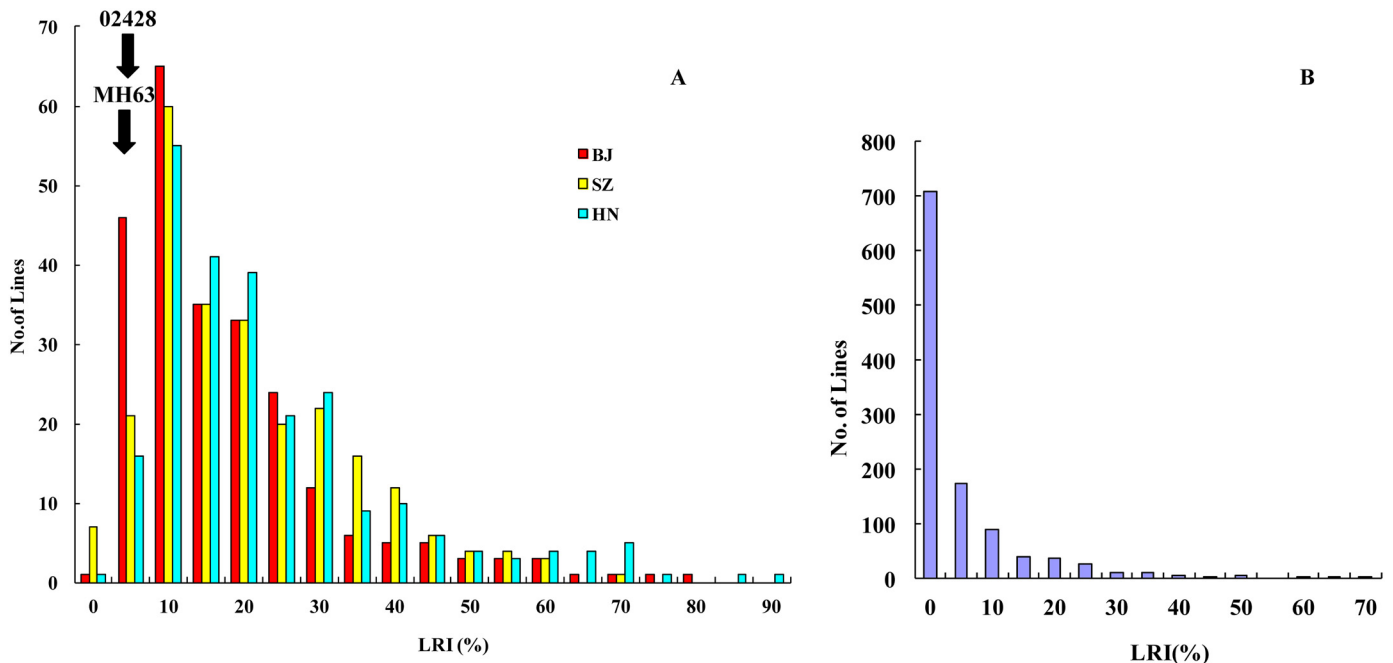


Fig 1. Distribution of leaf rolling index (LRI, %) in the MH63 × 02428 RIL population in three environments (A, BJ = Beijing, SZ = Shenzhen, and HN = Hainan) and a random subset of the 1127 germplasm panel at SZ (B). The average values throughout three environments of the two parents (02428 and MH63) for the RILs are indicated by two black arrows.

doi:10.1371/journal.pone.0158246.g001

These reverse effects of alleles from two parents at different loci may ultimately be responsible for the nearly flat leaves of MH63 and 02428.

The known loci/genes for the rolled leaf trait are as follows (some genes for abaxial rolling are underlined): *qRL-1*[32], *url1(t)*[33], *rl-4(rl-2)*[34], *SCL1*[35], *nal10*[36], *REL1*[37], *rl8(t)*[38], *qRL-2-1b*[32], *CFL1*[39], *DNAL1*[40], *rl(t)*[41], *OsAGO1a(LOC_Os02g45070)*[42], *Roc5*[43], *Nrl3(t)*[44], *s1-145*[45], *nal7(Os03g0162000)*[46], *NRL2(t)*[47], *RL3(t)*[44], *OsAGO7*[48], *rl-3(rl-5)*[34], *ACL1*[49], *SRL2*[50], *rl11(t)*[51], *nl(t)*[52], *qRL4-2*[22], *QFL4* and *QFL5*[53], *qRL5-9* and *qRL5-10*[22], *rl8*[54], *RL13*[55], *qRL-6*[32], *sd-sl*[56], *rl11(t)*[51], *SRL1*[57], *YABBY1*[8], *rfs*[34], *qRL-7*[32], *Nul1*[58], *QFL7*[53], *sl2*[59], *qRL-8-1*, *qRL-8-2* and *qRL-9*[32], *OsMYB103L*

Table 1. QTLs controlling leaf-rolling index (LRI) detected by linkage mapping in an RIL population at three planting sites (BJ = Beijing, HN = Hainan, and SZ = Shenzhen).

QTL	Chr	Pos (cM)	Flanking marker	HN			SZ			BJ			Reference ³⁾
				LOD	A (%) ¹⁾	PVE (%) ²⁾	LOD	A (%)	PVE (%)	LOD	A (%)	PVE (%)	
<i>qRI4</i>	4	103	M110-Bin_1720	3.8	3.6	4.7	4.9	2.4	4.8	4.1	3.5	6.2	<i>rl11(t)</i> [51], <i>SRL2</i> [50]
<i>qRI5</i>	5	98	M129-Bin_1976	9.2	6	12.9	9.5	3.6	10.4	3.9	3.7	6.6	<i>qRL5-10</i> [22], <i>rl8(t)</i> [38]
<i>qRI6</i>	6	78	M153-Bin_2361	4.1	-3.5	6.4	5	-2.4	4.6	3.2	-3.3	3.8	<i>qRL-6</i> [32]
<i>qRI7a</i>	7	46	Bin_2619-M163	5	-4.3	6.7	3.9	-2.2	4.1				<i>qRL-7</i> [32]
<i>qRI7b</i>	7	112	M177-Bin_2847				3	3	4.5	3.7	3.6	4.5	
<i>qRI9a</i>	9	28	Bin_3289-M198	3.7	3.6	4.5							<i>rl13(t)</i> [55]
<i>qRI9b</i>	9	79	M205-Bin_3476	6.3	-4.7	8.4	3.3	-2	3.3	3.6	-3.6	6.2	

¹⁾ The additive effect results from the effect of substitution of MH63 alleles with 02428 alleles.

²⁾ Phenotypic variance explained.

³⁾ Numbers in brackets are reference numbers, as listed in the reference section.

doi:10.1371/journal.pone.0158246.t001

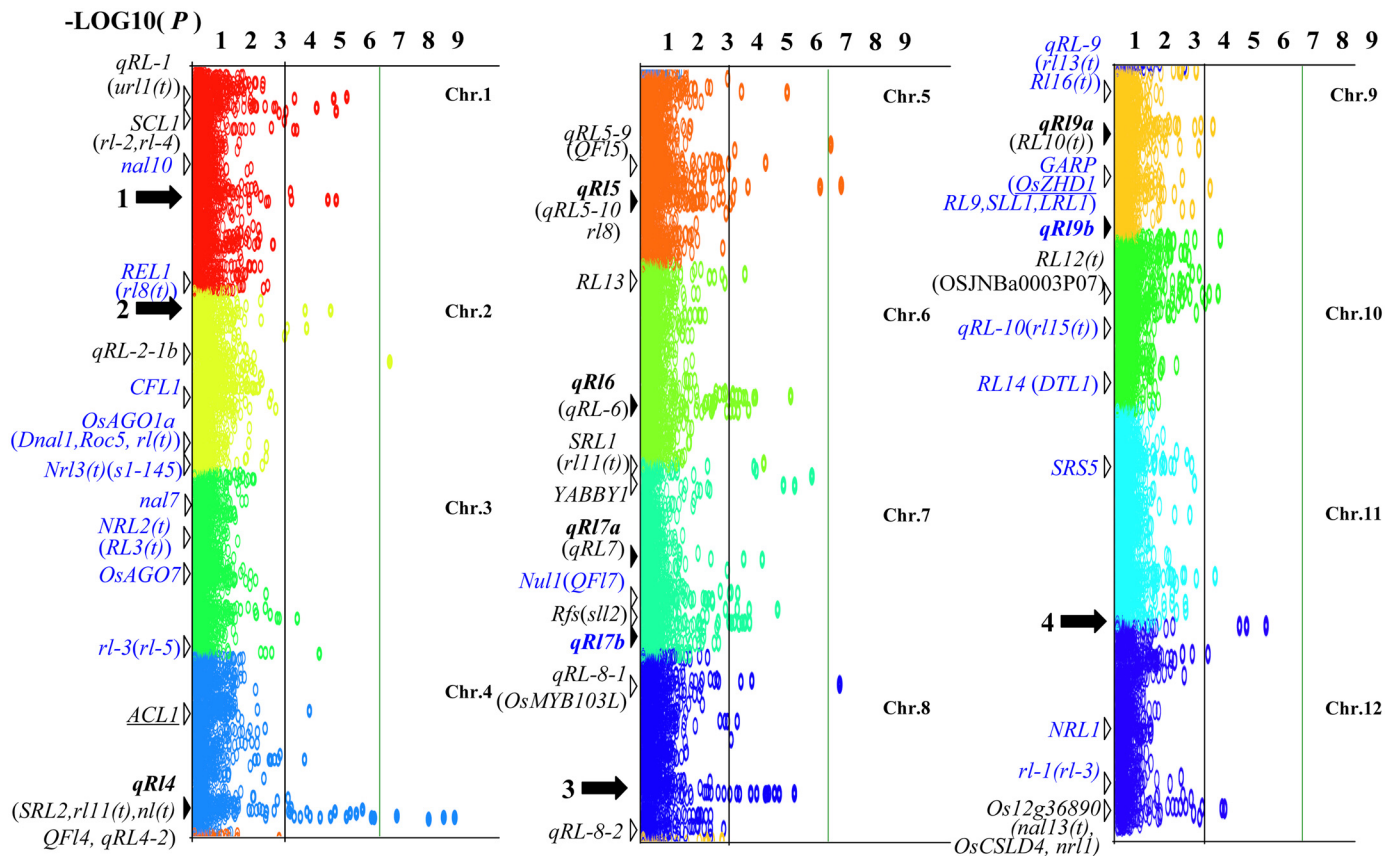


Fig 2. Distribution and comparison of QTLs for leaf-rolling index (LRI, %) identified by linkage mapping and GWAS (Manhattan plot) with those detected in previous studies. The reported loci/genes are indicated by white triangles and/or plain italic font, and QTLs detected by linkage mapping in our RIL population of MR63/02428 are indicated by black triangles and bold italic font. Several loci/genes with no significant support in the germplasm panel are shown in blue font. Five positions with possible new loci detected only by GWAS in the germplasm panel are indicated by black arrows and are numbered 1–5.

doi:10.1371/journal.pone.0158246.g002

[60], *rl13(t)*[55], *RL16(t)*[61], *RL10(t)*[62], *SLL1*[63], *RL9(rl9(t),GARP)*[64], *OSZHD1* [65], *LRL1* [66], *QF19*[53], *OSJNBa0003P07*[67], *RL12(t)*[68], *rl15(t)*[69], *qRL-10*[32], *DTL1*[70], *RL14*[71], *SRS5*[72], *NRL1(OSJNBa0027H05)*[73], *rl-1(rl-3)*[34], *nal3(t)*[74], and *nrl1*[46].

GWAS mapping of QTLs affecting the rolled leaf trait

A total of 18 significant loci were detected by GWAS using a combination of the relatively stringent threshold of $-\text{LOG}_{10}(P) = 5.0$ and the relatively low threshold of $-\text{LOG}_{10}(P) = 2.5$, with supporting evidence from either our linkage mapping with RILs or previous reports (Table 2, Fig 2). These QTLs are distributed throughout the genome, except for chromosome 3, 10, and 11. Of these QTLs, 14 (77.8%) are closely related to loci that were previously identified by comparative mapping or to QTLs identified by our linkage mapping of the MH63 × 02428 RIL population, whereas the other four (*gRL1-2*, *gRL5-1*, *gRL5-2*, and *gRL12-1*) are newly identified QTLs that are associated with LRI.

Allele mining for the rolled leaf trait

We mined favorable alleles and estimated their effects on LRI using a random subset of the 1127 3K panel. We ultimately detected a total of 33 favorable alleles for the 14 loci, which were

Table 2. QTLs affecting leaf-rolling index (LRI) detected by GWAS in a panel of 1,129 germplasms.

QTL	Range (bp)	-LOG ₁₀ (P)	QTL from linkage mapping	QTL from references
<i>gRI1-1</i>	4,832,637–4,832,637	5.3		<i>qRL-1</i> [32], <i>ur1(t)</i> [33], <i>rl-4(rl-2)</i> [34], <i>SCL1</i> [35]
<i>gRI1-2</i>	25,404,548–25,404,548	5.7		
<i>gRI2-1</i>	7,519,002–9,022,802	3.9		<i>qRL-2-1b</i> [40]
<i>gRI2-2</i>	14,150,759–20,850,818	6.8		<i>CFL1</i> [41]
<i>gRI4-1</i>	19,823,372–20,860,914	3.8		<i>ACL1</i> [49]
<i>gRI4-2</i>	31,977,228–32,592,463	9.1	<i>qRI4</i>	<i>rl11(t)</i> [51], <i>SRL2</i> [50], <i>qRL4-2</i> [22], <i>nl(t)</i> [52]
<i>gRI5-1</i>	3,117,099–3,117,099	5		
<i>gRI5-2</i>	11,228,990–11,228,990	6.6		
<i>gRI5-3</i>	17,598,480–19,569,643	6.9		<i>qRL5-9</i> [22]
<i>gRI5-4</i>	20,444,240–21,926,430	3.1	<i>qRI5</i>	<i>qRL5-10</i> [22], <i>rl8(t)</i> [38]
<i>gRI6-1</i>	20,532,308–22,944,285	5.1	<i>qRI6</i>	<i>qRL-6</i> [32]
<i>gRI7-1</i>	1,762,263–3,230,532	5.9		<i>YABBY1</i> [8]
<i>gRI7-2</i>	14,684,286–16,298,897	4.2	<i>qRI7a</i>	<i>qRL-7</i> [32]
<i>gRI7-3</i>	27,671,916–27,671,916	2.6	<i>qRI7b</i>	
<i>gRI8-1</i>	4,426,494–4,426,494	6.9		<i>qRL-8-1</i> [32]
<i>gRI8-2</i>	26,810,755–27,101,483	5.3		<i>qRL-8-2</i> [32]
<i>gRI9-1</i>	7,959,893–10,895,719	3.3	<i>qRI9a</i>	<i>rl13(t)</i> [55]
<i>gRI12-1</i>	363,480–363,480	5.2		

doi:10.1371/journal.pone.0158246.t002

consistently detected in GWAS and linkage mapping or comparative mapping (Table 3). Among these, 16 (48.5%) alleles were found in the five regions (*gRI4-2*, *gRI5-4*, *gRI6-1*, *gRI7-2*, and *gRI7-3*) associated with loci identified by linkage mapping of the RIL population, and 24 (72.7%) of the 33 alleles were donated by the favorable germplasms from the 3K Rice Genomes Project panel (Table 3). The average effect of the favorable alleles on LRI was 1.9%, ranging from 1.1% to 3.1%. Approximately 30.3% of the favorable alleles can improve the LRI by no less than 2%, with the maximum effects from the A alleles at the representative SNP at the position of 14,150,759 in the region of *gRI2-2*.

Subregional analysis of *gRI4-2*

The region of *gRI4-2* possessed the highest peak among all the rolled leaf loci detected by GWAS in this study. The *gRI4-2* was also consistently detected in the RILs derived from MH63 × 02428 across three environments. Whether this clustering was caused by tightly linked loci or a single locus remains unclear. To get more details in the candidate region, we extracted 89,349 SNP markers surrounding the above peaks with an average distance of 18.1 ± 55.2 bp between adjacent markers to perform a fine re-scanning of this region and to re-estimate all of the -LOG₁₀(P) values and additive effects.

It's notable that within this region of no more than 2 Mb (30,977,335–32,592,463 bp) at the end of the long arm of chromosome 4, at least three clusters of peaks were found (Fig 3). The first subregion (*gRI4-2_1*) covers nucleotides in a range from 30,980,707 to 30,994,770 bp (marked by an SNP peak of -LOG₁₀(P) = 5.2), which is consistent with our linkage mapping-derived locus *qLR4*, with an average value of 2.9% (ranging from 1.3–7.6%) favorable effects on LRI. The second subregion (*gRI4-2_2*) is marked by an SNP peak of -LOG₁₀(P) = 5.2 and comprises nucleotides in the range of 31,068,550–32,086,234 bp. The favorable allele effects averaged 1.6% (ranging from 1.1–2.2%) for LRI. The region *gRI4-2_2* harbors two previously reported genes, *nl(t)*[52] and *SRL2*[50]. The third subregion (*gRI4-2_3*) is located at the physical range of 32,156,194–32,452,869 bp, with an SNP peak of -LOG₁₀(P) = 13.7. The favorable

Table 3. Representative SNPs for favorable alleles and their effects on leaf rolling index (LRI, %) simultaneously detected by GWAS and linkage mapping in this study or previous studies.

QTL	QTL from linkage mapping	Physical position (bp)	Favorable SNP allele	Effect (%)	Top five accessions with favorable alleles ¹⁾
<i>gRI1-1</i>		4,832,637	T	2.1	IRIS_313-9759 (65.2), IRIS_313-8023 (60.3), IRIS_313-8027 (57.3), IRIS_313-8149 (55.4), IRIS_313-8129 (49.9)
<i>gRI2-1</i>		7,519,002	G	1.7	IRIS_313-9759 (65.2), IRIS_313-8023 (60.3), IRIS_313-8027 (57.3), IRIS_313-8075 (55.6), IRIS_313-8149 (55.4)
		9,022,802	T	1.6	IRIS_313-9759 (65.2), IRIS_313-8023 (60.3), IRIS_313-8027 (57.3), IRIS_313-8075 (55.6), IRIS_313-8149 (55.4)
<i>gRI2-2</i>		14,150,759	A	3.1	IRIS_313-9759 (65.2), IRIS_313-8023 (60.3), IRIS_313-8027 (57.3), IRIS_313-8075 (55.6), IRIS_313-8149 (55.4)
		20,850,818	A	1.5	Nipponbare
<i>gRI4-1</i>		19,823,372	T	1.6	IRIS_313-9759 (65.2), IRIS_313-8023 (60.3), IRIS_313-8027 (57.3), IRIS_313-8075 (55.6), IRIS_313-8149 (55.4)
		20,743,867	A	1.9	IRIS_313-9759 (65.2), IRIS_313-8023 (60.3), IRIS_313-8027 (57.3), IRIS_313-8149 (55.4), IRIS_313-8129 (49.9)
		20,786,774	T	1.4	IRIS_313-9759 (65.2), IRIS_313-8023 (60.3), IRIS_313-8027 (57.3), IRIS_313-8149 (55.4), IRIS_313-8129 (49.9)
		20,821,210	C	1.3	IRIS_313-9759 (65.2), IRIS_313-8023 (60.3), IRIS_313-8027 (57.3), IRIS_313-8149 (55.4), IRIS_313-8129 (49.9)
		20,860,914	A	1.3	IRIS_313-9759 (65.2), IRIS_313-8023 (60.3), IRIS_313-8027 (57.3), IRIS_313-8149 (55.4), IRIS_313-8129 (49.9)
<i>gRI4-2</i>	<i>qRI4</i>	31,977,228	A	2.4	Nipponbare
		32,086,234	A	1.1	IRIS_313-8135(47.0), B153 (28.1), CX15 (46.5), CX28 (35.0), CX314 (42.0)
		32,134,331	A	1.8	IRIS_313-8023 (60.3), IRIS_313-8027 (57.3), IRIS_313-8075 (55.6), IRIS_313-8149 (55.4), IRIS_313-8111 (48.7)
		32,227,059	G	2.4	IRIS_313-9759 (65.2), IRIS_313-8023 (60.3), IRIS_313-8027 (57.3), IRIS_313-8075 (55.6), IRIS_313-8149 (55.4)
		32,236,238	A	1.7	IRIS_313-9759 (65.2), IRIS_313-8023 (60.3), IRIS_313-8027 (57.3), IRIS_313-8075 (55.6), IRIS_313-8149 (55.4)
		32,271,965	C	2.7	IRIS_313-9759 (65.2), IRIS_313-8023 (60.3), IRIS_313-8027 (57.3), IRIS_313-8075 (55.6), IRIS_313-8149 (55.4)
		32,592,463	C	2.5	IRIS_313-9759 (65.2), IRIS_313-8023 (60.3), IRIS_313-8027 (57.3), IRIS_313-8075 (55.6), IRIS_313-8149 (55.4)
<i>gRI5-3</i>		17,598,480	T	2.4	IRIS_313-9759 (65.2), IRIS_313-8023 (60.3), IRIS_313-8027 (57.3), IRIS_313-8075 (55.6), IRIS_313-8149 (55.4)
<i>gRI5-4</i>		19,471,779	T	2.1	Nipponbare
	<i>qRI5</i>	19,569,643	C	2.1	Nipponbare
<i>gRI6-1</i>	<i>qRI6</i>	20,532,308	G	2.2	IRIS_313-8023 (60.3), IRIS_313-8027 (57.3), IRIS_313-8149 (55.4), IRIS_313-8129 (49.9), IRIS_313-8185 (49.8)
		22,661,800	G	1.8	IRIS_313-9759 (65.2), IRIS_313-8023 (60.3), IRIS_313-8075 (55.6), IRIS_313-8149 (55.4), IRIS_313-8129 (49.9)
		22,854,876	T	1.4	IRIS_313-9759 (65.2), IRIS_313-8149 (55.4), IRIS_313-8129 (49.9), IRIS_313-8075 (55.6), IRIS_313-8023 (60.3)
		22,944,285	T	2.6	IRIS_313-10084 (36.9), CX15 (46.5), CX288 (26.5), CX314 (42.0), CX361 (14.6)
<i>gRI7-1</i>		1,762,263	C	2.3	IRIS_313-9759 (65.2), IRIS_313-8023 (60.3), IRIS_313-8027 (57.3), IRIS_313-8075 (55.6), IRIS_313-8149 (55.4)
		3,230,532	T	1.9	Nipponbare
<i>gRI7-2</i>	<i>qRI7a</i>	14,684,286	C	1.7	Nipponbare

(Continued)

Table 3. (Continued)

QTL	QTL from linkage mapping	Physical position (bp)	Favorable SNP allele	Effect (%)	Top five accessions with favorable alleles ¹⁾
		16,298,897	A	1.4	IRIS_313–8023 (60.3), IRIS_313–8075 (55.6), IRIS_313–8185 (49.8), IRIS_313–8111 (48.7), B055 (48.0)
<i>gRI7-3</i>	<i>qRI7b</i>	27,671,916	T	1.4	Nipponbare
<i>gRI8-1</i>		4,426,494	C	2.9	IRIS_313–9759 (65.2), IRIS_313–8075 (55.6), IRIS_313–8149 (55.4), IRIS_313–8185 (49.8), IRIS_313–10083 (37.2)
<i>gRI8-2</i>		21,374,656	G	2.3	Nipponbare
<i>gRI9-1</i>		7,959,893	C	1.9	Nipponbare
		10,895,719	G	1.3	IRIS_313–9759 (65.2), IRIS_313–8075 (55.6), IRIS_313–8149 (55.4), IRIS_313–8129 (49.9), IRIS_313–8111 (48.7)

¹⁾ The accession ID can be found in the database (<http://www.rmbreeding.cn/snp3k>) for further details. Numbers in brackets are LRI values (%).

doi:10.1371/journal.pone.0158246.t003

allele effects averaged 3.5%, ranging from 2.4–4.1% for LRI, suggesting that a new locus or loci could be localized within this region.

Discussion

QTLs underlying segregation of the rolled leaf trait in the RIL population

In this study, we carried out both linkage mapping and GWAS analysis in order to perform accurate locus searching and to mine multiple favorable alleles for the rolled leaf trait, one of

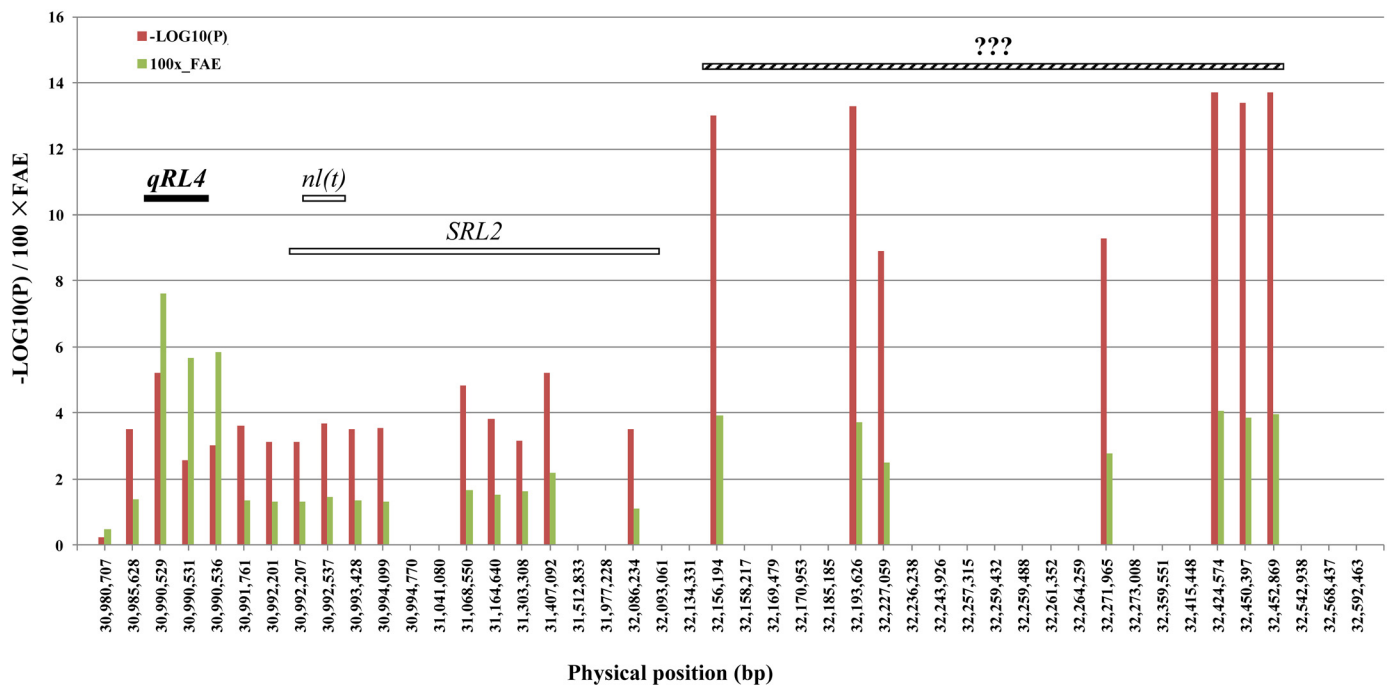


Fig 3. Subregional analysis of the region harboring *gRI4-2* by increasing marker density. The known loci, including the QTLs detected by linkage mapping in the MH63 × 02428 RIL population, the loci from the literature, and (likely) new clusters within subregions *gRI4-2_1*, *gRI4-2_2*, and *gRI4-2_3* are indicated by horizontal black, white, and striped bars, respectively. The vertical bars in red and green indicate the $-\text{LOG}_{10}(\text{P})$ and 100x favorable allele effect (FAE) values of the peak markers, respectively.

doi:10.1371/journal.pone.0158246.g003

the key components of plant architecture. The rolled leaf trait showed transgressive segregation in RILs derived from two parents with insignificant differences in this trait (LRI of no more than 10%, Fig 1). As shown in Table 1, favorable alleles at the detected QTLs are evenly dispersed in the two parents. Therefore, transgressive segregation of the rolled leaf trait in the RIL population can be partially explained by the reverse patterns of the allelic effects at the seven loci. Our results further support the observation that even though the germplasms themselves do not show prominent traits, they do harbor some excellent alleles, as previously observed for grain yield [75], salt tolerance [76], cold tolerance [13], and drought tolerance [12].

Of the seven loci identified for the rolled leaf trait, five were previously reported, and according to the comparative mapping, four of the above loci (*qRL4*, *qRL5*, *qRL6*, and *qRL7a*) are close to the previously reported loci *rl1(t)* [51], *qRL5-10* [22], *qRL-6* [32], and *qRL-7* [32], respectively. Two QTLs (*qRL7b* and *qRL9b*) are newly reported; the former was confirmed by GWAS analysis.

Previous studies have revealed multiple locus clusters, including regions on chromosome 4, 9, and 12 (Fig 2). Here, we found clustering of allele peaks based on the GWAS results, especially in *gRL4-2* at the end of chromosome 4, where at least three clusters of allele peaks were detected in the random subset of the 3K Rice germplasms panel (Fig 3).

Comparison of rolled leaf QTLs detected by GWAS with those revealed in earlier studies

Approximately six (33.3%) of the 18 GWAS loci were confirmed by our linkage mapping with an RIL population, five of which were previously reported (Table 2). An additional eight loci were supported by comparative mapping with the results from earlier reports. Taking together, these 14 (77.8%) GWAS loci are fairly reliable and appropriate for use in allele mining for breeding purposes. We identified at least 33 key SNP genotypes for favorable alleles at these loci, with an average of 2.4 SNP genotypes at each locus (Table 3).

Specifically, *gRL1-1* on chromosome 1 is located in the same region as the previously reported rolled leaf loci *qRL-1*, *url1(t)*, *rl-4(rl-2)*, and *SCL1* [32–35]. Two loci (*gRL2-1* and *gRL2-2*) on chromosome 2 were mapped together with *qRL-2-1b* [32] and *CFL1* [39], respectively. We identified two loci (*gRL4-1* and *gRL4-2*) on chromosome 4: the former is located in the same region as *ACL1*, for abaxial rolled leaves [49], and the latter is located in a region harboring *SRL2* [50], *rl11(t)* [51], *nl(t)* [52], and *qRL4-2* [22]. The latter locus also harbors QTL *qRL4*, as detected in our linkage map (Table 1). Two loci (*gRL5-3*, and *gRL5-4*) mapped together with *qRL5-9* [22], *qRL5-10* [22], and *rl8* [54], respectively, and *gRL5-4* was also detected on our linkage map (Table 1). Only one locus (*gRL6-1*) on chromosome 6, which was mapped together with *qRL6* detected by linkage mapping, was mapped in the same region with *qRL-6* from a previous report [32]. There are two significant loci on chromosome 7 and 8, respectively. Among these, *gRL7-1* and *gRL7-2* were mapped together with the loci *YABBY1* [8] and *qRL-7* [32] from previous reports, and *gRL8-1* and *gRL8-2* were mapped to the same regions as previously reported loci *qRL-8-1* and *qRL-8-2* [32], respectively. Only one locus (*gRL9*) on chromosome 9 was mapped together with *rl13(t)* from a previous study [55]. All of these candidate loci affecting the rolled leaf trait detected in different experiments are reliable and suitable for use in molecular breeding for plant type. Four loci (*gRL1-2*, *gRL5-1*, *gRL5-2*, and *gRL12-1*) were identified for the first time in this study and likely represent new QTLs for the rolled leaf trait.

To further explore the SNP markers with extremely higher density (average 18.1 ± 55.2 bp between adjacent markers), we split *gRL4-2* on chromosome 4 into at least three subregions. The second region, which is located in the region 31,068,550–32,086,234, was previously reported as *nl(t)* and *SRL2* [50,52]. The natural variations affect the LRI at a level of 1.7–2.2%, without significant correlation to leaf width, as detected in our diverse germplasm panel, although mutant

alleles, especially *nl(t)* at this locus, cause narrow leaves in addition to rolled leaves. The third region, located at 32,156,194–32,452,869, is thought to harbor new QTLs underlying the rolled leaf trait, with gene effects as high as 3.5% of the average gene effects for LRI.

By analyzing a highly diverse germplasm panel, we detected many new alleles, such as multiple alleles at the *ACL1* locus [49], in which the *ACL1* mutant has abaxial leaf rolling, while at least five favorable alleles from our 3K Rice germplasms improve the LRI by an average of 1.5%.

Implications for rice breeding for ideal plant type

The rolled leaf trait is a morphological character for which tremendous genetic variation exists among different rice genotypes, as shown in the current study. Suitably rolled leaves may allow rice plants to have greater effective leaf area per unit land without causing shading, thus likely resulting in extremely high yields due to higher rates of photosynthesis, as demonstrated in super hybrid rice [1]. Cultivars with moderately rolled leaves are suitable for relatively high density cultivation [11] and thought to be with better lodging and disease resistances at the population level [10]. Moreover, genotypes with partially rolled leaves may have better water use efficiency because rolled leaves are expected to have reduced leaf area [77]. Although both MH63 and 02428 have nearly flat leaves, the derived RILs showed various degrees of leaf rolling due to recombination of non-allelic parental alleles. Indeed, the most favorable LRI for *indica* cultivars is approximately 12% [78]. Therefore, it is possible to improve the leaf type of existing elite varieties by identifying “hidden” favorable alleles segregating in existing breeding populations and germplasms and introgressing and pyramiding them into elite backgrounds by MAS. The favorable alleles from IRIS_313–8023, IRIS_313–8027, IRIS_313–8149, IRIS_313–8129, and IRIS_313–8185, and perhaps even Nipponbare, at five QTLs (*qRl4*, *qRl5*, *qRl6*, *qRl7a*, and *qRl7b*) for the rolled leaf trait consistently identified in the RILs and in the re-sequenced germplasms (Table 3) in this study and in previous studies could be used to deploy allele combinations for ideal plant type in rice by MAS.

Conclusion

We identified seven main-effect QTLs underlying the transgressive segregation of the rolled leaf trait in rice in progenies from parents with minor differences in this trait. Five of these QTLs were previously reported, two (*qRl7b* and *qRl9b*) are newly identified, and one, *qRl7b*, was confirmed by GWAS analysis. Eighteen loci were found by GWAS: four are newly identified and the 14 other loci are consistent with QTLs from linkage mapping or comparative mapping. We carried out favorable allele mining for these 14 loci and identified possible elite donors for future plant type breeding programs. By performing subregional analysis, we identified a subregion (*gRl4-2_3*) with a possible new locus/loci and favorable alleles with an average effect of 3.5% for LRI, ranging from 2.4 to 4.1%. The favorable alleles at five QTLs (*qRl4*, *qRl5*, *qRl6*, *qRl7a*, and *qRl7b*) for the rolled leaf trait that were consistently identified in different populations could be used for breeding rice with ideal plant type by MAS.

Supporting Information

S1 Table. List of accessions used in this study.
(DOC)

Acknowledgments

We would like to thank the native English speaking scientists of Elixigen Company (Huntington Beach, California) for editing our manuscript.

Author Contributions

Conceived and designed the experiments: TQZ J LX ZKL. Performed the experiments: QZ LH. Analyzed the data: TQZ CCW. Contributed reagents/materials/analysis tools: QZ Nafisah CAJ. Wrote the paper: J LX WZZ TQZ.

References

1. Yuan LP Hybrid rice breeding for super high yield. *Hybrid Rice*.1997; 12: 1–6.
2. Liu CX, Pan GJ, Zhang XG Innovation practice on early-maturing and ideal plant type super rice Longjing-31 in cold region. *North Rice*.2014; 44: 1–4.
3. Bai L, Duan ZQ, Wang JM, An LZ, Zhao ZG, Chen KM Anatomical and chemical characteristics of a rolling leaf mutant of rice and its ecophysiological properties. *Rice Science*.2008; 15: 201–208.
4. Wang LF, Fu H, Ji YH Photosynthetic characterization of a rolled leaf mutant of rice (*Oryza sativa* L.). *African Journal of Biotechnology* 2012; 11: 6839–6846.
5. Hoang L QTL mapping for leaf rolling and yield component traits in a set of reciprocal introgression lines and RILs in rice (*Oryza sativa* L.). Beijing: Graduate School of Chinese Academy of Agricultural Sciences;2013.
6. Chen ZX, Hu J, Chen G, Pan XB Effects of rolled leaf gene *Rl(t)* on economic traits of hybrid rice. *Acta Agronomica Sinica*.2004; 30: 465–469.
7. Hu N, Lu CG, Yao KM, Zou JS Simulation on photosynthetically active radiation distributing in rice canopy with rolled leaves and its optimum leaf rolling index. *Chinese J Rice Sci*.2008; 22: 617–624.
8. Dai MQ, Zhao Y, Ma Q, Hu YF, Hedden P, Zhang QF, et al. The rice *YABBY1* gene is involved in the feedback regulation of gibberellin metabolism. *Plant Physiology*.2007; 144: 121–133. PMID: [17369428](https://pubmed.ncbi.nlm.nih.gov/17369428/)
9. Lu JF Effects of the rolling-leaf on biomass production and yield. Beijing: Yangzhou University;2002.
10. Lu CG, Zou JS Comparative analysis on plant type of two super hybrid rice and Shanyou63. *Scientia Agricultura Sinica*.2003; 36: 633–639.
11. Lang YZ, Zhang ZJ, Gu XY, Yang JC, Zhu QS Physiological and ecological effects of crimping leaf character in rice (*Oryza sativa* L.) I. Leaf orientation, canopy structure and light distribution. *Acta Agron Sin*.2004; 30: 806–810.
12. Ali AJ, Xu JL, Ismail AM, Fu BY, Vijaykumar CHM, Gao YM, et al. Hidden diversity for abiotic and biotic stress tolerances in the primary gene pool of rice revealed by a large backcross breeding program. *Field Crops Research*.2006; 97: 66–76.
13. He YX, Zheng TQ, Hao XB, Wang LF, Gao YM, Hua ZT, et al. Yield performances of *japonica* introgression lines selected for drought tolerance in a BC breeding programme. *Plant Breeding*.2010; 129: 167–175.
14. Li ZK, Zheng TQ Utilization of Exotic Germplasm. In: Zhang QF, Wing R, editors. *Genetics and Genomics of Rice*: Springer Science + Business Media, Inc.;2013. pp. 349–361.
15. Leung H, Raghavan C, Zhou B, Oliva R, Choi IR, Lacorte V, et al. Allele mining and enhanced genetic recombination for rice breeding. *Rice*.2015; 8: 34.
16. Lu YL, Zhang SH, Trushar S, Xie CX, Hao ZF, Li XH, et al. Joint linkage-linkage disequilibrium mapping is a powerful approach to detecting quantitative trait loci underlying drought tolerance in maize. *Proc Natl Acad Sci U S A*.2010; 107: 19585–19590. doi: [10.1073/pnas.1006105107](https://doi.org/10.1073/pnas.1006105107) PMID: [20974948](https://pubmed.ncbi.nlm.nih.gov/20974948/)
17. Zheng TQ, Yang J, Zhong WG, Zhai HQ, Zhu LH, Fan FJ, et al. Novel loci for field resistance to black-streaked dwarf and stripe viruses identified in a set of reciprocal introgression lines of rice (*Oryza sativa* L.). *Molecular Breeding*.2012; 29: 925–938.
18. The-3K-rice-genomes-project The 3,000 rice genomes project. *GigaScience*.2014; 3: 7.
19. Wu FX, Cai QH, Zhu YS, Zhang JF, Xie HA Application of indica restorer line, Minghui 63, for rice hybridization. *Fujian Journal of Agricultural Sciences*.2011; 26: 1101–1112.
20. Li HB A wide-compatibility rice germplasm—02428. *China Seed Industry*.1989: 42.
21. Nafisah QTL dissection of source and sink related traits using reciprocal introgression lines in rice (*Oryza sativa* L.) under different water regimes. Beijing: Graduate School of Chinese Academy of Agricultural Sciences;2013.
22. Gao YH, Lu CG, Wang MQ, Wang P, Yan XY, Xie K, et al. QTL mapping for rolled leaf gene in rice. *Jiangsu J of Agr Sci*.2007; 23: 5–10.

23. Huang X, Feng Q, Qian Q, Zhao Q, Wang L, Wang A, et al. High-throughput genotyping by whole-genome resequencing. *Genome Res.*2009; 19: 1068–1076. doi: [10.1101/gr.089516.108](https://doi.org/10.1101/gr.089516.108) PMID: [19420380](https://pubmed.ncbi.nlm.nih.gov/19420380/)
24. Kawahara Y, de la Bastide M, Hamilton JP, Kanamori H, McCombie WR, Ouyang S, et al. Improvement of the *Oryza sativa* Nipponbare reference genome using next generation sequence and optical map data. *Rice (N Y)*.2013; 6: 4.
25. Chen W, Chen HD, Zheng TQ, Yu R, Terzaghi WB, Li ZK, et al. Highly efficient genotyping of rice biparental populations by GoldenGate assays based on parental resequencing. *Theoretical Applied Genetics*.2014; 127: 297–307. doi: [10.1007/s00122-013-2218-2](https://doi.org/10.1007/s00122-013-2218-2) PMID: [24190103](https://pubmed.ncbi.nlm.nih.gov/24190103/)
26. Li H, Ye G, Wang J A modified algorithm for the improvement of composite interval mapping. *Genetics*.2007; 175: 361–374. PMID: [17110476](https://pubmed.ncbi.nlm.nih.gov/17110476/)
27. Miller MR, Dunham JP, Amores A, Cresko WA, Johnson EA Rapid and cost-effective polymorphism identification and genotyping using restriction site associated DNA (RAD) markers. *Genome Research*.2007; 17: 240–248. PMID: [17189378](https://pubmed.ncbi.nlm.nih.gov/17189378/)
28. cMap.Gramene database. Release 43#; 2015. Available: <http://www.gramene.org/>.
29. Zhang ZW, Ersoz E, Lai CQ, Todhunter RJ, Tiwari HK, Gore MA, et al. Mixed linear model approach adapted for genome-wide association studies. *Nat Genet*.2010; 42: 355–360. doi: [10.1038/ng.546](https://doi.org/10.1038/ng.546) PMID: [20208535](https://pubmed.ncbi.nlm.nih.gov/20208535/)
30. Lipka AE, Tian F, Wang Q, Peiffer J, Li M, Bradbury PJ, et al. GAPIT: genome association and prediction integrated tool. *Bioinformatics*.2012; 28: 2397–2399. PMID: [22796960](https://pubmed.ncbi.nlm.nih.gov/22796960/)
31. Li ZK QTL mapping in rice: a few critical considerations. In: Khush GS, Brar DS, Hardy B, editors. *Rice Genetics IV*. New Delhi (India) and Los Banos (Philippines): Science Publishers, Inc., and International Rice Research Institute;2001. pp. 153–172.
32. Guo Y, Cheng BS, Hong DL Construction of SSR linkage map and analysis of QTLs for rolled leaf in japonica rice. *Rice Science*.2010; 17: 28–34.
33. Yu D, Wu HB, Yang WT, Gong PT, Li YZ, Zhao DG Genetic analysis and mapping of the unilateral rolled leaf trait of rice mutant B157. *Molecular Plant Breeding*.2008; 6: 220–226.
34. Yoshimura A, Ideta O, Iwata N Linkage map of phenotype and RFLP markers in rice. *Plant Molecular Biology* 1997; 35: 49–60. PMID: [9291959](https://pubmed.ncbi.nlm.nih.gov/9291959/)
35. Shen NW Genetic analysis and gene mapping for the semi-curved leaf mutant in rice (*Oryza sativa* L.). Beijing: Graduate School of Chinese Academy of Agricultural Sciences;2011.
36. Fang YX, Zhu L, Pan JJ, Yu HP, Xue DW, Rao YC, et al. Identification and fine mapping of a narrow leaf mutant nal10 in rice *Chin J Rice Sci*.2015; 29: 587–594.
37. Chen QL, Xie QJ, Gao J, Wang WY, Sun B, Liu BH, et al. Characterization of Rolled and Erect Leaf 1 in regulating leave morphology in rice. *Journal of Experimental Botany*.2015; 66: 6047–6058. doi: [10.1093/jxb/erv319](https://doi.org/10.1093/jxb/erv319) PMID: [26142419](https://pubmed.ncbi.nlm.nih.gov/26142419/)
38. Gao J Cloning and functional analysis of leaf rolling related gene r18(t) of rice(*Oryza sativa* L.). Guangzhou: China Southern Agricultural University;2008.
39. Wu RH, Li SB, He S, Waßmann F, Yu CH, Qin GJ, et al. CFL1, a WW domain protein, regulates cuticle development by modulating the function of HDG1, a class IV homeodomain transcription factor, in rice and Arabidopsis. *The Plant Cell*.2011; 23: 3392–3411. doi: [10.1105/tpc.111.088625](https://doi.org/10.1105/tpc.111.088625) PMID: [21954461](https://pubmed.ncbi.nlm.nih.gov/21954461/)
40. Sang XC, Lin TT, He PL, Wang XW, Liao HX, Zhang XB, et al. Identification and gene mapping of a dominant narrow leaf mutant Dnal1 in rice (*Oryza sativa*). *Scientia Agricultura Sinica* 2014; 47: 1819–1827.
41. P C.H., L L., C Z.X., X X., Z Y.F., Z S.M., et al. Fine mapping of a rolled leaf gene *rl(t)* in rice. *Rice Science*.2011; 25: 455–460.
42. Li L, Xue X, Zuo SM, Chen ZX, Zhang YF, Li QQ, et al. Suppressed expression of OsAG01 a leads to adaxial leaf rolling in rice. *Chin J Rice Sci*.2013; 27: 223–230.
43. Zou LP, Sun XH, Zhang ZG, Liu P, Wu JX, Tian CJ, et al. Leaf rolling controlled by the homeodomain leucine zipper class IV gene *Roc5* in rice. *Plant Physiol*.2011; 156: 1589–1602. doi: [10.1104/pp.111.176016](https://doi.org/10.1104/pp.111.176016) PMID: [21596949](https://pubmed.ncbi.nlm.nih.gov/21596949/)
44. Zhang XH, Qin YZ, Zhang YX, Zhan XD, Zhang ZH, Shen XH, et al. Gene mapping of a narrow and rolled leaf mutant *Nrl3(t)* in rice. *Chin J Rice Sci*.2015; 29: 595–600.
45. Xie ZW, Sun W, Yin L, Zhao JF, Yuan SJ, Zhang WH, et al. Phenotypic and genetic analyses of a novel adaxially-rolled leaf mutant in rice. *Acta Agronomica Sinica*.2013; 39: 1970–1975.
46. Wu C, Fu Y, Hu G, Si H, Cheng S, Liu W Isolation and characterization of a rice mutant with narrow and rolled leaves. *Planta*.2010; 232: 313–324. doi: [10.1007/s00425-010-1180-3](https://doi.org/10.1007/s00425-010-1180-3) PMID: [20443024](https://pubmed.ncbi.nlm.nih.gov/20443024/)

47. Wang DZ, Sang XC, You XQ, Wang Z, Wang QS, Zhao FM, et al. Genetic analysis and gene mapping of a novel narrow and rolled leaf mutant *nrl2(t)* in rice (*Oryza sativa* L.). *Acta Agronomica Sinica* 2011; 37: 1159–1166.
48. Shi Z, Wang J, Wan X, Shen G, Wang X, Zhang J Over-expression of rice *OsAGO7* gene induces upward curling of the leaf blade that enhanced erect-leaf habit. *Planta*.2007; 226: 99–108.
49. Li L, Shi ZY, Li L, Shen GZ, Wang XQ, An LS, et al. Overexpression of *ACL1* (*abaxially curled leaf 1*) increased Bulliform cells and induced Abaxial curling of leaf blades in rice. *Mol Plant*.2010; 3: 807–817.
50. Lai KK Genetic and fine mapping analysis of a semi-dominant rolled leaf mutant. Hangzhou: Hangzhou Normal University;2012.
51. Zhou Y, Fang YX, Zhu JY, Li SQ, Gu F, Gu MH, et al. Genetic analysis and gene fine mapping of a rolling leaf mutant (*rl11(t)*) in rice (*Oryza sativa* L.). *Chinese Sci Bull*.2010; 55: 1763–1769.
52. Pan YL, Mao BG, Hu YY, Guo LQ, Peng Y, Shao Y, et al. Genetic analysis and gene mapping of a narrow-leaf mutant *nl(t)* in rice (*Oryza sativa* L.). *Journal of Biology*.2015; 32: 92–95.
53. Xu JL, Zhong DB, Yu SB, Luo LJ, Li ZK QTLs affecting leaf rolling and folding in rice. *Rice Genet Newsl*.1999; 16: 51–52.
54. Shao YJ, Chen ZX, Zhang YF, Chen EH, Qi DC, Miao J, et al. One major QTL mapping and physical map construction for rolled leaf in rice. *Yi Chuan Xue Bao*.2005; 32: 501–506. PMID: [16018261](#)
55. Chen YZ, Liu PQ, Bai DL, Li RB Genetic analysis and gene mapping of a new rolled leaf mutant in rice (*Oryza sativa* L.). *Guangxi Agricultural Sciences*.2010; 41: 403–407.
56. Xia L, Chen L, Guo CM, Zhang HX, Zhao Z, Shen MS, et al. Genetic analysis and mapping of rice (*Oryza sativa* L.) *sd-sl* mutant. *Journal of Xiamen University(Natural Science)*.2007; 46: 847–851.
57. Xiang JJ, Zhang GH, Qian Q, Xue HW Semi-rolled leaf1 encodes a putative glycosylphosphatidylinositol-anchored protein and modulates rice leaf rolling by regulating the formation of bulliform cells. *Plant Physiol*.2012; 159: 1488–1500. doi: [10.1104/pp.112.199968](#) PMID: [22715111](#)
58. Wang F, Tang YQ, Miao RL, Xu FF, Lin TT, He GH, et al. Identification and gene mapping of a narrow and upper-albino leaf mutant in rice (*Oryza sativa* L.). *Chinese Science Bulletin*.2012; 57: 3798–3803.
59. Alexandrov N, Tai S, Wang W, Mansueto L, Palis K, Fuentes RR, et al. SNP-Seek database of SNPs derived from 3000 rice genomes. *Nucleic Acids Res*.2015; 43: D1023–1027. doi: [10.1093/nar/gku1039](#) PMID: [25429973](#)
60. Yang CH, Li D, Liu X, Ji CJ, Hao LL, Zhao XF, et al. *OsMYB103L*, an R2R3-MYB transcription factor, influences leaf rolling and mechanical strength in rice (*Oryza sativa* L.). *BMC Plant Biology*.2014; 14: 158. doi: [10.1186/1471-2229-14-158](#) PMID: [24906444](#)
61. Liu C, Kong WY, You SM, Zhong XJ, Jiang L, Zhao ZG, et al. Genetic analysis and fine mapping of a novel rolled leaf gene in rice *Scientia Agricultura Sinica* 2015; 48: 2487–2496.
62. Luo Z, Yang Z, Zhong B, Li Y, Xie R, Zhao F, et al. Genetic analysis and fine mapping of a dynamic rolled leaf gene, *RL10(t)*, in rice (*Oryza sativa* L.). *Genome*.2007; 50: 811–817. PMID: [17893721](#)
63. Zhang GH, Xu Q, Zhu XD, Qian Q, Xue HW SHALLOT-LIKE1 is a KANADI transcription factor that modulates rice leaf rolling by regulating leaf abaxial cell development. *Plant Cell*.2009; 21: 719–735. doi: [10.1105/tpc.108.061457](#) PMID: [19304938](#)
64. Yan S, Yan CJ, Zeng XH, Yang YC, Fang YW, Tian CY, et al. *ROLLED LEAF 9*, encoding a GARP protein, regulates the leaf abaxial cell fate in rice. *Plant Mol Biol*.2008; 68: 239–250. doi: [10.1007/s11103-008-9365-x](#) PMID: [18594992](#)
65. Xu Y, Wang Y, Long Q, Huang J, Wang Y, Zhou K, et al. Overexpression of *OsZHD1*, a zinc finger homeodomain class homeobox transcription factor, induces abaxially curled and drooping leaf in rice. *Planta*.2014; 239: 803–816. doi: [10.1007/s00425-013-2009-7](#) PMID: [24385091](#)
66. Zhao FM, Wei X, Ma L, Sang XC, Wang N, Zhang CW, et al. Identification, gene mapping and candidate gene prediction of a late-stage rolled leaf mutant *lr1* in rice (*Oryza sativa* L.). *Chin Sci Bull*.2015; 60: 3133–3143.
67. Wang X, Gu FG, Sun BY Ds-tagged rice rolling leaf mutant abnormal in bulliform cells. *Journal of Soochow University(Natural Science Edition)*.2012; 28: 89–94.
68. Luo YZ, Zhao FM, Sang XC, Ling Y-h, Yang ZL, He GH Genetic analysis and gene mapping of a novel rolled leaf mutant *rl12(t)* in rice. *Acta Agronomica Sinica* 2009; 35: 1967–1972.
69. Zhang LX, Liu HQ, Yu X, Wang LY, Fan HH, Jin QS, et al. Molecular mapping and physiological characterization of a novel mutant *rl15(t)* in rice. *Scientia Agricultura Sinica* 2014; 47: 2881–2888.
70. Zhang FT, Fang J, Sun CH, Li RB, Luo XD, Xie JK, et al. Characterisation of a rice dwarf and twist leaf 1 (*dtl1*) mutant and fine mapping of *DTL1* gene. *Hereditas(Beijing)*.2012; 34: 79–86.

71. Fang L, Zhao F, Cong Y, Sang X, Du Q, Wang D, et al. Rolling-leaf14 is a 2OG-Fe (II) oxygenase family protein that modulates rice leaf rolling by affecting secondary cell wall formation in leaves. *Plant Biotechnol J.* 2012; 10: 524–532. doi: [10.1111/j.1467-7652.2012.00679.x](https://doi.org/10.1111/j.1467-7652.2012.00679.x) PMID: [22329407](https://pubmed.ncbi.nlm.nih.gov/22329407/)
72. Segami S, Kono I, Ando T, Yano M, Kitano H, Miura K, et al. Small and round seed 5 gene encodes alpha-tubulin regulating seed cell elongation in rice. *Rice.* 2012; 5: 4. doi: [10.1186/1939-8433-5-4](https://doi.org/10.1186/1939-8433-5-4) PMID: [24764504](https://pubmed.ncbi.nlm.nih.gov/24764504/)
73. Hu J, Zhu L, Zeng D, Gao Z, Guo L, Fang Y, et al. Identification and characterization of NARROW AND ROLLED LEAF 1, a novel gene regulating leaf morphology and plant architecture in rice. *Plant Mol Biol.* 2010; 73: 283–292. doi: [10.1007/s11103-010-9614-7](https://doi.org/10.1007/s11103-010-9614-7) PMID: [20155303](https://pubmed.ncbi.nlm.nih.gov/20155303/)
74. Wang D, Liu H, Li K, Li S, Tao Y Genetic analysis and gene mapping of a narrow leaf mutant in rice (*Oryza sativa* L.). *Chinese Science Bulletin.* 2009; 54: 752–758.
75. Zhang HJ, Wang H, Qian YL, Xia JF, Li ZF, Shi YY, et al. Simultaneous improvement and genetic dissection of grain yield and its related traits in a backbone parent of hybrid rice (*Oryza sativa* L.) using selective introgression. *Molecular Breeding.* 2012; 31: 181–194.
76. Zang J, Sun Y, Wang Y, Yang J, Li F, Zhou Y, et al. Dissection of genetic overlap of salt tolerance QTLs at the seedling and tillering stages using backcross introgression lines in rice. *SciChina CLife Sci.* 2008; 51: 583–591.
77. Dingkuhn M, Cruz R, O'Toole J, Dreyer K Net photosynthesis, water use efficiency, leaf water potential and leaf rolling as affected by water deficit in tropical upland rice. *Australian Journal of Agricultural Research.* 1989; 40: 1171–1181.
78. Lang YZ, Zhang ZJ, Gu XY, Yang JC, Zhu QS Physiological and ecological effects of crimp leaf character in rice (*Oryza sativa* L.) II. photosynthetic character, dry mass production and yield forming. *Acta Agronomica Sinica.* 2004; 30: 883–887.