

Advances in Cardiovascular Disease Lipid Research Can Provide Novel Insights Into Mycobacterial Pathogenesis

Shyamala Thirunavukkarasu and Shabaana A. Khader*

Department of Molecular Microbiology, Washington University in St. Louis School of Medicine, St. Louis, MO, United States

OPEN ACCESS

Edited by:

Stephane Canaan, Centre National de la Recherche Scientifique (CNRS), France

Reviewed by:

Xuwen Peng, Penn State Milton S. Hershey Medical Center, United States Evgeniya V. Nazarova, Immunology Discovery, Genentech, United States Natalie Garton, University of Leicester, United Kingdom

*Correspondence:

Shabaana A. Khader sakhader@wustl.edu

Specialty section:

This article was submitted to Molecular Bacterial Pathogenesis, a section of the journal Frontiers in Cellular and Infection Microbiology

Received: 20 November 2018 Accepted: 02 April 2019 Published: 18 April 2019

Citation:

Thirunavukkarasu S and Khader SA (2019) Advances in Cardiovascular Disease Lipid Research Can Provide Novel Insights Into Mycobacterial Pathogenesis. Front. Cell. Infect. Microbiol. 9:116. doi: 10.3389/fcimb.2019.00116 Cardiovascular disease (CVD) is the leading cause of death in industrialized nations and an emerging health problem in the developing world. Systemic inflammatory processes associated with alterations in lipid metabolism are a major contributing factor that mediates the development of CVDs, especially atherosclerosis. Therefore, the pathways promoting alterations in lipid metabolism and the interplay between varying cellular types, signaling agents, and effector molecules have been well-studied. Mycobacterial species are the causative agents of various infectious diseases in both humans and animals. Modulation of host lipid metabolism by mycobacteria plays a prominent role in its survival strategy within the host as well as in disease pathogenesis. However, there are still several knowledge gaps in the mechanistic understanding of how mycobacteria can alter host lipid metabolism. Considering the in-depth research available in the area of cardiovascular research, this review presents an overview of the parallel areas of research in host lipid-mediated immunological changes that might be extrapolated and explored to understand the underlying basis of mycobacterial pathogenesis.

Keywords: mycobacterium, tuberculosis, granuloma, lipid, cardiovascular, plaque, arachidonic acid, foam cell

INTRODUCTION

One of the leading causes of morbidity and mortality in westernized countries is cardiovascular disease (CVD), such as atherosclerosis (Yeates et al., 2015). Atherosclerosis is a complex, chronic, progressive, inflammatory disease involving different cell types resulting in the formation of an atheromatous plaque. Atherosclerosis is characterized by infiltration of the arterial intima by macrophages which scavenge oxidized low density lipoprotein (oxLDL), which further promotes alterations in cholesterol influx, esterification and efflux, ultimately resulting in the progression of the macrophage into a foam cell. The specific contributions of lipids and lipoproteins as well as the influence of cholesterol metabolism in the formation of atheromatous plaques has been extensively researched in the context of atherosclerosis and CVD (Chroni et al., 2011).

Mycobacterium tuberculosis (*Mtb*) is a successful human pathogen due to its ability to cause tuberculosis (TB) in almost 10 million individuals annually (Dye and Williams, 2010). Non-tuberculous mycobacteria (NTM) also cause diseases such as pulmonary and skin infections, in addition to being implicated as putative causative agents of Crohn's disease in humans (Thirunavukkarasu et al., 2017a). NTM share commonalities with tuberculous mycobacteria with regard to subversion of host macrophage immune responses (Whittington et al., 2012; Thirunavukkarasu et al., 2017a). A primary reason for the ubiquitous spread of mycobacteria

infection despite current control strategies is the ability of pathogenic mycobacteria to persist in a non-replicative state both within the host, and sometimes in the environment (Falkinham, 2009). Macrophages play a pivotal role in the immune response against mycobacteria (Pieters, 2008; Thirunavukkarasu et al., 2015; Mcclean and Tobin, 2016). One of the main mechanisms of the successful intra-macrophage survival of mycobacteria including Mtb, M. avium, M. bovis, M. paratuberculosis, M. ulcerans, and M. leprae is their capacity to manipulate the host cellular metabolism to utilize intracellular substrates including fatty acids and cholesterol (Mendum et al., 2015). This manipulation of the host macrophage lipid metabolic pathway is a hallmark of several mycobacterial infections including TB (Peyron et al., 2008; Russell et al., 2009; Almeida et al., 2012; Caire-Brändli et al., 2014). Dysregulated lipid metabolism resulting in foam cell formation in macrophages and other cell types, and its association with steroid hormones as well as granuloma lesion formation, is a critical aspect in understanding mycobacterial pathogenesis. However, current research on the contribution of host lipid metabolic pathways in disease pathogenesis is limited, unlike in cardiovascular research where it has been the focus of extensive studies (Tambo et al., 2016). The similarities in the immune responses in the kinetics of atherosclerotic plaque formation and a granuloma formation in TB is an exposition of how knowledge could be gained by extrapolating ideas from among these fields.

NIH has identified interdisciplinarity as an essential contributor to needed knowledge and made it an explicit priority in its roadmap. Considering the several areas of similarities between the immunopathology of atherosclerosis and mycobacteriosis, it would be applicable to explore and extrapolate the plethora of information available in this arena in CVD research to address the knowledge gaps in the area of host lipid metabolism in mycobacterial research. However, comprehensive review articles providing reference pools for promoting scientific knowledge in interdisciplinary applications between CVD and mycobacterial immunopathology are lacking. Therefore, the purpose of this review is to identify and put forth the similarities in relation to alterations in host lipid metabolism contributing to disease pathology induced by cardiovascular and mycobacterial diseases. Furthermore, we highlight the recent advances pertaining to host lipid metabolism in CVD immunopathology that could provide potential avenues to explore for researchers involved in studying mycobacterial pathogenesis.

MACROPHAGE INTRACELLULAR LIPID METABOLISM AND FOAM CELL FORMATION

A key feature in atherosclerosis and mycobacteriosis is the presence of lipid laden cells called foam cells (Kruth, 2001; Chen et al., 2008; Almeida et al., 2012; Bah et al., 2017). The process of foam cell formation is directly or indirectly influenced by uptake of native or modified lipoproteins by cell types present in atheromatous lesions especially macrophages, subsequent

processing and retention of intracellular lipids, and altered reverse cholesterol transport (Remmerie and Scott, 2018). Lipids are not soluble in plasma and circulating lipids are transported to various tissues in the form of lipoproteins. Additionally, lipids can also circulate in blood in an albumin bound manner and can be taken into macrophages via micropinocytosis in a receptor independent manner (Kruth, 2013).

Lipoproteins are complex particles made up of a central hydrophobic core of non-polar lipids, primarily cholesterol esters, and triacyl glycerides (TAG). This hydrophobic core is surrounded by a hydrophilic membrane consisting of phospholipids, free cholesterol, and apolipoproteins (Chistiakov et al., 2017). Based on size and concentration, lipoproteins are classified as chylomicrons, very low density lipoproteins (VLDL), low density lipoproteins (LDL), or high density lipoproteins (HDL). LDL particles transport cholesterol esters, by associating with apolipoproteins B-100 and C-III, and are taken up inside the cell via the low density lipoprotein receptor (LDLR) (Rosenson et al., 2011). Once inside, LDL is digested inside lysosomes and free cholesterol is released. Free cholesterol can then either move to the membrane surface and undergo efflux via ATPbinding cassette transporter (ABC) A1 or ABCG1, or enter the endoplasmic reticulum (ER) where they are esterified to form lipid droplets by acyl CoA acetyl transferase 1 (ACAT-1) (Chistiakov et al., 2017). Free cholesterol could also cause peristent stress to the ER which ultimately results in excess nuclear factor kappa B (NFkB) signaling and apoptosis. Excess free cholesterol inside the cells can also result in the formation of cholesterol crystals, leading to enrichment of the cell membrane lipid rafts causing toll-like receptor (TLR) activation, and downstream signaling events (Remmerie and Scott, 2018).

LDL also undergoes oxidation resulting in the formation OxLDL. The sub-endothelial space is considered to be the site of LDL oxidation as the antioxidant properties of blood prevents LDL oxidation while in circulation (Matsuura et al., 2008). LDL modification to OxLDL occurs by multiple mechanisms including oxidative and nitrosative stress (ROS and NOS) as well as enzymes like 12/15 lipoxygenases (LOX) and secretory phospholipase A2 (sPLA2) (Linton et al., 2015). OxLDL comprises of a variety of by-products arising due to the modification of both lipids and apolipoprotein B by lipid peroxidation (Matsuura et al., 2008). OxLDL is taken up preferentially by the macrophages by endocytosis, via scavenger receptors such as CD36, macrophage receptor with collagenous structure (MARCO), scavenger receptor B1 (SR-B1), and LOX1, as well as CD14-Toll like receptor 4-MD2 complex (Kruth, 2001). This induces ROS generation, cytokine secretion, production of myeloperoxidase, and 12/15 lipoxygenase secretion which further oxidize new LDL thus increasing the local pool of oxLDL. The components of oxLDL also act as ligands for the peroxisome proliferator activated receptor gamma (PPARy) which in turn increases the intake of more OxLDL via upregulation of CD36.

Abbreviations: CVD, cardiovascular disease; TB, tuberculosis; *Mtb*, *Mycobacterium tuberculosis*; AA, arachidonic acid; COX, cyclooxygenase; LOX, lipoxygenase; CYP, Cytochrome; oxLDL, oxidized low density lipoprotein; PGE2, Prostaglandin E2; IFN, interferon.

Moreover, accumulating oxLDL blocks the PPARy mediated liver X receptor (LXR) signaling which favors cholesterol efflux due to upregulation of ABCA1. The events result in the macrophage transforming into a foam cell (Figure 1). Internalization of OxLDL by CD36 decreases macrophage migration and activates focal adhesion kinases. Therefore, OxLDL contributes to both macrophage activation as well as retention (Parthasarathy et al., 2010). In addition, OxLDL increases platelet derived growth factor (PDGF) and other growth factors secretion thus amplifying cellular proliferation. The OxLDL mediated ROS generated triggers the cytochrome c mediated apoptosis cascade leading to cellular death and debris accumulation (Leiva et al., 2015). This coupled with a malfunctioning lipid metabolic pathway can impact the apoptotic cell clearance by other macrophages resulting in chronic inflammation (Szondy et al., 2014), which may further enhance foam cell formation. Mtb which enter the host macrophages has the capacity to utilize host cholesterol and also sequester host fatty acids in the form of TAG within intra-bacterial lipid inclusion bodies believed to occur via bacterial mce transporters mediated by Rv3723/LucA (Pandey and Sassetti, 2008; Daniel et al., 2011; Nazarova et al., 2017). This lipid accquisition is required for bacterial persistence especially in the chronic phase of disease (Neyrolles, 2014).

Since foam cell formation is an essential aspect of the immunopathology of both atherosclerosis and several mycobacterial infections, the knowledge regarding the lipid mediators, and signaling events that contribute to this are of interest.

Arachidonic Acid and Lipid Mediators

OxLDL activates phospholipase enzymes which act on membrane phospholipids releasing arachidonic acid (AA) (Akiba et al., 2003). Arachidonic acid is the substrate from which the biologically active lipid mediators belonging to the eicosanoid family are produced due to the activity of various enzymes (Akiba et al., 2003). These enzymes belong to three pathways, namely (a) the COX pathway (COX1, COX2) resulting in production of prostaglandins (PG), (b) the LOX pathway (5-LOX, 1LOX, 15LOX) resulting in production of leukotrienes (LTs) and lipoxins (LXs), and (c) the cytochrome P450 (Cyp) pathway which forms the hydroxyeicosotetranoic acids (HETEs) and the epoxyeicosotrienoic acids (EETEs) (Hanna and Hafez, 2018). Non-enzymatic lipid peroxidation of AA results in the production of isoprostanes and isoketals. PGs and LTs are the initial mediators which recruit neutrophils and monocytes to the site of inflammation (Dietzold et al., 2015). The importance and involvement of PGE2 and other lipid mediators and the signaling events of the arachidonic acid metabolic pathway (Figure 2) in CVDs in general and atherosclerosis in particular has been extensively explored utilizing several approaches which could offer insight for mycobacterial researchers to pursue.

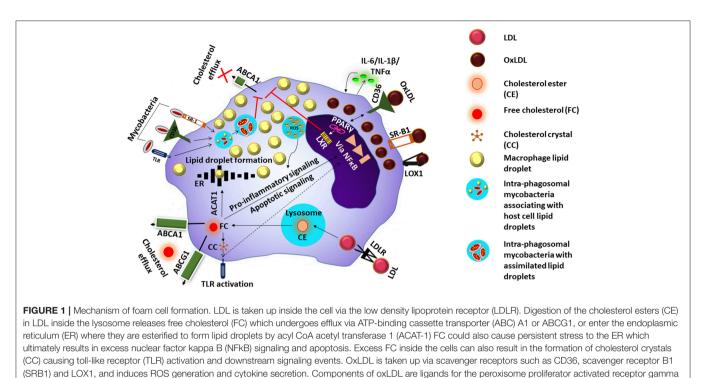
LESION: ATHEROSCLEROTIC PLAQUE VS. TB GRANULOMA

An atherosclerotic plaque formed as a result of dysregulated lipid metabolism is either stable and characterized by low

inflammatory cell infiltration with a thick fibrous cap or an unstable plaque characterized by a necrotic core of foamy macrophages, degradation of the collagenous fibrous cap by matrix metalloproteinases, and subsequent rupture of the plaque necrotic core (Kruth, 2001). Several enzymes including those belonging to the Cyp family play crucial roles in the generation of either a stable plaque or progression to an unstable plaque with poor prognosis for atherosclerosis outcome (Silvestre-Roig et al., 2014; Song et al., 2016; Stefanadis et al., 2017).

The stability of the atheromatous plaques formed directs disease outcome in atherosclerosis (Chen et al., 2016). Similarly, many mycobacterial infections including Mtb, M. paratuberculosis, M. bovis, and M. marinum are characterized by granuloma formation, the relative stability and organization of which impacts disease severity and outcome (Wangoo et al., 2005; Silva Miranda et al., 2012; Wu et al., 2012; Guirado and Schlesinger, 2013; Fernández et al., 2016). Granulomas are formed as organized aggregates of immune cells composed primarily of infected and uninfected macrophages, foamy macrophages, and other cell types including NK cells and DCs that are surrounded by a ring of B and T lymphocytes (Silva Miranda et al., 2012). Granulomas are formed as a means to control chronic mycobacterial infection, but progression to cavitary granuloma leads to subclinical and clinical disease (Guirado and Schlesinger, 2013). Thus, the TB granuloma is categorized as either stable wherein the bacteria are contained and infection is mostly latent or as a disorganized granuloma with a necrotic caseous core, matrix metalloproteinase-mediated loss of the collagen layer, and dissemination of infection leading to active TB with a much poorer prognosis (Kim et al., 2010; Russell et al., 2010). Although there are differences between an atherosclerotic plaque and TB granuloma regarding the role of T cells and B cells, similarities in dysregulated host lipid metabolism which contribute to disease pathogenesis are indeed conspicuous in both diseases (Figure 3).

Cholesterol catabolism, apart from providing a potent source of carbon for mycobacterial metabolism (Wilburn et al., 2018), also results in the production of propionyl CoA and methylmalonyl CoA that are used for the synthesis of mycobacterial cell wall virulence components such as phthiocerol dimycocerosate (PDIM) (Jain et al., 2007), and sulpholipids (Pandey and Sassetti, 2008; Griffin et al., 2012; Abuhammad, 2017). Since cholesterol modulates mycobacterial metabolism, identification of novel targets in the cholesterol metabolic pathway has been carried out for the purpose of chemotherapeutic intervention of TB (Vanderven et al., 2015). Moreover, the modulation of host cholesterol can have adverse effects on several host signaling mechanisms as well as cellular immune polarization. It is believed that lipid mediated signaling via the sterol regulatory element binding protein 1 leads to a pro-inflammatory M1 phenotype in macrophages while signaling via LXR leads to an anti-inflammatory M2 phenotype (Tangirala et al., 2002). Cholesterol also acts as a substrate for various enzyme activities including those belonging to the Cyp family (Pikuleva, 2006; Rogers et al., 2015). Furthermore, cholesterol is also a precursor for major sex hormones and a role for sex differences in the host immune responses to CVD (Jennings et al., 2014; Pingili et al., 2015) and mycobacterial



(PPARy) which increases the intake of more OxLDL via upregulation of CD36. Activation of PPARy triggers liver X receptor signaling which facilitates lipid efflux via upregulation of ACDJ/FC results in the macrophage transforming into a foam cell. Mycobacteria entering the cell via the cell surface receptors are taken up by phagosomes and associate with lipid droplets and assimilate them facilitating their survival in the host cell whose lipid metabolism is further impacted by the mycobacteria.

infections (Neyrolles and Quintana-Murci, 2009; Bini et al., 2014; Karunasena et al., 2014; Nhamoyebonde and Leslie, 2014; Mirsaeidi and Sadikot, 2015; Dibbern et al., 2017), has been of interest in recent years, with men reportedly being more susceptible than women to CVD as well as mycobacterial infection. In this context, it is important to note that although a role for sex steroid hormones in influencing susceptibility to pulmonary tuberculosis has also been established (Bini et al., 2014), it has not been researched indepth as is the case in CVD.

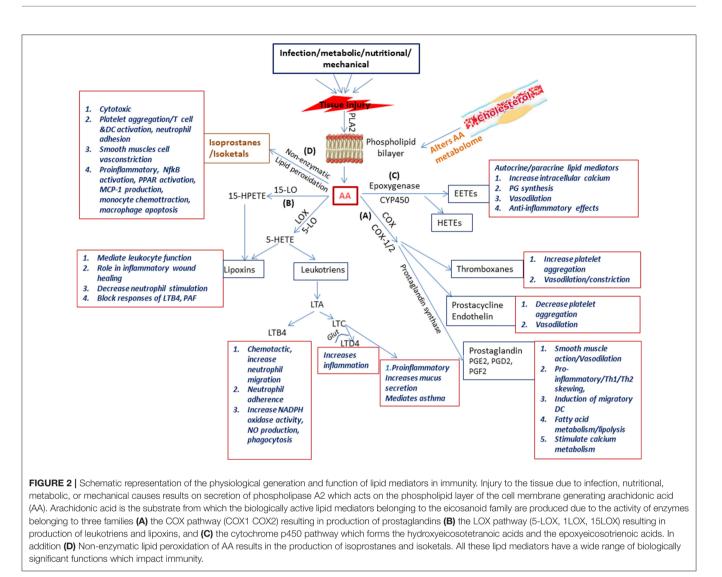
ROLE OF LIPID METABOLISM IN MYCOBACTERIAL PATHOGENESIS

The outcome of initial exposure of *Mtb* in the human host could either result in the pathogen progressing to primary active TB disease or efficient elimination through an innate and/or acquired immune response. Mycobacteria could also survive inside the host in a non-replicative state but capable of reactivation and subsequent progression to the active form of disease. The active phase of TB infection is characterized by the formation of caseous granulomas and cavities which are correlated with failure of control and disease relapse (Russell et al., 2010). The progression of human TB infection, from latent infection to active disease, occurs as a continuous spectrum of metabolic bacterial activity amidst the onslaught of protective host immune responses (Drain et al., 2018). Furthermore, the

asymptomatic but well-contained disease forms in the early stages of infection are classified as incipient disease while the asymptomatic disease at the latent end of the spectrum associated with loss of effective containment is termed as subclinical TB (Achkar and Jenny-Avital, 2011). Several studies have reported on the role of host and pathogen derived lipids both *in vitro* and *in vivo* in disease pathogenicity caused by mycobacterial sp. in general, and *Mtb* in particular (Guenin-Mace et al., 2009; Philips and Ernst, 2012; Lee et al., 2013; Thirunavukkarasu et al., 2014, 2016a). Thus, the modulation of the host lipid metabolism could have an impact on the different stages of the TB disease spectrum, including granuloma formation. In the following sections we provide a snapshot of the lipid mediators and pathways that affect TB disease progression and outcome.

Role of Lipid Mediators in TB Progression

Foamy macrophages formed during the course of mycobacterial infection are rich sites of PGE2 generation (Almeida et al., 2009) and therefore cross regulation of eicosanoid pathways could play a critical role in controlling the growth of pathogenic mycobacteria. PGE2 is an important lipid mediator as it has the potential to regulate its own production, while suppressing acute inflammatory mediators, resulting in its predominance at late/chronic stages of immunity during a wide range of autoimmune, metabolic, and infectious diseases (Kalinski, 2012). Therefore, inhibition of PGE2 production and signaling could potentially represent a therapeutic alternative to treat bacterial



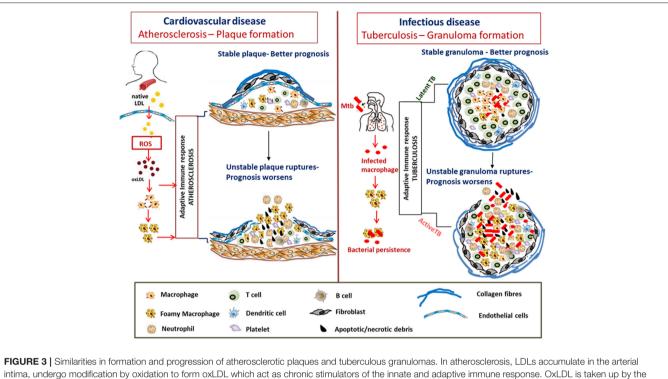
infections in the chronic phase (Agard et al., 2013), although augmentation of PGE2 levels reportedly improves protection against Mtb in the acute stages of infection (Mayer-Barber et al., 2014). Apart from PGE2, another area gaining importance in mycobacterial research in recent years is the lipoxygenase pathway. Mice deficient in 5-LOX were found to be more resistant to Mtb infection (Bafica et al., 2005; Chen et al., 2008). Moreover, LOX-derived eicosanoids are known to regulate DC maturation and function as well as a neutrophilic recruitment cascade during Mtb infection (Rothe et al., 2015; Mishra and Sassetti, 2016). Zebra fish larvae deficient in LTA4 hydrolase were found to have enhanced susceptibility to M. marinum during the chronic phase of infection, due to the utilization of eicosanoid substrates leading to the production of anti-inflammatory lipoxins and an increase in TNF- α levels (Tobin et al., 2010, 2012). Hence, it is likely that these pathways/mediators might have a differential effect depending on the stage of infection which could in turn define the final disease outcome.

Inhibition of apotosis is a mechanism by which mycobacteria can persist in the host and this is also reported to occur at the

level of lipid mediators (Dietzold et al., 2015). Virulent strains of *Mtb* are known to enhance production of leukotriens while suppressing cyclooxygenase (COX)2-induced PGE2 synthesis, which drives the cells toward necrosis (Rocca and Fitzgerald, 2002; Chen et al., 2008). Moreover, it is known that *Mtb* also utilizes host ceramide, which reduces the capacity of aveolar macrophages to clear apoptotic cells leading to persistent inflammation (Mattos et al., 2011). Of interest, ceramides are a known antagonist of foam cell formation via decreased transport of CD36 to the macrophage cell membrane (Chistiakov et al., 2016), therefore utilizing the host ceramide supply could indirectly drive foam cell formation and disease progression.

Cytokines Involved in Modulating Host Lipid Metabolism in *Mtb* Infection

Mycobacterial lipid antigens are known to both enhance as well as suppress secretion of pro-inflammatory cytokines such as TNF- α , interleukin (IL)-1 α , IL-1 β , and IL-6 by host macrophages (Lee et al., 2007; Thirunavukkarasu et al., 2013, 2017b; Howard et al., 2018). However, the role of these



intima, undergo modification by oxidation to form oxLDL which act as chronic stimulators of the innate and adaptive immune response. OxLDL is taken up by the macrophages via scavenger receptors forming foam cells. The plaque is composed of neutrophils, platelets, T cells, and DC along with foam cells and resultant secretion of pro-inflammatory cytokines that accentuate inflammation. Apoptosis and secondary necrosis of foam cells and SMCs result in necrotic core development, degradation of the collagen layer forming unstable plaque that eventually ruptures resulting in bad prognosis. *Mtb* infection is also characterized by foam cell formation which is crucial for bacterial peristence. The cellular composition of TB granulomatous lesions includes infected and uninfected macrophages, foamy macrophages, B and T lymphocytes, neutrophils, platelets, T cells, and fibroblasts. The production of bacteria worsening the prognosis.

cytokines in altering host lipid metabolism and promoting macrophage foam cell formation during *Mtb* infection has not been addressed. In contrast, both type I and type II interferon (IFN)-induced alterations in host lipid metabolism leading to either mycobacterial persistence or protective immunity has been previously addressed (Donovan et al., 2017; Knight et al., 2018). While type II signaling is associated with protective immunity and lipid droplet formation involving PGE2 and LXB4, type I (Knight et al., 2018), signaling suppresses IL-1 β induced protective PGE2 conferring susceptibility to mycobacterial infection (Mayer-Barber and Yan, 2016).

Recently there has been a renewed interest in the role of type I interferons in affecting *de novo* cholesterol and fatty acid synthesis as well as influencing uptake of exogenous lipids by cells during the course of infectious diseases (Blanc et al., 2011, 2013; York et al., 2016). The role of type I IFN in specific cellular subsets is also being explored with recent reports claiming that pDCs (plasmacytoid dendritic cells) which characteristically produce copious type I IFN (Swiecki and Colonna, 2015) alter cellular metabolism by increasing fatty acid oxidation and oxidative phosphorylation, which is mediated in an autocrine manner by the type I IFN secreted (Wu et al., 2016). Moreover, type I IFN signaling via signal transducer and activator of transcription 1 (STAT-1) enhances expression of the cholesterol 25 hydroxylase (*ch25h*) gene and production of 25-hydroxy cholesterol as

well as miR342-5p which suppresses the sterol biosynthetic pathway (Robertson et al., 2016). The lipophilic sterol metabolites produced by IFN signaling including oxysterols are substrates for Cyp enzymes and they profoundly influence cellular metabolism (Newmark et al., 2017) which in turn has been shown to influence *Mtb* replication inside cells (Huang et al., 2018). Since cytokine modulation of host-lipid metabolism seems to have a profound influence in mycobacterial persistance detailed studies to delineate the signaling mechanisms involved is recommended.

TB Granuloma and Host Lipid Metabolism

During disease progression in TB, granuloma is formed locally in the affected lungs. While a a solid granuloma is often associated with efficient containment of *Mtb*, a caseous granuloma is believed to favor rupture and dissemination of bacteria that likely results in clinical TB symptoms (Ehlers and Schaible, 2012). However, the underlying pathology at the site of infection that determines progression to active TB is yet to be fully estalished. Research focused in identifying the cellular mechanisms within TB granuloma have determined that host genes for lipid sequestration, synthesis, and metabolism are upregulated in caseous pulmonary TB granulomas (Kim et al., 2010). Moreover, biochemical analysis of these caseous granulomas identified LDL-derived host lipids such as cholesterol, cholesteryl esters, lactosylceramides, and TAG (Kim et al., 2010). In a different

perspective, it is also known that mycobacterial infection results in enhanced PPAR expression and resultant lipid droplet formation in the host macrophages (Almeida et al., 2012). This is believed to occur via the utilization of TAG resulting in the limiting of bacterial replication while promoting drug tolerance of Mtb inside foamy macrophages, which is a mechanism of disease perpetuation (Daniel et al., 2011). Moreover, Mtb is known to modulate the kinetics of cholesterol metabolism in host macrophages mainly by enhancing the expression of macrophage LXR (liver X receptor) genes, which eventually increases bacterial replication inside the foamy macrophages in granulomatous lesions (Vermeulen et al., 2017). In addition, the presence of an association between vitamin D receptor (VDR) and lipid metabolism in human tuberculosis and infected macrophages has also been reported (Salamon et al., 2014). When infected macrophages were treated with vitamin D, the accumulation of lipid droplets was abolished via a mechanism that involved VDR mediated downregulation of the proadipogenic PPARy in infected macrophages. Also administration of PPARy agonists reversed the antiadipogenic as well as the antimicrobial effects of VDR, thus establishing a role for vitamin D in macrophage lipid metabolism (Salamon et al., 2014).

Therefore, manipulation of the endogenous lipid mediators discussed above as well as cytokines involved in altering the lipid metabolic pathway of the host during mycobacterial infections has started to gain importance in studies addressing mycobacterial pathogenesis with the aim of identifying targets for vaccine development and drug discovery.

CARDIOVASCULAR DISEASE TO MYCOBACTERIAL INFECTION: RESEARCH AVENUES TO EXTRAPOLATE AND EXPLORE

Since macrophages are the primary cell type involved in foam cell formation in atherosclerosis and mycobacterial infections, the biogenesis of macrophage foam cell formation may likely be similar. While many of the receptors and enzymes involved in foam cell formation have been extensively researched in relation to CVD (Schumacher and Benndorf, 2017) their application in relation to mycobacterial pathogenicity is not yet fully understood.

Lipid Transporters, Cytokines, Lipid Mediators, and Foam Cell Research

Since mycobacteria are intracellular pathogens often infecting macrophages, the contribution of macrophage foam cell formation to mycobacterial pathogenesis has been addressed. However, there are several areas which could benefit from the application of relevant knowledge available in the CVD field. For example, the ABC transport proteins not only play a role in lipid transport in host macrophages but in *Mtb* a total of 26 complete ABC transporters have been cataloged with many of the ABC exporters potentially implicated in the transport of drugs, probably contributing to the resistance of *Mtb* to many antibiotics (Braibant et al., 2000). While the role of

mycobacterial ABC transport proteins and phospholipases has been explored (Braibant et al., 2000; Glass et al., 2017), the importance of host ABC proteins in mycobacterial diseases could be further explored. Recently Long et al. profiled the plasma membrane of macrophages infected with *Mtb* and reported an upregulation of ABCA1 which is involved in cholesterol efflux from the macrophages (Long et al., 2016). Similarly, few studies have reported on the importance of other transporters/receptors such as MARCO for induction of TNF- α , IL-1 β , IL-6, and other responses to mycobacterial trehalose dimycolate and *Mtb* (Benard et al., 2014; Thuong et al., 2016; Khan et al., 2017). These avenues could be explored further in the context of TB disease reactivation.

IL-6 is a key cytokine induced by Angiotensin-II which is involved in the development of several CVDs including atherosclerosis (Schieffer et al., 2004; Kokje et al., 2016; Akita et al., 2017). IL-6 causes an increase in macrophage uptake of oxLDL via increase in expression of scavenger receptors such as SR-A, lectin-like OxLDL receptor-I, and CD36 (Keidar et al., 2001; Hashizume and Mihara, 2012). Mycobacteria, as well as their cell wall components, have been established to induce IL-6 production by varying cell types including macrophages, dendritic cells, and platelets (Adams and Czuprynski, 1994; Champsi et al., 1995; Jang et al., 2004). Apart from systemic production of IL-6, its expression locally at the granulomatous lesion has been reported by Renshaw et al. who have confirmed that thrombocytosis associated with Mtb infection in the granuloma was accompanied by elevated levels of IL-6 (Renshaw and Gould, 2013). Moreover, mycobacteria infected murine macrophages reportedly produce 10,000-fold more IL-6 compared to uninfected controls (Vanheyningen et al., 1997). Therefore, it would be worthwhile to assess how IL-6 is associated with mycobacteria-induced foam cell formation and Mtb survival in a non-replicative state.

Among the lipid mediators, isoprostanes and isoketals produced by lipid peroxidation of AA have been studied in detail in CVD pathogenesis (Cracowski and Ormezzano, 2004; Roberts and Milne, 2009; Kirabo et al., 2014). F2 isoprostanes containing oxidized phospholipids rapidly adduct to the amino group of lysine residue of cell membrane proteins disrupting their structural and functional capacity resulting in cellular dysfunction (Brame et al., 2004; Sullivan et al., 2010). Isoketals in particular are highly cytotoxic and their production is enhanced by oxidative stress (Kirabo et al., 2014). Since the immune response to mycobacterial infection is characterized by an oxidative burst, it is likely that these agents are generated within the inflammatory milieu during infection which might influence the disease progression and outcome, however their importance in TB has not been explored. Isoketals increase binding and uptake of LDL (Sullivan et al., 2010) and thus also promote foam cell formation which is an important criterion promoting mycobacterial survival. Moreover, isoketals have a role in DC activation by crosslinking lysine residues on proteins, altering protein function causing them to become antigenic (Brame et al., 2004; Kirabo et al., 2014) which might be a source of persistant uncontrolled inflammation aggravating pulmonary tissue destruction resulting in fibrotic development (Mont et al., 2016).

Alternate Cell Types: Platelets

The importance of platelets in inflammation mediated diseases and disorders including atherosclerosis, arthritis, and cancer has now been established (Ferrer-Acosta et al., 2014; Papapanagiotou et al., 2016). Platelets have the capacity to facilitate inflammatory cell infiltration at the lesion site resulting in the release of a milieu of inflammatory mediators (Stokes and Granger, 2012). Moreover, in atherosclerosis LDL initiates a platelet-mediated signaling cascade and facilitates foam cell formation. Platelets initiate maturation of macrophages promoting internalization of oxLDL via scavenger receptors aiding in the formation of foam cells (Huo et al., 2003; Siegel-Axel et al., 2006; Von Hundelshausen and Weber, 2007; Von Hundelshausen and Schmitt, 2014). Interaction of platelets with monocytes results in an increase in the circulating population of CD14^{hi} CD16⁺ monocytes in humans (Passacquale et al., 2011; Tapp et al., 2011) and these are the subpopulations that generally transform to foam cells, as enzymatically degraded LDL preferentially binds to these cells via enhanced CD36 expression (Kapinsky et al., 2001). In a study by Huo et al. it was observed that injecting activated platelets resulted in the adhesion of monocytes to atherosclerotic lesions, leading to a reduction in the number of circulating monocytes, which suggests that platelet-monocyte adhesion plays a role in the formation of atherosclerotic lesions (Huo et al., 2003). Therefore, platelets are even being targeted for therapeutic development to reduce atherosclerosis and atherothrombosis (Lindemann et al., 2001, 2007).

One of the first studies assessing the direct involvement of platelets in bacterial pathogenesis was by Sullam et al. who reported through activation of the COX pathway and resultant production of thromboxane A2, platelets could directly kill *Streptococcus sanguis in vitro* (Sullam et al., 1993). Although the role of macrophages and T cells in protection against mycobacteria has been well-established (Ernst, 2012), the role of platelets in mycobacterial infections has lagged behind and the importance of these cells in relation to TB pathogenesis has gained priority only in the recent past (Lugo-Villarino and Neyrolles, 2014). Pulmonary TB due to *Mtb* infection is now associated with increased platelet infiltration and activity, coupled with enhanced platelet interaction with monocytes and T cells in granulomas, stressing a critical role for these cells in TB pathogenesis (Renshaw and Gould, 2013; Kullaya et al., 2018).

In atherosclerosis, apart from macrophages, other cell types including smooth muscle cells and endothelial cells lining the blood vessel can also transform into foam cells (Glukhova et al., 1987; Ivan and Antohe, 2010). Similarly, whether other cell types present locally at the site of granuloma formation eg. alveolar cells, airway smooth muscle cells (in *Mtb* infection) or intestinal smooth muscle cells (in *M. paratuberculosis* infection) could form foam cells and how that would drive the disease course has not been addressed. In this context the role of platelets in host lipid metabolism during mycobacterial infection was only recently addressed, wherein platelets were found to have the capacity to initiate the differentiation of monocytes into epithelioid-like multinucleated giant foam cells of the suppressive phenotype (Feng et al., 2014). However, other aspects of plateletmediated signaling cascades and associated alterations in lipid metabolism including the activation state of platelets, their interaction with monocyte subsets and T cells driving foam cell formation and/or antigen presentation and adaptive immunity during mycobacterial infections are avenues which could be further explored utilizing literature available in depth in this area in CVD research as guidelines.

Cytochrome P450 (Cyp) System

The Cyp system has been implicated in the pathogenesis of various CVDs including atherosclerosis and aneurysms (Song et al., 2016; Thirunavukkarasu et al., 2016b). The Cyp enzymes are heme containing monooxygenases involved in the pathways associated with the metabolism of fatty acids, steroids and other lipophilic molecules (Pikuleva, 2008). The loss of Cyp enzymes prevents Apolipoprotein (Apo) A1 synthesis thus affecting the removal of cholesterol from the cells to the liver for biliary excretion (Rubin et al., 1991; Nebert and Russell, 2002; Guan et al., 2003). The cholesterol metabolizing ability of Cyp family members varies depending on the physiological requirements of different organs and *in vivo* cholesterol levels. Since the activity of the Cyp enzymes can be modulated post-translationally, they are targets to manipulate cholesterol homeostasis (Luoma, 2008).

The importance of Cyp in mycobacteria is known. For example, *Mtb* has about 20 different Cyp enzymes, and several Cyp genes have been associated with mycobacterial viability (Mclean et al., 2010). Cole et al. have reported that the *Mtb* genome sequence has a very high number of Cyp enzymes, with Cyp128 contributing to synthesis of cell wall sulpholipid and being necessary for *in-vitro* growth of the bacterium (Cole et al., 1998; Holsclaw et al., 2008; Sogi et al., 2016). Iron is necessary for mycobacterial Cyp (Ouellet et al., 2002, 2011) activity and iron has been recognized as a crucial element for the survival of intracellular mycobacteria (Ratledge, 2004; Janagama et al., 2009; Lamont et al., 2013). Thus, the importance of Cyp from the pathogen perspective has been explored albeit not in depth.

From the host perspective, a transcriptomic analysis of TB granulomas has revealed a characteristic downregulation of several *Cyp* genes in cells within these granulomas (Eisenreich et al., 2013). Cyp monooxygenases are involved in biosynthesis of oxysterols (Pikuleva, 2006, 2008), however the dynamics between the expression of *Cyp* genes in granuloma formation and cholesterol metabolism has not been assessed in *Mtb* infection.

Cyp450 and Steroid Sex Hormones

Physiologically occurring steroid hormones which impact sexual characteristics are lipids that are synthesized from cholesterol in the gonads and adrenal glands. These sex steroid hormones have additional functions influencing metabolism, inflammation and immune responses. Apart from their direct role in modulating host cholesterol metabolism, members of the Cyp family such as Cyp4501b1(Cyp1b1) are involved in the metabolism of sex steroidal hormones with the resultant metabolites produced being implicated to play a role in the gender differences noticed to the susceptibility to cardiac and renal diseases (Jennings et al., 2014; Pingili et al., 2015, 2016; Song et al., 2016; Thirunavukkarasu et al., 2016b). While Cyp1b1 has been shown to have a protective role against cardiac and renal diseases in females, it aggravates the pathogenesis of these diseases in males, due to the action of this enzyme on estrogen and testosterone, respectively, as shown by ovariectomy and castration studies (Jennings et al., 2014; Pingili et al., 2016, 2017).

Sexual dimorphism in the relative susceptibility to Mtb and other mycobacterial infections has been reported, with the male to female ratio of susceptibility to Mtb infection reported to be 1.9/0.6 worldwide (Yamamoto et al., 1991; Nevrolles and Quintana-Murci, 2009; Karunasena et al., 2014; Nhamoyebonde and Leslie, 2014). Both in humans and animal models it has been shown that the male gender is more susceptible to TB than the female gender, and the differences in sex hormones could be a possible underlying mechanism for this difference (Svanberg, 1981; Tsuyuguchi et al., 2001; Ramsey et al., 2006; Dibbern et al., 2017). Intact female and castrated male mice have reduced severity of M. avium, M. marinum, and M. intracellulare infection, while ovariectomized females, or females and castrated males treated with testosterone had increased susceptibility to infection (Yamamoto et al., 1991; Tsuyuguchi et al., 2001). These findings have translational significance as in humans the death rate in castrated males due to TB was less compared to intact males and pre-menopausal females (Hamilton and Mestler, 1969). Moreover, there is almost a 2-fold increase in the severe lepromatous form of tuberculoid leprosy in males compared to females (Guerra-Silveira and Abad-Franch, 2013). These studies suggest that sex steroidal hormones might have a role in influencing the susceptibility to mycobacterial infections in mammals. Therefore, extrapolating from the evidence for Cyp1b1 in gender specific pathogeneis of CVDs, it would be worthwhile to explore further the role of Cyp family members in mycobacterial infections due to a necessity for a sex-tailored therapeutic approach in view of the gender related susceptibility to disease.

Technology Transfer

3D in vitro technology employing scaffolds made of biomaterials that could be engineered to simulate tissue degeneration or regeneration has revolutionized biomedical research in CVD medicine (Ou and Hosseinkhani, 2014). Such models could be employed in mycobacterial research as well, to assess, understand, and decipher novel mechanisms involved in the pathogenesis of these bacteria in a dynamic sense. Positive steps in this direction were initiated by devising a three dimensional in vitro granuloma mode utilizing PBMCs to study host-pathogen interactions, drug susceptibility as well as mycobacterial drug tolerance and persistence (Kapoor et al., 2013; Fitzgerald et al., 2014). Since foam cell formation is associated with Mtb survival during infection research in in vitro models of Mtb-infected foam cells is critical. Novel methods to generate and evaluate foam cells in vitro have been utilized in the CVD research field (Xu et al., 2010; Sengupta et al., 2013), and these techniques could be applied to assess the interaction of mycobacteria with foam cells under controlled conditions to gain a better understanding of the events that lead to persistence of the bacterium in a non-replicative state as reported by Santucci et al. (2016). Thus, incorporating the recent advances in *in vitro* technology employed in CVD research to study foam cell signaling in mycobacterial research should prove fruitful.

CONCLUSIONS

Decrease in cholesterol efflux, increase in uptake, and excess esterification of cholesterol compounded by the action of lipid peroxidation products generated by inflammatory processes contribute to foam cell formation in atherosclerosis. Since these are critical events in the pathogenesis of atherosclerosis, they have been extensively studied. On the contrary, although a role for host lipid metabolism in mycobacterial pathogenesis has been established, research in the different components that contribute to the alteration in host lipid metabolism and cholesterol homeostasis is still in the nascent stages. The specific cause for reactivation of mycobacterial disease from a controlled state in an infected host is still a critical question that remains inconclusive. It is speculated that the reversible lipid accumulation in foamy macrophages could be a possible phenomenon that is associated with disease reactivation (Caire-Brändli et al., 2014). Researchers attempting to decipher these loopholes in TB pathogenesis specifically targeting the host lipid pathway with the aim of developing novel therapeutics and vaccination strategies would be better served by utilizing the plethora of resources available in this area in the cardiovascular field (Kurth et al., 2004; Han et al., 2017; Rodriguez et al., 2017). The potential for PPAR regulators of airway inflammation as potential therapeutic targets for asthma and lung diseases (Standiford et al., 2005; Banno et al., 2018) has already been explored and their application for TB therapeutics could be further explored. Some studies have already reported on the application of statins and COX-2 inhibitors as adjuncts in the therapeutic strategy of mycobacterial infections which could be further exlpored in depth (Turull and Queralt, 2000; Brombacher et al., 2013; Lobato et al., 2014). Thus, extrapolation and application of relevant knowledge and techniques between these fields would prove to be beneficial and sustainable amidst an increasingly competitive research scenario beset by time and funding constraints.

AUTHOR CONTRIBUTIONS

ST performed the literature review and wrote the first draft of the manuscript. SK critically revised the paper. All authors read and approved the final version of the manuscript.

FUNDING

This work was supported by Washington University in St. Louis, NIH grants R01 AI134236, R01 AI111914, R01 HL105427, R01 AI123780.

ACKNOWLEDGMENTS

The authors would like to thank Ms. Nicole Howard for critical review of the manuscript.

REFERENCES

- Abuhammad, A. (2017). Cholesterol metabolism: a potential therapeutic target in Mycobacteria. Br. J. Pharmacol. 174, 2194–2208. doi: 10.1111/bph.13694
- Achkar, J. M., and Jenny-Avital, E. R. (2011). Incipient and subclinical tuberculosis: defining early disease states in the context of host immune response. J. Infect. Dis. 204 (Suppl. 4), S1179–S1186. doi: 10.1093/infdis/jir451
- Adams, J. L., and Czuprynski, C. J. (1994). Mycobacterial cell wall components induce the production of TNF-alpha, IL-1, and IL-6 by bovine monocytes and the murine macrophage cell line RAW 264.7. *Microb. Pathog.* 16, 401–411. doi: 10.1006/mpat.1994.1040
- Agard, M., Asakrah, S., and Morici, L. A. (2013). PGE(2) suppression of innate immunity during mucosal bacterial infection. *Front Cell. Infect. Microbiol.* 3, 45–45. doi: 10.3389/fcimb.2013.00045
- Akiba, S., Yoneda, Y., Ohno, S., Nemoto, M., and Sato, T. (2003). Oxidized LDL activates phospholipase A2 to supply fatty acids required for cholesterol esterification. J. Lipid Res. 44, 1676–1685. doi: 10.1194/jlr.M300012-JLR200
- Akita, K., Isoda, K., Sato-Okabayashi, Y., Kadoguchi, T., Kitamura, K., Ohtomo, F., et al. (2017). An interleukin-6 receptor antibody suppresses atherosclerosis in atherogenic mice. *Front. Cardiovasc. Med.* 4:84. doi: 10.3389/fcvm.2017.00084
- Almeida, P. E., Carneiro, A. B., Silva, A. R., and Bozza, P. T. (2012). PPAR gamma; expression and function in mycobacterial infection: roles in lipid metabolism, immunity, and bacterial killing. *PPAR Res.* 2012:383829. doi: 10.1155/2012/383829
- Almeida, P. E., Silva, A. R., Maya-Monteiro, C. M., Töröcsik, D., D[']Ávila, H., Dezsö, B., et al. (2009). *Mycobacterium bovis* bacillus calmette-guérin infection induces TLR2-dependent peroxisome proliferator-activated receptor γ expression and activation: functions in inflammation, lipid metabolism, and pathogenesis. J. Immunol. 183, 1337–1345. doi: 10.4049/jimmunol.0900365
- Bafica, A., Scanga, C. A., Serhan, C., Machado, F., White, S., Sher, A., et al. (2005). Host control of *Mycobacterium tuberculosis* is regulated by 5lipoxygenase-dependent lipoxin production. *J. Clin. Invest.* 115, 1601–1606. doi: 10.1172/JCI23949
- Bah, S. Y., Dickinson, P., Forster, T., Kampmann, B., and Ghazal, P. (2017). Immune oxysterols: role in mycobacterial infection and inflammation. J. Steroid Biochem. Mol. Biol. 169, 152–163. doi: 10.1016/j.jsbmb.2016.04.015
- Banno, A., Reddy, A. T., Lakshmi, S. P., and Reddy, R. C. (2018). PPARs: key regulators of airway inflammation and potential therapeutic targets in asthma. *Nuclear Receptor Res.* 5:101306. doi: 10.11131/2018/101306
- Benard, E. L., Roobol, S. J., Spaink, H. P., and Meijer, A. H. (2014). Phagocytosis of mycobacteria by zebrafish macrophages is dependent on the scavenger receptor Marco, a key control factor of pro-inflammatory signalling. *Dev. Comp. Immunol.* 47, 223–233. doi: 10.1016/j.dci.2014.07.022
- Bini, E. I., Mata Espinosa, D., Marquina Castillo, B., Barrios Payán, J., Colucci, D., Cruz, A. F., et al. (2014). The influence of sex steroid hormones in the immunopathology of experimental pulmonary tuberculosis. *PLoS ONE* 9:e93831. doi: 10.1371/journal.pone.0093831
- Blanc, M., Hsieh, W. Y., Robertson, K. A., Kropp, K. A., Forster, T., Shui, G., et al. (2013). The transcription factor STAT-1 couples macrophage synthesis of 25hydroxycholesterol to the interferon antiviral response. *Immunity* 38, 106–118. doi: 10.1016/j.immuni.2012.11.004
- Blanc, M., Hsieh, W. Y., Robertson, K. A., Watterson, S., Shui, G., Lacaze, P., et al. (2011). Host defense against viral infection involves interferon mediated down-regulation of sterol biosynthesis. *PLoS Biol.* 9:e1000598. doi: 10.1371/journal.pbio.1000598
- Braibant, M., Gilot, P., and Content, J. (2000). The ATP binding cassette (ABC) transport systems of *Mycobacterium tuberculosis*. *FEMS Microbiol Rev.* 24, 449–467. doi: 10.1111/j.1574-6976.2000.tb00550.x
- Brame, C. J., Boutaud, O., Davies, S. S., Yang, T., Oates, J. A., Roden, D., et al. (2004). Modification of proteins by isoketal-containing oxidized phospholipids. *J. Biol. Chem.* 279, 13447–13451. doi: 10.1074/jbc.M313349200
- Brombacher, F., Hurdayal, R., Guler, R., Parihar, S. P., Lang, D. M., Marais, A. D., et al. (2013). Statin therapy reduces the *Mycobacterium tuberculosis* burden in human macrophages and in mice by enhancing autophagy and phagosome maturation. *J. Infect. Dis.* 209, 754–763. doi: 10.1093/infdis/jit550
- Caire-Brändli, I., Papadopoulos, A., Malaga, W., Marais, D., Canaan, S., Thilo, L., et al. (2014). Reversible lipid accumulation and associated division arrest of *Mycobacterium avium* in lipoprotein-induced foamy macrophages may

resemble key events during latency and reactivation of tuberculosis. *Infect. Immun.* 82, 476–490. doi: 10.1128/IAI.01196-13

- Champsi, J., Young, L. S., and Bermudez, L. E. (1995). Production of TNF-alpha, IL-6 and TGF-beta, and expression of receptors for TNF-alpha and IL-6, during murine *Mycobacterium avium* infection. *Immunology* 84, 549–554.
- Chen, M., Divangahi, M., Gan, H., Shin, D. S., Hong, S., Lee, D. M., et al. (2008). Lipid mediators in innate immunity against tuberculosis: opposing roles of PGE2 and LXA4 in the induction of macrophage death. J. Exp. Med. 205, 2791–2801. doi: 10.1084/jem.20080767
- Chen, Y.-C., Huang Alex, L., Kyaw Tin, S., Bobik, A., and Peter, K. (2016). Atherosclerotic plaque rupture. *Arterioscler. Thromb. Vasc. Biol.* 36, e63–e72. doi: 10.1161/ATVBAHA.116.307993
- Chistiakov, D. A., Bobryshev, Y. V., and Orekhov, A. N. (2016). Macrophagemediated cholesterol handling in atherosclerosis. J. Cell. Mol. Med. 20, 17–28. doi: 10.1111/jcmm.12689
- Chistiakov, D. A., Melnichenko, A. A., Myasoedova, V. A., Grechko, A. V., and Orekhov, A. N. (2017). Mechanisms of foam cell formation in atherosclerosis. *J. Mol. Med.* 95, 1153–1165. doi: 10.1007/s00109-017-1575-8
- Chroni, A., Leondaritis, G., and Karlsson, H. (2011). Lipids and lipoproteins in atherosclerosis. *J. Lipids* 2011:160104. doi: 10.1155/2011/160104
- Cole, S. T., Brosch, R., Parkhill, J., Garnier, T., Churcher, C., Harris, D., et al. (1998). Deciphering the biology of *Mycobacterium tuberculosis* from the complete genome sequence. *Nature* 393, 537–544. doi: 10.1038/31159
- Cracowski, J. L., and Ormezzano, O. (2004). Isoprostanes, emerging biomarkers and potential mediators in cardiovascular diseases. *Eur. Heart J.* 25, 1675–1678. doi: 10.1016/j.ehj.2004.07.031
- Daniel, J., Maamar, H., Deb, C., Sirakova, T. D., and Kolattukudy, P. E. (2011). *Mycobacterium tuberculosis* uses host triacylglycerol to accumulate lipid droplets and acquires a dormancy-like phenotype in lipid-loaded macrophages. *PLOS Pathog*, 7:e1002093. doi: 10.1371/journal.ppat.1002093
- Dibbern, J., Eggers, L., and Schneider, B. E. (2017). Sex differences in the C57BL/6 model of *Mycobacterium tuberculosis* infection. *Sci. Rep.* 7:10957. doi: 10.1038/s41598-017-11438-z
- Dietzold, J., Gopalakrishnan, A., and Salgame, P. (2015). Duality of lipid mediators in host response against *Mycobacterium tuberculosis*: good cop, bad cop. *F1000Prime Rep.* 7:29. doi: 10.12703/P7-29
- Donovan, M. L., Schultz, T. E., Duke, T. J., and Blumenthal, A. (2017). Type I interferons in the pathogenesis of tuberculosis: molecular drivers and immunological consequences. *Front. Immunol.* 8:1633. doi: 10.3389/fimmu.2017.01633
- Drain, P. K., Bajema, K. L., Dowdy, D., Dheda, K., Naidoo, K., Schumacher, S. G., et al. (2018). Incipient and subclinical tuberculosis: a clinical review of early stages and progression of infection. *Clin. Microbiol. Rev.* 31:e00021-18. doi: 10.1128/CMR.00021-18
- Dye, C., and Williams, B. G. (2010). The population dynamics and control of tuberculosis. *Science* 328, 856–861. doi: 10.1126/science.1185449
- Ehlers, S., and Schaible, U. E. (2012). The granuloma in tuberculosis: dynamics of a host-pathogen collusion. *Front. Immunol.* 3:411. doi: 10.3389/fimmu.2012.00411
- Eisenreich, W., Heesemann, J., Rudel, T., and Goebel, W. (2013). Metabolic host responses to infection by intracellular bacterial pathogens. *Front. Cell. Infect. Microbiol.* 3:24. doi: 10.3389/fcimb.2013.00024
- Ernst, J. D. (2012). The immunological life cycle of tuberculosis. Nat. Rev. Immunol. 12, 581-591. doi: 10.1038/nri3259
- Falkinham, I. J. O. (2009). Surrounded by mycobacteria: nontuberculous mycobacteria in the human environment. J. Appl. Microbiol. 107, 356–367. doi: 10.1111/j.1365-2672.2009.04161.x
- Feng, Y., Dorhoi, A., Mollenkopf, H. J., Yin, H., Dong, Z., Mao, L., et al. (2014). Platelets direct monocyte differentiation into epithelioid-like multinucleated giant foam cells with suppressive capacity upon mycobacterial stimulation. J. Infect. Dis. 210, 1700–1710. doi: 10.1093/infdis/jiu355
- Fernández, M., Benavides, J., Castaño, P., Elguezabal, N., Fuertes, M., Muñoz, M., et al. (2016). Macrophage subsets within granulomatous intestinal lesions in bovine paratuberculosis. *Vet. Pathol.* 54, 82–93. doi: 10.1177/0300985816653794
- Ferrer-Acosta, Y., González, M., Fernández, M., and Valance, W. A. (2014). Emerging roles for platelets in inflammation and disease. J. Infect. Dis. Ther. 2:149. doi: 10.4172/2332-0877.1000149

- Fitzgerald, L. E., Abendaño, N., Juste, R. A., and Alonso-Hearn, M. (2014). Threedimensional *in vitro* models of granuloma to study bacteria-host interactions, drug-susceptibility, and resuscitation of dormant mycobacteria. *BioMed. Res. Int.* 2014:623856. doi: 10.1155/2014/623856
- Glass, L. N., Swapna, G., Chavadi, S. S., Tufariello, J. M., Mi, K., Drumm, J. E., et al. (2017). *Mycobacterium tuberculosis* universal stress protein Rv2623 interacts with the putative ATP binding cassette (ABC) transporter Rv1747 to regulate mycobacterial growth. *PLoS Pathog.* 13:e1006515. doi: 10.1371/journal.ppat.1006515
- Glukhova, M. A., Ornatsky, O. I., Frid, M. G., Kabakov, A. E., Adany, R. R., Muszbek, L., et al. (1987). Identification of smooth muscle-derived foam cells in the atherosclerotic plaque of human aorta with monoclonal antibody IIG10. *Tissue Cell* 19, 657–663. doi: 10.1016/0040-8166(87)90072-3
- Griffin, J. E., Pandey, A. K., Gilmore, S. A., Mizrahi, V., McKinney, J. D., Bertozzi, C. R., et al. (2012). Cholesterol catabolism by *Mycobacterium tuberculosis* requires transcriptional and metabolic adaptations. *Chem. Biol.* 19, 218–227. doi: 10.1016/j.chembiol.2011.12.016
- Guan, J. Z., Tamasawa, N., Murakami, H., Matsui, J., Yamato, K., and Suda, T. (2003). Clofibrate, a peroxisome-proliferator, enhances reverse cholesterol transport through cytochrome P450 activation and oxysterol generation. *Tohoku J. Exp. Med.* 201, 251–259. doi: 10.1620/tjem.201.251
- Guenin-Mace, L., Simeone, R., and Demangel, C. (2009). Lipids of pathogenic Mycobacteria: contributions to virulence and host immune suppression. *Transbound. Emerg. Dis.* 56, 255–268. doi: 10.1111/j.1865-1682.2009.01072.x
- Guerra-Silveira, F., and Abad-Franch, F. (2013). Sex bias in infectious disease epidemiology: patterns and processes. *PLoS ONE* 8:e62390. doi: 10.1371/journal.pone.0062390
- Guirado, E., and Schlesinger, L. S. (2013). Modeling the *Mycobacterium tuberculosis* granuloma the critical battlefield in host immunity and disease. *Front. Immunol.* 4:98. doi: 10.3389/fimmu.2013.00098
- Hamilton, J. B., and Mestler, G. E. (1969). Mortality and survival: comparison of eunuchs with intact men and women in a mentally retarded population. J. Gerontol. 24, 395–411. doi: 10.1093/geronj/24.4.395
- Han, L., Shen, W.-J., Bittner, S., Kraemer, F. B., and Azhar, S. (2017). PPARs: regulators of metabolism and as therapeutic targets in cardiovascular disease. Part II: PPAR- β/δ and PPAR- γ . *Future Cardiol.* 13, 279–296. doi: 10.2217/fca-2017-0019
- Hanna, V. S., and Hafez, E. A. A. (2018). Synopsis of arachidonic acid metabolism: a review. J. Adv. Res. 11, 23–32. doi: 10.1016/j.jare.2018.03.005
- Hashizume, M., and Mihara, M. (2012). Blockade of IL-6 and TNF-alpha inhibited oxLDL-induced production of MCP-1 via scavenger receptor induction. *Eur. J. Pharmacol.* 689, 249–254. doi: 10.1016/j.ejphar.2012.05.035
- Holsclaw, C. M., Sogi, K. M., Gilmore, S. A., Schelle, M. W., Leavell, M. D., Bertozzi, C. R., et al. (2008). Structural characterization of a novel sulfated menaquinone produced by stf3 from *Mycobacterium tuberculosis*. ACS Chem. Biol. 3, 619–624. doi: 10.1021/cb800145r
- Howard, N. C., Marin, N. D., Ahmed, M., Rosa, B. A., Martin, J., Bambouskova, M., et al. (2018). *Mycobacterium tuberculosis* carrying a rifampicin drug resistance mutation reprograms macrophage metabolism through cell wall lipid changes. *Nat. Microbiol.* 3, 1099–1108. doi: 10.1038/s41564-018-0245-0
- Huang, L., Nazarova, E. V., Tan, S., Liu, Y., and Russell, D. G. (2018). Growth of *Mycobacterium tuberculosis in vivo* segregates with host macrophage metabolism and ontogeny. J. Exp. Med. 215, 1–18. doi: 10.1084/jem.20172020
- Huo, Y., Schober, A., Forlow, S. B., Smith, D. F., Hyman, M. C., Jung, S., et al. (2003). Circulating activated platelets exacerbate atherosclerosis in mice deficient in apolipoprotein E. *Nat. Med.* 9, 61–67. doi: 10.1038/nm810
- Ivan, L., and Antohe, F. (2010). Hyperlipidemia induces endothelialderived foam cells in culture. J. Receptors Signal. Trans. 30, 106–114. doi: 10.3109/10799891003630606
- Jain, M., Petzold, C. J., Schelle, M. W., Leavell, M. D., Mougous, J. D., Bertozzi, C. R., et al. (2007). Lipidomics reveals control of *Mycobacterium tuberculosis* virulence lipids via metabolic coupling. *Proc. Natl. Acad. Sci. U.S.A.* 104, 5133–5138. doi: 10.1073/pnas.0610634104
- Janagama, H. K., Senthilkumar, T. M., Bannantine, J. P., Rodriguez, G. M., Smith, I., Paustian, M. L., et al. (2009). Identification and functional characterization of the iron-dependent regulator (IdeR) of *Mycobacterium avium* subsp. *paratuberculosis. Microbiology*. 155, 3683–3690. doi: 10.1099/mic.0.031948-0

- Jang, S., Uematsu, S., Akira, S., and Salgame, P. (2004). IL-6 and IL-10 induction from dendritic cells in response to *Mycobacterium tuberculosis* is predominantly dependent on TLR2-mediated recognition. *J. Immunol.* 173, 3392–3397. doi: 10.4049/jimmunol.173.5.3392
- Jennings, B. L., George, L. W., Pingili, A. K., Khan, N. S., Estes, A. M., Fang, X. R., et al. (2014). Estrogen metabolism by cytochrome P450 1B1 modulates the hypertensive effect of angiotensin II in female mice. *Hypertension* 64, 134–140. doi: 10.1161/HYPERTENSIONAHA.114.03275
- Kalinski, P. (2012). Regulation of immune responses by prostaglandin E2. J. Immunol. 188, 21–28. doi: 10.4049/jimmunol.1101029
- Kapinsky, M., Torzewski, M., Buchler, C., Duong, C. Q., Rothe, G., and Schmitz, G. (2001). Enzymatically degraded LDL preferentially binds to CD14(high) CD16(+) monocytes and induces foam cell formation mediated only in part by the class B scavenger-receptor CD36. Arterioscler. Thromb. Vasc. Biol. 21, 1004–1010. doi: 10.1161/01.ATV.21.6.1004
- Kapoor, N., Pawar, S., Sirakova, T. D., Deb, C., Warren, W. L., and Kolattukudy, P. E. (2013). Human granuloma *in vitro* model, for TB dormancy and resuscitation. *PLoS ONE* 8:e53657–e53657. doi: 10.1371/journal.pone.0053657
- Karunasena, E., Mcmahon, K. W., Chang, D., and Brashears, M. M. (2014). Host responses to the pathogen *Mycobacterium avium* subsp. *paratuberculosis* and beneficial microbes exhibit host sex specificity. *Appl. Environ. Microbiol.* 80, 4481–4490. doi: 10.1128/AEM.01229-14
- Keidar, S., Heinrich, R., Kaplan, M., Hayek, T., and Aviram, M. (2001). Angiotensin II administration to atherosclerotic mice increases macrophage uptake of oxidized ldl: a possible role for interleukin-6. Arterioscler. Thromb. Vasc. Biol. 21, 1464–1469. doi: 10.1161/hq0901.095547
- Khan, A., Mann, L., Papanna, R., Lyu, M. A., Singh, C. R., Olson, S., et al. (2017). Mesenchymal stem cells internalize *Mycobacterium tuberculosis* through scavenger receptors and restrict bacterial growth through autophagy. *Sci. Rep.* 7:15010. doi: 10.1038/s41598-017-15290-z
- Kim, M.-J., Wainwright, H. C., Locketz, M., Bekker, L.-G., Walther, G. B., Dittrich, C., et al. (2010). Caseation of human tuberculosis granulomas correlates with elevated host lipid metabolism. *EMBO Mol. Med.* 2, 258–274. doi: 10.1002/emmm.201000079
- Kirabo, A., Fontana, V., De Faria, A. P. C., Loperena, R., Galindo, C. L., Wu, J., et al. (2014). DC isoketal-modified proteins activate T cells and promote hypertension. *J. Clin. Invest.* 124, 4642–4656. doi: 10.1172/JCI 74084
- Knight, M., Braverman, J., Asfaha, K., Gronert, K., and Stanley, S. (2018). Lipid droplet formation in *Mycobacterium tuberculosis* infected macrophages requires IFN-γ/HIF-1α signaling and supports host defense. *PLoS Pathog.* 14:e1006874. doi: 10.1371/journal.ppat.1006874
- Kokje, V. B. C., Gäbel, G., Koole, D., Northoff, B. H., Holdt, L. M., Hamming, J. F., et al. (2016). IL-6: A Janus-like factor in abdominal aortic aneurysm disease. *Atherosclerosis.* 251, 139–146. doi: 10.1016/j.atherosclerosis.2016.06.021
- Kruth, H. S. (2001). Lipoprotein cholesterol and atherosclerosis. *Curr. Mol. Med.* 1, 633–653. doi: 10.2174/1566524013363212
- Kruth, S., H. (2013). Fluid-phase pinocytosis of LDL by macrophages: a novel target to reduce macrophage cholesterol accumulation in atherosclerotic lesions. *Curr.Pharm. Design* 19, 5865–5872. doi: 10.2174/1381612811319330005
- Kullaya, V., Van Der Ven, A., Mpagama, S., Mmbaga, B. T., De Groot, P., Kibiki, G., et al. (2018). Platelet-monocyte interaction in *Mycobacterium tuberculosis* infection. *Tuberculosis*. 111, 86–93. doi: 10.1016/j.tube.2018.05.002
- Kurth, T., Hennekens, C. H., Buring, J. E., and Gaziano, J. M. (2004). Aspirin, NSAIDs, and COX-2 inhibitors in cardiovascular disease: possible interactions and implications for treatment of rheumatoid arthritis. *Curr. Rheumatol. Rep.* 6, 351–356. doi: 10.1007/s11926-004-0009-0
- Lamont, E. A., Xu, W. W., and Sreevatsan, S. (2013). Host-Mycobacterium avium subsp. paratuberculosis interactome reveals a novel iron assimilation mechanism linked to nitric oxide stress during early infection. BMC Genomics 14, 1–22. doi: 10.1186/1471-2164-14-694
- Lee, K.-S., Dubey, V. S., Kolattukudy, P. E., Song, C.-H., Shin, A. R., Jung, S.-B., et al. (2007). Diacyltrehalose of *Mycobacterium tuberculosis* inhibits lipopolysaccharide- and mycobacteria-induced proinflammatory cytokine production in human monocytic cells. *FEMS Microbiol. Lett.* 267, 121–128. doi: 10.1111/j.1574-6968.2006.00553.x

- Lee, W., Vanderven, B. C., Fahey, R. J., and Russell, D. G. (2013). Intracellular Mycobacterium tuberculosis exploits host-derived fatty acids to limit metabolic stress. J. Biol. Chem. 288, 6788–6800. doi: 10.1074/jbc.M112.445056
- Leiva, E., Wehinger, S., Guzman, L., and Orrego, R. (2015). Role of oxidized LDL in atherosclerosis. *Open Access Peer Reviewed Chapter* [Online]. (accessed February 15, 2019).
- Lindemann, S., Krämer, B., Daub, K., Stellos, K., and Gawaz, M. (2007). Molecular pathways used by platelets to initiate and accelerate atherogenesis. *Curr. Opin. Lipidol.* 18, 566–573. doi: 10.1097/MOL.0b013e3282ef7c1e
- Lindemann, S., Tolley, N. D., Dixon, D. A., Mcintyre, T. M., Prescott, S. M., Zimmerman, G. A., et al. (2001). Activated platelets mediate inflammatory signaling by regulated interleukin 1β synthesis. *J. Cell Biol.* 154, 485–490. doi: 10.1083/jcb.200105058
- Linton, M. F., Yancey, P. G., and Davies, S. S. E. A. (eds.). (2015). The Role of Lipids and Lipoproteins in Atherosclerosis. S. Dartmouth, MA: MDText.com, Inc.
- Lobato, L. S., Rosa, P. S., Ferreira, J. D. S., Neumann, A. D. S., Da Silva, M. G., Do Nascimento, D. C., et al. (2014). Statins increase rifampin mycobactericidal effect. *Antimicrob. Agents Chemother.* 58, 5766–74. doi: 10.1128/AAC.01826-13
- Long, J., Basu Roy, R., Zhang, Y. J., Antrobus, R., Du, Y., Smith, D. L., et al. (2016). Plasma membrane profiling reveals upregulation of ABCA1 by infected macrophages leading to restriction of mycobacterial growth. *Front. Microbiol.* 7:1086. doi: 10.3389/fmicb.2016.01086
- Lugo-Villarino, G., and Neyrolles, O. (2014). Of clots and granulomas: platelets are new players in immunity to tuberculosis. J. Infect. Dis. 210, 1687–1690. doi: 10.1093/infdis/jiu356
- Luoma, P. V. (2008). Cytochrome P450 and gene activation—from pharmacology to cholesterol elimination and regression of atherosclerosis. *Eur. J. Clin. Pharmacol.* 64, 841–850. doi: 10.1007/s00228-008-0515-5
- Matsuura, E., Hughes, G. R., and Khamashta, M. A. (2008). Oxidation of LDL and its clinical implication. *Autoimmun. Rev.* 7, 558–566. doi: 10.1016/j.autrev.2008.04.018
- Mattos, K. A., Lara, F. A., Oliveira, V. G. C., Rodrigues, L. S., D'avila, H., Melo, R. C. N., et al. (2011). Modulation of lipid droplets by *Mycobacterium leprae* in Schwann cells: a putative mechanism for host lipid acquisition and bacterial survival in phagosomes. *Cell. Microbiol.* 13, 259–273. doi: 10.1111/j.1462-5822.2010.01533.x
- Mayer-Barber, K. D., Andrade, B. B., Oland, S. D., Amaral, E. P., Barber, D. L., Gonzales, J., et al. (2014). Host-directed therapy of tuberculosis based on interleukin-1 and type I interferon crosstalk. *Nature* 511, 99–103. doi: 10.1038/nature13489
- Mayer-Barber, K. D., and Yan, B. (2016). Clash of the Cytokine Titans: counterregulation of interleukin-1 and type I interferon-mediated inflammatory responses. *Cell. Mol. Immunol.* 14, 22–35. doi: 10.1038/cmi.2016.25
- Mcclean, C. M., and Tobin, D. M. (2016). Macrophage form, function, and phenotype in mycobacterial infection: lessons from tuberculosis and other diseases. *Pathog. Dis.* 74:ftw068-ftw068. doi: 10.1093/femspd/ftw068
- Mclean, K. J., Belcher, J., Driscoll, M. D., Fernandez, C. C., Le Van, D., Bui, S., et al. (2010). The *Mycobacterium tuberculosis* cytochromes P450: physiology, biochemistry & molecular intervention. *Future Med. Chem.* 2, 1339–1353. doi: 10.4155/fmc.10.216
- Mendum, T. A., Wu, H., Kierzek, A. M., and Stewart, G. R. (2015). Lipid metabolism and Type VII secretion systems dominate the genome scale virulence profile of *Mycobacterium tuberculosis* in human dendritic cells. *BMC Genomics* 16:372. doi: 10.1186/s12864-015-1569-2
- Mirsaeidi, M., and Sadikot, R. T. (2015). Gender susceptibility to mycobacterial infections in patients with non-CF bronchiectasis. *Int. J. Mycobacteriol.* 4, 92–96. doi: 10.1016/j.ijmyco.2015.05.002
- Mishra, B. B., and Sassetti, C. (2016). Neutrophilic inflammation promotes bacterial growth during Tuberculosis. J. Immunol. 196:63.
- Mont, S., Davies, S. S., Roberts Second, L. J., Mernaugh, R. L., Mcdonald, W. H., Segal, B. H., et al. (2016). Accumulation of isolevuglandin-modified protein in normal and fibrotic lung. *Sci. Rep.* 6:24919. doi: 10.1038/srep24919
- Nazarova, E. V., Montague, C. R., La, T., Wilburn, K. M., Sukumar, N., Lee, W., et al. (2017). Rv3723/LucA coordinates fatty acid and cholesterol uptake in *Mycobacterium tuberculosis. eLife* 6:e26969. doi: 10.7554/eLife.26969
- Nebert, D. W., and Russell, D. W. (2002). Clinical importance of the cytochromes P450. *Lancet* 360, 1155–1162. doi: 10.1016/S0140-6736(02)11203-7

- Newmark, H., Dantoft, W., and Ghazal, P. (2017). Evolutionary origin of the interferon-immune metabolic axis: the sterol-vitamin D link. *Front. Immunol.* 8:62. doi: 10.3389/fimmu.2017.00062
- Neyrolles, O. (2014). Mycobacteria and the greasy macrophage: getting fat and frustrated. *Infect. Immun.* 82:472. doi: 10.1128/IAI.01512-13
- Neyrolles, O., and Quintana-Murci, L. (2009). Sexual inequality in tuberculosis. *PLoS Med.* 6:199. doi: 10.1371/journal.pmed.1000199
- Nhamoyebonde, S., and Leslie, A. (2014). Biological differences between the sexes and susceptibility to tuberculosis. J. Infect. Dis. 209 (Suppl. 3), S100–106. doi: 10.1093/infdis/jiu147
- Ou, K.-L., and Hosseinkhani, H. (2014). Development of 3D in vitro technology for medical applications. Int. J. Mol. Sci. 15, 17938–17962. doi: 10.3390/ijms151017938
- Ouellet, H., Johnston, J. B., and De Montellano, P. R. (2011). Cholesterol catabolism as a therapeutic target in *Mycobacterium tuberculosis*. *Trends Microbiol*. 19, 530–539. doi: 10.1016/j.tim.2011.07.009
- Ouellet, H., Ouellet, Y., Richard, C., Labarre, M., Wittenberg, B., Wittenberg, J., et al. (2002). Truncated hemoglobin HbN protects *Mycobacterium bovis* from nitric oxide. *Proc. Natl. Acad. Sci. U.S.A.* 99, 5902–5907. doi: 10.1073/pnas.092017799
- Pandey, A. K., and Sassetti, C. M. (2008). Mycobacterial persistence requires the utilization of host cholesterol. *Proc. Natl. Acad. Sci. U.S.A.* 105, 4376–4380. doi: 10.1073/pnas.0711159105
- Papapanagiotou, A., Daskalakis, G., Siasos, G., Gargalionis, A., and Papavassiliou, A. G. (2016). The role of platelets in cardiovascular disease: molecular mechanisms. *Curr. Pharm. Des.* 22, 4493–4505. doi: 10.2174/1381612822666160607064118
- Parthasarathy, S., Raghavamenon, A., Garelnabi, M. O., and Santanam, N. (2010). Oxidized low-density lipoprotein. *Methods Mol. Biol.* 610, 403–417. doi: 10.1007/978-1-60327-029-8_24
- Passacquale, G., Vamadevan, P., Pereira, L., Hamid, C., Corrigall, V., and Ferro, A. (2011). Monocyte-platelet interaction induces a proinflammatory phenotype in circulating monocytes. *PLoS ONE* 6:e25595. doi: 10.1371/journal.pone.0025595
- Peyron, P., Vaubourgeix, J., Poquet, Y., Levillain, F., Botanch, C., Bardou, F., et al. (2008). Foamy macrophages from tuberculous patients' granulomas constitute a nutrient-rich reservoir for *M. tuberculosis* persistence. *PLoS Pathog.* 4:e1000204. doi: 10.1371/journal.ppat.1000204
- Philips, J. A., and Ernst, J. D. (2012). Tuberculosis pathogenesis and immunity. Annu. Rev. Pathol. 7, 353–384. doi: 10.1146/annurev-pathol-011811-132458
- Pieters, J. (2008). *Mycobacterium tuberculosis* and the macrophage: maintaining a balance. *Cell Host Microbe* 3, 399–407. doi: 10.1016/j.chom.2008.05.006
- Pikuleva, I. A. (2006). Cholesterol-metabolizing cytochromes P450. Drug Metab. Disposition 34, 513–520. doi: 10.1124/dmd.105.008789
- Pikuleva, I. A. (2008). Cholesterol-metabolizing cytochromes P450: implications for cholesterol lowering. *Expert Opin. Drug Metab. Toxicol.* 4, 1403–1414. doi: 10.1517/17425255.4.11.1403
- Pingili, A. K., Davidge, K. N., Thirunavukkarasu, S., Khan, N. S., Katsurada, A., Majid, D. S. A., et al. (2017). 2-Methoxyestradiol reduces angiotensin II-induced hypertension and renal dysfunction in ovariectomized female and intact male mice. *Hypertension* 69, 1104–1112. doi: 10.1161/HYPERTENSIONAHA.117.09175
- Pingili, A. K., Kara, M., Khan, N. S., Estes, A. M., Lin, Z., Li, W., et al. (2015). 6beta-hydroxytestosterone, a cytochrome P450 1B1 metabolite of testosterone, contributes to angiotensin II-induced hypertension and its pathogenesis in male mice. *Hypertension* 65, 1279–1287. doi: 10.1161/HYPERTENSIONAHA.115.05396
- Pingili, A. K., Thirunavukkarasu, S., Kara, M., Brand, D. D., Katsurada, A., Majid, D. S., et al. (2016). 6beta-hydroxytestosterone, a cytochrome P450 1B1-testosterone-metabolite, mediates angiotensin ii-induced renal dysfunction in male mice. *Hypertension* 67, 916–926. doi: 10.1161/HYPERTENSIONAHA.115.06936
- Ramsey, D. S., Coleman, J. D., Coleman, M. C., and Horton, P. (2006). The effect of fertility control on the transmission of bovine tuberculosis in wild brushtail possums. N. Z. Vet. J. 54, 218–223. doi: 10.1080/00480169.2006.36700
- Ratledge, C. (2004). Iron, mycobacteria and tuberculosis. *Tuberculosis* 84, 110–130. doi: 10.1016/j.tube.2003.08.012

- Remmerie, A., and Scott, C. L. (2018). Macrophages and lipid metabolism. Cell. Immunol. 330, 27–42. doi: 10.1016/j.cellimm.2018.01.020
- Renshaw, A. A., and Gould, E. W. (2013). Thrombocytosis is associated with *Mycobacterium tuberculosis* infection and positive acid-fast stains in granulomas. *Am. J. Clin. Pathol.* 139, 584–586. doi: 10.1309/AJCPCM1CKASVBMBP
- Roberts, L. J., and Milne, G. L. (2009). Isoprostanes. J. Lipid Res. 50, S219–S223. doi: 10.1194/jlr.R800037-JLR200
- Robertson, K. A., Hsieh, W. Y., Forster, T., Blanc, M., Lu, H., Crick, P. J., et al. (2016). An interferon regulated MicroRNA provides broad cell-intrinsic antiviral immunity through multihit host-directed targeting of the sterol pathway. *PLoS Biol.* 14:e1002364. doi: 10.1371/journal.pbio.1002364
- Rocca, B., and Fitzgerald, G. A. (2002). Cyclooxygenases and prostaglandins: shaping up the immune response. *Int. Immunopharmacol.* 2, 603–630. doi: 10.1016/S1567-5769(01)00204-1
- Rodriguez, F., Maron, D. J., Knowles, J. W., Virani, S. S., Lin, S., and Heidenreich, P. A. (2017). Association between intensity of statin therapy and mortality in patients with atherosclerotic cardiovascular disease. *JAMA Cardiol.* 2, 47–54. doi: 10.1001/jamacardio.2016.4052
- Rogers, M. A., Liu, J., Song, B.-L., Li, B.-L., Chang, C. C. Y., and Chang, T.-Y. (2015). Acyl-CoA:cholesterol acyltransferases (ACATs/SOATs): enzymes with multiple sterols as substrates and as activators. J. Steroid Biochem. Mol. Biol. 151, 102–107. doi: 10.1016/j.jsbmb.2014.09.008
- Rosenson, R. S., Brewer, H. B. Jr., Chapman, M. J., Fazio, S., Hussain, M. M., Kontush, A., et al. (2011). HDL measures, particle heterogeneity, proposed nomenclature, and relation to atherosclerotic cardiovascular events. *Clin. Chem.* 57, 392–410. doi: 10.1373/clinchem.2010.155333
- Rothe, T., Gruber, F., Uderhardt, S., Ipseiz, N., Rössner, S., Oskolkova, O., et al. (2015). 12/15-lipoxygenase-mediated enzymatic lipid oxidation regulates DC maturation and function. J. Clin. Invest. 125, 1944–1954. doi: 10.1172/JCI78490
- Rubin, E. M., Krauss, R. M., Spangler, E. A., Verstuyft, J. G., and Clift, S. M. (1991). Inhibition of early atherogenesis in transgenic mice by human apolipoprotein AI. *Nature* 353, 265–267. doi: 10.1038/353265a0
- Russell, D. G., Cardona, P.-J., Kim, M.-J., Allain, S., and Altare, F. (2009). Foamy macrophages and the progression of the human TB granuloma. *Nat. Immunol.* 10, 943–948. doi: 10.1038/ni.1781
- Russell, D. G., Vanderven, B. C., Lee, W., Abramovitch, R. B., Kim, M.-J., Homolka, S., et al. (2010). *Mycobacterium tuberculosis* wears what it eats. *Cell Host Microbe* 8, 68–76. doi: 10.1016/j.chom.2010.06.002
- Salamon, H., Bruiners, N., Lakehal, K., Shi, L., Ravi, J., Yamaguchi, K. D., et al. (2014). Cutting edge: vitamin D regulates lipid metabolism in *Mycobacterium tuberculosis* infection. *J. Immunol.* 193, 30–34. doi: 10.4049/jimmunol.14 00736
- Santucci, P., Bouzid, F., Smichi, N., Poncin, I., Kremer, L., De Chastellier, C., et al. (2016). Experimental models of foamy macrophages and approaches for dissecting the mechanisms of lipid accumulation and consumption during dormancy and reactivation of tuberculosis. *Front. Cell. Infect. Microbiol.* 6:122. doi: 10.3389/fcimb.2016.00122
- Schieffer, B., Selle, T., Hilfiker, A., Hilfiker-Kleiner, D., Grote, K., Tietge, U. J. F., et al. and (2004). Impact of interleukin-6 on plaque development and morphology in experimental atherosclerosis. *Circulation* 110, 3493–3500. doi: 10.1161/01.CIR.0000148135.08582.97
- Schumacher, T., and Benndorf, R. A. (2017). ABC transport proteins in cardiovascular disease-a brief summary. *Molecules* 22:589. doi: 10.3390/molecules22040589
- Sengupta, B., Narasimhulu, C. A., and Parthasarathy, S. (2013). Novel technique for generating macrophage foam cells for *in vitro* reverse cholesterol transport studies. J. Lipid Res. 54, 3358–3372. doi: 10.1194/jlr.M041327
- Siegel-Axel, D., Langer, H., Lindemann, S., and Gawaz, M. (2006). Role of platelets in atherosclerosis and inflammation. *Med. Klin.* 101, 467–475. doi: 10.1007/s00063-006-1066-0
- Silva Miranda, M., Breiman, A., Allain, S., Deknuydt, F., and Altare, F. (2012). The tuberculous granuloma: an unsuccessful host defence mechanism providing a safety shelter for the bacteria? *Clin. Dev. Immunol.* 2012:139127. doi: 10.1155/2012/139127
- Silvestre-Roig, C., De Winther, M. P., Weber, C., Daemen, M. J., Lutgens, E., and Soehnlein, O. (2014). Atherosclerotic plaque destabilization. *Circ. Res.* 114, 214–26. doi: 10.1161/CIRCRESAHA.114.302355

- Sogi, K. M., Holsclaw, C. M., Fragiadakis, G. K., Nomura, D. K., Leary, J. A., and Bertozzi, C. R. (2016). Biosynthesis and regulation of sulfomenaquinone, a metabolite associated with virulence in *Mycobacterium tuberculosis. ACS Infect. Dis.* 2, 800–806. doi: 10.1021/acsinfecdis.6b00106
- Song, C. Y., Ghafoor, K., Ghafoor, H. U., Khan, N. S., Thirunavukkarasu, S., Jennings, B. L., et al. (2016). Cytochrome P450 1B1 contributes to the development of atherosclerosis and hypertension in apolipoprotein E-deficient mice. *Hypertension* 67, 206–213. doi: 10.1161/HYPERTENSIONAHA.115.06427
- Standiford, T. J., Keshamouni, V. G., and Reddy, R. C. (2005). Peroxisome proliferator-activated receptor-{gamma} as a regulator of lung inflammation and repair. Proc. Am. Thorac. Soc. 2, 226–231. doi: 10.1513/pats.200501-010AC
- Stefanadis, C., Antoniou, C. K., Tsiachris, D., and Pietri, P. (2017). Coronary atherosclerotic vulnerable plaque: current perspectives. J. Am. Heart Assoc. 6:e005543. doi: 10.1161/JAHA.117.005543
- Stokes, K. Y., and Granger, D. N. (2012). Platelets: a critical link between inflammation and microvascular dysfunction. J. Physiol. 590, 1023–1034. doi: 10.1113/jphysiol.2011.225417
- Sullam, P. M., Frank, U., Yeaman, M. R., Tauber, M. G., Bayer, A. S., and Chambers, H. F. (1993). Effect of thrombocytopenia on the early course of streptococcal endocarditis. J. Infect. Dis. 168, 910–914. doi: 10.1093/infdis/168.4.910
- Sullivan, C. B., Matafonova, E., Roberts, L. J., Amarnath, V., and Davies, S. S. (2010). Isoketals form cytotoxic phosphatidylethanolamine adducts in cells. *J. Lipid Res.* 51, 999–1009. doi: 10.1194/jlr.M001040
- Svanberg, L. (1981). Effects of estrogen deficiency in women castrated when young. Acta Obstet. Gynecol. Scand. Suppl. 106, 11–15.
- Swiecki, M., and Colonna, M. (2015). The multifaceted biology of plasmacytoid dendritic cells. Nat. Rev. Immunol. 15, 471–485. doi: 10.1038/nri3865
- Szondy, Z., Garabuczi, E., Joos, G., Tsay, G. J., and Sarang, Z. (2014). Impaired clearance of apoptotic cells in chronic inflammatory diseases: therapeutic implications. *Front. Immunol.* 5:354. doi: 10.3389/fimmu.2014.00354
- Tambo, A., Roshan, M. H. K., and Pace, N. P. (2016). Testosterone and cardiovascular disease. Open Cardiovasc. Med. J. 10, 1–10. doi: 10.2174/1874192401610010001
- Tangirala, R. K., Bischoff, E. D., Joseph, S. B., Wagner, B. L., Walczak, R., Laffitte, B. A., et al. (2002). Identification of macrophage liver X receptors as inhibitors of atherosclerosis. *Proc. Natl. Acad. Sci. U.S.A.* 99, 11896–11901. doi: 10.1073/pnas.182199799
- Tapp, L., D., Shantsila, E., Wrigley, B., J., Pamukcu, B., and Lip, G. Y. H. (2011). The CD14++CD16+ monocyte subset and monocyte-platelet interactions in patients with ST-elevation myocardial infarction. *J. Thromb. Haemostasis* 10, 1231–1241. doi: 10.1111/j.1538-7836.2011.04603.x
- Thirunavukkarasu, S., De Silva, K., Begg, D. J., Whittington, R. J., and Plain, K. M. (2015). Macrophage polarization in cattle experimentally exposed to *Mycobacterium avium* subsp. *paratuberculosis. Pathog. Dis.* 73:ftv085. doi: 10.1093/femspd/ftv085.
- Thirunavukkarasu, S., De Silva, K., Plain, K. M., and Whittington, R. (2016a). Role of host- and pathogen-associated lipids in directing the immune response in mycobacterial infections, with emphasis on *Mycobacterium avium* subsp. *paratuberculosis. Crit. Rev. Microbiol.* 42, 262–275. doi: 10.3109/1040841X.2014.932327
- Thirunavukkarasu, S., Khan, N. S., Song, C. Y., Ghafoor, H. U., Brand, D. D., Gonzalez, F. J., et al. (2016b). Cytochrome P450 1B1 contributes to the development of angiotensin II-induced aortic aneurysm in male ApoE^{-/-} mice. Am. J. Pathol. 186, 2204–2219. doi: 10.1016/j.ajpath.2016.04.005
- Thirunavukkarasu, S., Plain, K. M., De Silva, K., Begg, D., Whittington, R. J., and Purdie, A. C. (2014). Expression of genes associated with cholesterol and lipid metabolism identified as a novel pathway in the early pathogenesis of *Mycobacterium avium* subspecies *paratuberculosis*-infection in cattle. *Vet. Immunol. Immunopathol.* 160, 147–157. doi: 10.1016/j.vetimm.2014.04.002
- Thirunavukkarasu, S., Plain, K. M., De Silva, K., Marais, B. J., and Whittington, R. J. (2017a). Applying the one health concept to mycobacterial research – overcoming parochialism. *Zoonoses and Public Health* 64, 401–422. doi: 10.1111/zph.12334
- Thirunavukkarasu, S., Plain, K. M., Eckstein, T. M., De Silva, K., and Whittington, R. J. (2013). Cellular and humoral immunogenicity of *Mycobacterium avium* subsp. *paratuberculosis* specific lipopentapeptide antigens. *Res. Vet. Sci.* 95, 123–129. doi: 10.1016/j.rvsc.2013.03.002

- Thirunavukkarasu, S., Plain, K. M., Purdie, A. C., Whittington, R. J., and De Silva, K. (2017b). IFN- γ fails to overcome inhibition of selected macrophage activation events in response to pathogenic mycobacteria. *PLoS ONE* 12, e0176400. doi: 10.1371/journal.pone.0176400
- Thuong, N. T., Tram, T. T., Dinh, T. D., Thai, P. V., Heemskerk, D., Bang, N. D., et al. (2016). MARCO variants are associated with phagocytosis, pulmonary tuberculosis susceptibility and Beijing lineage. *Genes Immun.* 17, 419–425. doi: 10.1038/gene.2016.43
- Tobin, D. M., Roca, F. J., Oh, S. F., Mcfarland, R., Vickery, T. W., Ray, J. P., et al. (2012). Host genotype-specific therapies can optimize the inflammatory response to mycobacterial infections. *Cell* 148, 434–446. doi: 10.1016/j.cell.2011.12.023
- Tobin, D. M., Vary, J. C., Ray, J. P., Walsh, G. S., Dunstan, S. J., Bang, N. D., et al. (2010). The lta4h Locus modulates susceptibility to mycobacterial infection in zebrafish and humans. *Cell* 140, 717–730. doi: 10.1016/j.cell.2010.02.013
- Tsuyuguchi, K., Suzuki, K., Matsumoto, H., Tanaka, E., Amitani, R., and Kuze, F. (2001). Effect of oestrogen on *Mycobacterium avium* complex pulmonary infection in mice. *Clin. Exp. Immunol.* 123, 428–434. doi: 10.1046/j.1365-2249.2001.01474.x
- Turull, A., and Queralt, J. (2000). Selective cyclooxygenase-2 (COX-2) inhibitors reduce anti-Mycobacterium antibodies in adjuvant arthritic rats. *Immunopharmacology* 46, 71–77. doi: 10.1016/S0162-3109(99)00159-9
- Vanderven, B. C., Fahey, R. J., Lee, W., Liu, Y., Abramovitch, R. B., Memmott, C., et al. (2015). Novel inhibitors of cholesterol degradation in *Mycobacterium tuberculosis* reveal how the bacterium's metabolism is constrained by the intracellular environment. *PLoS Pathog.* 11:e1004679. doi: 10.1371/journal.ppat.1004679
- Vanheyningen, T. K., Collins, H. L., and Russell, D. G. (1997). IL-6 produced by macrophages infected with Mycobacterium species suppresses T cell responses. *J. Immunol.* 158, 330–337.
- Vermeulen, I., Baird, M., Al-Dulayymi, J., Smet, M., Verschoor, J., and Grooten, J. (2017). Mycolates of *Mycobacterium tuberculosis* modulate the flow of cholesterol for bacillary proliferation in murine macrophages. J. Lipid Res. 58, 709–718. doi: 10.1194/jlr.M073171
- Von Hundelshausen, P., and Schmitt, M. M. N. (2014). Platelets and their chemokines in atherosclerosis—clinical applications. *Front. Physiol.* 5:294. doi: 10.3389/fphys.2014.00294
- Von Hundelshausen, P., and Weber, C. (2007). Platelets as immune cells: bridging inflammation and cardiovascular disease. *Circ. Res.* 100, 27–40. doi: 10.1161/01.RES.0000252802.25497.b7
- Wangoo, A., Johnson, L., Gough, J., Ackbar, R., Inglut, S., Hicks, D., et al. (2005). Advanced granulomatouslesions in *Mycobacterium bovis*-infected cattle are

associated with increased expression of type I procollagen, gammadelta (WC1+) T cells and CD 68+ cells. *J. Comp. Pathol.* 133, 223–234. doi: 10.1016/j.jcpa.2005.05.001

- Whittington, R. J., Begg, D. J., De Silva, K., Plain, K. M., and Purdie, A. C. (2012). Comparative immunological and microbiological aspects of paratuberculosis as a model mycobacterial infection. *Vet. Immunol. Immunopathol.* 148, 29–47. doi: 10.1016/j.vetimm.2011.03.003
- Wilburn, K. M., Fieweger, R. A., and Vanderven, B. C. (2018). Cholesterol and fatty acids grease the wheels of *Mycobacterium tuberculosis* pathogenesis. *Pathog. Dis.* 76, 1–13. doi: 10.1093/femspd/fty021
- Wu, D., Sanin, D. E., Everts, B., Chen, Q., Qiu, J., Buck, M. D., et al. (2016). Type 1 interferons induce changes in core metabolism that are critical for immune function. *Immunity* 44, 1325–1336. doi: 10.1016/j.immuni.2016. 06.006
- Wu, T.-S., Chiu, C.-H., Yang, C.-H., Leu, H.-S., Huang, C.-T., Chen, Y.-C., et al. (2012). Fish tank granuloma caused by *Mycobacterium marinum*. *PLoS ONE* 7:e41296. doi: 10.1371/journal.pone.0041296
- Xu, S., Huang, Y., Xie, Y., Lan, T., Le, K., Chen, J., et al. (2010). Evaluation of foam cell formation in cultured macrophages: an improved method with Oil Red O staining and DiI-oxLDL uptake. *Cytotechnology* 62, 473–481. doi: 10.1007/s10616-010-9290-0
- Yamamoto, Y., Saito, H., Setogawa, T., and Tomioka, H. (1991). Sex differences in host resistance to *Mycobacterium marinum* infection in mice. *Infect. Immun.* 59, 4089–4096.
- Yeates, K., Lohfeld, L., Sleeth, J., Morales, F., Rajkotia, Y., and Ogedegbe, O. (2015). A global perspective on cardiovascular disease in vulnerable populations. *Can. J. Cardiol.* 31, 1081–1093. doi: 10.1016/j.cjca.2015.06.035
- York, A. G., Williams, K. J., Liu, X., Argus, J. P., Tong, A.-J., Zhou, Q. D., et al. (2016). Rapid transcriptional reprogramming of the lipid biosynthesis genes in response to type I interferon. *J. Immunol.* 196, 202–214.

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Thirunavukkarasu and Khader. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.