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Supplemental information

Virus-encoded histone doublets are essential and form nucleosome-like structures

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Table S1. Primers used in the MV histone knockout generation cloning and analysis. Related to Figure 2.

Primer ID	Purpose	Sequence
HB112	5' homology arm for recombination 368	tctcactaagacatatgacagcgtgcctccaaggt
HB113	5' homology arm for recombination 368	gcccttgctcaccatgacaaaaccgcggatggga
HB114	3' homology arm for recombination 368	cgcgtggtacctctaaggccgccgagctcatgc
HB115	3' homology arm for recombination 368	ccgggtcgactctagagttggattgaagttttccagtcg
HB120	5' homology arm for recombination 369	tctcactaagacatacgagctgcatcgcaagaa
HB121	5' homology arm for recombination 369	gcccttgctcaccatagatttgatggtttagttccctct
HB122	3' homology arm for recombination 369	ttcacgcgtggtaccaggaccttgaaggcacgc
HB123	3' homology arm for recombination 369	ccgggtcgactctagattagtatccggggctcattcc
HB145	Histone 368 KO genotyping	cgttttcctcagcaggaggaag
HB146	Histone 368 KO genotyping	ctggtgacggccttggtc
HB147	Histone 369 KO genotyping	atgcatgagctcggcggcc
HB148	Histone 369 KO genotyping	cagaggacgatgcttgtgtctcgtc
HB155	pEF1 (Nourseo) for transcomplementation	taatctagagtcgacccggaac
HB156	pEF1 (Nourseo) for transcomplementation	atgggtggattggtggtgttg
HB157	pEF1-mel368 (Nourseo) for transcomplementation	caccaatccacccatatgtcaaaagcaggcaaaaagg
HB158	pEF1-mel368 (Nourseo) for transcomplementation	gtcgactctagattattatagcggacatgttcagc
HB159	pEF1-mel369 (Nourseo) for transcomplementation	caccaatccacccatatggcgacacaaaaggaaaccac
HB160	pEF1-mel369 (Nourseo) for transcomplementation	gtcgactctagattattagtatccggggctcattccttg
GFP Fo	Histone KO genotyping	aggtgaacttcaagatccgc
GFP Rev	Histone KO genotyping	gaacttgtagccgtttacgtc

Table S2. Analysis of AFM data. Samples were imaged by AFM with heights (nm) of nucleosome like particles (NLP) recorded separately from free DNA. Average heights and standard deviations were calculated from the number of incidences (n). In addition to the average height of the NLP, the amount of free DNA (as a percent of particles on the slide) is a good indication of the stability of MV-NLP. When not prepared by GraFix, NLP heights generally correspond with heights of the MV-Tetrasome (p-values from unpaired t-test), whereas GraFix preparation yields heights in agreement with the eNuc₁₄₇ system. Related to Figure 3.

	MV-H4-H3-DNA (MV-tetrasome)		eNuc ₁₄₇		MV-NLP ₁₄₇		MV-NLP ₂₀₇		MV-NLP ₂₀₇ GraFix	
Particle Class	Free DNA	MV-Tetrasome	Free DNA	eNuc	Free DNA	MV-NLP	Free DNA	MV-NLP	DNA	MV-NLP
Average Height (nm)	0.60	1.35	0.61	2.09	0.50	1.21	0.78	1.55	0.64	2.25
Heights σ (nm)	0.07	0.33	0.21	0.52	0.12	0.91	0.08	0.40	0.19	0.63
n	25	26	2	41	29	46	41	37	31	120
% Particle.	43.9	45.6	4.3	87.2	32.6	51.7	50.6	45.7	20.0	77.4
p-value (MV-Tet)	---	---	---	< 0.0001	---	0.4528	---	0.0403	---	< 0.0001
p-value (eNuc ₁₄₇)	---	< 0.0001	---	---	---	< 0.0001	---	< 0.0001	---	0.1452

Table S3. Summary of cryoEM data collection and refinement. Related to Figure 4.

	MV-NLP ₁₄₇ (EMD-24238) (PDB: 7N8N)	MV-NLP ₂₀₇
Data collection and processing		
Magnification	64,000	29,000
Voltage (kV)	300	200
Electron exposure (e-/Å ²)	50	80
Defocus range (μm)	0.8-2.0	1.0-2.5
Pixel size (Å)	1.065	1.219
Symmetry imposed	C1	C1
Initial particle images (no.)	2,648,493	2,131,563
Final particle images (no.)	34,418	377,702
Map resolution (Å)	3.95	6.1
FSC threshold	0.143	0.143
Map resolution range (Å)	3.9-6.9	6.0-10.0
Refinement		
Initial model used (PDB code)	1AOI, 3LZO	
Model resolution (Å)	4.1	
FSC threshold	0.143	
Model resolution range (Å)	4.1-7.3	
Map sharpening <i>B</i> factor (Å ²)		
Model composition		
Non-hydrogen atoms	11125	
Protein residues	789	
Nucleotide	250	
Ligands	0	
<i>B</i> factors (Å ²)		
Protein	n/a	
Nucleotide	n/a	
Ligand	n/a	
R.m.s. deviations		
Bond lengths (Å)	0.006	
Bond angles (°)	0.840	
Validation		
MolProbity score	2.38	
Clashscore	22.42	
Poor rotamers (%)	0.00	
Ramachandran plot		
Favored (%)	90.38	
Allowed (%)	9.62	
Disallowed (%)	0.00	

Table S4: Comparison of MV-NLP₁₄₇ with eNuc₁₄₇. “Global” root-mean-squared deviation (RMSD) values represent changes in positioning throughout the nucleosome-like complex and are calculated for each domain after least-squares fitting the cores of the MV-NLP₁₄₇ and eNuc₁₄₇ complexes, whereas “local” RMSDs are values calculated when least-squares fitting individual domains prior to calculation and thereby represent the degree of internal re-ordering of the domain. Distances are calculated as the separation of geometric centers of backbone positions (C, C_α, and N atoms), except for the H2A L1 Loops “closest distance”, which defines the smallest distance between heavy atoms in the two moieties. The H2B α_C orientation angle is calculated around the pseudo-dihedral defined in Figure S5A. Related to Figure 5.

RMSD Values (Å)	Global	Local
Particle Core	2.7	---
DNA (central 120 bp)	3.1	3.1
Octamer Fold	2.3	2.3
H4-H3 Tetramer	2.1	1.2
H3 α _N -helix (Chain A)	2.3	0.3
H3 α _N -helix (Chain E)	2.2	0.3
H2B-H2A Dimers (both)	2.5	2.1
H2B-H2A Dimer (Chain C,D)	2.5	1.6
H2B-H2A Dimer (Chain G,H)	2.5	1.6
H2B α _C -helix (chain D)	2.6	0.5
H2B α _C -helix (chain H)	3.4	0.9
H2A Docking Domain (Chain C)	8.5	3.9
H2A Docking Domain (Chain G)	6.2	3.3
Separation Distances (Å)	eNuc₁₄₇	MV-NLP₁₄₇
H4-H3 Dimers	32.6	33.3
H3-H3' 4HB Interface	10.1	10.4
H2B-H2A Dimers	36.3	38.6
H2B α _C -helices	49.1	52.2
H2A L1 Loops (centers)	7.2	9.7
H2A L1 Loops (closest distance)	2.9	5.3
Orientation Angle (°)	eNuc₁₄₇	MV-NLP₁₄₇
H2B α ₂ -helices	135.4	143.9