



# Establishment of Pediatric Reference Intervals for Routine Laboratory Tests in Korean Population: A Retrospective Multicenter Analysis

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**Background:** Reference intervals defined for adults or children of other ethnicities cannot be applied in the evaluation of Korean pediatric patients. Pediatric reference intervals are difficult to establish because children are in their growing stage and their physiology changes continuously. We aimed to establish reference intervals for routine laboratory tests for Korean pediatric patients through retrospective multicenter data analysis.

**Methods:** Preoperative laboratory test results from 1,031 pediatric patients aged 0 month–18 years who underwent minor surgeries in four university hospitals were collected. Age- and sex-specific reference intervals for routine laboratory tests were defined based on the Clinical and Laboratory Standards Institute (CLSI) EP28-A3c guidelines.

**Results:** The pediatric reference intervals determined in this study were different from existing adult reference intervals and pediatric reference intervals for other ethnicities. Most tests required age-specific partitioning, and some of those required sex-specific partitioning for at least one age-partitioned subgroup. Erythrocyte sedimentation rate, monocyte percentage, basophil percentage, activated partial thromboplastin time, glucose, cholesterol, albumin, bilirubin, chloride, and C-reactive protein did not show any difference between age- or sex-partitioned subgroups.

**Conclusions:** We determined Korean pediatric reference intervals for hematology, coagulation, and chemistry tests by indirect sampling based on medical record data from multiple institutions. These reference intervals would be valuable for clinical evaluations in the Korean pediatric population.

**Key Words:** Reference intervals, Indirect sampling, Korean pediatrics, Age-partitioned groups, Sex-partitioned groups, Multicenter

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## INTRODUCTION

Reliable reference intervals for laboratory tests are critical guides

for clinicians to diagnose diseases or make medical decisions [1]. The Clinical and Laboratory Standards Institute (CLSI) EP28-A3c guidelines describe procedures for establishing refer-

ence intervals in a clinical laboratory [2]. Although reference intervals established for a specific population using a specific platform can be transferred to other laboratories after appropriate verification, intervals for adults cannot be reliably adopted for the interpretation of test results of pediatric patients. Pediatric reference intervals should reflect not only age and sex, but also the ongoing growth and development of children [2-5]. Efforts have been made to establish reliable pediatric reference intervals, including studies from the Canadian Laboratory Initiative in Pediatric Reference Intervals (CALIPER) [6-8], German Health Interview and Examination Survey for Children and Adolescents [9, 10], and Children's Health Improvement through Laboratory Diagnostics in the USA [11, 12]. However, some ethnic group studies revealed that ethnicity influences certain test values, such as alanine aminotransferase (ALT) and total protein concentrations [8], and ferritin, total thyroxine, cobalamin, and 25-OH vitamin D in East Asians [13]; thus, reference intervals established for Caucasian populations cannot simply be extrapolated to Asian populations. Another prospective analysis revealed a significant effect of ethnicity on biochemical markers, including 25-OH vitamin D, ferritin, follicle-stimulating hormone, and immunoglobulin M [14]. Therefore, ethnic-specific reference intervals should be determined for certain biochemical markers. There are many previous efforts and some published studies about Korean pediatric reference intervals for biochemical markers including hematology, chemistry, and hormone [15-18]. However, these studies have limitations to assess dynamic age-dependent change in pediatric population. For instance, neither neonates nor infants were included in the reference population [15, 16], test samples taken at specific time point from children who underwent 1-year-old check-up, not from whole pediatric period were analyzed [17], or age group between neonate and adult was classified as 'children' without additional subdivision [18]. Korean pediatric reference intervals that cover whole age range have not been established to date.

Establishing pediatric reference intervals is challenging. Especially, collecting a sufficient number of blood samples from healthy children for reference interval establishment and validation are problematic. A small sample size renders the application of simple, nonparametric statistical methods less reliable and requires the use of complicated analytical methods [3, 5, 19, 20]. The CLSI EP28-A3c guidelines suggest conducting a multicenter trial to overcome these problems [2]. Data from multiple centers can be pooled to establish reference intervals when the comparability of the test methods is ensured.

Currently, pediatric reference intervals are rarely provided

when laboratory test results are reported in Korea, and when provided, they are usually based on studies in other ethnic populations. Moreover, existing reference intervals were established using old measurement methods, and instruments and are not applicable to current methods or instruments used in laboratories. Therefore, we aimed to establish reliable age- and sex-specific Korean pediatric reference intervals for hematology, coagulation, and chemistry tests.

## MATERIALS AND METHODS

### Data collection

Four university hospitals, i.e., Seoul National University Hospital (SNUH, Seoul), Seoul National University Bundang Hospital (SNUBH, Seongnam), Asan Medical Center (AMC, Seoul), and Kangnam Sacred Heart Hospital (KSHH, Seoul), Korea, participated in this retrospective study. Patients aged 0 month–18 years who visited any of the hospitals between June 2010 and August 2015 were selected for potential inclusion in this study. No analytical drift was noted in any of the hospitals during the period of data collection, including alteration in instrument or reagent, and a notable internal quality control issue.

In accordance with CLSI EP28-A3c guidelines, to obtain data from "presumed healthy" individuals [2, 21], patients who underwent routine preoperative laboratory evaluation for minor surgery in the departments of general surgery, orthopedics, plastic surgery, thoracic surgery, ophthalmology, otorhinolaryngology, urology, dermatology, and dental surgery were considered as the "healthy" pediatric population. Preoperative laboratory test results of these patients were collected as reference values, following the indirect sampling method [2]. Basic demographic and laboratory data were extracted from the medical records to exclude patients with "unhealthy" general conditions or any underlying diseases. Patients with the following conditions were considered "unhealthy" and were therefore excluded: those taking prescribed medicines, those with any other comorbidity, those with febrile disease within two weeks before blood sampling for laboratory tests, those whose planned surgery was canceled after assessment of the laboratory test results, and those with other diseases requiring treatment at the time of blood sampling. As bone-specific alkaline phosphatase is elevated when bone injuries occur [22, 23], ALP values from orthopedic patients were considered inappropriate for the present study and were excluded.

The patients included in the final analysis largely had mild diseases unlikely to affect the laboratory test results. Only results

from samples that passed the acceptance criteria of each hospital for each test, including volume, and were verified without warning or additional comment about hemolytic, icteric, or lipemic interferences, were included.

Referring to previous studies [8, 24] and considering the size of the reference group, the patients were divided into six subgroups by age: 0–3 months, 4–11 months, 1–6 years, 7–12 years, 13–15 years, and 16–18 years, and two subgroups by sex, to establish age- and sex-partitioned pediatric reference intervals. Finally, we extracted data for a total of 1,031 Korean pediatric patients from the four hospitals, including 401, 302, 228, and 100 patients from SNUH, AMC, SNUBH, and KSHH, respectively. The size of the aforementioned age-partitioned six subgroups was 44 (22 boys, 22 girls), 258 (140 boys, 118 girls), 415 (231 boys, 184 girls), 141 (76 boys, 65 girls), 101 (49 boys, 52 girls), and 72 (31 boys, 41 girls), respectively (Supplemental Data Table S1).

The study protocol was approved by the Institutional Review Board of each participating hospital. Informed consent was waived as this was a minimal risk study using only the test results stored in a database.

### Laboratory tests and analytical methods

Preoperative routine laboratory test results were used for the reference interval analysis. The laboratory tests included hematology: white blood cell count with absolute neutrophil count, percentage of neutrophils, lymphocytes, monocytes, eosinophils, and basophils; red blood cell (RBC) count with mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH), and mean corpuscular hemoglobin concentration (MCHC), hemoglobin (Hb), hematocrit (Hct), RBC distribution width (RDW) and erythrocyte sedimentation rate (ESR); platelet count with mean platelet volume (MPV), plateletcrit (PCT) and platelet distribution width (PDW); coagulation: prothrombin time (PT), activated partial thromboplastin time (aPTT), and fibrinogen; chemistry: calcium (Ca), phosphorus (P), blood urea nitrogen (BUN), creatinine (Cr), protein, albumin, uric acid, aspartate aminotransferase (AST), ALT, ALP, bilirubin, glucose, cholesterol, sodium (Na), potassium (K), chloride (Cl), total CO<sub>2</sub> (TCO<sub>2</sub>), and C-reactive protein (CRP).

For hematology tests, Sysmex XE-2100 (Sysmex Corporation, Kobe, Japan) was used in SNUH, SNUBH, and AMC, and Advia2120 (Siemens, Washington, DC, USA) in KSHH. For coagulation tests, ACL TOP (Instrumentation Laboratory, Bedford, MA, USA) was used in SNUH and KSHH, Sysmex CA-7000 (Sysmex Corporation, Kobe, Japan) in AMC, and STA-R MAX (Diagnos-

tica Stago Inc., Asnières, France) in SNUBH. For chemistry tests, TBA-c16000 (Toshiba, Tokyo, Japan) was used in SNUH, AU 5800 (Beckman Coulter, Brea, CA, USA) in SNUBH, COBAS 8000 (Roche Diagnostic International Ltd., Rotkreuz ZG, Switzerland) in AMC, and Hitachi 7600 (Hitachi, Tokyo, Japan) in KSHH.

### Comparison of test methods

We conducted a test method comparison and bias estimation to ensure comparability of the data from the different hospitals using diverse platforms and analytical methods, and to evaluate the possibility of establishing common reference intervals applicable to multiple analytical platforms. Fifty-one pooled serum samples were prepared in SNUH using residual serum stored at 4°C for three days, with 2-mL aliquots from each sample set distributed to the participating hospitals, and subsequently evaluated at the day of receipt for chemistry tests for Ca, P, BUN, Cr, protein, albumin, uric acid, AST, ALT, ALP, total bilirubin, glucose, cholesterol, Na, K, Cl, and TCO<sub>2</sub>. We compared only chemistry tests because of the difficulty in controlling the many factors that are to be considered when preparing pooled samples to compare hematology and coagulation test methods, such as blood type and isoagglutinin, unexpected antibody, and the condition of the plasma in which the cells were incubated. The hematology and coagulation tests from diverse platforms were indirectly compared by reviewing external quality assessment result, and analyzing the results using Mann-Whitney U test; however, this did not provide evidence enough to justify data combining. Equivalence of the chemistry test results was evaluated by Deming regression: when the 95% confidence interval (CI) of the slope contained 1 and that of the intercept contained 0, and reference material for each analyte was available in the Joint Committee for Traceability in Laboratory Medicine (JCTLM) database [25], the data from each hospital were regarded valid for reference interval determination and were merged for analysis. Chemistry tests results obtained from a single instrument, TBA-c16000, and from multiple equivalent platforms were analyzed to determine platform-specific and common reference intervals, respectively, whereas results obtained at SNUH using Sysmex XE-2100 and ACL TOP were used to determine platform-specific reference intervals for hematology and coagulation tests.

### Data analysis and reference interval determination

The data were analyzed as recommended by the CLSI EP28-A3c guidelines [2]. The approach to establish reference inter-

vals was similar to that used in the CALIPER study [8]. First, distribution tables and scatter plots were generated to globally assess the data. When the raw data were not normally distributed, they were subjected to logarithmic or Box-Cox transformation, and then, outliers for each test, confirmed by applying the Tukey test twice, were removed [26]. Partitioning of the reference interval according to age and sex was evaluated by Harris and Boyd's test, using the mean, standard deviation (SD), and sample size of the subgroups [1]. If no significant difference was found between the partitioned reference intervals of adjacent age or sex subgroups, these subgroups were merged and the reference intervals for the merged group were re-assessed against those for the adjacent subgroup using the Harris and Boyd's test, until no mergeable subgroups remained. When the sample size of the subgroup was  $\geq 120$ , we defined the 2.5th and 97.5th percentiles as the lower and upper reference limits, respectively, according to the nonparametric method, and the

90% CI of the limit was determined. When the sample size was  $< 120$ , the robust method reported by Horn, *et al.* [27] was used to set the reference intervals and calculate the 90% CI, except for a few tests, in which this method is inadequate to calculate the reference lower limit. The parametric method, which defines the range of mean  $\pm 2$ SD as reference interval, was applied for these tests after Shapiro-Wilk normality test. Statistical analyses were performed using Excel 2013 (Microsoft, Redmond, WA, USA), IBM SPSS Statistics 23 (IBM, Armonk, NY, USA), EP Evaluator 11 (Data Innovations, Burlington, VT, USA), and MedCalc version 14.8.1 (MedCalc Software, Ostend, Belgium).

## RESULTS

### Hematology

Details of the reference intervals for hematology testing using

**Table 1.** Age- and sex-specific pediatric reference intervals for hematology testing using the Sysmex XE-2100 system

Analyte (unit)	Total N	Age	Sex	N	Lower limit (90% CI)	Upper limit (90% CI)
WBC ( $\times 10^9/L$ )	385	0–3 mo	M	17	6.48 (4.74–8.10)	16.30 (14.52–17.68)
			F	17	5.78 (4.12–7.66)	15.80 (14.05–17.00)
		4–11 mo	Both	67	4.64 (3.78–5.42)	13.85 (12.98–14.77)
		1–6 yr	Both	138	4.85 (4.62–5.60)	11.50 (11.09–11.90)
		7–18 yr	Both	146	4.32 (2.72–4.62)	10.22 (9.57–10.20)
ANC ( $\times 10^9/L$ )	383	0–11 mo	Both	97	0.65* (N/A)	3.35* (N/A)
		1–18 yr	Both	286	1.40 (1.22–1.51)	5.76 (5.42–5.96)
Segmented neutrophils (%)	393	0–11 mo	Both	101	3.3 (1.1–5.5)	33.1 (30.1–35.6)
		1–6 yr	Both	147	16.9 (12.8–19.0)	62.2 (56.3–65.6)
		7–12 yr	Both	46	28.4 (25.1–32.2)	64.5 (60.5–68.3)
		13–18 yr	Both	99	38.3 (36.4–40.3)	67.7 (65.6–69.8)
Lymphocytes (%)	391	0–11 mo	Both	100	54.5 (52.0–57.6)	87.6 (85.2–89.8)
		1–6 yr	Both	147	26.9 (24.9–35.1)	73.8 (72.6–77.5)
		7–12 yr	Both	45	29.4 (26.2–32.8)	59.3 (56.3–62.0)
		13–18 yr	Both	99	21.5 (19.3–23.6)	51.5 (49.5–53.6)
Monocytes (%)	387	All	Both	387	3.5 (3.0–3.7)	10.4 (10.0–10.7)
Eosinophils (%)	375	All	M	181	0.3 (0.1–0.7)	6.9 (6.4–7.3)
			F	194	0.4 (0.1–0.7)	5.7 (4.9–5.9)
Basophils (%)	380	All	Both	380	0.1 (0.1–0.1)	0.8 (0.8–0.9)
RBC ( $\times 10^6/\mu L$ )	390	0–3 mo	Both	32	3.22 (3.07–3.41)	4.65 (4.44–4.82)
		4 mo–18 yr	M	174	4.16 (4.09–4.29)	5.49 (5.30–5.62)
			F	184	4.03 (3.86–4.10)	5.16 (5.03–5.29)
MCV (fL)	395	0–3 mo	Both	35	73.9 (71.2–76.6)	98.3 (95.2–100.9)
		4–11 mo	M	34	68.6 (66.9–70.6)	82.8 (81.0–84.5)
			F	35	70.8 (69.1–72.5)	84.6 (82.8–86.2)
		1–6 yr	M	73	72.3 (71.5–73.2)	83.4 (82.5–84.3)
			F	72	73.9 (72.8–75.0)	86.1 (85.2–87.0)

(Continued to the next page)

Table 1. Continued

Analyte (unit)	Total N	Age	Sex	N	Lower limit (90% CI)	Upper limit (90% CI)
		7–12 yr	M	22	74.7 (73.1–76.6)	87.8 (85.7–89.8)
			F	26	78.3 (77.0–79.9)	89.0 (87.6–90.3)
		13–15 yr	M	27	76.4 (74.3–78.7)	93.3 (91.1–95.3)
			F	35	80.5 (79.0–82.1)	94.3 (92.6–96.0)
		16–18 yr	M	18	80.3 (78.3–82.6)	95.2 (92.4–97.8)
			F	18	83.4 (81.6–85.6)	95.9 (93.9–97.5)
MCH (pg)	390	0–3 mo	Both	35	25.3 (24.4–26.4)	33.7 (32.7–34.6)
			M	32	23.5 (22.8–24.2)	28.6 (28.0–29.2)
		4–11 mo	F	31	24.9 (24.5–25.4)	28.2 (27.9–28.5)
			Both	146	24.6 (24.3–25.2)	29.1 (28.8–29.3)
		7–12 yr	Both	48	26.4 (26.0–26.7)	30.4 (29.9–30.8)
		13–15 yr	Both	62	26.4 (25.8–26.9)	32.2 (31.7–32.7)
		16–18 yr	Both	36	28.0 (27.5–28.6)	32.3 (31.8–32.8)
MCHC (g/L)	386	0–12 yr	Both	298	327 (325–329)	359 (357–364)
			M	44	325 (322–328)	357 (354–360)
		F	44	323 (321–326)	343 (341–345)	
Hb (g/L)	393	0–3 mo	Both	33	97 (92–101)	134 (128–140)
			Both	214	108 (107–111)	140 (138–143)
		4 mo–6 yr	M	23	117 <sup>†</sup> (112–123)	154 <sup>†</sup> (148–159)
			F	25	115 (110–120)	148 (143–152)
		7–12 yr	M	27	124 (119–129)	164 (158–169)
			F	34	118 (114–122)	150 (146–152)
		13–15 yr	M	27	124 (119–129)	164 (158–169)
			F	34	118 (114–122)	150 (146–152)
16–18 yr	M	18	138 <sup>†</sup> (132–143)	170 <sup>†</sup> (164–176)		
	F	19	119 <sup>†</sup> (114–123)	146 <sup>†</sup> (141–151)		
Hct (proportion of 1.0)	380	0–3 mo	Both	27	0.31 (0.30–0.31)	0.36 (0.36–0.37)
			Both	211	0.33 (0.32–0.33)	0.40 (0.39–0.41)
		4 mo–6 yr	Both	48	0.34 (0.34–0.35)	0.43 (0.42–0.44)
			M	27	0.36 (0.34–0.37)	0.48 (0.46–0.50)
		7–12 yr	F	34	0.36 (0.35–0.37)	0.44 (0.43–0.45)
			M	18	0.40 (0.39–0.42)	0.50 (0.49–0.52)
		F	15	0.36 <sup>†</sup> (0.35–0.37)	0.42 <sup>†</sup> (0.41–0.43)	
RDW (%)	390	0–3 mo	Both	35	11.7 (11.2–12.2)	16.3 (15.7–16.8)
			M	105	12.0 (11.8–12.1)	14.4 (14.2–14.6)
		4 mo–6 yr	F	106	11.7 (11.5–11.9)	14.1 (13.9–14.3)
			Both	144	11.7 (11.3–11.9)	13.8 (13.6–13.8)
7–18 yr	Both	144	11.7 (11.3–11.9)	13.8 (13.6–13.8)		
	Both	225	2 (2–2)	9 (8–9)		
ESR (mm/hr)	225	All	Both	225	2 (2–2)	9 (8–9)
Platelets ( $\times 10^9/L$ )	390	0–3 mo	Both	35	188 (143–243)	610 (558–663)
			Both	65	199 (174–224)	495 (467–520)
		4–11 mo	Both	192	178 (171–192)	414 (389–434)
			Both	98	165 (151–179)	365 (351–379)
1–12 yr	Both	192	178 (171–192)	414 (389–434)		
	Both	98	165 (151–179)	365 (351–379)		
13–18 yr	Both	98	165 (151–179)	365 (351–379)		
	Both	98	165 (151–179)	365 (351–379)		
PCT (%)	390	0–11 mo	Both	102	0.17 (0.15–0.19)	0.52 (0.49–0.54)
			Both	288	0.17 (0.15–0.18)	0.38 (0.36–0.39)
MPV (fL)	373	0–6 yr	Both	236	7.6 (7.5–8.0)	10.9 (10.5–11.1)
			Both	137	8.8 (8.6–9.1)	11.4 (11.1–11.8)
7–18 yr	Both	137	8.8 (8.6–9.1)	11.4 (11.1–11.8)		
	Both	137	8.8 (8.6–9.1)	11.4 (11.1–11.8)		
PDW (%)	358	All	Both	358	8.4 (8.1–8.6)	13.3 (12.9–13.7)

\*Although the sample size was <120, the values presented were established using a nonparametric method because the 90% CI of the reference lower limit calculated using the robust method contained a negative value and the data did not pass a normality test; <sup>†</sup>The 90% CI was not determined using the robust method; however, the data passed a normality test and a parametric method based on the normal distribution was applied.

Abbreviations: CI, confidence interval; N/A, not applicable; yr, year; mo, month; M, male; F, female; ANC, absolute neutrophil count; MCH, mean corpuscular hemoglobin; MCHC, mean corpuscular hemoglobin concentration; RDW, red cell distribution width; PCT, plateletcrit; MPV, mean platelet volume; PDW, platelet distribution width; WBC, white blood cells; RBC, red blood cells; MCV, mean corpuscular volume; ESR, erythrocyte sedimentation rate; Hct, hematocrit.



the Sysmex XE-2100 are shown in Table 1. Most hematology tests showed significantly different results between two or more adjacent age-partitioned subgroups. Especially, values for children younger than 3 months or 1 year tended to differ from those of older children. Sex-partitioned reference intervals were established for some tests in at least one age-partitioned subgroup. A few tests, including ESR, monocyte percentage, basophil percentage, and PDW, did not show any significant difference between age- or sex-partitioned subgroups; thus, all results for these tests were combined to establish common pediatric reference intervals.

### Coagulation

Reference intervals for coagulation tests established on the basis of test results obtained at SNUH using ACL TOP are shown Table 2. In the coagulation tests, we observed age-specific differences between children younger and older than one year for PT and younger and older than 13 years for fibrinogen concentration, whereas aPTT did not show age-specific differences. None of the coagulation tests showed sex-specific differences.

### Chemistry

Reference intervals for chemistry tests established using TBA-c16000 as well as from the various platforms are shown in Table 3. This table also presents pediatric reference intervals established in previous studies, including a single-center study in a Korean population established using Hitachi 7600 [15] and the CALIPER study using Abbott Architect c8000 [8]. Test items for which the multicenter results were equivalent to those obtained using the TBA-c16000 instrument in the method comparison test were combined to establish multicenter reference intervals.

For some test items, the comparison based on Deming regression revealed adequate equivalence (Supplemental Data Table S2) to results obtained from at least one of the other platforms, AU 5800, Cobas 8000, Hitachi 7600, and reference material was available in the JCTLM database (Supplemental Data Table S3); therefore, data obtained for these items were mergeable with those obtained using the TBA-c16000 instrument. Results of Ca testing on Cobas 8000 and Hitachi 7600, glucose testing on Cobas 8000, cholesterol testing on Hitachi 7600, Na testing on all instruments, and K and Cl testing on Cobas 8000 and AU 5800 were used to establish reference intervals (Table 3). For most chemistry tests, age-partitioned reference intervals were established. Glucose, cholesterol, albumin, bilirubin, Cl, and CRP did not show any difference between age-partitioned subgroups. For P, uric acid, protein, ALP, AST, and Cr, sex-specific reference intervals were established for at least one age-partitioned subgroup.

Hematology, coagulation, and chemistry tests that showed age-dependent changes are presented as scatter plots in Figs. 1, 2, and 3, respectively. In hematology, segmented neutrophil percentage, hemoglobin, and hematocrit tended to increase with age (Fig. 1B, 1I, and 1J), whereas WBC count, lymphocyte percentage, and platelets tended to decrease with age (Fig. 1A, 1C, and 1L). For chemistry, pediatric reference intervals established previously based mainly on a Caucasian population [8] are indicated as horizontal dashed lines in Fig. 3. Minor differences in reference intervals between ethnicities are indicated in Table 3; however, these differences were too small to be visible in Fig. 3. In Fig. 3, all mergeable multicenter data are plotted. BUN, uric acid, and Cr tended to increase with age (Fig. 3C, 3I, and 3D), whereas P, and K tended to decrease with age (Fig.

**Table 2.** Age-specific pediatric reference intervals for coagulation testing using the ACL TOP system

Analyte (unit)	Total N	Age	Sex	N	Lower limit (90% CI)	Upper limit (90% CI)
PT INR	389	0–11 mo	Both	102	0.87 (0.85–0.89)	1.14 (1.12–1.16)
		1–18 yr	Both	287	0.93 (0.91–0.94)	1.17 (1.15–1.19)
PT (%)	392	0–11 mo	Both	100	82 (79–84)	117 (114–120)
		1–18 yr	Both	292	78 (73–81)	111 (109–114)
PT (s)	387	0–11 mo	Both	100	9.6 (9.4–9.8)	12.3 (12.1–12.5)
		1–18 yr	Both	287	10.1 (10.0–10.3)	12.8 (12.6–13.1)
aPTT (s)	383	All	Both	383	28.3 (27.5–29.0)	41.3 (40.8–41.7)
Fibrinogen (g/dL)	67	0–12 yr	Both	56	1.46 (1.31–1.63)	3.21 (3.04–3.35)
		13–18 yr	Both	11	1.96* (1.54–2.37)	3.79* (3.38–4.21)

\*The 90% CI was not determined using a non-parametric method or robust method; however, the data passed a normality test and a parametric method based on the normal distribution was applied.

Abbreviations: PT, prothrombin time; INR, international normalized ratio; aPTT, activated partial thromboplastin time; yr, year; mo, month.

**Table 3.** Age- and sex-specific pediatric reference intervals for chemistry testing using the TBA-c16000 system and multiple platforms, including Cobas 800, Hitachi 7600, and AU 5800, in the present study, and comparison with reference intervals from another single-center study in a Korean population and CALIPER study that were converted from conventional units to SI units

Analyte (unit)	Total N	Age	Sex	N	This study		Severance Hospital [15]			CALIPER [8]		
					Lower limit (90% CI)	Upper limit (90% CI)	Age	Lower limit	Upper limit	Age†	Lower limit	Upper limit
Calcium (mmol/L)	387	0–11 mo	Both	101	2.44 (2.41–2.47)	2.85 (2.82–2.87)				0–1 yr	2.13	2.75
		1–6 yr	Both	143	2.28 (2.23–2.33)	2.61 (2.60–2.70)				1–19 yr	2.30	2.63
		7–18 yr	Both	143	2.23 (2.20–2.25)	2.53 (2.48–2.53)						
Multicenter combined (T+C+H)	780	0–11 mo	Both	270	2.43 (2.40–2.48)	2.81 (2.80–2.85)						
		1–6 yr	Both	280	2.28 (2.25–2.30)	2.67 (2.63–2.70)						
		7–18 yr	Both	230	2.20 (2.15–2.23)	2.58 (2.55–2.63)						
Phosphorus (mmol/L)	390	0–3 mo	Both	32	1.74 (1.63–1.85)	2.54 (2.45–2.61)	M: 2–16 yr	1.23	1.91			
		4–11 mo	Both	68	1.57 (1.51–1.63)	2.29 (2.23–2.35)	F: 2–14 yr	1.26	1.87			
		1–6 yr	Both	147	1.42 (1.26–1.45)	2.07 (2.00–2.16)						
Glucose (mmol/L)	377	All	Both	377	4.38 (4.11–4.44)	6.44 (6.33–6.49)	M: 2–16 yr	4.16	6.49			
		All	Both	655	4.38 (4.16–4.50)	6.42 (6.33–6.49)	F: 2–14 yr	4.11	6.72			
		0–11 mo	Both	101	0.40 (0.11–0.70)	5.17 (4.85–5.61)				0–14 d	1.00	8.21
Multicenter combined (T+C)	655	1–18 yr	Both	295	2.28 (2.14–2.50)	6.07 (6.07–6.43)				15 d–1 yr	1.21	6.00
			Both	33	0.08 (0.06–0.10)	0.25 (0.23–0.27)	M: 2–10 yr	0.15	0.35	1–10 yr	3.21	7.89
			Both	204	0.12 (0.11–0.14)	0.30 (0.29–0.34)	M: 11–16 yr	0.18	0.45	10–19 yr	M: 2.61, F: 2.61	M: 7.50, F: 6.78
BUN (mg/dL)	396	0–3 mo	Both	33	0.08 (0.06–0.10)	0.25 (0.23–0.27)						
		4 mo–6 yr	Both	204	0.12 (0.11–0.14)	0.30 (0.29–0.34)						
		7–12 yr	M	21	0.16 (0.14–0.19)	0.37 (0.33–0.40)	F: 2–14 yr	0.15	0.35			
Uric acid (mmol/L)	357	7–12 yr	F	25	0.12 (0.09–0.15)	0.37 (0.33–0.40)						
		13–18 yr	M	34	0.23 (0.21–0.27)	0.46 (0.43–0.49)						
			F	40	0.16 (0.14–0.18)	0.36 (0.34–0.39)						
Cholesterol (mmol/L)	388	All	Both	388	2.95 (2.82–3.06)	5.64 (5.46–5.83)	M: 2–16 yr	3.16	5.41	0–14 d	M: 1.09, F: 1.19	M: 2.82, F: 3.24
		All	Both	489	2.90 (2.82–3.03)	5.72 (5.52–5.85)	F: 2–14 yr	3.32	5.46	15 d–1 yr	1.66	6.14
			Both	32	53.6 (51.1–56.4)	75.0 (72.4–77.2)	M: 2–5 yr	61	74	1–19 yr	2.90	5.39
Multicenter combined (T+H)	489	0–3 mo	Both	32	53.6 (51.1–56.4)	75.0 (72.4–77.2)				0–14 d	53	83
		4–11 mo	Both	70	58.5 (57.0–59.7)	74.7 (73.0–76.2)	M: 6–16 yr	63	77	15 d–1 yr	44	71
		1–6 yr	Both	147	64.0 (61.0–64.0)	78.0 (76.0–79.0)	F: 2–14 yr	63	77	1–6 yr	61	75

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Table 3. Continued

Analyte (unit)	Total N	Age	Sex	N	This study		Severance Hospital [15]			GALIPER [8]		
					Lower limit (90% CI)	Upper limit (90% CI)	Age	Lower limit	Upper limit	Age <sup>†</sup>	Lower limit	Upper limit
Albumin <sup>§</sup> (g/L)	394	7–12 yr	M	20	68.5* (67.2–69.8)	76.2* (74.9–77.5)				6–9 yr	64	77
		13–18 yr	F	25	65.0* (63.0–67.0)	78.8* (76.7–80.8)				9–19 yr	65	81
		All	Both	394	40 (39–41)	50 (50–51)	M: 2–16 yr F: 2–14 yr	40 40	49 49	0–14 d 15 d–1 yr 1–8 yr 8–15 yr 15–19 yr	33 28 38 41 M: 41, F: 40	45 47 47 48 M: 51, F: 49
Total bilirubin (µmol/L)	367	All	Both	367	3.4 (3.4–3.4)	13.7 (13.7–13.7)	M: 2–8 yr M: 9–16 yr F: 2–9 yr F: 10–14 yr	3.4 3.4 3.4 3.4	13.7 17.1 13.7 17.1	0–14 d 15 d–1 yr 1–9 yr 9–12 yr 12–15 yr 15–19 yr	3.2 0.9 0.9 0.9 1.7 1.7	283.9 11.6 6.8 9.4 12.0 14.4
		0–12 yr	Both	212	2.01 (1.64–2.47)	5.75 (5.46–6.20)	M: 2–5 yr	1.77	4.35	0–14 d	1.50	4.55
		13–15 yr	M	13	1.04* (0.08–2.00)	5.66* (4.71–6.62)	M: 6–14 yr	1.97	5.68	15 d–1 yr	2.23	8.63
ALP (µkat/L)	268	16–18 yr	F	17	0.92 <sup>†</sup> (N/A)	2.76 <sup>†</sup> (N/A)	M: 15–16 yr	1.03	4.17	1–10 yr	2.60	6.15
		0–12 yr	M	13	0.96* (0.76–1.15)	1.89* (1.69–2.08)	F: 2–5 yr	1.95	4.35	10–13 yr	2.35	7.67
		13–18 yr	F	13	0.57* (0.35–0.78)	1.61* (1.39–1.82)	F: 6–12 yr F: 13–14 yr	2.05 1.02	5.50 6.65	13–15 yr 15–17 yr 17–19 yr	M: 2.12, F: 1.03 M: 1.48, F: 0.90 M: 0.98, F: 0.80	M: 8.62, F: 4.67 M: 6.08, F: 2.13 M: 2.73, F: 1.58
AST (GOT) (µkat/L)	376	0–11 mo	Both	97	0.25 (0.19–0.33)	1.15 (1.07–1.24)	M: 2–16 yr	0.25	0.65	0–14 d	0.53	2.70
		1–6 yr	Both	144	0.38 (0.35–0.40)	0.80 (0.75–0.85)	F: 2–7 yr	0.33	0.68	15 d–1 yr	0.30	1.12
		7–12 yr	M	19	0.27* (0.21–0.33)	0.63* (0.57–0.69)	F: 8–14 yr	0.23	0.57	1–7 yr	0.35	0.73
ALT (GPT) (µkat/L)	380	13–18 yr	F	25	0.22* (0.17–0.26)	0.51* (0.47–0.56)				7–12 yr	0.30	0.60
		0–11 mo	Both	98	0.17 (0.16–0.19)	0.42 (0.39–0.43)	M: 2–16 yr	0.13	0.65	0–1 yr	0.08	0.55
		1–18 yr	Both	282	0.12 (0.12–0.13)	0.45 (0.43–0.52)	F: 2–14 yr	0.12	0.52	1–13 yr 13–19 yr	0.15 M: 0.15, F: 0.13	0.42 M: 0.40, F: 0.37
Creatinine (µmol/L)	391	0–3 mo	Both	33	13.4 (11.3–15.8)	31.7 (29.7–34.2)	M: 2–5 yr	19.4	49.5	0–14 d	37.1	92.8
		4–11 mo	Both	69	18.5 (17.1–20.0)	33.4 (32.3–34.8)	M: 6–9 yr	28.3	61.9	15 d–1 yr	28.3	46.9
		1–6 yr	Both	147	23.0 (18.6–23.9)	46.5 (43.3–47.1)	M: 10–12 yr	34.5	70.7	1–4 yr	33.6	47.7
7–12 yr	Both	48	30.2 (27.4–33.5)	66.2 (62.0–70.0)	M: 13–16 yr	39.8	86.6	4–7 yr	38.9	56.6		

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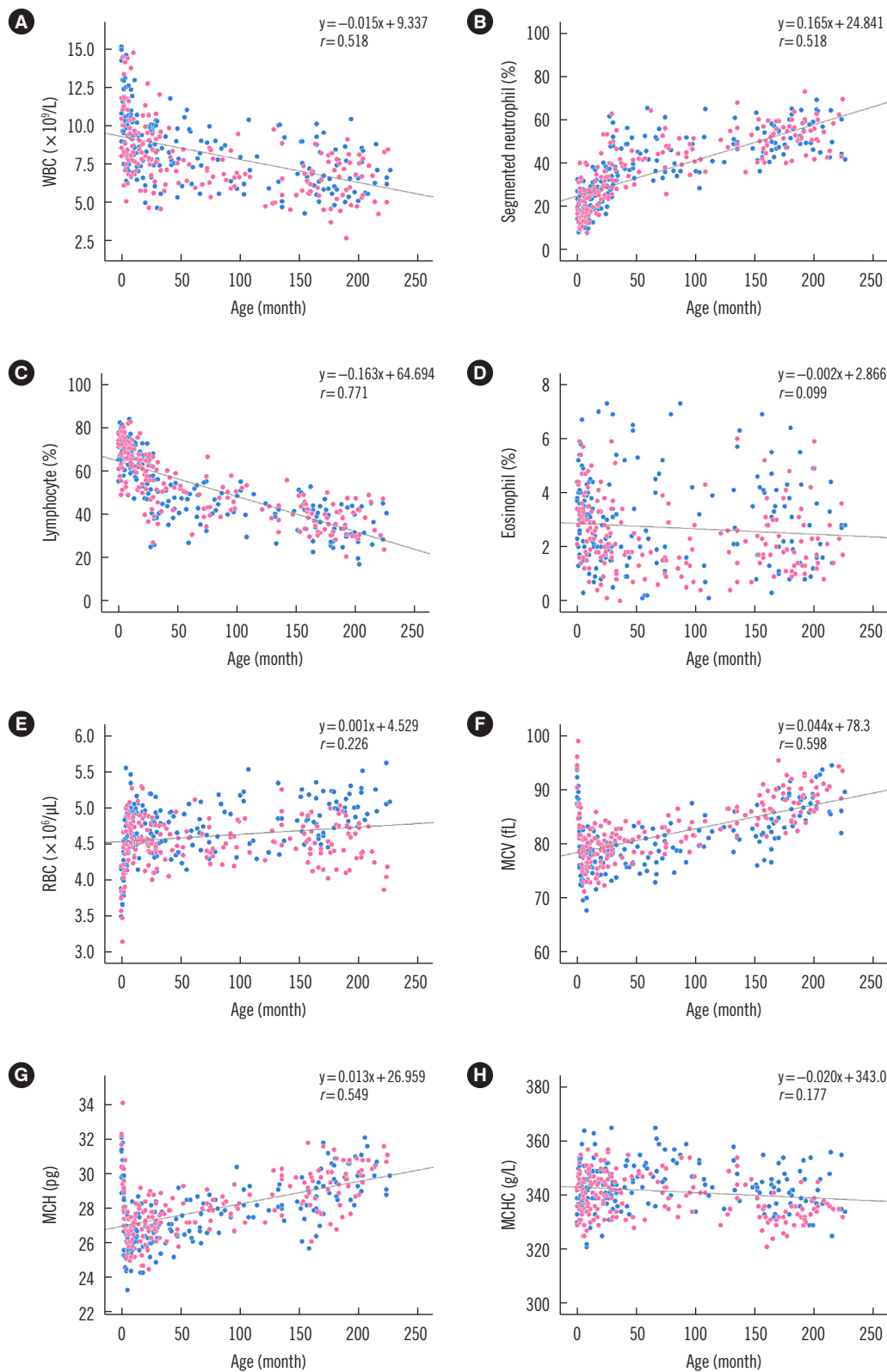


Table 3. Continued

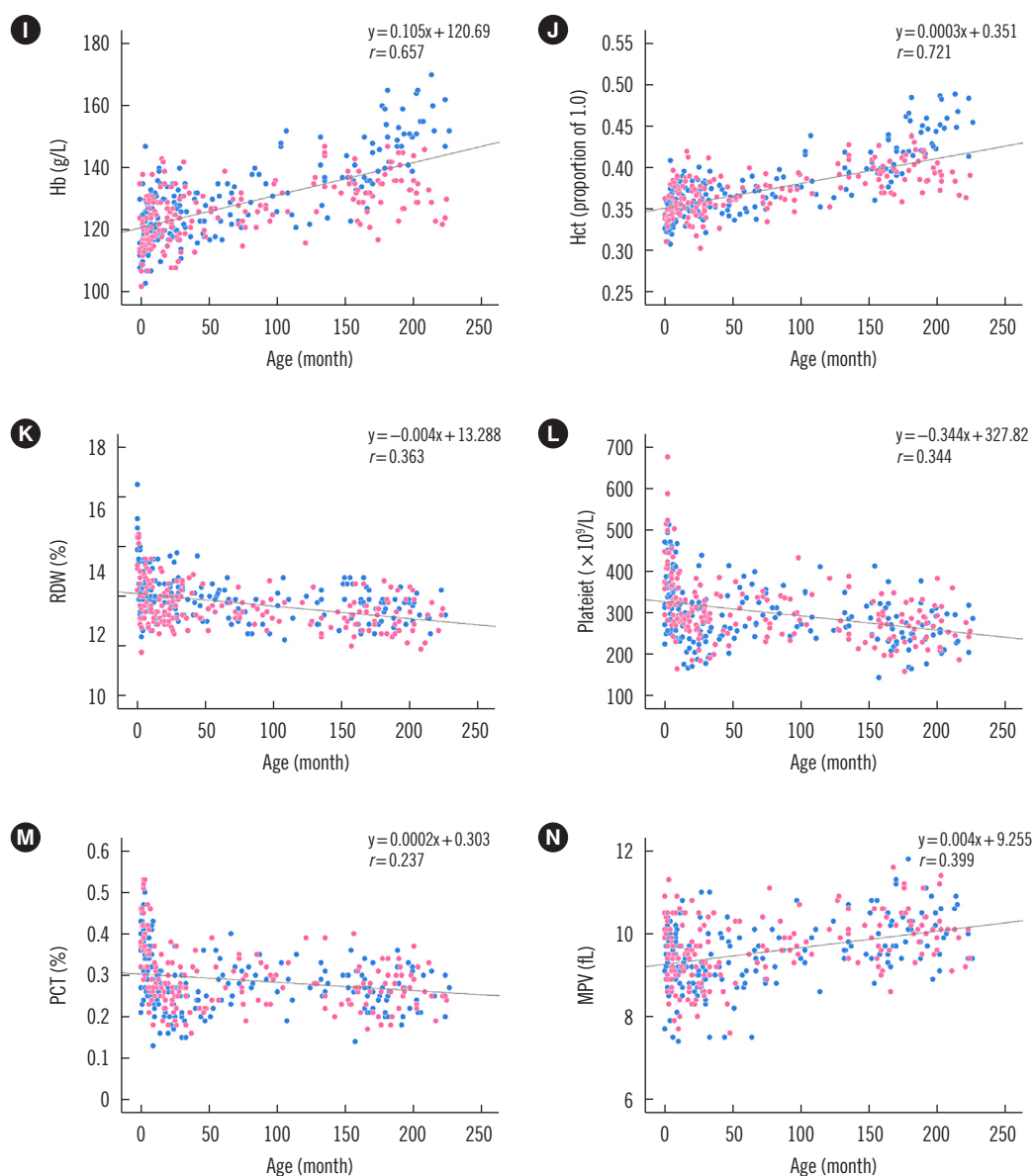
Analyte (unit)	Total N	Age	Sex	N	This study			Severance Hospital [15]			CALIPER [8]		
					Lower limit (90% CI)	Upper limit (90% CI)	Age	Lower limit	Upper limit	Age <sup>‡</sup>	Lower limit	Upper limit	
Na (mmol/L)	392	13–15 yr	M	27	38.7 (31.8–47.4)	97.1 (89.4–104.3)	F: 2–5 yr	18.6	46.9	7–12 yr	46.0	61.0	
		16–18 yr	F	30	44.4 (41.2–48.3)	71.7 (68.1–75.7)	F: 6–8 yr	28.3	56.6	12–15 yr	50.4	70.7	
		0–6 yr	M	18	61.7* (54.5–68.9)	102.8* (95.6–110.0)	F: 9–14 yr	32.7	63.6	15–17 yr	M: 58.3, F: 52.2	M: 91.9, F: 76.0	
		7–18 yr	F	19	48.9* (44.0–53.8)	77.8* (72.9–82.7)	M: 17–19 yr	M: 61.0, F: 53.0	M: 97.2, F: 77.8				
Multicenter combined (T+C+A)	909	0–6 yr	Both	248	134 (133–134)	141 (140–141)							
		7–18 yr	Both	144	136 (135–136)	142 (142–143)							
		0–3 mo	Both	42	132 (131–133)	142 (141–143)							
		4 mo–6 yr	Both	603	135 (135–136)	143 (142–143)							
K (mmol/L)	389	7–18 yr	Both	264	136 (135–136)	144 (143–144)							
		0–3 mo	Both	33	4.5 (4.2–4.7)	6.3 (6.1–6.5)							
		4–11 mo	Both	67	4.1 (4.0–4.2)	5.3 (5.2–5.4)							
		1–18 yr	Both	289	3.7 (3.6–3.7)	4.8 (4.7–4.9)							
Multicenter combined (T+C+A)	911	0–3 mo	M	20	4.7* (4.4–4.9)	6.2* (5.9–6.4)							
		4–11 mo	F	21	4.3* (4.0–4.6)	6.2* (5.9–6.5)							
		1–18 yr	Both	248	4.2 (4.0–4.3)	5.4 (5.3–5.5)							
		0–6 yr	Both	622	3.7 (3.7–3.8)	5.0 (4.9–5.1)							
Cl (mmol/L)	398	All	Both	398	100 (99–100)	108 (108–109)							
		0–6 yr	Both	890	101 (100–101)	107 (107–108)							
		7–12 yr	Both	248	17 (17–18)	26 (26–28)	0–14 d			0–14 d			
		13–18 yr	Both	47	21 <sup>†</sup> (N/A)	29 <sup>†</sup> (N/A)	15 d–1 yr			15 d–1 yr			
TCO2 (mmol/L)	393	0–6 yr	Both	98	21 (20–22)	31 (30–32)	1–5 yr			1–5 yr			
		7–12 yr	Both	241	0.10 (0.10–0.10)	1.80 (1.70–2.30)	5–15 yr			5–15 yr			
		13–18 yr	Both	241	0.10 (0.10–0.10)	1.80 (1.70–2.30)	15–19 yr			15–19 yr			
		All	Both	241	0.10 (0.10–0.10)	1.80 (1.70–2.30)	0–14 d			0–14 d			
CRP (mg/L)	241	0–6 yr	Both	241	0.10 (0.10–0.10)	1.80 (1.70–2.30)	0–14 d			0–14 d			
		7–12 yr	Both	241	0.10 (0.10–0.10)	1.80 (1.70–2.30)	15 d–15 yr			15 d–15 yr			
		13–18 yr	Both	241	0.10 (0.10–0.10)	1.80 (1.70–2.30)	15–19 yr			15–19 yr			
		All	Both	241	0.10 (0.10–0.10)	1.80 (1.70–2.30)	0–14 d			0–14 d			

\*The 90% CI was not determined using the robust method; however, the data passed a normality test and a parametric method based on the normal distribution was applied; <sup>†</sup>Although the sample size was <120, the values presented were established using a nonparametric method because the 90% CI calculated using the robust method contained a negative value; <sup>‡</sup>Age-specific group for the CALIPER study described as “A-B” means equal to or older than A, and younger than B; <sup>§</sup>Albumin concentration was measured using a bromocresol green dye-binding assay (AlbBCG).

<sup>†</sup>Creatinine was measured using the Jaffe reaction.  
Abbreviations: CALIPER, Canadian laboratory initiative on pediatric reference interval database; T, TBA-c16000; C, Cobas 800; H, Hitachi 7600; d, day; mo, month; yr, year; A, AU 5800; BUN, blood urea nitrogen; ALP, alkaline phosphatase; AST, aspartate aminotransferase; GOT, glutamate oxaloacetate transaminase; ALT, alanine aminotransferase; GPT, glutamate pyruvate transaminase; TCO2, total CO<sub>2</sub>; CRP, C-reactive protein.



**Fig. 1.** Scatter plots of hematology test results showing age-dependent changes. Results obtained at SNUH using the Sysmex XE-2100 platform after outlier exclusion are presented. (A–D) WBC count, percentage of segmented neutrophils, lymphocytes, and eosinophils. (E–K) RBC count, MCV, MCH, MCHC, hemoglobin concentration, hematocrit, and RDW. (Continued to the next page)



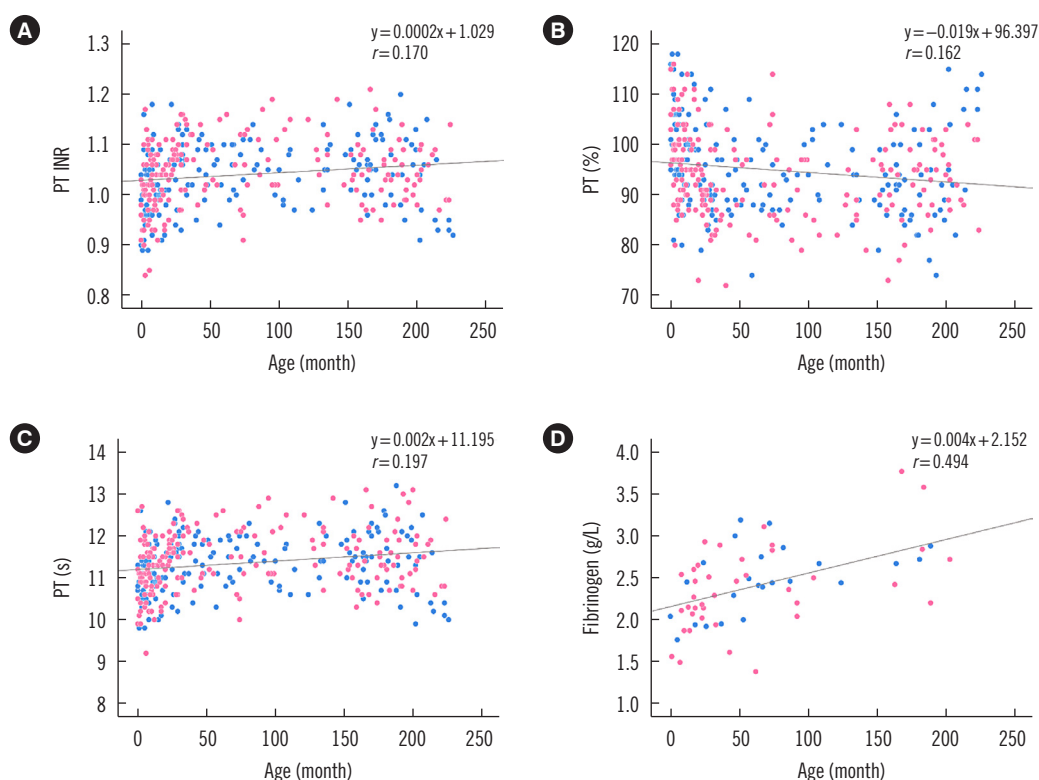
**Fig. 1.** Continued. (L–N) platelet count, PCT, and MPV. Blue and red dots represent male and female children enrolled in the present study. The gray line represents the trend line; its equation and *r* value are given. Abbreviations: SNUH, Seoul National University Hospital; WBC, white blood cell; MCV, mean corpuscular volume; MCH, mean corpuscular hemoglobin; MCHC, mean corpuscular hemoglobin concentration; RBC, red blood cell; RDW, RBC distribution width; PCT, plateletcrit; MPV, mean platelet volume; Hct, hematocrit.

3B and 3K). Reference intervals for tests that showed age-dependent changes largely differed from adult reference intervals. Tests that did not show age-dependent changes and for which no age-specific reference intervals were established are presented as scatter plots in Supplemental Data Figs. S1 and S2.

## DISCUSSION

Although indirect sampling for establishing reference intervals is

controversial [28, 29], and the CLSI working group does not endorse this method as a primary approach [2], some studies support well-designed indirect sampling methods [30–33]. While the possibility of including “unhealthy” patients in the reference group cannot be completely excluded owing to the intrinsic limitation of this approach, best efforts to derive reliable, sophisticated results from data have been made by careful medical record review and discussion by the international CALIPER research team, which leads studies on the establishment



**Fig. 2.** Scatter plots of coagulation test results showing age-dependent changes. Results obtained at SNUH using the ACL TOP analyzer (Instrumentation Laboratory, Bedford, MA, USA) after outlier exclusion are presented. (A–D) PT INR, PT percentage, PT, and fibrinogen. Blue and red dots represent male and female children enrolled in the present study. The gray line represents the trend line; its equation and  $r$  value are given.

Abbreviations: SNUH, Seoul National University Hospital; PT, prothrombin time; INR, international normalized ratio.

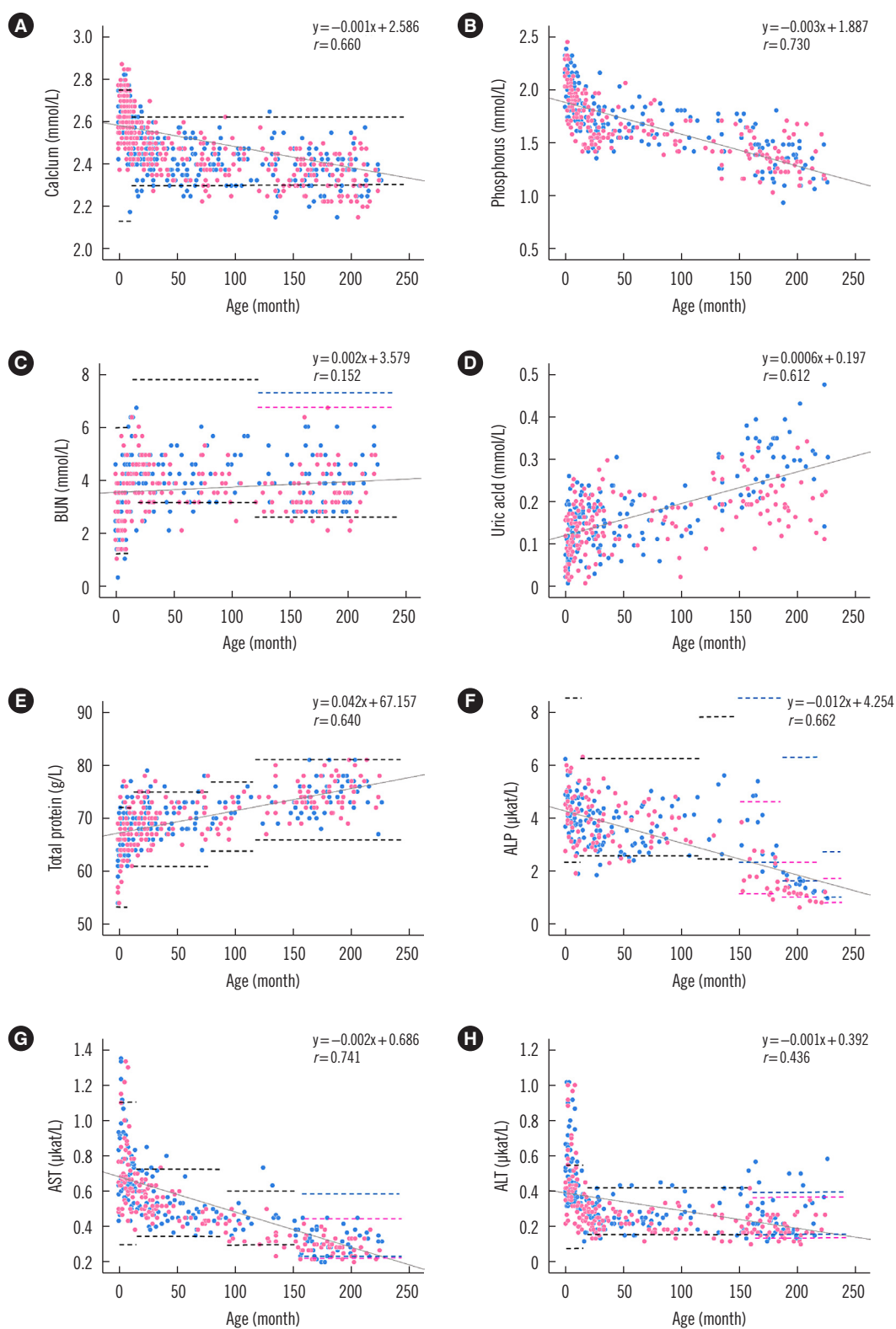
of authoritative pediatric reference intervals. While we did select “presumed healthy” patients on the basis of medical record review, various statistical techniques are available to extract “healthy” population data from a mixed dataset using visualization or formulation, under the assumption that the reference values from the healthy population have a near-Gaussian distribution [21].

Age-specific reference intervals were required for most tests, except glucose, cholesterol, albumin, bilirubin, Cl, and CRP, which did not require any partitioning. Age partitioning of children younger or older than 3 months for Na was confirmed only in the multicenter analysis, which included a larger sample size than the single-center analysis. Sex-specific reference intervals were established for P, uric acid, protein, ALP, AST, and Cr in some age-partitioned subgroups, and for K only in the multicenter analysis. Generally, reference intervals were more strongly affected by age than by sex, which is in line with findings in previous studies [7, 8, 24, 32]. However, the influence of sex on the reference intervals in the present study was not significant or substantially smaller than that in previous studies [7, 8]. For

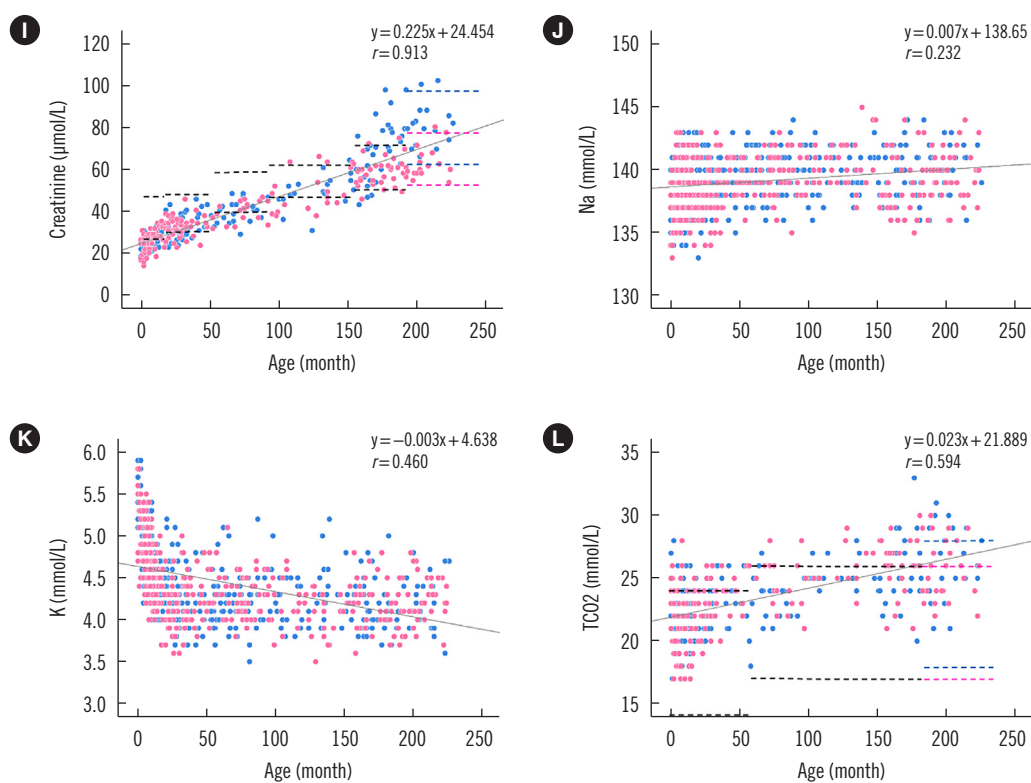
example, the reference intervals for calcium, BUN, cholesterol, albumin, bilirubin, and ALT were affected by sex in at least one age-partitioned subgroup in these previous studies, whereas we did not observe an effect of sex for these tests. This may be due to the relatively small sample sizes of the sex-partitioned subgroups, thus reducing statistical power, or due to age-partitioning based on chronological age regardless of individual growth and sexual development, such as the Tanner stage.

Age-dependent increasing tendencies were observed for segmented neutrophil count, hemoglobin, hematocrit, BUN, uric acid, and Cr, whereas decreasing tendencies were observed for WBC count, lymphocyte percentage, platelets, P, and K. Age-dependent increasing tendencies for Cr, uric acid, and decreasing tendencies for P have been observed in previous studies [8, 15, 28]. Figs. 1, 2, and 3 show age-dependent changes in pediatric laboratory test results, which can lead to misinterpretation of “abnormal” results from pediatric patients as “normal” in reference to adult intervals or vice versa. Thus, adult reference intervals are inappropriate for interpreting results of pediatric patients.

Despite the best efforts to minimize disparities among plat-



**Fig. 3.** Scatter plots of clinical chemistry results showing age-dependent changes. Reference intervals established in a previous study based mainly on a Caucasian population [8] are indicated as horizontal dashed lines. All test results that passed method comparison, considered mergeable, and subsequently used in multicenter analysis are plotted. (A–H) Calcium, phosphorus, BUN, uric acid, total protein, ALP, AST, and ALT. *(Continued to the next page)*



**Fig. 3.** Continued. (I–L), creatinine, sodium, potassium, and TCO<sub>2</sub>. Blue and red dots represent male and female children enrolled in the present study. Blue, red, and black dashed lines represent male, female, and common reference limits established in a previous study, respectively. The gray line represents the trend line; its equation and  $r$  value are given. Abbreviations: BUN, blood urea nitrogen; ALP alkaline phosphatase; AST, aspartate aminotransferase; ALT, alanine aminotransferase.

forms and methods, previous studies have reported such disparities arising from differences in instruments and testing principles used [5, 8], suggesting the necessity of establishing platform- or methodology-specific reference intervals. Our study was initially designed as a multicenter trial for the establishment of Korean pediatric reference intervals, which generally are applicable to all analytical platforms. However, based on a method comparison, only items that showed sufficient comparability of test methods were included in the multicenter reference interval analysis. This difference between instruments may have contributed to the loss of ethnical influence like lower ALT level in the Asian population that was revealed in a previous study but not observed in our study (Table 3). Therefore, the reference intervals determined in the present study are valid only for specific platforms (hematology testing using Sysmex XE-2100, coagulation testing using ACL TOP, and chemistry testing using TBA-c16000) and the availability can be limited now because one of the platforms, Sysmex XE-2100 has been discontinued. However, some chemistry test results were highly equivalent to those obtained from TBA-c16000 and thus can be transferred

to other platforms after validation.

The transferability of CALIPER reference intervals established using one platform to other platforms even if  $r^2$  is smaller than 0.95 has been previously assessed using statistical methods such as Deming regression rather than simple linear regression when  $r^2$  is larger than 0.70 [34]. Further, research groups have attempted to transfer CALIPER reference intervals to their own platform in practical trials [35, 36]. We assessed method comparability of different platforms using the protocol suggested in the CLSI EP09-A3 guidelines [37], which revealed a high correlation among the platforms, as shown in Supplemental Data Table S3. Thus, although results of a limited number of test items could be combined to determine pediatric reference intervals, most pediatric reference intervals for chemistry tests established in the present study can be adopted by other laboratories and utilized for patient evaluation after adequate validation [2], without the need for establishing institution-specific reference intervals.

Our study had some limitations. As a retrospective study, data were not obtained in a well-controlled condition. For example, blood samples for preoperative tests had not been taken under



strictly controlled conditions considering diet or a fasting period, which might have affected the results of certain tests. Therefore, the influence of these factors cannot be completely excluded, although one study interpreted this characteristic of indirect sampling to well reflect “routine” pre-analytical and analytical conditions [33]. Additionally, the small sizes of some subgroups might have led to analytical inaccuracies despite the application of robust statistical methods. Although the analytical approach we used was developed to allow for establishing reliable reference intervals for small reference groups, broadening of the CI for the upper and lower reference limits is inevitable. In practice, lower reference limits of some tests or the 90% CI of those determined using the robust method showed negative values because of a small subgroup size. Sample size affects not only statistical power but also subgroup partitioning per se. As shown in Table 3, in the CALIPER study, age-specific reference intervals were established for the early neonatal period (0–14 days). However, we could not establish reference intervals for this age group, as our study did not include reference individuals of this age.

Further, the method comparison test was performed only for clinical chemistry tests on the platforms used in the participating hospitals (TBA-c16000, AU 5800, COBAS 8000, and Hitachi 7600). Therefore, hematology and coagulation test results from other instruments were unavailable to establish multicenter reference intervals by merging with those obtained from Sysmex XE 2100 and ACL TOP, although some tests showed comparable results in the diagnostic hematology program of Korean Association of External Quality Assessment (Supplemental Data Table S4) and Mann-Whitney U test (Supplemental Data Table S5). In addition, chemistry test results from other platforms that were not standardized or harmonized cannot be combined for analysis, even if the comparison test showed acceptable equivalence.

Considering the limitations of the present study, future studies designed in a prospective manner with direct reference individual sampling based on healthy community children with a larger sample size, strict control of the sample collection condition, and matching platforms between hospitals are required to establish reliable pediatric reference intervals.

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## AUTHOR CONTRIBUTIONS

JY Sung and JD Seo drafted the manuscript. JY Sung, DH Ko,

MJ Park, and SM Hwang acquired data from each institution. JY Sung, JD Seo, and S Oh analyzed the data and interpreted the results. JY Sung, MJ Park, S Chun, MW Seong, J Song, SH Song, and SS Park designed the study. JD Seo and SH Song revised and approved the final manuscript. All authors reviewed the manuscript and provided critical feedback.

## CONFLICTS OF INTEREST

None declared.

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