## Review

## **Breast tumour angiogenesis**

Stephen B Fox<sup>1</sup>, Daniele G Generali<sup>2</sup> and Adrian L Harris<sup>3</sup>

Pathology, Peter MacCallum Cancer Centre, St Andrews Place, East Melbourne, Victoria, 3002, Australia

<sup>2</sup>Unità di Patologia Mammaria, Senology and Breast Cancer Unit, Breast Research Laboratory, Istituti Ospitalieri di Cremona, Viale Concordia, 26100 Cremona, Italy

3 Molecular Oncology, Cancer Research UK, Weatherall Institute of Molecular Medicine, John Radcliffe Hospital, Headley Way, Oxford, OX3 9DS, UK

Corresponding author: Stephen B Fox, stephen.fox@petermac.org

Published: 18 December 2007

This article is online at http://breast-cancer-research.com/content/9/6/216

© 2007 BioMed Central Ltd

Breast Cancer Research 2007, 9:216 (doi:10.1186/bcr1796)

#### **Abstract**

The central importance of tumour neovascularization has been emphasized by clinical trials using antiangiogenic therapy in breast cancer. This review gives a background to breast tumour neovascularization in *in situ* and invasive breast cancer, outlines the mechanisms by which this is achieved and discusses the influence of the microenvironment, focusing on hypoxia. The regulation of angiogenesis and the antivascular agents that are used in an antiangiogenic dosing schedule, both novel and conventional, are also summarized.

## Introduction

It has been 3 years since the last critical review of antiangiogenic therapy was published in Breast Cancer Research [1], and since then the central importance of tumour neovascularization has been emphasized by clinical trials in various tumour types, including breast cancer. Many of these trials have used bevacizumab (Avastin™; Genentech, South San Francisco, CA, USA), which was specifically designed to target vascular endothelial cell growth factor (VEGF). Bevacizumab is a recombinant VEGF antibody derived from a humanized murine monoclonal antibody that can recognize all known isoforms of VEGF-A and prevents receptor binding, thereby inhibiting angiogenesis and tumour growth. The critical contribution of this angiogenic factor in controlling many of the processes involved in angiogenesis and its importance as a paradigm for the rational design of an anticancer agent have been among the successes of antiangiogenic treatment, which was first suggested by Judah Folkman more than 35 years ago. The attractiveness of the antiangiogenic approach has always been the wide therapeutic window, since all tumours (including liquid such as leukaemias) are angiogenesis dependent, that angiogenesis is highly restricted in the adult, that endothelium of the vessels are accessible and that any treatment would be amplified through subsequent tumour infarction. Furthermore,

the erstwhile problem in oncology of resistance should not be an issue because endothelial cells are non-neoplastic and should have a stable genome [2].

Nevertheless, although these trials have demonstrated significant improvements in response rates, findings to date have not indicated substantial benefits in terms of survival. This is likely to be due to redundancy in breast tumours with an individual tumour being able to utilise several angiogenic pathways at any one time [3] with changes in this profile during tumour progression coupled with the use of other mechanisms to establish a blood supply. Indeed, the central tenet that tumours are angiogenesis dependent (in that for a tumour to grow, this must be preceded by a wave of angiogenesis to deliver nutrients and meet the metabolic requirements of the growing tumour) has been challenged. Thus, a number of nonangiogenic mechanisms may contribute to establishing tumour blood supply; these include cooption, vasculogenesis, vascular remodelling, intussusception and vascular mimicry.

A further important issue that has not been addressed is stratification of patients for appropriate treatment; specifically, individual patients given antiangiogenic agents have yet to be selected based on the characteristics of their tumour. It is therefore likely, as has been demonstrated for other targeted agents such as herceptin, that benefit will be restricted to those patients whose tumours rely largely on VEGF signalling for their angiogenic response. The administration of agents based on the biology of the individual tumour (so-called personalized medicine) will become increasingly important not only to generate maximum therapeutic benefit to the patient but also to realize the optimal economic advantage from the finite resources available.

FGF = fibroblast growth factor; HER = human epidermal growth factor receptor; HIF = hypoxia-inducible factor; PDGFR = platelet-derived growth factor receptor; TAM = tumour-associated macrophage; TKI = tyrosine kinase inhibitor; VEGF = vascular endothelial cell growth factor; VEGFR = vascular endothelial cell growth factor receptor.

## **Breast tumour neovascularization**

Angiogenesis in the normal human adult is highly restricted, largely to wound healing and reproduction. Sustained angiogenesis is pathological and is characteristic of many common diseases, including diabetes, psoriasis and rheumatoid arthritis [4]. Thus, in order to initiate neovascularization, a tumour must switch to an angiogenic phenotype. Evidence from transgenic models that have reproducible distinct tumour stages suggest that the acquisition of this phenotype occurs early in tumour development and that it is rate limiting with regard to tumour progression [5.6]. These experimental models are supported by findings in human tissues, in which 30% of transplanted human hyperplastic breast tissue samples were found to be angiogenic as compared with only 3% of samples from normal breast tissue [7-9]. Interestingly, normal breast adjacent to malignant breast induced angiogenesis twice as frequently as did tissues from non-neoplastic breast, suggesting that the angiogenic switch occurs before morphological changes are identifiable [10]. Using microvessel density as a surrogate for angiogenesis, benign lesions associated with high vascular density are correlated with increased risk for developing breast cancer. It has also been suggested that quantification of angiogenesis might help to predict the likelihood that in situ cancers will progress [11,12] or that a tumour will respond to treatment [13-17], and has been shown to correlate directly with the presence of bone marrow micrometastases [18] and survival [19,20].

Although it is likely that different tumour types use different genetic pathways to establish a blood supply, oncogenes and tumour suppressor genes that are frequently associated with transformation also appear to be important in activating the angiogenic switch. Thus, Ras, myc, raf, c-erbB-2, c-jun and src transformed cells exhibit a strong angiogenic phenotype [21-24]. However, the vessels formed under the influence of these pathways are abnormal, leaky with blind sacs, and have reversed and intermittent flow [25]. The result is that although there is an increase in formation of new vessels, drug and oxygen delivery is much poorer than in normal tissues. This leads to hypoxia and microenvironmental stresses that have been demonstrated to have profound effects on tumour biology and resistance to treatment [26].

## The microenvironmental influence of hypoxia

Hypoxia is the pathophysiological consequence of a structurally and functionally disturbed microcirculation [26], and it is therefore a common feature in solid tumours. Tumours respond to low oxygen tension by enhancing the hypoxia-inducible factor (HIF) response [27]. The HIF response is mediated through the transcription factor dimer inducible HIF-1 $\alpha$  and constitutively expressed HIF-1 $\beta$  (also known as aryl hydrocarbon nuclear translocator).

In normoxia, three prolyl hydroxylases (prolyl hydroxylase-1, -2 and -3) hydroxylate HIF-1 $\alpha$  at two proline residues in its

oxygen-dependent degradation domain, with oxoglutarate from the Krebs cycle, ascorbate and Fe2+ leading to recognition and binding of the  $\alpha$  domain by the von Hippel-Lindau protein. This interaction, and through binding of elongin C via von Hippel-Lindau β domain in turn, leads to ubiquitination and targeting for degradation through the proteasome. In conditions of hypoxia, however, molecular oxygen is not available for hydroxylation which results in HIF-1 $\alpha$  stabilization and translocation to the nucleus, where it binds to HIF-1β and consensus hypoxia response elements on gene promoters. Co-activators and polymerases are recruited and transcriptional activation of several gene pathways that are involved in angiogenesis, glycolysis, erythropoiesis and apoptosis occurs. The asparagine hydroxylase, factor inhibitor of HIF-1, and CITED4 (CBP p300-interacting transactivator 4), which interfere with coactivator binding, provide a further level of control [28,29].

Over-expression of HIF-1 a protein has been identified in various tumour types, with high levels influencing the growth rate and metastatic potential of these cancers. In breast cancers, the frequency of HIF-1 $\alpha$  positive cells increases in parallel with increasing clinical stage and is associated with poor prognosis [30-33]. In addition to HIF-1α, other isoforms have been identified, namely HIF-2 $\alpha$  and HIF-3 $\alpha$  [34,35]. The roles played by these isoforms are complex, but there is evidence that the latter antagonizes hypoxia-dependent gene expression, whereas the former can enhance the hypoxic response element. Interestingly, there is evidence that HIF-1 $\alpha$ and HIF- $2\alpha$  activate different sets of hypoxia-inducible genes [36], including those involved in glycolysis, cell survival and proliferation [37]. However, the clinical relevance to breast cancer is not known because there are only limited data on these HIF isoforms. Thus, in breast cancer HIF- $2\alpha$  has been reported to be expressed in both tumour cells [38] and in tumour-associated macrophages (TAMs) [39]. TAM HIF- $2\alpha$ expression was found to be related to tumour vascularity, suggesting that hypoxia induces TAM clustering and overexpression of TAM HIF-2α, thereby inducing an angiogenic phenotype, leading to induction of localized angiogenic hot spots [39]. Thus, hypoxic stress response through HIF is likely to be an important mechanism by which continued remodelling of vessels occurs.

Recognition that HIF plays a significant role in tumour behaviour, conferring an aggressive phenotype and contributing to resistance to both radiotherapy and chemotherapy [40], has led to efforts to target the HIF pathway. Several trials of agents that decrease and/or block HIF-1 $\alpha$  expression, including rapamycin/CCI779, quinocarmycin, topoisomerase inhibitors, anti-microtubular agents, YC-1, 17-AAG, thioredoxin inhibitors and 2ME2, have been conducted or are planned [41]. Other potential targets in HIF signalling include the molecules that are involved in oxygen sensing or transcription. For instance, obstructing the interaction between HIF-1 $\alpha$  and the co-activator CBP/p300 led to attenuation of

HIF-induced gene expression and inhibition of tumour growth in a xenograft model [42]. An alternative strategy is to use the HIF pathway to activate bioreductive drugs such as tirapazamine, which inhibits DNA repair under hypoxic condition [43] and has been shown to have an antiangiogenic effect as well as direct antitumour activity [44]. Correction of the hypoxic environment by reducing anaemia [45] using human recombinant erythropoietin is also a potentially effective approach.

## Mechanisms of neovascularization

Although sprouting-type angiogenesis is an important mechanism in tumour neovascularization, several other mechanisms by which tumours establish a blood supply have been identified; these include vascular remodelling, vasculogenesis, vascular mimicry and glomeruloid angiogenesis. Each may have significance in a particular tumour type or at a particular stage of tumour evolution, but the relative importance of each in human tumours is unknown. However, angiogenesis and vascular remodelling appear to be the major mechanisms in breast cancer with evidence that vascular mimicry may additionally play a role in inflammatory breast cancer. The acquisition of this type of biological information is likely to become more important as patients are treated in a more individuallized manner. Although microvessel density has been used as a surrogate for angiogenesis, many other parameters of tumour neovascularization have also been explored, including angiogenic factor expression, cell adhesion molecules, vessel maturation and endothelial cell proliferation [19]. These measures, including microvessel density, have associated problems [46] and none provides a reliable measure of blood flow, which is extremely variable because of shunting, stasis and even reverse flow occurring through the abnormal tumour vasculature [47].

## **Angiogenesis**

Angiogenesis is the generation of new blood vessels from the existing vasculature. It consists of multiple coordinated, sequential and interdependent steps. The angiogenic programme requires the degradation of the basement membrane, endothelial cell migration and invasion of the extracellular matrix, with endothelial cell proliferation and capillary lumen formation before maturation and stabilization of the new vasculature. The latter requires inhibition of further endothelial proliferation, reconstitution of the basement membrane, and junctional complex formation and organization of endothelial cells into a new luminal space.

## Vascular remodelling

In contrast to animal models in which endothelial cells proliferate 30-fold to 40-fold faster in tumour blood vessels than in the vasculature of normal tissue, irrespective of tumour type, growth rate, or size, endothelial cell proliferation in human breast tumours is relatively rare. The corollary of this finding is that vascular remodelling must be the dominant mechanism in establishing the neovasculature in breast

cancers [48-50]. This can occur through a variety of processes including co-option, in which tumours hijack the existing vasculature. This has been reported early in brain tumour development, in which existing blood vessels are used in the absence of an angiogenic response [51,52], although continued tumour growth results in angiogenesis. The tie2-angiopoietin and VEGF growth factor pathways may regulate these respective mechanisms of tumour vascularization. Thus, in some circumstances tumours are able to 'parasitize' the normal stroma and sinusoidal vasculature for its metabolic needs.

Intussusception of tumour columns has also been hypothesized to contribute to the establishment of a tumour blood supply. This process is independent of endothelial cell proliferation and is rapid, depending on insertion of tissue pillars into vessels, partitioning the vessel lumen into two or more channels [53,54]. This may be part of vascular remodelling, which may be the dominant mechanism in the establishment of the tumour vascular bed.

## Vasculogenesis

Vasculogenesis is the *de novo* generation of blood vessels from endothelial cell progenitors, as occurs in the embryo. In animal models it has been demonstrated that circulating endothelial cell precursors derived from the bone marrow lodge in the cancer vasculature, differentiate into endothelial cells and enhance tumour neovascularization through a combination of vasculogenesis and conventional angiogenesis [55-58]. There appear to be differences in the proportion of tumour neovascularization that can be apportioned to vasculogenesis, depending on the model (up to 90%). In orthoptic models this appears to account for <5% and in human tumours had an average of 4.9% (range 1-12%) when examining tumours from transplant recipients [59,60].

There is evidence to suggest that this process of tumour vascularization may be more frequent and/or significant in early tumour development because inhibition of stem cells or endothelial cell precursor mobilization prevents xenografts from inducing the initial angiogenic response [61]. Nevertheless, there is some debate as to whether such bone marrow derived endothelial cells are actually incorporated into the vasculature and whether they may be acting in a paracrine/support function [62]. The discrepancies between reported findings may be due to replacement of bone marrow derived cells with surrounding endothelial cells with tumour progression [63]. The two models may not be mutually exclusive because several different populations of cells have been reported that may be involved in a support and integral role (for instance, macrophages/monocytes, myeloid progenitors, platelet/megakaryocyte lineages, pericyte progenitors, neutrophils and so-called vascular leucocytes, which express mixed endothelial and white cell lineages). Regulation of these processes may be through angiogenic factors such as VEGF, which mobilizes precursor cells from the bone marrow, but there are some data to suggest that the subsequent retention of these progenitor cells may require additional factors such as stromal-derived factor 1 [64]. Interestingly, stromal-derived factor 1 is hypoxia inducible via HIF [65]. There are few data in breast, but it has been suggested that vasculogenesis occurs in inflammatory subtype breast tumours [66].

### Glomeruloid angiogenesis

Glomeruloid bodies that are characteristic of glioblastomas are also observed in invasive breast cancers [67]. These are highly complex vascular aggregates that resemble glomeruli of the kidney, composed of a network of capillaries that are variably lined by basement membrane and pericytes. Their presence is associated with a significantly shorter survival in breast (and other) cancers [68]. Their formation is related to VEGF, because this angiogenic factor is not only essential for their induction but also for maintenance of these bodies [69]. We have also observed these in breast cancer xenografts transfected with VEGF [70]. This type of tumour neovascularization may also represent vascular remodelling rather than classical sprouting-type angiogenesis [71].

## Vascular mimicry

Vascular mimicry is a neovascularization strategy that may largely be restricted to aggressive occular malignant melanomas and ovarian tumours [72], but it has also been reported in breast cancers [66]. Partial lining of the capillary surface by tumour cells has been known for many years [73] and was more recently reported in animal models using advanced techniques [74], but vascular mimicry is defined as a complete capillary network composed of tumour cells themselves rather than vascular endothelial cells that conducts blood [75,76]. The tumour cells not only take on the morphology of endothelial cells but they also acquire phenotypic characteristics of endothelium, expressing a number of vascular markers. It is important to recognize this type of neovascularization because the therapeutic implications of having mimicry as a dominant mechanism are that these tumours may not respond to conventional antiangiogenic agents.

## Angiogenic factors in breast cancer

Whatever the mechanism(s) that a tumour use(s) to establish a blood supply, similar regulatory factors are utilized (although some may preferentially be used in particular processes). The presence of a humoral mediator of tumour angiogenesis was suggested more than 60 years ago, but it was not until 1968 that it was demonstrated that a diffusible tumour-derived factor could induce capillary growth [77,78]. Folkman and coworkers [79] reported the first angiogenic factor, namely tumour angiogenesis factor; this discovery was followed by identification of numerous other angiogenic promoters and inhibitors [80,81]. Most have pleiotropic effects, and the role played by many in human tumours is unknown. However,

several important angiogenic pathways have been implicated in human tumour neovascularization.

The angiogenic promoters and inhibitors that underlie establishment of a tumour blood supply through the above-mentioned mechanisms of neovasculrization can originate from the neoplastic cell and/or from other tumour elements. Thus, neoplastic cells can recruit inflammatory cells such as macrophages and mast cells, both of which are rich sources of angiogenic factors and cytokines, or they can induce release of sequestered growth factors or their receptors from the extracellular matrix through protease degradation. Platelets, which also are a rich source of angiogenic factors and are often elevated in malignancy, can be activated by tumour endothelium or epithelium.

Invasive and *in situ* breast cancers express many angiogenic factors, including the VEGF family (see below), fibroblast growth factor (FGF)-1, FGF-2, placenta growth factor, transforming growth factor- $\beta_1$ , thymidine phosphorylase, pleiotrophin and adrenomedullin [3,82,83]. However, these are expressed preferentially at different stages of tumour development. Hence, thymidine phosphorylase is expressed in *in situ* [84] and T1 breast tumours [85], whereas VEGF expression occurs throughout the tumour stages. Furthermore, breast cancers are likely to express different angiogenic profiles, which will necessitate the use of a different spectrum of antiangiogenic agents.

## **VEGF** and anti-**VEGF** therapies

Studies have shown that the VEGF family plays a central role in many human tumour types (for review [81]). Comprising VEGF-A, VEGF-B, VEGF-C, VEGF-D and placenta growth factor, these polypeptides exist in a number of isoforms and can form homodimers and heterodimers. They bind variably to three high-affinity endothelial cell tyrosine kinase receptors, namely Flt-1 (VEGF receptor [VEGFR]1), KDR (VEGFR2) and Flt-4 (VEGFR3), which are responsible for initiating intracellular signalling. Receptor activation results in slightly different effects, with VEGFR1 promoting differentiation and vascular maintenance, VEGFR2 inducing endothelial cell proliferation and vascular permeability, and VEGFR3 stimulating lymphangiogenesis. Additional regulation is achieved through the isoform-specific receptors neuropilin-1 and neuropilin-2. The neuropilins bind not only class 3 semaphorins, which are involved in axonal growth, but also some isoforms of VEGF, where they function as co-receptors, increasing VEGF binding to VEGFR2 [86]. Further modulation is achieved by proteolytic processing and/or heparin, which is not only required for binding of VEGF (and basic FGF) but can also compete for receptor sites. This complex pathway enables the VEGFs to have numerous effects, including increasing vascular permeability (thereby augmenting tumour stroma formation), endothelial cell proliferation, endothelial cell survival and tube formation. Although VEGFR expression is largely endothelial (vascular and/or lymphatic), VEGFRs have also been reported on inflammatory cells such as macrophages and tumour cells themselves.

VEGF-A is highly expressed in many tumours of lung, brain, and gastrointestinal and urogenital tracts, as well as in situ and invasive breast cancers [81]. Expression in some studies is associated with microvessel density and prognosis, supporting the importance of VEGF-A in human malignancies. VEGF has a hypoxic response element in its promoter and is one of several genes that are upregulated in a low oxygen microenvironment to elicit a vascular phenotype. However, the role played by the other family members in human disease is still being elucidated and if tumours are unable to express VEGF-A, other VEGF homologues may be induced to augment neo-vessel formation. VEGF-B, VEGF-C and VEGF-D are also expressed in breast cancers, with some pathological correlates with nodal metastases, prognosis and lymphatic density [87-94]. This may be important for two reasons. First, there is large-scale redundancy in blood supply to any tumour, allowing them to switch angiogenic pathways; this suggests that there is a need for several or multifunction agents. Second, many of these angiogenic factors may synergize with each other (for instance, VEGF-A and FGF-2), at least in vitro.

#### Anti-VEGF therapy

Although there are many 'antiangiogenic' targets for anticancer therapies, many therapies have directly targeted the VEGF pathway because of its critical role in pathological angiogenesis and its profound influence of this growth factor on endothelial biology. Many points on the pathway can be targeted, including direct targeting of ligand and receptor (extracellular and intracellular tyrosine kinase domains) at the protein and mRNA levels, interfering with downstream intermediates, and indirect inhibition of upstream regulators of VEGF.

## Bevacizumab

The most investigated agent to date is bevacizumab (Avastin™; Genentech). Bevacizumab is a recombinant VEGF antibody derived from a humanized mouse monoclonal antibody (93% human) that is composed of the mouse VEGF-binding site joined to a human IgG framework. Bevacizumab recognizes all isoforms of VEGF-A and thereby prevents receptor binding, which leads to inhibition of angiogenesis and tumour growth. *In vitro* bevacizumab inhibits VEGF-induced endothelial cell proliferation and migration, and in xenograft models of a range of tumour types (including breast cancer) tumour growth is significantly decreased by bevacizumab [95,96]. In some human breast carcinoma models, treatment with bevacizumab is associated with a reduction in microvessel density [97].

In phase I/II clinical trial of 75 patients with metastatic breast cancer treated with bevacizumab [98] there was an overall

response rate of 9.3% (confirmed response rate 6.7%) with a median duration of 5.5 months (range 2.3 to 13.7 months); 16% had stable disease or an ongoing response at the end of the trial (after 22 weeks). These data supported the initiation of a phase III clinical trial that combined bevacizumab with capecitabine in patients previously treated with an anthracyclin and a taxane. Although the combination therapy resulted in a statistically significantly increased response rate (19.8% versus 9.1%), neither progression-free nor overall survival differed between arms. Although there are many explanations for the lack of success in terms of the primary end-point of the study, the absence of patient selection (specifically, of those patients whose tumours rely on VEGF) is probably of great importance, in that advanced tumours have redundancy in their ability to establish a blood supply and utilize many pathways. There is ongoing analysis of primary tumour samples for pathological factors that might predict response to bevacizumab in this study.

Nevertheless, a phase III first-line trial comparing bevacizumab plus paclitaxel versus paclitaxel alone in patients with locally recurrent or metastatic breast cancer [99] revealed that this combination increased significantly response rates in all patients (28.2% versus 14.2%; P<0.0001). It also resulted in an increase in median progression-free survival by 4.9 months (6.1 months versus 11 months), which was associated with improved overall survival (hazard ratio 0.674) in the combination arm relative to paclitaxel monotherapy, although this difference did not reach statistical significance. Furthermore, a pilot study conducted in patients with inflammatory breast cancer demonstrated a decrease in tumour cell VEGFR2 (KDR) phosphorylation and an increase in apoptosis after a single cycle of bevacizumab therapy. This response was maintained with the addition of chemotherapy. Results from other trials of bevacizumab are awaited, including studies evaluating neoadjuvant and adjuvant use of this agent, and promising results are emerging for patients with renal cell, colorectal, brain and lung cancers.

In a neoadjuvant study 39 patients with locally advanced breast cancer were treated with docetaxel with or without bevacizumab. There were five complete clinical responses and 24 partial responses, and the therapy was generally well tolerated. Recently, new results for the combination of bevacizumab with doxorubicin and docetaxel in the treatment of inflammatory breast cancer were reported [98]. After treatment, eight out of 13 patients experienced a confirmed partial response, with evidence of a decrease in vascular permeability on dynamic contrast-enhanced magnetic resonance imaging. An ongoing trial plans to evaluate the efficacy of bevacizumab in the adjuvant setting with low dose of methotrexate and cyclophosphamide for cases with residual cancer after neoadjuvant chemotherapy. There is also a planned Eastern Cooperative Oncology Group adjuvant feasibility trial, which will evaluate bevacizumab in combination with dose-dense doxorubicin and cyclophosphamide followed by paclitaxel in women with node-positive breast cancer.

Bevacizumab has a relatively long half-life, which allows intervals between intravenous administrations of up to 3 weeks. The tolerability profile of bevacizumab is generally acceptable in clinical trials, and the drug can readily be delivered with other chemotherapeutic agents that, in some circumstances, may be synergistic. Although phase I trials suggested no dose-limiting toxicity, common adverse events of any severity in patients include asthenia, adnominal pain, headache, hypertension, diarrhoea, nausea, vomiting, anorexia, stomatitis, constipation, upper respiratory infection, epistaxis and proteinuria. Most adverse events were mild to moderate in severity, and events such as hypertension, haemorrhage, or proteinuria were clinically manageable. Thus, bevacizumab provides a highly effective addition to standard chemotherapeutic regimens in several common solid tumours.

### Tyrosine kinase inhibitors of VEGF

Several receptor tyrosine kinase inhibitors (TKIs) that target the tyrosine kinase portion of VEGFR1 and VEGFR2 have been developed that are being investigated [100,101]. The orally administered VEGFR2 inhibitor ZD6474 was generally well tolerated but exhibited little activity in patients with refractory metastatic breast cancer [102]. A variety of other small molecule TKIs targeting the VEGFRs are being evaluated, as are ribozyme (catalytic RNA molecules that specifically cleave VEGFR mRNAs) and antisense strategies.

VEGF is regulated by other transmembrane receptor tyrosine kinases, the most relevant of which in breast cancer is the human epidermal growth factor receptor (HER)1 and HER2 (c-erbB-2, neu) [103-105]. Thus, attenuation of VEGF signalling and therapeutic synergy might be achieved through interference in these other receptor pathways with agents such as trastuzumab in combination with anti-VEGF therapy. The validity and potential of this strategy is supported by the positive correlation between HER2 and VEGF expression in a large cohort of breast cancers [106] and results from a phase I trial of trastuzumab and bevacizumab [107] that indicated a clinical response in five out of nine patients.

Inhibition of the VEGFR mRNA has been attempted both with ribozyme (catalytic RNA molecules), which specifically cleave the mRNAs for the primary VEGFRs [108], and antisense VEGF [109]. Angiozyme is a synthetic ribosome that cleaves the mRNA for the receptor VEGFR1/Flt-1. Preclinical studies confirmed inhibition of both primary tumour growth and metastasis [109]. In patients with refractory solid tumours, a phase I trial of angiozyme demonstrated [110] good tolerability without significant side effects, and phase II trials are ongoing. However, a phase II trial in breast cancer provided no evidence of clinical activity [111], although there was evidence of biological activity, with a decrease in serum VEGFR1 levels.

Recently, various small molecule TKIs targeting the VEGFRs and other critical signalling pathways (for instance, platelet-derived growth factor receptor [PDGFR] and epidermal growth factor receptor) in angiogenesis have been developed. Depending on tumour entity, oral multitargeted TKIs can exert both antiangiogenic and antitumour activities at the same time. As a consequence, they may improve the outcome of cancer patients as single-agent treatment.

A multireceptor targeting agent is PTK787/ZK 222584. It is a pan-VEGF, PDGFR, c-kit and c-Fos receptor TKI. It inhibited the growth of a broad panel of carcinomas in rodent models, with histological examination revealing inhibition of microvessel formation [112]. Patients with a variety of advanced cancers have received this agent and it has been well tolerated. A recent phase I/II study of PTK787/vatalanib in combination with trastuzumab in patients with newly diagnosed HER2-positive metastatic breast cancer has been initiated.

Many extracellular proteolytic enzymes and their inhibitors are active during angiogenesis. Expression of various matrix metalloproteinases has been found to be upregulated in virtually every type of human cancer, and this upregulation correlates with advanced stage, invasive and metastatic properties, and poor prognosis in general [113]. Marimastat, an orally bioavailable hydroxamate, was the most widely studied. E2196 was a phase III trial of 190 patients with metastatic breast cancer who had responding or stable disease after six to eight cycles of first-line chemotherapy for metastatic disease [114]. Patients were randomly assigned to receive marimastat or placebo after chemotherapy. There were no significant differences in median progression-free survival or overall survival, but important musculoskeletal toxicities were noted.

Other agents being evaluated in breast cancer include sunitinib and sorafenib. Future studies are being directed at evaluating these agents in combination with other targeted therapies as well as in the first-line metastatic and/or adjuvant setting. SU11248 (sunitinib malate) is an inhibitor of receptor tyrosine kinases for VEGFR1, VEGFR2, PDGFR, c-kit, and Flt-3. In January 2006, this drug was granted approval by the US Food and Drug Administration for treatment of gastrointestinal stromal tumour after disease progression on, or intolerance to, imatinib mesylate, as well as for the treatment of metastatic renal cell cancer. Sorafenib (BAY 43-9006) belongs chemically to a class described as bis-aryl ureas. It was selected for further pharmacological characterization based on potent inhibition of Raf-1 and its favourable kinase selectivity profile. Sorafenib exhibited significant activity against several receptor tyrosine kinases, including VEGFR2, VEGFR3, PDGFR-α, Flt-3, and c-kit. This molecule is currently being evaluated in phase III clinical trials for renal cell and hepatocellular carcinomas.

Based on these promising findings, these small molecular inhibitors of VEGFR tyrosine kinase activity are being tested in the breast cancer setting. Also, the combination of antiangiogenic drugs with one another and with other biological agents is also being explored in an attempt to improve efficacy and to overcome the drug resistance observed in the initial studies of antiangiogenic agents. In addition, selecting patients for treatment on the basis of their clinical features and tumour characteristics may be essential in optimizing outcomes with these agents.

# Novel use of conventional chemotherapy as antiangiogenic agents

It is likely that over the next few years the designer agents discussed above will be supplemented by conventional chemotherapeutic agents, including cyclophosphamide, paclitaxel, doxorubicin and vincristine, which appear to have antitumour effects through interfering with new vessel formation when used at 'metronomic' doses. This is where chemotherapy is administered frequently at low doses, which avoids myelosuppression and other dose-limiting side effects and which would otherwise require rest periods, but this approach inhibits tumour growth indirectly by damaging endothelial cells. This delivery strategy has several advantages over the conventional maximum tolerated dose approach, apart from reduced toxicity, in that a treatment response should occur irrespective of the resistance profile of the tumour cell population. Thus far, only a few clinical trials have tested this antiangiogenic schedule chemotherapy [115-118], and the findings of these studies suggest that tumour associated endothelial cells may be sensitive to protracted low-dose chemotherapy. Other chemotherapeutic agents, for example the camptothecin analogues, have also been shown to modulate angiogenesis as a secondary mechanism of action [119]. Placlitaxel, a microtubule inhibitor that is an active agent in the treatment of many different cancers, was shown to possess antiangiogenic properties that are independent of its antiproliferative action in in vivo models [120]. The level of expression of thymidine phosphorylase, a migration but nonmitogenic angiogenic enzyme that converts thymidine to thymine and 2-deoxyribose, may enhance survival in breast cancer through at least two mechanisms [121-123]. The first of these is by activation of intravenous 5-fluoruracil or oral capecitabine through conversion to active metabolites; the second mechanism is by abrogating thymidine rescue in methotrexate regimens and therefore salvaging methotrexate block on de novo DNA synthesis.

In addition to conventional chemotherapeutic drugs that have antivascular effects, hormonal therapies such as tamoxifen may also have antiangiogenetic properties. Oestrogen is known to enhance VEGF expression (which may be partly HIF mediated [124]), and tamoxifen inhibits VEGF and FGF stimulated angiogenesis, resulting in a decrease in microvessel density and an increase in necrosis in MCF-7

xenografts [125-130]. Tamoxifen may also downregulate the angiogenic inhibitor thrombospondin [131]. Other drugs that may be of interest in this setting are the cyclo-oxygenase-2 inhibitors and biphosphonates, which also appear to have antiangiogenic potency [132-134].

#### Other targets

At the time of writing there are 30 agents on the National Cancer Institute website included in antiangiogenesis trials that interfere with the neovascularization process at many levels. Many promising novel targets have yet to reach this stage of development, the discussion of which is beyond the scope of this review. However, two of these that are of great interest are agents targeting the TAMs (which act as 'conductors' of angiogenesis) and Notch signalling (which is involved in cell differentiation, proliferation and apoptosis).

TAMs in breast cancers are markers of poor prognosis [135]. Evidence suggests that tumours recruit and use macrophage functions to promote tumour growth and metastasis. They achieve this through modulation of immune function, matrix degradation, growth factor production and angiogenesis. TAMs are recruited to avascular areas in breast tumours probably through hypoxic stimulation, in which they can release a variety of potent angiogenic factors, including VEGF (itself a chemoattractant for macrophages), thymidine phosphorylase, cyclo-oxygenase-2 and tumour necrosis factorα. Matrix metalloproteinases such as urokinase plasminogen activator (also a prognostic factor in breast cancer) from TAMs can further increase local VEGF levels through cleavage from the matrix. Thus, the multifunctional and pivotal TAMs are a target for therapy. Indeed, macrophages are regulated by oestrogen, and crosstalk between the oestrogen receptor and cytokine-mediated pathways provide a potential role for selective oestrogen receptor modulators in prevention and/or treatment of breast cancer [136].

Notch signalling plays an important oncogenic role in breast tumour development in animal models. It is also significant in human breast cancers [137], with upregulation of several of the Notch pathway components occurring in human breast cancer cells; also, pathway members are expressed by endothelial cells in breast cancers. The Notch pathway is critical for angiogenesis, with mutations being associated with abnormal vascular development. Interestingly, like VEGF, haploinsufficiency for the endothelial-specific Notch ligand Delta-like 4 in mice is embryonic lethal. In view of the significance of Notch signalling in oncogenesis and tumour neovascularization, the pathway is a promising target for treatment. The pathway is complex but several points in the pathway may be target (reviewed by Shi and Harris [137]). Central to Notch activation is γ-secretase, which cleaves Notch, allowing its translocation to the nucleus where it activates target genes. Thus, inhibiting  $\gamma$ -secretase function would prevent Notch signal transduction; γ-secretase inhibitors have been developed that perform this function.

#### Conclusion

It has been more than 35 years since Judah Folkman suggested that the tumour vasculature would be a target for anticancer therapy, and in the interim there has been a huge increase in our understanding of the biology underlying tumour angiogenesis. Unfortunately, early enthusiasm for this approach based on strong preclinical data has not transferred simply to the clinic. Nevertheless, the latest generation of agents provides reason for optimism, such that antiangiogenic therapies are being integrated into routine oncology practice. There is still much to learn, with the full complexity of the mechanisms of tumour neovascularization and their regulators still to be defined, not only in individual tumour types but also in individual patients. Thus, more information in terms of biomarkers that are predictive of response is required so that tailored treatment can be offered. A huge amount of data should become available over the next few years that should help us to use these agents in the most effective manner.

## **Competing interests**

The authors declare that they have no competing interests.

### References

- Miller KD: Recent translational research: antiangiogenic therapy for breast cancer: where do we stand? Breast Cancer Res 2004, 6:128-132.
- Kerbel RS: A cancer therapy resistant to resistance. Nature 1997. 390:335-336.
- Relf M, LeJeune S, Scott PA, Fox S, Smith K, Leek R, Moghaddam A, Whitehouse R, Bicknell R, Harris AL: Expression of the angiogenic factors vascular endothelial cell growth factor, acidic and basic fibroblast growth factor, tumor growth factor beta-1, platelet-derived endothelial cell growth factor, placenta growth factor, and pleiotrophin in human primary breast cancer and its relation to angiogenesis. Cancer Res 1997, 57: 963-969
- Folkman J: Angiogenesis in cancer, vascular, rheumatoid and other disease. Nat Med 1995, 1:27-31.
- Hanahan D, Folkman J: Patterns and emerging mechanisms of the angiogenic switch during tumorigenesis. Cell 1996, 86: 353-364.
- Skobe M, Rockwell P, Goldstein N, Vosseler S, Fusenig NE: Halting angiogenesis suppresses carcinoma cell invasion. Nat Med 1997, 3:1222-1227.
- Brem SS, Jensen HM, Gullino PM: Angiogenesis as a marker of preneoplastic lesions of the human breast. Cancer 1978, 41: 239-244
- Jensen HM, Chen I, De VM, Lewis AE: Angiogenesis induced by 'normal' human breast tissue: a probable marker for precancer. Science 1982, 218:293-295.
- Lichtenbeld HC, Barendsz-Janson AF, van Essen H, Struijker Boudier H, Griffioen AW, Hillen HF: Angiogenic potential of malignant and non-malignant human breast tissues in an in vivo angiogenesis model. Int J Cancer 1998, 77:455-459.
- Deng G, Lu Y, Zlotnikov G, Thor A, Smith HS: Loss of heterozygosity in normal tissue adjacent to breast carcinomas. Science 1996, 274:2057-2059.
- Engels K, Fox SB, Whitehouse RM, Gatter KC, Harris AL: Distinct angiogenic patterns are associated with high-grade in situ ductal carcinomas of the breast. J Pathol 1997, 181:207-212.
- Guidi A, Fischer L, Harris J, Schnitt S: Microvessel density and distribution in ductal carcinoma in situ of the breast. J Natl Cancer Inst 1994, 86:614-619.
- Marson LP, Kurian KM, Miller WR, Dixon JM: The effect of tamoxifen on breast tumour vascularity. Breast Cancer Res Treat 2001, 66:9-15.
- 14. Gasparini G, Fox SB, Verderio P, Bonoldi E, Bevilacqua P, Borac-

- chi P, Dante S, Marubini E, Harris AL: Determination of angiogenesis adds information to estrogen receptor status in predicting the efficacy of adjuvant tamoxifen in node-positive breast cancer patients. Clin Cancer Res 1996, 2:1191-1198.
- Macaulay V, Fox SB, Zhang H, Whitehouse RM, Leek RD, Gatter K, Bicknell R, Harris AL: Breast cancer angiogenesis and tamoxifen resistence. Endocr Rel Cancer 1995. 2:1-8.
- Gasparini G, Toi M, Verderio P, Ranieri G, Dante S, Bonoldi E, Boracchi P, Fanelli M, Tominaga T: Prognostic significance of p53, angiogenesis, and other conventional features in operable breast cancer: subanalysis in node-positive and nodenegative patients. *Int J Oncol* 1998, 12:1117-1125.
- 17. Gasparini G, Biganzoli E, Bonoldi E, Morabito A, Fanelli M, Boracchi P: Angiogenesis sustains tumor dormancy in patients with breast cancer treated with adjuvant chemotherapy. *Breast Cancer Res Treat* 2001, **65**:71-75.
- Fox SB, Leek RD, Bliss J, Mansi JL, Gusterson B, Gatter KC, Harris AL: Association of tumor angiogenesis with bone marrow micrometastases in breast cancer patients. J Natl Cancer Inst 1997, 89:1044-1049.
- Fox SB: Quantitative angiogenesis in breast cancer. Methods Mol Med 2006, 120:161-187.
- Uzzan B, Nicolas P, Cucherat M, Perret GY: Microvessel density as a prognostic factor in women with breast cancer: a systematic review of the literature and meta-analysis. Cancer Res 2004, 64:2941-2955.
- Arbiser JL, Moses MA, Fernandez CA, Ghiso N, Cao Y, Klauber N, Frank D, Brownlee M, Flynn E, Parangi S, Byers HR, Folkman J: Oncogenic H-ras stimulates tumor angiogenesis by two distinct pathways. Proc Natl Acad Sci USA 1997, 94:861-866.
- Okada F, Rak JW, Croix BS, Lieubeau B, Kaya M, Roncari L, Shirasawa S, Sasazuki T, Kerbel RS: Impact of oncogenes in tumor angiogenesis: mutant K-ras up-regulation of vascular endothelial growth factor/vascular permeability factor is necessary, but not sufficient for tumorigenicity of human colorectal carcinoma cells. Proc Natl Acad Sci USA 1998, 95: 3609-3614.
- Dameron KM, Volpert OV, Tainsky MA, Bouck N: The p53 tumor suppressor gene inhibits angiogenesis by stimulating the production of thrombospondin. Cold Spring Harb Symp Quant Biol 1994, 59:483-489.
- Rodriguez-Manzaneque JC, Lane TF, Ortega MA, Hynes RO, Lawler J, Iruela-Arispe ML: Thrombospondin-1 suppresses spontaneous tumor growth and inhibits activation of matrix metalloproteinase-9 and mobilization of vascular endothelial growth factor. Proc Natl Acad Sci USA 2001, 98:12485-12490.
- Vaupel P, Mayer A, Briest S, Hockel M: Hypoxia in breast cancer: role of blood flow, oxygen diffusion distances, and anemia in the development of oxygen depletion. Adv Exp Med Biol 2005, 566:333-342.
- 26. Harris AL: Hypoxia: a key regulatory factor in tumour growth. Nat Rev Cancer 2002, 2:38-47.
- Maxwell PH: The HIF pathway in cancer. Semin Cell Dev Biol 2005, 16:523-530.
- Hewitson KS, McNeill LA, Riordan MV, Tian YM, Bullock AN, Welford RW, Elkins JM, Oldham NJ, Bhattacharya S, Gleadle JM, et al.: Hypoxia-inducible factor (HIF) asparagine hydroxylase is identical to factor inhibiting HIF (FIH) and is related to the cupin structural family. J Biol Chem 2002, 277:26351-26355.
- Fox SB, Bragança J, Turley H, Campo L, Han C, Gatter KC, Bhattacharya S, Harris AL: CITED4 inhibits hypoxia-activated transcription in cancer cells, and its cytoplasmic location in breast cancer is associated with elevated expression of tumor cell hypoxia-inducible factor 10. Cancer Res 2004, 64:6075-6081.
- Bos R, van der Groep P, Greijer AE, Shvarts A, Meijer S, Pinedo HM, Semenza GL, van Diest PJ, van der Wall E: Levels of hypoxia-inducible factor-1alpha independently predict prognosis in patients with lymph node negative breast carcinoma. Cancer 2003, 97:1573-1581.
- Schindl M, Schoppmann SF, Samonigg H, Hausmaninger H, Kwasny W, Gnant M, Jakesz R, Kubista E, Birner P, Oberhuber G; Austrian Breast and Colorectal Cancer Study Group: Overexpression of hypoxia-inducible factor 1alpha is associated with an unfavorable prognosis in lymph node-positive breast cancer. Clin Cancer Res 2002, 8:1831-1837.
- Trastour C, Benizri E, Ettore F, Ramaioli A, Chamorey E, Pouysségur J, Berra E: HIF-1alpha and CA IX staining in inva-

- sive breast carcinomas: prognosis and treatment outcome. *Int J Cancer* 2007, **120**:1451-1458.
- Dales JP, Garcia S, Meunier-Carpentier S, Andrac-Meyer L, Haddad O, Lavaut MN, Allasia C, Bonnier P, Charpin C: Overexpression of hypoxia-inducible factor HIF-1alpha predicts early relapse in breast cancer: retrospective study in a series of 745 patients. Int J Cancer 2005, 116:734-739.
   Wiesener MS, Jürgensen JS, Rosenberger C, Scholze CK,
- Wiesener MS, Jürgensen JS, Rosenberger C, Scholze CK, Hörstrup JH, Warnecke C, Mandriota S, Bechmann I, Frei UA, Pugh CW, et al.: Widespread hypoxia-inducible expression of HIF-2alpha in distinct cell populations of different organs. FASEB J 2003, 17:271-273.
- Maynard MA, Qi H, Chung J, Lee EH, Kondo Y, Hara S, Conaway RC, Conaway JW, Ohh M: Multiple splice variants of the human HIF-3 alpha locus are targets of the von Hippel-Lindau E3 ubiquitin ligase complex. J Biol Chem 2003, 278:11032-11040
- Raval RR, Lau KW, Tran MG, Sowter HM, Mandriota SJ, Li JL, Pugh CW, Maxwell PH, Harris AL, Ratcliffe PJ: Contrasting properties of hypoxia-inducible factor 1 (HIF-1) and HIF-2 in von Hippel-Lindau-associated renal cell carcinoma. Mol Cell Biol 2005. 25:5675-5686.
- 37. Ratcliffe PJ: HIF-1 and HIF-2: working alone or together in hypoxia? J Clin Invest 2007, 117:862-865.
- Giatromanolaki A, Sivridis E, Fiska A, Koukourakis MI: Hypoxiainducible factor-2 alpha (HIF-2 alpha) induces angiogenesis in breast carcinomas. Appl Immunohistochem Mol Morphol 2006, 14:78-82.
- Leek RD, Talks KL, Pezzella F, Turley H, Campo L, Brown NS, Bicknell R, Taylor M, Gatter KC, Harris AL: Relation of hypoxiainducible factor-2 alpha (HIF-2 alpha) expression in tumorinfiltrative macrophages to tumor angiogenesis and the oxidative thymidine phosphorylase pathway in Human breast cancer. Cancer Res 2002, 62:1326-1329.
- Generali D, Berruti A, Brizzi MP, Campo L, Bonardi S, Wigfield S, Bersiga A, Allevi G, Milani M, Aguggini S, et al.: Hypoxiainducible factor-1alpha expression predicts a poor response to primary chemoendocrine therapy and disease-free survival in primary human breast cancer. Clin Cancer Res 2006, 12: 4562-4568.
- Semenza GL: Targeting HIF-1 for cancer therapy. Nat Rev Cancer 2003, 3:721-732.
- Kung AL, Wang S, Klco JM, Kaelin WG, Livingston DM: Suppression of tumor growth through disruption of hypoxia-inducible transcription. *Nat Med* 2000, 6:1335-1340.
- 43. Rischin D, Peters L, Fisher R, Macann A, Denham J, Poulsen M, Jackson M, Kenny L, Penniment M, Corry J, et al.: Tirapazamine, cisplatin, and radiation versus fluorouracil, cisplatin, and radiation in patients with locally advanced head and neck cancer: a randomized phase II trial of the Trans-Tasman Radiation Oncology Group (TROG 98.02). J Clin Oncol 2005, 23:79-87.
- Nagasawa H, Yamashita M, Mikamo N, Shimamura M, Oka S, Uto Y, Hori H: Design, synthesis and biological activities of antiangiogenic hypoxic cytotoxin, triazine-N-oxide derivatives. Comp Biochem Physiol A Mol Integr Physiol 2002, 132:33-40.
- Boogaerts M, Mittelman M, Vaupel P: Beyond anaemia management: evolving role of erythropoietin therapy in neurological disorders, multiple myeloma and tumour hypoxia models.
   Oncology 2005, Suppl 2:22-30.
- Hlatky L, Hahnfeldt P, Folkman J: Clinical application of antiangiogenic therapy: microvessel density, what it does and doesn't tell us. J Natl Cancer Inst 2002, 94:883-893.
- Vaupel P, Kallinowski F, Okunieff P. Blood flow, oxygen consumption and tissue oxygenation of human tumors. Adv Exp Med Biol 1990, 277:895-905.
- Fox SB, Gatter KC, Bicknell R, Going JJ, Stanton P, Cooke TG, Harris AL: Relationship of endothelial cell proliferation to tumor vascularity in human breast cancer. Cancer Res 1993, 53:9161-9163.
- Kakolyris S, Giatromanolaki A, Koukourakis M, Leigh IM, Georgoulias V, Kanavaros P, Sivridis E, Gatter KC, Harris AL: Assessment of vascular maturation in non-small cell lung cancer using a novel basement membrane component, LH39: correlation with p53 and angiogenic factor expression. Cancer Res 1999, 59:5602-5607.
- Kakolyris S, Fox SB, Koukourakis M, Giatromanolaki A, Brown N, Leek RD, Taylor M, Leigh IM, Gatter KC, Harris AL: Relationship

- of vascular maturation in breast cancer blood vessels to vascular density and metastasis, assessed by expression of a novel basement membrane component, LH39. *Br J Cancer* 2000. **82**:844-851.
- Holash J, Maisonpierre PC, Compton D, Boland P, Alexander CR, Zagzag D, Yancopoulos GD, Wiegand SJ: Vessel cooption, regression, and growth in tumors mediated by angiopoietins and VEGF. Science 1999, 284:1994-1998.
- Holash J, Wiegand SJ, Yancopoulos GD: New model of tumor angiogenesis: dynamic balance between vessel regression and growth mediated by angiopoietins and VEGF. Oncogene 1999, 18:5356-5362.
- Patan S, Munn LL, Jain RK: Intussusceptive microvascular growth in a human colon adenocarcinoma xenograft: a novel mechanism of tumor angiogenesis. *Microvasc Res* 1996, 51: 260-272.
- Patan S: Vasculogenesis and angiogenesis as mechanisms of vascular network formation, growth and remodeling. J Neurooncol 2000, 50:1-15.
- Asahara T, Masuda H, Takahashi T, Kalka C, Pastore C, Silver M, Kearne M, Magner M, Isner JM: Bone marrow origin of endothelial progenitor cells responsible for postnatal vasculogenesis in physiological and pathological neovascularization. Circ Res 1999, 85:221-228.
- Gunsilius E, Duba HC, Petzer AL, Kähler CM, Grünewald K, Stockhammer G, Gabl C, Dirnhofer S, Clausen J, Gastl G: Evidence from a leukaemia model for maintenance of vascular endothelium by bone-marrow-derived endothelial cells. Lancet 2000, 355:1688-1691.
- Rafii S: Circulating endothelial precursors: mystery, reality, and promise. J Clin Invest 2000, 105:17-19.
- Asahara T, Takahashi T, Masuda H, Kalka C, Chen D, Iwaguro H, Inai Y, Silver M, Isner JM: VEGF contributes to postnatal neovascularization by mobilizing bone marrow-derived endothelial progenitor cells. EMBO J 1999, 18:3964-3972.
- Peters BA, Diaz LA, Polyak K, Meszler L, Romans K, Guinan EC, Antin JH, Myerson D, Hamilton SR, Vogelstein B, et al.: Contribution of bone marrow-derived endothelial cells to human tumor vasculature. Nat Med 2005, 11:261-262.
- Young PP, Vaughan DE, Hatzopoulos AK: Biologic properties of endothelial progenitor cells and their potential for cell therapy. Prog Cardiovasc Dis 2007, 49:421-429.
- Lyden D, Hattori K, Dias S, Costa C, Blaikie P, Butros L, Chadburn A, Heissig B, Marks W, Witte L, et al.: Impaired recruitment of bone-marrow-derived endothelial and hematopoietic precursor cells blocks tumor angiogenesis and growth. Nat Med 2001, 7:1194-1201.
- Ziegelhoeffer T, Fernandez B, Kostin S, Heil M, Voswinckel R, Helisch A, Schaper W: Bone marrow-derived cells do not incorporate into the adult growing vasculature. Circ Res 2004, 94:230-238
- Nolan DJ, Ciarrocchi A, Mellick AS, Jaggi JS, Bambino K, Gupta S, Heikamp E, McDevitt MR, Scheinberg DA, Benezra R, et al.: Bone marrow-derived endothelial progenitor cells are a major determinant of nascent tumor neovascularization. Genes Dev 2007, 21:1546-1558.
- Grunewald M, Avraham I, Dor Y, Bachar-Lustig E, Itin A, Jung S, Chimenti S, Landsman L, Abramovitch R, Keshet E: VEGFinduced adult neovascularization: recruitment, retention, and role of accessory cells. Cell 2006, 124:175-189.
- Ceradini DJ, Kulkarni AR, Callaghan MJ, Tepper OM, Bastidas N, Kleinman ME, Capla JM, Galiano RD, Levine JP, Gurtner GC: Progenitor cell trafficking is regulated by hypoxic gradients through HIF-1 induction of SDF-1. Nat Med 2004, 10:858-864.
- Shirakawa K, Wakasugi H, Heike Y, Watanabe I, Yamada S, Saito K, Konishi F: Vasculogenic mimicry and pseudo-comedo formation in breast cancer. *Int J Cancer* 2002, 99:821-828.
- Brat DJ, Van Meir EG: Glomeruloid microvascular proliferation orchestrated by VPF/VEGF: a new world of angiogenesis research. Am J Pathol 2001, 158:789-796.
- Straume O, Chappuis PO, Salvesen HB, Halvorsen OJ, Haukaas SA, Goffin JR, Bégin LR, Foulkes WD, Akslen LA: Prognostic importance of glomeruloid microvascular proliferation indicates an aggressive angiogenic phenotype in human cancers. Cancer Res 2002, 62:6808-6811.
- Sundberg C, Nagy JA, Brown LF, Feng D, Eckelhoefer IA, Manseau EJ, Dvorak AM, Dvorak HF: Glomeruloid microvascular

- proliferation follows adenoviral vascular permeability factor/vascular endothelial growth factor-164 gene delivery. *Am J Pathol* 2001. **158**:1145-1160.
- Zhang HT, Scott PA, Morbidelli L, Peak S, Moore J, Turley H, Harris AL, Ziche M, Bicknell R: The 121 amino acid isoform of vascular endothelial growth factor is more strongly tumorigenic than other splice variants in vivo. Br J Cancer 2000, 83: 63-68
- 71. Dome B, Hendrix MJ, Paku S, Tovari J, Timar J: Alternative vascularization mechanisms in cancer: Pathology and therapeutic implications. *Am J Pathol* 2007, **170**:1-15.
- Sood AK, Seftor EA, Fletcher MS, Gardner LM, Heidger PM, Buller RE, Seftor RE, Hendrix MJ: Molecular determinants of ovarian cancer plasticity. Am J Pathol 2001, 158:1279-1288.
- Warren B: The vascular morphology of tumors. In Tumor Blood Circulation. Edited by Peterson H. Boca Raton, FL: CRC Press; 1979:1-47.
- Chang YS, di Tomaso E, McDonald DM, Jones R, Jain RK, Munn LL: Mosaic blood vessels in tumors: frequency of cancer cells in contact with flowing blood. Proc Natl Acad Sci USA 2000, 97:14608-14613.
- 75. Folberg R, Hendrix MJ, Maniotis AJ: Vasculogenic mimicry and tumor angiogenesis. *Am J Pathol* 2000, **156**:361-381.
- McDonald DM, Munn L, Jain RK: Vasculogenic mimicry: how convincing, how novel, and how significant? Am J Pathol 2000, 156:383-388
- Greenblatt M, Shubik P: Tumour angiogenesis: transfilter diffusion studies in the hamster by the transparent chamber technique. J Natl Cancer Inst 1968, 41:111-124.
- Ehrmann RL, Knoth M: Choriocarcinoma. Transfilter stimulation of vasoproliferation in the hamster cheek pouch. Studied by light and electron microscopy. J Natl Cancer Inst 1968, 41: 1329-1341
- Folkman J, Merler E, Abernathy C, Williams G: Isolation of a tumor factor responsible for angiogenesis. J Exp Med 1971, 133:275-288.
- Neufeld G, Kessler O: Pro-angiogenic cytokines and their role in tumor angiogenesis. Cancer Metastasis Rev 2006, 25:373-385.
- Roskoski R Jr: Vascular endothelial growth factor (VEGF) signaling in tumor progression. Crit Rev Oncol Hematol 2007, 62: 179-213
- 82. Fox SB, Gasparini G, Harris AL: Angiogenesis: pathological, prognostic, and growth-factor pathways and their link to trial design and anticancer drugs. *Lancet Oncol* 2001, 2:278-289.
- Nikitenko LL, Fox SB, Kehoe S, Rees MC, Bicknell R: Adrenomedullin and tumour angiogenesis. Br J Cancer 2006, 94:1-7.
- 84. Engels K, Fox SB, Whitehouse RM, Gatter KC, Harris AL: Upregulation of thymidine phosphorylase expression is associated with a discrete pattern of angiogenesis in ductal carcinomas in situ of the breast. J Pathol 1997, 182:414-420.
- 85. Fox SB, Westwood M, Moghaddam A, Comley M, Turley H, Whitehouse RM, Bicknell R, Gatter KC, Harris AL: The angiogenic factor platelet-derived endothelial cell growth factor/thymidine phosphorylase is up-regulated in breast cancer epithelium and endothelium. Br J Cancer 1996, 73:275-280.
- Bielenberg DR, Pettaway CA, Takashima S, Klagsbrun M: Neuropilins in neoplasms: expression, regulation, and function. Exp Cell Res 2006, 312:584-593.
- Kurebayashi J, Otsuki T, Kunisue H, Mikami Y, Tanaka K, Yamamoto S, Sonoo H: Expression of vascular endothelial growth factor (VEGF) family members in breast cancer. *Jpn J Cancer Res* 1999, 90:977-981.
- Okada K, Osaki M, Araki K, Ishiguro K, Ito H, Ohgi S: Expression of hypoxia-inducible factor (HIF-1alpha), VEGF-C and VEGF-D in non-invasive and invasive breast ductal carcinomas. Anticancer Res 2005, 25:3003-3009.
- Gunningham SP, Currie MJ, Han C, Robinson BA, Scott PA, Harris AL, Fox SB: The short form of the alternatively spliced flt-4 but not its ligand VEGF-C is related to lymph node metastasis in human breast cancers. Clin Cancer Res 2000, 6: 4278-4286.
- Currie MJ, Hanrahan V, Gunningham SP, Morrin HR, Frampton C, Han C, Robinson BA, Fox SB: Expression of vascular endothelial growth factor D is associated with hypoxia inducible factor (HIF-1alpha) and the HIF-1alpha target gene DEC1, but not lymph node metastasis in primary human breast carcinomas.

- J Clin Pathol 2004, 57:829-834.
- Nakamura Y, Yasuoka H, Tsujimoto M, Yang Q, Imabun S, Nakahara M, Nakao K, Nakamura M, Mori I, Kakudo K: Prognostic significance of vascular endothelial growth factor D in breast carcinoma with long-term follow-up. Clin Cancer Res 2003, 9: 716-721.
- Kinoshita J, Kitamura K, Kabashima A, Saeki H, Tanaka S, Sugimachi K: Clinical significance of vascular endothelial growth factor-C (VEGF-C) in breast cancer. Breast Cancer Res Treat 2001, 66:159-164.
- 93. Yang W, Klos K, Yang Y, Smith TL, Shi D, Yu D: ErbB2 overexpression correlates with increased expression of vascular endothelial growth factors A, C, and D in human breast carcinoma. Cancer 2002, 94:2855-2861.
- Mylona E, Alexandrou P, Giannopoulou I, Liapis G, Sofia M, Keramopoulos A, Nakopoulou L: The prognostic value of vascular endothelial growth factors (VEGFs)-A and -B and their receptor, VEGFR-1, in invasive breast carcinoma. Gynecol Oncol 2007, 104:557-563.
- 95. Presta LG, Chen H, O'Connor SJ, Chisholm V, Meng YG, Krummen L, Winkler M, Ferrara N: Humanization of an anti-vascular endothelial growth factor monoclonal antibody for the therapy of solid tumors and other disorders. Cancer Res 1997, 57:4593-4599.
- Kim KJ, Li B, Winer J, Armanini M, Gillett N, Phillips HS, Ferrara N: Inhibition of vascular endothelial growth factor-induced angiogenesis suppresses tumour growth in vivo. Nature 1993, 362:841-844.
- Zhang W, Ran S, Sambade M, Huang X, Thorpe PE: A monoclonal antibody that blocks VEGF binding to VEGFR2 (KDR/Flk-1) inhibits vascular expression of Flk-1 and tumor growth in an orthotopic human breast cancer model. Angiogenesis 2002, 5:35-44.
- Wedam SB, Low JA, Yang SX, Chow CK, Choyke P, Danforth D, Hewitt SM, Berman A, Steinberg SM, Liewehr DJ, et al.: Antiangiogenic and antitumor effects of bevacizumab in patients with inflammatory and locally advanced breast cancer. J Clin Oncol 2006, 24:769-777.
- Miller KD: E2100: a phase III trial of paclitaxel versus paclitaxel/bevacizumab for metastatic breast cancer. Clin Breast Cancer 2003, 3:421-422.
- 100. Schneider BP, Miller KD: Angiogenesis of breast cancer. J Clin Oncol 2005, 23:1782-1790.
- 101. Schneider BP, Sledge GW Jr: Drug insight: VEGF as a therapeutic target for breast cancer. Nat Clin Pract Oncol 2007, 4: 181-189.
- 102. Miller KD, Trigo JM, Wheeler C, Barge A, Rowbottom J, Sledge G, Baselga J: A multicenter phase II trial of ZD6474, a vascular endothelial growth factor receptor-2 and epidermal growth factor receptor tyrosine kinase inhibitor, in patients with previously treated metastatic breast cancer. Clin Cancer Res 2005, 11:3369-3376.
- 103. Clarke K, Smith K, Gullick WJ, Harris AL: Mutant epidermal growth factor receptor enhances induction of vascular endothelial growth factor by hypoxia and insulin-like growth factor-1 via a PI3 kinase dependent pathway. Br J Cancer 2001, 84:1322-1329.
- 104. Maity A, Pore N, Lee J, Solomon D, O'Rourke DM: Epidermal growth factor receptor transcriptionally up-regulates vascular endothelial growth factor expression in human glioblastoma cells via a pathway involving phosphatidylinositol 3'-kinase and distinct from that induced by hypoxia. Cancer Res 2000, 60:5879-5886.
- 105. Yen L, You XL, Al Moustafa AE, Batist G, Hynes NE, Mader S, Meloche S, Alaoui-Jamali MA: Heregulin selectively upregulates vascular endothelial growth factor secretion in cancer cells and stimulates angiogenesis. Oncogene 2000, 19:3460-3469.
- 106. Konecny GE, Meng YG, Untch M, Wang HJ, Bauerfeind I, Epstein M, Stieber P, Vernes JM, Gutierrez J, Hong K, et al.: Association between HER-2/neu and vascular endothelial growth factor expression predicts clinical outcome in primary breast cancer patients. Clin Cancer Res 2004, 10:1706-1716.
- 107. Pegram MD, Konecny GE, O'Callaghan C, Beryt M, Pietras R, Slamon DJ. Rational combinations of trastuzumab with chemotherapeutic drugs used in the treatment of breast cancer. J Natl Cancer Inst 2004, 96:739-749.
- 108. Sandberg JA, Parker VP, Blanchard KS, Sweedler D, Powell JA,

- Kachensky A, Bellon L, Usman N, Rossing T, Borden E, et al.: Pharmacokinetics and tolerability of an antiangiogenic ribozyme (ANGIOZYME) in healthy volunteers. J Clin Pharmacol 2000, 40:1462-1469.
- 109. Im SA, Kim JS, Gomez-Manzano C, Fueyo J, Liu TJ, Cho MS, Seong CM, Lee SN, Hong YK, Yung WK: Inhibition of breast cancer growth in vivo by antiangiogenesis gene therapy with adenovirus-mediated antisense-VEGF. Br J Cancer 2001, 84: 1252-1257.
- 110. Weng DE, Usman N: Angiozyme: a novel angiogenesis inhibitor. Curr Oncol Rep 2001, 3:141-146.
- 111. Bergsland EK: Update on clinical trials targeting vascular endothelial growth factor in cancer. Am J Health Syst Pharm 2004, Suppl 5:S12-S20.
- 112. Wood JM, Bold G, Buchdunger E, Cozens R, Ferrari S, Frei J, Hofmann F, Mestan J, Mett H, O'Reilly T, et al.: PTK787/ZK 222584, a novel and potent inhibitor of vascular endothelial growth factor receptor tyrosine kinases, impairs vascular endothelial growth factor-induced responses and tumor growth after oral administration. Cancer Res 2000, 60:2178-2189.
- 113. Egeblad M, Werb Z: New functions for the matrix metalloproteinases in cancer progression. Nat Rev Cancer 2002, 2:161-174.
- 114. Sparano JA, Bernardo P, Stephenson P, Gradishar WJ, Ingle JN, Zucker S, Davidson NE: Randomized phase III trial of marimastat versus placebo in patients with metastatic breast cancer who have responding or stable disease after first-line chemotherapy: Eastern Cooperative Oncology Group trial E2196. J Clin Oncol 2004, 22:4683-4690.
- 115. Bottini A, Generali D, Brizzi MP, Fox SB, Bersiga A, Bonardi S, Allevi G, Aguggini S, Bodini G, Milani M, et al.: Randomized phase II trial of letrozole and letrozole plus low-dose metronomic oral cyclophosphamide as primary systemic treatment in elderly breast cancer patients. J Clin Oncol 2006, 24:3623-3628
- 116. Engelsman E, Klijn JC, Rubens RD, Wildiers J, Beex LV, Nooij MA, Rotmensz N, Sylvester R: 'Classical' CMF versus a 3-weekly intravenous CMF schedule in postmenopausal patients with advanced breast cancer. An EORTC Breast Cancer Co-operative Group Phase III Trial (10808). Eur J Cancer 1991, 27:966-970
- 117. Colleoni M, Orlando L, Sanna G, Rocca A, Maisonneuve P, Peruzzotti G, Ghisini R, Sandri MT, Zorzino L, Nolè F, et al.: Metronomic low-dose oral cyclophosphamide and methotrexate plus or minus thalidomide in metastatic breast cancer: antitumor activity and biological effects. Ann Oncol 2006, 17:232-238
- 118. Colleoni M, Rocca A, Sandri MT, Zorzino L, Masci G, Nolè F, Peruzzotti G, Robertson C, Orlando L, Cinieri S, et al.: Low-dose oral methotrexate and cyclophosphamide in metastatic breast cancer: antitumor activity and correlation with vascular endothelial growth factor levels. Ann Oncol 2002, 13:73-80.
- 119. O'Leary JJ, Shapiro RL, Ren CJ, Chuang N, Cohen HW, Potmesil M: Antiangiogenic effects of camptothecin analogues 9-amino-20(S)-camptothecin, topotecan, and CPT-11 studied in the mouse cornea model. Clin Cancer Res 1999, 5:181-187.
- 120. Klauber N, Parangi S, Flynn E, Hamel E, D'Amato RJ: Inhibition of angiogenesis and breast cancer in mice by the microtubule inhibitors 2-methoxyestradiol and taxol. Cancer Res 1997, 57: 81-86.
- 121. Fox S, Engels K, Comley M, Whitehouse R, Turley H, Gatter K, Harris AL: Relationship of elevated tumour thymidine phosphorylase in node positive breast carcinomas to the effects of adjuvant CMF. Annal Oncol 1997, 8:271-275.
- 122. Schwartz EL, Baptiste N, Wadler S, Makower D: Thymidine phosphorylase mediates the sensitivity of human colon carcinoma cells to 5-fluorouracil. J Biol Chem 1995, 270:19073-19077.
- 123. Haraguchi M, Furukawa T, Sumizawa T, Akiyama S: Sensitivity of human KB cells expressing platelet-derived endothelial cell growth factor to pyrimidine antimetabolites. Cancer Res 1993, 53:5680-5682.
- 124. Kazi AA, Koos RD: Estrogen-induced activation of hypoxiainducible factor 1α (HIF-1α), vascular endothelial growth factor (VEGF) expression, and edema in the uterus are mediated by the phosphatidylinositol 3-kinase (PI3K)/Akt pathway.

- Endocrinology 2007, 148:2363-2374.
- 125. Ruohola JK, Valve EM, Karkkainen MJ, Joukov V, Alitalo K, Harkonen PL: Vascular endothelial growth factors are differentially regulated by steroid hormones and antiestrogens in breast cancer cells. *Mol Cell Endocrinol* 1999, 149:29-40.
- 126. Mueller MD, Vigne JL, Minchenko A, Lebovic DI, Leitman DC, Taylor RN: Regulation of vascular endothelial growth factor (VEGF) gene transcription by estrogen receptors alpha and beta. Proc Natl Acad Sci USA 2000, 97:10972-10977.
- 127. McNamara DA, Harmey J, Wang JH, Kay E, Walsh TN, Bouchier-Hayes DJ. Tamoxifen inhibits endothelial cell proliferation and attenuates VEGF-mediated angiogenesis and migration in vivo. Eur J Surg Oncol 2001, 27:714-718.
- 128. Garvin S, Nilsson UW, Dabrosin C: Effects of oestradiol and tamoxifen on VEGF, soluble VEGFR-1, and VEGFR-2 in breast cancer and endothelial cells. *Br J Cancer* 2005, **93**:1005-1010.
- 129. Morena AM, Oshima CT, Gebrim LH, Egami MI, Silva MR, Segreto RA, Giannotti Filho O, Teixeira VP, Segreto HR: Early nuclear alterations and immunohistochemical expression of Ki-67, Erb-B2, vascular endothelial growth factor (VEGF), transforming growth factor (TGF-beta1) and integrine-linked kinase (ILK) two days after tamoxifen in breast carcinoma. Neoplasma 2004, 51:481-486.
- 130. Hyder SM: Sex-steroid regulation of vascular endothelial growth factor in breast cancer. Endocr Relat Cancer 2006, 13: 667-687.
- 131. Silva ID, Salicioni AM, Russo IH, Higgy NA, Gebrim LH, Russo J: Tamoxifen down-regulates CD36 messenger RNA levels in normal and neoplastic human breast tissues. Cancer Res 1997, 57:378-81.
- 132. Nie D, Honn KV: Eicosanoid regulation of angiogenesis in tumors. Semin Thromb Hemost 2004, 30:119-125.
- 133. Wood J, Bonjean K, Ruetz S, Bellahcène A, Devy L, Foidart JM, Castronovo V, Green JR: Novel antiangiogenic effects of the bisphosphonate compound zoledronic acid. J Pharmacol Exp Ther 2002, 302:1055-1061.
- 134. Santini D, Vincenzi B, Dicuonzo G, Avvisati G, Massacesi C, Battistoni F, Gavasci M, Rocci L, Tirindelli MC, Altomare V, et al.: Zoledronic acid induces significant and long-lasting modifications of circulating angiogenic factors in cancer patients. Clin Cancer Res 2003, 9:2893-2897.
- 135. Leek RD, Harris AL: Tumor-associated macrophages in breast cancer. J Mammary Gland Biol Neoplasia 2002, 7:177-189.
- 136. Harkonen PL, Vaananen HK: Monocyte-macrophage system as a target for estrogen and selective estrogen receptor modulators. Ann N Y Acad Sci 2006, 1089:218-227.
- 137. Shi W, Harris AL: Notch signaling in breast cancer and tumor angiogenesis: cross-talk and therapeutic potentials. J Mammary Gland Biol Neoplasia 2006, 11:41-52.