Diabetes Metab J 2011;35:309-316 http://dx.doi.org/10.4093/dmj.2011.35.4.309 pISSN 2233-6079 · eISSN 2233-6087 DIABETES & METABOLISM JOURNAL

Glycosphingolipid Modification: Structural Diversity, Functional and Mechanistic Integration of Diabetes

Tadashi Yamashita^{1,2}

¹Graduate School of Advanced Life Science, Hokkaido University, Sapporo, Japan, ²World Class University Program, Kyungpook National University School of Medicine, Daegu, Korea

Glycosphingolipids (GSLs) are present in all mammalian cell plasma membranes and intracellular membrane structures. They are especially concentrated in plasma membrane lipid domains that are specialized for cell signaling. Plasma membranes have typical structures called rafts and caveola domain structures, with large amounts of sphingolipids, cholesterol, and sphingomyelin. GSLs are usually observed in many organs ubiquitously. However, GSLs, including over 400 derivatives, participate in diverse cellular functions. Several studies indicate that GSLs might have an effect on signal transduction related to insulin receptors and epidermal growth factor receptors. GSLs may modulate immune responses by transmitting signals from the exterior to the interior of the cell. Guillain-Barré syndrome is one of the autoimmune disorders characterized by symmetrical weakness in the muscles of the legs. The targets of the immune response are thought to be gangliosides, which are one group of GSLs. Other GSLs may serve as second messengers in several signaling pathways that are important to cell survival or programmed cell death. In the search for clear evidence that GSLs may play critical roles in various biological functions, many researchers have made genetically engineered mice. Before the era of gene manipulation, spontaneous animal models or chemical-induced disease models were used.

Keywords: Cell membrane; Diabetes mellitus; Glycosphingolipids; Mouse model

INTRODUCTION

Glycosphingolipids (GSLs) are ubiquitous cell membrane components of eukaryotic cells [1,2]. They are amphipathic molecules consisting of a ceramide and sugar chains, including one or more glucose, galactose, and sialic acid residues, as shown Fig. 1. The ceramide portion is hydrophobic, and the sugar portion is hydrophilic as shown Fig. 2. These molecules are embedded in the leaflet of the membrane, as shown Fig. 3. GSLs are also present in intracellular components such as the Golgi apparatus, nuclear membrane, and mitochondria. GSLs, along with the associated membrane components, circulate through these organelles. In the Golgi apparatus, GSLs are newly synthesized by the addition of saccharides one by one. On the other hand, in lysosomes, GSLs are degraded by the remov-

Corresponding author: Tadashi Yamashita Graduate School of Advanced Life Science, Hokkaido University, N21, W11, Sapporo 001-0021, Japan E-mail: ty11106@sci.hokudai.ac.jp al of saccharides one by one. These products are the result of enzyme reactions [3]. More than 400 species of GSLs possessing different sugar structures have been reported, although only seven monosaccharides have mainly been found in vertebrate GSLs.

Microdomains called lipid rafts are functional units in cell membranes [4,5]. Such microdomains in cell membranes consisting of GSL-cholesterol, function as platforms for the attachment of lipid-modified proteins, such as glycosylphosphatidylinositol (GPI)-anchored proteins [6]. These specialized membrane microdomains may organize the assembly of signaling molecules, membrane protein trafficking, and regulate neurotransmission and receptor trafficking [6]. Rietveld and Simons [7] found lipid rafts in model membranes to be related to the immiscibility of ordered and disordered liquid phases.

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/3.0/) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.



Fig. 1. The synthetic pathway from ceramide. Following the synthesis of glucosylceramide, sphingomyelin, galactosylceramide, ceramide-1-phosphate, and sphingosine-1-phosphate, the derivatives are synthesized by enzyme activities predominantly in the Golgi apparatus.



Fig. 2. Illustration of a eukaryotic cell membrane. It composed of the lipid bilayer, consisting of hydrophobic tails and hydrophilic heads.

The cause of this immiscibility is uncertain, but the immiscibility may have a constant effect in events such as phosphorylation of the signal transduction molecules. These events include phosphorylation by tyrosine kinases and/or serine/threonine kinases. Lipid rafts have been found by researchers to be involved in many signal transduction processes, including T



Fig. 3. Glycosphingolipids (GSLs), cholesterol, sphingomyelin, and glycoproteins present in the cell membrane. Some viruses and bacteria might catch and use these molecules as the entry gate into cells. Cell-cell interaction might also occur through these molecules.

cell antigen receptor signaling [8], B cell antigen receptor signaling [9], epidermal growth factor receptor signaling [10], and insulin receptor signaling [11,12]. In this review, I summarize our understanding of the role of GSLs on the insulin receptor abnormality as one of the pathophysiology of diabetes, and rodent models for the research on insulin resistance.

CHARACTERIZATION OF GSLS

GSLs in a subclass that contains sialic acid residues are known as gangliosides, and are abundantly present in the nervous system. Other subclasses of GSLs such as the globo series and isoglobo series, are abundant in red blood cells and leukocytes, respectively. Neolacto series and asialo series are also known. The synthesis of many GSLs starts from ceramide. Glucosylceramide (GlcCer) synthase, the first enzyme required for the synthesis of GSLs, is a transmembrane protein located in the endoplasmic reticulum (ER) with its C-terminal catalytic domain located in the cytoplasm [13]. Following the synthesis of glucosylceramide, lactosylceramide, which consists of ceramide, glucose, and galactose, is synthesized in the Golgi apparatus as a key molecule in this synthetic pathway. Through this molecule, higher-order GSL structures are produced by a series of Golgi glycosyltransferases. As a regular pathway, GlcCer is synthesized on the cytosolic side and must translocate across to the Golgi lumen for LacCer synthesis. However, recent evidence has shown that GlcCer destined for glycolipid synthesis follows a different pathway and is transported back into the ER via the late Golgi protein FAPP2 [14]. GSLs have been implicated in many fundamental cellular processes, including cell growth, migration, differentiation, morphogenesis, early development, and functioning of the nervous system [15-20]. GSLs seem to construct plasma membrane microdomains, known as rafts and caveolae [21,22], which are rich in sphingolipids, cholesterol, and sphingomyelin. As for metabolism, Whitmore et al. [23] demonstrated significant cell-to-cell heterogeneity in the amounts of gangliosides and their metabolites. GSL expression varies among and within cell populations, during cellular differentiation, and between normal cells and their transformed counterparts. For the regulation of GSL metabolism, Memon et al. [24], have reported that endotoxins (lipopolysaccharide, LPS) and cytokines enhance hepatic sphingolipid synthesis, increase the activity and mRNA levels of serine palmitoyltransferase, the first committed step in sphingolipid synthesis, and increase the content of sphingomyelin, ceramide, and GlcCer in circulating lipoproteins of Syrian hamsters. Tettamanti described the metabolic turnover or re-

http://e-dmj.org Diabetes Metab J 2011;35:309-316

cycling of GSLs in a review [25]. Briefly, the events to be considered are de novo biosynthesis, in situ chemical modifications at the plasma membrane level, direct recycling to the plasma membrane from early endosomes, sorting to the Golgi apparatus from endosomes with subsequent glycosylation, degradation at the lysosomal/late endosomal level, salvage pathways, and complete degradation to waste products. The half-lives of GSLs seems to range from 2 to 6.5 hours, to three days depending on the cells or tissues used.

GSLS, DIABETES, AND INSULIN RESISTANCE

Insulin is an anabolic hormone with regard to carbohydrate and lipid metabolism. Insulin exerts its effects via the insulin receptor (IR), a transmembrane receptor tyrosine kinase with two α -subunits and two β -subunits [26]. The α -subunits contain the insulin-binding sites. The β-subunits form transmembrane and intracellular parts of the receptor. Following insulin binding, the IR undergoes a conformational change that allows trans phosphorylation of the catalytic sites on multiple tyrosine residues [27]. The most important sites for the action of insulin are the liver, adipose tissues, and muscles. The concentration of IR is high in these tissues, with more than 200,000 receptors on adipocytes and hepatocytes [27]. On the other hand, insulin resistance is defined as the reduced ability of a cell to respond to physiological concentrations of insulin. As a result of insulin stimulation, insulin binds to the IR and induces autophosphorylation of tyrosine kinase under normal physiological conditions. Following autophosphorylation, at least two signaling cascades are known. In the first, Akt together with protein kinase C promotes translocation of the glucose transporter (GLUT-4) to the plasma membrane enabling the uptake of glucose in the bloodstream. In the second, the mitogen activated protein kinase cascade is initiated [28]. The PI(3)K/Akt signaling pathway is central to proper insulin signaling [29]. As for cholesterol, Simons and Toomre [30] had reported that cholesterol depletion inhibited raft-dependent signaling. Therefore, cholesterol in the cell membrane seems to act as a positive effector for IR and IRS-1 phosphorylation. What then are the effects of IR and IRS-1 phosphorylation by GSLs, which are components of the cell membrane like cholesterol? Generally, GSLs may play a prominent role in cell signaling, acting as both first and second messengers in a variety of signaling and regulatory pathways [31]. Of all the sphingolipids, ceramide and sphingosine, together with their phos-

phorylated counterparts, have received the most attention with regard to bioactivity. As one of the structures that GSLs participate in, lipid rafts are required for effective PI(3)K/Akt signaling. Moreover, Vainio et al. [32] have provided evidence that the IR is recruited to detergent-resistant domains upon ligand binding and the clustering of GM2 ganglioside inhibits IR signaling, apparently by excluding the ligand-bound IR from these domains. GM2 ganglioside may be important for the dynamic recruitment of the ligand-bound IR into the raft.

Diabetes mellitus type 2, which is known as non insulin-dependent diabetes mellitus or adult-onset diabetes, is a metabolic disorder that is characterized by a high glucose level in the bloodstream, insulin resistance, and insulin deficiency. Type 2 diabetes is a lifestyle- and genetic-related disease. The importance of various factors in the onset and pathological symptoms of the disease has been emphasized. There are many drugs with different mechanisms of action for this disease. For example, sulfonylureas lower blood sugar by stimulating the pancreas to release more insulin. Biguanides improve insulin's ability to move sugar into cells, especially into muscle cells. Thiazolidinediones improve insulin's effectiveness in muscle and fat tissues. Alpha-glucosidase inhibitors block enzymes that help digest starches, slowing the rise in blood sugar. Dipeptidyl peptidase IV inhibitors work to lower blood sugar in patients with type 2 diabetes by increasing insulin secretion from the pancreas and reducing sugar production. Studies have shown that some diabetes pills may help prevent diabetes and diabetes-related complications. On the hand, Ottenhoff et al. [33] reported that pharmacological inhibition of glucosylceramide synthase enhanced insulin sensitivity. They developed a highly specific, small molecule inhibitor of glucosylceramide synthase, N-(5'-adamantane-1'-yl-methoxy)-pentyl-1deoxynojirimycin (AMP-DNM). AMP-DNM counteracts TNF induced abnormalities in GSL concentrations and concomitantly reverses abnormalities in insulin signal transduction. Due to the pharmacological actions mentioned above, AMP-DNM significantly reduces GSL but not ceramide, concentrations in various tissues in mice and rats. Treatment of ob/ob mice with AMP-DNM normalizes their elevated tissue glucosylceramide levels, markedly lowers circulating glucose levels, improves oral glucose tolerance, reduces glycated hemoglobin, and improves insulin sensitivity in muscle and liver tissues. It is typically reported that the density of GSLs is closely related to insulin sensitivity in vivo. D-threo-1-Phenyl-2decanoylamino-3-morpholino-1-propanol (D-PDMP) is a

well-known inhibitor of UDP-glucose, which is the first step of GSL synthesis [34]. This inhibitor has long been employed to study the roles of GSLs. The inhibitor is transported to the ER and Golgi apparatus [35]. Following treatment with D-PDMP, nearly all GSL production is dramatically reduced on the surfaces of cells. In some cell lines, exogenously added Lac-Cer rescues blocking by D-PDMP.

ANIMAL MODELS OF INSULIN RESISTANCE

Rat models

Though many animal models of diabetes have been established, it is impossible to completely reproduce the condition of human patients. However, the knowledge gained from model animals contributes to our understanding of genetic diseases, the effects of drugs, elucidation of pathological conditions, prevention, and treatments. Diabetes is a result of the lack of insulin. Amongst rodents, high-calorie feeding can induce hypertriglyceridemia, insulin resistance, and hypertension. Sprague-Dawley (SD) rats [36] and Wistar rats [37,38] are established models of sucrose-induced insulin resistance and hypertriglyceridemia. There are also high-fat rat models. The Goto-Gakizaki (GK) rat was established by genetic segregation based on the glucose level as early as eight days after birth. In GK rats, blood glucose level after oral glucose administration is high, with a severe glucose tolerance abnormality. GK rats also have metabolic-endocrine abnormalities, such as hyperglycemia and insulin resistance. In the pancreas, islet deformation, secretion abnormality, and gradual b-cell loss were observed [39]. The diabetic Zuker fatty rat was established as a fatty model rat [40,41]. An abnormal gain of weight is observed three weeks after birth and the body weight increases with age. Obese Zucker rats have high levels of lipids in their blood and are resistant to insulin. The Otsuka Long Evans Tokushima fatty (OLETF) rat is a useful animal model of type II diabetes with obesity [42,43]. The OLETF rat presents clinically relevant phenotypes of diabetes such as hyperinsulinemia, hyperglycemia, insulin resistance, hypertriglycemia, and mild obesity. In vivo insulin-stimulated glucose uptake as measured with a euglycemic clamp was reduced 37% compared with that in non-diabetic control rats. Morphological studies on the OLETF rat pancreas showed enlarged multi-lobulated fibrotic islets, and hyper states of insulin production and secretion. The Wister fatty rat (fa/fa) was developed by taking the fatty gene from the Zucker rat and transferring it to the Wistar-Kyoto rat, which is less sensitive to insulin, and less tolerant to glucose, than lean Zucker rats. Wistar fatty rats also exhibit obesity, hyperphagia, hyperlipidemia, hyperinsulinemia, and peripheral insulin resistance, similar to Zucker rats. The tolerance and insulin response to oral glucose are decreased with advancing age in males. Hypertrophy of pancreatic islets and degranulation of beta cells have also been observed.

Mouse models

In contrast to rats, mice are used less frequently as a model for sucrose/fructose-induced insulin resistance and hypertriglyceridemia. The response to high fructose/sucrose diets is very strain-dependent in mice [44], and commonly used strains like C57Bl/6 either do not develop insulin resistance or only develop it slowly [45]. When C57Bl/6 mice were fed a highfructose diet for eight weeks, they developed increased mean arterial pressure, reduced glucose tolerance, and increased plasma cholesterol that was attributed to the activation of the sympathetic and angiotensin systems [46]. Like the high dietary fructose used for the induction of diabetes, fat-based diets can induce diabetes and obesity [47]. The effects of fat and sucrose were compared separately and in combination in diabetes- and obesity-prone C57Bl/6J and diabetes- and obesityresistant A/J mice. After four months, the feed efficiency (weight gained divided by calories consumed), did not differ across diets in A/J mice. However, C57Bl/6J mice had significantly increased feed efficiency for the fat-based diet. The KK mouse was established in 1962 [48]. It shows a striking intolerance to glucose, hyperinsulinism, and obesity. Of particular interest is the development of mild to moderate glomerulosclerosis in the prediabetic stage. Later in life, progression to severe glomerulosclerosis and attendant proteinuria are observed [49]. The ob/ob mouse is a mutant strain that eats excessively and becomes obese. It is one of the typical animal models of type 2 diabetes. In adipose tissue, enlargement of cells and an increase of cell number were observed. Idenfication of a mutation in the ob gene led to the discovery of leptin, which is important for the control of food consumption. Since the ob/ob mouse cannot produce leptin, its food intake is uncontrolled [50]. A mouse similar to the *ob/ob* mouse, the *db/* db mouse, was eatablished [51]. This mouse exhibits severe diabetes with longstanding hyperglycemia. Moreover, within six weeks of age, *db/db* mice develop significant obesity, fasting hyperglycemia, and hyperinsulinemia. The *db/db* mice have a prominent peak in the low density lipoprotein (LDL) range.

http://e-dmj.org Diabetes Metab J 2011;35:309-316

dmj

Upon consumption of a 0.15% (wt/wt) cholesterol and 21% (wt/wt) fat "western" diet, db/db mice show elevated plasma cholesterol [52]. The non-obese diabetic (NOD) mouse was established in 1984. Diabetes develops in NOD mice as a result of insulitis, leukocytic infiltration of the pancreatic islets [53,54]. The incidences of spontaneous diabetes in the NOD mouse are 60% to 80% in females and 20% to 30% in males. The onset of diabetes also varies between males and females. The time difference of the onset may be attributed to the breeding environment. For chemical induction of diabetes, the N-nitro derivative of glucosamin (STZ) is used frequently. STZ is a glucosamine-nitrosourea compound that is particularly toxic to the insulin-producing β -cells in the pancreas in mammals. Multiple small doses of STZ produce a delayed, progressive increase in plasma glucose in mice within 5 to 6 days after the injections. High dose injection of STZ results in acute hyperglycemia. Alloxan, a crystalline substance produced by oxidation of uric acid and a toxic glucose analogue, selectively destroys β-cells in the pancreas when administrated to laboratory animals, including mice and rats [55]. This causes insulindependent diabetes mellitus similar to type 1 diabetes in human patients. This most likely occurs because of selective uptake of the compound due to its structual similarity to glucose, as well as the β -cell's highly efficient uptake mechanism [56].

Gene-manipulated mouse models

Knockout mouse technology is a valuable research tool and contributes to the clarification of biological questions. Via disruption of a specific gene in the mouse and observation of differences from normal behaviour or responses, researchers can infer its probable biological function. However, some limitations exist. With knockout mice, it is often difficult to determine a gene's function in relation to human diseases as the genes in mice may have different functions from the human genes. Investigators have established knockout mouse models for diabetes research. However mice carrying a targeted disruption of the diabetes-related gene might develop early onset diabetes and die perinatally. Detailed examination of the pancreas revealed proliferation arrest and cell death in pancreatic β -cells. As a consequence, it is likely that these cumulative effects cause diabetes.

Considering carbohydrate chains including GSLs, diabetic mice with P-selectin glycoprotein ligand-1 (PSGL-1) deficiency show a protective phenotype against obesity-related insulin resistance [57]. PSGL-1 knockout mice fed a high-fat diet (HFD)

show a remarkable reduction of macrophage accumulation and expression of pro-inflammatory genes. Moreover, adipocyte hypertrophy, insulin resistance, lipid metabolism, and hepatic fatty change were improved in PSGL-1 knockout mice compared with WT mice fed HFD. PSGL-1 is a crucial adhesion molecule for the recruitment of monocytes into adipose tissues in obese mice. Several studies have reported that blockade of PSGL-1 reduces inflammatory reactions [58,59]. Mice with gene disruption of GM3 are viable without major abnormalities, but have heightened sensitivity to insulin [60]. The basis for the increased insulin sensitivity in mutant mice was found to be enhanced IR phosphorylation in skeletal muscle. Importantly, the absence of the ganglioside GM3 confers protection from HDF-induced insulin resistance to the knockout mice. Gganglioside GM3 is a negative regulator of insulin signaling.

CONCLUSION

In this review, I summarized the molecular-based diversity of GSLs. Microdomains, termed lipid rafts, which are special structures of cell membranes are made of GSLs and protein receptors. These specialized membrane microdomains may compartmentalize cellular processes and function for the assembly of signaling molecules. Since many molecules, such as receptors and channels penetrate cell membranes, the positive or negative influence (e.g., regulation of signaling, internalization, and trafficking) of the cell membrane, including GSLs and other components, such as cholesterol, and sphingomyelin, is not small. On the other hand, model animals including mice with gene disruption are used for diabetes research. They provide an essential research tool for academic institutions and commercial companies by enabling the rapid screening of novel compounds, and testing of new gene-based therapies for the treatment of diabetes and diabetes-related symptoms.

CONFLICTS OF INTEREST

No potential conflict of interest relevant to this article was reported.

ACKNOWLEDGMENTS

This work was supported by Grant-in-Aid for Scientific Research KAKENHI (20570122) and the World Class University (WCU) program through the National Research Foundation of Korea funded by the Ministry of Education, Science and Technology of Korea (R32-10064).

REFERENCES

- 1. Kolter T, Sandhoff K. Recent advances in the biochemistry of sphingolipidoses. Brain Pathol 1998;8:79-100.
- 2. Hakomori S. Bifunctional role of glycosphingolipids. Modulators for transmembrane signaling and mediators for cellular interactions. J Biol Chem 1990;265:18713-6.
- 3. Gault CR, Obeid LM, Hannun YA. An overview of sphingolipid metabolism: from synthesis to breakdown. Adv Exp Med Biol 2010;688:1-23.
- 4. Kasahara K, Sanai Y. Functional roles of glycosphingolipids in signal transduction via lipid rafts. Glycoconj J 2000;17:153-62.
- 5. Korade Z, Kenworthy AK. Lipid rafts, cholesterol, and the brain. Neuropharmacology 2008;55:1265-73.
- 6. Pike LJ. The challenge of lipid rafts. J Lipid Res 2009;50 Suppl: S323-8.
- 7. Rietveld A, Simons K. The differential miscibility of lipids as the basis for the formation of functional membrane rafts. Biochim Biophys Acta 1998;1376:467-79.
- Saeki K, Fukuyama S, Ayada T, Nakaya M, Aki D, Takaesu G, Hanada T, Matsumura Y, Kobayashi T, Nakagawa R, Yoshimura A. A major lipid raft protein raftlin modulates T cell receptor signaling and enhances th17-mediated autoimmune responses. J Immunol 2009;182:5929-37.
- 9. Sohn HW, Pierce SK, Tzeng SJ. Live cell imaging reveals that the inhibitory FcgammaRIIB destabilizes B cell receptor membrane-lipid interactions and blocks immune synapse formation. J Immunol 2008;180:793-9.
- Kinoshita MO, Furuya S, Ito S, Shinoda Y, Yamazaki Y, Greimel P, Ito Y, Hashikawa T, Machida T, Nagatsuka Y, Hirabayashi Y. Lipid rafts enriched in phosphatidylglucoside direct astroglial differentiation by regulating tyrosine kinase activity of epidermal growth factor receptors. Biochem J 2009;419:565-75.
- Sanchez-Wandelmer J, Davalos A, de la Pena G, Cano S, Giera M, Canfran-Duque A, Bracher F, Martin-Hidalgo A, Fernandez-Hernando C, Lasuncion MA, Busto R. Haloperidol disrupts lipid rafts and impairs insulin signaling in SH-SY5Y cells. Neuroscience 2010;167:143-53.
- 12. Kim KB, Kim BW, Choo HJ, Kwon YC, Ahn BY, Choi JS, Lee JS, Ko YG. Proteome analysis of adipocyte lipid rafts reveals

that gC1qR plays essential roles in adipogenesis and insulin signal transduction. Proteomics 2009;9:2373-82.

- Yamashita T, Wada R, Sasaki T, Deng C, Bierfreund U, Sandhoff K, Proia RL. A vital role for glycosphingolipid synthesis during development and differentiation. Proc Natl Acad Sci U S A 1999;96:9142-7.
- 14. Godi A, Di Campli A, Konstantakopoulos A, Di Tullio G, Alessi DR, Kular GS, Daniele T, Marra P, Lucocq JM, De Matteis MA. FAPPs control Golgi-to-cell-surface membrane traffic by binding to ARF and PtdIns(4)P. Nat Cell Biol 2004;6:393-404.
- 15. Hakomori S, Igarashi Y. Functional role of glycosphingolipids in cell recognition and signaling. J Biochem 1995;118:1091-103.
- Nagai Y, Tsuji S. Significance of ganglioside-mediated glycosignal transduction in neuronal differentiation and development. Prog Brain Res 1994;101:119-26.
- 17. Ichikawa S, Hirabayashi Y. Glucosylceramide synthase and glycosphingolipid synthesis. Trends Cell Biol 1998;8:198-202.
- Schnaar RL. Glycosphingolipids in cell surface recognition. Glycobiology 1991;1:477-85.
- 19. Tettamanti G, Riboni L. Gangliosides and modulation of the function of neural cells. Adv Lipid Res 1993;25:235-67.
- 20. Simons K, Ikonen E. Functional rafts in cell membranes. Nature 1997;387:569-72.
- 21. Harder T, Simons K. Caveolae, DIGs, and the dynamics of sphingolipid-cholesterol microdomains. Curr Opin Cell Biol 1997;9:534-42.
- 22. Anderson RG. The caveolae membrane system. Annu Rev Biochem 1998;67:199-225.
- 23. Whitmore CD, Hindsgaul O, Palcic MM, Schnaar RL, Dovichi NJ. Metabolic cytometry. Glycosphingolipid metabolism in single cells. Anal Chem 2007;79:5139-42.
- Memon RA, Holleran WM, Uchida Y, Moser AH, Ichikawa S, Hirabayashi Y, Grunfeld C, Feingold KR. Regulation of glycosphingolipid metabolism in liver during the acute phase response. J Biol Chem 1999;274:19707-13.
- 25. Tettamanti G. Ganglioside/glycosphingolipid turnover: new concepts. Glycoconj J 2004;20:301-17.
- 26. White MF, Kahn CR. The insulin signaling system. J Biol Chem 1994;269:1-4.
- 27. Hubbard SR, Wei L, Ellis L, Hendrickson WA. Crystal structure of the tyrosine kinase domain of the human insulin receptor. Nature 1994;372:746-54.
- 28. Schinner S, Scherbaum WA, Bornstein SR, Barthel A. Molecular mechanisms of insulin resistance. Diabet Med 2005;22:674-

82.

- 29. Vander Haar E, Lee SI, Bandhakavi S, Griffin TJ, Kim DH. Insulin signalling to mTOR mediated by the Akt/PKB substrate PRAS40. Nat Cell Biol 2007;9:316-23.
- 30. Simons K, Toomre D. Lipid rafts and signal transduction. Nat Rev Mol Cell Biol 2000;1:31-9.
- Hannun YA, Obeid LM. Principles of bioactive lipid signalling: lessons from sphingolipids. Nat Rev Mol Cell Biol 2008;9:139-50.
- 32. Vainio S, Heino S, Mansson JE, Fredman P, Kuismanen E, Vaarala O, Ikonen E. Dynamic association of human insulin receptor with lipid rafts in cells lacking caveolae. EMBO Rep 2002;3:95-100.
- 33. Ottenhoff R, Powlson AS, Grefhorst A, van Eijk M, Dubbelhuis PF, Aten J, Kuipers F, Serlie MJ, Wennekes T, Sethi JK, O'Rahilly S, Overkleeft HS. Pharmacological inhibition of glucosylceramide synthase enhances insulin sensitivity. Diabetes 2007;56:1341-9.
- 34. Radin NS, Shayman JA, Inokuchi J. Metabolic effects of inhibiting glucosylceramide synthesis with PDMP and other substances. Adv Lipid Res 1993;26:183-213.
- 35. Kobayashi T, Takahashi M, Nagatsuka Y, Hirabayashi Y. Lipid rafts: new tools and a new component. Biol Pharm Bull 2006; 29:1526-31.
- Pagliassotti MJ, Prach PA, Koppenhafer TA, Pan DA. Changes in insulin action, triglycerides, and lipid composition during sucrose feeding in rats. Am J Physiol 1996;271(5 Pt 2):R1319-26.
- 37. Chicco A, D'Alessandro ME, Karabatas L, Pastorale C, Basabe JC, Lombardo YB. Muscle lipid metabolism and insulin secretion are altered in insulin-resistant rats fed a high sucrose diet. J Nutr 2003;133:127-33.
- 38. Robbez Masson V, Lucas A, Gueugneau AM, Macaire JP, Paul JL, Grynberg A, Rousseau D. Long-chain (n-3) polyunsaturated fatty acids prevent metabolic and vascular disorders in fructose-fed rats. J Nutr 2008;138:1915-22.
- 39. Ihara Y, Toyokuni S, Uchida K, Odaka H, Tanaka T, Ikeda H, Hiai H, Seino Y, Yamada Y. Hyperglycemia causes oxidative stress in pancreatic beta-cells of GK rats, a model of type 2 diabetes. Diabetes 1999;48:927-32.
- 40. Johnson PR, Zucker LM, Cruce JA, Hirsch J. Cellularity of adipose depots in the genetically obese Zucker rat. J Lipid Res 1971;12:706-14.
- 41. Schonfeld G, Felski C, Howald MA. Characterization of the plasma lipoproteins of the genetically obese hyperlipoprotein-

emic Zucker fatty rat. J Lipid Res 1974;15:457-64.

- Kawano K, Hirashima T, Mori S, Saitoh Y, Kurosumi M, Natori T. Spontaneous long-term hyperglycemic rat with diabetic complications. Otsuka Long-Evans Tokushima Fatty (OLETF) strain. Diabetes 1992;41:1422-8.
- 43. Shima K, Shi K, Sano T, Iwami T, Mizuno A, Noma Y. Is exercise training effective in preventing diabetes mellitus in the Otsuka-Long-Evans-Tokushima fatty rat, a model of spontaneous non-insulin-dependent diabetes mellitus? Metabolism 1993;42:971-7.
- 44. Nagata R, Nishio Y, Sekine O, Nagai Y, Maeno Y, Ugi S, Maegawa H, Kashiwagi A. Single nucleotide polymorphism (-468 Gly to A) at the promoter region of SREBP-1c associates with genetic defect of fructose-induced hepatic lipogenesis [corrected]. J Biol Chem 2004;279:29031-42.
- 45. Sumiyoshi M, Sakanaka M, Kimura Y. Chronic intake of highfat and high-sucrose diets differentially affects glucose intolerance in mice. J Nutr 2006;136:582-7.
- 46. Farah V, Elased KM, Chen Y, Key MP, Cunha TS, Irigoyen MC, Morris M. Nocturnal hypertension in mice consuming a high fructose diet. Auton Neurosci 2006;130:41-50.
- 47. Surwit RS, Feinglos MN, Rodin J, Sutherland A, Petro AE, Opara EC, Kuhn CM, Rebuffe-Scrive M. Differential effects of fat and sucrose on the development of obesity and diabetes in C57BL/6J and A/J mice. Metabolism 1995;44:645-51.
- 48. Nakamura M. A diabetic strain of the mouse. Proc Jpn Acad Ser B Phys Biol Sci 1962;38:348-52.
- 49. Reddi AS, Camerini-Davalos RA. Hereditary diabetes in the KK mouse: an overview. Adv Exp Med Biol 1988;246:7-15.
- 50. Friedman JM, Halaas JL. Leptin and the regulation of body weight in mammals. Nature 1998;395:763-70.
- Sima AA, Robertson DM. Peripheral neuropathy in mutant diabetic mouse [C57BL/Ks (db/db)]. Acta Neuropathol 1978; 41:85-9.
- 52. Kobayashi K, Forte TM, Taniguchi S, Ishida BY, Oka K, Chan

L. The db/db mouse, a model for diabetic dyslipidemia: molecular characterization and effects of Western diet feeding. Metabolism 2000;49:22-31.

- Kikutani H, Makino S. The murine autoimmune diabetes model: NOD and related strains. Adv Immunol 1992;51:285-322.
- 54. Wen L, Ley RE, Volchkov PY, Stranges PB, Avanesyan L, Stonebraker AC, Hu C, Wong FS, Szot GL, Bluestone JA, Gordon JI, Chervonsky AV. Innate immunity and intestinal microbiota in the development of type 1 diabetes. Nature 2008;455: 1109-13.
- 55. Chatzigeorgiou A, Halapas A, Kalafatakis K, Kamper E. The use of animal models in the study of diabetes mellitus. In Vivo 2009;23:245-58.
- 56. Lenzen S. The mechanisms of alloxan- and streptozotocin-induced diabetes. Diabetologia 2008;51:216-26.
- 57. Sato C, Shikata K, Hirota D, Sasaki M, Nishishita S, Miyamoto S, Kodera R, Ogawa D, Tone A, Kataoka HU, Wada J, Kajitani N, Makino H. P-selectin glycoprotein ligand-1 deficiency is protective against obesity-related insulin resistance. Diabetes 2011;60:189-99.
- 58. Inoue T, Tsuzuki Y, Matsuzaki K, Matsunaga H, Miyazaki J, Hokari R, Okada Y, Kawaguchi A, Nagao S, Itoh K, Matsumoto S, Miura S. Blockade of PSGL-1 attenuates CD14+ monocytic cell recruitment in intestinal mucosa and ameliorates ileitis in SAMP1/Yit mice. J Leukoc Biol 2005;77:287-95.
- Rivera-Nieves J, Burcin TL, Olson TS, Morris MA, McDuffie M, Cominelli F, Ley K. Critical role of endothelial P-selectin glycoprotein ligand 1 in chronic murine ileitis. J Exp Med 2006;203:907-17.
- 60. Yamashita T, Hashiramoto A, Haluzik M, Mizukami H, Beck S, Norton A, Kono M, Tsuji S, Daniotti JL, Werth N, Sandhoff R, Sandhoff K, Proia RL. Enhanced insulin sensitivity in mice lacking ganglioside GM3. Proc Natl Acad Sci U S A 2003;100: 3445-9.