

CASE REPORT

Open Access

# Lethal infection caused by *Tetratrichomonas gallinarum* in black swans (*Cygnus atratus*)



Shengyong Feng<sup>1,2</sup>, Han Chang<sup>1,2</sup>, Yutian Wang<sup>3</sup>, Fubing Luo<sup>4</sup>, Qiaoxing Wu<sup>5</sup>, Shuyi Han<sup>1</sup> and Hongxuan He<sup>1\*</sup>

## Abstract

**Background:** *Tetratrichomonas gallinarum* is parasitic protozoa with a wide host range. However, its lethal infection is rare reported.

**Case presentation:** Here, we described the first lethal cases of *T. gallinarum* infection in black swans in China. Five black swans died within a week in succession without obvious symptoms except mild diarrhea. At necropsy, severe lesions were observed in caeca with thickened caecal walls and hemorrhages in the mucosa. A large number of moving trophozoites were found in the contents of the cecum by microscopic examination. The livers were enlarged with multiple bleeding spots on the surface. Histopathology of the livers showed mononuclear cell infiltration and moderate hyperplasia of fibrous tissue. The histopathology of the cecum showed that the villi of the cecum were edematous. Finally, the presence of *T. gallinarum* was determined by specific PCR and in-situ hybridization assay. Additionally, common pathogens that can cause similar symptoms were excluded.

**Conclusions:** The death of the black swan was caused by *T. gallinarum*, suggesting that the parasite might be a new threat to the *Cygnus* birds.

**Keywords:** Black swan, Cecum, China, Liver, *T. gallinarum*

## Background

*Tetratrichomonas gallinarum* is parasitic protozoa with a wide host range [1]. Owing to sick birds are usually co-infected with other pathogens and artificially infected animals rarely develop symptoms, the pathogenicity of *T. gallinarum* is controversial [2–4]. Moreover, lesions caused by *T. gallinarum* in birds were sporadically reported in some countries, such as in chukar partridges, mockingbird, Waldrapp ibis and white pelican from America [5–8], in duck from Germany [9], in red-legged partridges from Great Britain [10], and in Layer chickens from the Netherlands [11]. Here, we described the first fatal case of black swans (*Cygnus atratus*) associated

with *T. gallinarum* infection in China, and the threat of the protozoa to *Cygnus* birds must be considered.

## Case presentation

In August 2019, five adult black swans from a wetland park of Beijing died within a week in succession. Before they died, no obvious symptoms were observed except mild diarrhea.

The fresh carcasses were sent to the National Research Center for Wildlife Borne Diseases for post-mortem and histopathological examination. At routinely pathological investigation, the ceca were swollen and the mucosa were hemorrhages and

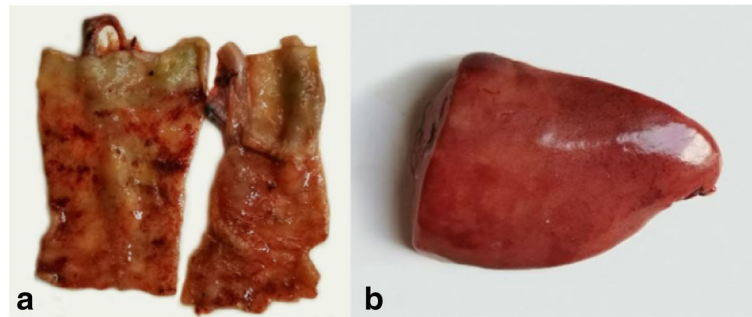
\* Correspondence: [hehx@ioz.ac.cn](mailto:hehx@ioz.ac.cn)

<sup>1</sup>National Research Center for Wildlife Borne Diseases, Institute of Zoology, Chinese Academy of Sciences, 1-5 Beichenxilu, Chaoyang District, Beijing 100101, PR China

Full list of author information is available at the end of the article



© The Author(s). 2021 **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

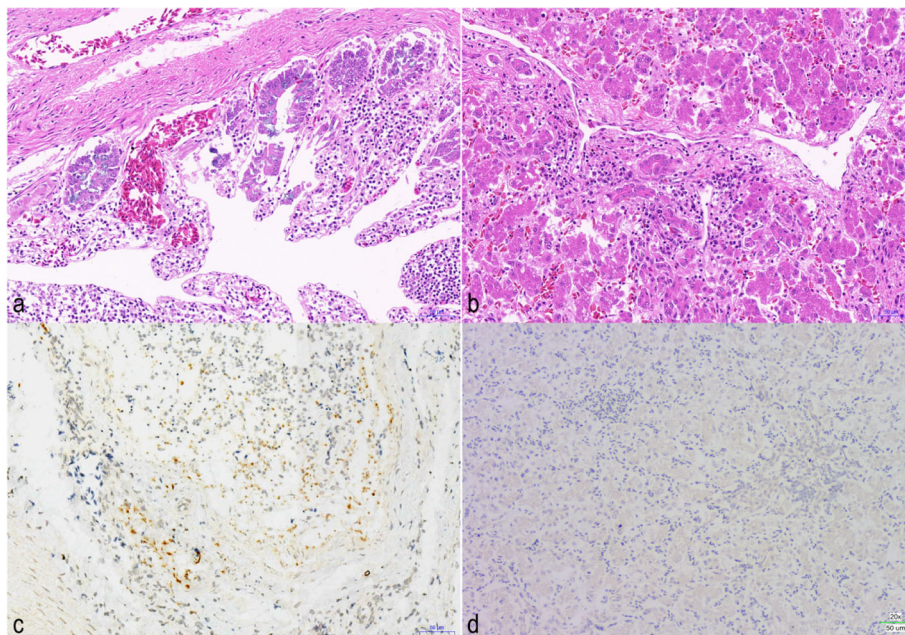


**Fig. 1** Pathological changes of cecum (a) and liver (b)

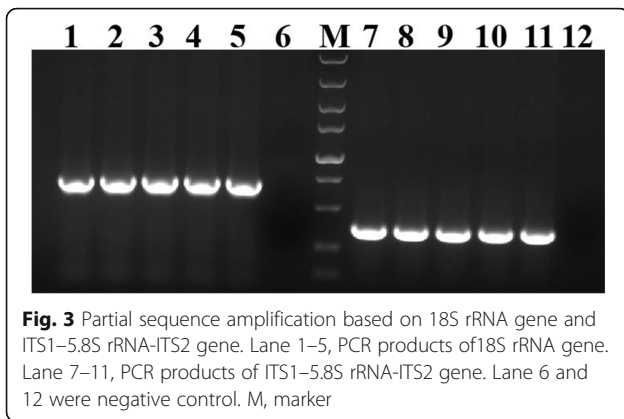
anabrosis (Fig. 1a). A large number of moving trophozoites were observed by microscopic examination. The livers were enlarged and accompanied by the color turned dark red and the edge was blunt (Fig. 1b). No visible lesions were found in other organs. Histopathological examination showed that cecal hemorrhage, intestinal villi edema, disordered arrangement, epithelial cells exfoliated, and many parasites were found in lamina propria (Fig. 2a). Vacuolar degeneration of hepatocytes and interlobular bile duct hyperplasia were observed in the liver tissues. A large number of mononuclear inflammatory cells infiltrated between the liver lobules, and the fibrous tissue proliferated moderately (Fig. 2b).

Histological sections from the livers and caeca of the birds were further processed for in situ hybridization (ISH) using the described probe specific for *T. gallinarum* and *H. meleagridis* [12, 13]. The positive signals with the *T. gallinarum* probe were found in the caeca (Fig. 2c) but not in the livers (Fig. 2d). The result of ISH in the caeca and livers showed no signal with the *H. meleagridis* probe.

Using two trichomonad primer sets, TFR1/R2 and 18S-F/R, the ITS and 18S rRNA region of the isolates were successfully amplified with specific single band size of approximately 350 bp and 600 bp in the gel [14, 15] (Fig. 3), respectively. Notably, the PCR products were subcloned into T-vectors before sequencing to ensure



**Fig. 2** Haematoxylin and eosin staining of the caecum (a) and the liver (b) of a dead black swans. ISH revealed the presence of *T. gallinarum* in the caecum (c) within the localizations as brown-stained cells. The signals of *T. gallinarum* probe in the liver was negative (d)



that the specific sequences be successfully sequenced. Both sequences were clustered with the reference sequences of *T. gallinarum* download from GenBank database under phylogenetic analyses (Fig. 4a, b).

Other potential pathogens, such as *Coccidia* spp., *Blastocystis* spp. and hepatitis E virus were negative using the method previously reported [16–18].

Taken together, after eliminating potential pathogens, such as *H. meleagridis*, *Coccidia*, *Blastocystis* spp., hepatitis E virus as well as pathogenic bacteria, the presence of *T. gallinarum* was eventually confirmed by microscopic examination, histopathology, specific PCR amplification and ISH. Therefore, the

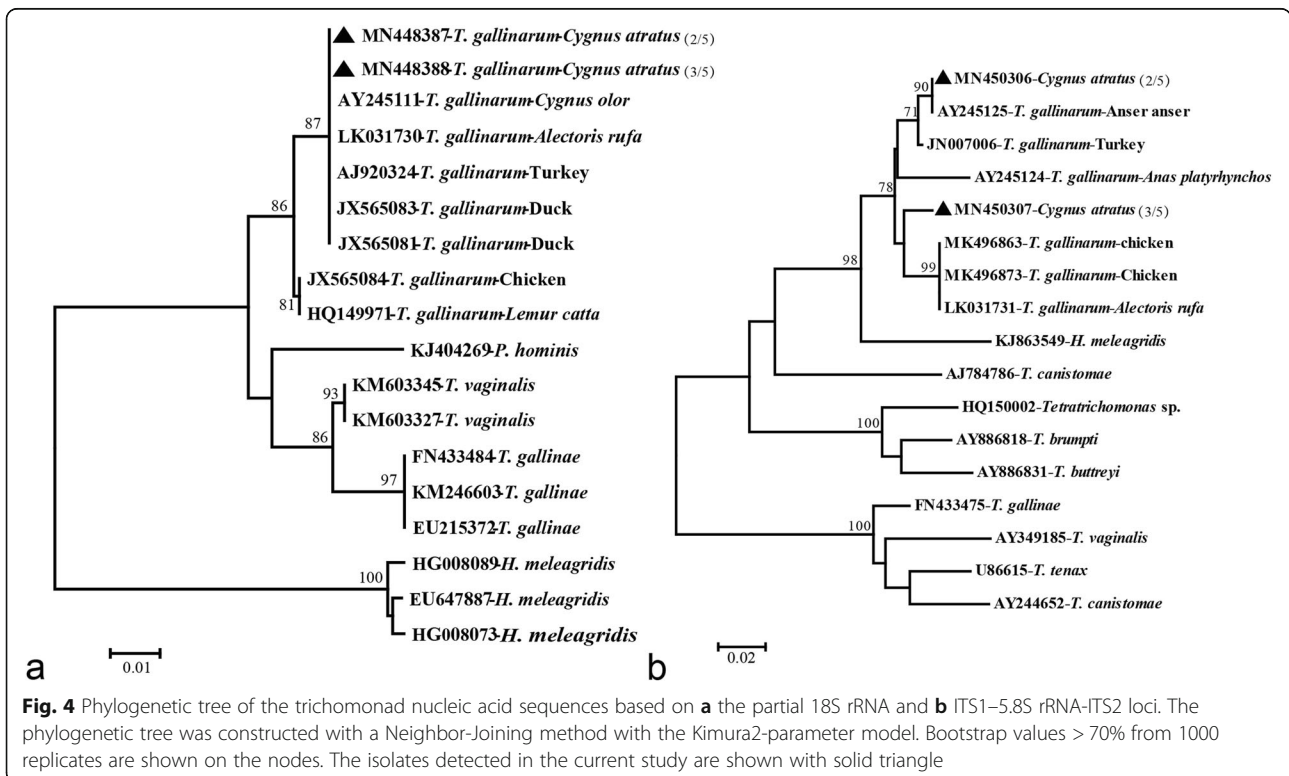
death of the black swan was likely to be caused by *T. gallinarum*.

### Discussion and conclusions

Though *T. gallinarum* is commonly found gallinaceous and anseriform birds, it seldom causes diseases [19]. The maturity of the immune system may be an important reason for the host to suffer from this parasite, as previous studies have found that most of the dead birds were juveniles or subadults [8, 9]. However, all the dead black swans in the present study were adult, thus the heterogeneity between *T. gallinarum* isolates might also be an important factor result in the differences in pathogenicity among hosts.

Studies conducted by Dimasuy and Rivera shown that *T. gallinarum* can be detected from healthy ducks (*Anas platyrhynchos*) [20], which suggested that the parasite might be commensal in some duck species. In the present study, some healthy ducks shared activity area with the black swans. Thus the *T. gallinarum* recovered from the black swans may be spillover from the ducks.

In conclusion, we described the first fatal case of black swans associated with *T. gallinarum* infection in China, suggesting that the protozoan might be a new threat to the *Cygnus* birds. A comprehensive epidemiological investigation of *T. gallinarum* in *Cygnus* birds is urgently needed in the future.





## Abbreviations

ISH: In situ hybridization; ITS: Internal transcribed spacer

## Acknowledgments

We acknowledge Dr. Ping Wang specifically for his excellent PS technical assistance.

## Authors' contributions

Experimental design was done by HH and SF. Collection of samples was done by HC and YW. The experiments were done by HC, SH, FL and QW. The manuscript was written by SF and revised by HH. All authors read and approved the final manuscript.

## Funding

This work was supported by the Strategic Priority Research Program of the Chinese Academy of Sciences (XDA19050204), Beijing Innovation Consortium of Agriculture Research System (BAIC04–2019); State Administration of Forestry and Grassland, China and Chinese Academy of Sciences (CZBZX-1).

## Availability of data and materials

The ITS and 18S nucleotide sequences of *T. gallinarum* generated in the present study have been deposited in GenBank database under the accession numbers MN448387 and MN448388 as well as MN450306 and MN450307, respectively.

## Declarations

### Ethics approval and consent to participate

This study was approved by the Animal Ethics Committee of the Institute of Zoology, Chinese Academy of Sciences. All samples were handled in accordance with good animal practices required by the Animal Ethics Procedures and Guidelines of the People's Republic of China.

### Consent for publication

Not applicable.

### Competing interests

The authors declare that they have no competing interests.

### Author details

<sup>1</sup>National Research Center for Wildlife Borne Diseases, Institute of Zoology, Chinese Academy of Sciences, 1-5 Beichenxilu, Chaoyang District, Beijing 100101, PR China. <sup>2</sup>College of Life Sciences, University of Chinese Academy of Sciences, Chaoyang District, Beijing 100101, China. <sup>3</sup>Beijing General Station of Animal Husbandry, Chaoyang District, Beijing 100107, China. <sup>4</sup>Beijing Center for Animal Disease Control, Beijing, China. <sup>5</sup>Shaanxi Institute of zoology, Xi'an 710032, Shaanxi, China.

Received: 14 December 2020 Accepted: 29 April 2021

Published online: 13 May 2021

## References

- Cepicka I, Hampf V, Kulda J, Flegr J. New evolutionary lineages, unexpected diversity, and host specificity in the parabasalid genus *Tetratrichomonas*. *Mol Phylogenet Evol*. 2006;39(2):542–51. <https://doi.org/10.1016/j.ympev.2006.01.005>.
- Norton RA. Pathogenicity of a strain of *Trichomonas gallinarum* in turkeys and its possible interaction with cecal coccidia. *Avian Dis*. 1997;41(3):670–5. <https://doi.org/10.2307/1592159>.
- Kemp RL, Reid WM. Pathogenicity studies on *Trichomonas gallinarum* in domestic poultry. *Poult Sci*. 1965;44(1):215–21. <https://doi.org/10.3382/ps.0440215>.
- Amin A, Liebhart D, Weissenböck H, Hess M. Experimental infection of turkeys and chickens with a clonal strain of *Tetratrichomonas gallinarum* induces a latent infection in the absence of clinical signs and lesions. *J Comp Pathol*. 2011;144(1):55–62. <https://doi.org/10.1016/j.jcpa.2010.06.002>.
- Wichmann RW, Bankowski RA. A report of *Trichomonas gallinarum* infection in chukar partridges (*Alectoris graeca*). *Cornell Vet*. 1956;46(3):367–9.
- Patton CS, Patton S. *Tetratrichomonas gallinarum* encephalitis in a mockingbird (*Mimus polyglottos*). *J Vet Diagn Invest*. 1996;8(1):133–7. <https://doi.org/10.1177/104063879600800126>.

- Laing ST, Weber ES, Yabsley MJ, et al. Fatal hepatic tetratrichomoniasis in a juvenile Waldraup ibis (*Geronticus eremita*). *J Vet Diagn Invest*. 2013;25(2):277–81. <https://doi.org/10.1177/1040638713476711>.
- Burns RE, Braun J, Armien AG, Rideout BA. Hepatitis and splenitis due to systemic tetratrichomoniasis in an American white pelican (*Pelecanus erythrorhynchos*). *J Vet Diagn Invest*. 2013;25(4):511–4. <https://doi.org/10.1177/1040638713488368>.
- Richter B, Schulze C, Kammerling J, Mostegl M, Weissenböck H. First report of typhlitis/typhlohepatitis caused by *Tetratrichomonas gallinarum* in three duck species. *Avian Pathol*. 2010;39(6):499–503. <https://doi.org/10.1080/03079457.2010.518137>.
- Liebhart D, Neale S, Garcia-Rueda C, Wood AM, Bilic I, Wernsdorf P, et al. A single strain of *Tetratrichomonas gallinarum* causes fatal typhlohepatitis in red-legged partridges (*Alectoris rufa*) to be distinguished from histomonosis. *Avian Pathol*. 2014;43(5):473–80. <https://doi.org/10.1080/03079457.2014.959435>.
- Landman WJ, Molenaar RJ, Cian A, van der Heijden HM, Viscogliosi E. Granuloma disease in flocks of productive layers caused by *Tetratrichomonas gallinarum*. *Avian Pathol*. 2016;45(4):465–77. <https://doi.org/10.1080/03079457.2016.1163325>.
- Richter B, Fragner K, Weissenböck H. Simultaneous detection of protozoa in the tissues of snakes by double in situ hybridization. *Microsc Res Tech*. 2008;71(4):257–9. <https://doi.org/10.1002/jemt.20546>.
- Liebhart D, Weissenböck H, Hess M. In-situ hybridization for the detection and identification of *Histomonas meleagridis* in tissues. *J Comp Pathol*. 2006;135(4):237–42. <https://doi.org/10.1016/j.jcpa.2006.08.002>.
- Felleisen RS. Comparative sequence analysis of 5.8S rRNA genes and internal transcribed spacer (ITS) regions of trichomonadid protozoa. *Parasitology*. 1997;115(Pt 2):111–9. <https://doi.org/10.1017/S0031182097001212>.
- Bilic I, Jaskulska B, Souillard R, Liebhart D, Hess M. Multi-locus typing of *Histomonas meleagridis* isolates demonstrates the existence of two different genotypes. *PLoS One*. 2014;9(3):e92438. <https://doi.org/10.1371/journal.pone.0092438>.
- Jarquín-Díaz VH, Balard A, Jost J, et al. Detection and quantification of house mouse *Eimeria* at the species level - challenges and solutions for the assessment of coccidia in wildlife. *Int J Parasitol Parasites Wildl*. 2019;10:29–40. <https://doi.org/10.1016/j.ijppaw.2019.07.004>.
- Scicluna SM, Tawari B, Clark CG. DNA barcoding of blastocystis. *Protist*. 2006;157(1):77–85. <https://doi.org/10.1016/j.protis.2005.12.001>.
- Sun ZF, Larsen CT, Dunlop A, Huang FF, Pierson FW, Toth TE, et al. Genetic identification of avian hepatitis E virus (HEV) from healthy chicken flocks and characterization of the capsid gene of 14 avian HEV isolates from chickens with hepatitis-splenomegaly syndrome in different geographical regions of the United States. *J Gen Virol*. 2004;85(Pt 3):693–700. <https://doi.org/10.1099/vir.0.19582-0>.
- Amin A, Bilic I, Liebhart D, Hess M. Trichomonads in birds—a review. *Parasitology*. 2014;141(6):733–47. <https://doi.org/10.1017/S0031182013002096>.
- Dimasuy KG, Rivera WL. Molecular characterization of trichomonads isolated from animal hosts in the Philippines. *Vet Parasitol*. 2013;196(3–4):289–95. <https://doi.org/10.1016/j.vetpar.2013.03.019>.

## Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more [biomedcentral.com/submissions](https://biomedcentral.com/submissions)

