

Emerging molecular biomarkers in osteoarthritis pathology

Amit Sandhu*, Jason S. Rocket*, Starlee Lively and Mohit Kapoor 

Ther Adv Musculoskelet Dis

2023, Vol. 15: 1–12

DOI: 10.1177/
1759720X231177116

© The Author(s), 2023.
Article reuse guidelines:
sagepub.com/journals-
permissions

Abstract: Osteoarthritis (OA) is the most common form of arthritis resulting in joint discomfort and disability, culminating in decline in life quality. Attention has been drawn in recent years to disease-associated molecular biomarkers found in readily accessible biofluids due to low invasiveness of acquisition and their potential to detect early pathological molecular changes not observed with traditional imaging methodology. These biochemical markers of OA have been found in synovial fluid, blood, and urine. They include emerging molecular classes, such as metabolites and noncoding RNAs, as well as classical biomarkers, like inflammatory mediators and by-products of degradative processes involving articular cartilage. Although blood-based biomarkers tend to be most studied, the use of synovial fluid, a more isolated biofluid in the synovial joint, and urine as an excreted fluid containing OA biomarkers can offer valuable information on local and overall disease activity, respectively. Furthermore, larger clinical studies are required to determine relationships between biomarkers in different biofluids, and their impacts on patient measures of OA. This narrative review provides a concise overview of recent studies of OA using these four classes of biomarkers as potential biomarker for measuring disease incidence, staging, prognosis, and therapeutic intervention efficacy.

Keywords: biomarkers, cartilage degradation, inflammatory biomarkers, metabolites, noncoding RNA, osteoarthritis

Received: 24 March 2022; revised manuscript accepted: 24 April 2023.

Introduction

Osteoarthritis (OA), the most prevalent form of arthritis, results in joint pain and disability. OA can affect any articulating joint in the body, often multiple joints at a time. The most common clinical symptoms of OA present as joint pain and stiffness, restricting mobility and function.¹ While the underlying causes of OA remain unknown, it is thought to begin with molecular, anatomical, and/or physiological anomalies in joint tissues. These perturbances initiate, usually over many years, a cascade of progressive, dynamic, whole-joint changes involving local inflammation, articular cartilage degradation, and bone remodeling.

Pathological tissue changes can be detected by different imaging modalities.² Primary assessment methods to diagnose OA are based on

clinical evaluations and patient-reported outcome measures^{3,4} combined with radiographic analysis of osteophytes, bone sclerosis, and joint space narrowing, using the Kellgren and Lawrence (KL) grading scale.⁵ Even though X-ray and magnetic resonance imaging (MRI) are employed to diagnose joint disorders, these have a limited ability to monitor very early molecular changes occurring in joint tissues that precede the morphological changes they detect.^{2,6,7} Moreover, findings from imaging modalities do not always correlate with patient symptoms.⁸ Joint alterations detected by radiographs generally identify structural modifications detected relatively late in disease pathogenesis. However, these late-stage alterations are the result of ongoing molecular changes that are only detectable using assays capable of detecting these specific molecular entities.⁹ With populations rapidly aging¹⁰ and the

Correspondence to:
Mohit Kapoor
Division of Orthopaedics,
Osteoarthritis Research
Program, Schroeder
Arthritis Institute,
University Health Network,
60 Leonard Avenue, 5th
Floor Krembil Discovery
Tower, Toronto, ON M5G
2C4, Canada

Krembil Research
Institute, University Health
Network, Toronto, ON,
Canada

Department of Surgery
and Department of
Laboratory Medicine and
Pathobiology, University
of Toronto, Toronto, ON,
Canada
mk Kapoor@uhnresearch.ca

Amit Sandhu
Jason S. Rocket
Starlee Lively
Division of Orthopaedics,
Osteoarthritis Research
Program, Schroeder
Arthritis Institute,
University Health Network,
Toronto, ON, Canada

Krembil Research
Institute, University Health
Network, Toronto, ON,
Canada

*Equal First Author
Contribution.

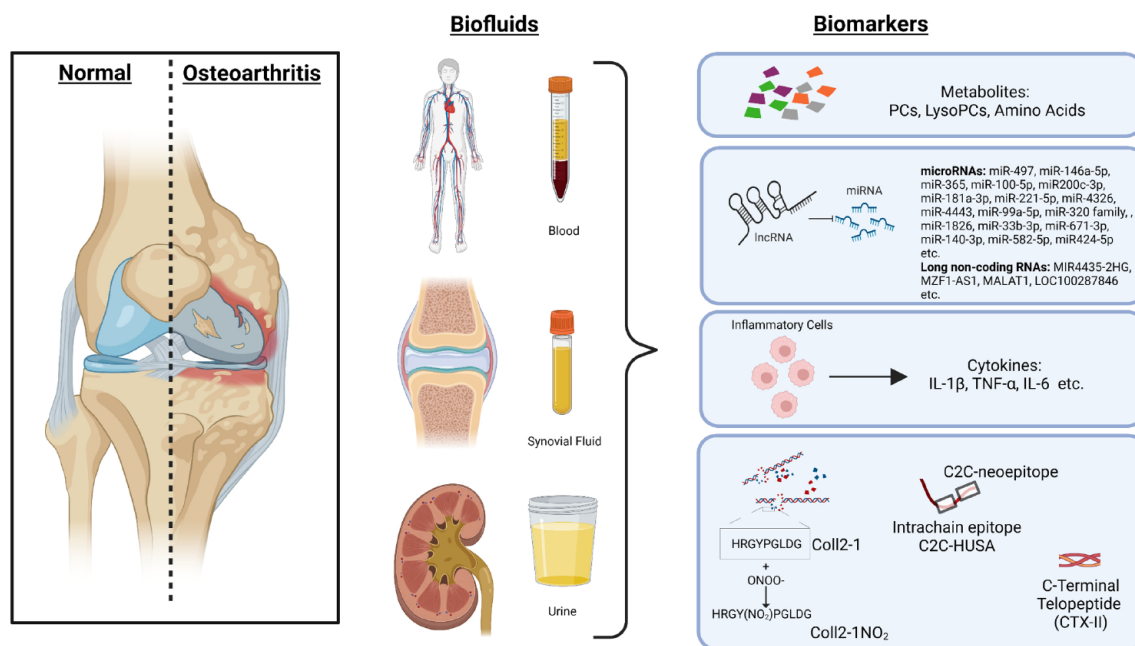


Figure 1. Diagrammatic representation of some key biomarkers in biofluids from patients with osteoarthritis. Metabolites, noncoding RNAs, inflammatory markers and type II collagen cleavage products have been investigated as potential biomarkers of OA diagnosis, progression and prognosis in multiple biofluids including blood isolates (plasma and serum), synovial fluid and urine. *Source:* Created by Biorender.

close association of OA prevalence with increasing age,^{1,11} there is an urgent need for cost-efficient and sensitive methods to identify molecular changes occurring in joints that precede imaging-detected pathologies. Use of these molecules as biomarkers would enable earlier detection and more effective treatment of OA. As OA appears to diagnose and prognose joint disease progression over weeks or months, as opposed to years. In this regard, recent studies suggest the association of biomarkers, such as noncoding RNAs,¹² metabolites,¹³ inflammatory markers,^{14–16} and cartilage degradation products^{17,18} with OA pathology (Figure 1), as well as other biomarker entities (e.g. epigenetic and microbial) reviewed in detail elsewhere.^{19–22} This narrative review includes discussion of some key emerging and classical molecular indicators found in human fluids that associate with OA diagnosis, progression, and prognosis, published in the past 5 years.

Emerging molecular biomarkers

Metabolites

Metabolites are small molecules produced as by-products of metabolic processes and include lipids,

amino acids, sugars, and their derivatives. A metabolome includes all metabolites found in a biological unit (i.e. any given cell, tissue, or biological fluid of interest). Methods to detect metabolite levels can be targeted or untargeted. Targeted approaches have the advantage of measuring concentrations of specific metabolites. Conversely, untargeted approaches more comprehensively measure metabolites within a metabolome and have the potential to uncover novel entities, but with limitations, as metabolite identification is generally based on comparison of measurement spectra to database records. In the past 5 years, multiple studies have investigated metabolomes of urine, synovial fluid, and blood (serum and plasma), as potential sources of biomarkers for OA.

Blood is a whole-body metabolome to which several tissues, including those in the joints, contribute. Blood-derived metabolites have been investigated as a source for diagnostic and prognostic biomarkers of OA. Untargeted metabolomics using gas chromatography, coupled with time-of-flight mass spectrometry (GC/TOF-MS), found 58 significantly changed metabolites in the plasma of OA patients compared to healthy volunteers, of which succinic acid, xanthurenic acid,

and L-tryptophan could potentially assist in OA diagnosis.²³ Gender-specific effects have also been observed. Using the Multicenter Osteoarthritis study, a comparison of the knee (K)OA progressors ($n=234$) versus nonprogressors ($n=322$) found that baseline plasma phenylalanine was associated with increased risk of KOA progression (unilateral or bilateral), particularly in women.²⁴ Furthermore, baseline serum ratio of lysophosphatidylcholine (lysoPC)18:2 to phosphatidylcholine (PC)44:3 was associated with lateral KOA cartilage loss examined over a 24-month period ($n=139$).²⁵ Finally, targeted serum metabolomics of end-stage knee or hip OA patients ($n=70$) compared to controls ($n=82$) showed decreased levels of C10:1, C10:2, C12, C12:1, C14, C14:2, C14:1-OH, carnitine palmitoyltransferase 1-ratio and total acylcarnitine/carnitine, with acylcarnitines also showing association to OA radiographic severity,²⁶ suggesting acylcarnitine levels could aid in grading of OA through nonimaging modalities. Thus blood is a rich source of OA-relevant metabolites that could potentially be used as diagnostic and prognostic measures of OA.

Up to 34% of patients with end-stage KOA do not show reduced pain or improved function after total knee arthroplasty (TKA).²⁷ Circulating biomarkers could contribute to the decision-making process involved in determining the likelihood of beneficial surgical outcomes. For instance, differential correlation network analysis of plasma metabolites from TKA patients ($n=461$) at baseline identified 12 and 23 metabolites that were positively associated with pain and function nonresponders, respectively.²⁸ In the same cohort, pain nonresponders were positively associated with ratios of acetyl-carnitine (C2) to PC acylalkyl (ae) C40:1, and negatively associated with PC diacyl (aa) C36:4 to isoleucine, while function nonresponders were negatively associated with the ratio of glutamine to isoleucine.²⁹ Thus, in addition to individual metabolites, their ratios might also be good candidates as biomarkers of OA prognosis.

Specific patient characteristics may also contribute to differences in the circulating metabolome. For instance, network analysis of plasma and synovial fluid metabolites from advanced KOA patients (males and females, both $n=50$) found sex-specific differences in synovial fluid versus plasma, with individual metabolites serving as key 'nodes' for the differential networks.³⁰

Furthermore, plasma cystine, uric acid, and tyrosine were associated with OA severity and effusion synovitis in female volunteers ($n=596$).³¹ However, a comparison to males is needed to see if these metabolite-OA disease associations are female-selective. Weight is another potential contributing factor to the circulating metabolome. LC/Q-TOF/MS/MS analyses of metabolites in the serum of control, normal weight KOA, and obese KOA subjects found differences in levels of 21 metabolites in KOA patients versus controls and 15 differences in metabolite levels between healthy weight and obese KOA patients.³² However, targeted nuclear magnetic resonance (NMR)-spectroscopy of serum from 31 KOA patients showed no detectable differences in metabolites between underweight and obese individuals.³³ This suggests that the contribution of weight to OA is complex. Age is another likely contributor to the metabolome of OA patients, as analysis of plasma from a cohort of 346 subjects (152 KOA, 194 healthy volunteers) by targeted metabolomics found that individual levels of select lysoPC and PC analogs were associated with individuals with KOA in older populations, particularly males.³⁴ Thus, multiple patient characteristics should be considered when investigating the role of metabolites in OA prognosis. The challenge lies in their identification, which is essential as their inclusion in computational modeling would further help improve metabolite discovery as well as biomarker accuracy for diagnosis and prognosis, optimizing precision medicine approaches.

While research has mainly focused on metabolite levels in the blood, examining more local fluids, like synovial fluid, may be a more direct, and potentially earlier, indicator of OA and its progression. OA-affected tissues (cartilage, synovium, osteophytes, fat pad, subchondral bone, etc.) secrete their metabolic by-products, directly contributing to the fluid's composition. Untargeted metabolite profiling identified 10 metabolites that were found at different levels in synovial fluid from primary KOA patients ($n=10$) versus control individuals ($n=5$).³⁵ Untargeted metabolomics of synovial fluid from OA and healthy individuals ($n=5$ each) identified 35 metabolites as potential biomarkers, including phosphatidylcholines, lysophosphatidylcholines, ceramides, myristate derivatives, and carnitine derivatives.³⁶ Synovial fluid metabolite levels of glutamine, 1,5-anhydroglucitol, gluconic lactone, tyramine, threonine, and 8-aminocaprylic acid,

detected by untargeted GC/TOF-MS, were significantly different in a sample of KOA patients ($n=25$) compared to controls ($n=10$). A validation set of 11 controls and 24 OA subjects with targeted metabolomics of the synovial fluid found glutamine, 1,5-anhydroglucitol, gluconic lactone and 8-aminocaprylic acid, or 1,5-anhydroglucitol and gluconic lactone in combination, could differentiate control and OA patients.³⁷ Metabolites might also be helpful in identifying the stage of OA, as levels of 28 metabolites were differentially detected by GC/TOF-MS in synovial fluid from KOA patients divided into KL1/2 ($n=8$) and KL3/4 ($n=7$), which related to fatty acid metabolism, glycerophospholipid metabolism, and glycerolipid metabolism.³⁸ Metabolites in synovial fluid offer the potential to expand our understanding of the local changes in joint metabolic processes during OA. However, this approach is limited by the increased chance of joint insult during acquisition, which could increase the risk of OA or exacerbate its progression. Thus, the need remains to identify OA-specific metabolites in more readily accessible biofluids, like blood.

Urine is an even easier and less invasive biofluid to acquire than blood, but metabolites in urine have been less frequently studied over the past 5 years as compared to plasma and synovial fluid. To our knowledge, only a single study was identified that used untargeted liquid chromatography-high resolution mass spectrometry of urine from KOA patients classified into inflammatory OA ($n=22$) and noninflammatory ($n=52$), compared to non-OA controls ($n=68$) to identify 26 metabolites that could distinguish inflammatory OA *versus* non-OA controls, but no metabolites that could distinguish noninflammatory OA *versus* non-OA controls.³⁹

Multiple studies indicate that metabolite levels in biofluids of OA patients may be helpful in providing diagnostic and prognostic information. However, future studies should focus on establishing congruency between joint metabolite changes associated with disease progression and their detection in systemic fluids while controlling for changes resulting from additional clinical, anthropometric, and demographic factors. This would help to better define the links between local pathogenesis and systemic biomarkers. Additional investigations probing putative urinary metabolite biomarkers and studying their possible interrelationships with other body fluids

may be helpful in understanding the relationships of metabolite biomarkers to OA.

Noncoding RNAs

In the past 5 years, several studies have provided intriguing data to support noncoding RNAs as biomarkers for OA, particularly microRNAs (miRNAs). These noncoding RNAs are encoded in DNA, transcribed into RNA but not translated into protein. MicroRNAs bind mRNA and either promote mRNA degradation or suppress protein translation.⁴⁰ Some studies have identified miRNAs as diagnostic of OA. For instance, higher levels of miR-497 were found in serum of women with hip OA (KL IV; $n=23$) compared to controls ($n=52$) and its levels were also an effective discriminator of individuals with OA (AUC = 0.89).⁴¹ Plasma miR-146a and miR-365 were also increased in KOA patients (KL I-III; $n=42$) compared to healthy controls ($n=28$) and could distinguish individuals with OA (AUC=0.8843 and 0.8182, respectively).⁴² However, not all miRNAs are increased with OA. Compared to trauma patients without KOA ($n=10$), serum levels of let-7e were lower in KOA patient patients ($n=10$).⁴³ Similarly, serum samples from primary KOA patients (KL III/IV) and healthy individuals (both groups, $n=12$) were screened by microarray and three miRNAs (miR-33b-3p, miR-671-3p, miR-140-3p) were found to be downregulated in KOA patients, with subsequent validation by qRT-PCR.⁴⁴ In a study using whole blood, miR-582-5p, and miR-424-5p were reduced in KOA patients compared to control individuals and were found to be reduced in expression in KOA patients with sclerotic ($n=6$) compared nonsclerotic ($n=4$) subchondral bone.⁴⁵ In a larger cohort study using synovial fluid, levels of miR-100-5p, miR-200c-3p, and miR-1826 were lower in KOA patients (KL II-IV, $n=150$) compared to controls ($n=150$), with each found to be fair to good at distinguishing individuals with KOA based on AUC analysis.⁴⁶ Overall, these studies suggest that miRNAs in different biofluids from OA patients may help in diagnosing OA, potentially even individual pathologies associated with the disease. However, a better understanding of the differences between healthy control subjects and OA patients at different stages of disease progression is needed.

Identification of biomarkers capable of distinguishing OA at early stages would help provide

interventions to slow disease progression in a timelier manner. Some studies have investigated miRNA levels in biofluids from early compared to late-stage OA patients. For instance, miR-210 in synovial fluid was lower in healthy volunteers ($n = 10$) compared to both KL I/II and KL III/IV KOA patients (both KOA groups, $n = 20$), suggesting that miR-210 may be a putative early diagnostic marker of OA.⁴⁷ In a separate study of early (KL 0/I; $n = 41$) and late (KL III/IV; $n = 50$) KOA subjects, a distinct panel of 11 plasma miRNAs was able to distinguish the 2 groups.⁴⁸ Finally, in women with KOA, serum miR-186-5p was found to be positively associated with incident KOA, while miR-146a-5p was also found to be positively associated with prevalent KOA in the same sample.⁴⁹ Further studies involving longitudinal and early OA patients would help identify novel miRNA biomarkers or validate already identified miRNA biomarkers of early disease, enabling earlier intervention to slow disease progression and prolong the time to potential surgical intervention.

Additional recent studies have focused on biofluid miRNAs as prognostic biomarkers of OA. Using hip and KOA subjects from Research Arthritis and Articular Cartilage (RAAK) ($n = 22$) and Genetics osteoArthritis and Progression (GARP) ($n = 71$) study cohorts, a panel of 7 miRNAs in plasma (miR140-3p, miR-1307-5p, miR-181a-3p, miR-221-5p, miR-4326, miR-443, and miR-99a-5p) was capable of predicting OA progression over 2 and 5 years.⁵⁰ Furthermore, in a KOA patient sample from the Osteoarthritis Initiative (OAI) cohort ($n = 106$), an association was found between plasma levels of the miR-320 family (miR-320b/c/d/e) and fast-progressing radiographic KOA subjects over a 4-year period.⁵¹ Finally, in a sample of KOA patients (KL II-IV, $n = 136$), baseline levels of 3 miRNAs (miR-146a-5p, miR-145-5p, and miR-130b-3p) were higher in individuals with lower pain relief, with miR-146a-5p trending as an independent predictor of postoperative pain relief.⁵² Taken together, these findings suggest that miRNAs may act as important predictors of OA progression.

In addition to predicting OA progression, biomarker tracking of therapeutic intervention efficacy could also be useful, alerting clinicians to when therapy is no longer effective and to try alternative interventions. For instance, in a study of OA patients (KL II/III, $n = 28$) who underwent

high tibial osteotomy for medial compartmental KOA, miR-30c-5p was reduced in 6-month postoperative synovial fluid compared to preoperative, which also associated with postoperative pain relief.⁵³ Additionally, miR-98 levels were higher in the serum of KOA patients meeting the American College of Rheumatology Classification Criteria for OA compared to controls (both groups, $n = 20$), which were reduced with treatment with glucosamine sulfate.⁵⁴ Future studies are required to determine if the decrease in miR-98 levels with treatment corresponded with improvements in patient outcome measures.

In addition to miRNAs, some of their regulators have also begun to be investigated as potential biomarkers of OA. Long noncoding (lnc) RNAs act as miRNA “sponges,” effectively inhibiting miRNA-RNA interactions, reducing the overall epigenetic contributions of miRNAs.⁴⁰ The lncRNA MIR4435-2HG was downregulated in both plasma and synovial fluid of OA patients (stage 3/4, $n = 78$) compared to healthy volunteers ($n = 58$), with synovial fluid levels being an excellent OA diagnostic indicator (AUC = 0.96). In this study, OA treatments, including exercise, reduction of joint burden, and nonsteroidal anti-inflammatory drugs, increased MIR4435-2HG plasma levels up to 3 months after treatment initiation.⁵⁵ The sustained elevation of this lncRNA suggests it might be a good biomarker of therapeutic efficacy; however, further study is needed to correlate its plasma levels to pain and function outcome measures. Additionally, a study of 20 preoperative KOA patients revealed that serum levels of MZF1-AS1, MALAT1, and LOC100287846 were lower preoperatively in the chronic postoperative pain group ($n = 10$) compared to the normal postoperative pain recovery group ($n = 10$).⁵⁶ Thus, in addition to miRNAs, biofluid lncRNAs may also be good candidate biomarkers for OA diagnosis, prognosis, and therapeutic efficacy tracking. More extensive studies investigating lncRNAs, their links to individual miRNAs, and changes with OA stage, symptoms and outcomes are necessary.

Overall, additional longitudinal validation studies with larger sample numbers are needed to confirm the applicability of these noncoding RNAs as biomarkers of OA. In addition to blood isolates, studies should also focus on other biofluids, like synovial fluid and urine, which may reveal connections between joint-specific production, presence in circulation, and eventual clearance,

helping to understand molecular dynamics of noncoding RNAs for biomarker purposes.

Classical molecular biomarkers: Recent progress

Inflammatory markers. Compared to rheumatoid arthritis (RA), inflammation in OA is considered a low-grade process. Despite this, inflammation is still an important contributor to the development and progression of OA, even in the early stages.¹⁸ Structural alterations in the OA-inflicted joint are hypothesized to be linked, at least in part, to multiple proinflammatory cytokines.⁵⁷ The role of inflammatory mediators in human OA has been extensively reviewed.^{15,58} Many of the identified cytokines are abundantly expressed by various cell types involved in OA, including immune cells, chondrocytes, osteoblasts, and synoviocytes, and have become attractive biomarkers for OA diagnostic and therapeutic intervention efficacy tracking purposes.⁵⁹ Of all the inflammatory mediators, interleukin (IL)-1 β , tumor necrosis factor (TNF)- α , and IL-6 have received notable attention due to their modulation of a myriad of OA-relevant signaling pathways.⁶⁰ Some recent studies provide support for further investigations into the potential of these inflammatory factors as biomarkers of OA.

A study of synovial fluid from 83 patients with primary KOA found negative correlations between IL-1 β , TNF- α , or IL-6 with patient-reported visual analog scale (VAS) pain scores or KL grade; however, a positive relationship between KL grade and VAS pain scores was identified.⁶¹ These findings suggest that these inflammatory mediators are larger drivers of pain in KOA patients with less severe radiographic damage compared to those at later stages, despite their chronic presence. In a study of 46 subjects, proinflammatory cytokines (IL-1 β , TNF- α , or IL-6) were detected at higher serum levels of both American College of Rheumatology Stage 3 primary and secondary KOA subtypes compared to healthy controls.⁶² Furthermore, exploratory analysis of patients with early KOA ($n=139$; $>80\%$ KL \leq II) showed that serum IL-6 was associated with increases in Western Ontario and McMaster Universities Arthritis Index (WOMAC) function and MRI features (osteophytes, synovitis, and meniscus extrusion), synovial fluid IL-6 was solely associated with increases in MRI effusion, and serum TNF- α was associated with increases

in osteophytes, cartilage loss, synovitis, and effusion MRI features.⁶³ In contrast, in later stage KOA subjects ($n=20$; $>70\%$ KL \geq II), only serum IL-6 was associated with increased WOMAC pain and decreased function.⁶³ Thus, proinflammatory mediators, like IL-1 β , TNF- α , and IL-6, have the potential to be used to help subclassify OA patients (early *versus* late or primary *versus* secondary), facilitating the administration of precision medicine procedures. Of note, in the later stages of KOA, the grading of the severity of OA has a considerable ceiling effect, reducing the capacity to uncover associations with increasing levels of proinflammatory biomarkers. Therefore, future studies should strive to evaluate KOA at earlier disease stages.

Evidence suggests that IL-6 may have interesting roles in regulating OA symptoms. IL-6 levels were reportedly higher in the serum of KOA patients undergoing TKA ($n=127$) compared to healthy participants ($n=39$).⁶⁴ Linear regression modeling of this patient sample demonstrated that circulating IL-6 levels were negatively correlated with pain intensity. However, in synovial fluid of KOA patients undergoing TKA ($n=104$), which is more closely situated to OA-affected tissues, IL-6 levels were positively associated with synovitis of the parapatellar subregion and pain (WOMAC or neuropathic).⁶⁵ Similarly, a study of KOA subjects ($n=70$; KL \geq II) also found IL-6 synovial fluid levels were positively associated with pain measured on the Likert scale during movement. Synovial TNF- α was also positively associated with WOMAC total pain and WOMAC or Likert scale scores for pain during movement or at rest, while IL-1 β was negatively associated with Likert scale scores for pain during movement.⁶⁶ Another study ($n=200$) reported a signature of 11 serum cytokines, which included IL-1 β and IL-6, were associated with increased WOMAC dysfunction scores and MRI-detected bone marrow lesions over 2 years.⁶⁷ Undoubtedly, IL-6 is likely a key mediator in OA; however, its detection in specific biofluids, association with distinct stages of OA pathology, and links to pain at different stages of disease require further investigation. Similarly, measures of other fluid-based cytokines, such as TNF- α and IL-1 β , and their relationships with one another must be better understood to uncover mechanisms and relationships with OA pain and pathology.

Another inflammatory mediator that has gained considerable attention is C-reactive protein

(CRP), an acute phase protein primarily expressed downstream in response to injury and the release of inflammatory cytokines. TNF- α and IL1- β trigger the synthesis of IL-6, which subsequently induces the production of CRP.⁶⁸ A comprehensive systematic review of 32 studies recently concluded that, compared to healthy controls, OA patient levels of serum high-sensitivity (hs-)CRP corresponded with increased pain and decreased physical function.⁶⁹ Similarly, CRP metabolite (CRPM), but not CRP itself, was predictive of KOA radiographic progression as KOA patients ($n=152$) with ≥ 9 ng/mL CRPM serum levels were more likely to develop contralateral KOA after 2 years (KL change ≥ 2).⁷⁰ A positive association between the levels of hs-CRP and KOA patients with MRI-defined synovitis ($n=58$), compared to healthy controls ($n=33$), has been reported, with no association to pain intensity as measured by VAS.⁷¹ In a separate study of KOA subjects with varying degrees of pain intensity measured by VAS ($n=281$), no relationship between either hs-CRP or CRPM, and clinical pain intensity was found, although hs-CRP did associate with KL grade.⁷² Furthermore, CRP was positively associated with VAS-measured pain severity in an independent cross-sectional study of a larger cohort of advanced OA subjects (knee or hip; $n=770$), but not with any items of the WOMAC questionnaire.⁷³ Finally, CRPM serum levels correlated with knee joint inflammation, particularly synovitis, as measured by etarfolatide radionucleotide imaging in KOA patients ($n=25$).⁶⁸ CRP and its metabolites appear to be involved in OA-related pain, but because measuring pain is typically a subjective measure, its analysis and outcome interpretations are complicated.⁶¹ Additional investigations are needed to clarify the associations between hs-CRP, CRPM, pain severity, and OA pathologic severity, to further support its use as a diagnostic or prognostic biomarker for OA.

Cartilage degradation markers. Collagen type II, the principle component of the extracellular matrix (ECM) of healthy hyaline cartilage, forms an extensive collagen fibrillar network.⁷⁴ Over time, it is susceptible to degradation, notably by collagenases, resulting in peptide fragments that can be readily assayed in biofluids. If the balance is shifted such that degradation occurs faster than anabolic processes, loss of cartilage ensues, exacerbating OA pathology. Various degradation products of type II collagen have been identified,

including the C-telopeptide of cross-linked type II collagen (CTX-II), 45-mer peptide (C2C), a specific 45-mer peptide containing the downstream C2C neoepitope and intrachain epitope (IB-C2C-HUSA), and a 9-mer peptide (¹⁰⁸HRGYPGLDG¹¹⁶) released during degradation from the triple helical region of type II collagen (Coll2-1). Immunoassays detecting these degradative type II collagen fragments have emerged as promising tools in the search for molecular biomarkers of OA.^{17,75,76}

In 2016, a longitudinal nested case-control study (194 cases and 406 OA comparators) of the OAI cohort was published that evaluated 18 biochemical molecules for their ability to predict OA progression between 24 and 48 months, defined as both progression of radiographic joint space loss and persistent progressive pain worsening.⁷⁵ This study identified that, baseline, 12-, and 24-month time-integrated levels of urinary CTX-II, and 24-month time-integrated level of urinary IB-C2C-HUSATM, were promising predictors of OA progression status. Furthermore, both time-integrated urinary CTX-II and urinary IB-C2C-HUSATM levels at baseline, 12 months, or 24 months were able to predict OA progression, as defined by radiographic joint space loss, persistent progressive pain worsening, or both.

CTX-II levels in urine, serum, or synovial fluid are increased in KOA patients with higher radiographic KL grades.^{66,76-82} Of note, no significant differences in urine CTX-II levels were detected between patients with early radiographic stages of OA (KL1) and healthy individuals,⁸⁰ suggesting that the rise of CTX-II levels might result from processes occurring at later stages of OA. Other patient characteristics, like age, sex, and body mass index, can also influence urine CTX-II levels. For instance, premenopausal women with early KOA ($n=146$; KL ≤ 1) had significantly higher urine CTX-II levels compared to controls ($n=108$), an effect not observed in men.⁸³ Urine CTX-II has also been found to be associated with pain duration and body weight of Mexican females with KOA ($n=155$).⁸² Moreover, a longitudinal study showed baseline urine CTX-II levels as a risk factor for the incidence of total joint replacement in postmenopausal women ($n=478$).⁸⁴ Thus, patient characteristics likely influence urine CTX-II levels, which may need to be considered when examining the role of this cartilage degradation product in OA studies.

Elevated IB-C2C-HUSA™ in urine was first reported in detail to discriminate subjects with early from advanced OA⁸⁵ and has been shown to predict the risk of emerging KOA over a 12-year span.⁸⁶ Higher baseline IB-C2C-HUSA™ levels were also shown to associate with, and increase the risk of, progressive radiographic cartilage loss over 3 years in the human population-based cohort study originally using IB-C2C-HUSA™.⁸⁷ Moreover, it was noted that IB-C2C-HUSA™ may be an effective preoperative marker of symptom outcome of total knee replacement in females.⁸⁸ Similarly, an analysis of 600 OA patient samples from the Foundation for the National Institutes of Health study suggested a positive association with MRI-measured infrapatellar fat pad signal intensities with IB-C2C-HUSA™ and uCTX-II.⁸⁹

Serum levels of another collagen degradation product, Coll2-1, and its nitrated form, Coll2-1NO₂, were reported to be associated with KOA features identified by whole-organ MRI scoring ($n = 121$)⁹⁰ or radiographic analysis ($n = 598$).⁷⁶ Overall, these recent findings further indicate that CTX-II, IB-C2C-HUSA™, and Coll2-1 (and its nitrated form) may be promising cartilage degradation biomarkers found in patient biofluids, which will further enhance diagnostic and prognostic accuracy. To support their usefulness in precision medicine, it will be imperative to consider how patient demographic, anthropometric, or clinical characteristics influence levels of these putative biomarkers in large-scale future studies.

Concluding remarks

It is evident that newly emerging and classical OA-related molecules, including metabolites, miRNAs, inflammatory cytokines, and type II collagen degradation fragments, could be helpful biomarkers for assessing disease incidence, staging, prognosis, and therapeutic intervention efficacy. However, to increase the chances of their success at adding value to patient care and advance prognostic and diagnostic precision medicine approaches, additional large, longitudinal, multi-center studies are required to uncover likely interrelationships between the various identified biomarkers and their levels in different biofluids. For example, although blood-based biomarkers tend to be frequently evaluated, the use of synovial fluid, a more isolated biofluid in the synovial joint, can provide insightful information on local disease activities. Similarly, urine is another worthwhile

biofluid for the exploration of biomarkers. As a result, studies investigating the convergence of biomarkers from all three fluids might also be of interest to intersect local, systemic, and excreted OA biomarkers. Finally, investigating OA populations to generate models using multiple molecular entities that link to OA metrics, while considering the influence of various patient characteristics, would be helpful to further define and refine OA biomarkers signatures best associated with specific OA pathologies.

Declaration

Ethics approval and consent to participate
Not applicable.

Consent for publication
Not applicable.

Author contribution(s)

Amit Sandhu: Conceptualization; Data curation; Formal analysis; Investigation; Methodology; Writing – original draft; Writing – review & editing.

Jason S. Rockel: Conceptualization; Data curation; Investigation; Methodology; Writing – review & editing.

Starlee Lively: Conceptualization; Data curation; Investigation; Methodology; Writing – review & editing.

Mohit Kapoor: Funding acquisition; Methodology; Project administration; Supervision; Validation; Writing – review & editing.

Acknowledgements

None.

Funding

The authors disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This work is supported in part by the Canada Research Chairs Program and Tony and Shari Fell Platinum Chair in Arthritis Research. AS is a recipient of a Krembil Research Institute postdoctoral fellowship award.

Competing interests

The authors declare that there is no conflict of interest.

Availability of data and materials

Not applicable.

ORCID iD

Mohit Kapoor  <https://orcid.org/0000-0001-9877-0772>

References

- Martel-Pelletier J, Barr AJ, Cicuttini FM, *et al.* Osteoarthritis. *Nat Rev Dis Primers* 2016; 2: 16072.
- Kraus VB, Blanco FJ, Englund M, *et al.* Call for standardized definitions of osteoarthritis and risk stratification for clinical trials and clinical use. *Osteoarthritis Cartilage* 2015; 23: 1233–1241.
- Roos EM and Lohmander LS. The Knee injury and Osteoarthritis Outcome Score (KOOS): from joint injury to osteoarthritis. *Health Qual Life Outcomes* 2003; 1: 64.
- Thumboo J, Chew LH and Soh CH. Validation of the Western Ontario and McMaster University osteoarthritis index in Asians with osteoarthritis in Singapore. *Osteoarthritis Cartilage* 2001; 9: 440–446.
- Kellgren JH and Lawrence JS. Radiological assessment of osteo-arthrosis. *Ann Rheum Dis* 1957; 16: 494–502.
- Kijowski R, Demehri S, Roemer F, *et al.* Osteoarthritis year in review 2019: imaging. *Osteoarthritis Cartilage* 2020; 28: 285–295.
- Mobasheri A and Henrotin Y. Biomarkers of (osteo)arthritis. *Biomarkers* 2015; 20: 513–518.
- Hayashi D, Roemer FW and Guermazi A. Imaging for osteoarthritis. *Ann Physical Rehabil Med* 2016; 59: 161–169.
- Kraus VB and Karsdal MA. Osteoarthritis: Current molecular biomarkers and the way forward. *Calcif Tissue Int* 2021; 109: 329–338.
- Bloom DE, Canning D and Lubet A. Global population aging: facts, challenges, solutions & perspectives. *Daedalus* 2015; 144: 80–92.
- O'Neill TW, McCabe PS and McBeth J. Update on the epidemiology, risk factors and disease outcomes of osteoarthritis. *Best Pract Res Clin Rheumatol* 2018; 32: 312–326.
- Rocha FAC and Ali SA. Soluble biomarkers in osteoarthritis in 2022: year in review. *Osteoarthritis Cartilage*. Epub ahead of print 1 October 2022. DOI: 10.1016/j.joca.2022.09.005.
- Rockel JS and Kapoor M. The Metabolome and osteoarthritis: possible contributions to symptoms and pathology. *Metabolites* 2018; 8: 92.
- Henrotin Y. Osteoarthritis in year 2021: biochemical markers. *Osteoarthritis Cartilage* 2022; 30: 237–248.
- Daghestani HN and Kraus VB. Inflammatory biomarkers in osteoarthritis. *Osteoarthritis Cartilage* 2015; 23: 1890–1896.
- van Spil WE and Szilagyfi IA. Osteoarthritis year in review 2019: biomarkers (biochemical markers). *Osteoarthritis Cartilage* 2020; 28: 296–315.
- Bay-Jensen AC, Mobasheri A, Thudium CS, *et al.* Blood and urine biomarkers in osteoarthritis – an update on cartilage associated type II collagen and Aggrecan markers. *Curr Opin Rheumatol* 2022; 34: 54–60.
- Rousseau JC, Chapurlat R and Garnero P. Soluble biological markers in osteoarthritis. *Ther Adv Musculoskelet Dis* 2021; 13: 1759720x211040300.
- Ratneswaran A and Kapoor M. Osteoarthritis year in review: genetics, genomics, epigenetics. *Osteoarthritis Cartilage* 2021; 29: 151–160.
- Fathollahi A, Aslani S, Jamshidi A, *et al.* Epigenetics in osteoarthritis: novel spotlight. *J Cell Physiol* 2019; 234: 12309–12324.
- Huang Z and Kraus VB. Does lipopolysaccharide-mediated inflammation have a role in OA? *Nat Rev Rheumatol* 2016; 12: 123–129.
- Berthelot JM, Sellam J, Maugars Y, *et al.* Cartilage-gut-microbiome axis: a new paradigm for novel therapeutic opportunities in osteoarthritis. *RMD Open* 2019; 5: e001037.
- Huang Z, He Z, Kong Y, *et al.* Insight into osteoarthritis through integrative analysis of metabolomics and transcriptomics. *Clin Chim Acta* 2020; 510: 323–329.
- Zhai G, Sun X, Randell EW, *et al.* Phenylalanine is a novel marker for radiographic knee osteoarthritis progression: the MOST Study. *J Rheumatol* 2021; 48: 123–128.
- Zhai G, Pelletier JP, Liu M, *et al.* Activation of The Phosphatidylcholine to lysophosphatidylcholine pathway is associated with osteoarthritis knee cartilage volume Loss over time. *Sci Rep* 2019; 9: 9648.
- Tootsi K, Kals J, Zilmer M, *et al.* Medium- and long-chain acylcarnitines are associated with

- osteoarthritis severity and arterial stiffness in end-stage osteoarthritis patients: a case-control study. *Int J Rheum Dis* 2018; 21: 1211–1218.
27. Beswick AD, Wylde V, Gooberman-Hill R, *et al.* What proportion of patients report long-term pain after total hip or knee replacement for osteoarthritis? A systematic review of prospective studies in unselected patients. *BMJ Open* 2012; 2: e000435.
 28. Costello CA, Hu T, Liu M, *et al.* Differential correlation network analysis identified novel metabolomics signatures for non-responders to total joint replacement in primary osteoarthritis patients. *Metabolomics* 2020; 16: 61.
 29. Costello CA, Hu T, Liu M, *et al.* Metabolomics signature for non-responders to total joint replacement surgery in primary osteoarthritis patients: the Newfoundland osteoarthritis Study. *J Orthop Res* 2020; 38: 793–802.
 30. Xu B, Su H, Wang R, *et al.* Metabolic networks of plasma and joint fluid base on differential correlation. *PLoS One* 2021; 16: e0247191.
 31. Sasaki E, Yamamoto H, Asari T, *et al.* Metabolomics with severity of radiographic knee osteoarthritis and early phase synovitis in middle-aged women from the Iwaki Health Promotion Project: a cross-sectional study. *Arthritis Res Ther* 2022; 24: 145.
 32. Senol O, Gundogdu G, Gundogdu K, *et al.* Investigation of the relationships between knee osteoarthritis and obesity via untargeted metabolomics analysis. *Clin Rheumatol* 2019; 38: 1351–1360.
 33. Schadler P, Lohberger B, Thauerer B, *et al.* The association of blood biomarkers and Body Mass Index in knee osteoarthritis: a cross-sectional study. *Cartilage* 2022; 13: 19476035211069251.
 34. Rockel JS, Zhang W, Shestopaloff K, *et al.* A classification modeling approach for determining metabolite signatures in osteoarthritis. *PLoS One* 2018; 13: e0199618.
 35. Nieminen P, Hämäläinen W, Savinainen J, *et al.* Metabolomics of synovial fluid and infrapatellar fat pad in patients with osteoarthritis or rheumatoid arthritis. *Inflammation* 2022; 45: 1101–1117.
 36. Carlson AK, Rawle RA, Adams E, *et al.* Application of global metabolomic profiling of synovial fluid for osteoarthritis biomarkers. *Biochem Biophys Res Commun* 2018; 499: 182–188.
 37. Zheng K, Shen N, Chen H, *et al.* Global and targeted metabolomics of synovial fluid discovers special osteoarthritis metabolites. *J Orthop Res* 2017; 35: 1973–1981.
 38. Kim S, Hwang J, Kim J, *et al.* Metabolite profiles of synovial fluid change with the radiographic severity of knee osteoarthritis. *Joint Bone Spine* 2017; 84: 605–610.
 39. Abdelrazig S, Ortori CA, Doherty M, *et al.* Metabolic signatures of osteoarthritis in urine using liquid chromatography-high resolution tandem mass spectrometry. *Metabolomics* 2021; 17: 29.
 40. Ali SA, Peffers MJ, Ormseth MJ, *et al.* The non-coding RNA interactome in joint health and disease. *Nat Rev Rheumatol* 2021; 17: 692–705.
 41. Pertusa C, Tarin JJ, Cano A, *et al.* Serum microRNAs in osteoporotic fracture and osteoarthritis: a genetic and functional study. *Sci Rep* 2021; 11: 19372.
 42. Wu W, Xuan Y, Ge Y, *et al.* Plasma miR-146a and miR-365 expression and inflammatory factors in patients with osteoarthritis. *Malays J Pathol* 2021; 43: 311–317.
 43. Feng L, Feng C, Wang CX, *et al.* Circulating microRNA let-7e is decreased in knee osteoarthritis, accompanied by elevated apoptosis and reduced autophagy. *Int J Mol Med* 2020; 45: 1464–1476.
 44. Ntoumou E, Tzetis M, Braoudaki M, *et al.* Serum microRNA array analysis identifies miR-140-3p, miR-33b-3p and miR-671-3p as potential osteoarthritis biomarkers involved in metabolic processes. *Clin Epigenetics* 2017; 9: 127.
 45. Wang P, Dong R, Wang B, *et al.* Genome-wide microRNA screening reveals miR-582-5p as a mesenchymal stem cell-specific microRNA in subchondral bone of the human knee joint. *J Cell Physiol* 2019; 234: 21877–21888.
 46. Lai Z and Cao Y. Plasma miR-200c-3p, miR-100-5p, and miR-1826 serve as potential diagnostic biomarkers for knee osteoarthritis: Randomized controlled trials. *Medicine (Baltimore)* 2019; 98: e18110.
 47. Xie W, Su W, Xia H, *et al.* Synovial Fluid MicroRNA-210 as a Potential Biomarker for Early Prediction of Osteoarthritis. *Biomed Res Int* 2019; 2019: 7165406.
 48. Ali SA, Gandhi R, Potla P, *et al.* Sequencing identifies a distinct signature of circulating microRNAs in early radiographic knee osteoarthritis. *Osteoarthritis Cartilage* 2020; 28: 1471–1481.

49. Rousseau JC, Millet M, Croset M, *et al.* Association of circulating microRNAs with prevalent and incident knee osteoarthritis in women: the OFELY study. *Arthritis Res Ther* 2020; 22: 2.
50. Ramos YFM, Coutinho de Almeida R, Lakenberg N, *et al.* Circulating microRNAs highly correlate to expression of cartilage genes potentially reflecting OA susceptibility – towards identification of applicable early OA biomarkers. *Biomolecules* 2021; 11.
51. Ali SA, Espin-Garcia O, Wong AK, *et al.* Circulating microRNAs differentiate fast-progressing from slow-progressing and non-progressing knee osteoarthritis in the Osteoarthritis Initiative cohort. *Ther Adv Musculoskelet Dis* 2022; 14: 1759720x221082917.
52. Giordano R, Petersen KK, Andersen HH, *et al.* Preoperative serum circulating microRNAs as potential biomarkers for chronic postoperative pain after total knee replacement. *Mol Pain* 2020; 16: 1744806920962925.
53. Kwak YH, Kwak DK, Kim NY, *et al.* Significant changes in synovial fluid microRNAs after high tibial osteotomy in medial compartmental knee osteoarthritis: identification of potential prognostic biomarkers. *PLoS One* 2020; 15: e0227596.
54. Zheng WD, Zhou FL, Lin N, *et al.* Investigation for the role of CTX-III and microRNA-98 in diagnosis and treatment of osteoarthritis. *Eur Rev Med Pharmacol Sci* 2018; 22: 5424–5428.
55. Xiao Y, Bao Y, Tang L, *et al.* LncRNA MIR4435-2HG is downregulated in osteoarthritis and regulates chondrocyte cell proliferation and apoptosis. *J Orthop Surg Res* 2019; 14: 247.
56. Giordano R, Petersen KK, Santoro M, *et al.* Circulating long non-coding RNA signature in knee osteoarthritis patients with postoperative pain one-year after total knee replacement. *Scand J Pain* 2021; 21: 823–830.
57. Kapoor M, Martel-Pelletier J, Lajeunesse D, *et al.* Role of proinflammatory cytokines in the pathophysiology of osteoarthritis. *Nat Rev Rheumatol* 2011; 7: 33–42.
58. Bay-Jensen AC, Thudium CS and Mobasher A. Development and use of biochemical markers in osteoarthritis: current update. *Curr Opin Rheumatol* 2018; 30: 121–128.
59. Kumavat R, Kumar V, Malhotra R, *et al.* Biomarkers of Joint Damage in Osteoarthritis: Current Status and Future Directions. *Mediators Inflamm* 2021; 2021: 5574582.
60. Molnar V, Matišić V, Kodvanj I, *et al.* Cytokines and chemokines involved in osteoarthritis pathogenesis. *Int J Mol Sci* 2021; 22.
61. Li L, Li Z, Li Y, *et al.* Profiling of inflammatory mediators in the synovial fluid related to pain in knee osteoarthritis. *BMC Musculoskelet Disord* 2020; 21: 99.
62. Rankothgedera S, Atukorala I, Fernando C, *et al.* A potential diagnostic serum immunological marker panel to differentiate between primary and secondary knee osteoarthritis. *PLoS One* 2021; 16: e0257507.
63. Rajandran SN, Ma CA, Tan JR, *et al.* Exploring the association of innate immunity biomarkers With MRI features in both early and late stages osteoarthritis. *Front Med* 2020; 7: 554669.
64. Giordano R, Petersen KK, Andersen HH, *et al.* Serum inflammatory markers in patients with knee osteoarthritis: a proteomic approach. *Clin J Pain* 2020; 36: 229–237.
65. Radojčić MR, Thudium CS, Henriksen K, *et al.* Biomarker of extracellular matrix remodelling C1M and proinflammatory cytokine interleukin 6 are related to synovitis and pain in end-stage knee osteoarthritis patients. *Pain* 2017; 158: 1254–1263.
66. Leung YY, Huebner JL, Haaland B, *et al.* Synovial fluid pro-inflammatory profile differs according to the characteristics of knee pain. *Osteoarthritis Cartilage* 2017; 25: 1420–1427.
67. Zhu J, Ruan G, Cen H, *et al.* Association of serum levels of inflammatory markers and adipokines with joint symptoms and structures in participants with knee osteoarthritis. *Rheumatology (Oxford)* 2022; 61: 1044–1052.
68. Alexander LC Jr., McHorse G, Huebner JL, *et al.* A matrix metalloproteinase-generated neoepitope of CRP can identify knee and multi-joint inflammation in osteoarthritis. *Arthritis Res Ther* 2021; 23: 226.
69. Jin X, Beguerie JR, Zhang W, *et al.* Circulating C reactive protein in osteoarthritis: a systematic review and meta-analysis. *Ann Rheum Dis* 2015; 74: 703–710.
70. Bay-Jensen AC, Bihlet A, Byrjalsen I, *et al.* Serum C-reactive protein metabolite (CRPM) is associated with incidence of contralateral knee osteoarthritis. *Sci Rep* 2021; 11: 6583.
71. Petersen KK, Siebuhr AS, Graven-Nielsen T, *et al.* Sensitization and serological biomarkers in knee osteoarthritis patients with different degrees of synovitis. *Clin J Pain* 2016; 32: 841–848.

72. Arendt-Nielsen L, Eskehave TN, Egsgaard LL, *et al.* Association between experimental pain biomarkers and serologic markers in patients with different degrees of painful knee osteoarthritis. *Arthritis Rheumatol* 2014; 66: 3317–3326.
73. Stürmer T, Brenner H, Koenig W, *et al.* Severity and extent of osteoarthritis and low grade systemic inflammation as assessed by high sensitivity C reactive protein. *Ann Rheum Dis* 2004; 63: 200–205.
74. Sophia Fox AJ, Bedi A and Rodeo SA. The basic science of articular cartilage: structure, composition, and function. *Sports Health* 2009; 1: 461–468.
75. Kraus VB, Collins JE, Hargrove D, *et al.* Predictive validity of biochemical biomarkers in knee osteoarthritis: data from the FNIH OA biomarkers consortium. *Ann Rheum Dis* 2017; 76: 186–195.
76. Liem Y, Judge A, Kirwan J, *et al.* Multivariable logistic and linear regression models for identification of clinically useful biomarkers for osteoarthritis. *Sci Rep* 2020; 10: 11328.
77. Wang P, Song J and Qian D. CTX-II and YKL-40 in early diagnosis and treatment evaluation of osteoarthritis. *Exp Ther Med* 2019; 17: 423–431.
78. Bihlet AR, Byrjalsen I, Bay-Jensen AC, *et al.* Associations between biomarkers of bone and cartilage turnover, gender, pain categories and radiographic severity in knee osteoarthritis. *Arthritis Res Ther* 2019; 21: 203.
79. Liu CX, Gao G, Qin XQ, *et al.* Correlation Analysis of C-terminal telopeptide of collagen type II and interleukin-1 β for early diagnosis of knee osteoarthritis. *Orthop Surg* 2020; 12: 286–294.
80. Xin L, Wu Z, Qu Q, *et al.* Comparative study of CTX-II, Zn²⁺, and Ca²⁺ from the urine for knee osteoarthritis patients and healthy individuals. *Medicine (Baltimore)* 2017; 96: e7593.
81. Garcia-Alvarado FJ, Delgado-Aguirre HA, Rosales-Gonzalez M, *et al.* Analysis of polymorphisms in the MATN3 and DOT1L genes and CTX-II urinary levels in patients with knee osteoarthritis in a Northeast Mexican-Mestizo population. *Genet Test Mol Biomarkers* 2020; 24: 105–111.
82. García-Alvarado FJ, González-Martínez MDR, Jaramillo-Rodríguez Y, *et al.* Increased urinary concentration of C-terminal telopeptide of Type II collagen and pain by radiographic grade in women with knee osteoarthritis in Northeastern Mexico: a cross-sectional study. *Biores Open Access* 2020; 9: 7–12.
83. Hu N, Zhang J, Wang J, *et al.* Biomarkers of joint metabolism and bone mineral density are associated with early knee osteoarthritis in premenopausal females. *Clin Rheumatol* 2022; 41: 819–829.
84. Garnero P, Sornay-Rendu E and Chapurlat R. The cartilage degradation marker, urinary CTX-II, is associated with the risk of incident total joint replacement in postmenopausal women. A 18 year evaluation of the OFELY prospective cohort. *Osteoarthritis Cartilage* 2020; 28: 468–474.
85. Kuhi L, Tamm AE, Tamm AO, *et al.* Cartilage collagen neoepitope C2C in urine as an integrative diagnostic marker for early knee osteoarthritis. *Osteoarthritis Cartilage Open* 2020; 2: 100096.
86. Kuhi L, Tamm AE, Tamm AO, *et al.* Risk assessment of the progression of early knee osteoarthritis by Collagen Neoepitope C2C: a longitudinal study of an Estonian middle-aged cohort. *Diagnostics (Basel)* 2021; 11: 1236.
87. Poole AR, Ha N, Bourdon S, *et al.* Ability of a Urine Assay of Type II Collagen cleavage by collagenases to detect early onset and progression of articular cartilage degeneration: results from a population-based cohort study. *J Rheumatol* 2016; 43: 1864–1870.
88. Kuhi L, Tamm AE, Kumm J, *et al.* Associations of urinary collagen II Neoepitope C2C with total knee replacement outcomes: is OA a systemic disease in rapidly Progressive Cases? *Applied Sciences* 2022; 12: 164.
89. Cen H, Yan Q, Han W, *et al.* Longitudinal association of infrapatellar fat pad signal intensity alteration with biochemical biomarkers in knee osteoarthritis. *Rheumatology (Oxford)* 2022; 62: 439–449.
90. Hick AC, Malaise M, Loeuille D, *et al.* Cartilage Biomarkers Coll2-1 and Coll2-1NO2 Are associated with knee OA MRI features and are helpful in identifying patients at risk of disease worsening. *Cartilage* 2021; 13: 1637s–1647s.