Research Article

Jianzhong Li, Jingjing Lan, Qing Qiao, Lei Shen, Guoyuan Lu* Effluent Osteopontin levels reflect the peritoneal solute transport rate

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Abstract: Long-term peritoneal dialysis (PD) is accompanied by low-grade intraperitoneal inflammation and may eventually lead to peritoneal membrane injury with a high solute transport rate and ultrafiltration failure. Osteopontin (OPN) is highly expressed through the stimulation of proinflammatory cytokines in many cell types. This study aimed to investigate the potential of OPN as a new indicator of peritoneal deterioration. One hundred nine continuous ambulatory PD patients were analyzed. The levels of OPN and IL-6 in peritoneal effluents or serum were analyzed by ELISA kits. The mean effluent OPN concentration was $2.39 \pm$ 1.87 ng/mL. The OPN levels in drained dialysate were correlated with D/P Cr (p < 0.0001, R = 0.54) and D/D0 glucose (p < 0.0001, R = 0.39). Logistic regression analysis showed that the OPN levels in peritoneal effluents were an independent predictive factor for the increased peritoneal solute transport rate (PSTR) obtained by the peritoneal equilibration test (p < 0.001). The area under the receiver operating characteristic curve of OPN was 0.84 (95% CI: 0.75-0.92) in predicting the increased PSTR with a sensitivity of 86% and a specificity of 67%. The joint utilization of effluent OPN with age, effluent IL-6, and serum albumin further increased the specificity (81%). Thus, OPN may be a useful indicator of peritoneal deterioration in patients with PD.

Keywords: peritoneal dialysis, Osteopontin, peritoneal solute transport rate

1 Introduction

Peritoneal dialysis (PD) is a vital replacement therapy for patients with end-stage kidney disease. Approximately 11% of the population undergoing dialysis uses PD worldwide [1,2]. During PD, the peritoneal membrane (PM) naturally removes waste products and excess fluid from the blood and transports them to the dialysis solution. However, long-term exposure to hyperglycemic, hyperosmotic, and acidic dialysis solutions often causes low-grade chronic inflammation and PM injury. The PM presents as progressive fibrosis, angiogenesis, and vasculopathy, which lead to the increased solute transport and ultrafiltration failure [3–6]. Peritoneal equilibration test (PET) is widely used to assess the PM transport function and includes three parameters, namely, D/P Cr, D/D0 glucose, and 4 h ultrafiltration volume. In the PET results, the faster solute transfer rate is closely related to higher mortality and hospitalization rate [7]. However, PET requires multiple collection of blood and peritoneal effluents, and patients need to be hospitalized. In this regard, detecting relevant effluent-based biomarkers that can reflect the solute transfer rate is a more convenient detection strategy. Previous studies showed that effluent concentrations of interleukin 6 (IL-6), cancer antigen125 (CA125), vascular endothelial growth factor (VEGF), plasminogen activator inhibitor-1 (PAI-1), tenascin-C, and MMP families in dialysate obtained using PET were correlated with the D/P Cr ratio [8-12].

Osteopontin (OPN) is a highly phosphorylated glycophosphoprotein that can be secreted from many cell types, including epithelial cells, macrophages, osteoclasts, and fibroblasts in different tissues under inflammatory milieu. OPN participates in diverse important biological functions including inflammation, biomineralization, cell viability, tissue epithelial-mesenchymal transition (EMT), and fibrosis [13-18]. Moreover, OPN is localized in peritoneum tissues in patients with CAPD and is highly expressed accompanied with calcification deposits in inflammatory cells and fibroblast-like cells in PM [19].

We suggest that OPN may be highly expressed and secreted from the PM through long-term and low-grade

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chronic inflammation stimulation during PD treatment. We investigated whether OPN may be a new potential indicator of PM injury characterized by high solute transport by analyzing the association between OPN levels in drained dialysate and peritoneal solute transport rate (PSTR) obtained with PET.

2 Patients and methods

2.1 Patients

From February 2018 to December 2018, 215 PD patients with end-stage renal disease at the PD unit of the First Hospital Affiliated of Soochow University were followed up regularly. We selected patients who were undergoing a CAPD prescription (four times exchanges per day). Other exclusion criteria included acute inflammatory processes, diagnosis of peritonitis or abdominal trauma within the past 6 months before the study, active autoimmune diseases, and tumors, with incomplete clinical characteristics. Eventually, 109 patients who were undergoing CAPD were analyzed. Table 1 summarized the clinical characteristics of the selected patients. All the patients in this study were dialyzed with the glucose-based PD fluid (Dianeal[®], Baxter).

Table 1: Clinical characteristics of CAPD patients without peritonitis (N = 109). Clinical values are expressed as mean \pm SD

Variable	Value
Gender (male/female)	57/52
Age (years)	49.14 ± 13.25
PD duration (months)	37.32 ± 35.01
D/P Cr	0.69 ± 0.11
D/D0 glucose	0.36 ± 0.07
4 h ultrafiltration volume (mL)	270.6 ± 138.4
Total Kt/V urea	$\textbf{1.82} \pm \textbf{0.40}$
Peritoneal Kt/V urea	1.47 ± 0.35
Renal Kt/V urea	0.35 ± 0.44
Cholesterol (mmol/L)	$\textbf{4.31} \pm \textbf{1.08}$
Triglyceride (mmol/L)	1.78 ± 1.33
Uric acid (µmol/L)	$\textbf{393.1} \pm \textbf{89.18}$
Creatinine (µmol/L)	960.3 ± 292.8
Serum albumin (g/L)	34.39 ± 4.91
Hemoglobin (g/L)	96.61 ± 16.60
Serum calcium (mmol/L)	$\textbf{2.19} \pm \textbf{0.58}$
Serum phosphate (mmol/L)	$\textbf{1.61} \pm \textbf{0.49}$
PTH (pg/mL)	419.9 ± 27.78

Abbreviations: PD: peritoneal dialysis; D/P Cr: dialysate/plasma ratio of creatinine; D/D0 glucose: 4 to 0 h dialysate glucose; OPN: Osteopontin; PTH: parathyroid hormone.

Ethics approval and consent to participate: This study was performed with the written informed consent of all patients, and the procedure was approved by the Ethics Committee of The First Affiliated Hospital Soochow University.

2.2 PET

Semiquantitative assessment of the PM transport function was assessed with PET. Overnight (12 h) intraabdominal fluid was drained, and the PD fluid containing 2.5% dextrose solution was injected intraperitoneally. Fourhour dialysate creatinine concentration (D) was divided by plasma creatinine (P) concentration to obtain D/P Cr. Four-hour dialysate glucose concentration (D) was also divided by that obtained immediately after the injection (D0) to obtain D/D0 glucose. Ultrafiltration volume was measured. The grouping standard of the PET results was as follows: high transport (D/P Cr = 0.82–1.03), high–average transport (D/P Cr = 0.65–0.81), low–average transport (D/P Cr = 0.50–0.64), and low transport (D/P Cr = 0.34–0.49).

2.3 ELISA of OPN and IL-6

All participants in the experiment were asked to perform dialysis exchange according to the usual overnight dialysis regimen before visiting the PD center. Overnight effluent was fully drained the next morning in the PD center. We collected a 10 mL effluent sample and a 10 mL blood sample from each patient. The samples were stored at -80° C immediately. The frozen blood samples were thawed on ice and centrifuged for 10 min at $1,500\times g$. The frozen peritoneal effluent samples were thawed on ice and centrifuged for 15 min at $2,500\times g$. OPN and IL-6 levels in peritoneal effluents or serum were quantified with the ELISA kit (CUSABIO China) according to the manufacturer's instructions.

2.4 Statistical analysis

Clinical data were presented as means \pm standard deviation (SD). Baseline characteristics of the study population were compared using Student's-test or Mann–Whitney *U* test for normally or nonnormally distributed variables. Relationships between clinical variable and D/P Cr level were analyzed with Spearman's correlation coefficient

test. Independent factors affecting PSTR were analyzed by the logistic regression analysis. In addition, a logistic regression model was constructed for the probability of increased PSTR to identify the possible predictors of increased PSTR. The equation is as follows: probability = $\exp(c)/[1 + \exp(c)]$, where *c* is $0.875 \times$ effluent OPN levels + $0.06 \times \text{age} - 0.139 \times \text{blood}$ albumin + $0.037 \times$ effluent IL-6 levels – 1.059. The optimal cutoff point was identified based on the maximum Youden index (sensitivity + specificity – 1). *p* < 0.05 (two tailed) was considered statistically significant. Statistical analyses were conducted using SPSS version 25.0 software (IBM Corp., USA).

3 Results

3.1 Patient characteristics

A total of 109 CAPD patients were enrolled in this study. Among these patients, 55.3% were males with a mean age of 49.14 \pm 13.25 years and a median PD duration of 37.32 \pm 35.01 months. OPN could be detected in the overnight peritoneal effluents with a mean concentration of 2.39 \pm 1.87 ng/mL. Table 1 presents the clinical characteristics of the study.

3.2 Correlation between systemic and dialysate OPN and peritoneal transport characteristics

The OPN concentration in overnight peritoneal effluents was correlated with PSTR determined using PET. OPN levels were correlated with D/P Cr (p < 0.0001, r = 0.54) and D/D0 glucose (p < 0.0001, r = -0.39) as determined through Spearman's correlation coefficient test (Figure 1). In the blood samples, no firm correlation was observed between OPN levels and PET results (Table 2).

3.3 Independent factors associated with peritoneal transport characteristics

The patients were divided into either low and low– average transport (L/LA) group or high and high–average transport (H/HA) group based on the PET results. The OPN concentration in peritoneal effluents was considerably higher in the H/HA group than in the L/LA group (3.05 ± 1.94 vs 1.25 ± 1.03 ng/mL, p < 0.0001). The H/HA group was more likely to be older (51.57 ± 13.14 vs $44.41 \pm$



Figure 1: Correlation between OPN level in the peritoneal effluents and peritoneal transport characteristics. Peritoneal solute transport was assessed with the PET, and the levels of OPN in the overnight peritoneal effluents were also analyzed with ELISA. (a) The D/P Cr ratio versus the OPN level. (b) The D/D0 glucose ratio versus the OPN levels. (c) The 4 h ultrafiltration volume versus the OPN levels. D/P Cr, dialysate/plasma ratio of creatinine; D/D0 glucose, 4 to 0 h dialysate glucose; OPN, osteopontin.

12.31, p = 0.007) and had lower serum albumin content (36.80 ± 3.17 vs 33.16 ± 5.19 g/L, p < 0.0001) and lower serum phosphate (1.51 ± 0.41 vs 1.79 ± 0.58 mmol/L, p = 0.004) and effluent IL-6 levels (45.8 ± 22.04 vs 74.13 ± 35.51 pg/mL; Table 3). Logistic regression was employed to identify independent predictive factors for increased PSTR (H/HA). OPN or IL-6 in effluents, age, and serum albumin were independent predictive factors for the increased

Table 2: Correlation between OPN level in serum and peritoneal transport characteristics. Peritoneal solute transport was assessed with the PET, the levels of OPN in serum were analyzed with ELISA

		4 h D/ P Cr	D/D0 glucose	4 h ultrafiltration volume
Serum	r	0.02	-0.13	-0.16
OPN	p	0.77	0.65	0.74

Abbreviations: D/P Cr: dialysate/plasma ratio of creatinine; D/D0 glucose: 4 to 0 h dialysate glucose; OPN: Osteopontin.

PSTR, whereas PD duration, gender, and serum phosphate were not (Table 4). The diagnostic accuracy of

the markers in effluents measured for identifying the increased PSTR was examined. Relative analysis showed

that the area under the receiver operating characteristic (ROC) curve of IL-6 was 0.76 (confidence interval (CI):

0.67–0.85, p < 0.001) in predicting increased PSTR with a sensitivity of 65% and a specificity of 81%. The ROC

curve of OPN was 0.84 (95% CI: 0.75–0.92, p < 0.001)

in predicting increased PSTR with a sensitivity of 86%

and a specificity of 67%. The OPN-PSTR model was con-

structed by using four variables (age, serum albumin, effluent

OPN, and effluent IL-6). The model showed increased speci-

ficity (81%; Figure 2).

Table 4: A logistic regression model for the predictors of PSTR in PDpatients, adjustment for age, gender, PD duration, serum albumin,serum phosphate, effluent OPN, and effluent IL-6

	В	Wald	p value	OR	95% CI	
					Lower	Upper
Age	0.060	5.431	0.020	1.062	1.010	1.116
Albumin	-0.139	4.406	0.036	0.870	0.764	0.991
OPN	0.875	9.345	0.002	2.400	1.369	4.206
IL-6	0.037	9.062	0.003	1.038	1.013	1.063
Constant	-1.059	0.139	0.709	0.347		

Abbreviation: OPN: Osteopontin.

4 Discussion

4.1 Principal findings and comparison with other studies

This study showed correlations between OPN levels in dialysate and D/P Cr or D/D0 glucose, and the OPN level in effluents was an independent predictive factor for the increased PSTR. However, we did not observe obvious relevance between OPN levels in effluents and 4 h ultra-filtration volume. On the basis of the results of a large retrospective study that enrolled more than 10,000

Table 3: Data are presented as means \pm standard deviation (SD) or *n* (%)

Variable	L/LA (<i>n</i> = 37)	H/HA (<i>n</i> = 72)	p	
Male gender (%)	0.54	0.51	0.794	
Serum albumin (g/L)	36.80 ± 3.17	33.16 ± 5.19	<0.0001	
Hemoglobin (g/L)	100.47 ± 18.62	94.62 ± 15.21	0.081	
PD duration (months)	30.09 ± 24.50	41.03 ± 38.96	0.123	
Age (years)	44.41 ± 12.31	51.57 ± 13.14	0.007	
Serum phosphate (mmol/L)	1.79 ± 0.58	1.51 ± 0.41	0.004	
Serum calcium (mmol/L)	2.25 ± 0.29	2.16 ± 0.67	0.454	
Triglyceride (mmol/L)	2.08 ± 1.37	1.62 ± 1.29	0.084	
Cholesterol (mmol/L)	4.37 ± 0.99	4.45 ± 1.17	0.715	
Uric acid (µmol/L)	391.31 ± 82.40	394.07 ± 92.02	0.879	
D/P Cr	0.57 ± 0.06	0.75 ± 0.07	<0.0001	
D/D0 glucose	0.43 ± 0.06	0.33 ± 0.05	<0.0001	
Ultrafiltration volume (mL)	316.22 ± 121.40	247.21 ± 141.45	0.01	
Creatinine (µmol/L)	990.78 ± 333.16	944 ± 270.94	0.438	
PTH (pg/mL)	470.72 ± 260.12	394.44 ± 294.68	0.197	
Total Kt/V	1.85 ± 0.42	1.79 ± 0. 39	0.373	
Peritoneal Kt/V	1.44 ± 0.32	1.49 ± 0.37	0.440	
Renal Kt/V urea	0.43 ± 0.50	0.32 ± 0.40	0.174	
Serum OPN (ng/mL)	32.45 ± 13.03	37.82 ± 17.31	0.771	
Serum IL-6 (pg/mL)	29.33 ± 20.11	33.98 ± 17.77	0.892	
Effluent OPN (ng/mL)	1.25 ± 1.03	3.05 ± 1.94	<0.0001	
Effluent IL-6 (pg/mL)	$\textbf{45.80} \pm \textbf{22.04}$	74.13 ± 35.51	<0.0001	

Abbreviations: PD: peritoneal dialysis; D/P Cr: dialysate/plasma ratio of creatinine; D/D0 glucose: 4 to 0 h dialysate glucose; OPN: Osteopontin; PTH: parathyroid hormone. Bold letters and values indicate statistically significant difference.



Marker	best cutoff	Sensitivity	Specificity
OPN	1.78ng/m1	0.86	0.67
IL-6	60.89pg/m1	0.65	0.81
OPN-PSTR model	0.56	0.86	0.81

Figure 2: AUROC analyses of the predictors for identifying increased PSTR. OPN: Osteopontin; PSTR, peritoneal solute transport rate.

patients with PD subjected to PET, a strong inverse association was found between D/P Cr and D/D0 glucose (r =0.84), and a linear association existed between these parameters and the hospitalization rate or all-cause mortality [7,20]. However, only a modest correlation was observed between 4-hour ultrafiltration volume and D/P creatinine or D/D0 glucose, and in addition, no correlation existed between 4-hour ultrafiltration volume and all-cause mortality [7]. Thus, D/P Cr and D/D0 glucose are superior to 4-hour ultrafiltration volume in identifying the increased PSTR at risk of adverse clinical outcomes. The lower predictive value of 4 h ultrafiltration volume maybe due to the following reasons: (1) except PM transport function, the variable residual renal function of CAPD patients is also an essential determinant of the 4 h ultrafiltration volume. (2) Since commercially available PD bags contain more dialysate volume than prefilling flushing, this is variable, resulting in inaccurate measurement [21]. Taken together, effluent OPN may be an excellent predictor for PET results and prognosis of patients with PD.

In this study, 66% of the patients followed up in our center were categorized into the H/HA group based on the PET results; this finding is similar to those reported by other multicenter studies (47.4–83%) [22–24]. The

comparison of H/HA and L/LA groups showed statistically significant differences between serum albumin and serum phosphate levels. Various studies reported that high peritoneal transport status is accompanied by high baseline albumin clearance [7,23,25,26]. Moreover, patients with fast transport present with severe systemic inflammation status, and pro-inflammatory cytokines could cause muscle wasting and suppress appetite [27,28]. Thus, patients with CAPD in the H/HA group had lower serum albumin compared with those in the L/LA group. Phosphate transport across the peritoneum is regulated by osmotic, chemical, and electrical gradients as well as by transmembranous active phosphate transporters; thus, this process is complex. Bernardo et al. reported that phosphate clearance increased in fast transporters compared with that in slow transporters in patients with CAPD and APD [29]; this finding is consistent with our results.

Increasing lines of evidence show that chronic inflammation might be an initial factor for PM injury. After prolonged PD treatment, MCs and peritoneal macrophages produce a wide array of inflammatory cytokines, such as IL-1 β , tumor necrosis factor- α (TNF- α), and IL-6, eventually leading to PM damage and functional abnormalities [5,27,30]. The secretion of OPN is enhanced in the inflammatory microenvironment, whereas TNFa and IL-1ß stimulate the transcription and the release of OPN genes [13]. Consistent with the previous studies, IL-6 levels in effluents were higher in the H/HA group than that in the L/LA group and could be an independent predictive factor for increased PSTR. No extreme change in the serum OPN levels was found as the PSTR increased. Overall, the high OPN concentration in the H/HA group may be attributed to the high number of pro-inflammatory cytokines released from the PM. The cellular source of PM, where OPN is synthesized and released, merits further investigation.

4.2 Limitations of study

This study features several important limitations. First, the cross-sectional nature of the study excluded the establishment of causal relationships and temporal trends between OPN levels in effluents and PSTR. Second, we did not investigate the association between OPN levels in effluents and adverse outcomes in PD. Third, this singlecenter study involved a small sample size, and selection bias might be present considering that a large number of subjects were excluded due to the strict inclusion and exclusion criteria.

4.3 Summary

In conclusion, we established the associations between OPN levels in the drained dialysate and D/P Cr and D/D0 glucose. Effluent OPN is also an independent predictive factor for the increased PSTR. The OPN-PSTR model, constructed by using four variables (age, serum albumin, effluent OPN, and effluent IL-6) demonstrated good diagnostic performance for accurately identifying increased PSTR.

Abbreviations

PD	peritoneal dialysis
OPN	Osteopontin
CAPD	continuous ambulatory PD
PET	peritoneal equilibration test
PSTR	peritoneal solute transport rate
ROC	receiver operating characteristic
PM	peritoneal membrane
EMT	epithelial-mesenchymal transition
IL-6	interleukin 6
CA125	cancer antigen 125
VEGF	vascular endothelial growth factor
PAI-1	plasminogen activator inhibitor-1
MMP	matrix metalloproteinase
D/P Cr	dialysate/plasma ratio of creatinine
D/D0 glucose	4-to 0-hour dialysate glucose

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Conflict of interest: The authors declare that they have no conflict of interest.

Data availability statement: The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

References

- Li PK, Chow KM, Van de Luijtgaarden MW, Johnson DW, Jager KJ, Mehrotra R, et al. Changes in the worldwide epidemiology of peritoneal dialysis. Nat Rev Nephrol. 2017;13:90–103.
- [2] Zimmerman AM. Peritoneal dialysis: increasing global utilization as an option for renal replacement therapy. J Glob Health. 2019;9:020316.
- [3] Mehrotra R, Devuyst O, Davies SJ, Johnson DW. The current state of peritoneal dialysis. J Am Soc Nephrol. 2016;27:3238–52.
- [4] Williams JD, Craig KJ, Topley N, Von Ruhland C, Fallon M, Newman GR, et al. Morphologic changes in the peritoneal membrane of patients with renal disease. J Am Soc Nephrol. 2002;13:470–9.
- [5] Twardowski ZJ. Pathophysiology of peritoneal transport. Contrib Nephrol. 2006;150:13–9.
- [6] Yanez-Mo M, Lara-Pezzi E, Selgas R, Ramirez-Huesca M, Dominguez-Jimenez C, Jimenez-Heffernan JA, et al. Peritoneal dialysis and epithelial-to-mesenchymal transition of mesothelial cells. N Engl J Med. 2003;348:403–13.
- [7] Mehrotra R, Ravel V, Streja E, Kuttykrishnan S, Adams SV, Katz R, et al. Peritoneal equilibration test and patient outcomes. Clin J Am Soc Nephrol. 2015;10:1990–2001.
- [8] Lopes Barreto D, Krediet RT. Current status and practical use of effluent biomarkers in peritoneal dialysis patients. Am J Kidney Dis. 2013;62:823–33.
- [9] Kawanishi H, Fujimori A, Tsuchida K, Takemoto Y, Tomo T, Minakuchi J, et al. Markers in peritoneal effluent for withdrawal from peritoneal dialysis: multicenter prospective study in Japan. Adv Perit Dial. 2005;21:134–8.
- [10] Hirahara I, Inoue M, Umino T, Saito O, Muto S, Kusano E. Matrix metalloproteinase levels in the drained dialysate reflect the peritoneal solute transport rate: a multicentre study in Japan. Nephrol Dial Transplant. 2011;26:1695–701.
- [11] Hirahara I, Inoue M, Okuda K, Ando Y, Muto S, Kusano E. The potential of matrix metalloproteinase-2 as a marker of peritoneal injury, increased solute transport, or progression to encapsulating peritoneal sclerosis during peritoneal dialysis – a multicentre study in Japan. Nephrol Dial Transplant. 2007;22:560–7.
- [12] Hirahara I, Kusano E, Imai T, Morishita Y, Inoue M, Akimoto T, et al. Effluent Tenascin-C levels reflect peritoneal deterioration in peritoneal dialysis: major in PD study. Biomed Res Int. 2015;2015:241098.
- [13] Shurin MR. Osteopontin controls immunosuppression in the tumor microenvironment. J Clin Invest. 2018;128:5209–12.
- [14] Kothari AN, Arffa ML, Chang V, Blackwell RH, Syn WK, Zhang J, et al. Osteopontin-A master regulator of epithelial-mesenchymal transition. J Clin Med. 2016;5:39.
- [15] Icer MA, Gezmen-Karadag M. The multiple functions and mechanisms of Osteopontin. Clin Biochem. 2018;59:17–24.
- [16] Giachelli CM, Steitz S. Osteopontin: a versatile regulator of inflammation and biomineralization. Matrix Biol. 2000;19:615-22.
- [17] Clemente N, Raineri D, Cappellano G, Boggio E, Favero F, Soluri MF, et al. Osteopontin bridging innate and adaptive immunity in autoimmune diseases. J Immunol Res. 2016;2016:7675437.

- [18] Trueblood NA, Xie Z, Communal C, Sam F, Ngoy S, Liaw L, et al. Exaggerated left ventricular dilation and reduced collagen deposition after myocardial infarction in mice lacking Osteopontin. Circ Res. 2001;88:1080–7.
- [19] Nakazato Y, Yamaji Y, Oshima N, Hayashi M, Saruta T. Calcification and Osteopontin localization in the peritoneum of patients on long-term continuous ambulatory peritoneal dialysis therapy. Nephrol Dial Transplant. 2002;17:1293–303.
- [20] Brimble KS, Walker M, Margetts PJ, Kundhal KK, Rabbat CG. Metaanalysis: peritoneal membrane transport, mortality, and technique failure in peritoneal dialysis. J Am Soc Nephrol. 2006;17:2591–8.
- [21] La Milia V, Pozzoni P, Crepaldi M, Locatelli F. Overfill of peritoneal dialysis bags as a cause of underestimation of ultrafiltration failure. Perit Dial Int. 2006;26:503–5.
- [22] Agarwal DK, Sharma AP, Gupta A, Sharma RK, Pandey CM, Kumar R, et al. Peritoneal equilibration test in Indian patients on continuous ambulatory peritoneal dialysis: does it affect patient outcome? Adv Perit Dial. 2000;16:148–51.
- [23] Chung SH, Chu WS, Lee HA, Kim YH, Lee IS, Lindholm B, et al. Peritoneal transport characteristics, comorbid diseases and survival in CAPD patients. Perit Dial Int. 2000;20:541–7.
- [24] Sezer S, Tutal E, Arat Z, Akcay A, Celik H, Ozdemir FN, et al. Peritoneal transport status influence on atherosclerosis/ inflammation in CAPD patients. J Ren Nutr. 2005;15:427–34.

- [25] John B, Tan BK, Dainty S, Spanel P, Smith D, Davies SJ. Plasma volume, albumin, and fluid status in peritoneal dialysis patients. Clin J Am Soc Nephrol. 2010;5:1463–70.
- [26] Lim PS, Chen HP, Chen CH, Wu MY, Wu CY, Wu TK. Association between redox status of serum albumin and peritoneal membrane transport properties in patients on peritoneal dialysis. Blood Purif. 2015;40:243–9.
- [27] Krediet RT, Struijk DG. Peritoneal changes in patients on longterm peritoneal dialysis. Nat Rev Nephrol. 2013;9:419–29.
- [28] Churchill DN, Thorpe KE, Nolph KD, Keshaviah PR, Oreopoulos DG, Page D. Increased peritoneal membrane transport is associated with decreased patient and technique survival for continuous peritoneal dialysis patients. The Canada-USA (CANUSA) peritoneal dialysis study group. J Am Soc Nephrol. 1998;9:1285–92.
- [29] Bernardo AP, Contesse SA, Bajo MA, Rodrigues A, Del Peso G, Ossorio M, et al. Peritoneal membrane phosphate transport status: a cornerstone in phosphate handling in peritoneal dialysis. Clin J Am Soc Nephrol. 2011;6:591–7.
- [30] Gao D, Zhao ZZ, Liang XH, Li Y, Cao Y, Liu ZS. Effect of peritoneal dialysis on expression of vascular endothelial growth factor, basic fibroblast growth factor and endostatin of the peritoneum in peritoneal dialysis patients. Nephrology (Carlton). 2011;16:736–42.